THE EFFECT OF LASER IRRADIATION ON THE VIABILITY OF HUMAN BREAST CANCER CELL, MDA-MB-231

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Abstract

This study compared the effects of different sources of laser phototherapy on the cell viability of the in vitro human breast cancer cell lines. Laser phototherapy is used in the breast cancer clinical treatment, despite the limited safety information of laser irradiation effect on the cancer cell behavior. This study contributed on the development of guidelines for safer laser usage in treating breast cancer and minimising the possibility of activating postmastectomy lymphedema. Cancer cell viability and morphology were studied in this research. Human breast cancer (MDA-MB-231) cell lines were cultured for 24 hours in CO\textsubscript{2} incubator and irradiated with different laser sources and number of pulse. The viability of cancer cells were assessed by MTT assay 24 hours after laser irradiation. The result showed that MDA-MB-231 cell viability increased after being irradiated by excimer 248 nm laser. However, the cancer cell viability slightly decreased after irradiation by both Nd:YAG 1064 and 532 nm. Although certain doses of laser may affect the MDA-MB-231 cell viability, additional laser exposures had nearly no effect. The research shows that Nd:YAG 1064 nm more effective in lowering cancer cell survivability than 532 nm and 248 nm. Further in vivo studies are needed for better understanding on the mechanism of laser-tissue interaction and improve the laser usage safety in photothermal therapy.

Keywords: Photothermal laser, MDA-MB-231, Excimer, postmastectomy lymphedema

Abstrak

1.0 INTRODUCTION

The clinical application of a thermal laser with low power outputs [1, 2] has been used now for many years in laser phototherapy to treat numbers of disease including epidermal abrasions, osteoarthritis and recently breast cancer-related lymphedema [3-5]. Both physical and psychological will get impacts of the untreated lymphoedema especially on an individual’s well-being. The signs and symptoms of lymphoedema are limb deformation and swelling, immobilization, restricted shoulder mobility, burning, elevated skin temperature, fibrosis, and social isolation [3, 6-9]. 30% of patients treated for breast cancer will have lymphoedema of the arm as it was one of the main side effects following breast cancer radiation therapy or surgery [6]. For the present time, there still no definite treatment for lymphoedema, although standard treatments for lymphedema include pressotherapy, exercise, manual lymphatic drainage, compression bandaging and skin care. However these treatments are generally applied in combination of each other known as Complete Decongestive Therapy (CDT) and it is very expensive, time consuming, require qualified medical professionals, less accepted by patients and limited effectiveness [10-12].

In previous studies [13-15], double blind and medicine-controlled trials have shown the effectiveness of Low Level Laser Therapy (LLLT) in improving the postmastectomy lymphedema symptoms. The treatments consisted of radiation with 904 nm wavelength, energy density of 1.5 J/cm², from 10 to 17 points distributed in the affected area and the number of sessions ranged from 8 to 36. A significant reduction in the size of the affected arm, volume of extracellular fluids and the tissue hardness, as well as improved the shoulder mobility and grip strength [16, 17].

Powell et al. [18] used four different cell line, human breast adenocarcinoma (MCF-7), human breast ductal, carcinoma with melanoma genotypic traits (MDA-MB-435S), immortalized human mammary epithelial (SVCT and Bre80hTERT), to study the effect of three different laser source, 780 nm continuos (50 mW), 830 nm continuos (30 mW) and 904 nm pulsed (90 mW) with range 1 to 3 laser exposure on cells proliferation. The result showed, although certain doses of laser will increase cell proliferation, however multiple doses exposures had either no effect or showed negative dose-response relationships. There was no sign of malignant modification and transformation of cells by laser phototherapy was observed under the applied conditions.

In other research [19], Schartinger et al. used human bronchial epithelial cells (BEAS-2B), human gingival fibroblasts and oral squamous cell carcinoma cells (SCC-25) to determine the cell proliferation after irradiate by GaAlAs, diode laser at 660 nm with power output 350 mW for 15 min and 3 repetition exposure in subsequent days. The result shows, there was increasing in cell proliferation for human gingival fibroblasts but decreasing for non-neoplastic epithelial cells and oral carcinoma cells SCC-25. However in SCC-25 cells, there is a proapoptotic effect of laser phototherapy was observed.

Gao et al. [20] studied the effect of two low power laser irradiation (LPLI) of He–Ne laser, 632.8 nm and 5 mw at fluence of 0.8 J/cm² and high fluence LPLI of He–Ne laser, 632.8 nm and 40 mw at fluence of 60 J/cm² on ASTC-a-1 cells. The study showed low fluence of LPLI will increase the cell proliferation and activated the protein kinase Cs (PKCs) in the cells. However, the study reported that high fluence of LPLI was decrease the cell viability and PKCs activity but increases the cell apoptosis.

These finding encourage more research on the application of laser phototherapy in cancer treatment such as chemotherapy and radiotherapy induced BCRL and oral mucositis. Few safety studies on the usage of laser in cancer treatment have been undertaken, although it has been by numbers of researches suggested that laser phototherapy has no effect on disease recurrence or survival in cancer patients [21,22] and it been suggested by Werneck et al. [23] "clinically, irradiating a cancer lesion may result in increased tumor progression". Although researches suggests that the growth of cancer cell in culture can be stimulated by laser phototherapy, and the clinical application laser in cancer treatment is increasing. The effect on cancer cells in vivo requires more critical review on the safety and efficiency of the laser application.

