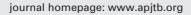


Review Article

Asian Pacific Journal of Tropical Biomedicine





doi: 10.4103/2221-1691.328054 Impact Factor: 1.55

Phytochemicals, pharmacological and ethnomedicinal studies of *Artocarpus*: A scoping review

Siti Mariam A Lathiff, Norzafneza M Arriffin, Shajarahtunnur Jamil

Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia, 81310 Johor Bahru, Johor, Malaysia

ABSTRACT

This article aims to review the scientific data on phytochemical and pharmacological studies of Artocarpus collected from Malaysia as well as to highlight their usage as ethnomedicine worldwide. About 55 Artocarpus species are distributed worldwide and 32 of the Artocarpus species can be found in Malaysia. Artocarpus species are well known worldwide for their edible fruits such as Artocarpus heterophyllus (jackfruit), Artocarpus integer (chempedak), and Artocarpus communis (breadfruit). Aside from its edible fruits, the timber is valued for light constructions, crates, large canoes, and boats. The literature for this review was searched using the term 'Artocarpus', 'Artocarpus Malaysia', 'Artocarpus extracts', 'Artocarpus traditional medicine' and 'Artocarpus ethnomedicine' from published books and scientific journals via various engines such as The Web of Science, PubMed, Science Direct, Scopus, Research Gate, and Google Scholar. The references cited from the retrieved articles were also scanned and cross-checked. All published studies on phytochemical and pharmacological activities of Malaysia's Artocarpus species up to January 2021 were included in this review. Articles on phytochemical studies of Malaysia's Artocarpus revealed the isolation of flavonoids as the major constituents. Research on pharmacological activities of the isolated phytochemicals showed that these compounds exhibited significant disease-linkedenzyme (tyrosinase, cholinesterase, glucosidase) inhibitors as well as antioxidant, anti-inflammatory, antimicrobial, and cytotoxic activities. The ethnomedicinal data gathered are useful to understand and prioritize Artocarpus species that can contribute to potent phytochemicals and possibly new drug leads. This review also provides valuable information for the future development of isolated compounds from Artocarpus species.

KEYWORDS: *Artocarpus*; Phytochemistry; Pharmacological activities; Ethnomedicine; Flavonoids

1. Introduction

Genus *Artocarpus* is one of the important groups of plants that belongs to the Moraceae family. A total of 55 *Artocarpus* species spread throughout East Asia, South Asia, Southeast Asia to the New Guinea and southern Pacific[1,2]. Up to 2020, a total of 32 *Artocarpus* species and another two varieties were discovered in Malaysia[2–4]. Some *Artocarpus* species have edible fruits that led to cultivations for the products. The fruits can be eaten as soon as it is ripe. Most are eaten fresh after they are ripe, fried with batter, or served as desserts. The seed can also be eaten after boiling, baking, roasting, or frying[5]. Our investigation on *Artocarpus* species started since we did our own phytochemical and pharmacological research. Although several reviews on *Artocarpus* species had been published, the objectives and focus are very much different from this article[6,7].

In this article, we focus on reviewing the phytochemistry and pharmacological studies of *Artocarpus* species available in Malaysia. But before we dive into that, we need to know the distribution and availability of *Artocarpus* species in Malaysia. Then from this data, we gathered the reported ethnomedicinal usage from all around the world related to these *Artocarpus* species. The ethnomedicinal data are important to understand its relationship with the pharmacological activities tested[8]. All the data are tabulated in Table 1 for better understanding.

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-Non Commercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical

For reprints contact: reprints@medknow.com

©2021 Asian Pacific Journal of Tropical Biomedicine Produced by Wolters Kluwer-Medknow. All rights reserved.

How to cite this article: Lathiff SMA, Arriffin NM, Jamil S. Phytochemicals, pharmacological and ethnomedicinal studies of *Artocarpus*: A scoping review. Asian Pac J Trop Biomed 2021; 11(11): 469-480.

Article history: Received 10 March 2021; Revision 7 April 2021; Accepted 27 August 2021; Available online 29 October 2021

 $^{^{\}mbox{\tiny \boxtimes}}$ To whom correspondence may be addressed. E-mail: shajarah@utm.my

Siti Mariam A Lathiff et al./ Asian Pacific Journal of Tropical Biomedicine 2021; 11(11): 469-480

2. Methodology

Literature was searched using various engines such as The Web of Science, PubMed, Science Direct, Scopus, Research Gate, and Google Scholar. The search term 'Artocarpus', 'Artocarpus Malaysia', 'Artocarpus extracts', 'Artocarpus traditional medicine' and 'Artocarpus ethnomedicine' were used without limitations. The references cited from the retrieved articles were scanned and cross-checked. Other than published articles, information on Artocarpus species were also obtained from books published by reliable sources. The distribution, morphology, and ethnobotanical information of Artocarpus species were obtained from books published under the Ministry of Agriculture, Malaysia, and Forest Research Institute of Malaysia. All published researches on phytochemistry and pharmacological activities of Malaysia's Artocarpus species up to January 2021 were included in this review.

3. Morphology and distributions

Artocarpus trees are mostly evergreen with thick white latex. Their leaves are spirally arranged or alternate. While Artocarpus fruits have different sizes from small to large with fleshy seeds that are mostly large and embedded in the head of the fruit and surrounded by a waxy or pulpy succulent layer. The sapwood has various shades of light yellow which can be differentiated from the heartwood that has different shades of brown and sometimes traces of olive green. Artocarpus produces two types of hardwood, light and medium hardwood. Other than its edible fruits, Artocarpus is also well known for its valuable timber in Malaysia. The light hardwood timber is known as terap in Peninsular Malaysia and Sabah or pudau in Sarawak. The medium hardwood is known as keledang in Peninsular Malaysia, beruni in Sabah and selangking in Sarawak[2,3]. Kochummen identified 20 species of Artocarpus in Malaysia including two incompletely known species and reported in Tree Flora of Malaya[3]. Another book was published specifically for Sabah and Sarawak (East Malaysia) that confirmed the availability of 20 species with one incompletely known species[2]. These two reports conclude a total of 32 Artocarpus species and another two varieties in Malaysia. Distribution of identified Artocarpus in Malaysia is listed in Table 1.

4. Ethnomedicine

Ethnomedicine is the study of the cultural concept of health, disease and illness using nature[9]. There are several published articles on ethnomedicine practices using different parts of *Artocarpus* by certain tribes or specific locations in the world[10–16]. Table 1 highlights available *Artocarpus* species discovered in Malaysia including their ethnomedicine practices worldwide.

5. Phytochemistry

A total of 61 compounds were isolated from *Artocarpus* species collected from different locations in Malaysia. These compounds fall under the flavonoids (chalcones, flavones, flavones, flavanones, flavonols), xanthones, stilbenoid as well as terpenoids, and sterols. Flavonoids are present in all *Artocarpus* species and proved to be the chemotaxonomic marker of *Artocarpus* plants. The structures of compounds 1-61 are shown in Supplementary Figures 1-5.

5.1. Phenolics secondary metabolites

5.1.1. Chalcones

Two separate studies of Artocarpus lowii (A. lowii) from Terengganu reported the isolation of two new dihydrochalcones. These chalcones are named as 2',4'-dihydroxy-4-methoxy-3'-prenyldihydrochalcone (1) and 2',4'-dihydroxy-3,4-(2",2"dimethylchromeno)-3'-prenyldihydrochalcone (2)[17,18]. Another two chalcones were also isolated and known as isobavachalcone (3) and 4-hydroxyonchocarpin (4)[17-19]. Isobavachalcone (3) was also isolated from Artocarpus anisophyllus (A. anisophyllus) collected from Johor[20]. In 2016, two new prenylated chalcones were reported from A. lowii collected from Selangor. These chalcones are identified as 2-hydroxyparatocarpin (5) and 2',3,4',4-tetrahydroxy-3'prenylchalcone (6)[21]. A study on the leaves of Artocarpus fulvicortex (A. fulvicortex) from Terengganu, Malaysia gave 2'-hydroxy-4,4',6'trimethoxychalcone (7)[22]. In 2013, two new dihydrochalcones named elastichalcone A (8) and elastichalcone B (9) were isolated from the leaves of Artocarpus elasticus (A. elasticus) collected from Selangor, Malaysia[23]. The structures of all chalcones are presented in Supplementary Figure 1.

5.1.2. Flavones

Artonin E (10), a known flavone with four hydroxyl groups, a prenyl, and a chromeno ring was isolated from the barks of *Artocarpus scortechinii* (*A. scortechinii*), *Artocarpus teysmanii* (*A. teysmanii*), and *A. elasticus* from Selangor[24–27]. An investigation of *A. elasticus* in 2019 also reported a new diprenylated flavone, artoflavone B (11)[27]. In 2010, two new prenylated flavones were reported from two different species collected from Sarawak[28,29]. These flavones are named as artosimmin (12) from *Artocarpus odoratissimus* (*A. odoratissimus*) and hydroxyartocarpin (13) from the stem bark of *Artocarpus altilis* (*A. altilis*)[28,29]. Another three flavones isolated from *A. altilis* were identified as artocarpin (14), morusin (15), and cycloartocarpin A (16)[29]. Artochamin A (17), a prenylated pyranoflavone was isolated from the stem bark of *Artocarpus kemando* (*A. kemando*) also from Sarawak[30].

