

BLOOD COMPATIBILITY ASSESSMENT OF METALLOCENE
POLYETHYLENE FOLLOWING NITRIC ACID TREATMENT

MUTHU VIGNESH VELLAYAPPAN

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Master of Philosophy (Biomedical Engineering)

Faculty of Biosciences and Medical Engineering

Universiti Teknologi Malaysia

JULY 2016

I dedicate this thesis to my beloved family:

My dearest parents, Mr. Vellayappan, Mrs. Meenakshi & fiancée Dr.Kamala.

ACKNOWLEDGEMENT

First of all, I wish to give my highest praise to God for giving me blessings and strength to complete this research. My deepest gratitude to my research supervisor, Dr. Saravana Kumar Jaganathan, Universiti Teknologi Malaysia (Faculty of Biosciences and Medical Engineering, UTM) for his continuous encouragement, guidance and support in the tenure of my study. I would like to thank my co-supervisors Prof. Ida Idayu Muhammad (Faculty of Chemical Engineering, UTM) and Dr. Ahmad Zahran Md. Khudzari (Faculty of Biosciences and Medical Engineering, UTM) for their guidance.

I would like to acknowledge Mr. Yong Lee Ming and Miss. Farah Nadiya binti Muhamad Sobri, MSI Technologies, Malaysia for their support to use Hirox 3D digital microscope KH-8700, and the Lab technicians of UTM for their assistance.

I am very much thankful to my lab-mates: Arunpandian, Aruna Priyadharshni, and Agnes Aruna John for their continued support. I am grateful to all faculties and non-teaching staffs of UTM.

This thesis would have been impossible without the unconditional love and support from my parents, Mr. M. Vellayappan and Mrs. V. Meenakshi. I would like to thank my brother Mr. V. Periakarruppan, sister Mrs. V. Umayal and my fiancée Dr. Kamala Swarnamani for being a constant source of motivation and encouragement.

Ultimately, my sincere appreciation goes to dean of Faculty of Biosciences & Medical Engineering, Prof. Dr. Jasmy bin Yunus and the IJN-UTM cardiovascular engineering center director Prof. Dr. -Ing. Eko Supriyanto for their constant support in helping me to complete my research.

ABSTRACT

Metallocene polyethylene (mPE) has been known for its excellent physical and mechanical properties, but its poor hemocompatibility limits its clinical application. Objective of this study was to analyze the physicochemical properties and blood compatibility of mPE following nitric acid (HNO₃) treatment. Characterization tests were performed using 3D Hirox, SEM, AFM, contact angle and FTIR. Blood compatibility of the sample was studied by conducting blood coagulation assays; hemolysis assay, PT, APTT, platelet adhesion and protein adsorption test. Result shows that the contact angle of the mPE treated with HNO₃ decreased from 86° to 69.7°. Surface of the mPE and the HNO₃ treated mPE investigated with FTIR revealed no major changes in its functional groups. 3D Hirox digital microscopy, SEM and AFM images show increased porosity and surface roughness. The protein adsorption studies show that the adsorbed albumin increased and adsorbed fibrinogen decreased in 60 minutes HNO₃ treated sample. Blood coagulation assays prothrombin time (PT) and activated partial thromboplastin time (APTT) were delayed significantly ($P < 0.05$) for the 60 minutes HNO₃ treated sample. Hemolysis assay and platelet adhesion of the treated surface resulted in reduced lysis of red blood cells and platelet adherence indicating improved hemocompatibility of HNO₃ treated mPE. To determine that HNO₃ does not deteriorate elastic modulus of mPE, the elastic modulus, tensile strength and tensile strength at break of mPE and HNO₃ treated mPE was compared and the result shows that HNO₃ treatment does not deteriorate the mechanical properties of mPE. To conclude, the overall observation suggests that the novel HNO₃ treated mPE may hold great potential to be exploited for various temporary blood contacting devices like catheters, endoscopy tip and etc.

ABSTRAK

Metallocene polietilena (mPE) lebih dikenali dengan sifat-sifat fizikal dan mekanikal yang mengagumkan, namun masalah keserasian dengan darah menghadkan aplikasi klinikal bahan ini. Objektif kajian ini adalah untuk menganalisis sifat-sifat fizikokimia dan keserasian darah mPE selepas rawatan asid nitrik (HNO_3). Ujian pencirian telah dilakukan dengan menggunakan 3D Hirox, SEM, AFM, sudut kenalan dan FTIR. Keserasian darah sampel telah dikaji dengan cerakin pembekuan darah seperti cerakin hemolisis, PT, APTT, bilangan platelet melekat dan ujian penyerapan protein. Sudut sentuhan bagi mPE yang ditindakbalaskan dengan HNO_3 telah menurun daripada 86° kepada 69.7° . Analisis FTIR menunjukkan tiada sebarang perbezaan yang ketara dari segi kumpulan berfungsi antara mPE dan mPE yang ditidakbalaskan dengan HNO_3 . Imej pada mikroskop digital 3D Hirox, SEM dan AFM telah menunjukkan peningkatan dari segi saiz liang dan kekasaran permukaan. Kajian penyerapan protein menunjukkan bahawa serapan albumin meningkat manakala penyerapan fibrinogen menurun bagi mPE yang ditindakbalaskan dengan HNO_3 selama 60 minit. Masa bagi cerakin pembekuan darah bagi prothrombin (PT) dan separa tromboplastin yang diaktifkan (APTT) mempunyai perbezaan yang ketara ($P < 0.05$) untuk mPE yang ditindakbalaskan dengan HNO_3 . Cerakin hemolisis dan lekatan platelet untuk permukaan yang ditindakbalas mengurangkan lisis sel-sel darah merah dan pematuhan platelet menunjukkan sifat keserasian dengan darah yang baik untuk mPE yang ditindakbalaskan dengan HNO_3 . Tiada perubahan dari segi sifat-sifat mekanik; elastik modulus dan kekuatan tegangan tercatat antara sampel mPE sebelum atau selepas ditidakbalaskan dengan HNO_3 . Oleh itu, analisis keseluruhan menunjukkan bahawa mPE yang ditindakbalaskan dengan HNO_3 mempunyai kebarangkalian yang positif untuk diaplikasikan sebagai peranti sementara yang bersentuhan dengan darah seperti kateter, tip endoskopi dan lain-lain lagi.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iv
	ACKNOWLEDGEMENTS	v
	ABSTRACT	vi
	ABSTRAK	vii
	TABLE OF CONTENTS	viii
	LIST OF TABLES	xi
	LIST OF FIGURES	xii
	LIST OF ABBREVIATIONS	xiv
	LIST OF SYMBOLS	xvi
	LIST OF APPENDICES	xvii
1	INTRODUCTION	
	1.1 Background	1
	1.2 Statement of Problem	5
	1.3 Objectives	5
	1.4 Scope of Study	6
	1.5 Significance of Study	6
	1.6 Thesis Outline	7
2	LITERATURE REVIEW	
	2.1 Biomaterials	8
	2.1.1 Blood Contact Activation	15
	2.1.2 Platelet Activation	16
	2.1.3 Protein Adsorption on Biomaterials	18
	2.2 Surface Treatment of Biomaterials	20

2.2.1	Types of Surface Modification Techniques	24
2.2.2	Chemical Surface Modification Technique	26
2.3	Nitric Acid Treatment	28
2.4	Summary	31
3	METHODOLOGY	
3.1	Chemicals and Instruments	33
3.2	Blood Procurement and Ethical Approval	34
3.3	Sample Preparation and Optimization of HNO ₃ Treatment Duration	34
3.4	Physical Characterization	37
3.4.1	Contact Angle Measurement	37
3.4.2	Attenuated Total Reflectance Fourier Transform Infrared Spectroscopy (ATR-FTIR)	37
3.4.3	3D-Hirox Digital Microscope	38
3.4.4	Scanning Electron Microscope (SEM)	40
3.4.5	Atomic Force Microscopy (AFM)	40
3.4.6	Tensile Testing	42
3.5	Blood Coagulation Assays	42
3.5.1	Protein Adsorption Test	42
3.5.2	Prothrombin Time (PT)	43
3.5.3	Activated Partial Thromboplastin Time (APTT)	43
3.5.4	Hemolysis Assay	44
3.5.5	Platelet Adhesion Assay	45
3.6	Statistical Analyses	45
4	RESULTS AND DISCUSSION	
4.1	Optimization of HNO ₃ Treatment Duration	46
4.2	Characterization of the Samples	48
4.2.1	Contact Angle Measurement	48

