

OPTIMIZATION OF MICROWAVE-ASSISTED EXTRACTION OF PECTIN  
FROM PINEAPPLE PEEL

NOORZETTY AKHTAR BINTI ZAKARIA

A thesis submitted in fulfilment of the  
requirements for the award of the degree of  
Master of Philosophy

School of Chemical and Energy Engineering  
Faculty of Engineering  
Universiti Teknologi Malaysia

DECEMBER 2019

## ACKNOWLEDGEMENT

Praised and thanks to Almighty Allah S.W.T. for giving me the strength, patience and capability to finish this study successfully.

With deep sense of gratitude, I would like to express my sincere thanks to my beloved supervisor, Associate Professor Dr. Roshanida A. Rahman for all her guidance, time and endless support for me in completing this research. Not forgetting my whole gratitude to my co-supervisor, Dr. Dayang Norulfairuz Abang Zaidel for sharing her knowledge, guidance and kindness.

This sincere appreciation and lot of thanks are express to my beloved family members especially my mother and my husband, Sa'idah Taip and Mohamad Haqim Suhaimy for all their encouragement and moral supports. My siblings, Muhammad Zarris Akhmar and Noor Zaireen Akhzar, who always supported me to do my best in all matters of life.

Not forgotten are the laboratory staffs at the N18 School of Chemical and Energy Engineering, Universiti Teknologi Malaysia (UTM) for their assistance and cooperation throughout this research from beginning and until the end. Last but not least to my fellow postgraduate friends for sharing their help and knowledge along the way in completing this research. Without their supports, this research might not be accomplished successfully.

## ABSTRACT

Pineapple peel has the potential to become one of the sources for pectin production due to its high pectin content in its dietary fibre composition. Pectin has been widely used in food industry as food thickener, stabilizer, emulsifier and gelling agent. The conventional extraction process with long operating hours at high temperature has been identified to cause thermal degradation of pectin molecules. Microwave technology application in pectin extraction is able to expedite the extraction process and produce higher yield. Therefore, this study was conducted to optimize the microwave-assisted extraction (MAE) process of pineapple peel pectin. The extracted pectin from optimal condition was subjected to physical characterization and chemical and physicochemical properties evaluation and it was also compared to the commercial pectin. Preliminary studies on the best solvent for pectin extraction from pineapple peel showed that hot solvent sulphuric acid resulted in higher recovery in pectin yield. The optimization of MAE of pectin from pineapple peel was performed using Box-behnken design under the response surface method. The optimum operating conditions obtained were pH 1.52 of solvent, 1:10 (w/v) of solid to liquid ratio and 600 W of microwave power with  $2.43 \pm 0.03$  % (w/w) of pectin yield and  $54.61 \pm 0.21$  % (w/w) of anhydrouronic acid (AUA) content. Under these optimal conditions, the degree of esterification (DE) of pectin was  $63.93 \pm 0.30$  % (w/w). The flow behaviour of pineapple peel pectin solutions at low concentration exhibited the Newtonian behaviour whilst at higher concentration it exhibited the pseudo-plastic behaviour. The elevation of temperature caused the pectin solution viscosity to decrease. The emulsifying activity of extracted pectin was 45 % and the emulsion prepared from pineapple peel pectin has high stability at 4 °C with 93.75 % and 88.24 % at 25 °C, after 30 days of storage. The Fourier transform infrared spectrum showed that the extracted pineapple peel pectin contained the same functional groups that exist in the commercial pectin. Through this study, it can be proven that extraction by using MAE could provide higher quantity and better quality of pectin from pineapple peel. Moreover, pectin produced using pineapple peel has comparable DE value, flow behaviour of pectin solution and emulsifying properties, to the commercial pectin. Hence, pineapple peel can be a potential source for pectin production with some extended studies in sample pre-treatment and pectin purification process.

## ABSTRAK

Kulit nanas mempunyai potensi menjadi salah satu sumber kepada penghasilan pektin kerana kandungan pektin yang tinggi dalam komposisi serat dietnya. Pektin telah digunakan dengan meluas dalam industri makanan sebagai pemekat, penstabil, pengemulsi dan agen penggelen makanan. Proses pengekstrakan konvensional dengan tempoh masa yang panjang pada suhu yang tinggi telah dikenal pasti menyebabkan degradasi haba dalam molekul pektin. Aplikasi teknologi gelombangmikro dalam pengekstrakan pektin menjadikan proses ini lebih cepat serta menghasilkan ekstrak yang lebih tinggi. Oleh itu, kajian ini dijalankan untuk mengoptimumkan pengekstrakan berbantu gelombangmikro (MAE) dalam pengekstrakan pektin daripada kulit nanas. Pektin yang terhasil daripada pengekstrakan yang optimum telah menjalani pencirian fizikal dan penilaian sifat kimia dan fizikokimia, dan potensinya telah dibandingkan dengan pektin komersial. Kajian awal terhadap pelarut terbaik dalam pengekstrakan pektin daripada kulit nanas menunjukkan bahawa larutan asid sulfurik panas memberikan hasil perolehan pektin yang lebih tinggi. Pengoptimuman proses pengekstrakan berbantu MAE pektin daripada kulit nanas telah dilaksanakan dengan menggunakan kaedah gerak balas permukaan menerusi reka bentuk Box-benhknen. Keadaan operasi pengekstrakan optimum yang diperolehi ialah pH 1.52 pelarut, 1:10 (w/v) nisbah pepejal-cecair dan 600 W kuasa gelombangmikro dengan sebanyak  $2.43 \pm 0.03$  % (w/w) hasil pektin dan  $54.61 \pm 0.21$  % (w/w) kandungan asid anhidrouronik (AUA). Pada keadaan optimum, darjah pengesteran (DE) pektin ialah  $63.93 \pm 0.30$  % (w/w). Kelakuan aliran larutan pektin kulit nanas pada kepekatan rendah menunjukkan sifat Newtonian manakala pada kepekatan tinggi ia menunjukkan sifat pseudo-plastik. Peningkatan suhu telah menyebabkan pengurangan pada kelikatan larutan pektin. Aktiviti emulsi pektin yang diekstrak ialah 45 % dan emulsi yang dihasilkan daripada pektin kulit nanas mempunyai kestabilan yang tinggi iaitu 93.75 % pada suhu 4 °C dan 88.24 % pada suhu 25 °C, selepas 30 hari penyimpanan. Spektrum inframerah jelmaan Fourier menunjukkan bahawa pektin kulit nanas mempunyai kumpulan berfungsi yang sama yang terkandung dalam pektin komersial. Kajian ini membuktikan bahawa pengekstrakan dengan menggunakan MAE boleh menghasilkan pektin kulit nanas dalam kuantiti yang lebih banyak dan kualiti yang lebih baik. Tambahan pula, pektin kulit nanas mempunyai kualiti setanding dengan pektin komersial dari segi nilai DE, kelakuan aliran larutan pektin dan sifat-sifat pengemulsi. Oleh itu, kulit nanas berpotensi menjadi sumber penghasilan pektin dengan beberapa kajian lanjutan dalam pra-rawatan sampel dan proses penulenan pektin.