A thorough investigation on the leaves of *A. fulvicortex* from Terengganu gave a unique new flavone bearing two chromeno rings[22]. The structure was identified as 5-hydroxy-(6:7,3':4')-di(2,2-dimethylpyrano)flavone (18). Carpachromene (19) together

Table 1. Distribution and availability of Artocarpus species in Malaysia and their ethnomedicinal uses.

C :					D C
	Synonyms	Common/Local names	Distribution in Malaysia	Ethnomedicine practices worldwide	References
		Keledang babi, Terap ikal		-	[2,3]
]	Boerl.; Artocarpus	(Sabah), Bintau, Bintawak,	available in Negeri		
	superba Becc.	Entawa, Mentawa, Kayo	Sembilan and Johor);		
		bibungan, Kelidang, Tawak	East Malaysia (available		
		(Sarawak)	widely in Sabah but		
			uncommon in Sarawak)		
Artocarpus anisophyllus var.	_	Terap ikal	West Malaysia (not	_	[2,3]
sessilifolius Kochummen		-	available); East Malaysia		
,g			(widely distributed in		
			Sabah but only single		
			collection in Kuching		
			_		
			Sarawak)		
A-t Iorrott		Bukoh, Patat	West Melevsie (not		[2 2]
Artocarpus annulatus Jarrett	_	bukon, Patat	West Malaysia (not	_	[2,3]
			available); East Malaysia		
			(only in Sarawak)		
, -	*			Leaves: Skin disease, toothache, enlarge spleen	
	& G. Forster; Artocarpus		over Malaysia	treatment (Indonesia); Diabetic treatment	8,12,13,15]
	camansi Blanco;			(Indonesia), eye ailments (Samoa and Futuna);	
	Artocarpus incisus			Treatment of chest pain and vomiting that are caused	
				by heart problem, muscle relaxant, remedy for fish	
				poisoning (South Pacific)	
				Shoots and flowers: Wrapped with Macaranga dioica	
				leaves to treat migraine and headache (Vanuatu);	
				Hemorrhoid treatment (Benin)	
				Barks: Treatment for chest pain and vomiting that	
				are caused by heart problem, bones pain, maternal	
				postpartum infections (South Pacific), stomach	
				aches, digestive tract problems (Samoa & Tonga),	
				relapsed illness (Tonga); Treatment for stomach	
				aches, heal wounds (Philippines); Treatment for	
				measles, scabies, dysentery (Benin)	
				Roots: Respiratory ailments include difficult &	
				painful breathing, remedy for weakness after birth,	
				blood vomiting and lung pain treatment (South	
				Pacific); Treatment of diarrhea and dysentery	
				(Indonesia); Treatment of typhoid fever, anaemia,	
				malaria, regulation of blood pressure, heart	
				palpitation (Benin) Seeds/Fruits: Help in giving birth, typhoid and other	
				1 0 0 11	
				fever treatment (China); As aphrodisiac (Indonesia)	
				Milky latex: Applied to rashes, abscesses, sores,	
				wounds (Tonga and Tahiti); Dysentery treatment	
				(Philippines); Mixed with equal amount of Ficus	
				adenosperm latex to treat menorrhagia (Vanuatu)	
				White sap: Treatment for eye puncture wounds	
				(Micronesia)	
Artocarpus corneri Kochummen	-	Talun	West Malaysia (not	-	[2,3]
			available); East Malaysia		
			(only in Sarawak)		
Artocarpus dadah Miq.	Artocarpus mollie Mia	Tampang bulu (Paningular)	West Malaysia (common	Leaves: Treating oedema (Southeast Asia)	[2,3,5,8]
	*	·	- ·	Bark: Sap from bark used to clean leg wound	
1	in ocurpus rujescens WIQ.;			•	
	Antooguess	uadak, meruni, selangking	open country/ villages		
	Artocarpus tampang		. 1 1 222		
1	Miq.; Artocarpus		-	Juice and seed: As laxative (Myanmar)	
]	Miq.; Artocarpus inconstantissima Miq.;		Malaysia); East Malaysia	Roots: Tonic for deobstruent	
]	Miq.; Artocarpus		-	Roots: Tonic for deobstruent	
1	Miq.; Artocarpus inconstantissima Miq.;		Malaysia); East Malaysia	Roots: Tonic for deobstruent	
] ; 1	Miq.; Artocarpus inconstantissima Miq.; Artocarpus lakoocha		Malaysia); East Malaysia (common in Sabah, rare	Roots: Tonic for deobstruent	
) , , ,	Miq.; Artocarpus inconstantissima Miq.; Artocarpus lakoocha Roxb.; Artocarpus		Malaysia); East Malaysia (common in Sabah, rare	Roots: Tonic for deobstruent	

Table 1. Distribution and availability of Artocarpus species in Malaysia and their ethnomedicinal uses (continued).

Species Species	Synonyms		Distribution in Malaysia	Ethnomedicine practices worldwide	References
•				· · · · · · · · · · · · · · · · · · ·	
	*			Leaves: Mix with rice and eaten for tuberculosis	[2,3,3]
	*	(Sabah), Kian, Pedalai,			
	*			Inner bark: As native bandages & poultice for ulcer	
	Artocarpus pubescens	(Sarawak)	East Malaysia	(Malaysia)	
	Willd.; Artocarpus			Bark: Pounded for lumbago (muscle and joint)	
	sericicarpus Jarrett.			treatment (Indonesia)	
				Latex: Treatment for dysentery (inflammation of	
				intestine) (Indonesia)	
4. 7. 7. 11			W . M 1		[0.0]
Artocarpus excelsus Jarrett	-	_	West Malaysia (not		[2,3]
			available); East Malaysia		
			(rarely available in		
			Sabah and Sarawak)		
Artocarpus fulvicortex Jarrett		Tampang gajah	West Malaysia (only		[2 2]
Artocarpus juivicoriex Janea	_	Tampang gajan	available in lowland		[2,3]
			forest of Perak, Pahang,		
			Negeri Sembilan,		
			Melaka); East Malaysia		
			(not available)		
Artocarnus alaucus Rhimo	Artocarnus danisania	Merubi, Pudau putih,	West Malaysia (orly		[2 3]
		Selangking	available in lowland		[2,3]
	King	Sciangking			
			and hill forest of Kedah,		
			Perak, Pahang, Selangor,		
			Johor); East Malaysia		
			(not available)		
Artocarpus gomezianus Wall. ex		Tampang hitam	West Malaysia (only	_	[2,3]
Trec.		rampang mam	available in lowland		[2,3]
TICC.			forest of Kedah,		
			Kelantan, Pahang,		
			Negeri Sembilan, Johor);		
			East Malaysia (not		
			available)		
Artocarpus heterophyllus Lamk.		Jackfruit (English) Nangka	Commonly available	Sap: Antisyphilitic (Southeast Asia); Vermifuge	[2 3 7 8 16
Artocarpus neterophytius Lanik.		(Malay)	-	(Southeast Asia); Ulcers, abscesses (Philippines,	[2,3,7,0,10]
		(Malay)	all over Malaysia	Myanmar, China)	
			ali ovei ivialaysia	i i	
				Pulp and seeds of fruits: Cooling tonic for pectoral	
				muscle (Philippines, Myanmar, China)	
				Leaves: Ulcers, wounds (Malaysia and Philippines);	
				Increase milk secretion for woman (Southeast Asia)	
				Barks: Ulcers (Malaysia); Sedative for convulsions	
				(Southeast Asia); Applied on skin to reduce swelling	
				(India)	
				Roots: Diarrhoea (Myanmar); Fever (Malaysia)	
Artagarnus himidus Igreatt		Temponek	West Malayeia (available		[2 3]
Artocarpus hispidus Jarrett.	_	Temponek	West Malaysia (available	-	[2,3]
			in lowland forest of		
			Penang, Perak, Pahang,		
			Terengganu, Selangor);		
			East Malaysia (not		
			21.1.1.5		
			available)		
Artacarnus integras Moss	Artocarpus chemned	Chemnedob		Seeds: Treatment for diarrhag	[2 3 6]
	Artocarpus chempeden	Chempedak	Commonly available	Seeds: Treatment for diarrhea	[2,3,6]
	Artocarpus chempeden Spreng	Chempedak	Commonly available because it is cultivated	Seeds: Treatment for diarrhea Roots: Treatment for malaria fever	[2,3,6]
		Chempedak	Commonly available		[2,3,6]
			Commonly available because it is cultivated all over Malaysia		
		Chempedak Bangkong	Commonly available because it is cultivated all over Malaysia Widely distributed in		[2,3,6]
			Commonly available because it is cultivated all over Malaysia Widely distributed in Malaysia but nowhere		
			Commonly available because it is cultivated all over Malaysia Widely distributed in		
Artocarpus integer var. silvestris		Bangkong	Commonly available because it is cultivated all over Malaysia Widely distributed in Malaysia but nowhere abundant	Roots: Treatment for malaria fever –	[2,3]
			Commonly available because it is cultivated all over Malaysia Widely distributed in Malaysia but nowhere abundant West Malaysia (not	Roots: Treatment for malaria fever - -	
Artocarpus integer var. silvestris Artocarpus jarriettiae		Bangkong	Commonly available because it is cultivated all over Malaysia Widely distributed in Malaysia but nowhere abundant	Roots: Treatment for malaria fever - -	[2,3]

Table 1. Distribution and availability of Artocarpus species in Malaysia and their ethnomedicinal uses (continued).

