

SORPTION OF HEAVY METALS ONTO POLYETHYLENE MICROBEADS  
AND ITS EFFECT ON SEABASS

NUR FARHAN BINTI ZON

UNIVERSITI TEKNOLOGI MALAYSIA

SORPTION OF HEAVY METALS ONTO POLYETHYLENE MICROBEADS  
AND ITS EFFECT ON SEABASS

NUR FARHAN BINTI ZON

A thesis submitted in fulfilment of the  
requirements for the award of the degree of  
Doctor of Philosophy

School of Civil Engineering  
Faculty of Engineering  
Universiti Teknologi Malaysia

JUNE 2021

## **DEDICATION**

Dedicated to my beloved little family, my parents, and my supportive supervisor –  
DR. SHAMILA BINTI AZMAN. Thank you very much for being positive, helpful,  
and understanding

## **ACKNOWLEDGEMENT**

Praise Be to Allah S.W.T, the Lord of the World

Foremost, I would like to express my sincere gratitude to my supervisor Dr. Shamila Azman for the continuous support of my study and research, for her patience, motivation, enthusiasm, and immense knowledge. Her guidance helped me in all the time of research and writing of this thesis. I could not have imagined having a better advisor and mentor for my study.

I also would like to convey an appreciation to Jabatan Perkhidmatan Awam Malaysia for funding my study under Program Pelajar Cemerlang 2017.

My sincere thanks also go to all staff, lectures, and individuals who directly or indirectly support me throughout completing this study. I am deeply thankful to my friends and officemates who always support and motivate me during ups and down.

Not least of all, special thanks to my supportive husband, Muhamad Azam, my cherished daughter, Humaira and my dearest mother, Rohani Rashid for their pray and always by my side through this journey.

## ABSTRACT

Microbeads are one of the causes of microplastic pollution that is currently polluting ocean environment. It enters food chain via ingestion of marine vertebrates and invertebrates. This study aims to elucidate the interactions between polyethylene microbeads and heavy metals as well as to determine the possibility of polyethylene microbeads as heavy metal vectors for juvenile seabass. Two parts of experiments performed in this study, i.e. adsorption and ingestion. For the adsorption study, 10 g of virgin polyethylene microbeads (300  $\mu\text{m}$ ) were submerged into 0.2, 0.4, 0.6, and 1.0  $\mu\text{g/mL}$  solutions of cadmium and chromium, and 0.4, 0.8, 1.2, 1.6, and 2.0  $\mu\text{g/mL}$  concentrations of lead in a batch of sorption experiments for 96 hours. In the ingestion experiment, seabass was exposed in control, single, co-exposure, and preloaded experiments. All exposure conditions were similar to the adsorption experiment with 10 g and 5 g of microbeads. Acid digestion and Atomic Absorption Spectroscopy were used to quantify the amount of heavy metal adsorbed on microbeads and accumulated in fish tissues. Maximum adsorption capacity of microbeads were 11  $\mu\text{g/g}$  for Cd, 1.7  $\mu\text{g/g}$  for Cr, and 9.0  $\mu\text{g/g}$  for Pb. The kinetic study concluded that the adsorption of polyethylene microbeads occurred at a pseudo-first-order reaction, which involves physical attraction. Adsorption isotherm fitted the Freundlich model signifying adsorption occurs rapidly and has the tendency to desorb due to weak binding. The rates of heavy metal adsorption onto microbeads were 11, 4.5, and 1.7  $\text{mL/g}$  for Cd, Pb, and Cr, respectively, suggesting that Cd had a higher affinity towards microbeads polyethylene than Pb and Cr. In the ingestion study, most of the heavy metal were detected at the skin layer. Control experiment validated that, seabass uptake exposed microbeads via ingestion. The single experiment concluded that uptake of heavy metals in seabass tissues increased with concentration and time. The higher the amount of exposed microbeads, the higher the uptake of heavy metals in the gastrointestinal tract after 48 hours of co-exposure. This indicates that heavy metals were first adsorbed on the microbeads followed by their ingestion by the seabass. In the preloaded experiment, the concentration of heavy metal ions detected in the gastrointestinal tract was higher than the direct exposure in single and co-exposure. The uptake values in the preloaded experiment increased steadily with concentration, time, and quantity of microbeads. Preloaded exposure in this study verified that microbeads-loaded heavy metals were incidentally ingested by seabass during foraging. The incorporation of the three heavy metals in the ingestion study was performed using Minitab 16.0 multi analysis of variance (MANOVA). This study proved that polyethylene microbeads possess the potential to accumulate, transport, and transfer heavy metals from water to intestinal organ, thus increasing risk, threatening the marine food web, and possibly harming other consumers.

## ABSTRAK

Manik mikro adalah salah satu sumber pencemaran mikroplastik, mencemarkan persekitaran lautan. Ia memasuki rantai makanan melalui pengambilan makanan vertebrata dan invertebrata. Kajian ini bertujuan untuk memperjelaskan interaksi antara manik mikro polietilena dan logam berat untuk menentukan kemungkinan manik mikro polietilena sebagai vektor logam berat kepada ikan siakap. Dua bahagian eksperimen dalam kajian ini, iaitu penjerapan dan pengambilan makanan. Untuk kajian penjerapan, 10 g manik mikro polietilena tulin (300  $\mu\text{m}$ ) direndam dalam larutan 0.2, 0.4, 0.6 dan 1.0  $\mu\text{g/mL}$  untuk kadmium dan kromium, manakala kepekatan plumbum adalah 0.4, 0.8, 1.2, 1.6 dan 2.0  $\mu\text{g/mL}$  dalam kumpulan eksperimen jerapan selama 96 jam. Dalam eksperimen pengambilan makanan, ikan siakap didedahkan dalam eksperimen kawalan, tunggal, pendedahan bersama dan pramuat. Semua keadaan adalah sama dengan eksperimen penjerapan dengan jumlah manik mikro yang digunakan adalah 10 g dan 5 g. Pencernaan asid dan Spektrofotometer Serapan Atom digunakan untuk mengukur jumlah pengambilan logam berat dalam manik mikro dan lapisan tisu ikan. Kapasiti penjerapan maksimum manik mikro adalah 11  $\mu\text{g/g}$  untuk Cd, 1,7  $\mu\text{g/g}$  Cr dan 9,0  $\mu\text{g/g}$  Pb. Kajian kinetik menyimpulkan bahawa penjerapan manik mikro polietilena berlaku mengikut pseudo-tertib-pertama, yang melibatkan tarikan fizikal. Model isoterm Freundlich yang menunjukkan bahawa penjerapan berlaku dengan cepat serta mempunyai ikatan yang lemah. Kadar penjerapan logam berat pada manik mikro adalah 11, 4.5 dan 1.7  $\text{mL/g}$  untuk Cd, Pb dan Cr, masing-masing menunjukkan bahawa Cd mempunyai tarikan yang lebih tinggi terhadap manik mikro polietilena berbanding Pb dan Cr. Dalam kajian pengambilan makanan, kebanyakan ion logam berat dikesan pada lapisan kulit ikan. Eksperimen kawalan menyimpulkan bahawa ikan siakap memakan manik mikro. Eksperimen tunggal menyimpulkan pengambilan logam berat dalam tisu ikan siakap meningkat dengan kepekatan dan masa. Semakin tinggi manik mikro yang terdedah, semakin tinggi pengambilan logam berat di saluran usus setelah 48 jam dalam eksperimen pendedahan bersama logam berat dan manik mikro. Ini menunjukkan bahawa manik mikro menyerap logam berat dari persekitaran, kemudian dicerna oleh ikan siakap. Dalam eksperimen pramuat, kepekatan ion logam berat yang dikesan di saluran usus lebih tinggi daripada pendedahan langsung dalam kawalan dan pendedahan bersama. Nilai pengambilan dalam eksperimen pramuat meningkat dengan stabil dengan kepekatan, masa dan jumlah manik mikro dengan jelas. Pendedahan yang dimuatkan dalam kajian ini mengesahkan bahawa, logam berat yang diserap oleh manik mikro secara tidak sengaja ditelan oleh ikan siakap semasa mencari makanan. Ketiga-tiga logam berat dalam kajian penjerapan dan pengambilan makanan dilakukan menggunakan Minitab 16.0 dalam analisis pelbagai varians (MANOVA). Kajian ini telah membuktikan bahawa manik mikro polietilena berpotensi untuk mengumpulkan, mengangkut, menjadi vektor logam berat di persekitaran laut ke organ pengambilan makanan, sehingga meningkatkan risiko dan mengancam jaringan makanan laut, dan mungkin berbahaya bagi pengguna lain.

## TABLE OF CONTENTS

	TITLE	PAGE
	<b>DECLARATION</b>	<b>iii</b>
	<b>DEDICATION</b>	<b>iv</b>
	<b>ACKNOWLEDGEMENT</b>	<b>v</b>
	<b>ABSTRACT</b>	<b>vi</b>
	<b>ABSTRAK</b>	<b>vii</b>
	<b>TABLE OF CONTENTS</b>	<b>viii</b>
	<b>LIST OF TABLES</b>	<b>xiii</b>
	<b>LIST OF FIGURES</b>	<b>xv</b>
	<b>LIST OF ABBREVIATIONS</b>	<b>xx</b>
	<b>LIST OF SYMBOLS</b>	<b>xxiii</b>
	<b>LIST OF APPENDICES</b>	<b>xxiv</b>
<b>CHAPTER 1</b>	<b>INTRODUCTION</b>	<b>1</b>
1.1	Introduction	1
1.2	Research Background	4
1.3	Problem Statement	9
1.4	Objectives	10
1.5	Scopes	10
1.6	Significance of the Study	12
<b>CHAPTER 2</b>	<b>LITERATURE REVIEW</b>	<b>13</b>
2.1	Introduction	13
2.2	Microbeads	13
2.3	Microplastics and Co-Contaminants	16
2.3.1	Microplastic Surface Characteristics: Diffusivity and Polarity	19
2.3.2	Inherent Contaminants	21
2.3.3	Exposure Condition	22

2.3.4	Summary of the Adsorption Ability of Microplastics	32
2.4	Microplastic Ingestion by Marine Organisms	34
2.5	Ingestion Factors of Microplastics: Organisms	37
2.5.1	Feeding Strategies and Foraging Behaviour	39
2.5.2	Habitats and Trophic Guilds/Trophic Levels	42
2.5.3	Growth Stages of Organisms	45
2.5.4	Summary of the Factors of Microplastic Ingestion in Organisms	50
2.6	Health Risks involving Microplastic Ingestion by Marine Biota	51
2.7	Trophic Transfer	53
2.8	Heavy Metal Pollution in the Marine Environment	55
2.9	Vector Effect Study in the Environment of Marine Organisms	57
2.9.1	Health Risks Involving Vector Effects	66
2.10	Summary of the Ingestion Effects of Microplastics and Contaminants on Organisms	67
2.11	Seabass or <i>Lates calcarifer</i>	69
2.11.1	Biological and Morphological Anatomies of Seabass	70
2.11.2	Life History	72
2.11.3	Behaviour of Seabass	72
2.11.4	Feeding Habits and Ecosystem Role	73
2.11.5	Seabass in Malaysia	74
2.11.6	Seabass as Bioindicator for Heavy Metal Pollution	75
2.12	Summary	78
<b>CHAPTER 3</b>	<b>RESEARCH METHODOLOGY</b>	<b>79</b>
3.1	Introduction	79
3.2	Sample Collections and Chemical Preparations	81
3.2.1	Reagents and Apparatus	81
3.2.2	In-situ Analysis	81
3.2.3	Artificial Seawater for Acclimatization	81



3.2.4	Sampling of Seabass	83
3.2.5	Acclimatization and Depuration of Seabass	84
3.2.6	Preparation of Heavy Metals-Preloaded Microbeads	85
3.3	Experimental Procedure	87
3.3.1	Adsorption of Heavy Metals onto Polyethylene Microbeads	88
3.3.2	Ingestion Experiment	89
3.4	Sample Analysis Procedure	92
3.4.1	Dissection of Gastrointestinal Tracts, Gills, and Skin of Seabass	92
3.4.2	Separation of Ingested Microbeads in Control Experiment	94
3.4.3	Microbeads Identification under Microscope	95
3.4.4	Microplastic Identification and Validation using ATR-FTIR	96
3.5	Heavy Metals Analysis	97
3.5.1	Digestion of Microbeads	97
3.5.2	Digestion of Tissue Samples	98
3.5.3	Digestion of Seawater	99
3.5.4	Atomic Adsorption Spectrophotometer (AAS)	99
3.6	Visual Observation	100
3.7	Data Analysis	100
3.7.1	Calculation of Heavy Metal Uptake	101
3.7.2	Kinetic and Isotherm Analyses of Microbeads Sorptive Study	101
3.7.3	Mathematical Approach for Ingestion Study	103
<b>CHAPTER 4</b>	<b>RESULTS AND DISCUSSION</b>	<b>105</b>
4.1	Introduction	105
4.2	Adsorption Study of Heavy Metals onto Low-Density Polyethylene Microbeads	106
4.2.1	Adsorption of Heavy Metals onto Polyethylene Microbeads	107
4.2.2	Kinetic Study of Heavy Metals onto Polyethylene Microbeads	117

4.2.3	Isotherm Study of Heavy Metals onto Polyethylene Microbeads	121
4.2.4	Adsorption Study of Cadmium, Chromium, and Lead	125
4.3	Ingestion Study	128
4.3.1	Preloaded Polyethylene Microbeads to Cadmium, Chromium, and Lead	130
4.3.2	Ingested Microbeads in Control Experiment	131
4.3.3	Characterization of the Microplastic Beads	134
4.3.4	Evaluating the Total Uptake of Cadmium, Chromium, and Lead by Seabass	135
4.3.5	Ingestion of Cadmium and Microbeads by Seabass	142
4.3.5.1	Effect of Time and Concentration in Single Experiments	142
4.3.5.2	Effect of Amount of Microbeads in Co-exposure and Preloaded Experiments	145
4.3.5.3	Effect of Time, Concentrations, and Microbeads in Co-exposure and Preloaded Experiments	148
4.3.5.4	Comparison of the Uptake of Tissues in Co-exposure and Preloaded Experiments	151
4.3.6	Ingestion of Chromium and Microbeads by Seabass	155
4.3.6.1	Effect of Time and Concentration in Single Experiments	155
4.3.6.2	Effect of Amount of Microbeads in Co-exposure and Preloaded Experiments	157
4.3.6.3	Effect of Time, Concentration, and Microbeads in Co-exposure and Preloaded Experiments	161
4.3.6.4	Comparison of the Uptake of Tissues in Co-exposure and Preloaded Experiments	164
4.3.7	Ingestion of Lead and Microbeads by Seabass	168

4.3.7.1	Effect of Time and Concentration in Single Experiments	168
4.3.7.2	Effect of Amount of Microbeads in Co-exposure and Preloaded Experiments	170
4.3.7.3	Effect of Time, Concentrations, and Microbeads in Co-exposure and Preloaded Experiments	173
4.3.7.4	Comparison of the Uptake of Tissues in Co-exposure and Preloaded Experiments	177
4.3.8	Summary of Heavy Metal Ingestion Study	180
4.4	Visual Observation on the Behaviour of Juvenile Seabass	182
4.5	Risk of Heavy Metals Adsorbed onto Microbeads with Uptake by Juvenile Seabass	184
4.6	Summary of Ingestion Effect of Microbeads-Heavy Metals into Seabass	188
<b>CHAPTER 5</b>	<b>CONCLUSION AND RECOMMENDATIONS</b>	<b>193</b>
5.1	Conclusion	193
5.2	Recommendations	195
<b>REFERENCES</b>		<b>199</b>
<b>APPENDIX</b>		<b>239</b>
<b>LIST OF PUBLICATIONS</b>		<b>250</b>

## LIST OF TABLES

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
Table 2.1	Adsorption of heavy metals onto microplastics	26
Table 2.2	Microplastic ingestion found in organisms	35
Table 2.3	Feeding strategies and foraging behaviour of organisms on microplastics	40
Table 2.4	Studies on habitat/trophic levels of microplastic ingestion in marine and laboratory	44
Table 2.5	Studies on early stages of fish ingestion	49
Table 2.6	Studies on microplastics as vectors in marine organisms	61
Table 2.7	Taxonomy of seabass (adapted from Mathew, 2009)	70
Table 2.8	List of previous studies on seabass as a bioindicator for heavy metal pollution	76
Table 3.1	Water condition in the acclimation tank	82
Table 3.2	Concentration used for preloaded microbeads preparation according to the ingestion exposure treatment	87
Table 3.3	Permissible limit of respective heavy metals based on Malaysia Food Regulation (1985) for fish	87
Table 3.4	Conditions of ingestion treatments	89
Table 4.1	Water condition in the experimental tank	106
Table 4.2	Adsorption capacity of cadmium ( $\mu\text{g/g}$ Cd w/w microbeads)	108
Table 4.3	Adsorption capacity of chromium ( $\mu\text{g/g}$ Cr w/w microbeads)	111
Table 4.4	Adsorption capacity of lead ( $\mu\text{g/g}$ Pb w/w microbeads)	114
Table 4.5	Parameters obtained from the pseudo-first-order for adsorption of microbeads onto cadmium	118
Table 4.6	Parameters obtained from the pseudo-first-order for adsorption of microbeads onto chromium	118
Table 4.7	Parameters obtained from the pseudo-first-order for adsorption of microbeads onto lead	118
Table 4.8	Rates of heavy metals onto polyethylene microbeads	120

Table 4.9	Isotherm Models' Constant and Correlation Coefficients for the Adsorption of Heavy Metals in Aqueous Solution	124
Table 4.10	Concentrations of microbeads for preloaded treatments ( $\mu\text{g/g}$ )	131
Table 4.11	Absorption band and functional groups of polyethylene	134

## LIST OF FIGURES

<b>FIGURE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
Figure 1.1	The increase in plastic production around the world from 1950-2017 (Plastics Europe, 2018)	1
Figure 1.2	Potential pathway for the transport of microplastics & their biological interactions (adapted from Wright et al., 2013)	5
Figure 2.1	Timeline for prohibitions of microbeads around the globe (adapted from Nelson et al., 2019)	15
Figure 2.2	Factors of adsorption between microplastics and contaminants	18
Figure 2.3	Factors of microplastic ingestion by marine organisms.	38
Figure 2.4	Heavy metals sorbed to ingest microbeads that are accumulated in fish from personal care products (adapted from Wardrop et al., 2016)	58
Figure 2.5	Seabass (adapted from Food and Agriculture Organization of the United States, 2006).	71
Figure 3.1	Research Methodology Flowchart	80
Figure 3.2	Acclimatization tank preparation before sampling	82
Figure 3.3	Aquaculture Fisheries Research Institute Malaysia, Gelang Patah	83
Figure 3.4	Juvenile seabass size sorting process	84
Figure 3.5	Seabass acclimatization process in the laboratory	85
Figure 3.6	Preloaded microbeads drying process at room temperature	86
Figure 3.7	Illustration of sorptive experimental tank set-up	88
Figure 3.8	Ingestion experiment tank	90
Figure 3.9	Ingestion of heavy metals-preloaded microbeads by seabass.	91
Figure 3.10	Seabass sampled every 24 hours	92
Figure 3.11	Seabass tissue analysis preparation	93
Figure 3.12	Parts of seabass used in this study	93
Figure 3.13	Seabass dissection process	94

