EXTRACTION AND IDENTIFICATION OF ANANAIN-LIKE PROTEASE FROM PINEAPPLE CROWN AND STEM

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DEDICATION

In the name of Allah the Most Gracious and the Most Merciful,

I would like to dedicate my deepest appreciation to the following people, who are very meaningful in my life, people that always inspires and lifting me up, support me emotionally with prayer, love and patience along the journey of this life. Each of them is the biggest spirit for me.

To My Great Parents and Siblings

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ABSTRACT

Pineapple (Ananas comosus) is the major tropical fruits and belongs to the family Bromeliaceae. The pineapple waste from pineapple industry has contributed to an increase of waste in Malaysia and around the world every year. Hence, there is a demand need to study the utilization of pineapple wastes such as pulp, peel, core, leaves and crown for industrial applications instead of disposing them, which may result in the loss of important biomass and nutrients. The pineapple contains endopeptidase enzymes that constitute the major components of bromelain extract which are stem bromelain, fruit bromelain, ananain and comasain. This study was conducted to extract and purify protease from crown and stem of MD2 pineapple. Protease was extracted and purified using anion exchange chromatography (IEX), gel filtration, desalting method before being identified using LC/MS. Proteolytic activity was determined using Casein Digestion Unit (CDU) and well diffusion method, whereas fibrinolytic activity was determined using fibrin suspension. In the present study, proteolytic assay showed 1 kg crown of MD2 cultivar produced the activity of 126.0 ± 3.86 U/mL, the specific activity of 3937.50 U/mg and total activity of 3.94×10^9 U. In another assessment, 1 kg stem from MD2 cultivar showed the proteolytic activity, specific activity and total activity of 118.5 ± 1.19 U/mg, 5925 U/mg and 5.925×10^9 U, respectively. The proteolytic activity and fibrinolytic activity of purified enzyme from the crown extract were 144.02 U/mL and 0.51×10^{-6} U/mL, respectively. The molecular weight of the purified enzyme was in the range of 25 to 35 kDa at the optimum condition of pH 7 at 37 °C. Meanwhile, a purified enzyme from stem extract was observed to contain high proteolytic activity and fibrinolytic activity which were 20.76 U/mL and 0.000272 U/mL, respectively. Purification of the extract yielded a band at molecular weight of between 20-25 kDa at the optimum conditions of pH 3 and 9 at 60 °C. From LC/MS analysis, the purified enzyme from the crown extract was similar to ananain under accession number A0A199VSS3 (according to Uniprot). It had seven unique peptides and covered 164/356 amino acids (44.9 percent coverage). The ananain (EC 3.4.22.31) is classified in the subfamilies of cysteine protease C1A (clan CA, family C1) that is peptidase family related to papain. In conclusion, protease was successfully extracted and identified as ananain-like protease from crown. However, protease extracted from the stem of MD2 cultivar was not determined. Therefore, further study on this enzyme needs to be explored in the near future.

ABSTRAK

Nanas (Ananas comosus) adalah buah tropika utama dan tergolong dalam keluarga Bromeliaceae. Sisa nanas daripada industri nanas telah menyumbang kepada peningkatan sisa di Malaysia dan di seluruh dunia setiap tahun. Oleh itu, terdapat keperluan untuk mengkaji penggunaan sisa nanas seperti pulpa, kulit, inti, daun dan jambul untuk aplikasi industri dan bukan membuangnya, yang boleh mengakibatkan kehilangan biojisim dan nutrien penting. Nanas mengandungi enzim endopeptidase yang merupakan komponen utama ekstrak bromelain iaitu bromelain batang, bromelain buah, ananain dan komasain. Kajian ini telah dijalankan untuk mengekstrak dan menulenan protease daripada jambul and batang nanas MD2. Protease telah diekstrak and ditulenkan menggunakan kromatografi pertukaran anion (IEX), penyaringan, kaedah penyahgaraman sebelum dikenal pasti menggunakan LC/MS. Aktiviti proteolitik telah ditentukan menggunakan Casein Digestion Unit (CDU) dan kaedah resapan agar, manakala aktiviti fibrinolitik ditentukan menggunakan ampaian fibrin. Keputusan kajian mendapati ujian proteolitik menunjukkan 1 kg jambul MD2 kultivar menunjukkan aktiviti 126.0 ± 3.86 U/mL, aktiviti khusus 3937.50 U/mg dan jumlah aktiviti 3.94×10^9 U. Dalam penilaian lain, 1 kg batang dari MD2 kultivar menunjukkan aktiviti proteolitik, aktiviti spesifik dan jumlah aktiviti masing-masing 118.5 ± 1.19 U/mL, 5925 U / mg dan 5.925×10^9 U. Aktiviti proteolitik dan aktiviti fibrinolitik enzim yang ditulenkan daripada ekstrak jambul masing-masing adalah 144.02 U/mL dan 0.51×10^{-6} U/mL. Berat molekul enzim tulen berada dalam julat 25 hingga 35 kDa di keadaan optimum pada pH 7 pada 37 °C. Sementara itu, enzim yang ditulenkan daripada ekstrak batang dilihat mengandungi aktiviti proteolitik tinggi dan aktiviti fibrinolitik masing-masing iaitu 20.76 U/mL dan 0.000272 U/mL. Penulenan ekstrak menghasilkan jalur pada berat molekul antara 20-25 kDa di keadaan optimum pH 3 dan 9 pada 60 °C. Berdasarkan hasil analisi LC/MS, enzim yang ditulenkan dari ekstrak jambul adalah menyamai dengan ananain dengan nombor aksesi A0A199VSS3 (menurut Uniprot). Ia mempunyai tujuh peptida unik dan merangkumi 164/356 asid amino (liputan 44.9 peratus). Ananain (EC 3.4.22.31) dikelaskan dalam subkeluarga sisteina protease C1A (klan CA, keluarga C1) iaitu keluarga peptidase yang berkaitan dengan papain. Kesimpulannya, enzim berjaya diekstrak dan dikenal pasti sebagai ananain-serupa protease daripada jambul. Walau bagaimanapun, protease vang diekstrak daripada batang kultivar MD2 masih belum ditentukan. Oleh itu, kajian lanjut berkaitan enzim ini perlu dikenal pasti dalam masa terdekat.