## TABLE OF CONTENT

	<b>TITLE</b>	<b>PAGE</b>
	<b>DECLARATION</b>	<b>ii</b>
	<b>DEDICATION</b>	<b>iii</b>
	<b>ACKNOWLEDGEMENT</b>	<b>iv</b>
	<b>ABSTRACT</b>	<b>v</b>
	<b>ABSTRAK</b>	<b>vi</b>
	<b>TABLE OF CONTENT</b>	<b>vii</b>
	<b>LIST OF TABLES</b>	<b>xi</b>
	<b>LIST OF FIGURES</b>	<b>xiii</b>
	<b>LIST OF ABBREVIATIONS</b>	<b>xvi</b>
	<b>LIST OF SYMBOLS</b>	<b>xvii</b>
<b>CHAPTER 1</b>	<b>INTRODUCTION</b>	<b>1</b>
1.1	Background of Study	1
1.2	Problem Statement	4
1.3	Objectives of Research	6
1.4	Scope of Study	6
1.5	Significant of Study	7
<b>CHAPTER 2</b>	<b>LITERATURE REVIEW</b>	<b>8</b>
2.1	Pectin and its Application	8
2.2	Pineapple Peel	12
2.3	Extraction of Pectin	15
	2.3.1 Dilute Acid Extraction	16
	2.3.2 Alkaline Extraction	18
	2.3.3 Chelating Agent Extraction	19

2.4	Conventional Extraction	20
2.5	Microwave-Assisted Extraction (MAE)	22
2.6	Pectin Characterization	26
2.6.1	Degree of Esterification (DE)	26
2.6.2	Viscosity	27
2.6.3	Emulsifying Properties	28
2.7	Optimization Analysis for Microwave-assisted Extraction (MAE) of Pectin	29
<b>CHAPTER 3</b>	<b>METHODOLOGY</b>	<b>31</b>
3.1	Introduction	31
3.2	Materials	31
3.3	Sample Collection and Preparation	33
3.4	Compositional Analysis of Pineapple Peel	34
3.4.1	Determination of Moisture Content	34
3.4.2	Determination of Ash Content	34
3.4.3	Determination of Fat Content	35
3.4.4	Determination of Total Protein Content	35
3.4.5	Determination of Total Dietary Fibre Content	36
3.5	Pectin Extraction	37
3.5.1	Conventional Extraction of Pectin with Various Type of Solvents	37
3.5.2	Extraction of Pectin by using Microwave-Assisted Extraction	38
3.5.3	Pectin Precipitation and Purification	39

3.6	Optimization of Microwave-Assisted Extraction of Pectin from Pineapple Peel by using Response Surface Method (RSM)	39
3.7	Pectin Characterization	41
3.7.1	Degree of Esterification and Anhydrouronic Acid Analysis	41
3.7.2	Viscosity Test	43
3.7.3	The Emulsifying Properties of Pectin	44
3.7.4	Structural Analysis by Fourier Transform Infrared (FTIR) Spectroscopy	45
3.8	Statistical Analysis	45
<b>CHAPTER 4</b>	<b>RESULTS AND DISCUSSION</b>	<b>46</b>
4.1	Compositional Analysis of Pineapple Peel	46
4.2	Screening in Pectin Extraction	48
4.2.1	Type of Extracting Solvents	48
4.2.2	Factors Influenced in Microwave-assisted Extraction (MAE)	50
4.2.3	Summary of Extraction Process	57
4.3	Optimization of Pectin Extraction Operating Conditions by Response Surface Methodology (RSM)	58
4.3.1	Analysis of Variance (ANOVA) and Statistical Analysis	59
4.3.2	The Effects of Factors on Pectin Yield and Anhydrouronic Acid (AUA) Content of Pineapple Peel Pectin	64
4.3.3	Prediction and Verification of Optimization Point	70

4.4	Comparison Yield of Conventional Extraction and MAE Methods	73
4.5	Pectin Characterization	75
4.5.1	Degree of Esterification (DE)	75
4.5.2	Viscosity Measurement of Pectin Solution	76
4.5.3	Emulsifying Properties of Pectin	79
4.5.4	Spectroscopy Analysis of Pectin by using Fourier Transform Infrared (FTIR)	82
4.5.5	Summary of Pectin Characterization	84
<b>CHAPTER 5</b>	<b>CONCLUSION AND RECOMMENDATION</b>	<b>85</b>
5.1	Conclusion	85
5.2	Recommendation	87
<b>REFERENCES</b>		<b>88</b>



## LIST OF TABLES

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
Table 2.1	Standard specification for pectin	10
Table 2.2	Proximate composition of the pineapple peels	14
Table 2.3	Chemical composition of pineapple shell	14
Table 2.4	The advantageous and disadvantageous of various solvents in pectin extraction	16
Table 2.5	Application of different type of acids in pectin extraction of various raw material	18
Table 2.6	Application of different type of chelating agents in pectin extraction of various raw material	20
Table 2.7	Soxhlet method in pectin extraction	21
Table 2.8	The advantageous and disadvantageous of different method in pectin extraction	22
Table 2.9	Microwave-assisted extraction in pectin extraction	25
Table 3.1	Independent variables for extraction of pectin from pineapple peel obtained from preliminary study	40
Table 3.2	Actual experiment design for pineapple peel pectin extraction by using MAE	41
Table 4.1	Proximate composition of the pineapple peels	48
Table 4.2	Box-Behnken design and the corresponding experimental responses	59
Table 4.3	Analysis of variance for quadratic model in response of pectin yield	61
Table 4.4	Analysis of variance for quadratic model in response of anhydrouronic acid (AUA) content	59
Table 4.5	Optimum experimental conditions for MAE of pectin from pineapple peel	70
Table 4.6	Verification experiments at the optimum process condition	73
Table 4.7	Pectin extraction from pineapple peel at optimum conditions for different extraction methods	74

Table 4.8	Chemical composition of pineapple peel pectin (PPP) by MAE and commercial pectin (CP)	76
Table 4.9	Emulsion activity of oil/0.5 % (w/v) pectin solutions	79
Table 4.10	Emulsion stability of oil/0.5 % (w/v) pectin solutions during storage one day and 30 days at different temperatures	81
Table 4.11	Chemical assignments of FTIR wavenumbers within the range 4000 – 400 cm <sup>-1</sup> of pineapple peel pectin (PPP) and commercial pectin (CP)	83

## LIST OF FIGURES

<b>FIGURE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
Figure 2.1	Structure and composition of plant cell wall	8
Figure 2.2	Pectin molecule and its functional group in repeating unit	9
Figure 2.3	Pectin as milk stabilizer preventing the formation of sedimentation and aggregation of casein	11
Figure 2.4	Schematic diagram of microwave apparatus connected to reflux system	23
Figure 2.5	Water-in-oil (W/O) emulsion and oil-in-water (O/W) emulsion	28
Figure 3.1	The flowchart of overall methodology	32
Figure 3.2	Sample of pineapple peels collected from the waste industry	33
Figure 4.1	Yield of pectin extracted from pineapple peel by using dilute sulphuric acid, citric acid and ammonium oxalate as extraction solvent. Values are mean $\pm$ SE of triplicate analysis; Means with different letters denote significant differences among pectin extracts ( $p < 0.05$ )	50
Figure 4.2	Effect of irradiation time of extraction towards the pectin yield extracted from pineapple peel. Values are mean $\pm$ SE of triplicate analysis; Means with different letters denote significant differences among pectin extracts ( $p < 0.05$ )	52
Figure 4.3	Effect of pH values towards pectin yield extracted from pineapple peel. Values are mean $\pm$ SE of triplicate analysis; Means with different letters denote significant differences among pectin extracts ( $p < 0.05$ )	53
Figure 4.4	Effect of solid to liquid ratio (g/mL) towards pectin yield extracted from pineapple peel. Values are mean $\pm$ SE of triplicate analysis; Means with different letters denote significant differences among pectin extracts ( $p < 0.05$ )	54
Figure 4.5	Effect of different temperatures ( $^{\circ}\text{C}$ ) towards pectin yield extracted from pineapple peel. Values are mean $\pm$ SE of duplicate analysis; Means with different letters denote significant differences among pectin extracts ( $p < 0.05$ )	55

