

OMEGA-3 FATTY ACIDS AND ASTAXANTHIN PRODUCED BY *Chlorella vulgaris* AND *Haematococcus pluvialis* CULTIVATED IN CHICKEN MANURE
MEDIUM

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DEDICATION

This thesis is dedicated to my parents, who taught me that the best kind of knowledge to have been that which is learned for its own sake. It is also dedicated to my mother, who taught me that even the largest task can be accomplished if it is done one step at a time.

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ABSTRACT

Chicken manure contains a high concentration of nutrients such as nitrogen and phosphate, which can be used to replace commercially available medium for growth of microalgae. Cultivating microalgae by using chicken manure as the nutrients can reduce the cost of cultivation and also increase the quality of fish pellet in aquaculture systems. This research was conducted to study the growth of *Chlorella vulgaris* and *Haematococcus pluvialis* in different concentrations of chicken manure on the growth of *C. vulgaris* and *H. pluvialis*. The concentration of omega-3 fatty acids in *C. vulgaris* and astaxanthin in *H. pluvialis* were determined by cultivating *C. vulgaris* and *H. pluvialis* in Bold Basal medium and Rudic medium respectively, and compared with chicken manure medium (CM). 50% of CM concentration was considered the best chicken manure concentration to replace commercial medium in *C. vulgaris* cultivation, although the Bold Basal Medium and 50% of CM has a comparable specific growth rate of 0.178 day^{-1} and 0.512 day^{-1} respectively. However, it will be much cost effective to use a waste product with high nutrients, i.e., CM instead of commercial medium to grow the microalgae. In *H. pluvialis* cultivation, the highest specific growth rate was obtained in 50% of CM and 40% of CM with 0.554 day^{-1} and 0.528 day^{-1} , respectively. For omega-3 fatty acid, the highest concentration was obtained in 14.8% *C. vulgaris* in 10% of CM due to depletion of nitrogen, which causes the increase of fatty acid production in the microalgae. For astaxanthin production, *H. pluvialis* grown in 50% of CM gave highest concentration of astaxanthin (0.082 mg/mL). The increase in astaxanthin was due to low concentration of phosphorus in the growth medium.

ABSTRAK

Najis ayam dikatakan mengandungi kepekatan nutrien yang tinggi seperti nitrogen dan fosfat, yang dapat digunakan untuk menggantikan medium mikroalga yang sudah dikomersial untuk tumbesaran mikroalga. Mengkultur mikroalga dengan menggunakan najis ayam sebagai nutrien dapat mengurangi kos mengkultur mikroalga dan dapat meningkatkan kualiti pelet ikan dalam sistem akuakultur. Penyelidikan ini dilakukan untuk mempelajari pertumbuhan *Chlorella vulgaris* dan *Haematococcus pluvialis* dalam kepekatan najis ayam yang berlainan. Kepekatan asid lemak omega-3 dalam *C. vulgaris* dan astaxanthin dalam *H. pluvialis* ditentukan dengan mengkultur *C. vulgaris* dan *H. pluvialis* dalam medium Bold Basal dan medium Rudic dan dibandingkan dengan medium najis ayam (CM). 50% kepekatan CM dianggap kepekatan najis ayam terbaik untuk menggantikan medium komersial dalam kultur *C. vulgaris* walaupun *Bold Basal Medium* dan 50% CM tidak mempunyai perbezaan yang banyak dalam pertumbuhan mikroalga iaitu 0.178 per hari dan 0.512 per hari masing-masing. Walau bagaimanapun, ia menjimatkan kos dengan menggunakan produk buangan yang mempunyai nutrien yang tinggi, iaitu CM sebagai nutrien untuk mengkultur mikroalga. Sementara dalam kultur *H. pluvialis*, kadar pertumbuhan tinggi diperoleh pada 50% CM dan 40% CM dengan 0.554 day⁻¹ dan 0.528 day⁻¹, masing-masing. Kepekatan asid lemak omega-3 tertinggi diperoleh pada *C. vulgaris* 14.8% dalam 10% CM disebabkan oleh kekurangan nitrogen yang menyebabkan peningkatan pengeluaran asid lemak dalam mikroalga. Untuk pengeluaran astaxanthin, 50% CM memberikan kepekatan astaxanthin tertinggi yang dihasilkan (0.082 mg / mL) dalam *H. pluvialis* kerana kepekatan fosforus yang rendah dalam medium.