Figure 3.14	Seabass gastrointestinal tract separation	94
Figure 3.15	Excised gastrointestinal tracts digestion process	95
Figure 3.16	Microscope (Olympus BX53M) used to visualize ingested microbeads in gastrointestinal tracts	96
Figure 3.17	ATR-FTIR equipped with analytical software to verify microbeads polymer	97
Figure 3.18	Gastrointestinal tracts were pooled and oven-dried to remove moisture content before the acid digestion process	98
Figure 3.19	Atomic Absorption Spectrophotometer (AAS) used in the analysis of heavy metals in digested samples	100
Figure 4.1	Time-dependent of cadmium adsorption	107
Figure 4.2	Data distribution analysis of cadmium for initial concentration	109
Figure 4.3	Data distribution analysis of cadmium for time of exposure	110
Figure 4.4	Time-dependent of chromium adsorption	110
Figure 4.5	Data distribution analysis of chromium for initial concentration	112
Figure 4.6	Data distribution analysis of chromium with respect to time of exposure	112
Figure 4.7	Time-dependent of lead adsorption	113
Figure 4.8	Data distribution analysis of lead for initial concentration	115
Figure 4.9	Data distribution analysis of lead with respect to time of exposure	115
Figure 4.10	Validity of adsorption isotherm of cadmium	122
Figure 4.11	Validity of adsorption isotherm of chromium	122
Figure 4.12	Validity of adsorption isotherm of lead	123
Figure 4.13	Preloaded concentrations of cadmium and chromium for 96 hours onto microbeads with 0.2 to 1.0 mg/L	130
Figure 4.14	Preloaded concentrations of lead for 96 hours onto microbeads with 0.4 to 2.0 mg/L	130
Figure 4.15	Microbeads ingested at 10 g for 96 hours in gastrointestinal tracts	132
Figure 4.16	Microbeads ingested at 5 g for 96 hours in gastrointestinal tracts	133

Figure 4.17	Infrared spectra for ethylene, the monomer for polyethylene	135
Figure 4.18	Total tissue uptake of cadmium with 10 g microbeads exposure for (a) co-exposure and (b) preloaded and with 5 g microbeads exposure for (c) co-exposure and (d) preloaded	137
Figure 4.19	Total tissue uptake of chromium with 10 g microbeads exposure for (a) co-exposure and (b) preloaded and with 5 g microbeads exposure for (c) co-exposure and (d) preloaded	138
Figure 4.20	Total tissue uptake of lead with 10 g microbeads exposure for (a) co-exposure and (b) preloaded and with 5 g microbeads exposure for (c) co-exposure and (d) preloaded	141
Figure 4.21	Adsorption capacity of cadmium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with no microbeads	144
Figure 4.22	Adsorption capacity of cadmium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (co-exposure experiments)	146
Figure 4.23	Adsorption capacity of cadmium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (co-exposure experiments)	147
Figure 4.24	Adsorption capacity of cadmium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (preloaded experiments)	149
Figure 4.25	Adsorption capacity of cadmium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (preloaded experiments)	150
Figure 4.26	Adsorption capacity of cadmium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (co-exposure experiments versus preloaded)	153
Figure 4.27	Adsorption capacity of cadmium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (co-exposure experiments versus preloaded)	154



Figure 4.28	Adsorption capacity of chromium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with no microbeads	156
Figure 4.29	Adsorption capacity of chromium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (co-exposure experiments)	159
Figure 4.30	Adsorption capacity of chromium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (co-exposure experiments)	160
Figure 4.31	Adsorption capacity of chromium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (preloaded experiments)	162
Figure 4.32	Adsorption capacity of chromium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (preloaded experiments)	163
Figure 4.33	Adsorption capacity of chromium into gastrointestinal tracts, gills, and skin at (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (co-exposure experiments versus preloaded)	166
Figure 4.34	Adsorption capacity of chromium into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (co-exposure experiments versus preloaded)	167
Figure 4.35	Adsorption capacity of lead into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with no microbeads	169
Figure 4.36	Adsorption capacity of lead into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (co-exposure experiments)	171
Figure 4.37	Adsorption capacity of lead into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (co-exposure experiments)	172
Figure 4.38	Adsorption capacity of lead into gastrointestinal tracts, gills, and skin at (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (preloaded experiments)	175

Figure 4.39	Adsorption capacity of lead into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (preloaded experiments)	176
Figure 4.40	Adsorption capacity of lead into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 10 g microbeads (co-exposure experiments versus preloaded)	178
Figure 4.41	Adsorption capacity of lead into gastrointestinal tracts, gills, and skin for (a) 24 hours, (b) 48 hours, (c) 72 hours, and (d) 96 hours with 5 g microbeads (co-exposure experiments versus preloaded)	179
Figure 4.42	Seabass showing signs of stress in preloaded treatments after 48 hours	183
Figure 4.43	Illustration of possible ingestion effect of microbeads-heavy metals on seabass	190

## LIST OF ABBREVIATIONS

AAS		Atomic Absorption Spectrophotometer
Ag	-	Silver
Al	-	Aluminium
As	-	Arsenic
ASW	-	Artificial seawater
Ba	-	Barium
Br	-	Bromine
Cd	-	Cadmium
Cl	-	Chlorine
Co	-	Cobalt
Cr	-	Chromium
Cu	-	Copper
DDT	-	Dichlorodiphenyltrichloroethane
DEHP	-	Di(2-ethylhexyl) Phthalate
DNA	-	Deoxyribonucleic Acid
DO	-	Dissolved Oxygen
EPA	-	Environmental Protection Agency
FAO	-	Food and Agriculture Organization of the United Nations
Fe	-	Iron
GST	-	Glutathione S-Transferase
HDPE	-	High Density Polyethylene
Hg	-	Mercury
HNO <sub>3</sub>	-	Nitric Acid
H <sub>2</sub> O <sub>2</sub>	-	Hydrogen Peroxide
HCl	-	Hydrochloric Acid
HOC	-	Halogenated Organic Carbons
HPLC	-	High Performance Liquid Chromatography
IDH	-	Isocitrate Dehydrogenase
LDH	-	Lactate Dehydrogenase
LDPE	-	Low Density Polyethylene

MANOVA	-	Multiple Analysis of Variance
Mn	-	Manganese
Mo	-	Molybdenum
MP	-	Microplastic
Ni	-	Nickel
PAHs	-	Polycyclic Aromatic Hydrocarbons
Pb	-	Plumbum/Lead
PBDEs	-	Polybrominated Diphenyl Ethers
PBTs	-	Persistent Bioaccumulative Toxic Substances
PCB	-	Polychlorinated biphenyl
PE	-	Polyethylene
PET	-	Polyethylene Terephthalate
PFASs	-	Perfluorooctanoic Acid
PFOS	-	Perfluorooctane Sulfonate
PMMA	-	Polymethylmethacrylate
POPs	-	Persistent Organic Pollutants
PP	-	Polypropylene
PS	-	Polystyrene
PVC	-	Polyvinyl chloride
$r^2$	-	Coefficient of Determination
Sb	-	Antimony
SD	-	Standard Deviation
Se	-	Selenium
SM	-	Synthetic Musks
Sn	-	Stannum/Tin
Sr	-	Strontium
SSE	-	Sum of Squared Estimate of Errors
TC	-	Tetracycline
Ti	-	Titanium
U	-	Uranium
UK	-	United Kingdom
UPM	-	Universiti Putra Malaysia
US	-	United States

USA	-	United States of America
UV	-	Ultraviolet
UV-B	-	Ultraviolet B-Rays
V	-	Vanadium
YSI	-	Yellow Springs Instrument
Zn	-	Zinc
$\alpha$ -HCHs	-	Hexachlorocyclohexanes

## LIST OF SYMBOLS

%	-	Percent
$\mu\text{m}$	-	Micrometer
$\text{cm}^2$	-	Centimeter Square
$\text{cm}^3$	-	Centimeter Cube
$\text{m}^2$	-	Meter Square
ng	-	Nanogram
$\mu\text{g}$	-	Microgram
ppm	-	Part Per Million
ppt	-	Part Per Trillion
$^{\circ}\text{C}$	-	Degree Celsius
J	-	Joule
$E_a$	-	Activation Energy
$K_d$	-	Distribution Coefficient
$q_e$	-	Heavy Metal Adsorbed per Unit Mass at Equilibrium
$k_t$	-	Pseudo-First-Rate Constant
$t$	-	Time
$q_m$	-	Energy Constant Related to The Heat of Adsorption
$C_e$	-	Concentration of Heavy Metals at Equilibrium
$k_f$	-	Adsorption Capacity
$n$	-	Adsorption Intensity

## LIST OF APPENDICES

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
Appendix A	General Linear Model (MANOVA) for Adsorption Study	239
Appendix B	General Linear Model (MANOVA) for Ingestion Study	240
Appendix C	Polyethylene Puenscrub Specification	246
Appendix D	Microbeads ingested at 10 g and 5 g for 24 to 72 hours in gastrointestinal tracts	247

# CHAPTER 1

## INTRODUCTION

### 1.1 Introduction

Plastics are one of the industrial products that are widely used and have successfully replaced several conventional materials such as glass, metal, and wood due to their cost of production, strong, durable, and lightweight characteristics, and easy to produce (DeArmitt, 2017; Thompson et al., 2009). Plastics are constructed through the linking of hydrocarbon monomers that created synthetic polymers. As shown in Figure 1.1, plastic production is growing steadily each year due to its demand and its production had been reported to be up to 348 million tonnes in 2017 (Plastics Europe, 2018), and estimated to climb up to around 33 billion tonnes by 2050 (Rochman et al., 2013).

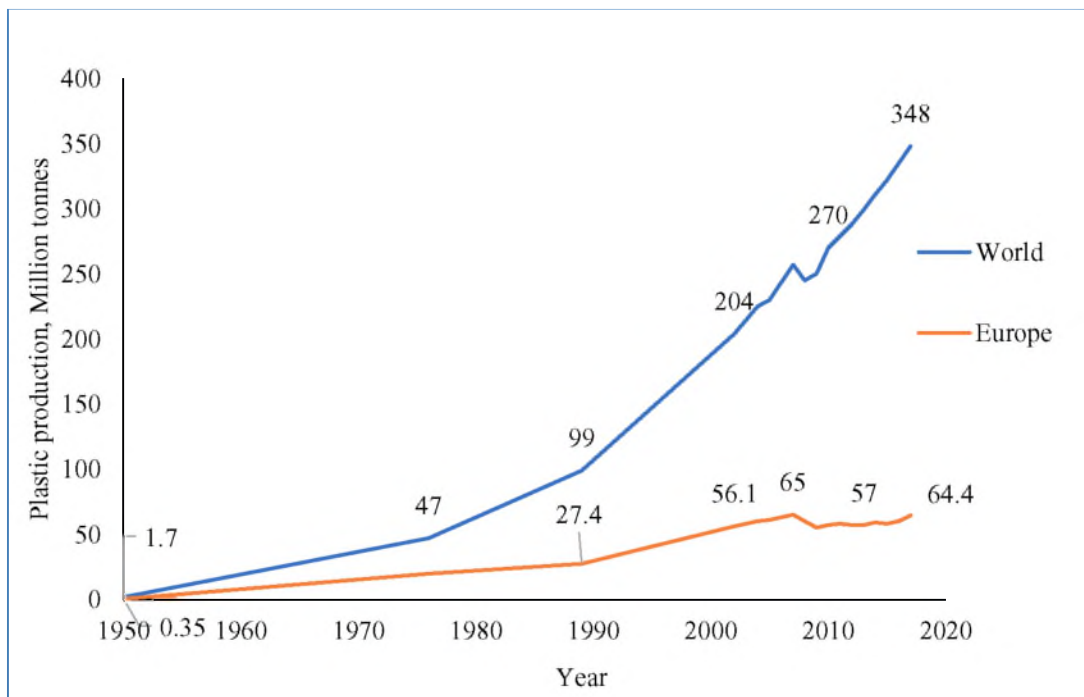


Figure 1.1 The increase in plastic production around the world from 1950-2017 (Plastics Europe, 2018)



The production of plastics grows rapidly each year due to industrial demands and their unique characteristics such as low cost, low density, and high durability in most industries, especially in the packaging industry. Being the most abundant and persistent debris found in marine (Cauwenberghe et al., 2015), plastics have a long lifespan and accumulate in the environment (Andrady, 2011; Cole et al., 2011; Galgani et al., 2013; Wright et al., 2013) despite the movement and campaign to reduce the use of plastics, which have been recognized as a threat to the marine ecosystems due to their abundance.

Based on estimations, 1.15 to 2.41 million tonnes of plastics are released into the marine environment annually (Lebreton et al., 2017). Thus, the alarming number of plastics and their persistent characteristics has led to environmental concerns (Paterson, 2019). As mentioned by previous researchers, consumer packaging is made from one-third of plastic production with 10% of municipal waste (Andrady, 2011), which mostly ends up in landfills and remains there for a long time (Barnes et al., 2009). Meanwhile, the remaining 90% are usually recycled or are not handled properly, which may end up in the environment via several routes. In general, plastics in all sizes, from meters to micrometers, are found in the environment (Barnes et al., 2009).

Most plastic polymers found in the environment are polypropylene (PP), polyethylene (PE), polystyrene (PS), polyethylene terephthalate (PET), and polyvinyl chloride (PVC) (Andrady & Neal, 2009; Andrady, 2017) due to their various applications. They can also be found in different levels of the water column due to their density. Generally, polyethylene and polypropylene are buoyant and have high mobility due to water currents and wind; hence, most of them will be permanently trapped or stranded in a location that cannot further move them with physical processes such as tidal and biofouling that can cause an increase in the density. Besides, polyethylene and polypropylene debris are abundantly found in the marine environment (Xu et al., 2020) or remote areas (Wang et al., 2018; Lusher, 2015; Nakashima et al., 2012). The denser fragment of PVC and PET is also readily settling out of suspension in the marine environment. Nonetheless, due to persistency character, all plastics that end up in the marine environment from years ago, either

transported, degraded, fragmented, fouled, or deposited, are presumed to exist until now (Thompson, 2015).

Plastics particles with a size less than 5 mm are classified as microplastics (Thompson, 2015; GESAMP, 2019), which have been considered as pollutants of high concern (Kögel et al., 2020; Kroon et al., 2020). Evidently, microplastics are widely found in marine sediments or water columns (Guo et al., 2020; Kik & Sici, 2020; Peng et al., 2020). In general, there are a few forms of microplastics detected in the environment such as fiber, pellets, beads, and fragments.

Primary microplastics are those introduced directly into the environment, mostly from proposed products, wastes from manufacturing processes, or derivatives from the erosion and tearing of large plastic products such as tires, wheels, and boards. On the other hand, plastic production uses powder or pellets as raw materials; therefore, these materials might accidentally end up in the environment during accidental release (Dris et al., 2016; Gasperi et al., 2019; Marnane et al., 2006; Vandermeersch et al., 2015), shipping, or cleaning machinery through shot blasting (Cole et al., 2011). These materials exist typically as resin pellets (Rocha-Santos & Duarte, 2015; Waller et al., 2017), microbeads (Yurtsever, 2019), microfiber (Mark Anthony Browne et al., 2011), and other forms.

The degradation of macro or mesoplastics in the environment under the physical, chemical, and biological forces with a size less than 5 mm is called secondary microplastics (Zhang et al., 2016; Thompson, 2015; Rocha-Santos & Duarte, 2015; Waller et al., 2017). These processes include heat, mechanical forces, ultraviolet (UV) light, oxidation, or biodegradation (Rillig et al., 2017). Macroplastics do not only impact the natural system but also the range of organisms in the environment through ingestions for large organisms and entanglement for smaller ones, especially birds and fish (Phuong et al., 2016; Compa et al., 2018; Provencher et al., 2018; Horn et al., 2019).

In addition, macroplastics potentially degrade and break down into smaller fragments based on their rate of degradation that is controlled by several environmental factors. As such, the carbon in the polymer can be transformed into carbon dioxide and incorporated into marine biomass, while complete mineralization is achieved when the polymer is transformed from the organic carbon (Andrady, 1994; Eubeler et al., 2009). Compared to microplastics, the chemical and physical effects of macroplastic debris are well-known globally. Previously, researchers have only focused on the contaminants released from plastics but not the ability of the plastics to absorb harmful contaminants from the environment.

Microplastics have been found in different water sources such as wastewater treatment plants, freshwater, and marine (Rezania et al., 2018). Previous studies have successfully identified that microplastic debris may exceed 100,000 items/m<sup>2</sup> in water surface and 100,000 pellets/m in beach sediment (Eerkes-Medrano, Thompson, & Aldridge, 2015; Cauwenberghe et al., 2015). According to previous researchers, microplastics are also suggested as a long-term sink in sediments (Cozar et al., 2014; Imhof et al., 2017; Coppock et al., 2017). The density of seawater is around 1.020 to 1.029 gcm<sup>-3</sup>; hence, plastics with a density higher than seawater will sink and potentially accumulate in the sediment. However, if the plastics' density is lower than the seawater density, they tend to float in the water or the surface column (Cauwenberghe et al., 2015).

## **1.2 Research Background**

Since the 1970s, plastics have been incidentally ingested by organisms such as fish (Wieczorek et al., 2018; Lv et al., 2019), invertebrates (Windsor et al., 2019; Horn et al., 2019), turtles (Nicolau et al., 2016), and seabirds (Basto et al., 2019). Thus, plastic ingestion may affect organisms physically or chemically (Mattsson et al., 2018; Mattsson et al., 2017). In terms of physical impact, organisms such as seabird will experience suppressed feeding activity when the plastic pellets in the gizzard indicate no fresh food is reaching the proventriculus; thus, this blocks the movement of food through the digestive tract and might reduce its appetite or change its food hunting

behaviour (Rochman et al., 2014; Law, 2017), thereby causing histopathological alterations in intestines, changes in behaviour and lipid metabolism, and potentially translocation to the liver (Jovanović, 2017).

