

INFLUENCE OF MICROBIAL INOCULANT ON THE OVIPOSITION AND
COMPLETION OF LIFE CYCLE OF *Chrysomya megacephala* AND *Chrysomya*
rufifacies INFESTING RABBIT CARCASSES

NUR AMALINA BINTI OMAR

A dissertation submitted in fulfilment of
the requirements for the awards of the degree of
Master of Science (Forensic Science)

Faculty of Science
Universiti Teknologi Malaysia

AUGUST 2018

Dedicated to my beloved parents and siblings.

ACKNOWLEDGEMENT

First and foremost, Alhamdulillah, I thank Allah S.W.T for blessing me with the strength and courage to complete my master's degree study. I would like to deliver my appreciation to my beloved father Omar Yusoff, mother Roslina Abdullah and my siblings for the endless supports given to me. Thank you for watching me completing my study with great patience and understanding.

I would like to express my deepest gratitude to my supervisor, Dr. Naji Arafat Mahat for his guidance, encouragement and critiques from the commence of the research until the writing phase that further improve the outcomes. This appreciation also goes to my co-supervisor, Dr. Heo Chong Chin for enlightening me with any shortcomings and furnished this research with relevant suggestions. For the time spent to review this writing from the beginning to the end, I could not have thanked you both enough.

Millions of thanks to my fellow classmates and seniors who have stick with me throughout this journey and assist me in any ways they can, lending me ears to listen and shoulders to rely on. Finally, I am grateful to all lecturers and staff from the Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia (UTM) for their advice and help. The friendliness and warmth were felt throughout my time here. Not to mention the well-equipped facilities within the faculty which have provided me with a convenient space for conducting this research.

ABSTRACT

Considering any surrounding factors as well as the cunning use of microbial inoculants to cover the smell of decomposing bodies could potentially alter (delay or speed up) the time of initial oviposition and developmental rate of insects infesting the corpses, thereby resulting in erroneous estimation of post mortem interval (PMI); such factors led to the initiation of this research. This research was conducted to assess the influence of a commercially available microbial inoculant (EM.1[®]) on the initial oviposition and developmental rate that lead to completion of life cycle of the two prevalent necrophagous flies (*Chrysomya megacephala* and *Chrysomya rufifacies*) in Malaysia infesting rabbit carcasses. In this research, nine rabbit carcasses were equally divided into control (C) and two treated (T1 and T2) groups. The T1 and T2 carcasses were individually sprayed with two different concentrations at 1:500 and 1:100 of the same microbial inoculant, respectively. The durations taken for each species of fly to first oviposit and complete their life cycle in treated carcasses were compared with that of control carcasses. Results revealed that single application of the microbial inoculant on both the treated carcasses did not statistically impede oviposition of both species as well as their subsequent developmental patterns (Kruskall-Wallis H: $P > 0.05$) when compared to the control carcasses. Since this is the first research that reported on the time of initial oviposition and completion of life cycle for *C. megacephala* and *C. rufifacies* in the presence of microbial inoculant, the results observed here may prove useful for estimating PMI in Malaysia particularly in cases where the use of microbial inoculant was suspected.