4.2.2	Attenuated Total Reflectance Fourier Transfer Infrared Spectroscopy (ATR-FTIR)	49
4.2.3	3D-Hirox Digital Microscopy	50
4.2.4	Scanning Electron Microscopy	55
4.2.5	Atomic Force Microscopy	56
4.2.6	Tensile Testing	58
4.3	Blood Coagulation Assays	60
4.3.1	Protein Adsorption Test	61
4.3.2	Prothrombin Time (PT)	64
4.3.3	Activated Partial Thromboplastin Time (APTT)	65
4.3.4	Hemolysis Assay	67
4.3.5	Platelet Adhesion Assay	69
4.3.6	Mechanism of Improved Blood Compatibility	71
5	CONCLUSION AND RECOMMENDATIONS	
5.1	Conclusion	73
5.2	Recommendations	74
	REFERENCES	76
	APPENDICES	93

LIST OF TABLES

TABLE NO.	TITLE	PAGE
4.1	Percentage weight loss of the HNO ₃ treated samples for optimization of acid treatment duration	46
4.2	Contact angle of the HNO ₃ treated samples for optimization of acid treatment duration	47
4.3	Tensile testing result of mPE before and after 30 minutes and 60 minutes HCl and HNO ₃ treatment	60

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Biomaterials used for different heart valve components	10
2.2	Circumferential direction	11
2.3	Structure of metallocene compound	14
2.4	Structure of polyethylene	14
2.5	Activation of the blood coagulation system initiated by biomaterial-protein interaction	17
2.6	Activation of platelets by artificial surfaces	17
2.7	Physico-chemical modification strategies	25
3.1	Schematic representation of series of characterization and blood compatibility experiments done	36
3.2	3D Hirox Microscope	39
3.3	Scanning Electron Microscopy	40
3.4	Atomic Force Microscopy	41
4.1	A representative FTIR spectra of untreated, 30 minutes and 60 minutes HNO ₃ treated metallocene polyethylene	50
4.2	Different three-dimensional representations using 3D Hirox digital microscope	52

4.3	Different three-dimensional representations using 3D Hirox digital microscopy with profiling line	52
4.4	The representative height of the pores of different samples measured using 3D-profiling of 3D Hirox digital microscopy	53
4.5	The average pore density of untreated, 30 minutes and 60 minutes HNO ₃ acid treated mPE.	54
4.6	Representative SEM micrographs of untreated, 30 minutes and 60 minutes HNO ₃ treated metallocene polyethylene.	56
4.7	Representative AFM images of untreated metallocene polyethylene, 30 minutes and 60 minutes HNO ₃ treated metallocene polyethylene	58
4.8	Comparison of normalized albumin adsorption of control and HNO ₃ treated metallocene polyethylene	63
4.9	Comparison of normalized fibrinogen adsorption of control and HNO ₃ treated metallocene polyethylene	63
4.10	Comparison of prothrombin time (PT) of control and HNO ₃ -treated metallocene polyethylene	65
4.11	Comparison Activated Partial Thromboplastin Time graph of control and HNO ₃ treated metallocene polyethylene	66
4.12	Comparison of hemolysis of control and HNO ₃ treated metallocene polyethylene	68
4.13	Comparison of platelet adhered on untreated and HNO ₃ treated metallocene polyethylene	70
4.14	Photomicrograph of platelets adhered on untreated, 30 minutes and 60 minutes HNO ₃ treated mPE at 40x magnification	70
4.15	Mechanism of improved hemocompatibility by HNO ₃ treatment	72

LIST OF ABBREVIATIONS

AFM	-	Atomic force microscopy
APTT	-	Activated partial thromboplastin time
ATR-FTIR	-	Attenuated total reflectance Fourier transformed infrared spectroscopy
BHV	-	Bioprosthetic heart valves
BMS	-	Bare-metal stents
DES	-	Drug-eluting stents
Fg	-	Fibrinogen
HA	-	Haemolysis assay
HCL	-	Hydrochloric acid
HI	-	Hemolytic index
HNO ₃	-	Nitric acid
IH	-	Intimal hyperplasia
ISR	-	In-stent restenosis
MHV	-	Mechanical heart valves
mPE	-	Metallocene polyethylene
NaOH	-	Sodium hydroxide
PANCMMA	-	Poly(acrylonitrile-co-maleic acid)s
PCL	-	Poly-3-caprolactone

PES	-	Polyethersulfone
PRP	-	Platelet rich plasma
PT	-	Prothrombin time
PTFE	-	Polytetrafluoroethylene
PU	-	Polyurethane
PVA	-	Poly vinyl alcohol
PVC	-	Poly vinyl chloride
RBC	-	Red blood cells
SD	-	Standard deviation
SEM	-	Scanning electron microscopy
ST	-	Stent thrombosis
SWCNT	-	Single-walled carbon nanotubes
THF	-	Tetrahydrofuran
VAD	-	Ventricular assist devices
vWF	-	Von willebrand factor
BCA	-	Bicinchoninic acid assay

LIST OF SYMBOLS

cm^{-1}	-	Per centimetre
mm^{-1}	-	Per millimetre
cm^2	-	Square centimetre
g/mol	-	Grams per mole
Hz	-	Hertz
MPa	-	Megapascal
mL	-	Millilitre
M	-	Molar
nm	-	Nanometre
mm	-	Millimeter
Ra	-	Average roughness
W	-	Watt
N	-	Newton
w/v	-	Weight per volume
μL	-	Microlitre
μm	-	Micrometre
μ^2	-	Micrometre square
$^{\circ}\text{C}$	-	Degree celsius
%	-	Percentage
\pm	-	Plus-minus sign

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Biomechanical properties of native and tissue engineered heart valve constructs	93
B	Properties of conventional blood contacting materials	95
C	Important human blood serum proteins and their key biological functions	102
D	Surface water contact angle and the cell types supported	104
E	Intensity of the FTIR peaks	106
F	Publications	107

CHAPTER 1

INTRODUCTION

1.1 Background

Materials which have been playing a vital role in replacing and mirroring the functions of various organs in human system [1] are collectively known as biomaterials. Biomaterial is the combination of substances originating from natural, inorganic or organic materials. These materials are biocompatible exactly or partially when it comes in contact with the body during the healing time. They involve complete or part of a living organism or biomedical device which perform, augments or replaces any natural functions. Biomaterials are commonly used in various medical devices and systems like synthetic skin [2], drug delivery systems [3], tissue cultures [4], hybrid organs [5], synthetic blood vessels [6], artificial heart valves, cardiac pacemakers [7], screws, plates, wires and pins for bone treatments, total artificial joint implants, skull reconstruction [8], dental and maxillofacial applications [9].

Biomaterials broadly fall into the four main types namely metals, ceramics, polymers and biological substances [10]. Metals have unique atomic structure which confers them characteristic strength and properties which enable them specifically for load-bearing applications like orthopaedics. However, the corrosion associated with the use of metals limits their utility. Ceramics have evolved as better biomaterials because of their bio-inertness and compatibility. However, due to brittleness and low impact strength [11], ceramics are losing popularity. Polymers

have widespread applications in the field of biomaterials. Properties of polymers are dependent on the unit macromolecule present in the long chain of the polymer.

Among all four types, the polymers have widespread application in the field of biomaterials because of excellent physico-chemical and mechanical properties. Moreover, polymers can be feasibly molded into desired shapes with desired mechanical characteristics. The most important application of polymers is cardiovascular based implants [7] and blood contacting devices [1]. The use of polymers in medical application ranges from vascular grafts, stents, prosthetic heart valves, catheters, heart assist devices, hemodialyser.