Species	Synonyms	Common/Local names	Distribution in Malaysia	Ethnomedicine practices worldwide	References
Artocarpus kemando Miq.	· ·	Pudu (Sabah), Pudau,		-	[2,3]
1	moore	Selibut, Pupud, Puduh,			[-,-]
		Puroh (Sarawak)	in lowland forest of		
		, , ,	Terengganu, Pahang,		
			Selangor); East Malaysia		
			(available widely in		
			Sabah and Sarawak)		
			ŕ		
Artocarpus lanceifolius Roxb.	-	Keledang	Widely distributed in	-	[2,3]
			Malaysia but nowhere		
			abundant		
Artocarpus lowii King	-	Miku	Widely distributed in		[2,3]
			Malaysia but nowhere		
			abundant		
		D 1			F2 23
Artocarpus maingayi King	_	Pudu	Available in lowland		[2,3]
			forest throughout		
			Malaysia		
Artagarnus malinavalus Gagnen		Pala mansoh, Pala tupai,	West Malaysia (not		[2 3]
Artocarpus melinoxylus Gagnep.	_	Temponek	available); East Malaysia		[2,3]
		remponek	(rarely available in		
			Sabah and Sarawak)		
			Saban and Sarawak)		
Artocarpus nitidus Trec.	Artocarpus lanceolate	Tampang (Peninsular),	Available in lowland	_	[2,3]
	•	Selangking, Beruni			[]
		(Sabah), Dadak, Empaka,			
		Karon, Ngidinuk, Sinojoh,			
	Artocarpus sampor				
	Gagnep.; Artocarpus				
	lingnanensis; Artocarpus				
	borneensis Merr.;				
	Artocarpus gomeziana				
	Wall.; Artocarpus griffithii				
	Merr.				
Artocarpus obtusus Jarrett	-	Sarawak and Sabah	-	-	[2,3]
			***	Y	FA 43
Artocarpus odoratissimus				Latex: Treat inflammation from wounds (Malaysia)	[2,3]
	*	(Sabah), Keiran Pingan	_		
	Becc.	Tekalong (Sarawak)	West Malaysia); East		
			Malaysia (growing naturally in Sabah and		
			-		
			Sarawak)		
Artocarpus peltatus Merr.	_	_	West Malaysia (not	_	[2,3]
, , , , , , , , , , , , , , , , , , ,			available); East Malaysia		[=,-]
			(commonly available		
			in Sarawak but rarely		
			available in Sabah)		
Artocarpus primackiana	-	Beruni (Sabah), Dadah	West Malaysia (not	-	[2,3]
Kochummen		(Sarawak)	available); East Malaysia		
			(rarely available in		
			Sabah and Sarawak)		
Artocarpus rigidus Blume	$Artocarpus\ rotunda;$	Temponek	Widely distributed	-	[2,3]
	Artocarpus cuspidatus;		throughout Malaysia		
	Artocarpus kertau;				
	Artocarpus varians				
	Miq; Artocarpus				
	dimorphophylla Miq.				

Table 1. Distribution and availability of Artocarpus species in Malaysia and their ethnomedicinal uses (continued).

Species	Synonyms	Common/Local names	Distribution in Malaysia	Ethnomedicine practices worldwide	References
Artocarpus sarawakensis Jarrett	-	Pedalai, pingan (Sarawak)	West Malaysia (not	-	[2,3]
			available); East Malaysia		
			(rarely available in		
			Sarawak)		
Artocarpus scortechinii King	-	Terap hitam	Scattered throughout	-	[2,3]
			Malaysia		
Artocarpus tamaran Becc.	-	Timbangan (Sabah),	West Malaysia (not	-	[2,3]
		Kakang, tamaran, tembaran,	available); East Malaysia		
		kalong, kihan (Sarawak)	(rarely available in		
			Sabah and Sarawak)		
Artocarpus teysmannii Miq.	Artocarpus peduncularis	Chempedak ayer	West Malaysia (available	-	[2,3]
			in Perak and Selangor);		
			East Malaysia (available		
			in Sabah and Sarawak)		
Artocarpus tomentosulus Jarrett	-	-	West Malaysia (not	-	[2,3]
			available); East Malaysia		
			(rarely available in		
			Sabah)		
Artocarpus sp. 'A'	-	Tampang FRI 17082	West Malaysia (Gunung	-	[2,3]
		(Known from only one	Tampin, Negeri		
		collection)	Sembilan)		
Artocarpus sp. 'B'	-	Tampang FRI 19234	West Malaysia (Gunung	_	[2,3]
		(Known from only one	Ledang, Johor)		
		collection)			
Artocarpus sp. A	-	SAN 82081 (Known from	East Malaysia (Sepilok,	-	[2,3]
		only one collection)	Sandakan Sabah)		

with cycloartocarpesin (20) and norartocarpetin (21) was isolated from *A. fulvicortex*[22]. Cycloartocarpesin (20) was also reported in a phytochemical study of *A. elasticus* from Selangor[23].

In 2015, two new and three known flavones were isolated from A. anisophyllus collected from Johor[20]. The new flavones were identified as 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8-γ,γ-dimethylallylflavone (22) and 3-hydroxycycloartocarpin (23) while the others are known as artocarpin (14), cycloartocarpin (24), and chaplashin (25)[20]. The following year, Abdullah et al. reported the isolation of artocarpin(14), 4',5-dihydroxy-6,7-(2,2dimethylpyrano)-2'-methoxy-8-γ,γ-dimethylallylflavone (22), and cycloheterophyllin (26) from A. lowii[18]. Whilst Arriffin et al. reported the isolation of 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8-γ,γ-dimethylallylflavone (22) from the bark of A. scortechinii in 2017[31]. Methoxycyclocommunol (27), cyclocommunol (28), and cudraflavone C (29) were present in the bark of Artocarpus integer var. silvestris (A. integer var. silvestris) from Pahang[32]. The structure of methoxycyclocommunol (27) was reported as a new derivative of cyclocommunol (28) due to the presence of a sharp peak indicating methoxy group in ¹HNMR.

5.1.3. Flavanones

Two flavanones were reported for the first time from Artocarpus

species characterized as 5,7-dihydroxy-4'-methoxy-8-prenylflavanone (30) and 5-hydroxy-7,8-(2,2-dimethylchromano)-4'-methoxyflavanone (31)[20,33]. In 2016, a phytochemical study on *A. integer* var. *silvestris* yielded heteroflavanone A (32) whilst two common flavanones named pinostrobin (33) and pinocembrin (34) were obtained from *A. odoratissimus*[32,34].

5.1.4. Flavanols

Flavanols are also referred to as dihydroflavonols. Shah *et al.* obtained a common flavanol named catechin (35) from the leaves of *A. fulvicortex* collected from Terengganu[35]. Arriffin *et al.* reported the isolation of two new dihydroflavonols named artoscortonol A (36) and artoscortonol B (37) from the leaves of *A. scortechinii*[36]. The structures of all flavanones (30-34) and flavanols (35-37) are presented in Supplementary Figure 3.

5.1.5. Xanthones and stilbenoid

Several articles reported the isolation of artobiloxanthone (38), cycloartobiloxanthone (39), and artonol B (40) from the barks of A. scortechinii and A. teysmanii[24,25,31]. Two separate study by Ee et al. and Hashim et al. on A. kemando from Sarawak also isolated cycloartobiloxanthone (39) and artonol B (40)[30,37]. In addition, Ee et al. highlights another two xanthones named as artomandin (41),

and artoindonesianin C (42) from *A. kemando*[30]. The first chemical investigation on the stem bark of *A. obstusus* in 2010 by Hashim *et al.* found two new xanthones named pyranocycloartobiloxanthone A (43) and dihydroartoindonesianin C (44)[38]. Afterwards, Hashim *et al.* reported the isolation of pyranocycloartobiloxanthone B (45) from *A. obstusus* and dihydroartoindonesianin C (44) from *A. kemando*[37,39].