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LIST OF ABBREVIATIONS

MPIB	-	Malaysian Pineapple Industry Board
MGI	-	Malaysian Genome Institute
CDU	-	Casein Digestion Unit
ATPS	-	Aqueous Two-Phase System
RME	-	Reverse Micelle Extraction
AEX	-	Anion Exchange Chromatoraphy
GF	-	Gel Filtration Chromatography
UTM	-	Universiti Teknologi Malaysia
LC/MS	-	Liquid Chromatography-Mass Spectrometry

LIST OF SYMBOLS

°C	-	Degree Celcius
kg	-	Kilogram
g	-	Gram
cm	-	Centimetre
mm	-	Milimetre
L	-	Litre
mL	-	Mililitre
%	-	Percent

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CHAPTER 1

INTRODUCTION

1.1 Introduction

Pineapple (Ananas comosus) is the most common fruit in the world and belongs to the Bromeliaceae family. Ananas comosus is grown in large numbers in tropical countries such as China, Malaysia, Indonesia, Thailand, Philippines, Kenya and India (Ramli et al., 2017). Today, world pineapple production continues to increase, including in Malaysia every year. In Malaysia, the three most common cultivars that are widely planted are Spanish (also known as Maspine, Josapine and Hybrid pineapple), Smooth Cayenne (Sarawak pineapple) and Queen (Hidayat et al., 2012). The introduction of MD2 (hybrid variety) into the pineapple market in Malaysia has increased its demand and a gradual shift from export of Smooth Cayenne to MD2 can also be seen. Moris, Sarawak and Josapine are cultivated mainly for the local fresh fruit consumption whereas MD2 and N36 are usually exported since they have a longer shelf life. Pineapple cultivars MD2 act as a benchmark in the development of future cultivars for fresh consumption. The average pineapple weight is about 1.5 kg each and has a shelf life similar to other fruits such as papaya (Thalip et al., 2015; Falah et al., 2015). According to the Malaysia Pineapple Industry Board (MPIB), in 2017, Malaysia has a total cultivation area of 13, 733 with the production of 394,000 tonnes and the value of RM515 million while exports of RM155 million for all varieties of pineapple. For MD2, the production has increased and reached 3,600 tonnes by the end of the year 2017. The MD2 pineapple production at 3,800 tonnes by 2019 saw an increase of 200 tonnes. Malaysia currently targets RM320 million for the export of pineapple products with a total production of 700,000 metric tonnes by 2020. 360 hectares of MD2 pineapple plants are being grown commercially in Ulu Tiram, Kluang and most recently in Tanah Abang, Mersing, Johor. The current planning is to expand the crop to 1,000 hectares by 2021. The aim is to produce 100 containers together with

1,800 tonnes of pineapple per month including 50 containers or 900 tonnes of pineapple would be exported.