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LIST OF ABBREVIATIONS

FAO	-	Food and Agriculture Organization of the United Nations
WHO	-	World Health Organization
DHA		Docosahexaenoic
EPA		Eicosapentaenoic
AA		Arachidonic acid
TN		Total nitrogen
TP		Total phosphorus
BBM		Bold basal medium
RM		Rudics medium
GHG		Greenhouse gas
GWP		Global warming potential
Gt		Gigatons
GCMS		Gas chromatograph mass spectrometer
HPLC		High performance liquid chromatograph
FM		Fish meal
FO		Fish oil
dH ₂ O		Distilled water
OD		Optical density

LIST OF SYMBOLS

A_{FAME}	-	Peak area of fatty acid methyl esters
$\sum A$	-	Total peak area of FAME chromatogram
A_{C17}	-	Peak area of the internal standard (C17:0)
W_{oil}	-	Oil weight
$W_{\text{raw material}}$	-	Weight of raw material

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CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Microalgae are aquatic plants that grow by consuming the nutrients in their environment and the energy provided by the sun (Gohman, 2021). Microalgae are live microorganisms made up of carbohydrates, proteins, lipid micronutrients, and other biologically active elements (Dani et al., 2016).

Microalgae has been demonstrated to be a long-term, sustainable source of biomass and oils for fuel, food, feed, and other co-products (Mitchell, 2021). Microalgae have shown potential in fulfilling the population's demand for a more sustainable food supply, particularly protein demand. These potential protein sources provide significant advantages over other currently used raw materials (Caporgno & Mathys, 2018).

Algae can be grown in fresh water or seawater, depending on the species (Morales et al., 2019). There were numerous applications for microalgae, including cosmetics, animal feeds, and supplements. Previous study found that, *Chlorella* and *Spirulina* species were used as health-food supermarkets and the other species like *Tetraselmis*, *Nannochloropsis* and *Scenedesmus* were used as animal feed (Koyande et al., 2019). In this research, the microalgae can be used in aquaculture systems since the medium of cultivation was chicken manure.

Microalgae, which was found near the aquatic food chain, are an essential part of the diet of marine animals such as fish. Aquaculture feeding with microalgae produced a primary fresh feed or addition, such as a source of colour. Algae contain almost all of the nutrients required for a fish diet.(Safafar, 2017).

The researchers were investigated microalgae as nutrients substitutes in fish feeds due to nutritional drawbacks and low fillet consistency. Microalgae have been considered alternatives nutrients in aquaculture as it is rich in amino acids, minerals, vitamins, and long-chain n-3 fatty acids (Sarker et al., 2020).

Two types of microalgae were selected for this study, *Chlorella vulgaris* and *Haematococcus pluvialis*. Omega-3 fatty acids is a sources of fatty acids that can be obtained in *C. vulgaris* (Rismani & Shariati, 2017). According to a recent Food and Agriculture Organization of the United Nations (FAO) and World Health Organization (WHO) expert consultation, when compared to children of women who do not consume fish, mothers who give birth to children who use fish high in omega-3 fatty acids have reduce risk of poor brain plus neural system development. There was strong evidence that it can reduce the risk of death from coronary heart disease (by 36%) (Globefish, 2021).

Astaxanthin was considered as super anti-oxidant, which can be obtained from highly in *H. pluvialis* (Shah et al., 2016). The importance of astaxanthin as an antioxidant for humans is only now being recognised, and fish tinted with natural astaxanthin could be a good supply of this nutrient for humans. In the United States, only two astaxanthin sources are currently permitted for direct human ingestion as a supplement: krill oil and the algae *H. pluvialis* (Valeri & C., 2007). Each microalga has special nutrients which are likely to improve the fish feed.

To produce high-quality fish, the microalgae must be grown under medium with nutrition-rich conditions. Medium is a chemical mixture that contains nutrients for microalgae to grow. This research used chicken manure as an alternative medium to commercial mediums such as Bold Basal medium (BBM) and Rudic`s medium (RM).

According to (M. A. Yaakob et al., 2021), nitrogen and phosphate were the essential main components in a medium for algae growth. As a result, this research emphasises the amount of nitrogen and phosphate necessary for microalgae culture and the advantages of nitrogen and phosphate in enhancing the biomass productivity of microalgae. Plant waste and animal waste were two promising alternative mediums.

Chicken manure can be used to substitute the typical culture medium since it is rich in nutrients (e.g., nitrogen and phosphate) (Tan et al., 2017). Chicken manure has long been a traditional organic alternative source of fertiliser that is more cost-effective than chemical fertilisers. It also reduces manure pollution caused by improper disposal, allowing much-needed nutrients to be recovered and re-used (Agwa & Abu, 2014). Chicken manure was a good choice of source because it is readily available, dissolved rapidly and had a well-defined composition (Sipaúba-Tavares et al., 2015).

Table 1.1 shows the total livestock for different states in Malaysia in 2018. Among 10 types of livestock, poultry was recorded the most, nearly 30 times higher compared to ducks, the second most popular livestock. Accordingly, management of poultry production can be challenging (Tan, 2013). Thus, it was the motivation of this study to utilize chicken manure as an alternative culture medium for microalgae rather than being discarded and becoming a source of pollution such as greenhouse effect (Chai et al., 2019).