In 2015, Lathiff *et al.* obtained pyranocycloartobiloxanthone A (43) from a rare *Artocarpus* species identified as *A. anisophyllus*[20]. The following year, a known xanthone called artonin F (46) was reported from *A. integer* var. *silvestris* from Pahang whereas the investigation on the stem bark of *A. altilis* yielded two known xanthones, cycloartobiloxanthone (39) and artoindonesianin V (47) [29,32]. An extensive investigation on *A. elasticus* in 2016 reported a new xanthone named elastixanthone (48) together with artobiloxanthone (38) and cycloartobiloxanthone (39)[26]. A recent investigation on *A. elasticus* in 2019 also reported the isolation of artobiloxanthone (38) and cycloartobiloxanthone (39), elastixanthone (48) as well as artoindonesianin P (49)[27]. Isolation of a common stilbenoid, oxyresveratrol (50) was reported from *A. scortechinii* and *A. fulvicortex*[31,35]. The structures of xanthones and stilbenoid (38-50) are presented in Supplementary Figure 4.

5.2. Other types of secondary metabolites

Isolation of friedelin (51), lupeol (52), and lupeol-3-acetate (53) was reported from separate study on A. fulvicortex from Terengganu[22,35]. A recent study showed A. odoratissimus roots contained two terpenoids, α and β amyrin acetate (54, 55)[34]. The structures can be distinguished from proton NMR and by comparison with literature. Traxateryl acetate (56) was also isolated from the stem bark and the roots of A. odoratissimus together with hexyl dodecanoate (57)[28,34]. Two common sterols, β-sitosterol (58) and stigmasterol (59) were reported from A. kemando and A. odoratissimus[29,34]. Phytochemical studies on the stem bark of A. kemando from Sarawak yielded 6,7-dimethoxycoumarin (60) and aurantiamide benzoate (61). Aurantiamide benzoate (61) was reported as the first dipeptide isolated from genus Artocarpus. It was crystallized from chloroform extracts of A. kemando's stem bark after a series of purification processes[37]. Structures of other secondary metabolites are shown in Supplementary Figure 5.

6. Pharmacological activities

The *Artocarpus* species have been exploited in various countries and revealed the medicinal variation possibility. As noted earlier, the most common ethnomedicinal usage was to treat wounds and ulcers as well as some skin problems. Extensive literature reported different pharmacological tests involved with the extracts and the isolated phytochemicals which exhibited anti-inflammatory, antioxidant,

antimicrobial, gastroprotective, cytotoxic, anti-proliferative activities and acted as selective enzymes inhibitors. Qualitative and quantitative phytochemical screening of the *Artocarpus* extracts revealed their high phenolic content[40–45].

6.1. Antioxidant activities

Several reports have revealed the positive correlation between high phenolic content and the antioxidant activities of the crude extracts and fractions[40,42-45]. Antioxidant activity for isolated compounds also had been reported. 2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (1), isobavachalcone (3) and 4-hydroxyonchocarpin (4) from A. lowii showed strong free radical scavenging activities against 2,2'-diphenyl-1-picrylhydrazyl (DPPH) with 50% inhibition (scavenging) concentrations (IC₅₀) of 0.03-0.24 mM using electron spin resonance spectrometry[17]. Ferric reducing antioxidant power assay (FRAP), 2,2'-azinobis (3-ethylbenzthiazoline-6-sulfonic acid) (ABTS) and DPPH scavenging assays for all the isolates, 2',4'-dihydroxy-4-methoxy-3'-prenyldihydrochalcone (1), 2',4'-dihydroxy-3,4-(2",2"-dimethylchromeno)-3'-prenyldihydrochalcone (2), isobavachalcone (3), 4-hydroxyonchocarpin (4), artocarpin (14), and cycloheterophyllin (26) from A. lowii were also conducted. Cycloheterophyllin (26) gave promising results compared with standards, butylated hydroxyanisole (BHA) with 50% scavenging concentration (SC₅₀) values of 0.320 mM (BHA=0.554 mM) (ABTS), 0.10 mM (BHA=0.082 mM) (DPPH) and (4.70±0.09) [Trolox=(2.8±0.09)] FRAP equivalent[19]. New chalcones, elastichalcones A-B (8-9) and cycloartocarpesin (20) from A. elasticus were tested using Thin Layer Chromaography (TLC) bioautography method for DPPH scavenging activity. Elastichalcone B (9) and cycloartocarpesin (20) showed colour changes and further evaluation using 96-wells with a microplate photometer supported the TLC results with IC₅₀ values of 11.30 and 11.89 µg/mL respectively[23].

Pyranocycloartobiloxanthone A (43) was obtained from Artocarpus obstusus for the first time together with dihydroartoindonesianin C (44) and pyranocycloartobiloxanthone B (45). Although they have similar backbones, only pyranocycloartobiloxanthone A (43) showed promising results against DPPH radical with IC₅₀ of 2.0 μg/mL while the other xanthones were considered inactive with IC₅₀ more than 500 μg/mL[38,39]. Isobavachalcone (3), artocarpin (14), 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8γ,γ-dimethylallylflavone (22), 5,7-dihydroxy-4'-methoxy-8prenylflavanone (30), 5-hydroxy-7,8-(2,2-dimethylchromano)-4'methoxyflavanone (31) and pyranocycloartobiloxanthone A (43) from A. anisophyllus were tested using DPPH scavenging assay with a microplate photometer. Pyranocycloartobiloxanthone A (43) demonstrated strong antioxidant activity with SC₅₀ of 20.2 µg/mL comparable with the positive control, BHA ($SC_{50}=17.5 \mu g/mL$)[20]. The structure activity relationship between flavonoids artosimmin (12), cycloartobiloxanthone (39), artonol B (40), artomandin (41), artoindonesinin C (42) and extracts of *A. odoratissimus* and *A. kemando* was evaluated using DPPH radical scavenging assay. All extracts of *A. odoratissimus* exhibited weak inhibition activity (>120 μg/mL) compared with *A. kemando* (<55 μg/mL). Artomandin (41) and artosimmin (12) from *A. kemando* exhibited the highest potential in DPPH radical scavenging assay with IC₅₀ of 38.0 and 32.1 μg/mL respectively[46]. The antioxidant capacities of *Artocarpus* secondary metabolites and extracts might lead to alleviation of diabetes mellitus and link to inhibition of tumour or cancer cell with potential antiproliferative activities[47,48].

6.2. Anti-inflammatory activity

The anti-inflammatory activity of 2',4'-dihydroxy-4methoxy-3'-prenyldihydrochalcone (1), isobavachalcone (3), 4-hydroxyonchocarpin (4), and cycloheterophyllin (26) from A. lowii was investigated using cyclooxygenase-2 (COX-2) and 15-lipoxygenase (15-LOX) screening kit. Isobavachalcone (3) was found to possess potent anti-inflammatory activity via COX-2 mechanism (IC₅₀=0.95 μM). However, no activities were shown by other compounds towards COX-2 and 15-LOX[49]. In 2017, seven flavonoids i.e. 2',4'-dihydroxy-4-methoxy-3'-prenyldihydrochalcone (1), isobavachalcone (3), artonin E (10), artocarpin (14), 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8γ,γ-dimethylallylflavone (22), cycloheterophyllin (26) and oxyresveratrol (50) from several Artocarpus species were investigated on inhibitory effect against the production of prostaglandin E₂ (PGE₂) in human plasma. The level of PGE₂ in plasma was determined using radioimmunoassay technique. Artocarpin (14) showed the highest inhibition of 68.1% followed by artonin E (10) (66.8%) compared with the positive control, indomethacin (79.2%). 2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (1), artonin E (10), artocarpin (14), and 4',5-dihydroxy-6,7-(2,2dimethylpyrano)-2'-methoxy-8-γ,γ-dimethylallylflavone (22) were the most potent inhibitors with IC₅₀ values of 8.99 μ M, 8.98 μ M, 11.66 μM, and 7.04 μM, respectively (indomethacin: IC₅₀=2.27 μM). These compounds reduced PGE₂ production in human blood which directly inhibited COX-2 enzymatic activity[50]. Isolated compounds methoxycyclocommunol (27), cyclocommunol (28), cudraflavone C (29), heteroflavanone A (32), and artonin F (46) from A. integer var. silvestris were tested for their antiinflammatory activity using 15-LOX screening kit and inhibitory effects on production of PGE2 in whole blood. All compounds showed weak inhibition against 15-LOX even at a concentration of 100 µM. Only methoxycyclocommunol (27), cudraflavone C (29), heteroflavanone A (32), and artonin F (46) were screened for inhibitory effects on production of PGE2 in whole blood using radioimmunoassay methods. All compounds except artonin F (46) showed more than 55% inhibition which then proceeded to serial dilution method to determine the IC₅₀ value. Cudraflavone C (29) showed the most potent COX-2 inhibition on PGE₂ with IC₅₀ of 0.07 μM which showed lower concentration needed for 50% inhibition

compared with the positive control, indomethacin (IC $_{50}$ =0.2 μ M). Methoxycyclocommunol (27) and heteroflavanone A (32) also showed remarkable COX-2 inhibition on PGE $_2$ with IC $_{50}$ values of 4.3 and 0.8 μ M, respectively[32]. These results indicated that isolated flavonoids from *Artocarpus* do not respond towards 15-LOX mechanisms but act as selective inhibitors through COX-2 pathway.