Figure 4.6	Effect of microwave power (W) towards pectin yield extracted from pineapple peel. Values are mean $\pm$ SE of duplicate analysis; Means with different letters denote significant differences among pectin extracts ( $p < 0.05$ )	57
Figure 4.7	Predicted values against experimental data (actual values) for (a) pectin yield and (b) anhydrouronic acid (AUA) content	58
Figure 4.8	Perturbation plot of effect of process variables for (a) pectin yield and (b) AUA content (A: Microwave power (X1); B: pH value (X2); C: S/L ratio (X3))	63
Figure 4.9	Interaction between (a) microwave power and pH value at centre point of 1:20 (w/v) S/L ratio, (b) microwave power and S/L ratio at centre point of pH 2.0 and (c) pH value and S/L ratio at centre point of 500 W microwave power for pectin yield	66
Figure 4.10	The surface response plots of the effects of microwave power, pH value and S/L ratio on the pectin yield (a) constant parameter of S/L ratio at 1:20 (w/v), (b) constant parameter of pH value at pH 2.0 and (c) constant parameter of microwave power at 500 W	67
Figure 4.11	Interaction between (a) microwave power and pH value at centre point of 1:20 (w/v) S/L ratio, (b) microwave power and S/L ratio at centre point of pH 2.0 and (c) pH value and S/L ratio at centre point of 500 W microwave power for AUA content	71
Figure 4.12	The surface response plots of the effects of microwave power, pH value and S/L ratio on the AUA content (a) constant parameter of S/L ratio at 1:20 (w/v), (b) constant parameter of pH value at pH 2.0 and (c) constant parameter of microwave power at 500 W	72
Figure 4.13	The flow behaviour of a dilute solution of PPP at various concentrations and 0.5 % (w/v) of commercial pectin (CP)	77
Figure 4.14	The flow behaviour of 2.0 % (w/v) PPP at different temperatures	78
Figure 4.15	The emulsion activity of oil-in-water emulsions prepared by (a) pineapple peel pectin (PPP) and (b) commercial pectin (CP)	80
Figure 4.16	The emulsion stability of oil-in-water emulsions prepared by (a) PPP and (b) CP at 25 °C temperature and (c) PPP and (d) CP at 4 °C temperature after 30 days of storage	81

Figure 4.17 The Fourier transform infrared spectrum of (a) pineapple peel pectin (PPP) extracted under optimal condition and (b) commercial pectin (CP)

83

## LIST OF ABBREVIATIONS

ANOVA	-	Analysis of Variance
AUA	-	Anhydrouronic Acid
BBD	-	Box-Behnken Design
CCD	-	Central Composite Design
CP	-	Commercial Pectin
DE	-	Degree of Esterification
FTIR	-	Fourier Transform Infrared
GalA	-	Galacturonic Acid
HCl	-	Hydrochloric Acid
H <sub>2</sub> SO <sub>4</sub>	-	Sulphuric Acid
HMP	-	High Methoxyl Pectin
LMP	-	Low Methoxyl Pectin
MAE	-	Microwave-Assisted Extraction
MeO	-	Methoxyl
MW	-	Molecular Weight
NaCl	-	Sodium Chloride
NaOH	-	Sodium Hydroxide
O/W	-	Oil-in-Water
PPP	-	Pineapple Peel Pectin
RSM	-	Response Surface Methodology
S/L	-	Solid to Liquid
UAE	-	Ultrasound Assisted Extraction

## LIST OF SYMBOLS

$\alpha$	-	Alpha
$\beta$	-	Beta
$^{\circ}\text{C}$	-	Degree Celcius
%	-	Percentage
>	-	More than
<	-	Less than
cm	-	Centimeter
g	-	Gram
g/mL	-	Gram per millilitre
mg	-	Miligram
mm	-	Milimeter
mL	-	Mililitre
min	-	Minute
N	-	Normality
pH	-	Power of hydrogen
$R^2$	-	Determination coefficient
rpm	-	Revolutions per minute
s	-	Second
w/w	-	Weight per weight
w/v	-	Weight per volume
W	-	Watt power

# CHAPTER 1

## INTRODUCTION

### 1.1 Background of Study

Pineapple is one of the tropical fruit that can be considered as the most important in the world market and become the top three most preferable fruit juices after orange and apple (Upadhyay *et al.*, 2010). Pineapple can be consumed in various way either by raw itself, cooked in meal or processed into jam or juices. The exportation of pineapple from Malaysia mainly involved fresh fruit or canned product. According to the statistical data by Food and Agriculture Organization (FAO), Malaysia has ranked the 8<sup>th</sup> place in Asia for the greatest pineapple producer after Philippines, Thailand, India, Indonesia, China, Vietnam and Taiwan (FAO, 2017). The pineapple industry shows exportation of canned pineapple increase enormously with 8,474.90 tons metric per year in 2014 to 8,853.37 tons metric per year in 2015 (LPNM, 2016). Demand of pineapple in worldwide also has shown a good growth of production from 2013 to 2014 with 24.5 million tons per year to 25.4 million tons per year of production (FAO, 2017).

Activity from pineapple industry has led to the huge waste production. Wastes from pineapple usually accounted 50 % (w/w) from the whole fruit mass which 29-40 % (w/w) came from peel, 9-10 % (w/w) from core, 2-5 % (w/w) from stem and 2-4 % (w/w) from crown (Ketnawa *et al.*, 2012). This waste has potential to become a renewable resources due to its high biodegradable content. The peel itself can be a promising resources since it is the largest proportion of the waste. According to Ketnawa *et al.* (2012), other than bromelain extraction, the peel could be utilized for further industrial application of its fiber. On the other hand, according to Huang *et al.* (2011), the fiber-rich fractions in pineapple peel potential to be exploited for food functional ingredients. The fiber-rich fractions were said to exhibit



great water- and oil- holding capacity, swelling properties and cation-exchange capacities that can modify the food texture, stabilize foods with high percentage of fat, and destabilize, entrap, and disintegrate the micelles formed by emulsion of lipid (Huang *et al.*, 2011).

Previous studies have been reported that pineapple peel can be a potential source for pectin (Ukiwe and Alinnor, 2011; Karim *et al.*, 2014). Pectin is a complex polysaccharides that made up from monomers of D-galacturonic acid joined by  $\alpha$ -(1-4) glycosidic linkages (Schols and Voragen, 1996). The galacturonic acid (GalA) or anhydrouronic acid (AUA) content in pectin was suggested to be not less than 65 % to be specified as pectin (FAO, 2007). Pectin can be divided into two main groups according to its degree of esterification (DE) which were high-methoxyl pectin (HMP) (DE > 50 %) and low-methoxyl pectin (LMP) (DE < 50 %) which dependent on the source of raw material and method of extraction (Thakur *et al.*, 1997; Wang *et al.*, 2016). Commercial pectin usually produced from either citrus peel or apple pomace. Citrus peel and apple pomace produce different types of pectin which respectively are HMP and LMP that make them preferable to specific applications (May, 1990). Different types of pectin also has different method preparations for gel formation.

Pectin in food industry uses as food thickener, stabilizer, emulsifier and gelling agent (May, 1990). The addition of pectin polysaccharide into foods and beverages also help to improve the mouthfeel and increase the satiety, thus can reduce the consumption of foods and beverages (Thakur *et al.*, 1997). Pectin usually added in fruit jams, confectionary jellies, fruit juices and dairy products. In pharmaceutical industry, pectin has been reported able to lower the cholesterol levels in blood, treat diarrheal disease and duodenal ulcers extensively, and remove mercury and lead from respiratory organs and gastrointestinal tracts effectively. The application of pectin in tablet formulations can act as a binding agent and delayed the drug delivery (Sriamornsak, 2003).