Recent advances in the pineapple industry have led to an increase in waste in Malaysia and around the world every year. The waste proportions are twice the proportion of fruit for flesh, core, peel and crown of Ananas comosus (Nor et al., 2015). Disposal of pineapple waste leads to the loss of valuable biomass and nutrients that contribute to industrial uses (Mirabella et al., 2014). Pineapple waste has been discovered as a possible source of proteases like bromelain (Abreu & Figueiredo 2019). Bromelain is a protease enzyme extracted from the pineapple plant that is mostly composed of proteinase and non-enzymatic components (Arshad et al., 2017). The major part of bromelain is from proteases such as stem bromelain (EC 3.4.22.32), fruit bromelain (EC 3.4.22.33), ananain (EC 3.4.22.31) and comosain whereas nonproteases consist of carbohydrates, glucosidases, cellulases, phosphatases, peroxidases and glycoproteins (Sahoo & Das 2017). Furthermore, aside from the pineapple stem and fruit as a source of bromelain, it was revealed that several other parts of the pineapple, such as the core, peel, and crown were also sources of protease enzyme (Ketnawa et al., 2012). This suggested that protease enzyme extracted from pineapple can be utilized to fulfill the demand in industrial processes including foods, textile and medical cosmeceutical industries. Protease enzyme has been shown to aid in the tenderization of meat (Arshad et al., 2017), the relaxation and suppression of shrinkage in baking industries (Sahoo & Das 2017), the improvement of silk and wool quality (Arshad et al., 2014), and the removal of tooth stains in tooth whitening products (Münchow et al., 2016). In addition, topical application of protease enzyme is introduced in the market mainly for debridement of skin wounds and burns (Muhammad et al., 2017), it acts as a fibrinolytic agent which increases fibrinolysis by activating the conversion of plasminogen to plasmin (Kwatra 2019), anti-cancer agents act as a suppressor for carcinogenesis and prevent cancer development (Rathnavelu et al., 2016) and anti-inflammatory agent to treat patients in the third molar extraction postoperative period (Barrera-Núñez et al., 2014). However, enzyme purity is necessary for industrial and commercial applications (Nor et al., 2015). Since a high degree of purity is required, entire downstream processing is necessary, including extraction, purification using ion-exchange chromatography, and

identification of purified enzyme. Therefore, pineapple stems, fruits and waste products such as crowns, leaves, peel and core are extracted and purified with different homogenization processes and addition of different extraction buffers to produce beneficial products (Youryon *et al.*, 2018; Wan *et al.*, 2016).

1.2 Problem Statement

Growing pineapple crops in Malaysia generates millions of tons of plant waste. During harvesting activities, a large amount of leftover from pineapple growing, known as agro-waste was created in the agricultural process. According to Nor et al., (2015), the waste proportions are twice the proportion of fruit for the flesh, core, peel and crown of the Smooth Cayenne pineapple. In addition, 998 million tons of agricultural waste are produced globally each year, with 1.2 million tons of agricultural waste disposed of into landfills in Malaysia each year (Neh and Ali, 2020). Agro-waste is predicted to account for 15% of total waste created in Asia with agricultural waste produced in Malaysia reaching 0.122 (kg/cap/day) in 2009 and expected to reach 0.210 (kg/cap/day) by 2025 (Ngoc and Schnitzer, 2009). Generally, harvest leftovers from crops such as pineapple waste are usually just burnt or left to rot, releasing carbon dioxide and methane gas into the atmosphere. These wastes are disposed of unethically in the environment, due to the high price of transportation and lack of land to be used for waste disposal. Consequently, disposal of pineapple waste causes environmental issues and leads to depletion of valuable biomass and nutrients from fruit processing waste (Mirabella et al., 2014). It was revealed that several other parts of the pineapple, such as the core, peel, and crown were sources of protease enzyme (Ketnawa et al., 2012). These protease enzyme have been shown to be applicable in many industrial processes including the foods, cosmetics and pharmaceutical industries (Nu`n ez et al., 2005; Muhammad & Ahmad, 2017). Hence, there is an increasing need to study the sustainable utilisation of pineapple wastes for industrial applications that can be commercialised as an environment-friendly alternative for carbon source utilization.

1.3 Objectives

The objectives of the research are:

- (a) To determine the proteolytic activity in stem and crown of pineapple and compare with different cultivars in Malaysia
- (b) To purify and characterise the protease extract from pineapple stem and crown from the best cultivar
- (c) To determine fibrinolytic activity of protease extract from pineapple stem and crown of the best cultivar

1.4 Research Scope

The area of this research was at Bahagian Pembangunan Teknologi Nanas Lembaga Perindustrian Nanas Malaysia, Pekan Nenas Johor, Laboratory of Plant Biotechnology at T02 in Faculty Science, University Technology Malaysia, Johor Bahru and Malaysian Genome Institute. This research applied the viable method for extraction, purification using ion-exchange chromatography, and identification of purified enzyme from different parts of pineapples specifically crown and stem in different cultivars. Besides that, this study also covered the characterization of protease enzyme extract and the fibrinolytic activity.

1.5 Significance of Study

In this study, significant biomass and nutrients were identified in pineapple waste, such as the crown and stem, for industrial uses. This study determined the best cultivar and pineapple waste, either the crown or stems that generated the maximum proteolytic activity. Finally, the protease enzyme was purified and tested for fibrinolytic activity for industrial usage.