ABSTRAK

Mengambil kira sebarang faktor di sekeliling serta inokulan mikrob yang boleh digunakan dengan licik oleh pembunuh untuk melindungi bau mayat yang mereput berpotensi untuk mengubah (melewatkan atau mempercepat) masa permulaan pengovipositan dan kadar perkembangan serangga yang menginfestasi mayat, seterusnya mengakibatkan penganggaran sela-masa kematian (PMI) yang tidak tepat; faktor-faktor tersebut membawa kepada permulaan penyelidikan ini. Penyelidikan ini dijalankan untuk menilai pengaruh inokulan mikrob yang boleh didapati secara komersial (EM.1[®]) ke atas permulaan pengovipositan dan kadar perkembangan yang membawa kepada kitaran hidup lengkap bagi dua lalat nekrofagus yang lazim di Malaysia (*Chrysomya megacephala* dan *Chrysomya rufifacies*) yang menginfestasi bangkai arnab. Dalam penyelidikan ini, sembilan bangkai arnab telah dibahagikan kepada kumpulan kawalan (C) dan dua kumpulan dirawat (T1 dan T2). Bangkai-bangkai T1 dan T2 disemur dengan inokulan mikrob yang sama pada dua kepekatan berbeza iaitu 1:500 dan 1:100, masing-masing. Tempoh yang diambil oleh setiap spesies lalat untuk ovipositi yang pertama dan melengkapkan kitaran hidup mereka pada bangkai dirawat dibandingkan dengan bangkai kawalan. Hasil penyelidikan menunjukkan bahawa aplikasi tunggal inokulan mikrob pada kedua-dua bangkai yang dirawat secara statistiknya tidak menjejaskan pengovipositan kedua-dua spesies serta corak perkembangan mereka (Kruskall-Wallis H: $P > 0.05$) apabila dibandingkan dengan bangkai kawalan. Oleh kerana ini adalah penyelidikan pertama yang melaporkan mengenai permulaan pengovipositan dan kitaran hidup lengkap untuk *C. megacephala* dan *C. rufifacies* dengan kehadiran inokulan mikrob, dapatan yang diperoleh di sini mungkin berguna untuk menganggarkan PMI di Malaysia terutamanya di dalam kes di mana penggunaan inokulan mikrob disyaki.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	x
	LIST OF FIGURES	xi
	LIST OF ABBREVIATIONS	xiii
	LIST OF APPENDICES	xiv
1	INTRODUCTION	
	1.1 Background of Study	1
	1.2 Problem Statement	3
	1.3 Objectives and Hypotheses	4
	1.4 Scope of Study	5
	1.5 Significance of Study	5
2	LITERATURE REVIEW	
	2.1 Forensic Entomology	6
	2.1.1 Forensic Entomology in Malaysia	7
	2.1.2 Forensic Entomology for Estimating PMI	8

2.2	Life Cycle of Fly	9
2.3	Factors Influencing Insects Colonization, Oviposition and Development	12
2.3.1	Insects Attraction to Decomposing Corpse/ Carrion	12
2.3.2	Factors Affecting Oviposition and Development of Insects	12
2.4	Microbe-Insect Interactions	14
2.4.1	EM.1 Microbial Inoculant	15
2.4.2	Influence of Bacterial Isolates on Oviposition and Larval Development	16
3	MATERIALS AND METHOD	
3.1	Materials	18
3.1.1	Instruments and Apparatus	18
3.1.2	Solutions/ Chemicals	20
3.2	Methodology	21
3.2.1	Experimental Design	21
3.2.2	Decomposition Site and Entomological Observation	25
3.2.3	Sample Collection and Preservation	26
3.2.4	Mounting of Larvae and Taxonomic Identification	27
3.3	Statistical Analysis	28
4	RESULTS AND DISCUSSION	
4.1	Temperature and Rainfall at the Decomposition Site	29
4.2	Taxonomic Identification	34
4.3	Influences of Microbial Inoculant on Initial Oviposition and Completion of Life Cycle of <i>C. megacephala</i> and <i>C. rufifacies</i>	38

5	CONCLUSION	
	5.1 Conclusions	45
	5.2 Limitations and Recommendations	46
	REFERENCES	47
	Appendix A	54

LIST OF TABLES

TABLE NO.	TITLE	PAGE
3.1	List of instruments used.	19
3.2	List of apparatus used.	19
3.3	List of chemicals used.	20
3.4	Details of rabbit carcasses that formed one set.	22
4.1	Daily ambient temperature during the three replicate experiments.	30
4.2	Overall ambient temperature and rainfall for the three replicate experiments.	31
4.3	Day and time for the first observation of the different stages of life cycle for <i>C. megacephala</i> in control, treated 1 and treated 2 carcasses for all three replicate experiments.	40
4.4	Day and time for the first observation of the different stages of life cycle for <i>C. rufifacies</i> in control, treated 1 and treated 2 carcasses for all three replicate experiments.	41
4.5	Comparison of the durations for initial oviposition and the onset of other stages of life cycle for <i>C. megacephala</i> and <i>C. rufifacies</i> among the three groups of carcasses.	42

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
1.1	Research conceptual framework.	4
2.1	Life cycle of a blowfly; a) eggs, b) larvae, c) pupae, d) adult.	10
2.2	Spiracular slits in the posterior spiracles of (a) 1st instar, (b) 2nd instar and (c) 3rd instar larvae of <i>Chrysomya</i> species.	11
3.1	The commercially available microbial inoculant used in this present research.	21
3.2	Decomposition site and the distance between two carcasses.	23
3.3	A rabbit carcass enclosed with slotted plastic basket with 3-4 bricks on top.	23
3.4	Flowchart of experimental design.	24
3.5	Location of the decomposition site within UTM, Johor Bahru Campus.	26
4.1	Data on daily total rainfall (mm) recorded outside the decomposition site during the three replicate experiments.	32
4.2	Taxonomic traits for identifying adult fly of <i>Chrysomya megacephala</i> using keys provided by Kurahashi <i>et al.</i> (1997) and Nazni <i>et al.</i> (2011b).	35