Modern revolution in polymer technology like metallocene single-site catalyst introduced a new class of polyolefins with improved performance properties like enhanced toughness, sealability, clarity, and elasticity [12]. The metallocene consists of two cyclopentadienyl anions (Cp,) which are bound to a metal center (M) [12] which has an oxidation state II, thereby resulting in a general formula $M(C_5H_5)_2$. One among the polymers developed through metallocene technology is metallocene polyethylene (mPE). mPE typically finds applications in disposable bags, storage bottles, blood bags, and syringe tubes. Even though mPE has outstanding permeability to oxygen and acts as a barrier towards ammonia and water, mPE lacks blood compatibility [13] to be used for blood contacting biomedical implants.

Biocompatibility is a prime factor which determines the quality of a biomaterial and its application in various arenas. There are different definitions for biocompatibility. It may be defined as the ability of the material to perform at a specific body site with an appropriate host reaction. Biocompatibility may also be defined as the ability of a biomaterial to perform its desired function with respect to a medical therapy, without eliciting any undesirable local or systemic effects in the recipient or beneficiary of that therapy, but generating the most appropriate beneficial cellular or tissue response to that specific situation and optimizing the clinically relevant performance of that therapy [14].

Biocompatibility has been mentioned in many works with increasing interest in evaluating the characteristics of medical materials and devices and also the responses caused by its components. The ideal pattern for determining these properties has not yet been determined; but, various methods have been suggested for this purpose. Biocompatibility encompasses many aspects of the material, including its physical, mechanical and chemical properties, and potential cytotoxic, mutagenic and allergenic effects [15], so that no significant injuries or toxic effects on the biological function of cells and individuals arise. Until the biocompatibility of a material is proven, it must be subjected to various studies ranging from in vitro assays to clinical trials, involving distinct areas such as pharmaceuticals, biology, chemistry and toxicology to justify its use as a biomaterial.

The term biocompatibility has been defined by consensus, but not blood compatibility. The interactions between blood and a surface depend on the blood composition, the blood flow and the surface of the material defined by its physicochemical feature. The design of bloodcompatible materials is clearly a challenge to increase success in all medical devices that come in contact with blood and to answer unsolved problems in vascular reconstruction.

To explain blood compatibility from a different perspective, consider a material that is not blood compatible, i.e. a thrombogenic material. Such material would produce specific adverse reactions when placed in contact with blood: formation of clot or thrombus composed of various blood elements; shedding or nucleation of emboli (detached thrombus); the destruction of circulating blood components and activation of the complement system and other immunologic pathways [10]. Thus, we can define blood compatibility as the ability of the material to work in a particular place without eliciting any of the above mentioned blood related complications. Indeed, biocompatibility of blood contacting devices relates mainly to the thrombotic response induced by the materials.

Several distinct but interrelated thrombotic and antithrombotic systems exist to prevent the formation of intravascular clots expected in response to vascular

trauma. Haemostasis is the sum of these mechanisms and serves to limit blood loss following injury. Once regulation is initiated, these same mechanisms combine first to localize the clot at the site of injury, then to terminate coagulation and finally to remove the clot once it has served its purpose. These haemostatic mechanisms include platelet activation, coagulation, fibrinolysis and local vascular effects. Blood clotting, platelet adhesion and giant cell formations are major problems associated with blood clotting devices. These problems frequently arise in cardiovascular implants since the material is always in contact with blood and its components [7].

The hypothesis of this thesis was that the HNO_3 treatment on mPE may improve the blood compatibility of the mPE polymer, to be utilized for different temporary blood contacting devices application like endscope tip, catheters and etc. The rationale for this hypothesis was that, when the mPE surface is subjected to HNO_3 treatment it was expected to etch the mPE surface which may improve the wettability or hydrophilicity of mPE. It may be expected that improved hydrophilicity may alter the protein adsorption and blood coagulation time resulting in enhanced blood compatibility. Besides that, it is also hypothesized that the HNO_3 treatment on mPE will not affect the mechanical strength of mPE.

1.2 Statement of Problem

Although mPE has excellent physico-chemical and mechanical properties, it fails as a promising biomaterial because of its poor blood compatibility. Biocompatibility is a vital factor which determines the quality of a biomaterial and its application in various arenas. It may be defined as the ability of the material to perform at a specific region with the appropriate host reaction. The events occur when the blood comes in contact with the implant is collectively called as blood mediated reactions or blood compatibility. Whenever the blood comes in contact with the implants (biomaterial) it will lead to following complications:

1. Blood components interaction with surfaces resulting in protein and water adsorption.
2. Blood cells interfere with the surface of biomaterial.
3. These actions lead to the hemostasis and coagulation.

To solve these issues, different surface modification techniques have been studied yet most of them are complex and limited to certain family of polymers. In recent times, millions of dollars was invested in advanced biomaterial research which includes discovery of new alternatives. However, in order to cater the high demand, more research needs to be encouraged to enhance the properties of the existing medical materials using feasible, eco-friendly and affordable modification technique. Hence, nitric acid surface modification technique on mPE was performed for improving its blood compatibility and it is the research gaph which will be addressed in this research.

1.3 Objectives

This research was carried out to determine the potential of nitric acid (HNO_3) treated metallocene polyethylene (mPE) as a biomaterial, for blood contacting device application. The following are the objectives of this study:

1. To investigate the physico-chemical changes induced on the surface of the mPE after HNO_3 treatment.
2. To determine the changes in the blood compatibility of the mPE subjected to acid treatment.

1.4 Scope of Study

This study consists of two parts. The first part of the study is focused mainly on the sample preparation, optimization of the nitric acid treatment on the mPE, and the characterization of mPE and HNO₃ treated mPE surfaces. Various methods were utilized; contact angle, attenuated total reflectance Fourier Transform Infrared Spectroscopy (ATR-FTIR), 3D Hirox digital microscopy, scanning electron microscopy (SEM), and atomic force microscopy (AFM) for the determination of the surface characteristic of the mPE and HNO₃ treated sample. To examine whether the HNO₃ treatment affect the tensile strength, elastic modulus and break strength of mPE, mechanical testing of mPE and HNO₃ treated mPE was compared.

In the second part, blood coagulation assays were carried out. The reason for this step is to ascertain the thromboresistance property of mPE and HNO₃ treated mPE surfaces when they are utilized as biomaterials, particularly for blood contacting devices. The blood coagulation assays like prothrombin test (PT), activated partial thromboplastin time (APTT), hemolysis assay (HA), and platelet adherence test is performed to investigate the blood compatibility of the mPE and HNO₃ treated mPE. Protein adsorption assay was carried out for determining the specific proteins albumin and fibrinogen adsorption on mPE and HNO₃ treated mPE. The mPE and the HNO₃ treated mPE blood compatibility results were compared with the conventional blood contacting materials for positioning this research with related studies in this field.

1.5 Significance of Study

The result of this study will provide an account on the improved blood compatibility of mPE by nitric acid treatment. In addition to that, the effect of the HNO₃ on the mPE polymer which was performed for the first time may kindle the enthusiasm of the other researchers to further explore the alternative acids available, to enhance the blood compatibility of polymers. Besides that, as nitric acid is low in

cost, a cost effective method for the blood compatibility enhancement of polymers can be introduced.

1.6 Thesis outline

This thesis is divided into five chapters. In Chapter 1, a brief explanation about the biomaterials and the research background of this study is given. Further, the objectives of this study have been presented in the context of solving the clinical complications mentioned. Finally, the importance of the proposed method and its influence in encouraging future researches is also projected.

In Chapter 2, a brief explanation about polymers and the problems observed during its contact with biological environment is summarized. In addition, the importance of surface modification techniques in solving those issues and some of the previous researches reported in that viewpoint is also discussed.

In Chapter 3, the research methodology and characterization studies followed in this research are given in detail. The discussions mainly cover the particulars, procedures and the need for characterization studies.

In Chapter 4, the results obtained from characterization and blood compatibility studies have been discussed and compared with the previous study's results. This section is the heart of the thesis, since it evidently reflects the achievement and the effectiveness of proposed idea.

In Chapter 5, a short summary of the whole work and its effectiveness in approaching blood compatibility problems are projected. Moreover, some suggestion about future research in the proposed perspective is also presented.