REFERENCES

- Abbas, S., Shanbhag, T. and Kothare, A. (2021) 'Saudi Journal of Biological Sciences Applications of bromelain from pineapple waste towards acne', *Saudi Journal of Biological Sciences*. The Authors, 28(1), pp. 1001–1009.
- Abreu, D. C. A. and Figueiredo, K. C. de S. (2019) 'Bromelain Separation And Purification', *Brazilian Journal of Chemical Engineering Chemical*, 36(02), pp. 1029–1039.
- Agustanti, A., Ramadhani, S. A., Adiatman, M., Rahardjo, A., Callea, M., Yavuz, I. and Maharani, D. A. (2017) 'Efficacy test of a toothpaste in reducing extrinsic dental stain', *Journal of Physics*.
- Ahsan, F., Ansari, T. M., Usmani, S., Bagga, P. and Cocoon, S. (2017) 'An Insight on Silk Protein Sericin : From Processing to Biomedical Application Sericulture Silk transformation', 68, pp. 317–327.
- Ali, Sharjahan Mohamed, Yee Shin Tan, Jegadeesh Raman, Hariprasath Lakshmanan, Tau Chuan Ling, Chia Wei Phan, and Vikineswary Sabaratnam. 2017. "Do Culinary Mushrooms Have Fibrinolytic Activities?" *Biomedical Reviews* 28 (April): 91–99.
- Amini, A., Masoumi-moghaddam, S., Ehteda, A. and Morris, D. L. (2014) 'Bromelain and N-acetylcysteine inhibit proliferation and survival of gastrointestinal cancer cells in vitro: significance of combination therapy', *Journal of Experimental & Clinical Cancer Research*, 33(92), pp. 1–15.
- Arshad, Z. I., Amid, A., Yusof, F., Sulaiman, S. Z., Mudalip, S. K. A., Man, R. C. and Shaarani, S. M. (2017) 'Comparison of purification methods to purify recombinant bromelain from Escherichia coli BL21-A1', *Malaysian Journal* of Analytical Sciences, 21(4), pp. 958–971.
- Arshad, Z. I. M., Amid, A., Yusof, F., Jaswir, I., Ahmad, K. and Loke, S. P. (2014)
 'Bromelain: An overview of industrial application and purification strategies', *Applied Microbiology and Biotechnology*, 98(17), pp. 7283–7297.
- Asim, M., Abdan, K., Jawaid, M., Nasir, M., Dashtizadeh, Z., Ishak, M. R. and Hoque,
 M. E. (2015) 'A Review on Pineapple Leaves Fibre and Its Composites', *International Journal of Polymer Science*, 2015, pp. 16.

- Babagana, K. and Bala, M. (2015) 'Comparative Study on The Extraction and Purification The Stem and Fruit Bromelain From Pineapple (Ananas comosus)', *Journal of Natural Sciences Research*, 5(18), pp. 30–35.
- Bala, M., Mel, M., Saedi Jami, M., Amid, A. and Mohd Salleh, H. (2013) 'Kinetic studies on recombinant stem bromelain', *Advances in Enzyme Research*, 01(03), pp. 52–60.
- Barrera-Núñez, M. del C. de la, Yáñez-Vico, R.-M., Batista-cruzado, A., Heurtebise-Saavedra, J.-M., Oyagüe, R. C. and Torres-Lagares, D. (2014) 'Prospective double-blind clinical trial evaluating the effectiveness of bromelain in the third molar extraction postoperative period', *Journal section: Oral Surgery*, 19(2), pp. 157–62.
- Botinestean, C., Gomez, C., Gomez, C., Nian, Y., Auty, M. A. E., Kerry, J. P. and Hamill, R. M. (2017) 'consumers using fruit-derived proteolytic enzymes Possibilities for developing texture-modified beef steaks suitable for older consumers using fruit-derived proteolytic enzymes', (April 2018).
- Chaurasiya, R. S., Sakhare, P. Z., Bhaskar, N. and Hebbar, H. U. (2015) 'Efficacy of reverse micellar extracted fruit bromelain in meat tenderization', *Journal of Food Science and Technology*, 52(6), pp. 3870–3880.
- Cordts, T., Horter, J., Vogelpohl, J., Kremer, T. and Kneser, U. (2016) 'Enzymatic debridement for the treatment of severely burned upper extremities – early single center experiences', *BMC Dermatology*. BMC Dermatology, 16(8), pp. 1–7.
- Corzo, C. A., Waliszewski, K. N. and Welti-Chanes, J. (2012) 'Pineapple fruit bromelain affinity to different protein substrates', *Food Chemistry*. Elsevier Ltd, 133(3), pp. 631–635.
- Costa, H. B., Fernandes, P. M. B., Romão, W. and Ventura, J. A. (2014) 'A new procedure based on column chromatography to purify bromelain by ion exchange plus gel filtration chromatographies', *Industrial Crops and Products*. Elsevier B.V., 59, pp. 163–168.
- Danso, K., Ayeh, K., Oduro, V., Amiteye, S. and Amoatey, H. (2008) 'Effect of 6-Benzylaminopurine and -Naphthalene Acetic Acid on In vitro Production of MD2 Pineapple Planting Materials', *World Applied Sciences Journal*, 3(4), pp. 614–619.