4.3	Taxonomic traits for identifying adult fly of <i>Chrysomya rufifacies</i> using keys provided by Kurahashi <i>et al.</i> (1997) and Nazni <i>et al.</i> (2011b).	36
4.4	Taxonomic traits for identifying third instar larva of <i>Chrysomya megacephala</i> using keys provided by Omar (2002)	37
4.5	Taxonomic traits for identifying third instar larva of <i>Chrysomya rufifacies</i> using keys provided by Omar (2002).	39

LIST OF ABBREVIATIONS

a.m.	-	Ante meridiem
ANOVA	-	Analysis of Variance
C	-	Control
<i>C. megacephala</i>	-	<i>Chrysomya megacephala</i>
<i>C. rufifacies</i>	-	<i>Chrysomya rufifacies</i>
DC	-	Dorsal cornu
EM	-	Effective microorganisms
G	-	Gena
KOH	-	Potassium hydroxide
mPMI	-	Minimum Postmortem interval
PG	-	Postgena
PE	-	Peritreme
p.m.	-	Post meridiem
PMI	-	Postmortem interval
S	-	Spiracular slits
SD	-	Standard deviation
T1	-	Treated 1 (1:500)
T2	-	Treated 2 (1:100)
VC	-	Ventral cornu
VOC	-	Volatile organic compound
<i>Viz.</i>	-	Videlicet (namely)

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	SPSS Output	56

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Application of entomology in death investigations involves interpretation of the insects' developmental data, particularly for providing accurate estimation of minimum post mortem interval (mPMI) (Mahat *et al.*, 2014). In addition, entomological evidence is also useful for revealing the possibility of postmortem relocation of bodies, suggesting the possible cause of death as well as identification of the deceased (Gennard, 2007; Goff, 2009; Mahat and Jayaprakash, 2013). Forensic entomological evidence becomes particularly important in cases which the time since death is more than 72 hours, whereby during this period pathological changes (e.g. rigor mortis, algor mortis, and livor mortis) become less reliable for estimating PMI (Gennard, 2007). This can be done by examining the necrophagous insects infesting the corpse, observing their current life cycle stage, calculating the age of their immatures (e.g. instar larvae), and subsequently estimate the time when female flies first arrived and laid eggs (Rajagopal *et al.*, 2008). Pertinently, the accuracy of mPMI estimation is based on the ability to accurately identify the oldest necrophagous insect species (particularly Calliphoridae), prior to utilizing the appropriate growth data.