REFERENCES

1. John, A., Subramanian, A., Vellayappan, M., Balaji, A., Jaganathan, S., Mohandas, H., Paramalinggam, T., Supriyanto, E. and Yusof, M. Review: physico-chemical modification as a versatile strategy for the biocompatibility enhancement of biomaterials. *RSC Advances*. 2015. 5(49): 39232-39244.
2. MacNeil, S. Biomaterials for tissue engineering of skin. *Materials Today*. 2008. 11(5): 26-35.
3. Buckles, R.G. Biomaterials for drug delivery systems. *J Biomed Mater Res*. 1983. 17(1): 109-28.
4. Stark, Y., Bruns, S., Stahl, F., Kasper, C., Wesemann, M., Grothe, C. and Scheper T. A study on polysialic acid as a biomaterial for cell culture applications. *J Biomed Mater Res A*. 2008. 85(1): 1-13.
5. Woerly, S., Plant, G. W. and Harvey, A.R. Neural tissue engineering: from polymer to biohybrid organs. *Biomaterials*. 1996. 17(3): 301-10.
6. Ravi, S. and Chaikof, E.L. Biomaterials for vascular tissue engineering. *Regenerative medicine*. 2010. 5(1): 107.
7. Jaganathan, S.K., Eko, S., Selvakumar, M., Arunpandian, B. and Manjesh, A. Biomaterials in Cardiovascular Research: Applications and Clinical Implications. *BioMed Research International*. 2014. 2014:11.
8. Stevens, M.M., Biomaterials for bone tissue engineering. *Materials Today*. 2008. 11(5): 18-25.
9. Galler, K.M., D'Souza, R.N. and Hartgerink, J.D. Biomaterials and their potential applications for dental tissue engineering. *Journal of Materials Chemistry*. 2010. 20(40): 8730-8746.

10. Jaganathan, S.K., Balaji, A., Vellayappan, M.V., Subramanian, A.P., John A.A., Manjesh, A. and Eko, S. Review: Radiation-induced surface modification of polymers for biomaterial application. *Journal of Materials Science*. 2014. 1-12.
11. Niinomi, M. Recent metallic materials for biomedical applications. *Metallurgical and Materials Transactions A*. 2002. 33(3): 477-486.
12. Kealy, T.J. and Pauson P. L. A. New Type of Organo-Iron Compound. *Nature*. 1951. 168(4285): 1039-1040.
13. Mohandas, H., Sivakumar, G., Kasi, P., Jaganathan, S.K. and Eko, S. Microwave-Assisted Surface Modification of Metallocene Polyethylene for Improving Blood Compatibility. *BioMed Research International*. 2013. 2013: 7.
14. Williams, D.F. On the mechanisms of biocompatibility. *Biomaterials*. 2008. 29(20): 2941-53.
15. Lemons, J. and Natiella, J. Biomaterials, biocompatibility, and peri-implant considerations. *Dent Clin North Am*. 1986. 30(1): 3-23.
16. Schieker, M., Hermann, S., Inga, D., Sebastian, S. and Wolf. M. Biomaterials as Scaffold for Bone Tissue Engineering. *European Journal of Trauma*. 2006. 32(2): 114-124.
17. Hutmacher, D.W., Sittinger, M. and Risbud, M.V. Scaffold-based tissue engineering: rationale for computer-aided design and solid free-form fabrication systems. *Trends Biotechnol*. 2004. 22(7): 354-62.
18. Gomes, M.E., Godinho, J.S., Tchalamov, D., Cunha, A.M. and Reis R.L. Alternative tissue engineering scaffolds based on starch: processing methodologies, morphology, degradation and mechanical properties. *Materials Science and Engineering: C*. 2002. 20(1-2): 19-26.
19. Anselme, K. Osteoblast adhesion on biomaterials. *Biomaterials*. 2000. 21(7): 667-81.
20. Vo-Dinh, T. and Cullum, B. Biosensors and biochips: advances in biological and medical diagnostics. *Fresenius J Anal Chem*. 2000. 366(6-7): 540-51.
21. Rossen, L., Pernille, N., Kim, H. and Ole, F. R. Inhibition of PCR by components of food samples, microbial diagnostic assays and DNA-extraction solutions. *International Journal of Food Microbiology*. 1992. 17(1): 37-45.

22. Ryou, M. and Thompson, C.C. Tissue Adhesives: A Review. *Techniques in Gastrointestinal Endoscopy*. 2006. 8(1): 33-37.
23. Vellayappan, M.V., Balaji, A., Subramanian, A.P., John A.A., Jaganathan, S.K., Murugesan, S., Supriyanto, E. and Yusof, M. Multifaceted prospects of nanocomposites for cardiovascular grafts and stents. *Int J Nanomedicine*. 2015. 10: 2785-803.
24. Formanek, G., Frech, R, S. and Amplatz, K. Arterial thrombus formation during clinical percutaneous catheterization. *Circulation*. 1970. 41(5): 833-9.
25. Roată, C.E. Clinical practice guidelines for vascular access. *Am J Kidney Dis*. 2006. 48 (1): 176-247.
26. Vellayappan, M.V., Balaji, A., Subramanian, A.P., John A.A., Jaganathan, S.K., Murugesan, S., Hemanth, M., Supriyanto, E. and Yusof, M. Tangible nanocomposites with diverse properties for heart valve application. *Science and Technology of Advanced Materials*. 2015. 16(3): 033504.
27. Mohammadi, H. and Mequanint, K. Prosthetic aortic heart valves: Modeling and design. *Medical Engineering & Physics*. 2011. 33(2): 131-147.
28. Anwarul, H., Kim, R., Wojciech, S., Šeila, S., Arghya, P., Gulden, C., Mohammad R.K. and Ali .K .Biomechanical properties of native and tissue engineered heart valve constructs. *Journal of Biomechanics*. 2014. 47(9): 1949-1963.
29. Balguid, A., Rubbens M.P., Mol, A., Bank R.A., Bogers A.J., Kats J.P., Mol B.A., Baaijens, F.P. and Bouten, C.V. The role of collagen cross-links in biomechanical behavior of human aortic heart valve leaflets--relevance for tissue engineering. *Tissue Eng*. 2007. 13(7): 1501-11.
30. Mavrilas, D. and Missirlis, Y. An approach to the optimization of preparation of bioprosthetic heart valves. *Journal of Biomechanics*. 1991. 24(5): 331-339.
31. Conte, M.S. The ideal small arterial substitute: a search for the Holy Grail? *Faseb Journal*. 1998. 12(1): 43-5.
32. Kannan, R.Y., Salacinski, H.J., Butler, P.E., Hamilton, G. and Seifalian, A.M. Current status of prosthetic bypass grafts: a review. *J Biomed Mater Res B Appl Biomater*. 2005. 74(1): 570-81.

33. Desai, N.P. and Hubbell, J.A. Biological responses to polyethylene oxide modified polyethylene terephthalate surfaces. *J Biomed Mater Res.* 1991. 25(7): 829-43.
34. Schmedlen, R.H., Elbjeirami, W.M., Gobin, A.S., West, J.L. Tissue engineered small-diameter vascular grafts. *Clin Plast Surg.* 2003. 30(4): 507-17.
35. Balasubramanian, V., Grusin, N.K., Bucher, R.W., Turitto, V.T. and Slack, S.M. Residence-time dependent changes in fibrinogen adsorbed to polymeric biomaterials. *J Biomed Mater Res.* 1999. 44(3): 253-60.
36. Ballyk P.D., Walsh, C., Butany, J. and Ojha, M. Compliance mismatch may promote graft-artery intimal hyperplasia by altering suture-line stresses. *J Biomech.* 1998. 31(3): 229-37.
37. Fager, G. Thrombin and proliferation of vascular smooth muscle cells. *Circ Res.* 1995. 77(4): 645-50.
38. Roqué, M., Reis, E.D., Fuster, V., Padurean, A., Fallon, J.T, Taubman, M.B., Chesebro, J.H. and Badimon, J.J. Inhibition of tissue factor reduces thrombus formation and intimal hyperplasia after porcine coronary angioplasty. *J Am Coll Cardiol.* 2000. 36(7): 2303-10.
39. Hansson, G.K. and Libby.P. The immune response in atherosclerosis: a double-edged sword. *Nat Rev Immunol.* 2006. 6(7): 508-19.
40. Stone, G.W., Ellis, S.G., Cannon, L., Mann, J.T., Greenberg, J.D., Spriggs, D., DeMaio, S., Hall, P., Popma, J.J., Koglin, J. and Russell, M.E. Comparison of a polymer-based paclitaxel-eluting stent with a bare metal stent in patients with complex coronary artery disease: a randomized controlled trial. *Jama.* 2005. 294(10): 1215-23.
41. Daemen, J., Wenaweser, P., Tsuchida, K., Abrecht, L., Vaina, S., Morger, C., Kukreja, N., Jüni, P., Sianos, G., Hellige, G., vanDomburg, R.T., Hess, O.M., Boersma, E., Meier, B., Windecker, S. and Serruys, P.W. Early and late coronary stent thrombosis of sirolimus-eluting and paclitaxel-eluting stents in routine clinical practice: data from a large two-institutional cohort study. *Lancet.* 2007. 369(9562): 667-78.