Table 1.1 Livestock population by states in year 2018 (Jabatan Perkhidmatan Veterinar Malaysia, 2018)

MALAYSIA : BILANGAN TERNAKAN MENGIKUT NEGERI, 2018 ^E										
Malaysia : Livestock Population By States, 2018 ^E										
NEGERI State	Kerbau Buffalo	Lembu Cattle	Kambing Goat	Bebiri Sheep	Babi Swine	Ayam Poultry	Itik Duck	Burung Unta Ostrich	Puyuh Quail	Rusa Deer
Perlis	249	6,718	3,176	1,644		1,784,417	7,838	5	10,203	287
Kedah	5,219	54,297	48,459	10,833	1,588	56,529,903	561,819	11	237,883	1,205
Pulau Pinang	608	11,934	11,008	2,811	320,165	11,342,444	343,397	1	142,930	78
Perak	9,174	45,516	27,167	3,008	545,258	39,897,038	7,463,087		43,197	5,251
Selangor	2,896	25,143	21,515	3,117	262,060	18,064,933	5,236	213	127,110	48
N. Sembilan	1,899	39,667	33,986	16,084	241	21,187,714	32,068	101	384,402	1,108
Melaka	4,408	29,272	38,121	6,902	44,025	27,000,465	84,314		381,794	422
Johor	3,788	105,712	42,463	21,870	228,639	72,686,534	1,057,353	297	1,570,996	1,749
Pahang	11,604	124,530	32,433	18,826	3,893	12,915,104	11,284	1	87,029	2,061
Terengganu	8,935	85,155	29,798	6,892		2,749,331	26,453		78,916	745
Kelantan	5,425	96,109	41,501	33,589	645	3,727,520	62,144		9,349	230
W. Persekutuan		210	108							47
Jumlah S. M'sia Total For P. M'sia	54,205	624,263	329,735	125,576	1,406,514	267,885,403	9,654,993	629	3,073,809	13,231
Sabah	54,680	75,766	56,346	2,597	101,840	9,814,509	54,861	59	16,762	
Sarawak	5,436	10,452	12,964	1,821	334,074	34,278,682	298,400		340,917	1,121
JUMLAH BESAR Grand Total	114,321	710,481	399,045	129,994	1,842,428	311,978,594	10,008,254	688	3,431,488	14,352

The study began with the *C. vulgaris* and *H. pluvialis* being grown on a chicken manure medium, followed by analyses of omega-3 fatty acids in *C. vulgaris* and astaxanthin in *H. pluvialis* using both chicken manure and commercial media (Bold Basal Medium and Rudic's Medium). This study focused on the development and analysis of omega-3 fatty acids and astaxanthin contained in microalgae inside chicken manure medium. Results obtained from this study can be formed into a product such as a fish pellet in an aquaponic system. Figure 1.1 shows the flowchart of this study.

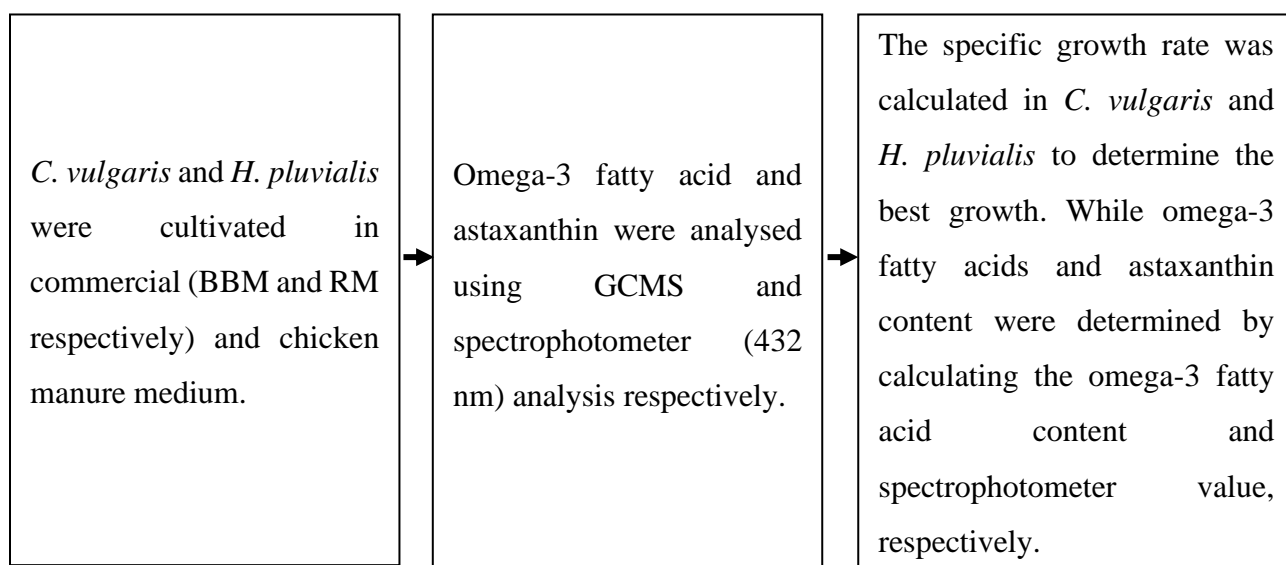


Figure 1.1 The flowchart of the research.