In the case of microplastics, a study has found that seabass larvae ingest polyethylene microbeads that went through the digestive tract during fish fed diet despite its high egestion behavior (Mazurais et al., 2015). Meanwhile, in a highly contaminated area, microplastics with co-contaminants might be ingested by zooplankton and then fish larvae (Mazurais et al., 2015; Moira et al., 2015; Khan et al., 2017). Thus, large predators with complex digestive tracts such as crustaceans, ctenophores, and medusae, or vertebrates such as fishes might also ingest the fish larvae even before the egestion process (Cole et al., 2013; Carlos et al., 2018). Consequently, the bioaccumulation process could harm organisms, especially the top predators because the microplastic co-contaminants have a potential for biomagnification (Teuten et al., 2009). Figure 1.2 shows potential pathway for the transport of microplastics & their biological interactions in environment.

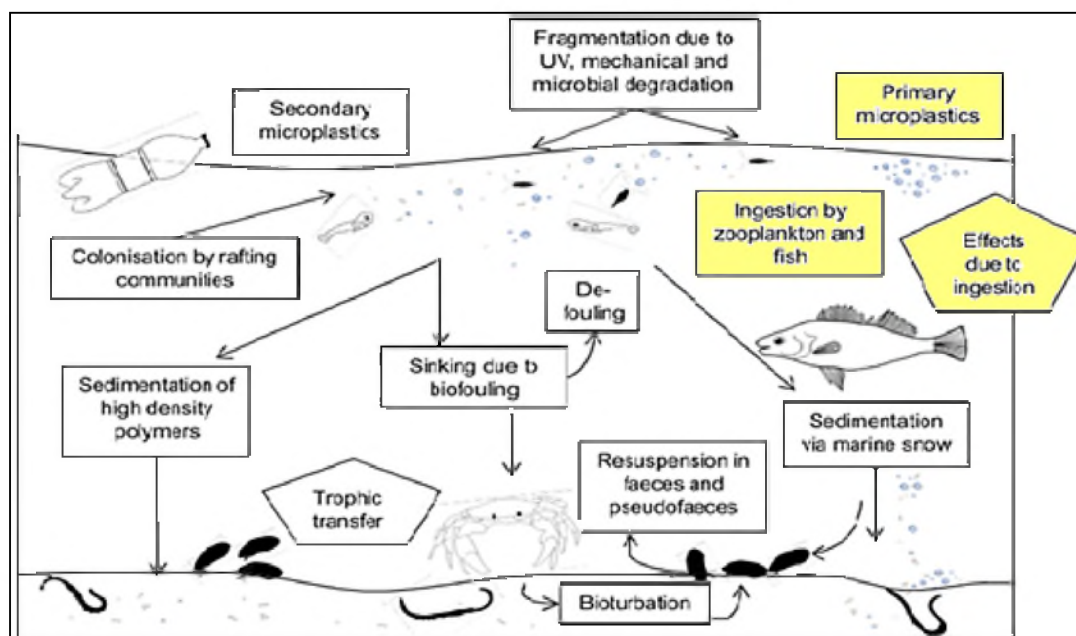


Figure 1.2 Potential pathway for the transport of microplastics & their biological interactions (adapted from Wright et al., 2013)

Research has been paying attention to investigating the impacts of toxic chemicals, especially on the relationship involving microplastic debris. Microplastics might act as a source or sink of toxic chemicals (Cole et al., 2011) since they have the ability to adsorb persistent, bioaccumulative, and toxic substances (PBTs) such as heavy metals, polychlorinated biphenyls (PCBs), and dioxins from water or sediment. These PBTs may then be released when the microplastics are ingested by aquatic life (Rochman et al., 2014; Karami et al., 2017), and the accumulation of microplastics is likely ingested by a wider range of organisms, thus raising concern in the marine ecosystem.

Recent studies have found that microplastics might be mobilized from digestive organ to other internal systems and trophic transferred from prey to predator (Dawson et al., 2018), and it has also been proven that toxic pollutants such as polychlorinated biphenyls (PCBs), dichlorodiphenyltrichloroethane (DDT), polycyclic aromatic hydrocarbons (PAHs), dioxin, and heavy metals tend to adsorb onto microbeads and yield higher concentration compared to water and sediments (Brennecke, Duarte, Paiva, Caçador, & Canning-Clode, 2016). Therefore, the presence of microbeads in the marine ecosystem poses a threat to food safety. Current research suggests that, despite the significant uncertainty and complexity in the kinetics and thermodynamics of the interaction, plastic debris also appears to act as a vehicle, transferring PBTs from the water to the food web, thus snowballing risk to the entire marine food web including humans. Due to the extremely long lifetime of plastics and PBTs in the ocean, prevention strategies are, therefore, crucial to minimize these risks (Engler, 2012; Turner, 2016).

Microbeads are classified as a primary source for microplastic pollution since the size of microbeads is in the range of microplastics (Thompson, 2004). Microbeads are manufactured and designed as demanded such as facial cleansers (Fendall & Sewell, 2009; Brennecke et al., 2016; Boucher et al., 2016). These materials are also widely used in cleaning products, printer toner, and abrasive media for plastic blasting, textile printing, and automotive molding (Pettipas et al., 2016). Subsequently, after treated and discharged, microbeads tend to accumulate, persist, and potentially act as

vectors for contaminants in the environment (Imhof et al., 2016; Brennecke et al., 2016; Smith, 2018) and poses a threat to food safety.

The main concern is that, when organisms ingest plastic-co-contaminants, the bound contaminants will likely be released due to chemical or physical conditions in the avian gizzard or the digestive tract of the organisms. To date, research on the ingestion effect of adsorbed heavy metals on microplastics in the marine organism is still lacking. The adsorption of metals to microplastics raises the potential for chemical transfer to marine animals that falsely ingest microplastics, which is identified as a “vector effect” (Syberg et al., 2015; Ory et al., 2018).

As plastics become vectors due to their ability to deploy and concentrate contaminants through ingestion to organisms (Khan et al., 2015), other than bioaccumulation, the contaminants also allow for biomagnification within the environment system, which starts from the low trophic predators until the top predators (Graham & Thompson, 2009). As such, various trophic level organisms that ingest microplastics might affect the vast ecosystem through the biomagnification of microplastic co-contaminants or microplastics alone. While the focus is directed towards the persistent organic pollutants (POPs) possibility concerning microplastics that carry heavy metals and ingested by the organism; therefore, it is important to investigate some organisms that appear to not only be unintentionally ingesting but also selectively consuming the floating microplastics (Ory et al., 2018; Hall et al., 2015).

The term “heavy metals” could be explained as the elements with high density and toxic to the environments even at low concentrations (Carolin et al., 2017). Heavy metals have been a major threat to the environment due to their flexibility, accumulation, endurance, and non-biodegradable characteristics (Raval et al., 2016). Usually, heavy metal pollution is caused by discharging untreated wastewater from industries such as pesticides, tanneries, metal plating industries, or mining operations.

Some heavy metals have become the main concern in the existing environment such as cadmium, chromium, cobalt, copper, lead, manganese, mercury, nickel, silver, and zinc (Bhattacharyya & Gupta, 2008; Turner & Millward, 2002). The concentrations of heavy metal ions in plastic particles are significantly higher than in the water column, which makes them likely to become toxic. As stated by Holmes (2013), heavy metals adsorbed to plastic pellets are highly bioavailable; therefore, toxic elements are expected to be extracted by the acidic digestive tract environment (Holmes & Thompson, 2014). As such, the ability of heavy metals to release back to the marine environment is more readily in a soluble form (Holmes et al., 2012).

Recognizing that there are species-specific, pollutant and polymer specific, as well as experimental differences between the studies, varied results demonstrated that the impact of microplastics on the uptake and accumulation of pollutants is far from consistent. One of the key determinants may be whether pollutants and microplastics encounter each other before organism exposure or whether or not they are introduced as a co-exposure.

To date, scientists have discovered a new pathway for heavy metal pollution by a carrier vector effect with microplastics as the vector (Kalčíková et al., 2017; Bradney et al., 2019; Hodson et al., 2017). For example, Vedolin et al. (2018) demonstrated the ability of microplastics to adsorb heavy metals as the concentration of the adsorbed heavy metals in the collected pellets were higher than the original particles. Following the ban by the US, UK, Canada, and other countries, it is crucial to investigate the ability of pristine polyethylene microbeads to adsorb heavy metals such as cadmium, chromium, and lead. Thus, this study focuses on the adsorption of heavy metals to the microbeads to prove the vector effect of heavy metals on the marine vertebrates.

### 1.3 Problem Statement

There have been fewer studies about ingestions of microbeads from cosmetic usage regardless of their occurrences in wastewater treatment plants (Rezania et al., 2018). In Malaysia, the only study of microbeads occurrence was conducted by Praveena et al. (2018). It is estimated that about 0.199 trillion microbeads particles from facial and personal care products were reported to enter the wastewater treatment plants (WWTPs) in Malaysia per day; hence, this problem is at an alarming point and needs sustainable solutions (Praveena et al., 2018). However, the data on possibility ingestion effect on microbeads to organism in Malaysia was still scarce.

Studies have mostly proven the ability of a general type of microplastics to adsorb heavy metals from the aqueous environment (Tchounwou et al., 2014; Boucher et al., 2016; Alomar et al., 2017; Peng et al., 2018; Munier & Bendell, 2018; Prunier et al., 2019). Specifically, studies on microbeads in the cosmetic industry and their ability to sorb contaminants are scarce.

Based on the literature review, microbeads have the potential to be a vector for heavy metals in the marine environment (Bayo et al., 2017; Zon et al., 2018). As the microbeads have the ability to carry hazardous ions from the domestic wastewater system into the marine environment, this increases the potential of heavy metal ions to pollute the water system and interact with marine animals. These phenomena increase the mobility of heavy metals into the water system. As the mobility of the heavy metals increases, the marine animals are highly likely to consume the heavy metal ions, thus transferring the pollution into the food web up to the final consumers, which are the humans. Therefore, this study will focus on the adsorption and desorption abilities of microbeads onto heavy metal ions.

This study was focused on juvenile seabass. To date, there is no study performed in evaluating heavy metals with microbeads effects on the uptake of juvenile seabass species specifically *Lates calcarifer*. Juvenile fish is predominantly exposed to pollutions in the environment, especially heavy metal ions (Morán et al., 2018). Therefore, any changes in this stage will positively affect the growth of the fish.



On the other hand, the bio-accumulation of heavy metal ions would cause a significant effect on the tissue growth for some period; thus, the polluted organs may affect the consumers. The early monitoring of the pollution is vital, especially in the juvenile animal class so the initial prevention acts can be planned and executed to save the environment. Therefore, further investigation was carried out for the effect of adsorbed microbeads with heavy metals into juvenile seabass as a marine organism model in different exposure condition. This experiment is crucial to prove the vector effect of microbeads and identify the pollution level in the vital organs of juvenile seabass.

#### **1.4 Objectives**

This study aims to clarify the interactions between polyethylene microbeads and heavy metals in the environment and to determine the possibility of polyethylene microbeads to become heavy metal vectors for juvenile seabass, also known as seabass. As such, the following objectives are addressed in this study:

1. To determine the adsorption ability of polyethylene microbeads in cadmium, chromium, and lead using a batch approach.
2. To determine the kinetic and isotherm characteristics for the adsorption of cadmium, chromium, and lead onto polyethylene microbeads.
3. To quantify the uptake of cadmium, chromium, and lead within gastrointestinal tracts, gills, and skin of juvenile seabass.
4. To determine the effect of polyethylene microbeads on seabass via direct and preloaded exposure with cadmium, chromium, and lead.

#### **1.5 Scopes**

The following research scopes have been summarized to accomplish the research aims. The scopes are listed as follows:

- a) Microbeads were used as a model of microplastics in this study. Primarily, it is the second-most obviously ingested type of microplastics and evidently found within every species of marine animals. Moreover, polyethylene microbeads have also been found in scrubs/peelings, shower/bath products, facial cleaners, toothpaste, bubble bath, lotions, and sunscreens. Besides, other types of polymer and wastewater discharge might be the ultimate ways to enter the aquatic ecosystem. The source of microbeads was obtained from a local private company that produces microbeads for cosmetic usage, purposely with a diameter of ~300  $\mu\text{m}$ .
- b) Cadmium (Cd), chromium (Cr), and lead (Pb) were selected because they are common contaminants to the marine ecosystems (Yunus, 2020). In addition, these heavy metals are known in industrial, domestic, agricultural, medical, and technological applications and are widely distributed in the environment; hence, this has raised concerns over their toxicology to human health and the environment.
- c) Juvenile seabass was collected from the Aquaculture Fisheries Research Institute, Johor Bahru with a size of 6-7 cm in length. While they are highly demanded food with a high market value in the aquaculture industry, they are also important for commercial purposes and game fish. Briefly, they are valuable both as recreational and commercial fish with a high, fairly stable price. Thus, juvenile seabass was chosen because it potentially ingests microplastics in the environment, mistaking the microplastics as prey or unable to distinguish between prey and food. Another vital factor for the selection of the juvenile class is its feeding behaviour. Even though recent studies have focused on the adult class, the bioaccumulation and distribution of heavy metals in the adult's organs and tissues are questionable since the adult diets are more selective, involving a wide range of animals. Therefore, the various sources or vectors of pollutants may be consumed by the adults. As a comparison, juveniles only have a plankto-phytophage diet, which only consumes diatoms, zooplankton, or green algae (Markovic, 2007). Since the size of microbeads is similar to plankton, the chance is higher for this proposed vector to be consumed by the test subjects.

## **1.6 Significance of the Study**

Environmental and health consequences are the main concern of this study since there is a high potential for microbeads and heavy metals to bio-accumulate in organisms, thus climbing up to the top predator through the food chain. As such, the ability of microbeads to accumulate heavy metals higher than seawater has been proven in previous studies (Khan et al., 2015; Brennecke et al., 2016). In general, heavy metals adsorbed onto microbeads are highly available in the environment. Microbeads with heavy metals tend to be desorbed in organisms via acidic conditions in their digestive tracts after ingestion (Holmes & Thompson, 2014; Khan et al., 2017). Consequently, microbeads have become a crucial concern since they appear in small sizes and can easily be ingested by a broader range of marine organisms. Besides, the bioavailability of vector-heavy metal microbeads increases the possibility to transfer harmful chemicals to the food chain.

While varied research results have demonstrated microplastics and contaminants adsorption due to the ‘specific polymer adsorbed specific pollutant’ characteristic, the same situation has been predicted for different organisms for different microplastic-heavy metals intake. Thus, the issue should begin with finding the relation involving the presence of microbeads in heavy metal exposure, including the exposure condition that favours the vector effect of microbeads carrying heavy metals into juvenile seabass via ingestion. In Malaysia, a website called [cleanmalaysia.com](http://cleanmalaysia.com) was established to create the awareness of microbeads pollutant as an effort to bring the light of environmental danger since 2016. As the occurrence of microbeads in the marine ecosystem poses a threat to food safety and results in the banning of microbeads by the US, UK, and Canada, it is essential to study the sorption of heavy metal pollutants on microbeads and their ingestion effects on marine organisms. Thus, raising the awareness of microbeads usage in cosmetic products and followed the banned action in the same way as other countries.

## REFERENCES

- Abbasi, S., Soltani, N., Keshavarzi, B., Moore, F., Turner, A., & Hassanaghaei, M. (2018). Microplastics in different tissues of fish and prawn from the Musa Estuary, Persian Gulf. *Chemosphere*, 205, 80–87.
- Ahmed, M., Ahmad, T., Liaquat, M., Abbasi, K. S., Farid, I. B. A., & Jahangir, M. (2016). Tissue specific metal characterization of selected fish species in Pakistan. *Environmental Monitoring and Assessment*, 188(4).
- Akhbarizadeh, R., Moore, F., & Keshavarzi, B. (2018). Investigating a probable relationship between microplastics and potentially toxic elements in fish muscles from northeast of Persian. *Environmental Pollution*, 232, 154–163.
- Ali Azadi, N., Mansouri, B., Spada, L., Sinkakarimi, M. H., Hamesadeghi, Y., & Mansouri, A. (2018). Contamination of lead (Pb) in the coastal sediments of north and south of Iran: a review study. *Chemistry and Ecology*, 34(9), 884–900.
- Aljaibachi, R., & Callaghan, A. (2018). Impact of polystyrene microplastics on *Daphnia magna* mortality and reproduction in relation to food availability. *PeerJ*, 6, e4601.
- Alomar, C., Sureda, A., Capó, X., Guijarro, B., Tejada, S., & Deudero, S. (2017). Microplastic ingestion by *Mullus surmuletus* Linnaeus, 1758 fish and its potential for causing oxidative stress. *Environmental Research*, 159, 135–142.
- Amanda, M. (2018). *There's Plastic in Your Poop: Study Warns Microplastics May Be Hurting Us All*. Retrieved from <https://www.health.com/home/microplastics-human-poop>
- Amat, M. K. A. bin. (2017). *Synthesis and characterization of molybdenum carbide from oil palm based activated carbon for carbon dioxide reduction*. Universiti Teknologi Malaysia.
- Anderson, J. C., Park, B. J., & Palace, V. P. (2016). Microplastics in aquatic environments: Implications for Canadian ecosystems. *Environmental Pollution*, 218, 269–280.

- Andersson, K. I., Eriksson, M., & Norgren, M. (2011). Removal of Lignin from Wastewater Generated by Mechanical Pulping Using Activated Charcoal and Fly Ash: Adsorption Isotherms and Thermodynamics. *Industrial & Engineering Chemistry Research*, 50(13), 7722–7732.
- Andrady, A. L., & Neal, M. A. (2009). Applications and societal benefits of plastics. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 364(1526), 1977–1984.
- Andrady, A. L. (2017). The plastic in microplastics: A review. *Marine Pollution Bulletin*, 119(1), 12–22.
- Andrady, Anthony L. (1994). Assessment of Environmental Biodegradation of Synthetic Polymers. *Journal of Macromolecular Science, Part C*, 34(1), 25–76.
- Andrady, Anthony L. (2011). Microplastics in the marine environment. *Marine Pollution Bulletin*, 62(8), 1596–1605.
- Arantes, F. P., Savassi, L. A., Santos, H. B., Gomes, M. V. T., & Bazzoli, N. (2016). Bioaccumulation of mercury, cadmium, zinc, chromium, and lead in muscle, liver, and spleen tissues of a large commercially valuable catfish species from Brazil. *Annals of the Brazilian Academy of Sciences*, 88(1), 137–147.
- Artham, T., Sudhakar, M., Venkatesan, R., Nair, C. M., Murty, K. V. G. K., & Doble, M. (2009). Biofouling and stability of synthetic polymers in seawater. *International Biodeterioration & Biodegradation*, 63, 884–890.
- Asefnejad, A., Khorasani, M. T., Behnamghader, A., Farsadzadeh, B., & Bonakdar, S. (2011). Manufacturing of biodegradable polyurethane scaffolds based on polycaprolactone using a phase separation method: physical properties and in vitro assay. *International Journal of Nanomedicine*, 6, 2375–2384.
- Ashton, K., Holmes, L., & Turner, A. (2010). Association of metals with plastic production pellets in the marine environment. *Marine Pollution Bulletin*, 60(11), 2050–2055.
- Auta, H. S., Emenike, C. U., & Fauziah, S. H. (2017). Distribution and importance of microplastics in the marine environment: A review of the sources, fate, effects, and potential solutions. *Environment International*, 102, 165–176.