- Dhaneshwar, A. D., Chaurasiya, R. S. and Hebbar, H. U. (2014) 'Process optimization for reverse micellar extraction of stem bromelain with a focus on back extraction', *Biotechnology Progress*, 30(4), pp. 845–855.
- Ding, X., Cai, J. and Guo, X. (2014) 'Effect of Surfactant Structure on Reverse Micellar Extraction of Ovalbumin', *Process Biochemistry*. Elsevier Ltd, 11(013), pp. 1–28.
- Doneva, M., Miteva, D., Dyankova, S., Nacheva, I., Metodieva, P. and Dimov, K. (2015) 'Efficiency of Plant Protease Bromelain and Papain on Turkey Meat Tenderness', *Biotechnology in Animal Husbandry*, 31(3), pp. 407–413.
- Engwerda, C., Peek, K., Gb, E. and Mtimkulu, A. (2017) '(12) United States Patent (10) Patent No .':, 2(12).
- Falah, M. A. F., Dima, M. D. and Suryandono, A. (2015) 'Effects of Storage Conditions on Quality and Shelf-life of Fresh-cut Melon (Cucumis melo L.) and Papaya (Carica papaya L.)', *Italian Oral Surgery*. Elsevier Srl, 3, pp. 313–322.
- Ferreira, J. F., Carlos, J., Santana, C. and Tambourgi, E. B. (2011) 'The Effect of pH on Bromelain Partition from Ananas comosus by PEG4000 / Phosphate ATPS', *Brazilian Archives of Biology and Technology*, 54(1), pp. 125–132.
- Gioia, L. C., Ganancio, J. R. and Steel, C. J. (2017) 'We are IntechOpen , the world ' s leading publisher of Open Access books Built by scientists , for scientists TOP 1 %'.
- Gokoglu, N., Yerlikaya, P., Ucak, I. and Aydan, H. (2016) 'Effect of bromelain and papain enzymes addition on physicochemical and textural properties of squid (Loligo vulgaris)', *Journal of Food Measurement and Characterization*. Springer US, pp. 1–7.
- Hale, L. P., Greer, P. K., Trinh, C. T. and James, C. L. (2005) 'Proteinase activity and stability of natural bromelain preparations', *International Immunopharmacology*, 5, pp. 783–793.
- Han, J., Cai, Y., Xie, X., Wang, Y., Wang, L., Li, C., Rao, W. and Ni, L. (2018) 'A simple method for purification of bromelain in a thermosensitive triblock copolymer-based protection system and recycling of phase components', *Separation Science and Technology (Philadelphia)*, 53(4), pp. 636–644.
- Hebbar, U. H., Hemavathi, A. B., Sumana, B. and Raghavarao, K. S. M. S. (2011) 'Reverse micellar extraction of bromelain from pineapple (Ananas comosus L.

Merryl) waste: Scale-up, reverse micelles characterization and mass transfer studies', *Separation Science and Technology*, 46(10), pp. 1656–1664.

- Hebbar, U. H., Sumana, B., Hemavathi, A. B. and Raghavarao, K. S. M. S. (2012)
 'Separation and Purification of Bromelain by Reverse Micellar Extraction Coupled Ultrafiltration and Comparative Studies with Other Methods', *Food* and Bioprocess Technology, 5(3), pp. 1010–1018.
- Hebbar, U. H., Sumana, B. and Raghavarao, K. S. M. S. (2008) 'Use of reverse micellar systems for the extraction and purification of bromelain from pineapple wastes', *Bioresource Technology*, 99(11), pp. 4896–4902.
- Helen, M., S.2, T. D., Godwin, J. J., Jinto, J. and Anitha, C. (2019) 'Premilinary Phytochemical Screening and Antioxidant Activity of Leaf, Stem and Fruit of Ananas Comosus', *World Journal of Pharmaceutical Research*, 8(5), pp. 1407–1416.
- Hidayat, T., Abdullah, F. I., Kuppusamy, C., Samad, A. A. and Wagiran, A. (2012)
 'Molecular Identification of Malaysian Pineapple Cultivar based on Internal Transcribed Spacer Region', *APCBEE Procedia*, 4, pp. 146–151.
- Indrajeet, Ojha, S. and Singh, S. (2017) 'Extraction and Purification of Bromelain from Pineapple Fruit Pulp and Peel and Comparative Study of Enzymatic Activities', *International Journal of Basic and Applied Biology*, 4(1), pp. 4–7.
- Kahiro, S. ., Kagira, J. ., Maina, N., Karanja, S. . and Njonge, F. . (2017) 'Enzymatic Activity of Bromelain from Crude Extracts of Crown, Peels and Stem of Pineapples from Different Agro-ecological Zones of Thika Region, Kenya', *Asian Journal of Biotechnology and Bioresource Technology*, 1(2), pp. 1–6.
- Kaur, A. and Chakraborty, J. N. (2015) 'Optimization of Bromelain Treatment pH with Wool for Antifelting and Reduced Pilling Behaviour : Objective Assessment Approach', 2015.
- Kaur, T., Kaur, A. and Grewal, R. K. (2015) 'Kinetics studies with fruit bromelain (Ananas comosus) in the presence of cysteine and divalent ions', *Journal of Food Science and Technology*, 52(9), pp. 5954–5960.
- Ketnawa, S., Chaiwut, P. and Rawdkuen, S. (2012) 'Pineapple wastes: A potential source for bromelain extraction', *Food and Bioproducts Processing*. Institution of Chemical Engineers, 90(3), pp. 385–391.