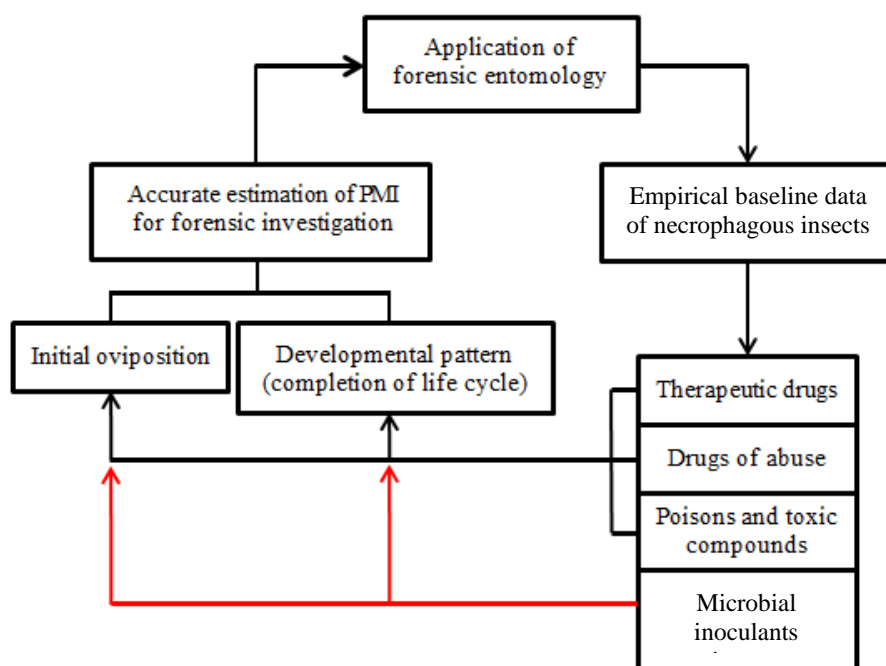
In Malaysia, *Chrysomya megacephala* (Fabricius) has been identified as the first and dominant necrophagous species infesting corpses and carcasses, followed by *Chrysomya rufifacies* (Macquart) (Mahat *et al.*, 2016). There are many factors that can affect initial oviposition, development and succession of species that include the biogeoclimatic factors (e.g. rain) (Mahat *et al.*, 2009), presence of therapeutic drugs and toxic substances (e.g. paracetamol) (O'brien and Turner, 2004), as well as physical barriers such as covering or wrapping (Goff, 1992). Review of literature reveals that while chemicals such as malathion may delay initial oviposition and prolong the development of blowfly species (Mahat *et al.*, 2009), faster development of necrophagous insects has been associated with cocaine (Goff *et al.*, 1989) and diazepam (Carvalho *et al.*, 2001). Besides, there are also chemicals (e.g. bleach) that do not affect the development of necrophagous insects (Auberton *et al.*, 2015). In this context, it is pertinent to indicate here that any factors that alter oviposition and development pattern of necrophagous insects would subsequently result in erroneous estimation of PMI. Hence, exploring the different chemicals and/or formulations that are commonly available in the market, in view of their potential influences on the oviposition and duration for completing life cycle among necrophagous insects, merits forensic consideration.

Microbial inoculants have long been widely used in farming and other agricultural activities for promoting livestock's growth, increasing nutrient level in soil, as well as improving waste management and sanitation (Kyan *et al.*, 1999). Often, livestock rearing farms emanate offensive odor from the waste or manure that can adversely affect the environment that humans live in, creating inconveniences. To overcome this issue, the use of microbial inoculants that contain effective microorganisms (EM) that are proven to be effective odor reducing agents for the environment has been suggested (Sakawi and Ismail, 2015). Some of the selected species of microorganisms included in microbial inoculants that are widely sold in the market are lactic acid bacteria (*Lactobacillus casei*), photosynthetic bacteria (*Rhodospseudomonas palustris*) and yeast (Higa and Parr, 1994). This has attracted a specific attention to forensic entomologists since alteration in odor may indirectly alter the activities of flies in the surrounding areas. Therefore, when applied in forensic entomology context, it means that microbial inoculants can be one of the

contributing factors that may affect oviposition of necrophagous flies on corpse found in the area where these agents are applied in its surrounding. Moreover, the fact that microbial inoculants can also be cunningly used by the murderer to reduce the odor of a decaying body to confuse forensic entomological assessments of mPMI cannot be excluded.

1.2 Problem Statement

The accuracy of estimating PMI relies largely on understanding the intrinsic and extrinsic factors that can alter the oviposition and developmental pattern of necrophagous insects. Because scientific information has already become a public domain, and since criminals nowadays are getting better-informed, the possibility that these cunning criminals may resort to the use of various chemicals/ formulations for delaying necrophagous insect activity cannot be ruled out. Leaving alone its routine application for various purposes such as controlling bad odor in animal farming, microbial inoculants that consist of various EM can be further utilized by murderers to confuse forensic entomologists at estimating mPMI. This can subsequently result in the miscarriage of justice. The fact that such agents are readily accessible in the market and since review of literature does not reveal any specific study focusing on its influence on oviposition and development pattern of necrophagous insects, this specific research that was designed to elucidate such aspect on *C. megacephala* and *C. rufifacies* appears forensically relevant. Based on the current information available, it is expected that the initial oviposition and completion of life cycle for *C. megacephala* and *C. rufifacies* may be prolonged in carcasses sprayed with EM.1[®] Microbial Inoculant. The conceptual framework depicting the gap of knowledge in the body of literature examined in this present research is presented in Figure 1.1.



Note: Black line represents the established forensic entomology empirical baseline data in Peninsular Malaysia, while the red one indicates the gap of knowledge addressed by this present research.

Figure 1.1: Research conceptual framework.