- Auta, H. S., Emenike, C. U., Jayanthi, B., & Fauziah, S. H. (2018). Growth kinetics and biodeterioration of polypropylene microplastics by *Bacillus* sp. and *Rhodococcus* sp. isolated from mangrove sediment. *Marine Pollution Bulletin*, 127, 15–21.
- Authman, M. M., Zaki, M. S., Khallaf, E. A., & Abbas, H. H. (2015). Use of fish as bio-indicator of the effects of heavy metals pollution. *Journal of Aquaculture Research & Development*, 6(4), 1000328.
- Avery-gomm, S., Borrelle, S. B., & Provencher, J. F. (2018). Linking plastic ingestion research with marine wildlife conservation. *Science of the Total Environment*, 637–638, 1492–1495.
- Avio, C. G., Gorbi, S., & Regoli, F. (2015). Experimental development of a new protocol for extraction and characterization of microplastics in fish tissues: First observations in commercial species from Adriatic Sea. *Marine Environmental Research*, 111, 18–26.
- Ayangbenro, A. S., & Babalola, O. O. (2017). A New Strategy for Heavy Metal Polluted Environments: A Review of Microbial Biosorbents. *International Journal of Environmental Research and Public Health*, 14(1).
- Bakir, A., Rowland, S. J., & Thompson, R. C. (2012). Competitive sorption of persistent organic pollutants onto microplastics in the marine environment. *Marine Pollution Bulletin*, 64(12), 2782–2789.
- Bakir, A., Rowland, S. J., & Thompson, R. C. (2014). Enhanced desorption of persistent organic pollutants from microplastics under simulated physiological conditions. *Environmental Pollution*, 185, 16–23.
- Banerjee, S., Dubey, S., Gautam, R. K., Chattopadhyaya, M. C., & Sharma, Y. C. (2019). Adsorption characteristics of alumina nanoparticles for the removal of hazardous dye, Orange G from aqueous solutions. *Arabian Journal of Chemistry*, 12(8), 5339–5354.
- Barbier, O., Jacquillet, G., Tauc, M., Cougnon, M., & Poujeol, P. (2005). Effect of Heavy Metals on, and Handling by, the Kidney. *Nephron Physiology*, 99(4), p105–p110.
- Barboza, Luís Gabriel Antão, Carolina, B., & Gimenez, G. (2015). Microplastics in the marine environment: Current trends and future perspectives. *Marine Pollution Bulletin*, 97, 5–12.

- Barboza, Luís Gabriel Antão, Russo, L., Branco, V., Figueiredo, N., Carvalho, F., Carvalho, C., & Guilhermino, L. (2018). Microplastics cause neurotoxicity, oxidative damage and energy-related changes and interact with the bioaccumulation of mercury in the European seabass, *Dicentrarchus labrax* (Linnaeus, 1758). *Aquatic Toxicology*, 195, 49–57.
- Barboza, Luís Gabriel Antão, Vethaak, A. D., Lavorante, B. R. B. O., Lundebye, A., & Guilhermino, L. (2018). Marine microplastic debris : An emerging issue for food security, food safety and human health. *Marine Pollution Bulletin*, 133, 336–348.
- Barboza, Luís Gabriel Antão, Vieira, L. R., & Guilhermino, L. (2018). Single and combined effects of microplastics and mercury on juveniles of the European seabass (*Dicentrarchus labrax*): Changes in behavioural responses and reduction of swimming velocity and resistance time\*. *Environmental Pollution*, 236, 1014–1019.
- Barnes, D. K. A., Galgani, F., Thompson, R. C., & Barlaz, M. (2009). Accumulation and fragmentation of plastic debris in global environments. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 364(1526), 1985–1998.
- Basto, M. N., Nicastro, K. R., Tavares, A. I., Mcquaid, C. D., Casero, M., Azevedo, F., & Zardi, G. I. (2019). Plastic ingestion in aquatic birds in Portugal. *Marine Pollution Bulletin*, 138, 19–24.
- Batel, A., Linti, F., Scherer, M., Erdinger, L., & Braunbeck, T. (2016). Transfer of benzo[a]pyrene from microplastics to *Artemia nauplii* and further to zebrafish via a trophic food web experiment: CYP1A induction and visual tracking of persistent organic pollutants. *Environmental Toxicology and Chemistry*, 35(7), 1656–1666.
- Bayo, J., Martínez, A., Guillén, M., Olmos, S., Roca, M.-J., & Alcolea, A. (2017). Microbeads in Commercial Facial Cleansers: Threatening. *Clean Soil Air Water*, 45(7), 1600683.
- Bervoets, L., Blust, R., & Verheyen, R. (2001). Accumulation of Metals in the Tissues of Three Spined Stickelback (*Gasterosteus aculeatus*) from Natural Fresh Waters. *Ecotoxicology and Environmental Safety*, 48(2), 117–127.

- Besseling, E., Wegner, A., Foekema, E. M., Heuvel-greve, M. J. Van Den, & Koelmans, A. A. (2013). Effects of Microplastic on Fitness and PCB Bioaccumulation by the Lugworm *Arenicola marina* (L.). *Environmental Sciences & Technology*, 47(1), 593–600.
- Bhatnagar, A., & Sillanpää, M. (2009). Applications of chitin- and chitosan-derivatives for the detoxification of water and wastewater — A short review. *Advances in Colloid and Interface Science*, 152(1), 26–38.
- Bhattacharyya, K. G., & Gupta, S. Sen. (2008). Adsorption of a few heavy metals on natural and modified kaolinite and montmorillonite: A review. *Advances in Colloid and Interface Science*, 140(2), 114–131.
- Boerger, C. M., Lattin, G. L., Moore, S. L., & Moore, C. J. (2010). Plastic ingestion by planktivorous fishes in the North Pacific Central Gyre. *Marine Pollution Bulletin*, 60(12), 2275–2278.
- Bonner, T. H., Thomas, C., Williams, C. S., & Karges, J. P. (2005). Temporal Assessment of a West Texas Stream Fish Assemblage. *The Southwestern Naturalist*, 50(1), 74–78.
- Bosch, A. C., O'Neill, B., Sigge, G. O., Kerwath, S. E., & Hoffman, L. C. (2016). Heavy metals in marine fish meat and consumer health: A review. *Journal of the Science of Food and Agriculture*, 96(1), 32–48.
- Botterell, Z. L. R., Beaumont, N., Dorrington, T., Steinke, M., Thompson, R. C., & Lindeque, P. K. (2020). Bioavailability and effects of microplastics on marine zooplankton: A review. *Environmental Pollution*, 245, 98–110.
- Boucher, C., Morin, M., & Bendell, L. I. (2016). The influence of cosmetic microbeads on the sorptive behavior of cadmium and lead within intertidal sediments: A laboratory study. *Regional Studies in Marine Science*, 3, 1–7.
- Bour, A., Giacomo, C., Gorbi, S., & Regoli, F. (2018). Presence of microplastics in benthic and epibenthic organisms: Influence of habitat, feeding mode and trophic level\*. *Environmental Pollution*, 243, 1217–1225.



- Bour, A., Hossain, S., Taylor, M., Sumner, M., & Carney Almroth, B. (2020). Synthetic Microfiber and Microbead Exposure and Retention Time in Model Aquatic Species Under Different Exposure Scenarios. *Frontiers in Environmental Science*, Vol. 8, p. 83.
- Brack, H. G. (2015). *Where Have All the Plastics Gone?* (Vol. 5). Pennywheel Press.
- Bradney, L., Wijesekara, H., Niroshika, K., & Kirkham, M. B. (2019). Particulate plastics as a vector for toxic trace-element uptake by aquatic and terrestrial organisms and human health risk. *Environment International*, 131, 104937.
- Bravo Rebolledo, E. L., Van Franeker, J. A., Jansen, O. E., & Brasseur, S. M. J. M. (2013). Plastic ingestion by harbour seals (*Phoca vitulina*) in The Netherlands. *Marine Pollution Bulletin*, 67(1), 200–202.
- Brennecke, D., Duarte, B., Paiva, F., Caçador, I., & Canning-Clode, J. (2016). Microplastics as vector for heavy metal contamination from the marine environment. *Estuarine, Coastal and Shelf Science*, 178, 189–195.
- Browne, Mark A. (2015). Source and pathways of microplastics to habitats. In A. A. Koelmans, E. Besseling, & W. J. Shim (Eds.), *Marine Anthropogenic Litter* (pp. 229–244). Springer.
- Browne, Mark A., Dissanayake, A., Galloway, T. S., Lowe, D. M., & Thompson, R. C. (2008). Ingested Microscopic Plastic Translocates to the Circulatory System of the Mussel, *Mytilus edulis* (L.). *Environmental Science & Technology*, 42(13), 5026–5031.
- Browne, Mark Anthony. (2008). Ingested Microscopic Plastic Translocates to the Circulatory System of the Ingested Microscopic Plastic Translocates to the Circulatory System of the Mussel, *Mytilus edulis* (L.). *Environmental Science & Technology*, 42, 5026–5031.
- Browne, Mark Anthony, Crump, P., Niven, S. J., Teuten, E., Tonkin, A., Galloway, T., & Thompson, R. (2011). Accumulation of Microplastic on Shorelines Worldwide: Sources and Sinks. *Environmental Science & Technology*, 45(21), 9175–9179.
- Burton G. Allen, J. (2002). Sediment quality criteria in use around the world. *Limnology*, 3(2), 65–76.
- CABI. (2019). *Lates calcarifer*. In: *Invasive Species Compendium*. Wallingford, UK: CAB International. Retrieved from [www.cabi.org/isc](http://www.cabi.org/isc)

- Campanale, C., Massarelli, C., Savino, I., Locaputo, V., & Uricchio, V. F. (2020). A Detailed Review Study on Potential Effects of Microplastics and Additives of Concern on Human Health. *International Journal of Environmental Research and Public Health*, 17(4), 1212.
- Carbery, M., Connor, W. O., & Thavamani, P. (2018). Trophic transfer of microplastics and mixed contaminants in the marine food web and implications for human health. *Environmental International*, 115, 400–409.
- Carolin, C. F., Kumar, P. S., Saravanan, A., Joshiba, G. J., & Naushad, M. (2017). Efficient techniques for the removal of toxic heavy metals from aquatic environment: A review. *Journal of Environmental Chemical Engineering*, 5(3), 2782–2799.
- Caron, A. G. M., Thomas, C. R., Berry, K. L. E., Motti, C. A., Ariel, E., & Brodie, J. E. (2018). Ingestion of microplastic debris by green sea turtles (*Chelonia mydas*) in the Great Barrier Reef: Validation of a sequential extraction protocol. *Marine Pollution Bulletin*, 127, 743–751.
- Carpenter, E. J., Anderson, S. J., Harvey, G. R., Miklas, H. P., & Peck, B. B. (1972). Polystyrene Spherules in Coastal Waters. *Science*, 178(4062), 749–750.
- Carreras-colom, E., Constenla, M., Soler-membrives, A., Cartes, J. E., & Baeza, M. (2018). Spatial occurrence and effects of microplastic ingestion on the deep-water shrimp *Aristeus antennatus*. *Marine Pollution Bulletin*, 133, 44–52.
- Cauwenberghe, L. Van, Devriese, L., Galgani, F., Robbens, J., & Janssen, C. R. (2015). Microplastics in sediments: A review of techniques, occurrence and effects. *Marine Environmental Research*, 111, 5–17.
- Cauwenberghe, L. Van, & Janssen, C. R. (2014). Microplastics in bivalves cultured for human consumption. *Environmental Pollution*, 193, 65–70.
- Chagnon, C., Thiel, M., Antunes, J., Lia, J., Sobral, P., & Christian, N. (2018). Plastic ingestion and trophic transfer between Easter Island flying fish (*Cheilopogon rapanouiensis*) and yellow fin tuna (*Thunnus albacares*) from Rapa Nui (Easter Island)\*. *Environmental Pollution*, 243, 127–133.
- Chang, M., & Chang, M. (2016). Reducing microplastics from facial exfoliating cleansers in wastewater through treatment versus consumer product decisions through treatment versus consumer product decisions. *Marine Pollution Bulletin*, 101(1), 330–333.

- Chen, L., Du, S., Song, D., Zhang, P., & Zhang, L. (2019). Ecotoxicology of Heavy Metals in Marine Fish. In D. S. Pei & M. Junaid (Eds.), *Marine Pollution: Current Status, Impacts and Remedies* (pp. 173–230). Singapore: Bentham Science Publishers.
- Chércoles Asensio, R., San Andrés Moya, M., de la Roja, J. M., & Gómez, M. (2009). Analytical characterization of polymers used in conservation and restoration by ATR-FTIR spectroscopy. *Analytical and Bioanalytical Chemistry*, 395(7), 2081–2096.
- Cheung, P. K., & Fok, L. (2016). Evidence of microbeads from personal care product contaminating the sea. *Marine Pollution Bulletin*, 109(1), 582–585.
- Chisada, S., Yoshida, M., & Karita, K. (2019). Ingestion of polyethylene microbeads affects the growth and reproduction of medaka, *Oryzias latipes*\*. *Environmental Pollution*, 254, 113094.
- Chua, E. M., Shimeta, J., Nugegoda, D., Morrison, P. D., & Clarke, B. O. (2014). Assimilation of Polybrominated Diphenyl Ethers from Microplastics by the Marine Amphipod, *Allorchestes Compressa*. *Environmental Science & Technology*, 48(14), 8127–8134.
- Cole, M., Lindeque, P., Fileman, E., Halsband, C., Goodhead, R., Moger, J., & Galloway, T. S. (2013). Microplastic Ingestion by Zooplankton. *Environmental Science & Technology*, 47(12), 6646–6655.
- Cole, M., Lindeque, P., Halsband, C., & Galloway, T. S. (2011). Microplastics as contaminants in the marine environment: A review. *Marine Pollution Bulletin*, 62(12), 2588–2597.
- Collard, F., Gilbert, B., Comp, P., Eppe, G., Das, K., Jauniaux, T., & Parmentier, E. (2017). Microplastics in livers of European anchovies (*Engraulis encrasicolus*, L)\*. *Environmental Pollution*, 229, 1000–1005.
- Collicutt, B., Juanes, F., & Dudas, S. E. (2019). Microplastics in juvenile Chinook salmon and their nearshore environments on the east coast of Vancouver Island\*. *Environmental Pollution*, 244, 135–142.
- Compa, M., Ventero, A., Iglesias, M., & Deudero, S. (2018). Ingestion of microplastics and natural fibres in *Sardina pilchardus* (Walbaum, 1792) and *Engraulis encrasicolus* (Linnaeus, 1758) along the Spanish Mediterranean coast. *Marine Pollution Bulletin*, 128, 89–96.

- Coppock, R. L., Cole, M., Lindeque, P. K., Queir, A. M., & Galloway, T. S. (2017). A small-scale, portable method for extracting microplastics from marine sediments. *Environmental Pollution*, 230, 829–837.
- Cozar, A., Echevarria, F., Gonzalez-Gordillo, J. I., Irigoien, X., Ubeda, B., Hernandez-Leon, S., Palma, A. T., Navarro, S., Garcia-de-Lomas, J., Ruiz, A., Fernandez-de-Puelles, M. L., & Duarte, C. M. (2014). Plastic debris in the open ocean. *Proceedings of the National Academy of Sciences*, 111(28), 10239–10244.
- Crawford, C. B., & Quinn, B. (2017). The interactions of microplastics and chemical pollutants. In *Microplastic Pollutants* (pp. 131–157). Elsevier Inc.
- Dauvergne, P. (2018). The power of environmental norms: marine plastic pollution and the politics of microbeads. *Environmental Politics*, 27(4), 579–597.
- Davarpanah, E., & Guilhermino, L. (2015). Estuarine, Coastal and Shelf Science Single and combined effects of microplastics and copper on the population growth of the marine microalgae *Tetraselmis chuii*. *Estuarine, Coastal and Shelf Science*, 167, 269–275.
- Davis, T. L. O. (1982). Maturity and sexuality in Barramundi, *Lates calcarifer* (Bloch), in the Northern Territory and south-eastern Gulf of Carpentaria. *Marine and Freshwater Research*, 33(3), 529–545.
- Davis, T. L. O. (1984). A Population of Sexually Precocious Barramundi, *Lates calcarifer*, in the Gulf of Carpentaria, Australia. *Copeia*, 1984(1), 144–149.
- Davis, T. L. O. (1986). Migration patterns in barramundi, *Lates calcarifer* (Bloch), in Van Diemen Gulf, Australia, with estimates of fishing mortality in specific areas. *Fisheries Research*, 4(3), 243–258.
- Dawson, A., Huston, W., Kawaguchi, S., King, C., Cropp, R., Wild, S., Eisenmann, P., Townsend, K., & Bengtson Nash, S. M. (2018). Uptake and Depuration Kinetics Influence Microplastic Bioaccumulation and Toxicity in Antarctic Krill (*Euphausia superba*). *Environmental Science and Technology*, 52(5), 3195–3201.
- DeArmitt, C. (2017). 23 - Functional Fillers for Plastics. In M. B. T.-A. P. E. H. (Second E. Kutz (Ed.), *Plastics Design Library* (pp. 517–532). William Andrew Publishing.

- Demim, S., Drouiche, N., Aouabed, A., Benayad, T., Dendene-Badache, O., & Semsari, S. (2013). Cadmium and nickel: Assessment of the physiological effects and heavy metal removal using a response surface approach by L. gibba. *Ecological Engineering*, 61, 426–435.
- Ding, N., An, D., Yin, X., & Sun, Y. (2020). Detection and evaluation of microbeads and other microplastics in wastewater treatment plant samples. *Environmental Science and Pollution Research*, 27, 15878–15887.
- Dris, R., Gasperi, J., Saad, M., Mirande, C., & Tassin, B. (2016). Synthetic fibers in atmospheric fallout: A source of microplastics in the environment? *Marine Pollution Bulletin*, 104(1), 290–293.
- Eckert, E. M., Di, A., Therese, M., Arias-andres, M., Fontaneto, D., Grossart, H., & Corno, G. (2018). Microplastics increase impact of treated wastewater on freshwater microbial community\*. *Environmental Pollution*, 234, 495–502.
- Eerkes-Medrano, D., Thompson, R. C., & Aldridge, D. C. (2015). Microplastics in freshwater systems: A review of the emerging threats, identification of knowledge gaps and prioritisation of research needs. *Water Research*, 75, 63–82.
- Endo, S., Takizawa, R., Okuda, K., Takada, H., Chiba, K., Kanehiro, H., Ogi, H., Yamashita, R., & Date, T. (2005). Concentration of polychlorinated biphenyls (PCBs) in beached resin pellets: Variability among individual particles and regional differences. *Marine Pollution Bulletin*, 50(10), 1103–1114.
- Engler, R. E. (2012). The complex interaction between marine debris and toxic chemicals in the ocean. *Environmental Science and Technology*, 46(22), 12302–12315.
- Eriksen, M., Mason, S., Wilson, S., Box, C., Zellers, A., Edwards, W., Farley, H., & Amato, S. (2013). Microplastic pollution in the surface waters of the Laurentian Great Lakes. *Marine Pollution Bulletin*, 77(1–2), 177–182.
- Eriksson, C., & Burton, H. (2003). Origin and biological accumulation of small plastic particles in fur-seal scats from Macquarie Island. *Ambio*, 32(6), 380–384.
- Espinosa, C., Cuesta, A., & Esteban, M. Á. (2017). Effects of dietary polyvinylchloride microparticles on general health, immune status and expression of several genes related to stress in gilthead seabream (*Sparus aurata* L.). *Fish and Shellfish Immunology*, 68, 251–259.