1.2 Problem Statement

The usage of chicken manure as a medium for microalgae is known to be cost-effective. Microalgae cultivation necessitates a considerable medium volume, resulting in high operating costs and adverse environmental effects due to nutrient leakage (Tan et al., 2017). This finding was in agreements with the study by Agwa and Abu (2014) who showed using chicken manure as an alternative medium for the cultivation of microalgae was economical.

Chicken manure can be used to grow microalgae as it provides a source of nutrients to cultivate microalgae. These nutrient sources are an effective technique to boost microalgae growth since the waste includes high concentrations of nutrients such as nitrate and phosphate (Tan et al., 2017) These elements are harmful to the environment if released untreated (Imran et al., 2007). Here, the application of chicken manure to cultivate microalgae, providing a cheap source of nutrients, will be studied.

Secondly, it is due to the nutritional deficiency and poor fillet quality in the aquaculture system. According to Young K. (2009), tilapia contains a high concentration of omega-6 fatty acids (n-6) and a low concentration of omega-3 fatty acids (n-3), this is a potentially unhealthy ratio for humans. Astaxanthin improves the fillet quality by providing colorants of pigmentation to the fish. Besides their role as colorants in food, it also have antioxidant properties, preventing the deterioration caused by oxidation in food products (Elisa et al., 2014).

Hence, this research focused on two primary nutrients, which were omega-3 fatty acids in *C. vulgaris* and astaxanthin in *H. pluvialis*. The omega-3 fatty acids and astaxanthin content in the cultivation of microalgae using chicken manure were investigated and compared with the commercial medium before it can be introduced for fish pellets in the future.

1.3 Objectives of Study

1. To study the effect of different concentration of chicken manure towards the growth of *C. vulgaris* and *H. pluvialis*.
2. To examine and compare the amount of omega-3 fatty acids produced in *C. vulgaris* cultivated in Bold Basal medium and chicken manure medium.
3. To examine and compare the amount of astaxanthin produced in *H. pluvialis* cultivated in Rudic`s medium and chicken manure medium.

1.4 Scope of the Study

The content of nitrogen and phosphorus were determined in commercially available mediums, which were Bold Basal Medium, Rudic`s and chicken manure medium. The growth of microalgae was monitored, and the optical density of microalgae was measured at 750 nm using a spectrophotometer. Omega-3 fatty acids in the *C. vulgaris* was extracted and identified using a gas chromatograph mass spectrometer (GCMS). The content of astaxanthin in the sample was quantified using a spectrophotometer at a wavelength of 432 nm by using the dry mass of *H. pluvialis*.

1.5 Significance of the study

Environmental issues are one of the significant concerns in Malaysia and need to be carefully addressed. Hence, recycling waste can help to reduce pollution. The waste from poultry can be used for nutrients in the cultivation of microalgae. For this study, chicken manure medium was used as a replacement for commercial mediums in the growth of microalgae. Chicken manure contributes to the high growth of *C. vulgaris* and *H. pluvialis* because of the nitrogen and phosphorus elements inside the chicken manure medium itself. Many countries use animal manure to raise plankton, resulting in much more fresh food for fish to eat and increased fish production. (Renalda & Hieromin, 2021). Studies have shown that, by using chicken manure as a culture medium, it can enhance the omega-3 fatty acids and astaxanthin contained in microalgae cells. Thus, it can be used to substitute the nutrients in fish feed.