1.3 Objectives and Hypotheses

The aim of this research was to investigate the influence of commercially available microbial inoculant on initial oviposition and development of *C. megacephala* and *C. rufifacies* using rabbit carcasses. Specifically, the objective of this research was to determine and compare the initial oviposition and duration for completing life cycles for *C. megacephala* and *C. rufifacies* infesting rabbit carcasses sprayed with two different concentrations of a microbial inoculant with that of control carcasses. It was hypothesized that significant differences in time of initial oviposition and completion of life cycle for *C. megacephala* and *C. rufifacies* shall be observed among the treated and control groups.

REFERENCES

- Abd Rashid, R., Osman, K., Ismail, M. I., Zuha, R. M. & Abu-Hassan, R. (2008). Determination of malathion levels and the effect of malathion on the growth of *Chrysomya megacephala* (Fabricius) in malathion-exposed rat carcasses. *Tropical Biomedicine*, 25(3), 184-190.
- Ab Muttalib, S. A., Syed Ismail, S. N., & Praveena, S. M. (2016). Application of effective microorganism (EM) in food waste composting: a review. *Asia Pacific Environmental and Occupational Health Journal*, 2(1).
- Adair, T. (2012). Aspects influencing the entomological postmortem interval in crime scene reconstruction, *J Assoc Crime Scene Reconstr*, 18(3), 17-19.
- Adams, Z. J. O., & Hall, M. J. R. (2003). Methods used for the killing and preservation of blowfly larvae, and their effect on post-mortem larval length. *Forensic Science International*, 138, 50–61.
- Ahmad, A., Broce, A., & Zurek, L. (2006). Evaluation of significance of bacteria in larval development of *Cochliomyia macellaria* (Diptera: Calliphoridae). *Journal of Medical Entomology*, 43(6), 1129-1133.
- Ahmad, A. & Ahmad, A.B. (2009). A preliminary study on the decomposition and dipteran associated with exposed carcasses in an oil palm plantation in Bandar Baharu, Kedah, Malaysia. *Tropical Biomedicine*, 26(1), 1-10.
- Ahmad, N. W., Lim, L. H., Dhang, C. C., Chin, H. C., Ag, A., Wan, W., & Azirun, S. M. (2011). Comparative insect fauna succession on indoor and outdoor monkey carrions in a semi-forested area in Malaysia. *Asian Pacific Journal of Biomedicine*, 32–38.
- Amendt, J., Krettek, R. & Zehner, R. (2004). Forensic entomology. *Naturwissenschaften*, 91, 51-65.
- Amendt, J., Campobasso C. P., Gaudry, E., Reiter, C., LeBlanc, H. N. & Hall, M. J. R. (2007). Best practice in forensic entomology- standards and guidelines. *International Journal of Legal Medicine*, 121(2), 90–104.
- Amendt, J., Richards, C. S., Campobasso, C. P., Zehner, R., & Hall, M. J. (2011). Forensic entomology: applications and limitations. *Forensic Science, Medicine and Pathology*, 7(4), 379-392.
- Anderson, G. S. (2010). Factors that influence insect succession on carrion. In: J. H., Byrd, & J. L., Castner, eds. *Forensic Entomology: The Utility of Arthropods*

- in Legal Investigation 2nd edition*. Boca Raton, Florida: CRC Press. pp 201-250.
- Ashworth, J. R., & Wall, R. (1994). Responses of the sheep blowflies *Lucilia sericata* and *L. cuprina* to odour and the development of semiochemical baits. *Medical and Veterinary Entomology*, 8(4), 303-309.
- Auberon, C., Devigne, C., Hedouin, V., Gosset, D., & Charabidze, D. (2015). In vitro effects of household products on Calliphoridae larvae development: implication for forensic entomology. *Journal of Forensic Sciences*, 60(1), 226-232.
- Azwandi, A., Nina Katerina, H., Owen, L.C., Nurizzati, M.D. & Omar, B. (2013). Adult carrion arthropod community in a tropical rainforest of Malaysia: analysis on three common forensic entomology animal models. *Tropical Biomedicine*, 30(3), 481-494.
- Badenhorst, R., & Villet, M. H. (2018). The uses of *Chrysomya megacephala* (Fabricius, 1794) (Diptera: Calliphoridae) in forensic entomology. *Forensic Sciences Research*, 3(1), 2-15.
- Barnes, K. M., & Gennard, D. E. (2011). The effect of bacterially-dense environments on the development and immune defences of the blowfly *Lucilia sericata*. *Physiological Entomology*, 36(1), 96-100.
- Beans, C. (2018). News Feature: can microbes keep time for forensic investigators? *Proceedings of the National Academy of Sciences*, 115(1), 3-6.
- Bourel, B. (2003). Flies eggs: a new method for the estimation of short-term postmortem interval? *Forensic Science International*, 135, 27–34.
- Campobasso, C. P., Di Vella, G., & Introna, F. (2001). Factors affecting decomposition and Diptera colonization. *Forensic Science International*, 120(1), 18-27.
- Carvalho, L. M., Linhares, A. X., & Trigo, J. R. (2001). Determination of drug levels and the effect of diazepam on the growth of necrophagous flies of forensic importance in southeastern Brazil. *Forensic Science International*, 120(1), 140-144.
- Charabidze, D., Bourel, B., Hedouin, V., & Gosset, D. (2009). Repellent effect of some household products on fly attraction to cadavers. *Forensic Science International*. 189, 28–33.