- Ketnawa, S., Rawdkuen, S. and Chaiwut, P. (2010) 'Two phase partitioning and collagen hydrolysis of bromelain from pineapple peel Nang Lae cultivar', *Biochemical Engineering Journal*. Elsevier B.V., 52(2–3), pp. 205–211.
- Kowalska, H., Czajkowska, K., Cichowska, J. and Lenart, A. (2017) 'SC', *Trends in Food Science & Technology*. Elsevier Ltd.
- Krishnan V, A. and Gokulakrishnan (2015) 'Extraction and purification of bromelain from pineapple and determination of its effect on bacteria causing peridontitis', *International Journal of Pharmaceutical Sciences and Research*, 6(12), pp. 5284–5294.
- Kurnia, S. D., Setiasih, S., Handayani, S. and Hudiyono, S. (2018) 'Kinetic studies of partially purified bromelain from Bogor pineapple (Ananas comosus [L.] Merr) core with ion exchange chromatography and in vitro evaluation of its antiplatelet activity Kinetic Studies of Partially Purified Bromelain from Bogor Pinea', *AIP Conference Proceedings*, 020077.
- López-García, B., Hernández, M. and Segundo, B. S. (2012) 'Bromelain, a cysteine protease from pineapple (Ananas comosus) stem, is an inhibitor of fungal plant pathogens', *Letters in Applied Microbiology*, 55(1), pp. 62–67.
- Lourenço, C. B., Ataide, J. A., Cefali, L. C., Novaes, L. C. d. L., Moriel, P., Silveira, E., Tambourgi, E. B. and Mazzola, P. G. (2016) 'Evaluation of the enzymatic activity and stability of commercial bromelain incorporated in topical formulations', *International Journal of Cosmetic Science*, 38(5), pp. 535–540.
- Manea, I., Manea, L., Marinescu, V., Science, F. and Street, S. (2016) 'Study Concerning The Proteolytic Activity of Vegetable Enzyme', 201(2).
- Mark, A., Joseph, P. and Ruth, M. (2017) 'UCC Library and UCC researchers have made this item openly available . Please let us know how this has helped you . Thanks ! Downloaded on 2019-09-30T08 : 53 : 25Z Possibilities for developing texture-modified beef steaks suitable for older consumers usin', *Journal of Texture Studies*.
- Martins, B. C., Rescolino, R., Coelho, D. F., Zanchetta, B., Tambourgi, E. B. and Silveira, E. (2014) 'Characterization of bromelain from ananas comosus agroindustrial residues purified by ethanol factional precipitation', *Chemical Engineering Transactions*, 37, pp. 781–786.

- Mirabella, N., Castellani, V. and Sala, S. (2014) 'Current options for the valorization of food manufacturing waste: a review', *Journal of Cleaner Production*. Elsevier Ltd, 65, pp. 28–41.
- Misran, E., Idris, A., Hajar, S., Sarip, M. and Ya, H. (2019) 'Biocatalysis and Agricultural Biotechnology Properties of bromelain extract from di ff erent parts of the pineapple variety Morris', *Biocatalysis and Agricultural Biotechnology*, 18.
- Mohan, R. and Sivakumar, V. (2016) 'Optimisation of Bromelain Enzyme Extraction from Pineapple (Ananas comosus) and Application in Process Industry', *American Journal of Biochemistry and Biotechnology*, 12(3), pp. 188–195.
- Muhammad, Z. A. and Ahmad, T. (2017) 'Therapeutic uses of pineapple-extracted bromelain in surgical care — a review', *Journal of the Pakistan Medical* Association, 67(1), pp. 121–125.
- Münchow, E. A., Hamann, H. J., Carvajal, M. T., Pinal, R. and Bottino, M. C. (2016) 'Stain removal effect of novel papain- and bromelain-containing gels applied to enamel', *Clinical Oral Investigations*. Clinical Oral Investigations, 20, pp. 2315–2320.
- Napper, A. D., Bennett, S. P., Borowski, M., Holdridge, M. B., Leonard, M. J. C., Rogers, E. E., Duan, Y., Laursen, R. A., Reinhold, B., Shames, S. L., Corporation, G. and Square, O. K. (1994) 'Purification and characterization of multiple forms of the pineapple-stem-derived cysteine proteinases ananain and comosain', *Biochemical Journal*, 301(3), pp. 727–735.
- Nasruddin, A., Amid, A., Sulaiman, S. and Othman, M. E. F. (2018) 'Biosynthesis of Nanoparticles Using Mushrooms', *Biological and Natural Resources Engineering Journal*, 1(1), pp. 351–360.
- Neh, A. and Ali, N. E. H. (2020) 'Agricultural Waste Management System [AWMS] in Malaysian', Open Access Journal of Waste Management & Xenobiotics, 3(2), pp. 1–2.
- Ngoc, U. N. and Schnitzer, H. (2009) 'Sustainable solutions for solid waste management in Southeast Asian countries', *Waste Management*. Elsevier Ltd, 29(6), pp. 1982–1995.
- Nguyen, Tham, and Cong Ha Nguyen. 2020. "Determination of Factors Affecting the Protease Content Generated in Fermented Soybean by Bacillus Subtilis 1423." *Energy Reports* 6. Elsevier Ltd: 831–36.