- Espinosa, Cristóbal, Beltrán, J. M. G., Esteban, M. A., & Cuesta, A. (2018). In vitro effects of virgin microplastics on fish head-kidney leucocyte activities\*. *Environmental Pollution*, 235, 30–38.
- Estahbanati, S., & Fahrenfeld, N. L. (2016). Influence of wastewater treatment plant discharges on microplastic concentrations in surface water. *Chemosphere*, 162, 277–284.
- Eubeler, J. P., Zok, S., Bernhard, M., & Knepper, T. P. (2009). Environmental biodegradation of synthetic polymers I. Test methodologies and procedures. *TrAC Trends in Analytical Chemistry*, 28(9), 1057–1072.
- Farhan R. Khan, Shashoua, Y., Crawford, A., Drury, A., Sheppard, K., Stewart, K., & Sculthorp, T. (2020). ‘The Plastic Nile’: First Evidence of Microplastic Contamination in Fish from the Nile River (Cairo, Egypt). *Toxics*, 8(2), 22.
- Farrell, P., & Nelson, K. (2013). Trophic level transfer of microplastic: *Mytilus edulis* (L.) to *Carcinus maenas* (L.). *Environmental Pollution*, 177, 1–3.
- Fendall, L. S., & Sewell, M. A. (2009). Contributing to marine pollution by washing your face: Microplastics in facial cleansers. *Marine Pollution Bulletin*, 58(8), 1225–1228.
- Filipič, M. (2012). Mechanisms of cadmium induced genomic instability. *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 733(1), 69–77.
- Fisner, M., Taniguchi, S., Moreira, F., Bicego, M. C., & Turra, A. (2013). Polycyclic aromatic hydrocarbons (PAHs) in plastic pellets: Variability in the concentration and composition at different sediment depths in a sandy beach. *Marine Pollution Bulletin*, 70(1), 219–226.
- Foekema, E. M., De Gruijter, C., Mergia, M. T., van Franeker, J. A., Murk, A. J., & Koelmans, A. A. (2013). *Plastic in North Sea Fish*. *Environmental Science & Technology*, 47(15), 8818–8824.
- Foo, K. Y., & Hameed, B. H. (2010). Insights into the modeling of adsorption isotherm systems. *Chemical Engineering Journal*, 156, 2–10.
- Food and Agriculture Organization of the United States. (2006). *Cultured Aquatic Species Information Programme. *Lates calcarifer**. FAO Fisheries and Aquaculture Department.

- Fossi, M. C., Coppola, D., Bains, M., Giannetti, M., Guerranti, C., Marsili, L., Panti, C., de Sabata, E., & Clò, S. (2014). Large filter feeding marine organisms as indicators of microplastic in the pelagic environment: The case studies of the Mediterranean basking shark (*Cetorhinus maximus*) and fin whale (*Balaenoptera physalus*). *Marine Environmental Research*, 100, 17–24.
- Fotopoulou, K. N., & Karapanagioti, H. K. (2012). Surface Properties of Beached Plastic Pellets. *Marine Environmental Research*, 81, 70–77.
- Fotopoulou, K. N., & Karapanagioti, H. K. (2015). Surface properties of beached plastics. *Environmental Science and Pollution Research*, 22(14), 11022–11032.
- Freije, A. M. (2015). Heavy metal, trace element and petroleum hydrocarbon pollution in the Arabian Gulf: Review. *Journal of the Association of Arab Universities for Basic and Applied Sciences*, 17, 90–100.
- Fu, F., & Wang, Q. (2011). Removal of heavy metal ions from wastewaters: A review. *Journal of Environmental Management*, 92(3), 407–418.
- Galgani, F., Hanke, G., Werner, S., Oosterbaan, L., Nilsson, P., Fleet, D. M., Kinsey, Susan., Thompson, R. C., Franeker, J. A. V., Vlachogianni, T., Scoullou, M., Veiga, J. M., Palatinus, A., Matiddi, M., Maes, T., Korpinen, S., Budziak, A., Leslie, H., Gago, J., & Liebezeit, G. (2013). Guidance on monitoring of marine litter in European seas. In G. Hanke, S. Werner, F. Galgani, J. Veiga, & M. Ferreira (Eds.), *MSFD Technical Subgroup on Marine Litter* (EUR – Scie, Vol. 70). Luxembourg: Publications Office of the European Union.
- Gambardella, C., Morgana, S., Bramini, M., Rotini, A., Manfra, L., Migliore, L., Piazza, V., Garaventa, F & Faimali, M. (2018). Ecotoxicological effects of polystyrene microbeads in a battery of marine organisms belonging to different trophic levels. *Marine Environmental Research*, 141, 313–321.
- Gambardella, C., Morgana, S., Ferrando, S., Bramini, M., Piazza, V., Costa, E., Garaventa, F & Faimali, M. (2017). Ecotoxicology and Environmental Safety Effects of polystyrene microbeads in marine planktonic crustaceans. *Ecotoxicology and Environmental Safety*, 145, 250–257.
- Gao, F., Li, J., Sun, C., Zhang, L., Jiang, F., Cao, W., & Zheng, L. (2019). Study on the capability and characteristics of heavy metals enriched on microplastics in marine environment. *Marine Pollution Bulletin*, 144, 61–67.

- Gasperi, J., Wright, S. L., Dris, R., Collard, F., Guerrouache, M., Langlois, V., Kelly, F. J., & France Mandin, C. (2019). Microplastics in air: Are we breathing it in? *Current Opinion in Environmental Science & Health*, 1, 1–5.
- Gilbert, J. M., Reichelt-Brushett, A. J., Bowling, A. C., & Christidis, L. (2016). Plastic ingestion in marine and coastal bird species of southeastern Australia. *Marine Ornithology*, 44, 21–26.
- Glover, C. N., & Hogstrand, C. (2002). In vivo characterisation of intestinal zinc uptake in freshwater rainbow trout. *Journal of Experimental Biology*, 205(1), 141–150.
- Godoy, V., Blázquez, G., Calero, M., Quesada, L., & Martín-Lara, M. A. (2019). The potential of microplastics as carriers of metals. *Environmental Pollution*, 255, 113363.
- Godoy, V., Calero, M., & Blázquez, G. (2019). Physical-chemical characterization of microplastics present in some exfoliating products from Spain. *Marine Pollution Bulletin*, 139(December 2018), 91–99.
- Goss, H., Jaskiel, J., & Rotjan, R. (2018). *Thalassia testudinum* as a potential vector for incorporating microplastics into benthic marine food webs. *Marine Pollution Bulletin*, 135, 1085–1089.
- Gouin, T., Avalos, J., Brunning, I., Brzuska, K., De, J. G., Kaumanns, J., Konong, T., Meyberg, M., Rettinger, K., Schlatter, H., Thomas, J., Van, R. W., & Wolf, T. (2015). Use of micro-plastic beads in cosmetic products in Europe and their estimated emissions to the North Sea environment. *SOFW Journal*, 1–33.
- Govoni, S., Lucchi, L., Missale, C., Memo, M., Spano, P. F., & Trabucchi, M. (1986). Effect of lead exposure on dopaminergic receptors in rat striatum and nucleus accumbens. *Brain Research*, 381(1), 138–142.
- Graham, E. R., & Thompson, J. T. (2009). Deposit- and suspension-feeding sea cucumbers (Echinodermata) ingest plastic fragments. *Journal of Experimental Marine Biology and Ecology*, 368(1), 22–29.
- Grigorakis, S., Mason, S. A., & Drouillard, K. G. (2017). Chemosphere Determination of the gut retention of plastic microbeads and microfibers in gold fish (*Carassius auratus*). *Chemosphere*, 169, 233–238.
- Grosell, M., Blanchard, J., Brix, K. V., & Gerdes, R. (2007). Physiology is pivotal for interactions between salinity and acute copper toxicity to fish and invertebrates. *Aquatic Toxicology*, 84(2), 162–172.



- Guardiola, M., Alvaro, A., Vallvé, J. C., Rosales, R., Solà, R., Girona, J., Serra, N., Duran, P., Esteve, E., Masana, L., & Ribalta, J. (2012). APOA5 gene expression in the human intestinal tissue and its response to in vitro exposure to fatty acid and fibrates. *Nutrition, Metabolism and Cardiovascular Diseases*, 22(9), 756–762.
- Guidelli, E. J., Ramos, A. P., Zaniquelli, M. E. D., & Baffa, O. (2011). Green synthesis of colloidal silver nanoparticles using natural rubber latex extracted from *Hevea brasiliensis*. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 82(1), 140–145.
- Guo, J., Huang, X., Xiang, L., Wang, Y., Li, Y., Li, H., & Cai, Q. (2020). Source, migration and toxicology of microplastics in soil. *Environment International*, 137, 105263.
- Guo, X., & Wang, J. (2019). The chemical behaviors of microplastics in marine environment: A review. *Marine Pollution Bulletin*, 142, 1–14.
- Güven, O., Gökdağ, K., Jovanović, B., & Kıdeys, A. E. (2017). Microplastic litter composition of the Turkish territorial waters of the Mediterranean Sea, and its occurrence in the gastrointestinal tract of fish. *Environmental Pollution*, 223, 286–294.
- Hall, N. M., Berry, K. L. E., Rintoul, L., & Hoogenboom, M. O. (2015). Microplastic ingestion by scleractinian corals. *Marine Biology*, 162(3), 725–732.
- Halle, A. ter, Ladirat, L., Martignac, M., Françoise, A., Boyron, O., & Perez, E. (2017). To what extent are microplastics from the open ocean weathered?\*. *Environmental Pollution*, 227, 167–174.
- Hämer, J., Gutow, L., Köhler, A., & Saborowski, R. (2014). Fate of Microplastics in the Marine Isopod *Idotea emarginata*. *Environmental Science & Technology*, 48(22), 13451–13458.
- Hansen, E., Nilsson, N. H., Lithner, D., & Lassen, C. (2013). *Hazardous substances in plastic materials*. In REACH. Denmark.
- Hayat, N. M., Shamaan, N. A., Sabullah, M. K., Shukor, M. Y., Syed, M. A., Khalid, A., Dahalan, F. A., & Ahmad, S. A. (2016). The use of *Lates calcarifer* as a biomarker for heavy metals detection. *Rendiconti Lincei*, 27(3), 463–472.
- Hemmadi, V. (2016). Metallothionein - A potential biomarker to assess the metal contamination in marine fishes. *International Journal of Bioassay*, 5(4), 4961 – 4973.

- Herzke, D., Anker-Nilssen, T., Nøst, T. H., Götsch, A., Christensen-Dalsgaard, S., Langset, M., Fangel, K., & Koelmans, A. A. (2016). Negligible Impact of Ingested Microplastics on Tissue Concentrations of Persistent Organic Pollutants in Northern Fulmars off Coastal Norway. *Environmental Science and Technology*, 50(4), 1924–1933.
- Hodson, M. E., Duffus-hodson, C., Clark, A., Prendergast-miller, M., & Thorpe, K. L. (2017). Plastic bag derived-microplastics as a vector for metal exposure in terrestrial invertebrates. *Environmental Sciences & Technology*, 51(8), 4714–4721.
- Holmes, Luke A., & Thompson, R. C. (2014). Interactions between trace metals and plastic production pellets under estuarine conditions. *Marine Chemistry*, 167, 25–32.
- Holmes, Luke A, Thompson, R. C., & Turner, A. (2020). In vitro avian bioaccessibility of metals adsorbed to microplastic pellets. *Environmental Pollution*, 261, 114107.
- Holmes, Luke A, Turner, A., & Thompson, R. C. (2012). Adsorption of trace metals to plastic resin pellets in the marine environment. *Environmental Pollution*, 160, 42–48.
- Holmes, Luke Alexander. (2013). *Interactions of Trace Metals with Plastic Production Pellets in the Marine Environment*. Doctor of Philosophy, School of Geography, Earth and Environmental Sciences, University of Plymouth.
- Horn, D., Miller, M., Anderson, S., & Steele, C. (2019). Microplastics are ubiquitous on California beaches and enter the coastal food web through consumption by Pacific mole crabs. *Marine Pollution Bulletin*, 139, 231–237.
- Horton, A. A., Vijver, M. G., Lahive, E., Spurgeon, D. J., Svendsen, C., Heutink, R., Bodegom, P. M. V., & Baas, J. (2018). Ecotoxicology and Environmental Safety Acute toxicity of organic pesticides to *Daphnia magna* is unchanged by co-exposure to polystyrene microplastics. *Ecotoxicology and Environmental Safety*, 166, 26–34.
- Hossain, M. S., Sobhan, F., Uddin, M. N., Sharifuzzaman, S. M., Chowdhury, S. R., Sarker, S., & Chowdhury, M. S. N. (2019). Microplastics in fishes from the Northern Bay of Bengal. *Science of the Total Environment*, 690, 821–830.
- Houde, E. D. (1997). Patterns and trends in larval-stage growth and mortality of teleost fish\*. *Journal of Fish Biology*, 51, 52–83.

- Hu, J.-Q., Yang, S.-Z., Guo, L., Xu, X., Yao, T., & Xie, F. (2017). Microscopic investigation on the adsorption of lubrication oil on microplastics. *Journal of Molecular Liquids*, 227, 351–355.
- Hu, J., Chen, C., Zhu, X., & Wang, X. (2009). Removal of chromium from aqueous solution by using oxidized multiwalled carbon nanotubes. *Journal of Hazardous Materials*, 162(2), 1542–1550.
- Iannilli, V., Di Gennaro, A., Lecce, F., Sighicelli, M., Falconieri, M., Pietrelli, L., Poeta, G & Battisti, C. (2018). Microplastics in *Talitrus saltator* (Crustacea, Amphipoda): new evidence of ingestion from natural contexts. *Environmental Science and Pollution Research*, 25(28), 28725–28729.
- Imhof, H. K., Laforsch, C., Wiesheu, A. C., Schmid, J., Anger, P. M., Niessner, R., & Ivleva, N. P. (2016). Pigments and plastic in limnetic ecosystems: A qualitative and quantitative study on microparticles of different size classes. *Water Research*, 98, 64–74.
- Imhof, H. K., Sigl, R., Brauer, E., Feyl, S., Giesemann, P., Klink, S., Leupolz, K., Löder, M. G. J., Löschel, L. A., Missun, J., Muszynski, S., Ramsperger, A. F. R. M., Schrank, I., Speck, S., Steibl, S., Trotter, B., Winter, I., & Laforsch, C. (2017). Spatial and temporal variation of macro-, meso- and microplastic abundance on a remote coral island of the Maldives, Indian Ocean. *Marine Pollution Bulletin*, 116(1), 340–347.
- Jambeck, J. R., Geyer, R., Wilcox, C., Siegler, T. R., Perryman, M., Andrady, A., Narayan, R., & Law, K. L. (2015). Plastic waste inputs from land into the ocean. *Science*, 347(6223), 768–771.
- Jemec, A., Kunej, U., & Skalar, T. (2018). Screening study of four environmentally relevant microplastic pollutants: Uptake and effects on *Daphnia magna* and *Artemia franciscana*. *Chemosphere*, 208, 522–529.
- Jinhui, S., Sudong, X., Yan, N., Xia, P., Jiahao, Q., & Yongjian, X. (2019). Effects of microplastics and attached heavy metals on growth, immunity, and heavy metal accumulation in the yellow seahorse, *Hippocampus kuda* Bleeker. *Marine Pollution Bulletin*, 149, 110510.
- Joint Group of Experts on, & Protection, the S. A. of M. E. (2019). Guidelines for the monitoring and assessment of plastic litter in the ocean. London, UK.
- Jones, M. N., & Bryan, N. D. (1998). Colloidal properties of humic substances. *Advances in Colloid and Interface Science*, 78(1), 1–48.

- Jovanović, B. (2017). Ingestion of microplastics by fish and its potential consequences from a physical perspective: Potential Consequences of Fish Ingestion of Microplastic Invited Commentary Ingestion of Microplastics. *Integrated Environmental Assessment and Management*, 13(3), 510–515.
- Juliano, C., & Magrini, G. A. (2017). Cosmetic Ingredients as Emerging Pollutants of Environmental and Health Concern. A Mini-Review. *Cosmetic*, 4(11), 1–18.
- Jung, M. R., Horgen, F. D., Orski, S. V, Rodriguez C., V., Beers, K. L., Balazs, G. H., Jones, T. T., Work, T. M., Brignac, K. C., Royer, S-J., Hyrenbach, K. D., Jensen, B., & Lynch, J. M. (2018). Validation of ATR FT-IR to identify polymers of plastic marine debris, including those ingested by marine organisms. *Marine Pollution Bulletin*, 127, 704–716.
- Kalčíková, G., Žgajnar Gotvajn, A., Kladnik, A., & Jemec, A. (2017). Impact of polyethylene microbeads on the floating freshwater plant duckweed *Lemna minor*. *Environmental Pollution*, 230, 1108–1115.
- Kalyoncu, L., Kalyoncu, H., & Arslan, G. (2012). Determination of heavy metals and metals levels in five fish species from Işıklı Dam Lake and Karacaören Dam Lake (Turkey). *Environmental Monitoring and Assessment*, 184(4), 2231–2235.
- Kamaruzzaman, B. Y., Akbar, B., Jalal, K. C. A., & Shahbudin, S. (2010). Accumulation of Metals in the Gills of Tilapia Fingerlings (*Oreochromis niloticus*) from in vitro Toxicology Study. *Journal of Fisheries and Aquatic Science*, 5(6), 503–509.
- Kamaruzzaman, B. Y., Ong, M. C., & Rina, S. Z. (2010). Concentration of Zn, Cu and Pb in some selected marine fishes of the Pahang coastal waters, Malaysia. *American Journal of Applied Sciences*, 7(3), 309–314.
- Kane, I. A., & Clare, M. A. (2019). Dispersion, Accumulation, and the Ultimate Fate of Microplastics in Deep-Marine Environments: A Review and Future Directions. *Frontiers in Earth Science*, Vol. 7, p. 80.
- Karami, A., Golieskardi, A., Keong, C., Romano, N., Bin, Y., & Salamatina, B. (2017). A high-performance protocol for extraction of microplastics in fish. *Science of the Total Environment*, 578, 485–494.
- Karami, A., Groman, D. B., Wilson, S. P., Ismail, P., & Neela, V. K. (2017). Biomarker responses in zebra fish (*Danio rerio*) larvae exposed to pristine low-density polyethylene fragments\*. *Environmental Pollution*, 223, 466–475.