6.3. Antimicrobial activity

Pyranocycloartobiloxanthone A (43), dihydroartoindonesianin C (44), and pyranocycloartobiloxanthone B (45) were screened for their antimicrobial properties using disc diffusion method. Only pyranocycloartobiloxanthone A (43) showed inhibition against most of the bacteria and fungi tested. Among all the microbes, pyranocycloartobiloxanthone A (43) showed the most promising result against methicillin resistant *Staphylococcus aureus* (*S. aureus*) (MRSA) with a 20 mm inhibition zone. Dihydroartoindonesianin C (44), and pyranocycloartobiloxanthone B (45) were found to be inactive[39]. Pyranocycloartobiloxanthone A (43) was also tested against two strains of *Helicobacter pylori*, NCTC 11637 (ATCC 43504) and J99 (ATCC 700824) with MIC values of > 250 μ g/mL and 62.5 μ g/mL, respectively. *Helicobacter pylori* is a bacterium that can cause ulcers in the stomach[51].

Crude extracts and ten isolated flavonoids, 2',4'-dihydroxy-4methoxy-3'-prenyldihydrochalcone (1), 2',4'-dihydroxy-3,4-(2",2"dimethylchromeno)-3'-prenyldihydrochalcone (2), isobavachalcone (3), 4-hydroxyonchocarpin (4), artocarpin (14), 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8-γ,γ-dimethylallylflavone (22), cycloheterophyllin (26), 5,7-dihydroxy-4'-methoxy-8prenylflavanone (30), 5-hydroxy-7,8-(2,2-dimethylchromano)-4'methoxyflavanone (31) and pyranocycloartobiloxanthone A (43) from A. lowii and A. anisophyllus were screened for antimicrobial activities against four bacteria (Bacillus subtilis, S. aureus, Escherichia coli, Pseudomonas putida) and two fungi (Candida albicans, Candida glabrata) via disc diffusion method and determination of minimum inhibitory concentration as well as minimum microbicidal concentration. The crude extracts showed inhibition against Grampositive bacteria but a negative response toward fungi. Artocarpin (14) showed the most promising result as an antimicrobial agent with more than 11 mm inhibition zone and a minimum microbicidal concentration value of 0.45 mg/mL[19].

In 2015, ultrastructural changes of *S. aureus* were compared against artonin E (10) and streptomycin (positive control) using two methods, standard antimicrobial technique, and transmission electron microscopy. The minimum inhibitory concentration of 3.9 μg/mL and minimum microbicidal concentration of 7.81 μg/mL against *S. aureus* proved that artonin E (10) is active against Grampositive bacteria. In addition, the transmission electron microscope images of *S. aureus* before and after artonin E (10) treatment were shown and the original shape, cocci colonies of grape shape walls were missing, shredded and broken which led to distorted shape and focally thickened outer membrane indicating severe damage[52].

6.4. Tyrosinase inhibitory activity

Preliminary screening of pyranocycloartobiloxanthone A (43) from Artocarpus obtusus showed significant tyrosinase inhibitory activity with 80% inhibition comparable to kojic acid with 96% inhibition[39]. Further investigation by Lathiff et al. proved that aside from acting as a potent antioxidant, pyranocycloartobiloxanthone A (43) also acted as a tyrosinase inhibitor with IC_{50} of 60.5 µg/mL (kojic acid = 31.2 μg/mL)[20]. As expected, the ethyl acetate extract of A. anisophyllus heartwood where the compound was isolated exhibited a low IC₅₀ value (155.4 μg/mL). Other isolated flavonoids i.e. isobavachalcone (3), hydroxyartocarpin (13), artocarpin (14) and 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8-γ,γ-dimethylallylflavone (22) were also evaluated for their inhibitory effect against tyrosinase enzyme in vitro with IC_{50} values of more than 200 µg/mL[20]. Cycloheterophyllin (26) from A. lowii demonstrated significant tyrosinase inhibitory activity against mushroom tyrosinase with IC_{50} of 52.5 µg/mL comparable with the positive control, kojic acid (IC₅₀=31.2 µg/mL). Other flavonoids 2',4'-dihydroxy-4methoxy-3'-prenyldihydrochalcone (1), 2',4'-dihydroxy-3,4-(2",2"dimethylchromeno)-3-prenyldihydrochalcone (2), isobavachalcone (3), 4-hydroxyonchocarpin (4), artocarpin (14), and 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8-γ,γ-dimethylallylflavone (22) were also tested but found inactive[53].

6.5. Cholinesterase inhibitory activity

Isobavachalcone (3), 5,7-dihydroxy-4'-methoxy-8prenylflavanone (30) and 5-hydroxy-7,8-(2,2-dimethylchromano)-4'-methoxyflavanone (31) from A. anisophyllus were selected for in silico bioactivity analysis. 5,7-Dihydroxy-4'-methoxy-8prenylflavanone (30) was found nontoxic in absorption, digestion, metabolism, and excretion study. 5,7-Dihydroxy-4'-methoxy-8prenylflavanone (30) also followed the drug-likeness properties in Molsoft described by Lipinski Rule of 5 (RO5) for orally administered drugs. In this study, acetylcholinesterase (AChE) was chosen as target while 5,7-dihydroxy-4'-methoxy-8-prenylflavanone (30) as ligand. Molecular docking was performed between ligand and target along with known inhibitors and drug molecules available on the market. The docking score of ligand-target binding complex (-13.5762) was more significant than phase 4 drugs but lower than donepezil (-15.4974) and some other known inhibitors. The IC₅₀ value for 5,7-dihydroxy-4'-methoxy-8-prenylflavanone (30) was 1659 nM against AChE in QSAR analysis. In vitro experiment was done to validate the in silico result using TLC bioautographic method and 5,7-dihydroxy-4'-methoxy-8-prenylflavanone (30) showed potential as an AChE inhibitor with a detection limit of 125 µg/mL[33].

Dichloromethane and methanol extracts of *A. fulvicortex* heartwoods demonstrated AChE inhibitory activity with a detection limit of 2 mg/mL. Catechin (35) and oxyresveratrol (50) from *A. fulvicortex* exhibited moderate AChE inhibitory activity using TLC bioautographic method and microplate assay. Oxyresveratrol (50)

acted as a potent inhibitor with IC $_{50}$ value of 6.25 mM comparable to galanthamine as a positive control[35]. In 2019, cholinesterase inhibition screening of methanol extracts of *A. altilis* and *A. integer* leaves against AChE and BChE at 200 µg/mL was done with physostigmine as the positive control. Both species showed inhibition percentage of more than 80%. Further investigation revealed acidic fraction (via acid-base fractionation of methanol extract) of *A. altilis* leaves showed higher inhibition compared to ethyl actetate fraction (via liquid-liquid fractionation of methanol extract)[54].

6.6. Alpha glucosidase activity

Six compounds from A. elasticus were screened using alpha glucosidase-UV assay. Artonin E (10), artoflavone B (11) and four dihydrobenzoxanthone known as artobiloxanthone (38), cycloartobiloxanthone (39), elastixanthone (48) and artoindonesianin P (49) showed promising results as alpha glucosidase inhibitors with IC $_{50}$ between 7.6-25.4 μM . Overall, all compounds showed dose dependent inhibition of enzyme. A detailed kinetic analysis on the inhibition of artobiloxanthone (38), cycloartobiloxanthone (39), elastizanthone (48) and artoindonesianin P (49) was carried out using double reciprocal plots. These analyses determined the kinetic profile of elastixanthone (48) with IC $_{50}$ value of 7.6 μM as a slow binding inhibitor due to the residual activity of the enzyme which decreases as a function of preincubation time. Molecular docking was also conducted between Saccharomyces cerevisiae alpha-glucosidase and the xanthones as ligand. The docking results revealed that all compounds have similar binding confirmations stabilized by interactions[27].

6.7. Cytotoxicity

Cytotoxicity in natural product is an important aspect to be measured as it requires to produce effects only on targeted cells without harming the host[55]. Artosimmin (12) and artomandin (41) exhibited strong inhibition activity against HL-60 human promyelocytic leukemia with IC $_{50}$ of 1.1 and 2.4 µg/mL, respectively, and against MCF-7 human breast adenocarcinoma cancer cell with IC $_{50}$ of 3.4 and 3.1 µg/mL, respectively[28,30]. The cytotoxicity of A. odoratissimus ethanol fruit extract was tested against human liver cancer cells (HepG2), human colon cancer cells (HT-29), and human ovarian cancer cells (Caov3) and was found inactive towards all tested cells[41]. A. altilis methanol pulp extract showed some toxicity against HeLa cells with IC $_{50}$ of 50 µg/mL. Cell proliferation and viability decreased as the concentration of the A. altilis methanol pulp extract increased[56].