42. Niccoli, G., Montone, R.A., Ferrante, G. and Crea, F. The evolving role of inflammatory biomarkers in risk assessment after stent implantation. *J Am Coll Cardiol.* 2010. 56(22): 1783-93.
43. Byrne, R.A., Joner, M., Tada, T. and Kastrati, A. Restenosis in bare metal and drug-eluting stents: distinct mechanistic insights from histopathology and optical intravascular imaging. *Minerva Cardioangiol.* 2012. 60(5): 473-89.
44. Bonaventura, K., Leber, A.W., Sohns, C., Roser, M., Boldt, L.H., Kleber, F.X., Haverkamp, W. and Dorenkamp, M. Cost-effectiveness of paclitaxel-coated balloon angioplasty and paclitaxel-eluting stent implantation for treatment of coronary in-stent restenosis in patients with stable coronary artery disease. *Clin Res Cardiol.* 2012. 101(7): 573-84.
45. Liistro, F., Angioli, P., Porto, I., Ricci, L., Ducci, K., Grotti, S., Falsini, G., Ventruzzo, G., Turini, F., Bellandi, G. and Bolognese, L. Paclitaxel-eluting balloon vs. standard angioplasty to reduce recurrent restenosis in diabetic patients with in-stent restenosis of the superficial femoral and proximal popliteal arteries: the debate-isr study. *J Endovasc Ther.* 2014. 21(1): 1-8.
46. Thierry, B., Winnik, F.M., Merhi, Y., Silver, J. and Tabrizian, M. Bioactive coatings of endovascular stents based on polyelectrolyte multilayers. *Biomacromolecules.* 2003. 4(6): 1564-71.
47. Pokasermson, P. and Praserttham, P. Comparison of Activity of Ziegler-Natta Catalysts Prepared by Recrystallization and Chemical Reaction Methods towards Polymerization of Ethylene. 2009. 13(1): 1-8.
48. Global Polyolefin Industry Development [<http://blogs.eci99.com/global-polyolefin-industry-development/>]. Retrieved date: 4, April, 2016.
49. Ahmad Shamiri, H.C., Shah, J., Mohd, H., Walter, K., Purushothaman, V. and Wageeh, A. The Influence of Ziegler-Natta and Metallocene Catalysts on Polyolefin Structure, Properties, and Processing Ability. *Materials.* 2014. 2014(7): 5069-108.
50. Harrington, B.A.W. and Glenn, M. Comparison of Ziegler-Natta and Metallocene ethylene elastomer-products. *EBSCO Host.* 2004. 230(2): 20.

51. Kaewarsa, P. Polymerization of ethylene over the supported Ziegler-Natta and metallocene catalysts on magnesium hydroxide and magnesium hydroxychloride: PhD Thesis. Khonkaen University; 2005.
52. Nitta, K.H. and Tanaka, A. Dynamic mechanical properties of metallocene catalyzed linear polyethylenes. *Polymer*. 2001. 42(3): 1219-26.
53. Hay, M.R. Thermal and dynamic mechanical properties of metallocene polyethylene polymer. *Polymer*. 2001. 42(21): 8621-7.
54. Bubeck, R.A. Structure–property relationships in metallocene polyethylenes. *Materials Science and Engineering: R: Reports*. 2002. 39(1): 1-28.
55. Sunny, K.E. Studies on Metallocene Polyolefin and Polyvinyl Chloride for Blood and Blood Component Storage Applications. PhD Thesis. Cochin University of Science and Technology; 2006.
56. Vogler, E.A. and Siedlecki, C.A. Contact activation of blood-plasma coagulation. *Biomaterials*. 2009. 30(10): 1857-69.
57. Gorbet, M.B. and M.V. Sefton, Biomaterial-associated thrombosis: roles of coagulation factors, complement, platelets and leukocytes. *Biomaterials*. 2004. 25(26): 5681-703.
58. Furie, B. and Furie, B.C. Mechanisms of Thrombus Formation. *New England Journal of Medicine*. 2008. 359(9): 938-949.
59. Godo, M.N. and Sefton, M.V. Characterization of transient platelet contacts on a polyvinyl alcohol hydrogel by video microscopy. *Biomaterials*. 1999. 20(12): 1117-26.
60. Sheppard, J.I., McClung, W.G. and Feuerstein, I.A. Adherent platelet morphology on adsorbed fibrinogen: effects of protein incubation time and albumin addition. *J Biomed Mater Res*. 1994. 28(10): 1175-86.
61. Wachtfogel, Y.T., Hack, C.E., Nuijens, J.H., Kettner, C., Reilly, T.M., Knabb, R.M., Bischoff, R., Tschesche, H., Wenzel, H. and Kucich, U. Selective kallikrein inhibitors alter human neutrophil elastase release during extracorporeal circulation. *Am J Physiol*. 1995. 268(3 2): 1352-57.
62. Hakim, R.M. and Schafer, A.I. Hemodialysis-associated platelet activation and thrombocytopenia. *Am J Med*. 1985. 78(4): 575-80.

63. Dee, K.C., Puleo, D.A. and Bizios, R. *Protein-Surface Interactions. An Introduction To Tissue-Biomaterial Interactions*. John Wiley & Sons. 2003.
64. Anderson, J.M., Rodriguez, A. and Chang D.T. Foreign body reaction to biomaterials. *Seminars in immunology*. 2008. 20(2): 86-100.
65. Kim, M.S., Khang, G. and Lee, H.B. Gradient polymer surfaces for biomedical applications. *Progress in Polymer Science*. 2008. 33(1): 138-64.
66. Sun, S., Yue, Y., Huang, X. and Meng, D. Protein adsorption on blood-contact membranes. *Journal of Membrane Science*. 2003. 222(1-2): 3-18.
67. Kinnari, T.J., Salonen, E.M. and Jero, J. Durability of the binding inhibition of albumin coating on tympanostomy tubes. *International Journal of Pediatric Otorhinolaryngology*. 2003. 67(2): 157-64.
68. Jung, F., Braune, S. and Lendlein, A. Haemocompatibility testing of biomaterials using human platelets. *Clinical hemorheology and microcirculation*. 2013. 53(1-2): 97-115.
69. Kottke, K., Anderson, J.M., Umemura, Y. and Marchant R.E. Effect of albumin coating on the in vitro blood compatibility of Dacron® arterial prostheses. *Biomaterials*. 1989. 10(3): 147-55.
70. DeQueiroz, A.A., Barrak, E.R, Gil, H.A. and Higa, O.Z. Surface studies of albumin immobilized onto PE and PVC films. *Journal of biomaterials science Polymer edition*. 1997. 8(9): 667-81.
71. Baican, M., Pâslaru, E., Hitruc, E.G. and Vasile, C. Albumin immobilization on polyvinylidene fluoride surfaces. *Dig J Nanometer Bios*. 2011. 6(3): 1053-64.
72. Ratner, B.D.H., Allan, S., Frederick, J. and Lemons, J. *Biomaterials Science: An Introduction to Materials in Medicine*. 2 ed. San Diego / London: Elsevier Academic Press; 2004.
73. Turci, F., Ghibaudi, E., Colonna, M., Boscolo, B., Fenoglio, I. and Fubini, B. An Integrated Approach to the Study of the Interaction between Proteins and Nanoparticles. *Langmuir : the ACS journal of surfaces and colloids*. 2010. 26(11): 8336-46.