- Nielson, C. B., Duethman, N., Howard, J. M., Moncure, M. and Wood, J. G. (2013)
 'Burns: Pathophysiology of Systemic Complications and Current Management', *Journal of Burn Care & Research*, pp. 1–13.
- Ninpetch, U., Tsukada, M. and Promboon, A. (2015) 'Mechanical Properties of Silk Fabric Degummed with Bromelain', 10(3).
- Nor, A., Ramli, M., Hasmaliana, N., Manas, A., Azzar, A., Hamid, A., Hamid, H. A. and Illias, R. (2018) 'Comparative structural analysis of fruit and stem bromelain from Ananas comosus', *Food Chemistry*.
- Novaes, L. C. de L., Jozala, A. F., Lopes, A. M., de Carvalho Santos-Ebinuma, V., Mazzola, P. G. and Pessoa Junior, A. (2016) 'Stability, purification, and applications of bromelain: A review', *Biotechnology Progress*, 32(1), pp. 5– 13.
- Nu`n^ez, O., Moyano, E. and Galceran, M. T. (2005) 'LC MS / MS analysis of organic toxics in food', *Trends in Analytical Chemistry*, 24(7), pp. 683–703.
- Nurul, A. I. and Azura, A. (2012) 'Differential scanning calorimetry as tool in observing thermal and storage stability of recombinant bromelain', *International Food Research Journal*, 19(2), pp. 727–731.
- Omotoyinbo, O. V and Sanni, D. M. (2017) 'Characterization of Bromelain from Parts of Three Different Pineapple Varieties in Nigeria', American Journal of Bioscience, 5(3), pp. 35–41.
- Osman, Y. (2017) 'Insights into the clinical effectiveness of whitening products . Part 2 Dentist-supervised-at-home LED gel bleaching product', pp. 30–31.
- Padmapriya, M. and Williams, B. C. (2012) 'Purification and characterization of neutral protease enzyme from Bacillus Purification and characterization of neutral protease enzyme from Bacillus Subtilis', *Journal of Microbiology and Biotechnology Research*, (June 2012), pp. 612–618.
- Palao, R., Aguilera-Sáez, J., Serracanta, J., Collado, J. M. and Bruce Patrik Dos Santos,J. P. B. (2017) 'World Journal of', 6190(2).
- Pandee, Patcharaporn, Aran H-Kittikul, Ohsugi Masahiro, and Yaowaluk Dissara.
 2008. "Production and Properties of a Fibrinolytic Enzyme by Schizophyllum Commune BL23." *Songklanakarin Journal of Science and Technology* 30 (4): 447–53.

- Patil, P., Ankola, A., Hebbal, M. and Patil, A. (2014) 'Comparison of effectiveness of abrasive and enzymatic action of whitening toothpastes in removal of extrinsic stains – a clinical trial', *International Journal of Dental Hygiene*, (2), pp. 3–7.
- Pk, C. and Acharya, S. (2012) 'Efficacy of Extrinsic Stain Removal by Novel Dentifrice Containing Papain and Bromelain Extracts', *Journal of Young Pharmacists*, 4(4), pp. 245–249.
- Praveen, N. C., Rajesh, A., Madan, M., Rampratap Chaurasia, V., Hiremath, N. V and Sharma, A. M. (2014) 'In vitro Evaluation of Antibacterial Efficacy of Pineapple Extract (Bromelain) on Periodontal Pathogens', *Journal of International Oral Health*, 6(March), pp. 96–98.
- Prawan, K., Dalei2, J. and Kumari, K. S. (2016) 'Extraction, Purification and Kinetic Studies of Bromelain Enzyme From Ananas Comosus', *International Journal* of Plant, Animal and Environmental Sciences, 6(4), pp. 152–160.
- Ramalingam, C., Srinath, R. and Islam, N. N. (2012) 'Isolation and characterization of Bromelain from pineapple (Ananas Comosus) and comparing its antibrowning activity on apple juice with commercial anti- browning agents', *Elixir Food Science*, 45, pp. 7822–7826.
- Ramli, A. N. M., Aznan, T. N. T. and Illias, R. M. (2017) 'Bromelain: from production to commercialisation', *Journal of the Science of Food and Agriculture*, 97(5), pp. 1386–1395.
- Rathnavelu, V., Alitheen, N. B., Sohila, S., Kanagesan, S. and Ramesh, R. (2016)
 'Potential role of bromelain in clinical and therapeutic applications (Review)', *Medical reports*, 5, pp. 283–288.
- Raveendran, S., Kuruvilla, A. and Rebello, S. (2018) 'Applications of Microbial Enzymes in Food Industry', *Food Technology & Biotechnology*, 56(1), pp. 16– 30.
- Rose, L. F. and Chan, R. K. (2016) 'The Burn Wound Microenvironment', *Wound Healing Society*, 5(3), pp. 106–118.
- Rosenberg, L., Krieger, Y., Bogdanov-berezovski, A., Silberstein, E., Shoham, Y. and Singer, A. J. (2013) 'ScienceDirect A novel rapid and selective enzymatic debridement agent for burn wound management : A multi-center RCT', *Burns*. Elsevier Ltd and International Society of Burns Injuries, 40(3), pp. 466–474.