Pyranocyloartobiloxanthone A (43) exhibited strong antiproliferative activity against K562 human chronic myeloid leukemia cell with IC $_{50}$ of 0.5 µg/mL and moderate inhibition against HL-60 human promyelocytic leukemia cell with IC $_{50}$ of 2.0 µg/mL and MCF7 positive breast cancer cell with IC $_{50}$ of 5.0 µg/mL in MTT assay[57]. Pyranocycloartobiloxanthone A (43) showed significant

gastroprotective efficacy using ethanol-induced ulcer model in rats. The 50% lethal dosage (LD₅₀) value of pyranocycloartobiloxanthone A (43) was more than 300 mg/kg in acute toxicity analysis. The possible side effect to other organs was also analyzed using liver function test. This study revealed that pre-treatment with pyranocycloartobiloxanthone A (43) significantly protects and reduces gastric mucosa from ethanol-induced gastric lesions as well as restores the depleted gluthione, non-protein sulfhydryl compound and nitric oxide levels in gastric homogenate[51].

Several pieces of research were conducted to investigate the cytotoxicity and inhibitory mechanism of artonin E (10) towards ovarian and breast cancer cells. Rahman et al. reported that artonin E (10) induced antiproliferative effect that led to S phase cell cycle arrest in a time-dependent manner and apoptosis by dysregulating mitochondrial pathways in SKOV-3 ovarian cancer cells[58]. Antiproliferative effects of artonin E (10) on various cell lines were evaluated using MTT assay. The IC₅₀ value for human ovarian adenocarcinoma cells (SKOV-3) was dramatically decreased after 24 h, comparable to the positive control, carboplatin, and paclitaxel. However, the normal human ovarian surface epithelial cells (T1074) showed more resistance towards artonin E. Artonin E also showed potential to inhibit aggressive triple-negative breast cancer cell (MDA-MB-231) by effectively reducing the apoptosis evading capacity, causing a half-maximal growth inhibition at low concentrations (14.3, 13.9 and 9.8 μM) after 24, 48 and 72 h respectively[59]. Furthermore, artonin E (10) helped in delaying quadruple tumor growth by more than 5 days compared to the untreated control group in female mice bearing 4T1 mammary tumors[60].

An *in vitro* study demonstrated jacalin, a lectin purified from protein extracts of *Artocarpus heterophyllus* seeds inhibited the viability of cancer cell MCF7 and H1299. The cancer cell viability was significantly decreased within 24 h upon treatment with purified and standard jacalin. The difference of 10% of cell viability between the purified jacalin and the extracts showed that it is necessary to purify the protein extract. At the highest concentration (10 μ L), the purified and standard jacalin showed almost equal proliferation activity with only 0.98% difference[61].

6.8. Other pharmacological activities

Isolated compounds, 2', 4'-dihydroxy-4-methoxy-3'-prenyldihydrochalcone (1), isobavachalcone (3), artonin E (10), cycloheterophyllin (26), artobiloxanthone (38), cycloartobiloxanthone (39) and artonol B (40) from *A. lowii*, *A. scortechinii*, and *A. teysmanii* were investigated for their ability to inhibit arachidonic acid, collagen, and adenosine diphosphate–induced platelet aggregation in human whole blood. Cycloheterophyllin (26) inhibited arachidonic acid with IC₅₀ of 100.9 μ M and showed strong inhibition against adenosine diphosphate-induced aggregation with IC₅₀ of 57.1 μ M[62].

7. Conclusions and recommendations

This review highlights the phenolic compounds from selected *Artocarpus* in Malaysia, as well as their pharmacological activities. To date, 61 distinct compounds had been isolated from Malaysia's *Artocarpus* and 18 were reported as new ones. Although there are 32 *Artocarpus* species identified in Malaysia, the published articles showed that research in Malaysia focuses on twelve *Artocarpus* species *i.e. A. altilis*, *A. anisophyllus*, *A. elasticus*, *A. fulvicortex*, *Artocarpus heterophyllus*, *A. integer* var. *silvestris*, *A. kemando*, *A. lowii*, *Artocarpus obtusus*, *A. odoratissimus*, *A. scortechinii* and *A. teysmanii*. Based on literature review, *A. altilis* is the most widely utilized *Artocarpus* species. Data gathered highlights the potential of *Artocarpus* as an important source of secondary metabolites to inhibit a certain enzyme, and act as antioxidant, antimicrobial, anti-inflammatory as well as cytotoxic and gastroprotective agents.

Further research such as *in silico* and *in vivo* assay is recommended for better understanding on how the isolated natural products work. Only two *in silico* studies of *Artocarpus* from Malaysia were conducted and valuable data were gathered[27,33]. Pro-oxidant activity of isolated compounds in the presence of copper ions may also be explored as there are evidence associated with anticancer activity[63]. Furthermore, other parts of the plant such as flower can also be explored as there are recent articles that highlighted the flower of *Artocarpus lakoocha* and *Artocarpus heterophyllus* as a source of bioactive compounds[55].

Conflict of interest statement

We declare that there is no conflict of interest.

Acknowledgments

The authors would like to thank the Ministry of Higher Education for the financial support under (Q.J130000.2554.21H57) and the Faculty of Science, Universiti Teknologi Malaysia for sources and research facilities. Lathiff SMA would like to acknowledge Zamalah Universiti Teknologi Malaysia for providing the scholarship.

Funding

This work is supported by the Ministry of Higher Education (Q.J130000.2554.21H57).

Authors' contributions

SMAL and SJ created the concept and designed the structural

and intellectual content. SMAL as the main author involved in the preparation and review of the manuscript. Both SMAL and SJ also involved in the final version of the manuscript. While SMAL and NMA contributed to literature search, data and statistical analysis as well as manuscript editing. SJ as a corresponding author also acted as a guarantor and supervised the project.

References

- [1] Verheij EWM, Coronel RE. Plant resources of South Asia No. 2. Edible fruits and nuts. Pudoc Wageningen Netherlands for Prosea Foundation, Bogor; 1992, p. 86-91.
- [2] Kochummen KM, Go R. Moraceae in tree flora of Sabah and Sarawak. Vol 3. Ampang Press Sdn. Bhd, Kuala Lumpur; 2000, p. 181-212.
- [3] Kochummen KM. Moraceae in tree Flora of Malaya. Vol. 3. Forest Research Institute Malaysia (FRIM), Malaysia; 1978, p. 119-134.
- [4] Kochummen KM. New species and varieties of Moraceae from Malaysia. Gard Bull (Singapore) 1998; 50: 197-219.
- [5] Suwardi AB, Navia ZI, Harmawan T, Syamsuardi, Mukhtar E. Ethnobotany and conservation of indigenous edible fruit plants in South Aceh, Indonesia. *Biodiversitas* 2020; 21(5): 1850-1860.
- [6] Hakim EH, Achmad SA, Juliawaty LD, Makmur L, Shah YM, Aimi N, et al. Prenylated flavonoids and related compounds of the Indonesian Artocarpus (Moraceae). J Nat Med 2006; 60: 161-184.
- [7] Jagtap UV, Bapat VA. Artocarpus: A review of its traditional uses, phytochemistry and pharmacology. J Ethnopharmacol 2010; 129: 142-166
- [8] Perry LM. Medicinal plants of East and Southeast Asia: Attributed properties and uses. The MIT Press, United States; 1980, p. 269-270.
- [9] Pieroni A, Price LL, Vandebroek I. Welcome to Journal of Ethnobiology and Ethnomedicine. J Ethnobiol Ethnomed 2005; 1: 1. Doi: 10.1186/1746-4269-1-1.
- [10]Situmorang ROP, Harianja AH, Silalahi J. Karo's local wisdom: The use of woody plants for traditional diabetic medicines. *Indones J For Res* 2015; 2(2): 121-131.
- [11]World Health Organization. Medicinal plants in the South Pacific: Information on 102 commonly used medicinal plants in the South Pacific. Manila: WHO Regional Office for the Western Pacific; 1998, p. 19-20.
- [12]Labouisse JP. Ethnobotany of breadfruit in Vanuatu: Review and prospects. *Ethnobiol Lett* 2016; **7**(1): 14-23.
- [13]Bradacs G, Heilmann J, Weckerle CS. Medicinal plant use in Vanuatu: A comparative ethnobotanical study of three islands. *J Ethnopharmacol* 2011; 137: 434-448.
- [14]Bourdy G, Walter A. Maternity and medicinal plants in Vanuatu I. The cycle of reproduction. *J Ethnopharmacol* 1992; **37**: 179-196.
- [15]Dossa AJ, Amanoudo JM, Houetchegnon T, Ouinsavi C. Socioeconomic and ethnobotanical importance of the breadfruit tree (*Artocarpus communis* J.G & G. Forster) in Benin. Eur Sci J 2018; 14(24): 447-463.
- [16]Sukumaran S, Sujin RM, Geetha VS, Jeeva S. Ethnobotanical study of medicinal plants used by the Kani tribes of Pechiparai hills, Western Ghats, India. Acta Ecol Sin 2020. [In press].