Pectin usually obtained from two steps production which were acid extraction of raw material and isolation of pectin from the extracted solution with alcohol

precipitation or aluminum salts (May, 1990). Commercially, the acid extraction use hot dilute mineral acid such as nitric acid, hydrochloric acid and sulphuric acid at pH around 1-3 with various length of time. There are also some studies reported the uses of organic acid such citric acid and salt such as ammonium oxalate as extracting solvents in order to improve the pectin yield and physicochemical properties of pectin (Mohd. Ismail *et al.*, 2012). The alternative citric acid extractors have been said to carry better in economic value and more environmental friendly (Canteri-Schemin *et al.*, 2005).

The traditional extraction method can be time consuming that took around 6 to 10 hours in soxhlet method and 60 to 120 minutes in hot dilute acid extraction. These method has less efficiency and produce limited yield of pectin. Excessive time of extraction somehow can lead to pectin degradation (Wang *et al.*, 2016). Therefore, new technologies of microwave-assisted extraction (MAE) and ultrasound-assisted extraction (UAE) have been introduced earlier in pectin extraction in order to increase the pectin yield and its quality (Maran *et al.*, 2014; Wang *et al.*, 2016). The MAE technique is a powerful technique and has potential to replace the existing conventional method since it has the ability to extract samples with shorter time of extraction, less solvent consumption and higher rate of extraction with low cost equipment due to its localized heat mechanism (Mandal *et al.*, 2007; Maran *et al.*, 2013). MAE also offered protection to thermolabile constituents and can extract the targeted bioactive compound from raw materials efficiently (Mandal *et al.*, 2007). UAE however required significant longer time of extraction compared to MAE and has less pectin recovery of pectin extraction from dragon fruit peel (Rahmati *et al.*, 2015; Chua *et al.*, 2018). One of the shortcoming in MAE is the loss of active compounds due to thermal degradation from the utilization of high microwave irradiation. However, this problem can be prevented with intermittent microwave extraction. Intermittent extractions can keep out samples from being overheated by balancing the heat and mass transfer besides enhance the efficacy of extraction (Swamy and Muthukumarappan, 2017).

This study was aimed to optimize the pectin extracted from pineapple peel by using MAE method. The extraction method can influence the pectin quality. Hence,

characterization of optimum extract from MAE were performed to evaluate the chemical and physicochemical properties of the extract. Pectin extracted at optimum condition was subjected to determination of AUA and DE value and Fourier Transform Infrared Spectroscopy (FTIR) to compare the chemical composition of extracted pectin with commercial pectin. On the other hand, determination of emulsifying properties and viscosity measurement to prove the ability of the extract to serve as an alternative for commercial pectin.

## 1.2 Problem Statement

Commercialized pectin is usually obtained either from citrus peel or apple pomace after juice extraction. According to May (1990), limited sources for potential food pectin was known due to its standard requirement to contain at least 65 % galacturonic acid of substance. Pectin has been an important food additive in food industry and its demand in world market is currently has reached approximately 45,000 tons annually with 4-5 % growing per annum (Raji *et al.*, 2017). In recent days, many studies have been done to utilize food wastes as an alternative resources for pectin production which are from mango peels (Rehman *et al.* 2004; Berardini *et al.*, 2005), passion fruit peels (Pinheiro *et al.*, 2008; Kliemann *et al.*, 2009; Liew *et al.*, 2014), dragon fruit peels (Mohd Ismail *et al.*, 2012) and jackfruit and chempedak fruit rind (Leong *et al.*, 2016).

Pineapple peel has been said to be an excellent potential in food application due to its composition high in dietary fiber which content was 42.2 % (w/w) to produce pectin (Huang *et al.*, 2011). Several studies on extracting pectin from pineapple peel have been done earlier by Ukiwe and Ainnor (2011), Karim *et al.* (2014) and Sun *et al.* (2014). However, the information of pineapple peel pectin on its purity, physicochemical, structural and functional properties are still unclear. The purity of pectin has become one of the challenges in product market after Food and Agriculture Organization (FAO) has defined the minimum galacturonic acid content required must not be less than 65 % (FAO, 2007). Meanwhile, characterization of pectin is crucial in further assessment to perform an extensive research on its

rheological properties and emulsifying properties to evaluate its potential becoming a marketable product. Apart of its high availability from canned processing activity in this country, pineapple peel waste in return would provide an alternative sources for commercial pectin and thus would help to reduce the number of pollutants in our environment.

Conventionally, it took long operating time of six hours to complete the extraction process of pectin from fruit wastes by using the soxhlet method. The hot dilute mineral acid extraction via water bath on the other hand requires one to two hours of operation that can lead to pectin degradation. Longer time of extraction also would causes more energy and cost consumption of production. Various method of extraction have been studied earlier in order to improve the recovery of valuable crude extracts from the biomass wastes such as microwave-assisted extraction (MAE), ultrasonic-assisted extraction (UAE), pressurized solvent extraction (PSE) and super-critical fluid extraction (SFE). The MAE has been said can be a promising as new extraction technique. The attractive features of MAE to its ability of performance of high and fast extraction, low solvent usage and offered protection to the thermolabile compounds become preferable as an alternative to the conventional method.

Optimization of operating condition for pectin extraction by MAE was important in order to obtain the optimum pectin yield and pectin quality. The quality of pectin was according to the AUA content that referred to the purity of extracted pectin. According to Kute *et al.* (2015), several parameters were considered to be optimized in pectin extraction by MAE which were pH, solid to liquid ratio, microwave power and time. These parameters had gave significant effect on the pectin yield. The optimum condition of MAE obtained in literature by other studies however are restricted since it was limited to a specific microwave system and specific scale that can be used solely as a reference for the extraction by using microwave. This was due to the fact that different microwave system have different efficiency in microwave heating and extraction performance (Chan *et al.*, 2014). Hence, optimization of MAE is required to obtain an optimum condition based on the parameters involved.

### **1.3 Objectives of Research**

This study aims to achieve the objectives:

1. To determine the significant parameters for pectin extracted from pineapple peel.
2. To optimize the pectin yield extracted from pineapple peel by using microwave-assisted extraction.
3. To characterize and evaluate the chemical and physicochemical properties of the extracted pineapple peel pectin obtained from the optimized pectin yield.

### **1.4 Scope of Study**

In order to achieve objectives, the scope of this research has been identified and divided into few parts.

- a) Preliminary study on best method of pectin extraction from pineapple peel was carried out by using three types of solvents which were citric acid, sulphuric acid, and ammonium oxalate. Different parameters (time, pH value, temperature, microwave power and solid to liquid ratio) were screened out in pectin extraction by using microwave-assisted extraction (MAE).
- b) The optimization was performed by using Box-behnken Design (BBD) of Response Surface Methodology (RSM) for three parameters which were microwave power (400, 500 and 600 W), pH value (pH 1.5, 2.0 and 2.5) and solid to liquid ratio (1:10, 1:20 and 1:30 g/mL) in response to pectin yield and anhydrouronic acid (AUA) content.

- c) The pectin obtained from the optimal extraction was analysed its characteristics which were degree of esterification (DE) value and AUA content of pectin, pectin flow behaviour, emulsifying properties of pectin and structural analysis by using Fourier Transform Infrared (FTIR). The pectin characterization was also compared to the commercial pectin in terms of DE and AUA value, flow behaviour of pectin solution, emulsifying properties and structural analysis.

## **1.5 Significant of Study**

The data obtained from the optimized extraction of pectin from pineapple peel by using MAE would be very useful because none of pectin extraction from pineapple peel by using MAE method has been carried out to date. MAE can be a promising and alternative extraction method over conventional to improve better pectin yield and quality.