- Karaoğlu, K., & Gül, S. (2020). Characterization of microplastic pollution in tadpoles living in small water-bodies from Rize, the northeast of Turkey. *Chemosphere*, 255, 126915.
- Karapanagioti, H.K., Endo, S., Ogata, Y., & Takada, H. (2011). Diffuse pollution by persistent organic pollutants as measured in plastic pellets sampled from various beaches in Greece. *Marine Pollution Bulletin*, 62(2), 312–317.
- Karapanagioti, H K, & Werner, D. (2019). Sorption of Hydrophobic Organic Compounds to Plastics in the Marine Environment: Sorption and Desorption Kinetics. In *Handbook of Environmental Chemistry* (Vol. 78, pp. 205–219). Department of Chemistry, University of Patras, Patras, Greece: Springer Verlag.
- Karapanagioti, Hrisi K., & Klontza, I. (2008). Testing phenanthrene distribution properties of virgin plastic pellets and plastic eroded pellets found on Lesbos island beaches (Greece). *Marine Environmental Research*, 65(4), 283–290.
- Karbalaei, S., Golieskardi, A., Hamzah, H. B., Abdulwahid, S., Hanachi, P., Walker, T. R., & Karami, A. (2019). Abundance and characteristics of microplastics in commercial marine fish from Malaysia. *Marine Pollution Bulletin*, 148(July), 5–15.
- Kartar, S., Milne, R. A., & Sainsbury, M. (1973). Polystyrene waste in the Severn Estuary. *Marine Pollution Bulletin*, 4(9), 144.
- Khan, F. R., Boyle, D., Chang, E., & Bury, N. R. (2017). Do polyethylene microplastic beads alter the intestinal uptake of Ag in rainbow trout (*Oncorhynchus mykiss*)? Analysis of the MP vector effect using in vitro gut sacs\*. *Environmental Pollution*, 231, 200–206.
- Khan, F. R., Syberg, K., Shashoua, Y., & Bury, N. R. (2015). Influence of polyethylene microplastic beads on the uptake and localization of silver in zebrafish (*Danio rerio*). *Environmental Pollution*, 206, 73–79.
- Kienle, C., Köhler, H.-R., & Gerhardt, A. (2009). Behavioural and developmental toxicity of chlorpyrifos and nickel chloride to zebrafish (*Danio rerio*) embryos and larvae. *Ecotoxicology and Environmental Safety*, 72(6), 1740–1747.
- Kik, K., Bukowska, B., & Sicińska, P. (2020). Polystyrene nanoparticles: Sources, occurrence in the environment, distribution in tissues, accumulation and toxicity to various organisms☆. *Environmental Pollution*, 262, 114297.

- Kim, D., Chae, Y., & An, Y.-J. (2017). Mixture Toxicity of Nickel and Microplastics with Different Functional Groups on *Daphnia magna*. *Environmental Science & Technology*, 51(21), 12852–12858.
- Koelmans, A. A., Besseling, E., Wegner, A., & Foekema, E. M. (2013). Plastic as a Carrier of POPs to Aquatic Organisms: A Model Analysis. *Environmental Science & Technology*, 47(14), 7812–7820.
- Kögel, T., Bjørøy, Ø., Toto, B., Marcel, A., & Sanden, M. (2020). Science of the Total Environment Micro- and nanoplastic toxicity on aquatic life: Determining factors. *Science of the Total Environment*, 709(5817), 136050.
- Kolandhasamy, P., Su, L., Li, J., Qu, X., Jabeen, K., & Shi, H. (2018). Adherence of microplastics to soft tissue of mussels: A novel way to uptake microplastics beyond ingestion ☆. *Science of the Total Environment*, 610–611, 635–640.
- Kontrick, A. V. (2018). Microplastics and Human Health: Our Great Future to Think About Now. *Journal of Medical Toxicology*, 14(2), 117–119.
- Kroon, F. J., Berry, K. L. E., Brinkman, D. L., Kookana, R., Leusch, F. D. L., Melvin, S. D., Neale, P. A., Negri, A. P., Puotinen, M., Tsang, J. J., Merwe, J. P. V. D., & Williams, M. (2020). Sources, presence and potential effects of contaminants of emerging concern in the marine environments of the Great Barrier Reef and Torres Strait, Australia. *Science of the Total Environment*, 719, 135140.
- Kühn, S., Franeker, J. A. V., Donoghue, A. M. O., Swiers, A., Starkenburg, M., Werven, B. V., Foekema, E., Hermsen, E., Egelkraut-holtus, M., & Lindeboom, H. (2019). Details of plastic ingestion and fibre contamination in North Sea. *Environmental Pollution*, 257, 113569.
- Laing, G. Du, Rinklebe, J., Vandecasteele, B., Meers, E., & Tack, F. M. G. (2008). Trace metal behaviour in estuarine and riverine floodplain soils and sediments: A review. *Science of the Total Environment*, 407(13), 3972–3985.
- Law, K. L. (2017). Plastics in the Marine Environment. *Annual Review of Marine Science*, 9(1), 205–229.
- Lebreton, L. C. M., Zwet, J. Van Der, Damsteeg, J., Slat, B., Andrady, A., & Reisser, J. (2017). River plastic emissions to the world's oceans. *Nature Communications*, 8(1), 15611.

- Lee, H., Joon, W., & Kwon, J. (2014). Sorption capacity of plastic debris for hydrophobic organic chemicals. *Science of the Total Environment*, 470–471, 1545–1552.
- Lee, J., Hong, S., Song, Y. K., Hong, S. H., Jang, Y. C., Jang, M., Heo, N. W., Han, G. M., Lee, M. J., Kang, D., & Shim, W. J. (2013). Relationships among the abundances of plastic debris in different size classes on beaches in South Korea. *Marine Pollution Bulletin*, 77(1–2), 349–354.
- Li, Jia, Zhang, K., & Zhang, H. (2018). Adsorption of antibiotics on microplastics\*. *Environmental Pollution*, 237, 460–467.
- Li, Jiana, Green, C., Reynolds, A., Shi, H., & Rotchell, J. M. (2018). Microplastics in mussels sampled from coastal waters and supermarkets in the United Kingdom\*. *Environmental Pollution*, 241, 35–44.
- Li, J., Lusher, A. L., Rotchell, J. M., Deudero, S., Turra, A., Lise, I., Bråte, N., Sun, C., Hossain, M. S., Li, Q., Kolandhasamy, P., & Shi, H. (2019). Using mussel as a global bioindicator of coastal microplastic. *Environmental Pollution*, 244, 522–533.
- Limousin, G., Gaudet, J. P., Charlet, L., Szenknecta, S., Barthèsa, V., & Krimissa, M. (2007). Sorption isotherms: A review on physical bases, modeling and measurement. *Applied Geochemistry*, 22, 249–275.
- Liu, H., Liu, K., Fu, H., Ji, R., & Qu, X. (2020). Sunlight mediated cadmium release from colored microplastics containing cadmium pigment in aqueous phase\*. *Environmental Pollution*, 263, 114484.
- Llorca, M., Schirinzi, G., Martínez, M., Barceló, D., & Farré, M. (2018). Adsorption of perfluoroalkyl substances on microplastics under environmental conditions. *Environmental Pollution*, 235, 680–691.
- Lopes, C., Raimundo, J., Caetano, M., & Garrido, S. (2020). Microplastic ingestion and diet composition of planktivorous fish. *Limnology and Oceanography Letters*, 5(1), 103–112.
- Lu, K., Qiao, R., An, H., & Zhang, Y. (2018). Influence of microplastics on the accumulation and chronic toxic effects of cadmium in zebrafish (*Danio rerio*). *Chemosphere*, 202, 514–520.

- Lu, Y., Zhang, Y., Deng, Y., Jiang, W., Zhao, Y., Geng, J., Ding, L., & Ren, H. (2016). Uptake and Accumulation of Polystyrene Microplastics in Zebrafish (*Danio rerio*) and Toxic Effects in Liver. *Environmental Science and Technology*, 50(7), 4054–4060.
- Luís, L. G., Ferreira, P., Fonte, E., Oliveira, M., & Guilhermino, L. (2015). Does the presence of microplastics influence the acute toxicity of chromium (VI) to early juveniles of the common goby (*Pomatoschistus microps*)? A study with juveniles from two wild estuarine populations. *Aquatic Toxicology*, 164, 163–174.
- Lusher, A. (2015a). Microplastics in the Marine Environment: Distribution, Interactions and Effects. In M. Bergmann, L. Gutow, & M. Klages (Eds.), *Marine Anthropogenic Litter* (pp. 245–307). Cham: Springer International Publishing.
- Lusher, A. (2015b). Microplastics in the Marine Environment: Distribution, Interactions and Effects. In M. Bergmann, L. Gutow, & M. Klages (Eds.), *Marine Anthropogenic Litter* (pp. 1–447). Galway, Ireland: Marine and Freshwater Research Centre, Galway-Mayo Institute of Technology.
- Lusher, A. L., McHugh, M., & Thompson, R. C. (2013). Occurrence of microplastics in the gastrointestinal tract of pelagic and demersal fish from the English Channel. *Marine Pollution Bulletin*, 67(1–2), 94–99.
- Lv, W., Zhou, W., Lu, S., Huang, W., Yuan, Q., Tian, M., Lv, W., & He, D. (2019). Microplastic pollution in rice- fish co-culture system: A report of three farmland stations in Shanghai, China. *Science of the Total Environment*, 652, 1209–1218.
- Ma, P., Wei Wang, M., Liu, H., Feng Chen, Y., & Xia, J. (2019). Research on ecotoxicology of microplastics on freshwater aquatic organisms. *Environmental Pollutants and Bioavailability*, 31(1), 131–137.
- MacMillan, A. (2018). *There's plastic in your poop: study warns microplastics may be hurting us all*. Retrieved from <https://www.health.com/home/microplastics-human-poop>



- Magni, S., Gagné, F., André, C., Della, C., Auclair, J., Hanana, H., Carla, C., Bonasoro, F., & Binelli, A. (2018). Evaluation of uptake and chronic toxicity of virgin polystyrene microbeads in freshwater zebra mussel *Dreissena polymorpha* (Mollusca: Bivalvia). *Science of the Total Environment*, 631–632, 778–788.
- Maharajan, A., Kitto, M. R., Paruruckumani, P. S., & Ganapiriya, V. (2016). Histopathology biomarker responses in Asian sea bass, *Lates calcarifer* (Bloch) exposed to copper. *The Journal of Basic & Applied Zoology*, 77, 21–30.
- Mai, L., Bao, L.-J., Shi, L., Wong, C. S., & Zeng, E. Y. (2018). A review of methods for measuring microplastics in aquatic environments. *Environmental Science and Pollution Research*, 25(12), 11319–11332.
- Mani, T., Blarer, P., Storck, F. R., Pittroff, M., Wernicke, T., & Burkhardt-holm, P. (2019). Repeated detection of polystyrene microbeads in the Lower Rhine. *Environmental Pollution*, 245, 634–641.
- Mao, R., Lang, M., Yu, X., Wu, R., Yang, X., & Guo, X. (2020). Aging mechanism of microplastics with UV irradiation and its effects on the adsorption of heavy metals. *Journal of Hazardous Materials*, 393, 122515.
- Mao, Z., Cao, Y., Jin, F., Cong, Y., Wang, L., & Zhang, W. (2019). Effects of ingested polystyrene microplastics on brine shrimp, *Artemia*. *Environmental Pollution*, 244, 715–722.
- Markovic, G. (2007). A Contribution to Data on Chub (*Leuciscus cephalus* L., Cyprinidae, PISCES) Diet in River Conditions. *Acta Agriculturae Serbica*, XII(24), 13–18.
- Marnane, R. N., Elliot, D. A., & Coulson, S. A. (2006). A Pilot Study to Determine the Background Population of Foreign Fibre Groups on a cotton/polyester T-shirt. *Science & Justice*, 46(4), 215–220.
- Massos, A., & Turner, A. (2017). Cadmium, lead and bromine in beached microplastics\*. *Environmental Pollution*, 227, 139–145.
- Mathew, G. (2009). Taxonomy, identification and biology of Seabass (*Lates calcarifer*). *Central Marine Fisheries Research Institute*, (1603), 38–43.

- Mathieu-denoncourt, J., Wallace, S. J., Solla, S. R. De, & Langlois, V. S. (2015). General and Comparative Endocrinology Plasticizer endocrine disruption: Highlighting developmental and reproductive effects in mammals and non-mammalian aquatic species. *General and Comparative Endocrinology*, 219, 74–88.
- Mato, Y., Isobe, T., Takada, H., Kanehiro, H., Ohtake, C., & Kaminuma, T. (2001). Plastic Resin Pellets as a Transport Medium for Toxic Chemicals in the Marine Environment. *Environmental Science & Technology*, 35(2), 318–324.
- Mattsson, K., Ekvall, M., Hansson, L., Linse, S., Malmendal, A., & Cedervall, T. (2015). Altered Behavior, Physiology, and Metabolism in Fish Exposed to Polystyrene Nanoparticles. *Environmental Science & Technology*, 49(1), 553–561.
- Mattsson, K., Jovic, S., Doverbratt, I., & Hansson, L.-A. (2018). Chapter 13 - Nanoplastics in the Aquatic Environment. In E. Zeng (Ed.), *Microplastic Contamination in Aquatic Environments* (pp. 379–399). Elsevier.
- Mattsson, K., Johnson, E. V, Malmendal, A., & Linse, S. (2017). Brain damage and behavioural disorders in fish induced by plastic nanoparticles delivered through the food chain. *Scientific Reports*, 7, 11452.
- Mazurais, D., Ernande, B., Quazuguel, P., Severe, A., Huelvan, C., Madec, L., Mouchel, O., Soudant, P., Robbens, J., Huvet, A., & Zambonino-Infante, J. (2015). Evaluation of the impact of polyethylene microbeads ingestion in European sea bass (*Dicentrarchus labrax*) larvae. *Marine Environmental Research*, 112, 78–85.
- McGrath, M. (2018). *Plastic microbead ban: what impact will it have?* Retrieved from <https://www.bbc.com/news/science-environment-42621388>
- Menéndez-Pedriza, A., & Jaumot, J. (2020). Interaction of environmental pollutants with microplastics: A critical review of sorption factors, bioaccumulation and ecotoxicological effects. *Toxic*, 8(2), 40.
- Miranda, T., Vieira, L. R., & Guilhermino, L. (2019). Neurotoxicity, Behavior, and Lethal Effects of Cadmium , Microplastics, and Their Mixtures on *Pomatoschistus microps* Juveniles from Two Wild Populations Exposed under Laboratory Conditions — Implications to Environmental and Human Risk Assessment. *International Journal of Environmental Research and Public Health*, 16, 2857.

- Miretzky, P., & Cirelli, A. F. (2010). Cr(VI) and Cr(III) removal from aqueous solution by raw and modified lignocellulosic materials: A review. *Journal of Hazardous Materials*, 180(1), 1–19.
- Moira, J. W. D., Peter, G., & Ross, P. S. (2015). Ingestion of Microplastics by Zooplankton in the Northeast Pacific Ocean. *Archives of Environmental Contamination and Toxicology*, 69(3), 320–330.
- Morán, P., Cal, L., Cobelo-García, A., Almécija, C., Caballero, P., & Garcia de Leaniz, C. (2018). Historical legacies of river pollution reconstructed from fish scales. *Environmental Pollution*, 234, 253–259.
- Morgana, S., Ghigliotti, L., Est, N., Stifanese, R., Wieckzorek, A., Doyle, T., Christiansen, J. S., Faimali, M., & Garaventa, F. (2018). Microplastics in the Arctic: A case study with sub-surface water and fish samples off Northeast Greenland\*. *Environmental Pollution*, 242, 1078–1086.
- Munier, B., & Bendell, L. I. (2018). Macro and micro plastics sorb and desorb metals and act as a point source of trace metals to coastal ecosystems. *PLoS ONE*, 13(2), e0191759.
- Murphy, F., Russell, M., Ewins, C., & Quinn, B. (2017). The uptake of macroplastic & microplastic by demersal & pelagic fish in the Northeast Atlantic around Scotland. *Marine Pollution Bulletin*, 122(1–2), 353–359.
- Murray, F., & Cowie, P. R. (2011). Plastic contamination in the decapod crustacean *Nephrops norvegicus* (Linnaeus, 1758). *Marine Pollution Bulletin*, 62(6), 1207–1217.
- Nadal, M. A., Alomar, C., & Deudero, S. (2016). High levels of microplastic ingestion by the semipelagic fish bogue *Boops boops* (L.) around the Balearic Islands. *Environmental Pollution*, 214, 517–523.
- Nakashima, E., Isobe, A., Kako, S., Itai, T., & Takahashi, S. (2012). Quantification of Toxic Metals Derived from Macroplastic Litter on Ookushi Beach, Japan. *Environmental Science & Technology*, 46(18), 10099–10105.
- Nan, B., Su, L., Kellar, C., Craig, N. J., Keough, M. J., & Pettigrove, V. (2020). Identification of microplastics in surface water and Australian freshwater shrimp *Paratya australiensis* in Victoria, Australia. *Environmental Pollution*, 259, 113865.