74. Amarnath, L.P., Srinivas, A. and Ramamurthi, A. In vitro hemocompatibility testing of UV-modified hyaluronan hydrogels. *Biomaterials*. 2006. 27(8): 1416-24.
75. VanKooten, T.G., Schakenraad, J.M., VanderMei, H.C. and Busscher, H.J. Influence of substratum wettability on the strength of adhesion of human fibroblasts. *Biomaterials*. 1992. 13(13): 897-904.
76. VanWachem, P.B., Beugeling, T., Feijen, J., Bantjes, A., Detmers, J.P. and Aken, W.G. Interaction of cultured human endothelial cells with polymeric surfaces of different wettabilities. *Biomaterials*. 1985. 6(6): 403-8.
77. Dalby M.J. Cellular response to low adhesion nanotopographies. *Int J Nanomed*. 2007. 2(3): 373-81.
78. Wachem, P.B., Hogt, A.H., Beugeling, T., Feijen, J., Bantjes, A. and Detmers, J.P. Adhesion of cultured human endothelial cells onto methacrylate polymers with varying surface wettability and charge. *Biomaterials*. 1987. 8(5): 323-8.
79. Nakaoka, R., Yamakoshi, Y., Isama, K. and Tsuchiya, T. Effects of surface chemistry prepared by self-assembled monolayers on osteoblast behavior. *Journal of Biomedical Materials Research Part A*. 2010. 94(2): 524-32.
80. Xu, L.C. and Siedlecki, C.A. Effects of surface wettability and contact time on protein adhesion to biomaterial surfaces. *Biomaterials*. 2007. 28(22): 3273-83.
81. Joseph, C., Dongwoo, K. and Thomas, J.W. Nanometer polymer surface features: the influence on surface energy, protein adsorption and endothelial cell adhesion. *Nanotechnology*. 2008. 19(50): 505103.
82. Deligianni, D.D., Katsala, N., Ladas, S., Sotiropoulou, D., Amedee, J. and Missirlis Y.F. Effect of surface roughness of the titanium alloy Ti-6Al-4V on human bone marrow cell response and on protein adsorption. *Biomaterials*. 2001. 22(11): 1241-51.
83. Ma, M., and Hill, R.M. Superhydrophobic surfaces. *Current Opinion in Colloid & Interface Science*. 2006. 11(4): 193-202.
84. Sigal, G.B., Mrksich, M. and Whitesides, G.M. Effect of Surface Wettability on the Adsorption of Proteins and Detergents. *Journal of the American Chemical Society*. 1998. 120(14): 3464-73.

85. Jaganathan, S.K., Mohandas, H., Sivakumar, G., Kasi, P., Sudheer, T., and Avineri S. Enhanced Blood Compatibility of Metallocene Polyethylene Subjected to Hydrochloric Acid Treatment for Cardiovascular Implants. *BioMed Research International*. 2014. 2014: 7.
86. Arima, Y. and Iwata, H. Effect of wettability and surface functional groups on protein adsorption and cell adhesion using well-defined mixed self-assembled monolayers. *Biomaterials*. 2007. 28(20): 3074-82.
87. Chang, H.I. and Wang, Y. *Cell Responses to Surface and Architecture of Tissue Engineering Scaffolds*. Intech. 2011.
88. Bahram, S.J., Julien, P., Rong, Q., Shuling, G., Eric, H., Gschweng, B., Stiles, K., Tzy, M., Owen, N., Witte, X., Bruce, D. and Hong, W. Hydrophobic surfaces for enhanced differentiation of embryonic stem cell-derived embryoid bodies. *Proceedings of the National Academy of Sciences of the United States of America*. 2008. 105(38): 14459-64.
89. Ranella, A., Barberoglou, M., Bakogianni, S., Fotakis, C. and Stratakis, E. Tuning cell adhesion by controlling the roughness and wettability of 3D micro/nano silicon structures. *Acta biomaterialia*. 2010. 6(7): 2711-20.
90. Ishizaki, T., Saito, N. and Takai, O. Correlation of Cell Adhesive Behaviors on Superhydrophobic, Superhydrophilic, and Micropatterned Superhydrophobic/Superhydrophilic Surfaces to Their Surface Chemistry. *Langmuir : the ACS journal of surfaces and colloids*. 2010. 26(11): 8147-54.
91. Neto, A.I., Custodio, C.A., Song, W. and Mano, J.F. High-throughput evaluation of interactions between biomaterials, proteins and cells using patterned superhydrophobic substrates. *Soft Matter*. 2011. 7(9): 4147-51.
92. Song, W., Veiga, D.D., Custódio, C.A. and Mano, J.F. Bioinspired Degradable Substrates with Extreme Wettability Properties. *Advanced Materials*. 2009. 21(18): 1830-34.
93. Zheng, J., Li, L., Tsao, H.K., Sheng, Y.J., Chen, S. and Jiang S. Strong Repulsive Forces between Protein and Oligo (Ethylene Glycol) Self-Assembled Monolayers: A Molecular Simulation Study. *Biophysical Journal*. 2005. 89(1): 158-66.

94. Arima, Y. and Iwata, H. Effects of surface functional groups on protein adsorption and subsequent cell adhesion using self-assembled monolayers. *Journal of Materials Chemistry*. 2007. 17(38): 4079-87.
95. Flemming, R.G., Murphy, C.J., Abrams, G.A., Goodman, S.L and Nealey, P.F. Effects of synthetic micro- and nano-structured surfaces on cell behavior. *Biomaterials*. 1999. 20(6): 573-88.
96. Curtis, A. and Wilkinson, C. Nantotechniques and approaches in biotechnology. *Trends in biotechnology*. 2001. 19(3): 97-101.
97. Chou, L., Firth, J.D., Uitto, V.J. and Brunette D.M. Substratum surface topography alters cell shape and regulates fibronectin mRNA level, mRNA stability, secretion and assembly in human fibroblasts. *Journal of cell science*. 1995. 108 (4): 1563-73.
98. Rosales , J.I., Rodríguez, M.A., Mazzaglia, G., Ramón, P.J., Díaz, L., García, M., Vallecillo, M., Ruizc, C. and Cabrerizo, M.A. Effect of roughness, wettability and morphology of engineered titanium surfaces on osteoblast-like cell adhesion. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2010. 365(1–3): 222-29.
99. Curtis, A. and Wilkinson, C. Topographical control of cells. *Biomaterials*. 1997. 18(24): 1573-83.
100. Clark, P., Connolly, P., Curtis, A.S., Dow, J.A. and Wilkinson, C.D. Topographical control of cell behaviour: II. Multiple grooved substrata. *Development*. 1990. 108(4): 635-44.
101. Tiaw K.S., Goh S.W., Hong, M., Wang, Z., Lan, B. and Teoh, S.H. Laser surface modification of poly(epsilon-caprolactone) (PCL) membrane for tissue engineering applications. *Biomaterials*. 2005. 26(7): 763-9.
102. Lampin, M., Warocquier, C., Legris, C., Degrange, M. and Sigot, M.F. Correlation between substratum roughness and wettability, cell adhesion, and cell migration. *Journal of biomedical materials research*. 1997. 36(1): 99-108.
103. Wan, Y., Wang, Y., Liu, Z., Qu, X., Han, B., Bei, J. and Wang, S. Adhesion and proliferation of OCT-1 osteoblast-like cells on micro- and nano-scale topography structured poly(L-lactide). *Biomaterials*. 2005. 26(21): 4453-59.