Furthermore, the data obtained from the characterization of pineapple peel pectin would have benefited to food industries, pharmaceuticals and cosmetics. The data would provide information of the pectin properties and its application in product development. In food industry for instance, type of pectin influenced the product preparation and it is crucial to understand the pectin type to ensure that the food product will be developed successfully according to the desired requirements.

## REFERENCES

- Abang Zaidel, D. N., Hamidon, N. H., and Mat Zahir, N. (2016) 'Extraction and characterization of pectin from sweet potato (*Ipomoea batatas*) peels using alkaline extraction method', In *III International Conference on Agricultural and Food Engineering*, 1152, 211-218.
- Abdullah, and Hanafi Mat, H. (2008) 'Characterisation of solid and liquid pineapple waste', *Reaktor*, 12(1), 48-52.
- AOAC. *Official methods of analysis*. Maryland. Association of Official Analytical Chemists International. (1995).
- AOAC. *Official methods of analysis*. Maryland. Association of Official Analytical Chemists International. (2000)
- Bagherian, H., Ashtiani, F. Z., Fouladitajar, A., and Mohtashamy, M. (2011) 'Comparisons between conventional, microwave- and ultrasound-assisted methods for extraction of pectin from grapefruit', *Chemical Engineering and Processing*, 50, 1237– 1243.
- Barbero, G. F., Palma, M., and Barroso, C. G. (2006) 'Determination of capsaicinoids in peppers by microwave-assisted extraction–high-performance liquid chromatography with fluorescence detection', *Analytica Chimica Acta*, 578(2), 227–233.
- Bartolomé, A. P., Rupérez, P., and Fúster, C. (1995) 'Pineapple fruit: morphological characteristics, chemical composition and sensory analysis of Red Spanish and Smooth Cayenne cultivars', *Food Chemistry*, 53(1), 75-79.
- BeMiller, J. N. (1986) *An Introduction to Pectins: Structure and Properties*, in Fishman, M. L. and Jen, J. J. (eds.), *Chemistry and Function of Pectins*. Washington: American Chemical Society, pp. 2-12.
- Berardini, N., Knödler, M., Schieber, A., and Carle, R. (2005) 'Utilization of mango peels as a source of pectin and polyphenolics', *Innovative Food Science and Emerging Technologies*, 6(4), 442-452.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., and Escaleira, L. A. (2008) 'Response surface methodology (RSM) as a tool for optimization in analytical chemistry', *Talanta*, 76(5), 965–977.

- Bochek, A. M., Zabivalova, N. M., and Petropavlovskii, G. A. (2001) 'Determination of the esterification degree of polygalacturonic acid', *Russian Journal of Applied Chemistry*, 74(5), 796-799.
- Canteri-Schemin, M. H., Fertoni, H. C. R., Waszczynskyj, N., and Wosiacki, G. (2005) 'Extraction of pectin from apple pomace', *Brazilian Archives of Biology and Technology*, 48(2), 259-266.
- Castillo-Israel, K. A. T., Baguio, S. F., Diasanta, M. D. B., Lizardo, R. C. M., Dizon, E. I., and Mejico, M. I. F. (2015) 'Extraction and characterization of pectin from Saba banana [*Musa'saba*(*Musa acuminata* x *Musa balbisiana*)] peel wastes: a preliminary study', *International Food Research Journal*, 22(1), 202-207.
- Chaharbaghi, E., Khodaiyan, K., and Hosseini, S. S. (2017) 'Optimization of pectin extraction from pistachio green hull as a new source', *Carbohydrate Polymers*, 173, 107–113.
- Chan, C.-H., Yusoff, R., and Ngoh, G.-C. (2014) 'Optimization of microwave-assisted extraction based on absorbed microwave power and energy', *Chemical Engineering Science*, 111, 41-47.
- Chawla, R., and Patil, G. R. (2010) 'Soluble dietary fiber', *Comprehensive Reviews in Food Science and Food Safety*, 9(2), 178-196.
- Chen, R. H. and Chen, W. Y. (2001) 'Rheological properties of the water-soluble mucilage of a green laver, *Monostroma nitidum*', *Journal of Applied Phycology*, 13, 481–488.
- Chen, G., and Tao, D. (2005) 'An experimental study of stability of oil–water emulsion', *Fuel Processing Technology*, 86(5), 499–508.
- Chen, J., Liang, R., Liu, W., Luo, S., Liu, C., Wu, S., and Wang, Z. (2014) 'Extraction of pectin from *Premna microphylla* turcz leaves and its physicochemical properties', *Carbohydrate Polymers*, 102, 376–384.
- Chua, B. L., Ng, Y. K., and Ali, A. (2018) 'Ultrasound assisted extraction of pectin from dragon fruit peels', *Journal of Engineering Science and Technology*, 7, 65-81.
- Ciriminna, R., Fidalgo, A., Delisi, R., Ilharco, L. M., and Pagliaro, M. (2016) 'Pectin production and global market', *Agro Food Industry Hi-Tech*, 27(5), 17-20.
- Czitrom, V. (1999) 'One-factor-at-a-time versus designed experiments', *The American Statistician*, 53(2), 126–131.



- Dalev, P. G., and Simeonova, L. S. (1995) 'Emulsifying properties of protein–pectin complexes and their use in oil-containing foodstuffs', *Journal of the Science of Food and Agriculture*, 68(2), 203–206.
- Dhingra, D., Michael, M., Rajput, H., and Patil, R. T. (2012) 'Dietary fibre in foods: a review', *Journal of Food Science and Technology*, 49(3), 255–266.
- Dickinson, E. (2009) 'Hydrocolloids as emulsifiers and emulsion stabilizers', *Food Hydrocolloids*, 23(6), 1473-1482.
- El-Nawawi, S. A., and Shehata, F. R. (1988) 'Effect of the extraction temperature on the quality characteristics of pectin extracted from Egyptian orange peel', *Biological Wastes*, 24(4), 307-311.
- Food and Agriculture Organization (FAO). (1969) Nutrition meetings of the FAO, pp. 133.
- Food and Agriculture Organization (FAO). (2003) *Food energy: methods of analysis and conversion factors*. Report of Technical Workshop (FAO Food and Nutrition paper 77). Food and Agriculture Organization: Rome.
- Food and Agriculture Organization (FAO). (2007) *Compendium of food additive specifications*. Food and Agriculture Organization: Rome.
- Food and Agriculture Organization (FAO). (2017) Food and agriculture data. *FAOSTAT*. [Online]. Available from World Wide Web: from <http://www.fao.org/faostat/en/#data>.
- Faravash, R. S., and Ashtiani, F. Z. (2007) 'The effect of pH, ethanol volume and acid washing time on the yield of pectin extraction from peach pomace', *International Journal of Food Science & Technology*, 42(10), 1177–1187.
- Feumba, D. R., Ashwini Rani, P., and Manohar, R. S. (2016) 'Chemical composition of some selected fruit peels', *European Journal of Food Science and Technology*, 4(4), pp.12-21.
- Fishman, M. L., Chau, H. K., Hoagland, P., and Ayyad, K. (2000) 'Characterization of pectin, flash-extracted from orange albedo by microwave heating, under pressure', *Carbohydrate Research*, 323, 126–138.
- Fishman, M. L., Chau, H. K., Cooke, P. H., and Hotchkiss Jr, A. T. (2008) 'Global structure of microwave-assisted flash-extracted sugar beet pectin', *Journal of Agricultural and Food Chemistry*, 56(4), 1471-1478.
- Ferreira, S. L. C., Bruns, R. E., Ferreira, H. S., Matos, G. D., David, J. M., Brandao, G. C., da Silva, E. G. P., Portugal, L. A., dos Reis, P. S., Souza, A. S., and dos