REFERENCES

- Abdo, S., Ali, G., & El-Baz, F. (2015). Potential production of omega fatty acids from microalgae. *International Journal of Pharmaceutical Sciences Review and Research*, 34, 210-215.
- Agwa, O. K., & Abu, G. O. (2014). Utilization of poultry waste for the cultivation of *Chlorella* sp. for biomass and lipid production. *International Journal of Current Microbiology and Applied Sciences*, 3, 1036-1047.
- Allaguvatova, R., Myasina, Y., Zakharenko, V., & Gaysina, L. (2019). A simple method for the cultivation of algae *Chlorella vulgaris* Beijerinck. *IOP Conference Series: Earth and Environmental Science*, 390, 012020.
- Badar, S. N., Yaakob, Z., & Timmiati, S. (2017). Growth Evaluation of Microalgae Isolated from Palm Oil Mill Effluent in Synthetic Media (Penilaian Pertumbuhan Mikroalga yang telah Dipencil dari Effluen Kilang Minyak Sawit dalam Media Sintetik). *Malaysian Journal of Analytical Sciences*, 21, 82-94.
- Basak, R., Wahid, K., & Dinh, A. (2021). Estimation of the Chlorophyll-A Concentration of Algae Species Using Electrical Impedance Spectroscopy. *Water*, 13, 1223.
- Bugter, M. (2015). Iron chelating agents and their effects on the growth of *Pseudokirchneriella subcapitata*, *Chlorella vulgaris*, *Phaeodactylum tricornerutum* and *Spirulina platensis* in comparison to Fe- EDTA. 2229 – 6905, 6, 56.
- Butler, T. O., McDougall, G. J., Campbell, R., Stanley, M. S., & Day, J. G. (2017). Media Screening for Obtaining *Haematococcus pluvialis* Red Motile Macrozooids Rich in Astaxanthin and Fatty Acids. *Biology*, 7(1), 2.
- Camacho-Rodríguez, J., Macías-Sánchez, M. D., Cerón-García, M. C., Alarcón, F. J., & Molina-Grima, E. (2018). Microalgae as a potential ingredient for partial fish meal replacement in aquafeeds: nutrient stability under different storage conditions. *Journal of Applied Phycology*, 30(2), 1049-1059.
- Campo, J. A., Rodríguez, H., Moreno Fernández, J., Vargas, M. A., Rivas, J., & Guerrero, M. (2004). Accumulation of astaxanthin and lutein in *Chlorella zofingiensis* (Chlorophyta). *Applied microbiology and biotechnology*, 64, 848-854.
- Chai, R., Ye, X., Ma, C., Wang, Q., Tu, R., Zhang, L., & Gao, H. (2019). Greenhouse gas emissions from synthetic nitrogen manufacture and fertilization for main upland crops in China. *Carbon Balance and Management*, 14(1), 20.
- Corro, G., Tellez, N., Jimenez, T., Tapia, A., Banuelos, F., & Vazquez-Cuchillo, O. (2011). Biodiesel from waste frying oil. Two step process using acidified SiO₂ for esterification step. *Catalysis Today*, 166(1), 116-122.
- Da, S. F. V., & Sant'Anna, C. (2017). Impact of culture conditions on the chlorophyll content of microalgae for biotechnological applications. *World J Microbiol Biotechnol*, 33(1), 20.
- Dani, D., Thirugnanamurthy, S., Kandasamy, S., Shalini, B., Baruah, A., R, K., & Dam Roy, S. (2016). A Review on Microalgae as Potential Fish Feed Ingredient. *Journal of Andaman Science Association*, 21, 140-144.
- Ding, E. L., Hutfless, S. M., Ding, X., & Girotra, S. (2006). Chocolate and prevention of cardiovascular disease: a systematic review. *Nutrition & metabolism*, 3, 2-2.
- Eggers, L. F., & Schwudke, D. (2016). Liquid Extraction: Folch. In M. R. Wenk (Ed.), *Encyclopedia of Lipidomics* (pp. 1-6). Dordrecht: Springer Netherlands.
- El-Mohsnawy, E., El-Sheekh, M., Mabrouk, M., & Zohir, W. (2020). Enhancing Accumulation of Omega 3 and 9 Fatty Acids in *Chlorella vulgaris* Under Mixotrophic Nutrition. *Journal of Animal and Plant Sciences*, 30, 485-492.