- Sadh, P. K., Duhan, S. and Duhan, J. S. (2018) 'Agro industrial wastes and their utilization using solid state fermentation: a review', *Bioresources and Bioprocessing*. Springer Berlin Heidelberg, pp. 1–15.
- Sahoo, R. and Das, P. (2017) 'Bromelain: Applications and Purification Strategies', *PharmaTutor*, 5(11), pp. 40–48.
- Sangkharak, K., Wangsirikul, P., Pichid, N., Yunu, T. and Prasertsan, P. (2016) 'Partitioning of Bromelain from Pineapple Stem (Smooth Cayenne) by Aqueous Two Phase System and Its Application for Recovery and Purification of Polyhydroxyalkanoate', *Chiang Mai J. Sci*, 43(4), pp. 794–807.
- Segovia, F. and Pablos, P. A. (2016) 'Pineapple Waste Extract for Preventing Oxidation in Model Food Systems', 81(7), pp. 1622–1628.
- Soares, P. A. G., Vaz, A. F. M., Correia, M. T. S., Pessoa, A. and Carneiro-Da-Cunha, M. G. (2012) 'Purification of bromelain from pineapple wastes by ethanol precipitation', *Separation and Purification Technology*. Elsevier B.V., 98, pp. 389–395.
- Sunar, K., Kumar, U. and Deshmukh, S. K. (2016) Recent Applications of Enzymes in Personal Care Products, Agro-Industrial Wastes as Feedstock for Enzyme Production. Elsevier Inc.
- Syahrin, S. (2011) 'Consumer preferences towards pineapple cultivars in Malaysia', *Acta Horticulturae*, 902, pp. 595–600.
- Tap, F. M., Majid, F. A. A. and Khairudin, N. B. A. (2016) 'Structure prediction of stem bromelain from pineapples (Ananas comosus) using procaricain enzyme as a modelling template', *International Journal of Applied Engineering Research*, 11(9), pp. 6109–6111.
- Thalip, A. A., Tong, P. S. and Ng, C. (2015) 'The MD2 "Super Sweet" pineapple (Ananas comosus)', *Utar Agriculture Science Journal*, 1(4), pp. 14–17.
- Thu, Nguyen T A, Nguyen T M Khue, Nguyen D Huy, Nguyen Q D Tien, and Nguyen H Loc. 2020. "Characterizations and Fibrinolytic Activity of Serine Protease from Bacillus Subtilis C10." *Current Pharmaceutical Biotechnology*. Netherlands, 21(2), pp. 110-116.
- Urdiales-ga, F., Mira, V. L. M., Moreno, A., Orti, F. and Romero-a, R. R. N. (2017) 'Treatment of Soft Tissue Filler Complications: Expert Consensus Recommendations', *Aesth Path Surg*.

- Vicente, F. A., Lario, L. D., Pessoa, A. and Ventura, S. P. M. (2016) 'Recovery of bromelain from pineapple stem residues using aqueous micellar two-phase systems with ionic liquids as co-surfactants', *Process Biochemistry*. Elsevier Ltd, 51(4), pp. 528–534.
- Vijayaraghavan, P., Gnana, S. and Vincent, P. (2013) 'A simple method for the detection of protease activity on agar plates using bromocresolgreen dye', *Journal of Biochemical*, 4, pp. 628–630.
- Vilanova Neta, J. L., Da Silva Lédo, A., Lima, A. A. B., Santana, J. C. C., Leite, N. S., Ruzene, D. S., Silva, D. P. and De Souza, R. R. (2012) 'Bromelain enzyme from pineapple: In vitro activity study under different micropropagation conditions', *Applied Biochemistry and Biotechnology*, 168(2), pp. 234–246.
- Wali, N. (2018) Pineapple (Ananas comosus), Nonvitamin and Nonmineral Nutritional Supplements. Elsevier Inc.
- Wan, J., Guo, J., Miao, Z. and Guo, X. (2016) 'Reverse micellar extraction of bromelain from pineapple peel – Effect of surfactant structure', *Food Chemistry*. Elsevier Ltd, 197, pp. 450–456.
- Wu, W. C., Ng, H. S., Sun, I. M. and Lan, J. C. W. (2017) 'Single step purification of bromelain from Ananas comosus pulp using a polymer/salt aqueous biphasic system', *Journal of the Taiwan Institute of Chemical Engineers*. Elsevier B.V., 79, pp. 158–162.
- Youryon, P., Supapvanich, S., Kongtrakool, P. and Wongs-Aree, C. (2018) 'Calcium chloride and calcium gluconate peduncle infiltrations alleviate the internal browning of Queen pineapple in refrigerated storage', *Horticulture Environment and Biotechnology*, 59(2), pp. 205–213.