- Santos, W. N. L. (2007) 'Box-behnken design: an alternative for the optimization of analytical methods', *Analytica Chimica Acta*, 597(2), 179-186.
- Frey, D. D. and Wang, H. (2006) 'Adaptive one-factor-at-a-time experimentation and expected value of improvement', *Technometrics*, 48(3), 418-431.
- Georgiev, Y., Ognyanov, M., Yanakieva, I., Kussovski, V., and Kratchanova, M. (2012) 'Isolation, characterization and modification of citrus pectins', *Journal of BioScience & Biotechnology*, 1(3), 223-233.
- Girma, E., and Worku, T. (2016) 'Extraction and characterization of pectin from selected fruit peel waste', *International Journal of Scientific and Research Publications*, 6(2), 447-454.
- Guo, X., Han, D., Xi, H., Rao, L., Liao, X., Hu, X., and Wu, J. (2012) 'Extraction of pectin from navel orange peel assisted by ultra-high pressure, microwave or traditional heating: a comparison', *Carbohydrate Polymers*, 88(2), 441-448.
- Gupta, B. S., and Ako, J. E. (2005) 'Application of guar gum as a flocculant aid in food processing and potable water treatment', *European Food Research and Technology*, 221(6), 746-751.
- Hemalatha, R., and Anbuselvi, S. (2013) 'Physicochemical constituents of pineapple pulp and waste', *Journal of Chemical and Pharmaceutical Research*, 5(2), 240-242.
- Huang, J. M. G. (1973) 'Improved method for the extraction of pectin', *Florida State Horticultural Society*, 260.
- Huang, Y.-L., Chow, C.-J., and Fang, Y.-J. (2011) 'Preparation and physicochemical properties of fiber-rich fraction from pineapple peels as a potential ingredient', *Journal of Food and Drug Analysis*, 19(3), 318-323.
- Hosseini, S. S., Khodaiyan, F., and Yarmand, M. S. (2016) 'Optimization of microwave assisted extraction of pectin from sour orange peel and its physicochemical properties', *Carbohydrate Polymers*, 140, 59-65.
- Jaji, K., Man, N., and Nawi, N. M. (2018) 'Factors affecting pineapple market supply in Johor, Malaysia', *International Food Research Journal*, 25(1), 366-375.
- Joy, P. P. (2010) 'Benefits and uses of pineapple', *Pineapple Research Station*, Kerala Agricultural University, Kerala, India.
- Kalapathy, U., and Proctor, A. (2001) 'Effect of acid extraction and alcohol precipitation conditions on the yield and purity of soy hull pectin', *Food chemistry*, 73(4), 393-396.

- Kanmani, P., Dhivya, E., Aravind, J., and Kumaresan, K. (2014) 'Extraction and analysis of pectin from citrus peels: augmenting the yield from citrus limon using statistical experimental design', *Iranica Journal of Energy & Environment*, 5(3), 303-312.
- Karim, R., Burhan Uddin, M., and Fahad Jubayer, M. (2014) 'Optimization of pectin isolation method from pineapple (*Ananas comosus* L.) waste', *Carpathian Journal of Food Science and Technology*, 6(2), 116-122.
- Kar, F., and Arslan, N. (1999) 'Effect of temperature and concentration on viscosity of orange peel pectin solutions and intrinsic viscosity–molecular weight relationship', *Carbohydrate Polymers*, 40(4), 277–284.
- Ketnawa, S., Chaiwut, P., and Rawdkuen, S. (2012) 'Pineapple wastes: a potential source for bromelain extraction', *Food and Bioproducts processing*, 90(3), 385-391.
- Khan, M., Bibi, N., and Zeb, A. (2015) 'Optimization of process conditions for pectin extraction from citrus peel', *Science, Technology and Development*, 34 (1), 9-15.
- Kliemann, E., de Simas, K. N., Amante, E. R., Prudêncio, E. S., Teófilo, R. F., Ferreira, M. M. C., and Amboni, R. D. (2009) 'Optimisation of pectin acid extraction from passion fruit peel (*Passiflora edulis flavicarpa*) using response surface methodology' *International Journal of Food Science and Technology*, 44(3), 476-483.
- Koh, P. C., Leong, C. M., and Noranizan, M. A. (2014) 'Microwave-assisted extraction of pectin from jackfruit rinds using different power levels', *International Food Research Journal*, 21(5), 2091-2097.
- Koubala, B. B., Kansci, G., Mbome, L. I., Crépeau, M. - J., Thibault, J. F., and Ralet, M. - C. (2008a) 'Effect of extraction conditions on some physicochemical characteristics of pectins from "Améliorée" and "Mango" mango peels', *Food Hydrocolloids*, 22(7), 1345-1351.
- Koubala, B. B., Mbome, L. I., Kansci, G., Mbiapo, F. T., Crepeau, M. J., Thibault, J. F., and Ralet, M. C. (2008) 'Physicochemical properties of pectins from ambarella peels (*Spondias cytherea*) obtained using different extraction conditions', *Food Chemistry*, 106(3), 1202-1207.
- Kute, A., Mohapatra, D., Babu, B., and Sawant, B. P. (2015) 'Optimization of Microwave Assisted Extraction of Pectin from Orange Peel Using Response

- Surface Methodology’, *Journal of Food Research and Technology*, 3(2), 62-70.
- Kratchanova, M., Pavlova, E., and Panchev, I. (2004) ‘The effect of microwave heating of fresh orange peels on the fruit tissue and quality of extracted pectin’, *Carbohydrate Polymers*, 56(2), 181-185.
- Li, J., Zu, Y.-G., Fu, Y.-J., Yang, Y.-C., Li, S.-M., Li, Z.-N., and Wink, M. (2010) ‘Optimization of microwave-assisted extraction of triterpene saponins from defatted residue of yellow horn (*Xanthoceras sorbifolia* Bunge.) kernel and evaluation of its antioxidant activity’, *Innovative Food Science and Emerging Technologies*, 11(4), 637–643.
- Li, D.-Q., Jia, X., Wei, Z., and Liu, Z.-Y. (2012) ‘Box–Behnken experimental design for investigation of microwave-assisted extracted sugar beet pulp pectin’, *Carbohydrate Polymers*, 88(1), 342–346.
- Liew, S. Q., Chin, N. L., and Yusof, Y. A. (2014) ‘Extraction and characterization of pectin from passion fruit peels’, *Agriculture and Agricultural Science Procedia*, 2, 231-236.
- Lim, B. T., and Khatijah, I. (1995) ‘Determination of dietary fibre in selected fruits and vegetables by prosky method’, *MARDI Res. J.*, 23(2), 191–197.
- Leong, C. M., Noranizan, M. A., Kharidah, M., and Choo, W. S. (2016) ‘Physicochemical properties of pectin extracted from jackfruit and chempedak fruit rinds using various acids’, *International Food Research Journal*, 23(3), 973-978.
- Letellier, M., Budzinski, H., Charrier, L., Capes, S., and Dorthe, A. M. (1999) ‘Optimization by factorial design of focused microwave assisted extraction of polycyclic aromatic hydrocarbons from marine sediment’, *Fresenius’ Journal of Analytical Chemistry*, 364(3), 228–237.
- Levigne, S., Ralet, M.-C., and Thibault, J.-F. (2002) ‘Characterisation of pectins extracted from fresh sugar beet under different conditions using an experimental design’, *Carbohydrate polymers*, 49(2), 145-153.
- Loix, C., Huybrechts, M., Vangronsveld, J., Gielen, M., Keunen, E., and Cuypers, A. (2017) ‘Reciprocal interactions between cadmium-induced cell wall responses and oxidative stress in plants’, *Frontiers in Plant Science*, 8, 1-19.