- Elisa, H. G. P., Thiago, L. M. G., Edson, F. E. S., Marcelo, T. S. M., Jefferson, F. C., & Marcos, F. P. (2014). *Color and Carotenoids in Tilapia Fish Fed Different Carotenoid*. Paper presented at the 60th International Congress of Meat Science and Technology, Punta Del Este, Uruguay.
- Gammone, M. A., Riccioni, G., Parrinello, G., & D'Orazio, N. (2018). Omega-3 Polyunsaturated Fatty Acids: Benefits and Endpoints in Sport. *Nutrients*, *11*(1), 46.
- Globefish. (2021). Farmed fish: a major provider or a major consumer of omega-3 oils? Retrieved from <http://www.fao.org/in-action/globefish/fishery-information/resource-detail/en/c/338773/>
- Gong, X., & Chen, F. (1997). Optimization of culture medium for growth of *Haematococcus pluvialis*. *Journal of Applied Phycology*, *9*, 437-444.
- Gordillo, F. J. L., Jiménez, C., Figueroa, F. L., & Niell, F. X. (1998). Effects of increased atmospheric CO₂ and N supply on photosynthesis, growth and cell composition of the cyanobacterium *Spirulina platensis* (Arthrospira). *Journal of Applied Phycology*, *10*(5), 461.
- Griffiths, M. J., Garcin, C., Van Hille, R. P., & Harrison, S. T. L. (2011). Interference by pigment in the estimation of microalgal biomass concentration by optical density. *Journal of Microbiological Methods*, *85*(2), 119-123.
- Guerin, M., Huntley, M. E., & Olaizola, M. (2003). *Haematococcus astaxanthin*: applications for human health and nutrition. *Trends Biotechnol*, *21*(5), 210-216.
- Han, X., Rusconi, N., Ali, P., Pagkatipunan, K., & Chen, F. (2017). Nutrients Extracted from Chicken Manure Accelerate Growth of Microalga *Scenedesmus obliquus* HTB1. *Green and Sustainable Chemistry*, *7*, 101-113.
- Ilavarasi, A., Mubarakali, D., Praveenkumar, R., Baldev, E., & Thajuddin, N. (2011). Optimization of Various Growth Media to Freshwater Microalgae for Biomass Production. *Biotechnology for Biofuels*, *10*, 540-545.
- Illman, A. M., Scragg, A. H., & Shales, S. W. (2000). Increase in *Chlorella* strains calorific values when grown in low nitrogen medium. *Enzyme and Microbial Technology*, *27*(8), 631-635.
- Imamoglu, E., Fazilet, v.-s., & Dalay, M. (2007). Effect of different culture media and light intensities on growth of *Haematococcus pluvialis*. *Int J Nat Eng Sci*, *1*, 5-9.
- Imran, M., Ullah, H., & Imran, M. (2007). Nitrate and phosphate pollution in surface and ground water in Western Malaysia. *Journal- Chemical Society of Pakistan*, *29*, 315-320.
- Jabatan Perkhidmatan Veterinar Malaysia. (2018). Livestock Population By States. Retrieved from [http://www.dvs.gov.my/dvs/resources/user_1/2019/BP/Perangkaan%20Ternakan%2020182019/1\)_Malaysia_Perangkaan_Ternakan.pdf](http://www.dvs.gov.my/dvs/resources/user_1/2019/BP/Perangkaan%20Ternakan%2020182019/1)_Malaysia_Perangkaan_Ternakan.pdf)
- Jay, M. I., Kawaroe, M., & Effendi, H. (2018). Lipid and fatty acid composition microalgae *Chlorella vulgaris* using photobioreactor and open pond. *IOP Conference Series: Earth and Environmental Science*, *141*, 012015.
- Ju, Z. Y., Davis, S., Ramm, K., Steck, M., Soller, F., & Fox, B. K. (2017). Effects of microalgae-added diets on growth performance and meat composition of tilapia (*Oreochromis mossambicus*). *Aquaculture Research*, *48*(9), 5053-5061.
- Karim, A., Islam, M. A., Khalid, Z. B., Faizal, C. K. M., Khan, M. M. R., & Yousuf, A. (2020). Chapter 9 - Microalgal Cell Disruption and Lipid Extraction Techniques for Potential Biofuel Production. In A. Yousuf (Ed.), *Microalgae Cultivation for Biofuels Production* (pp. 129-147): Academic Press.