- Napper, I. E., Bakir, A., Rowland, S. J., & Thompson, R. C. (2015). Characterisation, quantity and sorptive properties of microplastics extracted from cosmetics. *Marine Pollution Bulletin*, 99(1–2), 178–185.
- Naser, H. A. (2013). Assessment and management of heavy metal pollution in the marine environment of the Arabian Gulf: A review. *Marine Pollution Bulletin*, 72(1), 6–13.
- Nasyitah, N., Ahmad, A., Khairul, M., Ley, L., & Kyoung-woong, K. (2018). Bioaccumulation of heavy metals in maricultured fish, *Lates calcarifer* (Barramudi), *Lutjanus campechanus* (red snapper) and *Lutjanus griseus* (grey snapper). *Chemosphere*, 197, 318–324.
- Nelson, Sellers, D., Mackenzie, K., Weinberg, S., & Nadine. (2019). Microbeads—a Case Study in How Public Outrage Fueled the Emergence of New Regulations. *Current Pollution Reports*, 5, 172 – 179.
- Ngoc, N., Poirier, L., Tuan, Q., Lagarde, F., & Zalouk-vergnoux, A. (2018). Factors in fluencing the microplastic contamination of bivalves from the French Atlantic coast: Location, season and/or mode of life? *Marine Pollution Bulletin*, 129(2), 664–674.
- Nicolau, L., Marçalo, A., Ferreira, M., Sá, S., Vingada, J., & Eira, C. (2016). Ingestion of marine litter by loggerhead sea turtles, *Caretta caretta*, in Portuguese continental waters. *Marine Pollution Bulletin*, 103(1–2), 179–185.
- Nriagu, J. O. (1988). Production and Uses of Chromium. In J. O. Nriagu & E. Nieboer (Eds.), *Chromium in the Natural and Human Environments*. Wiley, New York.
- Nur, T. K. A. T., Hing, L. S., Sim, S. F., Pradit, S., Ahmad, A., & Ong, M. C. (2020). Comparative study of raw and cooked farmed sea bass (*Lates calcarifer*) in relation to metal content and its estimated human health risk. *Marine Pollution Bulletin*, 153, 111009.
- Obbard, R. W., Sadri, S., Wong, Y. Q., Khitun, A. A., Baker, I., & Thompson, R. C. (2014). Global warming releases microplastic legacy frozen in Arctic Sea ice. *Earth's Future*, 2(6), 315–320.
- Oliveira, M., Ribeiro, A., Hylland, K., & Guilhermino, L. (2013). Single and combined effects of microplastics and pyrene on juveniles (0+ group) of the common goby *Pomatoschistus microps* (Teleostei, Gobiidae). *Ecological Indicators*, 34, 641–647.

- Oliveira, P., Gabriel, L., Barboza, A., Branco, V., Figueiredo, N., Carvalho, C., & Guilhermino, L. (2018). Effects of microplastics and mercury in the freshwater bivalve *Corbicula fluminea* (Müller, 1774 ): Filtration rate, biochemical biomarkers and mercury bioconcentration. *Ecotoxicology and Environmental Safety*, 164, 155–163.
- Ory, N., Chagnon, C., Felix, F., Fernández, C., Lia, J., Gallardo, C., Garcés, O., Henostroza, A., Laaz, E., Mizraji, R., Mojica, H., Murillo, V., Ossa, L., Preciado, M., Sobral, P., Urbina, M. A., & Thiel, M. (2018). Low prevalence of microplastic contamination in planktivorous fish species from the southeast Pacific Ocean. *Marine Pollution Bulletin*, 127, 211–216.
- Paruruckumani, P. S., Maha Rajan, A., Ganapiriya, V., & Kumarasamy, P. (2015). Bioaccumulation and ultrastructural alterations of gill and liver in Asian sea bass, *Lates calcarifer* (Bloch) in sublethal copper exposure. *Aquat. Living Resour.*, 28(1), 33–44.
- Patel, M. M., Goyal, B. R., Bhadada, S. V., Bhatt, J. S., & Amin, A. F. (2009). Getting into the Brain. *CNS Drugs*, 23(1), 35–58.
- Paterson, H. (2019). Plastic habits – an overview for the collection “Plastics and Sustainable Earth.” *Sustainable Earth*, 2(10), 6–13.
- Paul-Pont, I., Lacroix, C., González Fernández, C., Hégaret, H., Lambert, C., Le Goïc, N., Frère, L., Cassone, A. L., Sussarellu, R., Fabioux, C., Guyomarch, J., Albentosa, M., Huvet, A., & Soudant, P. (2016). Exposure of marine mussels *Mytilus* spp. to polystyrene microplastics: Toxicity and influence on fluoranthene bioaccumulation. *Environmental Pollution*, 216, 724–737.
- Pedà, C., Caccamo, L., Fossi, M. C., Gai, F., Andaloro, F., Genovese, L., Perdichizzi, A., Romeo, T., & Maricchiolo, G. (2016). Intestinal alterations in European sea bass *Dicentrarchus labrax* (Linnaeus, 1758) exposed to microplastics: Preliminary results. *Environmental Pollution*, 212, 251–256.
- Peng, G., Xu, P., Zhu, B., Bai, M., & Li, D. (2018). Microplastics in freshwater river sediments in Shanghai, China: A case study of risk assessment in mega-cities\*. *Environmental Pollution*, 234, 448–456.
- Peng, L., Fu, D., Qi, H., Lan, C. Q., Yu, H., & Ge, C. (2020). Micro- and nano-plastics in marine environment: Source , distribution and threats — A review. *Science of the Total Environment*, 698, 134254.

- Peng, X., Chen, M., Chen, S., Dasgupta, S., Xu, H., Ta, K., Du, M., Li, J., Guo, Z., & Bai, S. (2018). Microplastics contaminate the deepest part of the world's ocean. *Geochemical Perspectives Letters*, 9, 1–5.
- Perez-Venegas, D. J., Toro-Valdivieso, C., Ayala, F., Brito, B., Iturra, L., Arriagada, M., Seguel, M., Barrios, C., Sepúlveda, M., Oliva, D., Cárdenas-Alayza, S., Urbina, M. A., Jorquera, A., Castro-Nallar, E., & Galbán-Malagón, C. (2020). Monitoring the occurrence of microplastic ingestion in Otariids along the Peruvian and Chilean coasts. *Marine Pollution Bulletin*, 153, 110966.
- Peter Munk, & Nielsen, J. (2007). *Eggs and Larvae of North Sea Fishes*. Frederiksberg, Denmark: Biofolia.
- Peters, C. A., & Bratton, S. P. (2016). Urbanization is a major influence on microplastic ingestion by sunfish in the Brazos River Basin, Central Texas, USA. *Environmental Pollution*, 210, 380–387.
- Peters, C. A., Hendrickson, E., Minor, E. C., Schreiner, K., Halbur, J., & Bratton, S. P. (2018). Pyr-GC / MS analysis of microplastics extracted from the stomach content of benthivore fish from the Texas Gulf Coast. *Marine Pollution Bulletin*, 137, 91–95.
- Peters, C. A., Thomas, P. A., Rieper, K. B., & Bratton, S. P. (2017). Foraging preferences influence microplastic ingestion by six marine fish species from the Texas Gulf Coast. *Marine Pollution Bulletin*, 124(1), 82–88.
- Pettipas, S., Bernier, M., & Walker, T. R. (2016). A Canadian policy framework to mitigate plastic marine pollution. *Marine Policy*, 68, 117–122.
- Phillips, M. B., & Bonner, T. H. (2015). Occurrence and amount of microplastic ingested by fishes in watersheds of the Gulf of Mexico. *Marine Pollution Bulletin*, 100(1), 264–269.
- Phuong, N. N., Zalouk-Vergnoux, A., Poirier, L., Kamari, A., Châtel, A., Mouneyrac, C., & Lagarde, F. (2016). Is there any consistency between the microplastics found in the field and those used in laboratory experiments? *Environmental Pollution*, 211, 111–123.
- Pittura, L., Avio, C. G., Giuliani, M. E., D'Errico, G., Keiter, S. H., Cormier, B., Gorbi, S., & Regoli, F. (2018). Microplastics as vehicles of environmental PAHs to marine organisms: Combined chemical and physical hazards to the mediterranean mussels, *Mytilus galloprovincialis*. *Frontiers in Marine Science*, 5, 103.

- Plastics Europe. (2018). *Plastics – the Facts: An analysis of European plastics production, demand and waste data*. Association of Plastic Manufacturers & European Association of Plastic Recycling and Recovery Organisation, Belgium.
- Possatto, F. E., Barletta, M., Costa, M. F., Ivar do Sul, J. A., & Dantas, D. V. (2011). Plastic debris ingestion by marine catfish: An unexpected fisheries impact. *Marine Pollution Bulletin*, 62(5), 1098–1102.
- Praveena, S. M., Shaifuddin, S. N. M., & Akizuki, S. (2018). Exploration of microplastics from personal care and cosmetic products and its estimated emissions to marine environment: An evidence from Malaysia. *Marine Pollution Bulletin*, 136, 135–140.
- Pritchard, G. (1998). An A-Z reference. In G. Pritchard (Ed.), *Plastics Additives* (pp. XX, 633). Dordrecht: Springer Netherlands.
- Procter, J., Hopkins, F. E., Fileman, E. S., & Lindeque, P. K. (2019). Smells good enough to eat: Dimethyl sulfide (DMS) enhances copepod ingestion of microplastics. *Marine Pollution Bulletin*, 138, 1–6.
- Provencher, J. F., Vermaire, J. C., Avery-Gomm, S., Braune, B. M., & Mallory, M. L. (2018). Garbage in guano? Microplastic debris found in faecal precursors of seabirds known to ingest plastics. *Science of The Total Environment*, 644, 1477–1484.
- Prunier, J., Maurice, L., Perez, E., Gigault, J., Pierson Wickmann, A.-C., Davranche, M., & Halle, A. ter. (2019). Trace metals in polyethylene debris from the North Atlantic subtropical gyre\*. *Environmental Pollution*, 245, 371–379.
- Qiao, R., Lu, K., Deng, Y., Ren, H., & Zhang, Y. (2019). Combined effects of polystyrene microplastics and natural organic matter on the accumulation and toxicity of copper in zebrafish. *Science of the Total Environment*, 682, 128–137.
- Ramos, J. A. A., Barletta, M., & Costa, M. F. (2012). Ingestion of nylon threads by gerreidae while using a tropical estuary as foraging grounds. *Aquatic Biology*, 17(1), 29–34.
- Raval, N. P., Shah, P. U., & Shah, N. K. (2016). Adsorptive removal of nickel(II) ions from aqueous environment: A review. *Journal of Environmental Management*, 179, 1–20.

- Rehse, S., Kloas, W., & Zarfl, C. (2016). Short-term exposure with high concentrations of pristine microplastic particles leads to immobilisation of *Daphnia magna*. *Chemosphere*, 153, 91–99.
- Renzi, M., Bla, A., Bernardi, G., & Russo, G. F. (2018). Plastic litter transfer from sediments towards marine trophic webs: A case study on holothurians. *Marine Pollution Bulletin*, 135, 376–385.
- Rezania, S., Park, J., Fadhil, M., Mat, S., Talaiekhosani, A., Kumar, K., & Kamyab, H. (2018). Microplastics pollution in different aquatic environments and biota: A review of recent studies. *Marine Pollution Bulletin*, 133, 191–208.
- Ribeiro, F., Garcia, A. R., Pereira, B. P., Fonseca, M., Mestre, N. C., Fonseca, T. G., Mestre, N. C., Fonseca, T. G., Ilharco, L. M., & João, M. (2017). Microplastics effects in *Scrobicularia plana*. *Marine Pollution Bulletin*, 122, 379–391.
- Rillig, M. C., Ingraffia, R., & De Souza Machado, A. A. (2017). Microplastic incorporation into soil in agroecosystems. *Frontiers in Plant Science*, 8, 1805.
- Rios Mendoza, L. M., & Jones, P. R. (2015). Characterisation of microplastics and toxic chemicals extracted from microplastic samples from the North Pacific Gyre. *Environmental Chemistry*, 12(5), 611–617.
- Rist, S., Baun, A., & Hartmann, N. B. (2017). Ingestion of micro- and nanoplastics in *Daphnia magna* – Quantification of body burdens and assessment of feeding rates and reproduction. *Environmental Pollution*, 228, 398–407.
- Rocha-Santos, T., & Duarte, A. C. (2015). A critical overview of the analytical approaches to the occurrence, the fate and the behavior of microplastics in the environment. *TrAC Trends in Analytical Chemistry*, 65, 47–53.
- Rochman, C. M., Hentschel, B. T., & Teh, S. J. (2014). Long-Term Sorption of Metals Is Similar among Plastic Types: Implications for Plastic Debris in Aquatic Environments. *PLoS ONE*, 9(1), 1–10.
- Rochman, C. M., Hoh, E., Kurobe, T., & Teh, S. J. (2013). Ingested plastic transfers hazardous chemicals to fish and induces hepatic stress. *Scientific Report*, 3, 3263.
- Rochman, C. M., Kross, S. M., Armstrong, J. B., Bogan, M. T., Darling, E. S., Green, S. J., Smyth, A. R., & Veríssimo, D. (2015). Scientific Evidence Supports a Ban on Microbeads. *Environmental Science & Technology*, 49(18), 10759–10761.



- Rochman, C. M., Kurobe, T., Flores, I., & Teh, S. J. (2014). Early warning signs of endocrine disruption in adult fish from the ingestion of polyethylene with and without sorbed chemical pollutants from the marine environment. *Science of The Total Environment*, 493, 656–661.
- Rochman, C. M., Tahir, A., Williams, S. L., Baxa, D. V, Lam, R., Miller, J. T., Teh, F-C., Werorilangi, S., & Teh, S. J. (2015). Anthropogenic debris in seafood: Plastic debris and fibers from textiles in fish and bivalves sold for human consumption. *Scientific Reports*, 5, 14340.
- Rodrigues, S. M., Almeida, C. M. R., Silva, D., Cunha, J., Antunes, C., Freitas, V., & Ramos, S. (2019). Science of the Total Environment Microplastic contamination in an urban estuary: Abundance and distribution of microplastics and fish larvae in the Douro estuary. *Science of the Total Environment*, 659, 1071–1081.
- Roesijadi, G. (1996). Metallothionein and its role in toxic metal regulation. *Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology*, 111(2), 117–123.
- Romeo, T., Pietro, B., Pedà, C., Consoli, P., Andaloro, F., & Fossi, M. C. (2015). First evidence of presence of plastic debris in stomach of large pelagic fish in the Mediterranean Sea. *Marine Pollution Bulletin*, 95(1), 358–361.
- Rummel, C. D., Löder, M. G. J., Fricke, N. F., Lang, T., Griebeler, E., Janke, M., & Gerdts, G. (2016). Plastic ingestion by pelagic and demersal fish from the North Sea and Baltic Sea. *Marine Pollution Bulletin*, 102(1), 134–141.
- Sá, L. C. de, Luís, L. G., & Guilhermino, L. (2015). Effects of microplastics on juveniles of the common goby (*Pomatoschistus microps*): Confusion with prey, reduction of the predatory performance and efficiency , and possible influence of developmental conditions. *Environmental Pollution*, 196, 359–362.
- Sá, L. C. de, Oliveira, M., Ribeiro, F., Lopes, T., & Norman, M. (2018). Studies of the effects of microplastics on aquatic organisms: What do we know and where should we focus our efforts in the future? *Science of the Total Environment*, 645, 1029–1039.
- Sanchez, W., Bender, C., & Porcher, J.-M. (2014). Wild gudgeons (*Gobio gobio*) from French rivers are contaminated by microplastics: Preliminary study and first evidence. *Environmental Research*, 128, 98–100.

- Sankhla, M. S., Kumari, M., Nandan, M., Kumar, R., & Agrawal, P. (2016). Heavy Metals Contamination in Water and Their Hazardous Effect on Human Health- A Review. *International Journal of Current Microbiology and Applied Sciences*, 5(10), 759 – 766.
- Santana, M. F. M., Moreira, F. T., & Turra, A. (2017). Trophic transference of microplastics under a low exposure scenario: Insights on the likelihood of particle cascading along marine food-webs. *Marine Pollution Bulletin*, 121, 154–159.
- Sarijan, S., Azman, S., Ismid, M., & Said, M. (2019). *Environment Asia*. 12(3), 75–84.
- Satapathy, S., & Panda, C. R. (2017). Toxic metal ion in seafood: Meta-analysis of human carcinogenic and non-carcinogenic threat assessment, a geomedical study from Dhamra and Puri, Odisha. *Human and Ecological Risk Assessment: An International Journal*, 23(4), 864–878.
- Schnurr, R. E. J., Alboiu, V., Chaudhary, M., Corbett, R. A., Quanz, M. E., Sankar, K., Srain, H. S., Thavarajah, V., Xanthos, D., & Walker, T. R. (2018). Reducing marine pollution from single-use plastics (SUPs): A review. *Marine Pollution Bulletin*, 137, 157–171.
- Scopetani, C., Cincinelli, A., Martellini, T., Lombardini, E., Cio, A., Fortunati, A., Pasquali, V., Ciattini, S., & Ugolini, A. (2018). Ingested microplastic as a two-way transporter for PBDEs in *Talitrus saltator*. *Environmental Research*, 167, 411–417.
- Sellers, K. (2015). *Product stewardship: life cycle analysis and the environment*. Boca Raton, Florida: CRC Press Taylor & Francis Group.
- Setälä, O., Fleming-Lehtinen, V., & Finnish, M. L. (2013). Ingestion and transfer of microplastics in the planktonic food web. *Environmental Pollution*, 185, 77–83.
- Shahid, M., Shamshad, S., Rafiq, M., Khalid, S., Bibi, I., Niazi, N. K., Dumat, C., & Rashid, M. I. (2017). Chromium speciation, bioavailability, uptake, toxicity and detoxification in soil-plant system: A review. *Chemosphere*, 178, 513–533.
- Simate, G. S., Maledi, N., Ochieng, A., Ndlovu, S., Zhang, J., & Walubita, L. F. (2016). Coal-based adsorbents for water and wastewater treatment. *Journal of Environmental Chemical Engineering*, 4(2), 2291–2312.

- Sinha, S., Saxena, R., & Singh, S. (2005). Chromium induced lipid peroxidation in the plants of *Pistia stratiotes* L.: role of antioxidants and antioxidant enzymes. *Chemosphere*, 58(5), 595–604.
- Sleight, V. A., Bakir, A., Thompson, R. C., & Henry, T. B. (2017). Assessment of microplastic-sorbed contaminant bioavailability through analysis of biomarker gene expression in larval zebrafish. *Marine Pollution Bulletin*, 116(1–2), 291–297.
- Smith, L. E. (2018). Plastic ingestion by *Scyliorhinus canicula* trawl captured in the North Sea. *Marine Pollution Bulletin*, 130, 6–7.
- Smith, M., Love, D. C., Rochman, C. M., & Neff, R. A. (2018). Microplastics in Seafood and the Implications for Human Health. *Current Environment Health Report*, 5(3), 375–386.
- So, W. K., Chan, K., & Not, C. (2018). Abundance of plastic microbeads in Hong Kong coastal water. *Marine Pollution Bulletin*, 133, 500–505.
- Steer, M., Cole, M., Thompson, R. C., & Lindeque, P. K. (2017). Microplastic ingestion in fish larvae in the western English Channel\*. *Environmental Pollution*, 226, 250–259.
- Su, L., Deng, H., Li, B., Chen, Q., Pettigrove, V., Wu, C., & Shi, H. (2019). The Occurrence of Microplastic in Specific Organs in Commercially Caught Fishes from Coast and Estuary Area of East China. *Journal of Hazardous Materials*, 365, 716–724.
- Sun, X., Chen, B., Li, Q., Liu, N., Xia, B., Zhu, L., & Qu, K. (2018). Toxicities of polystyrene nano- and microplastics toward marine bacterium *Halomonas alkaliphila*. *Science of the Total Environment*, 642, 1378–1385.
- Syberg, K., Khan, F. R., Selck, H., Palmqvist, A., Banta, G. T., Daley, J., Sano, L., & B. Duhaimek, M. (2015). Microplastics: Addressing Ecological Risk Through Lessons Learned. *Environmental Toxicology and Chemistry*, 34(5), 945–953.
- Tagg, A. S., & Ivar, J. A. (2019). Is this your glitter? An overlooked but potentially environmentally-valuable microplastic. *Marine Pollution Bulletin*, 146, 50–53.
- Talvitie, J., Mikola, A., Setälä, O., Heinonen, M., & Koistinen, A. (2017). How well is microlitter purified from wastewater? - A detailed study on the stepwise removal of microlitter in a tertiary level wastewater treatment plant. *Water Research*, 109, 164–172.

