

SYNTHESIS AND CHARACTERIZATION OF BIOCERS AS HIGH-PERFORMANCE
BIOSORBENTS FOR DYE REMOVAL PROCESS

WAN NURUL IZYANI BT WAN MOHD ZAWAWI

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (*Chemical Engineering*)

School of Chemical and Energy Engineering
Faculty of Engineering
Universiti Teknologi Malaysia

AUGUST 2019

To my supportive husband, Hafiz, my beautiful children, Husna, Ayra, Adam, and my beloved parents, Wan Mohd Zawawi and Sarah.

ACKNOWLEDGEMENTS

First and foremost, all praises to Allah, the Almighty, on whom ultimately, we depend for sustenance, guidance and also for giving me strength and ability to complete this study.

I would like to express my utmost appreciation to my supervisor Assoc. Prof. Dr. Hanapi bin Mat for his continuous advices, support and time in the completion of this thesis. The guidance I received from him motivated me to push forward and guided me to the correct directions. Special thanks to Dr Helen, Dr Sikin and Dr Fadziyana for their encouragements and helps during my time of needs.

I am grateful to my husband, Hafiz, who have provided me through moral and emotional support throughout my Ph.D. journey. I am also grateful to my father, mother and other family members who have supported me along the way.

I would like to acknowledge the financial support obtained from the Ministry of Higher Education Malaysia (MOHE), MyBrain 15 scholarships and the Fundamental Research Grant Scheme (4F218).

And finally, last but by no means least, to everyone in the AMPEN Lab. It was great sharing laboratory with all of you during last six years.

ABSTRACT

The immobilization of the biological species such as prokaryotic and eukaryotic cell systems (e.g. bacteria, animal and plant cells, and fungi) and cell-derived subunits (e.g. proteins and enzymes) within inorganic oxide matrices as a class of nanocomposite materials especially through sol-gel technique attracts increasing interests for biocatalysis, bioremediation and structured material templates applications. These interests are justified due to the unique approach to explore the richness of the biological structures for technical uses. In this study, Biocers (biologically modified ceramics) of *Trametes versicolor* (TV) embedded in silica matrices was synthesized according to the sol-gel method using tetraethyl orthosilicate (TEOS) as a precursor. The synthesized materials namely free silica (SS), hybrid silica with PVA (hSS), TV Biocers (TVB), hybrid TV Biocers with PVA (hTVB), acclimatized TV Biocers (TVB/AC) and PVA-hybrid acclimatized TV Biocers (hTVB/AC) were characterized using scanning electron microscope, transmission electron microscope, Fourier transform infrared spectroscopy, nitrogen adsorption-desorption measurement and laccase enzyme catalytic activity assay. The performance of the TV Biocers as biosorbents was evaluated using dyes as model emerging organic micropollutants carried out in batch (i.e. shake flask) and continuous (packed-bed) systems. It was observed that the dye removal performance, η (mmol/g) of the TV Biocers for methylene blue (MB) and malachite green (MG) respectively was 7.400 and 5.569 mmol/g which was 18 and 128 % higher than the SS and free TV cells. These results demonstrated that the TV Biocers can offer better dye removal performance through a combination of adsorption and biodegradation processes. The optimization experiment was carried out using MG due to its higher removal performance (adsorption and biodegradation) than MB. The dye removal performance by hTVB was found maximum when operated at pH 6, temperature of 30 °C, 150 rpm of agitation speed, and a biocers/dye ratio of 30 % (w/v) with MG at concentration of 0.5mM. The hTVB was also able to remove MB, methyl orange, and reactive red. The acclimatized TV Biocers (i.e. TVB/AC and hTVB/AC) was studied for dye removal performance in batch and packed-bed process. The highest dye removal by the hTVB/AC was at concentration of 0.2 mM, temperature of 30 °C, and pH 7 for batch process and fastest at concentration of 0.05 mM, bed height of 1 cm and flow rate of 3.0 mL/min for continuous process. The biosorption thermodynamics and kinetics as well as biodegradation kinetic studies were conducted for all biocers (i.e. SS, hSS, TVB, hTVB, TVB/AC and hTVB/AC) for both batch and continuous processes. High correlation coefficients favour Langmuir isotherm and Elovich model for batch and continuous process for biosorption. Meanwhile, the biodegradation fitted the Haldane model well for batch and continuous process. This is the first reported study on the immobilization of the TV cells in silica matrices for removal of emerging organic micropollutants.