104. Puckett, S., Pareta, R. and Webster, T.J. Nano rough micron patterned titanium for directing osteoblast morphology and adhesion. *Int J Nanomed.* 2008. 3(2): 229-41.
105. Yim, E.K., Pang, S.W. and Leong, K.W. Synthetic nanostructures inducing differentiation of human mesenchymal stem cells into neuronal lineage. *Experimental cell research.* 2007. 313(9): 1820-9.
106. Dalby, M.J., Riehle, M.O., Yarwood, S.J., Wilkinson, C.D. and Curtis, A.S. Nucleus alignment and cell signaling in fibroblasts: response to a micro-grooved topography. *Experimental cell research.* 2003. 284(2): 274-82.
107. Mirzadeh, H., Dadsetan, M. and Sharifi, N. Platelet adhesion on laser-induced acrylic acid-grafted polyethylene terephthalate. *Journal of Applied Polymer Science.* 2002. 86(13): 3191-3196.
108. Mittal, K.L. *Polymer International.* Taylor and Francis Group, 2009.
109. Vladkova, T.G. Surface Engineered Polymeric Biomaterials with Improved Biocontact Properties. *International Journal of Polymer Science.* 2010. 1-22.
110. Frederick, M.F. *Contact Angle, Wettability, and Adhesion.* American Chemical Society. 1964.
111. Lim, J.Y. and Donahue, H.J. Cell sensing and response to micro- and nanostructured surfaces produced by chemical and topographic patterning. *Tissue Eng.* 2007. 13(8): 1879-91.
112. Miller, D.C., Thapa, A., Haberstroh, K.M. and Webster, T.J. Endothelial and vascular smooth muscle cell function on poly(lactic-co-glycolic acid) with nanostructured surface features. *Biomaterials.* 2004. 25(1): 53-61.
113. Balakrishnan, B., Kumar, D.S., Yoshida, Y., and Jayakrishnan, A. Chemical modification of poly(vinyl chloride) resin using poly(ethylene glycol) to improve blood compatibility. *Biomaterials.* 2005. 26(17): 3495-3502.
114. Aldenhoff, Y.B. and Koole, L.H.. Platelet adhesion studies on dipyridamole coated polyurethane surfaces. *Eur Cell Mater.* 2003. 5: 61-67.
115. Xiang, T., Wen, Y., Rui, W., Su, L., Shu-Dong, S. and Chang-Sheng, Z. Surface hydrophilic modification of polyethersulfone membranes by surface-initiated

- ATRP with enhanced blood compatibility. *Colloids Surf B Biointerfaces*. 2013. 110: 15-21.
116. Mona, J., Kuo, C.J., Perevedentseva, E., Priezzhev, A.V. and Cheng, C.L. Adsorption of human blood plasma on nanodiamond and its influence on activated partial thromboplastin time. *Diamond and Related Materials*. 2013. 39: 73-7.
117. Park, H.D., Lee, W.K., Ooya, T., Park, K.D., Kim, Y.H. and Yui, N. *J Biomed Mater Res*. 2003. 66(3): 596-604.
118. Xu, Z.K., Nie, F.Q., Qu, C., Wan, L.S., Wu, J. and Yao, K. Tethering poly(ethylene glycol)s to improve the surface biocompatibility of poly(acrylonitrile-co-maleic acid) asymmetric membranes. *Biomaterials*. 2005. 26(6): 589-598.
119. Akelah, A. *Polymeric Materials: Preparation and Properties, in Functionalized Polymeric Materials in Agriculture and the Food Industry*. Springer. 2013.
120. Nie, W.Z. and Li, J. Effects of plasma and nitric acid treatment of carbon fibers on the mechanical properties of thermoplastic polymer composites. *Mechanics of Composite Materials*. 2010. 46(3): 251-256.
121. Cagiao, M.E., Rueda, D.R. and Baltá-Calleja, F.J. Degradation of nitric acid-treated bulk polyethylene. *Polymer Bulletin*. 1980. 3(5): 305-310.
122. Cagiao, M.E., Rueda, D.R. and Baltá Calleja, F.J. Hardness of nitric acid treated polyethylene followed by recrystallization. *Colloid and Polymer Science*. 1987. 265(1): 37-41.
123. Hock, C.W. Selective oxidation with nitric acid reveals the microstructure of polypropylene. *Journal of Polymer Science Part B: Polymer Letters*. 1965. 3(7): 573-576.
124. Hay, I.L. and Keller, A. Morphology of Synthetic Fibres; A Study on Drawn Polyethylene. *Nature*. 1964. 204(4961): 862-864.
125. Maxim, N., Tchoul, W.T.F., Giulio, L., Daniel, E. and Sivaram, A. Effect of Mild Nitric Acid Oxidation on Dispersability, Size, and Structure of Single-Walled Carbon Nanotubes. *Chemistry of Materials*. 2007. 19(23): 5765-5772.

126. Cagliao, M.E., Rueda, D. R. and Baltá, F. J. Degradation of Nitric Acid-Treated Bulk Polyethylene. III. Melting Behavior. *Colloid and Polymer Science*. 1983. 261: 626-630.
127. Ferro-Garcia, M. A., Joly, J.P., Bautista-Toledo, I., Carrasco-Marin, F. and Rivera-Utrilla, J. Activated Carbon Surface Modifications by Nitric Acid, Hydrogen Peroxide, and Ammonium Peroxydisulfate Treatments. *Langmuir*. 1995. 11(11): 4386-4392.
128. ChenboDong, A.S.C., Reem, E., Gabriela, P., Yon, R. and Cerasela, Z. Effects of acid treatment on structure, properties and biocompatibility of carbon nanotubes. *Applied Surface Science*. 2013. 264: 261–268.
129. Hanford. W.E. *Oxidized polymers and process for their preparation*. US2360673A. 1944.
130. Fu, Y.C. and Gunnell, T.J. *Nitric acid treatment of carbon black*. US3336148A. 1967.
131. Gomathi, N., Mishra, D., Maiti, T.K. and Neogi, S. Helium Plasma Treatment to Improve Biocompatibility and Blood Compatibility of Polycarbonate. *Journal of Adhesion Science and Technology*. 2010. 24(13-14): 2237-55.
132. Qi, P., Maitz, M.F., and Huang, N. Surface modification of cardiovascular materials and implants. *Surface and Coatings Technology*. 2013. 233: 80-90.
133. Pereira, C. Nondestructive characterization and enzyme cleaning of painted surfaces: assessment from the macro to nano level. *Microsc Microanal*. 2013. 19(6): 1632-44.
134. Roozbahani, F., Sultana, N., Almasi, D. and Naghizadeh, F. Effects of Chitosan Concentration on the Protein Release Behaviour of Electrospun Poly(ϵ -caprolactone)/Chitosan Nanofibers. *Journal of Nanomaterials*. 2015. 2015: 11.
135. Wagner, A., Poursorkhabi, V., Mohanty, A.K. and Misra, M. Analysis of Porous Electrospun Fibers from Poly(l-lactic acid)/Poly(3-hydroxybutyrate-co-3-hydroxyvalerate) Blends. *ACS Sustainable Chemistry & Engineering*. 2014. 2(8): 1976-82.
136. Pelagade, S.M., Rane, R.S., Mukherjee, S., Deshpande, U.P., Ganesan, V. and Shripathi, T. Investigation of Surface Free Energy for PTFE Polymer by Bipolar

- Argon Plasma Treatment. *Journal of Surface Engineered Materials and Advanced Technology*. 2012. 2(2): 132-136.
137. Roy, R.K., Choi, H.W., Yi, J.W., Moon, M.W., Lee, K.R. and Han, D.K. Hemocompatibility of surface-modified silicon-incorporated diamond-like carbon films. *Acta biomaterialia*. 2009. 5(1): 249-56.
138. Kwok, S.C.H., Wang, J. and Chu, P.K. Surface energy, wettability, and blood compatibility phosphorus doped diamond-like carbon films. *Diamond and Related Materials*. 2005. 14(1): 78-85.
139. Gomathi, N., Rajasekar, R., Rajesh, R., Debasish, M. and Neogi, S. Development of bio/blood compatible polypropylene through low pressure nitrogen plasma surface modification. *Materials Science and Engineering: C*. 2012. 32(7): 1767-1778.
140. Chau, T.T., Bruckard, W.J., Koh, P.T.L. and Nguyen, A.V. A review of factors that affect contact angle and implications for flotation practice. *Advances in Colloid and Interface Science*. 2009. 150(2): 106-115.
141. Hardy Paul W. Nitric acid treatment of steel. US2915420A. 1959.
142. Mirzadeh, H. and Dadsetan, M. Influence of laser surface modifying of polyethylene terephthalate on fibroblast cell adhesion. *Radiation Physics and Chemistry*. 2003. 67(3-4): 381-5.
143. Makphon, P., Ratanatongchai, W., Chongkum, S., Tantayanon, S. and Supaphol, P. Polycarbonate microfilters by nuclear tracking and chemical etching (track-etching) technique: Preparation and characterization. *Journal of Applied Polymer Science*. 2006. 101(2): 982-90.
144. Gui-qiu, M., Ben, L., Chen, L., Dinghai, H. and Jing, S. Plasma modification of polypropylene surfaces and its alloying with styrene in situ. *Applied Surface Science*. 2012. 258(7): 2424-2432.
145. Cesca, F., Limongi, T., Accardo, A., Rocchi, A., Orlando, M., Shalabaev, V., Di Fabrizio, E. and Benfenati, F. Fabrication of biocompatible free-standing nanopatterned films for primary neuronal cultures. *RSC Advances*. 2014. 4(86): 45696-45702.