- [17]Jamil S, Sirat HM, Jantan I, Aimi N, Kitajima M. A new prenylated dihydrochalcone from the leaves of *Artocarpus lowii*. J Nat Med 2008; 62: 321-324
- [18] Abdullah SA, Jamil S, Basar N, Lathiff SMA, Arriffin NM. Flavonoids from the leaves and heartwoods of *Artocarpus lowii* King and their bioactivities. *Nat Prod Res* 2016; 31(10): 1113-1120.
- [19]Jamil S, Lathiff SMA, Abdullah SA, Jemaon N, Sirat HM. Antimicrobial flavonoids from *Artocarpus anisophyllus* Miq. and *Artocarpus lowii* King. *J* Teknol 2014; 71(1): 95-99.
- [20] Lathiff SMA, Jemaon N, Abdullah SA, Jamil S. Flavonoids from Artocarpus anisophyllus and their bioactivities. Nat Prod Commun 2015; 10(3): 393-396.
- [21]Jamil S, Sirat HM, Jantan I. Two new prenylated chalcones from the leaves of *Artocarpus lowii* King. *J Teknol* 2016; **78**(10): 113-116.
- [22]Jamil S, Taher M, Sirat HM, Othman NA. Flavonoids and triterpenes from the leaves of *Artocarpus fulvicortex*. Nat Prod Commun 2012; 7(12): 1587-1588.
- [23]Ramli F, Rahmani M, Kassim NK, Hashim N, Sukari MA, Akim AA, et al. New diprenylated dihydrochalcones from leaves of *Artocarpus elasticus*. *Phytochem Lett* 2013; 6: 582-585.
- [24]Jamil S, Sirat HM, Aimi N, Kitajima M. Flavones from *Artocarpus scortechinii* King. *ACGC Chem Res Commun* 2004; **17**: 3-8.
- [25]Jamil S, Sirat HM, Aimi N, Kitajima M. Flavonoids from *Artocarpus teysmanii* Miq. *Malays J Sci* 2005; **24**: 99-103.
- [26]Ramli F, Rahmani M, Ismail IS, Sukari MA, Rahman MA, Zajmi A, et al. A new bioactive secondary metabolite from *Artocarpus elasticus*. *Nat Prod Commun* 2016; 11(8): 1103-1106.
- [27]Jenis J, Baiseitova A, Yoon SH, Park C, Kim JY, Li ZP, et al. Competitive α-glucosidase inhibitors, dihydrobenzoxanthones, from the barks of *Artocarpus elasticus*. *J Enzyme Inhib Med Chem* 2019; **34**(1): 1623-1632.
- [28]Ee GCL, Teo SH, Rahmani M, Lim CK, Lim YM, Bong CFJ. Artosimmin-A potential anti-cancer lead compound from Artocarpus odoratissimus. Lett Org Chem 2010; 7: 240-244.
- [29]Shamaun SS, Rahmani M, Hashim NM, Ismail HBM, Sukari MA, Ee GCL, et al. Prenylated flavones from Artocarpus altilis. J Nat Med 2010; 64: 478-481
- [30]Ee GCL, Siow HT, Rahmani M, Lim CK, Lim YM, Go R. Artomandin, a new xanthone from *Artocarpus kemando* (Moraceae). *Nat Prod Res* 2011; 25(10): 995-1003.
- [31] Arriffin NM, Jamil S, Basar N, Khamis S, Abdullah SA, Lathiff SMA. Phytochemical studies and antioxidant activities of *Artocarpus scortechinii* King. Rec Nat Prod 2017; 11(3): 299-303.
- [32]Shah MKK, Sirat HM, Jamil S, Jalil J. Flavonoids from the bark of Artocarpus integer var. silvestris and their anti-inflammatory properties. Nat Prod Commun 2016; 11(9): 1275-1278.
- [33]Das S, Laskar MA, Sarker SD, Choudhury MD, Choudhury PR, Mitra A, et al. Prediction of anti-Alzheimer's activity of flavonoids targeting acetylcholinesterase in silico. Phytochem Anal 2017; 28: 324-331.
- [34]Khong HY, Nyokat N, Kutoi CJ, Hamzah AS, Fong IL. Chemical constituents of *Artocarpus odoratissimus* from Sarawak. *J App Pharm Sci* 2017; 7(8): 137-141.

- [35]Shah MKK, Sirat HM, Jamil S. Cholinesterase inhibitors from heartwood of Artocarpus fulvicortex F.M. Jarret (Moraceae). J Teknol 2016; 78(6): 185-189.
- [36]Arriffin NM, Jamil S, Basar N. Two new dihydroflavonols from the leaves of *Artocarpus scortechinii* King. *Phytochem Lett* 2021; **41**: 139-141.
- [37]Hashim N, Rahmani M, Shamaun SS, Ee GCL, Sukari MA, Ali AM, et al. Dipeptide and xanthones from *Artocarpus kemando* Miq. *J Med Plant Res* 2011; 5(17): 4224-4230.
- [38]Hashim N, Rahmani M, Sukari MA, Ali AM, Alitheen N, Go R, et al. Two new xanthones from *Artocarpus obtusus*. J Asian Nat Prod Res 2010; 12(2): 106-112.
- [39]Hashim N, Rahmani M, Shamaun SS, Ee GCL, Sukari MA, Yahayu M, et al. Antioxidant, antimicrobial and tyrosinase inhibitory activities of xanthones isolated from *Artocarpus obtusus* F.M. Jarrett. *Molecules* 2012; 17: 6071-6082.
- [40] Abu Bakar MF, Mohamed M, Rahmat A, Fry JR. Phytochemicals and antioxidant activity of different parts of bambangan (*Mangifera panjang*) and tarap (*Artocarpus odoratissimus*). Food Chem 2009; 113: 479-483.
- [41] Abu Bakar MF, Mohamed M, Rahmat A, Fry J. Cytotoxicity and polyphenol diversity in selected parts *Mangifera panjang* and *Artocarpus* odoratissimus fruits. Nutr Food Sci 2010; 40(1): 29-38.
- [42] Abu Bakar MF, Karim FA, Perisamy E. Comparison of phytochemicals and antioxidant properties of different fruit parts of selected *Artocarpus* species from Sabah, Malaysia. *Sains Malays* 2015; 33(3): 355-363.
- [43] Arriffin NM, Jamil S, Basar N. Antioxidant activities of extracts from the leaves and stem bark of *Artocarpus scortechinii* King. *J Teknol* 2015; 77(2): 1-5.
- [44]Daud MNH, Fatanah DN, Abdullah N, Ahmad R. Evaluation of antioxidant potential of *Artocarpus heterophyllus* L. J33 variety fruit waste from different extraction methods and identification of phenolic constituents by LCMS. *Food Chem* 2017; 232: 621-632.
- [45]Daud MNH, Wibowo A, Abdullah N, Ahmad R. Bioassay-guided fractionation of Artocarpus heterophyllus L. J33 variety fruit waste extract and identification of its antioxidant constituents by TOF–LCMS. Food Chem 2018; 266: 200-214.
- [46]Ee GCL, Teo SH, Go R, Lim CK, Lim YM, Bong CFJ. Free radical scavenging effect of Artocarpus kemando and Artocarpus odoratissimus: Structure activity relationship of flavonoids derivatives. Asian J Chem 2012; 24(1): 231-234.
- [47]Sarian MN, Ahmed QU, So'ad SZM, Alhassan AM, Murugesu S, Perumal V, et al. Antioxidant and antidiabetic effects of flavonoids: A structure activity relationship based study. *Biomed Res Int* 2017; 2017: 8386065.
- [48] Grigalius I, Petrikaite V. Relationship between antioxidant and anticancer activity of trihydroxyflavones. *Molecules* 2017; **22**(12): 2169.
- [49]Rosdi MNM, Sirat HM, Abdullah SA, Jamil S, Muhamad II, Zulkifli RM. Inhibitory effect of Artocarpus lowii King compounds on COX-2 and