ABSTRAK

Immobilisasi spesies biologi seperti sistem sel prokariot dan eukariot (contohnya sel bakteria, sel haiwan, sel tumbuhan, dan kulat) dan subunit sel yang diperolehi (contohnya protein dan enzim) dalam matriks oksida bahan bukan organik sebagai bahan nano terutamanya melalui teknik sol-gel semakin menarik minat untuk dijadikan aplikasi biomangkin, biopemulihan dan templat bahan terstruktur. Ini adalah wajar kerana pendekatannya yang unik dalam meneroka kekayaan struktur biologi bagi kegunaan teknikal. Dalam kajian ini, Biocers (seramik terubahsuai secara biologi) daripada *Trametes versicolor* (TV) tertanam dalam matriks silika telah disintesis mengikut kaedah sol-gel menggunakan tetraetilortosilikat (TEOS) sebagai prapenanda. Bahan-bahan yang disintesis iaitu silika bebas (SS), silika hibrid dengan PVA (hSS), Biocers TV (TVB), Biocers TV hibrid (hTVB), Biocers TV tersuai diri (TVB/AC) dan Biocers TV hibrid-tersuai diri (hTVB/AC) telah dicirikan menggunakan mikroskop imbasan elektron, mikroskop penghantaran elektron, spektroskopi transformasi inframerah Fourier, penjerapan nyah-gerapan nitrogen dan aktiviti pemangkin cerakin pengukuran enzim lakase. Prestasi TV Biocers sebagai bio-penjerap dinilai menggunakan pencelup sebagai model bahan pencemar mikro organik yang dijalankan dalam sistem kelompok (kelalang goncang) dan berterusan (lapisan terpadat). Diperhatikan bahawa prestasi penyingkiran pencelup, η (mmol / g) Biocers TV untuk MB dan MG masing-masing adalah 7,400 dan 5,569 mmol / g di mana 18 % dan 128% lebih tinggi daripada SS dan sel TV bebas. Keputusan ini menunjukkan bahawa Biocers TV boleh menawarkan prestasi penyingkiran pencelup yang lebih baik melalui gabungan proses penjerapan dan biodegradasi. Eksperimen pengoptimuman dijalankan menggunakan MG kerana prestasi penyingkirannya (penjerapan dan biodegradasi) yang tinggi berbanding MB. Penyingkiran pewarna oleh hTVB ditemui maksimum pada pH 6, suhu 30 °C, kelajuan pengadukan 150 rpm, dan nisbah biocer/pencelup sebanyak 30% (w/v) pada kepekatan MG 0.5 mM. hTVB juga dapat menyingkirkan MB, metil jingga dan reaktif merah. TV menyesuaikan diri (TVB/AC dan hTVB/AC) telah diuji untuk prestasi penyingkiran pencelup dalam proses kelompok dan berterusan. Penyingkiran pencelup oleh hTVB / AC adalah tertinggi pada kepekatan 0.2 mM, suhu 30 °C dan pH 7 untuk proses kelompok dan terpanjang pada kepekatan 0.05 mM, ketinggian lapisan 1 cm dan kadar aliran 3.0 ml/min untuk proses berterusan. Termodinamik dan kinetik proses penjerapan dan kajian kinetik biodegradasi dilakukan untuk kesemua biocer (SS, hSS, TVB, hTVB, TVB/AC dan hTVB/AC) bagi kedua-dua proses kelompok dan berterusan. Pekali korelasi yang tertinggi memihak kepada model garis sesuhu Langmuir dan model kinetik Elovich untuk proses kelompok dan berterusan bagi penjerapan. Sementara itu, biodegradasi pula padan dengan model Haldane bagi proses kelompok dan berterusan. Ini adalah kajian yang pertama dilaporkan mengenai immobilisasi sel TV dalam matriks silika bagi penyingkiran bahan pencemar mikro organik.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	TITLE PAGE	i
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xii
	LIST OF FIGURES	xv
	LIST OF ABBREVIATIONS	xix
	LIST OF SYMBOLS	xxi
	LIST OF APPENDICES	xxiii
CHAPTER 1	INTRODUCTION	1
1.1	Background of Research	1
1.2	Problem Statement	3
1.3	Objectives and Scopes of Study	5
1.4	Thesis Outline	8
1.5	Summary	8
CHAPTER 2	LITERATURE REVIEW	11
2.1	Biocers	11
2.1.1	Introduction to Biocers	11
2.1.2	Synthesis of Biocers	12
2.1.3	Properties of Biocers	18
2.1.4	Potential Applications of Biocers	22
2.1.5	Advantages and Disadvantages of Biocers	24
2.1.6	Additive Effects on Biocers	25