146. Tverdokhlebov, S.I., Bolbasov, E.N., Shesterikov, E.V., Antonova, L.V., Golovkin, A.S., Matveeva, V.G., Petlin, D.G. and Anissimov, Y.G. Modification of polylactic acid surface using RF plasma discharge with sputter deposition of a hydroxyapatite target for increased biocompatibility. *Applied Surface Science*. 2015. 329(0): 32-39.
147. Wanke, C.H. Tuning of polypropylene wettability by plasma and polyhedral oligomeric silsesquioxane modifications. *Polymer*. 2011. 52(8): 1797-1802.
148. Slepíčka, P. Surface characterization of plasma treated polymers for applications as biocompatible carriers. *Express Polymer Letters*. 2013. 7(6): 535-545.
149. Poletti, G., Orsini, F., Lenardi, C., and Barborini, E. A comparative study between AFM and SEM imaging on human scalp hair. *Journal of Microscopy*. 2003. 211: 249–255.
150. Josep, F.B., Stefan, G. G., Notni, J.S., Jean, M. B. and Angela, D. Surface characterization techniques for determining the root-mean-square roughness and power spectral densities of optical components. *Applied Optics*. 2002. 41 (1). 1-23.
151. Jolly, W.L. *Modern Inorganic Chemistry*: McGraw-Hill; 1984.
152. Bell, R.P. *The Proton in Chemistry*: Cornell University Press; 1973.
153. Schopka, S. Current Strategies in Cardiovascular Biomaterial Functionalization. *Materials*. 2010. 3(1): 638-655.
154. Ma, W.J., Ruys, A.J., Mason, R.S., Martin, P.J., Bendavid, A. and Liu, Z. DLC coatings: Effects of physical and chemical properties on biological response. *Biomaterials*. 2007. 28(9): 1620-8.
155. Bohnert, J.L. Changes in adsorbed fibrinogen and albumin interactions with polymers indicated by decreases in detergent elutability. *J Coll Interf Sci*. 1986. 111(2): 363-78.
156. Jeyachandran, Y.L., Mielczarski, E. Rai, B. and Mielczarski. J.A. Quantitative and Qualitative Evaluation of Adsorption/Desorption of Bovine Serum Albumin on Hydrophilic and Hydrophobic Surfaces. *Langmuir : the ACS journal of surfaces and colloids*. 2009. 25(19): 11614-20.

157. Seeger, J.M., Ingegno, M.D., Bigatan, E., Klingman, N. Amery, D. and Widenhouse, C. Hydrophilic surface modification of metallic endoluminal stents. *Journal of vascular surgery*. 1995. 22(3): 327-36.
158. Wu, Y., Simonovsky, F.I., Ratner, B.D. and Horbett, T.A. The role of adsorbed fibrinogen in platelet adhesion to polyurethane surfaces: a comparison of surface hydrophobicity. protein adsorption. monoclonal antibody binding. and platelet adhesion. *Journal of biomedical materials research Part A*. 2005. 74(4): 722-38.
159. Vroman, L. and Adams, A.L. Findings with the recording ellipsometer suggesting rapid exchange of specific plasma proteins at liquid/solid interfaces. *Surface Science*. 1969. 16: 438-46.
160. Hemmerlé, J., Altmann, S.M., Maaloum, M., Hörber, J.K.H., Heinrich, L. and Voegel, J.C. Direct observation of the anchoring process during the adsorption of fibrinogen on a solid surface by force-spectroscopy mode atomic force microscopy. *Proceedings of the National Academy of Sciences of the United States of America*. 1999. 96(12): 6705-10.
161. Lu, D.R. and Park, K. Effect of surface hydrophobicity on the conformational changes of adsorbed fibrinogen. *Journal of colloid and interface science*. 1991. 144(1): 271-81.
162. Iwamoto, G.K., Winterton, L.C., Stoker, R.S., VanWagenen, R.A., Andrade, J.D. and Mosher, D.F. Fibronectin adsorption detected by interfacial fluorescence. *Journal of colloid and interface science*. 1985. 106(2): 459-64.
163. Lin, W.C., Liu, T.Y. and Yang, M.C. Hemocompatibility of polyacrylonitrile dialysis membrane immobilized with chitosan and heparin conjugate. *Biomaterials*. 2004. 25(10): 1947-57.
164. Wen, C.L., Da-Guang, Y. and Ming-Chien, Y. Blood compatibility of thermoplastic polyurethane membrane immobilized with water-soluble chitosan/dextran sulfate. *Colloids and Surfaces B: Biointerfaces*. 2005. 44: 82–92.
165. Gomathi, N. and Sudarsan, N. Investigation on Argon–Oxygen Plasma Induced Blood Compatibility of Polycarbonate and Polypropylene. *Journal of Adhesion Science and Technology*. 2009. 23: 1811-1826.

166. Wang, J., Pan, C.J., Huang, N., Sun, H., Yang, P., Leng, Y.X., Chen, J.Y., Wan, G.J. and Chu, P.K. Surface characterization and blood compatibility of poly(ethylene terephthalate) modified by plasma surface grafting. *Surface & Coatings Technology*. 2005. 196: 307–311.
167. Zhengbao, Z., YanMa, X., YueMeng, L. and Zhifei, D. Self-assembled hemocompatible coating on poly (vinyl chloride) surface. *Applied Surface Science*. 2009. 256: 805–814.
168. Fazley, M. Hemocompatibility of surface modified silk fibroin materials: a review. *Reviews on Advanced Materials Science*. 2014. 38:148-159.
169. Bailly, A.L., Lautier, A., Guiffant, G., Dufaux, J., Houdart, E. and Labarre, D. M. Thrombosis of angiographic catheters in humans: experimental study. *The International journal of Artificial Organs*. 1999. 22: 690-700.
170. Wenzhong, S., Li, Z. and Liu, Y. Surface chemical functional groups modification of porous carbon. *Recent Patents on Chemical Engineering* 2008. 1(1): 27-40.
171. Habibzadeh, S., Li, L., Shum-Tim, D., and Davis, E.C. Omanovic. S. Electrochemical polishing as a 316L stainless steel surface treatment method: Towards the improvement of biocompatibility. *Corrosion Science*. 2014. 87(0): 89-100.
172. Lee, J.H. and Lee, H.B. Platelet adhesion onto wettability gradient surfaces in the absence and presence of plasma proteins. *Journal of biomedical materials research*. 1998. 41(2): 304-11.
173. Zhao, T., Li, Y., Gao, Y., Xiang, Y., Chen, H. and Zhang, T. Hemocompatibility investigation of the NiTi alloy implanted with tantalum. *Journal of materials science Materials in medicine*. 2011. 22(10): 2311-18.
174. Zingg, W., Neumann, A.W., Strong, A.B. Hum, O.S. and Absolom, D.R. Platelet adhesion to smooth and rough hydrophobic and hydrophilic surfaces under conditions of static exposure and laminar flow. *Biomaterials*. 1981. 2(3): 156-68.
175. Zingg, W., Neumann, A.W., Strong, A.B. and Hum, O.S. Absolom. D.R. Effect of surface roughness on platelet adhesion under static and under flow conditions. *Canadian journal of surgery*. 1982. 25(1): 16-9.