- Tchounwou, P. B., Yedjou, C. G., Patlolla, A. K., & Sutton, D. J. (2012). Heavy Metals Toxicity and the Environment. *Experientia Supplementum*, 101, 133–164.
- Tekin-Özan, S., & Aktan, N. (2012). Relationship of Heavy Metals in Water, Sediment and Tissues with Total Length, Weight and Seasons of *Cyprinus carpio* L., 1758 From Jşikli Lake (Turkey). *Pakistan Journal of Zoology*, 44, 1405–1416.
- Teuten, E. L., Saquing, J. M., Knappe, D. R. U., Barlaz, M. A., Jonsson, S., Bjorn, A., Rowland, S. J., Thompson, R. C., Galloway, T. S., Yamashita, R., Ochi, D., Watanuki, Y., Moore, C., Viet, P. H., Tana, T. S., Prudente, M., Boonyatumanond, R., Zakaria, M. P., Akkhavong, K., Ogata, Y., Hirai, H., Iwasa, S., Mizukawa, K., Hagino, Y., Imamura, A., Saha, M., & Takada, H. (2009). Transport and release of chemicals from plastics to the environment and to wildlife. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 364(1526), 2027–2045.
- Thanomsit, C., Nanuam, J., Prasatkaew, W., Meemon, P., & Nanthanawat, P. (2016). Histological Alterations in Asian seabass (*Lates calcarifer*) during Exposed to Non-Essential and Essential Elements. *Environment Asia*, 9(2), 72–79.
- Thiel, M., Luna-jorquera, G., Álvarez-varas, R., & Gallardo, C. (2018). Impacts of Marine Plastic Pollution From Continental Coasts to Subtropical Gyres — Fish, Seabirds, and Other Vertebrates in the SE Pacific. *Frontiers in Marine Science*, 5, 238.
- Thomposon, R. C. (2004). Lost at Sea: Where Is All the Plastic? *Science*, 304(5672), 838–838.
- Thompson, R. C. (2015). Microplastics in the Marine Environment: Sources, Consequences and Solutions. In M. Bergmann, L. Gutow, & M. Klages (Eds.), *Marine Anthropogenic Litter* (pp. 185–200). Springer International Publishing.
- Thompson, R. C., Swan, S. H., Moore, C. J., & vom Saal, F. S. (2009). Our plastic age. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 364(1526), 1973–1976.
- Tien, C. J., & Chen, C. S. (2013). Patterns of metal accumulation by natural river biofilms during their growth and seasonal succession. *Archives of Environmental Contamination and Toxicology*, 64(4), 605–616.
- Turner, A., & Holmes, L. A. (2015). Adsorption of trace metals by microplastic pellets in fresh water. *Environmental Chemistry*, 12(5), 600–610.

- Turner, A., & Millward, G. E. (2002). Suspended Particles: Their Role in Estuarine Biogeochemical Cycles. *Estuarine, Coastal and Shelf Science*, 55(6), 857–883.
- Turner, Andrew. (2016). Heavy metals, metalloids and other hazardous elements in marine plastic litter. *Marine Pollution Bulletin*, 111(1–2), 136–142.
- Turner, Andrew, Crussell, M., Millward, G. E., Cobelo-garcia, A., & Fisher, A. S. (2006). Adsorption Kinetics of Platinum Group Elements in River Water. *Environmental Science & Technology*, 40(5), 1524–1531.
- Van Cauwenberghe, L., Claessens, M., Vandegehuchte, M. B., & Janssen, C. R. (2015). Microplastics are taken up by mussels (*Mytilus edulis*) and lugworms (*Arenicola marina*) living in natural habitats. *Environmental Pollution*, 199, 10–17.
- Van Cauwenberghe, L., Vanreusel, A., Mees, J., & Janssen, C. R. (2013). Microplastic pollution in deep-sea sediments. *Environmental Pollution*, 182, 495–499.
- van Franeker, J. A., Blaize, C., Danielsen, J., Fairclough, K., Gollan, J., Guse, N., Hansen, P-L., Heubeck, M., Jensen, J-K., Le Guillou, G., Olsen, B., Olsen, K-O., Pedersen, J., Stienen, E. W. M., & Turner, D. M. (2011). Monitoring plastic ingestion by the northern fulmar *Fulmarus glacialis* in the North Sea. *Environmental Pollution*, 159(10), 2609–2615.
- Vandermeersch, G., Lourenço, H. M., Alvarez-Muñoz, D., Cunha, S., Diogène, J., Cano-Sancho, G., Sloth, J. J., Kwadijk, C., Barcelo, D., Allegaert, W., Bekaert, K., Fernandes, J. O., Marques, A., & Robbens, J. (2015). Environmental contaminants of emerging concern in seafood – European database on contaminant levels. *Environmental Research*, 143, 29–45.
- Vedolin, M. C., Teophilo, C. Y. S., Turra, A., & Figueira, R. C. L. (2018). Spatial variability in the concentrations of metals in beached microplastics. *Marine Pollution Bulletin*, 129(2), 487–493.
- Velez, J. F. M., Shashoua, Y., Syberg, K., & Khan, F. R. (2018). Chemosphere Considerations on the use of equilibrium models for the characterisation of HOC-microplastic interactions in vector studies. *Chemosphere*, 210, 359–365.
- Vesilind, P. A. (2003). *Wastewater treatment plant design*. Alexandria, Va.; London: Water Environment Federation ; IWA Publishing.
- Waller, C. L., Griffiths, H. J., Waluda, C. M., Thorpe, S. E., Loaiza, I., Moreno, B., ... Duarte, A. C. (2017). Microplastics in the Antarctic marine system: An emerging area of research. *Science of The Total Environment*, 598, 220–227.

- Wang, Fayuan, Yang, W., Cheng, P., Zhang, S., & Zhang, S. (2019). Adsorption characteristics of cadmium onto microplastics from aqueous solutions. *Chemosphere*, 235, 1073–1080.
- Wang, Fei, Shih, K. M., & Li, X. Y. (2015). The partition behavior of perfluorooctanesulfonate (PFOS) and perfluorooctanesulfonamide (FOSA) on microplastics. *Chemosphere*, 119, 841–847.
- Wang, Fen, Wong, C. S., Chen, D., Lu, X., Wang, F., & Zeng, E. Y. (2018). Interaction of toxic chemicals with microplastics : A critical review. *Water Research*, 139, 208–219.
- Wang, J., Peng, J., Tan, Z., Gao, Y., Zhan, Z., Chen, Q., & Cai, L. (2017). Microplastics in the surface sediments from the Beijiang River littoral zone: Composition, abundance, surface textures and interaction with heavy metals. *Chemosphere*, 171, 248–258.
- Wang, J., Tan, Z., Peng, J., Qiu, Q., & Li, M. (2016). The behaviors of microplastics in the marine environment. *Marine Environmental Research*, 113, 7–17.
- Wang, Q., Zhang, Y., Wangjin, X., Wang, Y., Meng, G., & Chen, Y. (2020). The adsorption behavior of metals in aqueous solution by microplastics effected by UV radiation. *Journal of Environmental Sciences*, 87, 272–280.
- Wang, W., Gao, H., Jin, S., Li, R., & Na, G. (2019). The ecotoxicological effects of microplastics on aquatic food web, from primary producer to human: A review. *Ecotoxicology and Environmental Safety*, 173, 110–117.
- Wang, W., & Wang, J. (2018). Chemosphere Comparative evaluation of sorption kinetics and isotherms of pyrene onto microplastics. *Chemosphere*, 193, 567–573.
- Ward, J. E., Zhao, S., Holohan, B. A., Mladinich, K. M., Griffin, T. W., Wozniak, J., & Shumway, S. E. (2019). Selective Ingestion and Egestion of Plastic Particles by the Blue Mussel (*Mytilus edulis*) and Eastern Oyster (*Crassostrea virginica*): Implications for Using Bivalves as Bioindicators of Microplastic Pollution. *Environmental Science & Technology*, 53(15), 8776–8784.
- Wardrop, P., Shimeta, J., Nugegoda, D., Morrison, P. D., Miranda, A., Tang, M., & Clarke, B. O. (2016). Chemical Pollutants Sorbed to Ingested Microbeads from Personal Care Products Accumulate in Fish. *Environmental Science & Technology*, 50, 4037–4044.

- Watts, A. J. R., Urbina, M. A., Corr, S., Lewis, C., & Galloway, T. S. (2015). Ingestion of Plastic Microfibers by the Crab *Carcinus maenas* and Its Effect on Food Consumption and Energy Balance. *Environmental Science and Technology*, 49(24), 14597–14604.
- Welden, N. A. C., & Cowie, P. R. (2016). Environment and gut morphology influence microplastic retention in langoustine, *Nephrops norvegicus*. *Environmental Pollution*, 214, 859–865.
- Wen, B., Jin, S., Chen, Z., & Gao, J. (2018). Single and combined effects of microplastics and cadmium on the cadmium accumulation, antioxidant defence and innate immunity of the discus fish (*Symphysodon aequifasciatus*)\*. *Environmental Pollution*, 243, 462–471.
- Wesch, C., Bredimus, K., Paulus, M., & Klein, R. (2016). Towards the suitable monitoring of ingestion of microplastics by marine biota: A review. *Environmental Pollution*, 218, 1200–1208.
- Wieczorek, A. M., Morrison, L., Croot, P. L., Allcock, A. L., Macloughlin, E., Savard, O., Brownlow, H., & Doyle, T. K. (2018). Frequency of Microplastics in Mesopelagic Fishes from the Northwest Atlantic. *Frontiers in Marine Science*, 5, 39.
- Windsor, F. M., Tilley, R. M., Tyler, C. R., & Ormerod, S. J. (2019). Microplastic ingestion by riverine macroinvertebrates. *Science of the Total Environment*, 646, 68–74.
- Witeska, M., Jezierska, B., & Wolnicki, J. (2006). Respiratory and hematological response of tench, *Tinca tinca* (L.) to a short-term cadmium exposure. *Aquaculture International*, 14(1–2), 141–152.
- Wood, C., Farrell, A., & Brauner, C. (2012). *Fish physiology: homeostasis and toxicology of essential metals*. Academic Press, London.
- Woodall, L. C., Sanchez-vidal, A., Paterson, G. L. J., Coppock, R., Sleight, V., Calafat, A., Rogers, A. D., Narayanaswamy, B. E., & Thompson, R. C. (2014). The deep sea is a major sink for microplastic debris. *Royal Society Open Science*, 1, 140317.
- Wright, S. L., Thompson, R. C., & Galloway, T. S. (2013). The physical impacts of microplastics on marine organisms: A review. *Environmental Pollution*, 178, 483–492.

- Wu, W.-M., Yang, J., & Criddle, C. S. (2016). Microplastics pollution and reduction strategies. *Frontiers of Environmental Science & Engineering*, 11(1), 6.
- Wu, X.-Y., & Yang, Y.-F. (2010). Accumulation of heavy metals and total phosphorus in intensive aquatic farm sediments: comparison of tilapia *Oreochromis niloticus* × *Oreochromis aureus*, Asian seabass *Lateolabrax japonicus* and white shrimp *Litopenaeus vannamei* farms. *Aquaculture Research*, 41(9), 1377–1386.
- Xanthos, D., & Walker, T. R. (2017). International policies to reduce plastic marine pollution from single-use plastics (plastic bags and microbeads): A review. *Marine Pollution Bulletin*, 118(1), 17–26.
- Xiong, X., Chen, X., Zhang, K., Mei, Z., Hao, Y., Zheng, J., Wu, C., Wang, K., Ruan, Y., Lam, P. K. S., & Wang, D. (2018). Microplastics in the intestinal tracts of East Asian finless porpoises (*Neophocaena asiaeorientalis sunameri*) from Yellow Sea and Bohai Sea of China. *Marine Pollution Bulletin*, 136, 55–60.
- Xu, B., Liu, F., Brookes, P. C., & Xu, J. (2018). The sorption kinetics and isotherms of sulfamethoxazole with polyethylene microplastics. *Marine Pollution Bulletin*, 131, 191–196.
- Xu, S., Ma, J., Ji, R., Pan, K., & Miao, A. (2020). Microplastics in aquatic environments: Occurrence, accumulation, and biological effects. *Science of the Total Environment*, 703, 134699.
- Xu, X.-Y., Wong, C. Y., Tam, N. F. Y., Liu, H. M., & Cheung, S. G. (2020). Barnacles as potential bioindicator of microplastic pollution in Hong Kong. *Marine Pollution Bulletin*, 154, 111081.
- Yu, F., Yang, C., Zhu, Z., Bai, X., & Ma, J. (2019). Adsorption behavior of organic pollutants and metals on micro/nanoplastics in the aquatic environment. *Science of The Total Environment*, 694, 133643.
- Yu, Y., Ma, R., Qu, H., Zuo, Y., Yu, Z., Hu, G., Li, Z., Chen, H., Lin, B., Wang, B., & Yu, G. (2020). Enhanced adsorption of tetrabromobisphenol A (TBBPA) on cosmetic-derived plastic microbeads and combined effects on zebrafish. *Chemosphere*, 248, 126067.
- Yunus, K. (2020). A review on the accumulation of heavy metals in coastal sediment of Peninsular Malaysia. *Ecofeminism and Climate Change*, 1(1), 21–35.
- Yurtsever, M. (2019). Glitters as a Source of Primary Microplastics: An Approach to Environmental Responsibility and Ethics. *Journal of Agricultural and Environmental Ethics*, 32(3), 459–478.



- Yurtsever, M., & Yurtsever, U. (2019). Use of a convolutional neural network for the classification of microbeads in urban wastewater. *Chemosphere*, 216, 271–280.
- Zaki, M. R. M., Zaid, S. H. M., Zainuddin, A. H., & Aris, A. Z. (2021). Microplastic pollution in tropical estuary gastropods: Abundance, distribution and potential sources of Klang River estuary, Malaysia. *Marine Pollution Bulletin*, 162(November 2020), 111866.
- Zettler, E. R., Mincer, T. J., & Amaral-Zettler, L. A. (2013). Life in the “plastisphere”: microbial communities on plastic marine debris. *Environmental Science & Technology*, 47(13), 7137–7146.
- Zhang, H., Wang, J., Zhou, B., Zhou, Y., Dai, Z., Zhou, Q., Dai, Z., Zhou, Q., Christie, P., & Luo, Y. (2018). Enhanced adsorption of oxytetracycline to weathered microplastic polystyrene: Kinetics, isotherms and influencing factors \*. *Environmental Pollution*, 243, 1550–1557.
- Zhang, K., Su, J., Xiong, X., Wu, X., Wu, C., & Liu, J. (2016). Microplastic pollution of lakeshore sediments from remote lakes in Tibet plateau, China. *Environmental Pollution*, 219, 450–455.
- Zhang, W., Zhang, L., Hua, T., Li, Y., Zhou, X., Wang, W., You, Z., Wang, H., & Li, M. (2020). The mechanism for adsorption of Cr(VI) ions by PE microplastics in ternary system of natural water environment. *Environmental Pollution*, 257, 113440.
- Zhao, F.-J., & Huang, X.-Y. (2018). Cadmium Phytoremediation: Call Rice CAL1. *Molecular Plant*, 11(5), 640–642.
- Zhao, S., Zhu, L., & Li, D. (2016). Microscopic anthropogenic litter in terrestrial birds from Shanghai, China: Not only plastics but also natural fibers. *Science of the Total Environment*, 550, 1110–1115.
- Zhu, J., Yu, X., Zhang, Q., Li, Y., Tan, S., Li, D., Yang, Z., & Wang, J. (2019). Cetaceans and microplastics: First report of microplastic ingestion by a coastal delphinid, *Sousa chinensis*. *Science of the Total Environment*, 659, 649–654.
- Zitko, V., & Hanlon, M. (1991). Another source of pollution by plastics: Skin cleaners with plastic scrubbers. *Marine Pollution Bulletin*, 22(1), 41–42.
- Zocchi, M., & Sommaruga, R. (2019). Microplastics modify the toxicity of glyphosate on *Daphnia magna*. *Science of the Total Environment*, 697, 134194.

- Zon, N. F., Iskendar, A., Azman, S., Sarijan, S., & Ismail, R. (2018). Sorptive behaviour of chromium on polyethylene microbeads in artificial seawater. *MATEC Web of Conferences*, 250.
- Zou, J., Liu, X., Zhang, D., & Yuan, X. (2020). Adsorption of three bivalent metals by four chemical distinct microplastics. *Chemosphere*, 248, 126064.

## LIST OF PUBLICATIONS

### Indexed Conference Proceedings

Zon, N. F., Iskendar, A., Azman, S., Sarijan, S., & Ismail, R. (2018). Sorptive behaviour of chromium on polyethylene microbeads in artificial seawater. *MATEC Web of Conferences*, 250. doi.org/10.1051/mateconf/201825006001

**(Indexed by Scopus)**

Zon, N. F., Azman, S., Abdullah, N. H., & Supian, N. S. (2020). Kinetics and Isotherm of Cadmium Adsorption onto Polyethylene Microbeads in Artificial Seawater. *IOP Conference Series: Earth and Environmental Science*, 012130. IOP Publishing. doi.org/10.1088/1755-1315/476/1/012130 **(Indexed by Scopus)**

### Book Chapter

Zon, N. F., Azman, S., & Supian, N. S. (2020). Microplastic Pollution in Marine Environment. In *Issues and Technology in Water Contaminants*. Penerbit Utm Press. **(Non-indexed)**

Zon, N. F., Azman, S., & Jamal, M. H. (2021). Physicochemical Properties of Microplastic for Adsorption of Heavy Metals in Environment. In *Adsorption for Water and Wastewater Treatment*. Penerbit Utm Press. **(Indexed by WoS)**