- Koyande, A. K., Chew, K. W., Rambabu, K., Tao, Y., Chu, D. T., & Show, P. L. (2019). Microalgae: A potential alternative to health supplementation for humans. *Food Science and Human Wellness*, 8(1), 16-24.
- Laurens, L. M., Quinn, M., Van Wycken, S., Templeton, D. W., & Wolfrum, E. J. (2012). Accurate and reliable quantification of total microalgal fuel potential as fatty acid methyl esters by in situ transesterification. *Anal Bioanal Chem*, 403(1), 167-178.
- Lim, K. C., Yusoff, F. M., Shariff, M., & Kamarudin, M. S. (2018). Astaxanthin as feed supplement in aquatic animals. *Reviews in Aquaculture*, 10(3), 738-773.
- Liu, X. J., Wu, Y. H., Zhao, L. C., Xiao, S. Y., Zhou, A. M., & Liu, X. (2011). Determination of astaxanthin in *Haematococcus pluvialis* by first-order derivative spectrophotometry. *J AOAC Int*, 94(6), 1752-1757.
- Mandal, S., & Mallick, N. (2009). Microalga *Scenedesmus obliquus* as a potential source for biodiesel production. *Appl. Microbiol. Biotechnol*, 84, 281-291.
- Mendes, L., & Vermelho, A. (2013). Allelopathy as a potential strategy to improve microalgae cultivation. *Biotechnology for Biofuels*, 6.
- Mohsenpour, S., & Willoughby, N. (2016). Effect of CO₂ aeration on cultivation of microalgae in luminescent photobioreactors. *Biomass and Bioenergy*, 85, 168-177.
- Morales, M., Collet, P., Lardon, L., Helias, A., Steyer, J. P., & Bernard, O. (2019). *Biofuels from Algae* (2nd ed.).
- Nigam, S., Prakash, M., & Sharma, R. (2011). Effect of Nitrogen on Growth and Lipid Content of *Chlorella pyrenoidosa*. *American Journal of Biochemistry and Biotechnology*, 7, 126-131.
- Nzayisenga, J. C., Farge, X., Groll, S. L., & Sellstedt, A. (2020). Effects of light intensity on growth and lipid production in microalgae grown in wastewater. *Biotechnology for Biofuels*, 13(1), 4.
- Oh, M. J., Nam, J. J., Lee, E. O., Kim, J. W., & Park, C. S. (2016). A synthetic C16 omega-hydroxyphytoceramide improves skin barrier functions from diversely perturbed epidermal conditions. *Arch Dermatol Res*, 308(8), 563-574.
- Oleic Acid. (2005). Retrieved from <https://www.webmd.com/vitamins/ai/ingredientmono-1614/oleic-acid>
- Oo, Y., Su, M., & Kyaw, K. T. (2017). *Extraction And Determination Of Chlorophyll Content From Microalgae*.
- Orosa, M., Valero, J. F., Herrero, C., & Abalde, J. (2001). Comparison of the accumulation of astaxanthin in *Haematococcus pluvialis* and other green microalgae under N-starvation and high light conditions. *Biotechnology Letters*, 23(13), 1079-1085.
- Oslan, S. N. H., Shoparwe, N. F., Yusoff, A. H., Rahim, A. A., Chang, C. S., Tan, J. S., . . . Mohamed, M. S. (2021). A Review on *Haematococcus pluvialis* Bioprocess Optimization of Green and Red Stage Culture Conditions for the Production of Natural Astaxanthin. *Biomolecules*, 11(2).
- Othman, F., Jamaluddin, H., Ibrahim, Z., Hara, H., Ashyikin, N., Iwamoto, K., & Mohamad, S. (2019). Production of α -linolenic Acid by an Oleaginous Green Algae *Acutodesmus obliquus* Isolated from Malaysia. *Journal of Pure and Applied Microbiology*, 13, 1297-1306.
- Pan, A., Chen, M., Chowdhury, R., Wu, J. H. Y., Sun, Q., Campos, H., & Hu, F. B. (2012). α -Linolenic acid and risk of cardiovascular disease: a systematic review and meta-analysis. *The American journal of clinical nutrition*, 96(6), 1262-1273.
- Panahi, Y., Yari Khosroushahi, A., Sahebkar, A., & Heidari, H. R. (2019). Impact of Cultivation Condition and Media Content on *Chlorella vulgaris* Composition. *Advanced pharmaceutical bulletin*, 9(2), 182-194.

- Panis, G., & Rosales Carreón, J. (2016). Commercial astaxanthin production derived by green alga *Haematococcus pluvialis*: A microalgae process model and a techno-economic assessment all through production line. *Algal Research*, 18, 175-190.
- Prečanica, M. (2020). *Upotreba otpadne vode od uzgoja riba u RAS-u za ugoj mikroalga (Master's thesis)*. (2021). Retrieved from <https://urn.nsk.hr/urn:nbn:hr:155:504956> (25 September)
- Renalda, N. M., & Hieromin, A. L. (2021). Animal Waste and Agro-by-Products: Valuable Resources for Producing Fish at Low Costs in Sub-Saharan Countries. *IntechOpen*.
- Rinawati, M., Sari, L. A., & Pursetyo, K. T. (2020). Chlorophyll and carotenoids analysis spectrophotometer using method on microalgae. *IOP Conference Series: Earth and Environmental Science*, 441, 012056.
- Rismani, R., & Shariati, M. (2017). Changes of the Total Lipid and Omega-3 Fatty Acid Contents in two Microalgae *Dunaliella Salina* and *Chlorella Vulgaris* Under Salt Stress. *Brazilian Archives of Biology and Technology*, 60.
- Safar, H. (2017). *Micro -Algae Biomass as an Alternative Resource for Fishmeal and Fish Oil in The Production of Fish Feed*. Technical University of Denmark,
- Safi, C., Zebib, B., Merah, O., Pontalier, P.-Y., & Vaca-Garcia, C. (2014). Morphology, composition, production, processing and applications of *Chlorella vulgaris*: A review. *Renewable and Sustainable Energy Reviews*, 35, 265-278.
- Santos Ballardo, D., Rossi, S., Hernández, V., Vázquez, R., Unceta, C., Caro-Corrales, J., & Valdez, A. (2015). A simple spectrophotometric method for biomass measurement of important microalgae species in aquaculture. *Aquaculture*, 448.
- Sarker, P. K., Kapuscinski, A. R., McKuin, B., Fitzgerald, D. S., Nash, H. M., & Greenwood, C. (2020). Microalgae-blend tilapia feed eliminates fishmeal and fish oil, improves growth, and is cost viable. *Scientific Reports*, 10(1), 19328.
- Shah, M. M. R., Liang, Y., Cheng, J. J., & Daroch, M. (2016). Astaxanthin-Producing Green Microalga *Haematococcus pluvialis*: From Single Cell to High Value Commercial Products. *Frontiers in plant science*, 7, 531-531.
- Singh, G., Shamsuddin, M. R., Aqsha, & Lim, S. W. (2018). Characterization of Chicken Manure from Manjung Region. *IOP Conference Series: Materials Science and Engineering*, 458, 012084.
- Sipaúba-Tavares, L., Berchielli-Morais, F., & Scardoeli-Truzzi, B. (2015). Growth of *Haematococcus pluvialis* Flotow in alternative media. *Brazilian journal of biology = Revista brasleira de biologia*, 75.
- Tan, C. (2013). Issues on being nation's largest chicken producer. *The Star*. Retrieved from <https://www.thestar.com.my/news/community/2013/12/09/issues-on-being-nations-largest-chicken-producer-poor-livestock-site-management-meant-housefly-pro>
- Tan, X. B., Uemura, Y., Lim, J. W., & Lam, M. K. (2017). Cultivation of *Chlorella Vulgaris* in photobioreactor by using compost as a nutrient source for biomass production. *Journal of Fundamental and Applied Sciences*, 9(6S), 288-297.
- Tharek, A., Mohamad, S., Iwamoto, K., Hara, H., Ashyikin, N., Kaha, M., & Jamaluddin, H. (2020). Astaxanthin production by tropical microalgae strains isolated from environment in malaysia. *Asian Journal of Microbiology, Biotechnology and Environmental Sciences*, 22, 168-173.
- Toppe, J. (2021). Farmed fish: a major provider or a major consumer of omega-3 oils? Retrieved from <http://www.fao.org/in-action/globefish/fishery-information/resource-detail/en/c/338773/>