- López-García, B., Hernández, M., and Segundo, B. S. (2012) 'Bromelain, a cysteine protease from pineapple (*Ananas comosus*) stem, is an inhibitor of fungal plant pathogens', *Letters in Applied Microbiology*, 55(1), 62-67.
- Lembaga Perindustrian Nanas Malaysia (LPNM) *Pengeluaran Buah Nanas* (2016) [Online]. Available from World Wide Web: <http://www.mpib.gov.my/pasaran/>
- Liu, Y., Shi, J., and Langrish, T. A. G. (2006) 'Water-based extraction of pectin from flavedo and albedo of orange peels', *Chemical Engineering Journal*, 120, 203-209.
- Madureira, A. R., Atatoprak, T., Çabuk, D., Sousa, F., Pullar, R. C., and Pintado, M. (2018) 'Extraction and characterisation of cellulose nanocrystals from pineapple peel', *International Journal of Food Studies*, 7, 24–33.
- Mandal, V., Mohan, Y., and Hemalatha, S. (2006) 'Microwave assisted extraction – an innovative and promising extraction tool for medicinal plant research', *Pharmacognosy Reviews*, 1(1), 7-18.
- Maran, J. P., Sivakumar, V., Thirugnanasambandham, K., and Sridhar, R. (2013) 'Optimization of microwave assisted extraction of pectin from orange peel', *Carbohydrate Polymers*, 97(2), 703-709.
- Maran, J. P., Sivakumar, V., Thirugnanasambandham, K., and Sridhar, R. (2014) 'Microwave assisted extraction of pectin from waste *Citrullus lanatus* fruit rinds', *Carbohydrate Polymers*, 101, 786-791.
- Maran, J. P., Swathi, K., Jeevitha, P., Jayalakshmi, J., and Ashvini, G. (2015) 'Microwave-assisted extraction of pectic polysaccharide from waste mango peel', *Carbohydrate Polymers*, 123, 67-71.
- May, C. D. (1990) 'Industrial pectins: sources, production and applications', *Carbohydrate Polymers*, 12(1), 79-99.
- Ministry of Agriculture (MOA) *Nanas* (2003) [Online]. Available from World Wide Web: <http://www.moa.gov.my/nanas>
- Mohd. Ismail, N. S., Ramli, N., Mohd. Hani, N., and Meon, Z. (2012) 'Extraction and characterization of pectin from dragon fruit (*Hylocereus polyrhizus*) using various extraction conditions', *Sains Malaysiana*, 41(1), 41-45.
- Morais, D. R., Rotta, E. M., Sargi, S. C., Bonafe, E. G., Suzuki, R. M., Souza, N. E., Matsushita, M., and Visentainer, J. V. (2017) 'Proximate composition, mineral contents and fatty acid composition of the different parts and dried peels of

- tropical fruits cultivated in Brazil', *Journal of The Brazilian Chemical Society*, 28(2), 308-318.
- Müller-Maatsch, J., Bencivenni, M., Caligiani, A., Tedeschi, T., Bruggeman, G., Bosch, M., Petrusand, J., Droogenbroecke, B. V., Elst, K., and Sforza, S. (2016) 'Pectin content and composition from different food waste streams', *Food Chemistry*, 201, 37-45.
- Nielsen, S. S. (2010) *Food Analysis Laboratory Manual*. 2<sup>nd</sup> Ed. New York: Springer
- Ngouémazong, E. D., Christiaens, S., Shpigelman, A., Van Loey, A., and Hendrickx, M. (2015) 'The emulsifying and emulsion-stabilizing properties of pectin: a review', *Comprehensive Reviews in Food Science and Food Safety*, 14(6), 705-718.
- National Research Council (1989) *Recommended Dietary Allowances*. 10<sup>th</sup> Ed. Washington: The National Academies Press.
- Nor, M. Z. M., Ramchandran, L., Duke, M. and Vasiljevic, T. (2015) 'Characteristic properties of crude pineapple waste extract for bromelain purification by membrane processing', *J Food Sci Technol*, 52(11), 7103–7112.
- Oliveira, T. Í. S., Rosa, M. F., Cavalcante, F. L., Pereira, P. H. F., Moates, G. K., Wellner, N., Mazzetto, S. E., Waldron, K. W., and Azeredo, H. M. (2016) 'Optimization of pectin extraction from banana peels with citric acid by using response surface methodology', *Food Chemistry*, 198, 113-118.
- Owens, H. S., McCready, R. M., Shepard, A. D., Schultz, T. H., Pippen, E. L., Swenson, H. A., Miers, J. C., Erlandsen, R. F., and Maclay, W. D. (1952) *Methods used at Western Regional Research Laboratory for Extraction of Pectic Materials*. United States: Agricultural Research Administration.
- Pan, X., Niu, G., and Liu, H. (2002) 'Comparison of microwave-assisted extraction and conventional extraction techniques for the extraction of tanshinones from *Salvia miltiorrhiza bunge*', *Biochemical Engineering Journal*, 12, 71-77.
- Pagán, J., Ibarz, A., Llorca, M., and Coll, L. (1999) 'Quality of industrial pectin extracted from peach pomace at different pH and temperatures', *Journal of The Science of Food and Agriculture*, 79(7), 1038-1042.
- Pagán, J., Ibarz, A., Llorca, M., Pagan, A., and Barbosa-Cánovas, G. V. (2001) 'Extraction and characterization of pectin from stored peach pomace', *Food Research International*, 34(7), 605-612.

- Pardo, M. E. S., Cassellis, M. E. R., Escobedo, R. M., and García, E. J. (2014) 'Chemical characterisation of the industrial residues of the pineapple (ananas comosus)', *Journal of Agricultural Chemistry and Environment*, 3, 53-56.
- Pasandide, B., Khodaiyan, F., Mousavi, Z. E. and Hosseini, S. S. (2017) 'Optimization of aqueous pectin extraction from *Citrus medica* peel', *Carbohydrate Polymers*, 178, 27-33.
- Pinheiro, E. R., Silva, I. M. D. A., Gonzaga, L. V., Amante, E. R., Teófilo, R. F., Ferreira, M. M. C., and Amboni, R. D. M. C. (2008) 'Optimization of extraction of high-ester pectin from passion fruit peel (*Passiflora edulis* flavicarpa) with citric acid by using response surface methodology', *Bioresource Technology*, 99(13), 5561-5566.
- Rahmati, S., Abdullah, A., Momeny, E., and Kang, O. L. (2015) 'Optimization studies on microwave assisted extraction of dragon fruit (*Hylocereus polyrhizus*) peel pectin using response surface methodology', *International Food Research Journal*, 22(1), 233-239.
- Raji, Z., Khodaiyan, F., Rezaei, K., Kiani, H., and Hosseini, S. S. (2017) 'Extraction optimization and physicochemical properties of pectin from melon peel', *International Journal of Biological Macromolecules*, 98, 709-716.
- Rehman, Z. U., Salariya, A. M., Habib, F., and Shah, W. H. (2004) 'Utilization of mango peels as a source of pectin', *Journal-Chemical Society of Pakistan*, 26, 73-76.
- Renard, C. M., and Thibault, J. F. (1993) 'Structure and properties of apple and sugar-beet pectins extracted by chelating agents', *Carbohydrate Research*, 244(1), 99-114.
- Rodríguez, R., Jimenez, A., Fernández-Bolaños, J., Guillén, R., and Heredia, A. (2006) 'Dietary fibre from vegetable products as source of functional ingredients', *Trends in Food Science and Technology*, 17(1), 3-15.
- Rungrodnimitchai, S. (2011) 'Novel source of pectin from young sugar palm by microwave assisted extraction', *Procedia Food Science*, 1, 1553-1559.
- Sarah, M., Hanum, F., Rizky, M., and Hisham, M. F. Microwave-assisted extraction of pectin from cocoa peel. *IOP Conference Series: Earth and Environmental Science*. February, 2018. IOP Publishing. 1-7.
- Sayah, M. Y., Chabir, R., Benyahia, H., Kandri, Y. R., Chahdi, F. O., Touzani, H., and Errachidi, F. (2016) 'Yield, esterification degree and molecular weight