In seeking to identify the effective and safe dose parameters for the treatment of cancer, the aims of this research were to study the response of human breast cancer cell lines MDA-MB-231 in vitro with commonly used laser phototherapy at wavelengths of 248 nm, 532 nm, and 1064 nm and varied the number of doses exposure. This finding would lead to determination the relationship of wavelength-response, energy-response and dose-response after the human breast cancer irradiated with a range of laser parameters.
2.0 EXPERIMENTAL

2.1 Chemicals and Cell Line

MDA-MB-231, human breast cancer cells were a kind gift on Dr. Pauliena Bohari (Faculty of Biosciences and Medical Engineering, Universiti Teknologi Malaysia). Cells were grown in Dulbecco’s Modified Eagle’s Medium (DMEM), with 10% FBS, Fetal Bovine Serum and 1% Penicillin-Streptomycin Liquid and kept at 37°C in incubator with 5% CO2. The viability of the cells were measured with MTT assay, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide.

2.2 In Vitro Treatment Protocol

MDA-MB-231 cells were seeded in 24-well plate with 6×10⁴ seeding density and incubated for 24 hours. After 24 hours, the medium for all cells were changed with new medium to make sure all cell have enough food and not to die because of the insufficient supplement. After the medium were renewed, cells were treated by the lasers and incubated for another 24 hours before the cell viability determined by the MTT assay.

2.3 Laser treatment

The treatments have been performed with two different lasers. The Excimer laser, which available with average powers up to 35 W with KrF at 248 nm and Nd:YAG laser, which have power up to 500W at 1064 nm and 532 nm. For Excimer laser, KrF at 248 nm, a single pulse with constant frequency 20 Hz was used and the energy was varied in range 1 to 6 mJ delivered to the cells. The laser pulses were manually started using external trigger. A pulse of Nd:YAG laser at both 1064 and 532 nm were varied in energy from 200 to 1200 mJ. The three different wavelength lasers were set with constant energy but the number of pulses was increase from 1 to 6 pulses. 24 hours before treatment, cells were seeded in 24-wellplate with seeding density 6×10⁴. For laser irradiation, the 24-wellplates were placed on a stationary platform and the 4 treatments were carried out 24 hours away from each other and the experiment was repeated 3 times. The treated samples were compared with controls cancer cells maintained in the same conditions, without laser irradiation.

Figure 1 The viability (percentage of total cells) of human breast cancer cell, MDA-MB-231 after irradiated by (a)Excimer KrF 248nm (b)Nd:YAG 532nm (c)Nd:YAG 1064nm lasers with different energy

Figure 2 The viability (percentage of total cells) of human breast cancer cell, MDA-MB-231 after irradiated by (a)Excimer KrF 248nm (b)Nd:YAG 532nm (c)Nd:YAG 1064nm lasers with different number of pulse
of human corneal limbal epithelial HLE cells. In other research, Olivia Serdarevic et al. [25] reported at the 248-nm wavelength, the thermal effects of laser become more dominant therefore; all the fungal elements will be ablated. These ranges of laser wavelength are suitable for healing treatment acting as sterilization agent.

The result in Figure 1 (b) & (c) shows after irradiate by both Nd:YAG at 1064nm and 532nm the percentage of total cell are lesser compare to the control untreated breast cancer cells. However, when the cells were treated by Nd:YAG with same energy 200mJ but different wavelength, it shows that at wavelength 1064nm, there are more cell death which have only 74% of cell survived after the treatment compare to the cell treated by 532 nm which have 84%. As the laser energy of both 532 and 1064 nm Nd:YAG laser were increased, the percentage of cell survive were decreased.

Base on literature, there are records on therapeutic effects by low-level of laser in range of visible to near-infrared wavelengths, 500 – 1000 nm. The therapeutic effects caused by laser phototherapy, such as increasing cellular activity not only influenced by the laser wavelength, doses of exposure, power density , as well as the laser frequency of treatment and type of injury and tissue condition of the breast cancer patient [24].

The mechanism of laser-tissue interaction was explained by Welch et al. [27]. In laser photothermal reactions, the laser light will absorb by cell chromophores and will convert to heat. This causes a rise in temperature of the cells and the heat will diffuse through the cell causing a rise in temperature in the surrounding cell. If the heat is high enough, it will result in denaturization, and even vaporization. The damage done to the cell depends on the temperature that is reached.

3.2 The Effect of Number of Laser Pulse on Cell Viability

Although using different source of laser will increase and decrease the MDA-MB-231 cell viability, additional pulses of laser exposures had nearly no effect on the result as shown in Figure 2 (a) (b) and (c). When MDA-MB-231 cells were treated with more than one exposure of laser irradiation at three different wavelengths, there were no significant changes on viability of the cells. This result proved the study conducted by Powell et al. as they reported that no change in proliferation of the breast cancer cell MCF-7 when irradiated with more than one exposure doses of laser [28].

The morphology of MDA-MB-231 after treatment was rounded compare to controlled breast cancer cell, which the morphology is epithelial. The spherical structure of cell was indicating the death cell as reported by a few studies on the morphology of death cell for MDA-MB-231 [29-31]. Figure 2 (a) (b) & (c) show there are more spherical shape in morphology of cell treated by 1064 nm laser, followed by 532 nm. The cell death is due to heat absorbed by cell chromophores
from the laser light. If the electric field is very high, a small region of plasma will occur with high electric fields, associate with dielectric breakdown, the formation of shockwave, and tissue rupture [29].

4.0 CONCLUSION

Although these results does not show any significant laser-induced modification of cancer cell behaviour, further in vivo studies with high methodology quality are needed to better understand on the action mechanism of laser and improve the safely applying laser therapy to prevent and minimize the postmastectomy lymphedema.

Acknowledgement

We wish to thank to Gavornment of Malaysia through FRGS grant vote 4F543 and MyBrain 15 for the scholarship to the first author. We also thanks Faculty of Biosciences and Medical Engineering, Universiti Teknologi Malaysia (UTM) for providing facilities and services.

References