- Clarkson, C. A., Hobischak, N. R., & Anderson, G. S. (2004). A comparison of the developmental rate of *Protophormia terraenovae* (Robineau-Desvoidy) raised under constant and fluctuating temperature regimes. *Can. Soc. Forensic Science Journal*, 37(2), 95-101.
- Dadour I. R., Cook D. F., Fissioli J. N., & Bailey W.J. (2001). Forensic entomology: application, education and research in Western Australia. *Forensic Science International*, 120, 48–52.
- Gennard, D. E. (2007). *Forensic Entomology*, New Jersey: John Wiley and Sons.
- Goff, M. L. (1992). Problems in Estimation of Postmortem Interval Resulting from Wrapping of the Corpse: A Case Study from Hawaii, *Journal of Agric. Entomology*, 9(4), 237-243.
- Goff, M. L. (2009). Forensic Entomology. In: Resh, V.H. and Cardé, R.T. (editors). *Encyclopedia of Insects*. *Academic Press*. pp. 381-386.
- Greenberg, B. & Kunich, J. C. (2002). Entomology and the law – Flies as forensic indicator. *Cambridge, U.K: Cambridge University Press*.
- Heo, C. C., Marwi, M. A., Mohd Salleh, A. F., Jeffery, J. & Omar, B. (2007). A preliminary study of insect succession on a pig carcass in a palm oil plantation in Malaysia. *Tropical Biomedicine*, 24(2), 23-27
- Heo, C. C., Marwi, M. A., Jeffery, J. & Omar, B. (2008a). Insect succession on a decomposing piglet carcass placed in a man-made freshwater pond in Malaysia. *Tropical Biomedicine*, 25(1), 23-29.
- Heo, C. C., Marwi, M. A., Mohd. Salleh, A. F., Jeffery, J., Kurahashi, H. & Omar, B. (2008b). Study of insect succession and rate of decomposition on a partially burned pig carcass in an oil palm plantation in Malaysia. *Tropical Biomedicine*, 25(3), 202–208.
- Heo, C. C., Sulaiman, S., Othman, H., Jeffery, J., Kurahashi, H. & Omar, B. (2010). Insect succession associated with a hanging pig carcass placed in an oil palm plantation in Malaysia. *Sains Malaysiana*, 39(6), 921-926.
- Higa, T., & Parr, J. F. (1994). Beneficial and effective microorganisms for a sustainable agriculture and environment (Vol. 1). *International Nature Farming Research Center, Japan*.
- Hough–Goldstein, J. A., & M. A. Bassler. 1988. Effects of bacteria on oviposition by seed corn maggots (Diptera: Anthomyidae). *Environ. Entomol*, 17, 7-12.