- 15-LO activities. J Teknol 2015; 76(1): 299-302.
- [50]Sazali SNM, Jalil J, Arriffin NM, Abdullah SA, Jamil S. In vitro inhibitory effects of flavonoids from the extracts of Artocarpus species on prostaglandin E₂ (PGE₂) production in human plasma. J Innov Pharm Biol Sci 2017; 4(4); 106-111.
- [51]Sidahmed HMA, Hashim NM, Amir J, Abdulla MA, Hadi AHA, Abdelwahab SI, et al. Pyranocycloartobiloxanthone A, a novel gastroprotective compound from Artocarpus obtusus Jarret, against ethanol-induced acute gastric ulcer in vivo. Phytomed 2013; 20: 834-843.
- [52]Zajmi A, Hashim NM, Noordin MI, Khalifa SAM, Ramli F, Ali HM, et al. Ultrastructural study on the antibacterial activity of artonin E versus streptomycin against *Staphylococcus aureus* strains. *PLoS One* 2015; 10(6): e0128157.
- [53]Jamil S, Abdullah SA, Lathiff SMA, Sirat HM. Tyrosinase inhibitory activity of flavonoids from *Artocarpus lowii* King. *J Teknol* 2014; 71(1): 55-58.
- [54] Amir Rawa MS, Hassan Z, Murugaiyah V, Nogawa T, Wahab HA. Anticholinesterase potential of diverse botanical families from Malaysia: Evaluation of crude extract and fractions from liquid-liquid extraction and acid-base extraction. *J Ethnopharmacol* 2019; 245: 112160.
- [55]Gupta AK, Rather MA, Jha AK, Shashank A, Singhal S, Sharma M, et al. Artocarpus lakoocha Roxb. and Artocarpus heterophyllus Lam. flowers: New sources of bioactive compound. Plants 2020; 9(10): 1329.
- [56]Jamil MMA, Ganeson S, Mammam HB, Wahab RA. Artocarpus altilis extract effect on cervical cancer cells. Mater Today 2018; 5: 15559-15566.
- [57]Hashim N, Rahmani M, Ee GCL, Sukari MA, Yahayu M, Oktima W, et al. Antiproliferative activity of xanthones isolated from *Artocarpus obtusus*. J Biomed Biotechnol 2012; 2012: 130627.
- [58]Rahman MA, Ramli F, Karimian H, Deghan F, Nordin N, Ali HM, et al. Artonin E induces apoptosis via mitochondrial dysregulation in SKOV-3 ovarian cancer cells. PLoS One 2016; 11(3): e0151466.
- [59]Etti IC, Abdullah R, Kadir A, Hashim NM, Yeap SK, Imam MU. The molecular mechanism of the anticancer effect of artonin E in MDA-MB 231 triple negative breast cancer cells. *PLoS One* 2017; 12(8): e0182357.
- [60]Etti IC, Abdullah R, Hashim NM, Arifah K, Yeap SK, Sani D, et al. Reduction of breast tumor burden in mice by a prenylated flavonoid, Artonin E. AMJ 2017; 10(8): 681-693.
- [61]Ali ZM, Munaim MSA, Sulaiman WMAW. Effect of lectin from Artocarpus heterophyllus seed on cancer cell lines. J Life Sci Tech 2014; 2(2): 55-59.
- [62]Jantan I, Yasin YHM, Jamil S, Sirat H, Basar N. Effect of prenylated flavonoids and chalcones isolated from *Artocarpus* species on platelet aggregation in human whole blood. *J Nat Med* 2010; 64: 365-369.
- [63]Radapong S, Sarker SD, Ritchie KJ. Oxyresveratrol possesses DNA damaging activity. *Molecules* 2020; 25: 2577.

Siti Mariam A Lathiff, Norzafneza M Arriffin, Shajarahtunnur Jamil $^{\boxtimes}$

Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia,

81310 Johor Bahru, Johor, Malaysia

Corresponding address: shajarah@utm.my

2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone

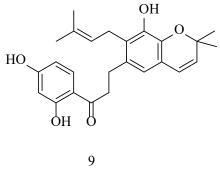
2',4'-Dihydroxy-3,4-(2",2"-dimethylchromeno)-3',-prenyldihydrochalcone

3: R = H; Isobavacalcone 6: R = OH; 2',3,4',4-Tetrahydroxy-3'prenylchalcone

4-Hydroxyonchocarpin

2-Hydroxyparatocarpin

2'-Hydroxy-4,4',6'-trimethoxychalcone



Elastichalcone B

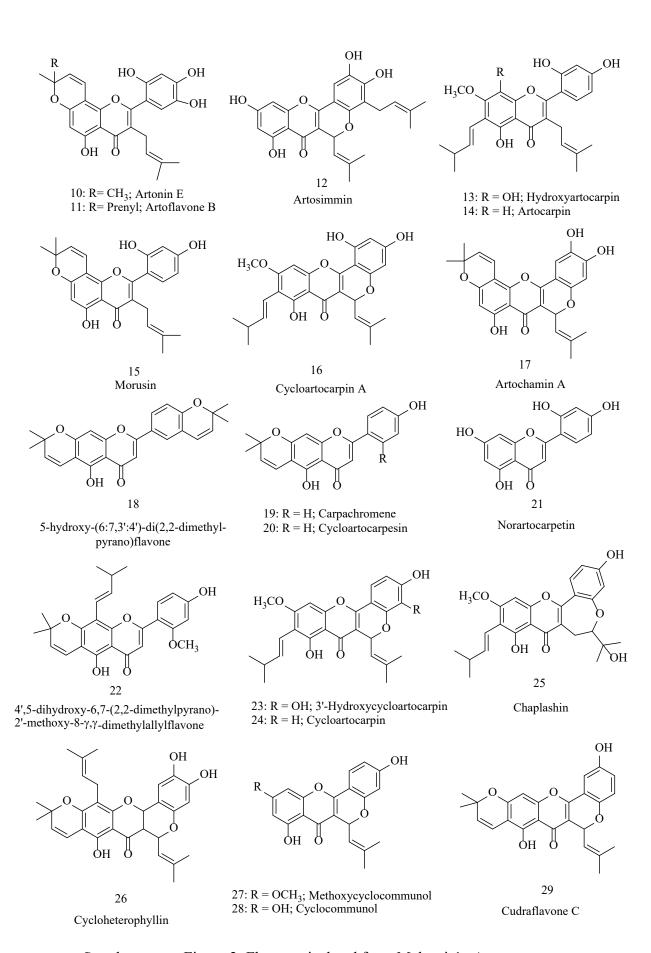
Supplementary Figure 1: Chalcones isolated from Malaysia's Artocarpus

Siti Mariam A Lathiff, Norzafneza M Arriffin, Shajarahtunnur Jamil[™]

Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia,

81310 Johor Bahru, Johor, Malaysia

Corresponding address: shajarah@utm.my



Supplementary Figure 2: Flavones isolated from Malaysia's Artocarpus

Siti Mariam A Lathiff, Norzafneza M Arriffin, Shajarahtunnur Jamil[™]

Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia,

81310 Johor Bahru, Johor, Malaysia

Corresponding address: shajarah@utm.my

5,7-Dihydroxy-4'-methoxy-8-prenylflavanone

5-Hydroxy-7,8-(2,2-dimethyl-chromano)-4'-methoxyflavanone

Heteroflavanone A

Pinostrobin

Pinocembrin

Catechin

Artoscortonol A

Artoscortonol B

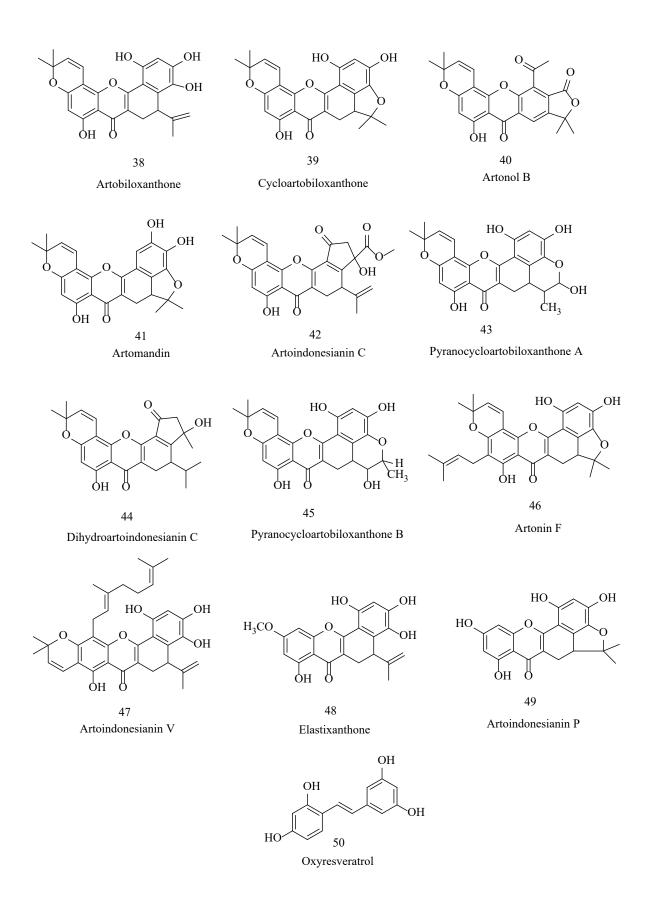
Supplementary Figure 3: Flavanones and Flavanols isolated from Malaysia's Artocarpus

Siti Mariam A Lathiff, Norzafneza M Arriffin, Shajarahtunnur Jamil[™]

Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia,

81310 Johor Bahru, Johor, Malaysia

Corresponding address: shajarah@utm.my



Supplementary Figure 4: Xanthones and Stilbenoid isolated from Malaysia's Artocarpus

Siti Mariam A Lathiff, Norzafneza M Arriffin, Shajarahtunnur Jamil[™]

Department of Chemistry, Faculty of Science, Universiti Teknologi Malaysia,

81310 Johor Bahru, Johor, Malaysia

Corresponding address: shajarah@utm.my

Supplementary Figure 5: Other secondary metabolites isolated from Malaysia's *Artocarpus*