- evaluation of pectins isolated from orange and grapefruit peels under different conditions’, *PLoS one*, 11(9), 1-16.
- Schols, H. A., and Voragen, A. G. J. (1996) Complex pectins: structure elucidation using enzymes. *Proceedings of an International Symposium*. December 3-7, 1995. The Netherlands: Elsevier. 3-19.
- Shaha, R. K., Punichelvana, Y. N. A., and Afandi, A. (2013) ‘Optimized extraction condition and characterization of pectin from kaffir lime (*Citrus hystrix*)’, *Research Journal of Agriculture and Forestry Sciences*, 1(2), 1-11.
- Simpson, B. K., Egyankor, K. B., and Martin, A. M. (1984) ‘Extraction, purification and determination of pectin in tropical fruits’, *Journal of Food Processing and Preservation*, 2, 63-72.
- Singthong, J., Cui, S. W., Ningsanond, S., and Douglas Goff, H. (2004) ‘Structural characterization, degree of esterification and some gelling properties of Krueo Ma Noy (*Cissampelos pareira*) pectin’, *Carbohydrate Polymers* 58, 391–400.
- Singthong, J., Ningsanond, S., Cui, S. W., and Douglas Goff, H. (2005) ‘Extraction and physicochemical characterization of Krueo Ma Noy pectin’, *Food Hydrocolloids*, 19, 793–801.
- Slavin, J. L., and Lloyd, B. (2012) ‘Health benefits of fruits and vegetables’, *Advances in Nutrition*, 3(4), 506–516.
- Sriamornsak, P. (2003) ‘Chemistry of pectin and its pharmaceutical uses: a review’, *Silpakorn University International Journal*, 3, 206-228.
- Srivastava, P., and Malviya, R. (2011) ‘Sources of pectin, extraction and its applications in pharmaceutical industry – an overview’, *Indian Journal of Natural products and Resources*, 2(1), 10-18.
- Sun, Y., Jing, H., Ren, T.-Q., and Jia, Z.-H. (2014) ‘Research on ultrasonic extraction of pectin in pineapple peel’, *Fujian Journal of Agricultural Sciences*, 29(1), 78-81.
- Swamy, G. J., and Muthukumarappan, K. (2017) ‘Optimization of continuous and intermittent microwave extraction of pectin from banana peels’, *Food Chemistry*, 220, 108–114.
- Tatke, P., and Jaiswal, Y. (2011) ‘An overview of microwave assisted extraction and its applications in herbal drug research’, *Research Journal of Medicinal Plant*, 5(1), 21-31.



- Thakur, B. R., Singh, R. K., Handa, A. K. and Rao, M. A. (1997) 'Chemistry and uses of pectin – a review', *Critical Reviews in Food Science and Nutrition*, 37(1), 47-73.
- Thibault, J. F., and Rombouts, F. M. (1986) 'Effects of some oxidising agents, especially ammonium peroxy sulfate, on sugar-beet pectins', *Carbohydrate Research*, 154(1), 205-215.
- Udonne, J. D., Ajani, O. O., and Akinyemi, O. P. (2016) 'A comparative study of extraction of pectin from wet and dried peels using water based and microwave methods', *International Journal of Scientific and Engineering Research*, 7(3), 416-432
- Ukiwe, L. N., and Alinnor, J. I. (2011) 'Extraction of pectin from pineapple (*Ananas comosus*) peel using inorganic/organic acids and aluminum chloride', *Fresh Produce*, 5(1), 80-83.
- Upadhyay, A., Lama, J. P., and Tawata, S. (2010) 'Utilization of pineapple waste: a review', *Journal of Food Science and Technology Nepal*, 6, 10-18.
- Virk, B. S., and Sogi, D. S. (2004) 'Extraction and characterization of pectin from apple (*Malus Pumila*. Cv Amri) peel waste', *International Journal of Food Properties*, 7(3), 693-703.
- Wandee, Y., Uttapap, D., and Mischnick, P. (2019) 'Yield and structural composition of pomelo peel pectins extracted under acidic and alkaline conditions', *Food Hydrocolloids*, 87, 237–244.
- Wang, L., and Weller, C. L. (2006) 'Recent advances in extraction of nutraceuticals from plants', *Trends in Food Science and Technology*, 17(6), 300-312.
- Wang, S., Chen, F., Wu, J., Wang, Z., Liao, X., and Hu, X. (2007) 'Optimization of pectin extraction assisted by microwave from apple pomace using response surface methodology', *Journal of Food Engineering*, 78(2), 693-700.
- Wang, M., Huang, B., Fan, C., Zhao, K., Hu, H., Xu, X., Pan, S., and Liu, F. (2016) 'Characterization and functional properties of mango peel pectin extracted by ultrasound assisted citric acid', *International Journal of Biological Macromolecules*, 91, 794–803.
- Willats, W. G. ., Knox, J. P., and Mikkelsen, J. D. (2006) 'Pectin: new insights into an old polymer are starting to gel', *Trends in Food Science & Technology*, 17(3), 97–104.

- Yamada, H. Contribution of pectins on health care. *Proceedings of an International Symposium*. December 3-7, 1995. The Netherlands: Elsevier. 173-190.
- Yan, M.-M., Liu, W., Fu, Y.-J., Zu, Y.-G., Chen, C.-Y., and Luo, M. (2010) 'Optimisation of the microwave-assisted extraction process for four main astragalosides in Radix Astragali', *Food Chemistry*, 119(4), 1663–1670.
- Yapo, B. M., Robert, C., Etienne, I., Wathelet, B., and Paquot, M. (2007) 'Effect of extraction conditions on the yield, purity and surface properties of sugar beet pulp pectin extracts', *Food Chemistry*, 100, 1356–1364.
- Yapo, B. M. (2009a) 'Biochemical characteristics and gelling capacity of pectin from yellow passion fruit rind as affected by acid extractant nature', *Journal of Agricultural and Food Chemistry*, 57(4), 1572-1578.
- Yapo, B. M. (2009b) 'Pineapple and banana pectins comprise fewer homogalacturonan building blocks with a smaller degree of polymerization as compared with yellow passion fruit and lemon pectins: implication for gelling properties', *Biomacromolecules*, 10(4), 717-721.
- Yeoh, S., Shi, J. T. A. G., and Langrish, T. A. G. (2008) 'Comparisons between different techniques for water-based extraction of pectin from orange peels', *Desalination*, 218(1-3), 229-237.
- Zhang, B., Yang, R. and Liu, C.-Z. (2008) 'Microwave-assisted extraction of chlorogenic acid from flower buds of *Lonicera japonica* Thunb', *Separation and Purification Technology*, 62(2), 480-483.
- Zhongdong, L., Guohua, W., Yunchang, G., and Kennedy, J. F. (2006) 'Image study of pectin extraction from orange skin assisted by microwave', *Carbohydrate Polymers*, 64, 548–552.
- Ziari, H., Ashtiani, F. Z., and Mohtashamy, M. (2010) 'Comparing the effectiveness of processing parameters in pectin extraction from apple pomace', *Afinidad*, 67(549), 374-379.