- Towers, L. (2013). The Use of Algae in Fish Feeds as Alternatives to Fishmeal. Retrieved from <https://thefishsite.com/articles/the-use-of-algae-in-fish-feeds-as-alternatives-to-fishmeal>
- Toyub, M., Miah, M., Habib, M. A., & Rahman, M. (2012). Growth Performance And Nutritional Value Of *Scenedesmus Obliquus* Cultured In Different Concentrations Of Sweetmeat Factory Waste Media. *Bangladesh Journal of Animal Science*, 37.
- Ulmer, C. Z., Jones, C. M., Yost, R. A., Garrett, T. J., & Bowden, J. A. (2018). Optimization of Folch, Bligh-Dyer, and Matyash sample-to-extraction solvent ratios for human plasma-based lipidomics studies. *Analytica chimica acta*, 1037, 351-357.
- Valeri, L. H., & C., G. R. (2007). Astaxanthin the most prevalent carotenoid in the marine environment. Retrieved from <https://www.globalseafood.org/advocate/astaxanthin-feed-supplement-enhances-pigmentation-health/>
- Wayama, M., Ota, S., Matsuura, H., Nango, N., Hirata, A., & Kawano, S. (2013). Three-Dimensional Ultrastructural Study of Oil and Astaxanthin Accumulation during Encystment in the Green Alga *Haematococcus pluvialis*. *PLoS one*, 8, e53618.
- Wong, Y.-K., Ho, Y.-H., Ho, k. C., Ho Man, L., & Yung, K. (2017). Growth Medium Screening for *Chlorella vulgaris* Growth and Lipid Production. *J Aquac Mar Biol*, 6, 00143.
- Yaakob, Maizatul, A. M., Radin, M. S. R., Al-Gheethi, A., Aswathnarayana Gokare, R., & Ambati, R. R. (2021). Influence of Nitrogen and Phosphorus on Microalgal Growth, Biomass, Lipid, and Fatty Acid Production: An Overview. *Cells*, 10(2), 393.
- Yaakob, M. A., Mohamed, R., Al-Gheethi, A., Gokare, R., & Ambati, D. R. R. (2021). Influence of Nitrogen and Phosphorus on Microalgal Growth, Biomass, Lipid, and Fatty Acid Production: An Overview. *Cells*, 10, 393.
- Yahya, N. A., Suhaimi, N., Kaha, M., Hara, H., Zakaria, Z., Sugiura, N., & Iwamoto, K. (2018). Lipid production enhancement in tropically isolated microalgae by azide and its effect on fatty acid composition. *Journal of Applied Phycology*, 30(6), 3063-3073.
- Yeesang, C., & Cheirsilp, B. (2011). Effect of nitrogen, salt, and iron content in the growth medium and light intensity on lipid production by microalgae isolated from freshwater sources in Thailand. *Bioresource Technol.*, 102, 3034-3040.
- Zhu, Z., Jiang, J., & Fa, Y. (2020). Overcoming the Biological Contamination in Microalgae and Cyanobacteria Mass Cultivations for Photosynthetic Biofuel Production. *Molecules (Basel, Switzerland)*, 25(22), 5220.