- Kavitha, R., Nazni, W.A., Tan, T.C. Lee, H.L. & Azirun, M.S. (2013). Review of forensically important entomological specimens collected from human cadavers in Malaysia, *Journal of Forensic and Legal Medicine*, 20, 480-482.
- Kokdener, M. (2016). Application of entomology in forensic sciences. *Turkish Bulletin of Entomology*, 6(3), 269-275.
- Kurahashi, H. Benjaphong, N. & Omar, B. (1997). Blowflies (Insecta: Diptera: Calliphoridae) of Malaysia and Singapore. *Raffles Bulletin Zoology*; 5(Supplement), 1-88.
- Kyan, T., Shintani, M., Kanda, S., Sakurai, M., Ohashi, H., Fujisawa, A., & Pongdit, S. (1999). Kyusei nature farming and the technology of effective microorganisms. *Bangkok, TH, International Nature Farming Research Center, Atami, Japan and Asia Pacific Natural Agricultural Network 44p*.
- Lee, H. L. (1989). Recovery of Forensically Important Entomological Specimens from Human Cadavers in Malaysia-an update. *Mal. J. Pathology*, 11, 33-36.
- Lee, H. L., Krishnasamy, M., Abdullah, A. G., & Jeffery, J. (2004). Review of Forensically Important Entomological Specimens in the Period of 1972-2002. *Tropical Biomedicine*, 1, 69-75.
- Li, W., & Ni, Y. (2001). Use of effective microorganisms to suppress malodors of poultry manure, *Journal of Crop Production*, 3(1), 215-221.
- Lord, W. D., & Rodriguez, W. C. (1989). Forensic entomology: the use of insects in the investigation of homicide and untimely death. *The Prosecutor. Winter*, 41-48.
- Mahat, N. A., Zafarina, Z., & Jayaprakash, P. T. (2009). Influence of Rain and Malathion on the Oviposition and Development of Blowflies (Diptera: Calliphoridae) infesting rabbit carcasses in Kelantan, Malaysia. *Forensic Science International*, 192, 19-28.
- Mahat, N.A. & Jayaprakash, P.T. (2013). Forensic Entomology in Malaysia. *Malaysian Journal of Forensic Sciences*. Vol. 4, No. 1.
- Mahat, N. A., Yin, C. L., & Jayaprakash, P. T. (2014). Influence of Paraquat on *Chrysomya megacephala* (Fabricius) (Diptera: Calliphoridae) Infesting Minced-beef Substrates in Kelantan, Malaysia. *Journal of Forensic Sciences*, 59(2), 529-532.
- Mahat, N. A., Zainol-Abidin, N. L., Nordin, N. H., Abdul-Wahab, R. & Jayaprakash, P. T. (2016). Pattern of oviposition and development of *Chrysomya*

- megacephala* (Fabricius) (Diptera: Calliphoridae) and *Chrysomya rufifacies* (Macquart) (Diptera: Calliphoridae) on burned rabbit carcasses. *Forensic Science International*, 260, 9-13.
- Mohd Salleh, A. F., Talib, A., Marwi, M. A., Mohd-Isa, N. H., Abdullah, S. R., RajaKamal-Bashah, R. M. Z. & Omar, B. (2009). Effects of temperatures on larval development of *Chrysomya megacephala* (Fabricius) and *Chrysomya rufifacies* (Macquart) (Diptera: Calliphoridae): application in forensic science. *Malaysian Journal of Health Sciences*, 7(2), 89-96.
- Nazni, W.A., Lim, L. H., Dhang, C. C., Chin, H. C., Abdullah, A. G., Wan Norjuliana, W. M., Kian, C. W., Jeffery, J., Hashim, R. & Azirun, S.M. (2011a). Comparative insect fauna succession on indoor and outdoor monkey carrions in a semi-forested area in Malaysia. *Asian Pacific Journal of tropical Biomedicine*. S232-S238.
- Nazni, W. A., Jeffrey, J., Heo, C. C., Chew, W. K. & Lee, H. L. (2011b). Illustrated key to adult flies of forensic importance in Malaysia. *Institute for Medical Research, Kuala Lumpur, Malaysia*. Bulletin No. 25.
- O'Brien, C., & Turner, B. (2004). Impact of paracetamol on *Calliphora vicina* larval development. *International Journal of Legal Medicine*, 118(4), 188-189.
- Omar, B., Marwi, M. A., Oothuman, P. & Othman, H. F. (1994b). Observations on the behaviour of immatures and adults of some Malaysian Sarcosaprophagous flies. *Tropical Biomedicine*, 11, 149-153.
- Omar, B., Marwi, M. A., Suleiman, S. & Oothuman, P. (1994a). Dipteran succession in monkey carrion at a rubber tree plantation in Malaysia. *Tropical Biomedicine* 11, 77-82.
- Omar, B. (2002). Key to third instar larvae of flies of forensic importance in Malaysia. In: B., Greenberg and J.C., Kunich, editors. Entomology and the law- flies as forensic indicators. *Cambridge: Cambridge University Press*. pp 120-127.
- Ouda, M. S. (2014). Biological control by microorganisms and ionizing radiation. *International Journal of Advanced Research*, 2(5), 314-356.
- Rajagopal, K., Dhang, C. C., Lim, L. H., Ahmad, N. W., Ibrahim, S., & Aris, E. M. (2008). Estimated post-mortem intervals (PMI) of pathologist and entomologist in Malaysia: a comparison. *Proc ASEAN Congr Trop Med Parasitol*, 3, 21-7.

- Rivers, D. B. & Dahlem, G. A. (2014). *The Science of Forensic Entomology*. John Wiley and Sons Ltd. West Sussex, United Kingdom.
- Sakawi, Z., & Ismail, L. (2015). Managing odour pollution from livestock sources in Malaysia: Issues and challenges. *Geografia- Malaysian Journal of Society and Space*, 11(13), 96-103.
- Schroeder, H., Klotzbach, H., & Pu, K. (2003). Insects' Colonization of Human Corpses in Warm and Cold Season. *Legal Medicine*. 5, 372–374.
- Silahuddin, S. A., Latif, B., Kurahashi, H., Walter, D. E., & Heo, C. C. (2015). The importance of habitat in the ecology of decomposition on rabbit carcasses in Malaysia: implications in forensic entomology. *Journal of Medical Entomology*. 52(1), 9-23.
- Smith, K. G. V. (1986). *A Manual of Forensic Entomology*. New York: British Museum (Natural History) and Cornell University Press.
- Thompson, C. R., Brogan, R. S., Scheifele, L. Z., & Rivers, D. B. (2013). Bacterial interactions with necrophagous flies. *Annals of the Entomological Society of America*, 106(6), 799-809.
- Tomberlin, J. K., Mohr, R., Benbow, M. E., Tarone, A. M. & VanLaerhoven, S. (2011). A roadmap for bridging basic and applied research in forensic entomology. *Annual Review of Entomology*. 56, 401-421.
- Tomberlin, J. K., Crippen, T. L., Tarone, A. M., Singh, B., Adams, K., Rezenom, Y. H., Benbow, M. E., Flores, M., Longnecker, M., Pechal, J. L., Russell, D. H., Beier, R. C. & Wood, T. K. (2012). Interkingdom responses of flies to bacteria mediated by fly physiology and bacterial quorum sensing. *Animal Behavior*. 84, 1449-1456.
- Tomberlin, J. K., Crippen, T. L., Tarone, A. M., Chaudhury, M. F., Singh, B., Cammack, J. A., & Meisel, R. P. (2017). A review of bacterial interactions with blow flies (Diptera: Calliphoridae) of medical, veterinary, and forensic importance. *Annals of the Entomological Society of America*, 110(1), 19-36.
- Wells, J. D., & Stevens, J. R. (2008). Application of DNA-based methods in Forensic Entomology. *Annual Review Entomology*. 53, 103-120.
- Wells, J. D. & LaMotte, L. R. (2010) Estimating the Postmortem Interval. In: Byrd JH, Castner JL, editors. *Forensic Entomology: the utility of arthropods in legal investigation*. 2nd Edition. Boca Raton, FL: CRC Press, 201-50.

- Yu, G., Cheng, P., Chen, Y., Li, Y., Yang, Z., Chen, Y., & Tomberlin, J. K. (2011). Inoculating poultry manure with companion bacteria influences growth and development of black soldier fly (Diptera: Stratiomyidae) larvae. *Environmental Entomology*, 40(1), 30-35.
- Zheng, L., Crippen, T. L., Holmes, L., Singh, B., Pimsler, M. L., Benbow, M. E., ... & Wood, T. K. (2013). Bacteria mediate oviposition by the black soldier fly, *Hermetia illucens* (L.) (Diptera: Stratiomyidae). *Scientific reports*, 3, 2563.
- Zhu, J. (2000). A review of microbiology in swine manure odor control. *Agriculture, Ecosystems & Environment*, 78(2), 93-106.
- Zhu, G. H., Ye, G. Y., Hu, C., Xu, X. H., & Li, K. (2006). Developmental changes of cuticular hydrocarbons in *Chrysomya rufifacies* larvae; potential for determining larval age. *Medical and Veterinary Entomology*, 20, 438-444.
- Zhu, G. H., Yu, X. J., Xie, L. X., Luo, H., Wang, D., Lv, J. Y., & Xu, X. H. (2013). Time of death revealed by hydrocarbons of empty puparia of *Chrysomya megacephala* (Fabricius) (Diptera: Calliphoridae): A field experiment. *PLoS ONE*, 8(9), 1-7.