2.2	White Rot Fungi Biocers	26
2.2.1	Introduction to White Rot Fungi (WRF)	26
2.2.2	Overview of <i>Trametes versicolor</i> Fungus	31
2.2.3	<i>Trametes versicolor</i> Biocers	34
2.2.4	Potential Applications of <i>Trametes versicolor</i> Biocers	35
2.3	Dye Removal Using Biocers	36
2.3.1	Introduction to Dyes	36
2.3.2	Dye Removal by Biocers	39
2.4	Bioreactors for Biocers	56
2.4.1	Introduction	56
2.4.2	Stirred Tank	57
2.4.3	Fixed-bed Reactor	58
2.4.5	Fluidized Bed	59
2.4.3	Others	60
2.5	Summary	60
CHAPTER 3	MATERIALS AND METHODS	63
3.1	Introduction	63
3.2	Materials	66
3.2.1	Chemicals	66
3.2.2	Strain	66
3.3	Synthesis of TV Biocers	67
3.3.1	Preparation of <i>T. versicolor</i>	67
3.3.2	Acclimatization of <i>T. versicolor</i> in Dye	68
3.3.3	Preparation of the TV Biocers	68
3.4	Characterization of TV Biocers	69
3.4.1	Physical Characterization	70
	3.4.1.1 Surface Morphology	70
	3.4.1.2 Pore and Surface Characteristics	71
3.4.2	Chemical Characterization	71
	3.4.2.1 Functional Group Determination	71
3.4.3	Biological Characterization	72

	3.4.3.1 Optimum Growth of <i>T. versicolor</i>	72
	3.4.3.2 Laccase Catalytic Activity	72
3.5	Dye Removal Performance	73
	3.5.1 Batch Dye Removal Experiments	73
	3.5.2 Continuous Dye Removal Experiments	75
	3.5.3 Modelling of dye removal process	78
3.6	Summary	79
CHAPTER 4	RESULTS AND DISCUSSION	81
4.1	Introduction	81
4.2	Synthesis and Characterization of TV Biocers (TVB)	82
	4.2.1 TV Cells Growth and Catalytic Activity Assay	82
	4.2.2 TV Biocers (TVB) and Free Silica (SS)	85
	4.2.2.1 Physical Properties	85
	4.2.2.2 Chemical Properties	89
	4.2.2.3 Biological Properties	90
	4.2.3 Hybrid TV Biocers (hTVB) and Silica (hSS)	91
	4.2.3.1 Physical Properties	92
	4.2.3.2 Chemical Properties	94
	4.2.3.3 Biological Properties	96
	4.2.4 Dye-acclimatized TV Biocers (TVB/AC) and Hybrid TV Biocers (hTVB/AC)	99
	4.2.4.1 Introduction	99
	4.2.4.2 Physical Properties	99
	4.2.4.3 Chemical Properties	103
	4.2.4.4 Biological Properties	104
4.3	Dye Removal Performance in Batch Process	106
	4.3.1 Batch Dye Removal Parameters	106
	4.3.1.1 Effect of TV Cells	106
	4.3.1.2 Effect of Initial pH	109

	4.3.1.3 Effect of Temperature	113
	4.3.1.4 Effect of Agitation Speed	116
	4.3.1.5 Effect of Initial Dye Concentrations	119
	4.3.1.6 Effect of the Biocers Loading	122
	4.3.1.7 Types of dye	123
	4.3.1.8 Effect of TV Cells Acclimatization in Dye	125
	4.3.2 Modelling of Batch Dye Removal Process	128
	4.3.2.1 Biosorption Thermodynamic Model Analysis	128
	4.3.2.2 Biosorption Kinetic Model Analysis	136
	4.3.2.3 Biodegradation Kinetic Analysis	149
4.4	Dye Removal Performance in Continuous Process	154
	4.4.1 Introduction	154
	4.4.2 Fixed-bed Dye Removal Parameters	154
	4.4.2.1 Effect of Dye Initial Concentration (C_0)	154
	4.4.2.2 Effect of Bed Heights (Z)	157
	4.4.2.3 Effect of Feed Flow Rate (F)	158
	4.4.3 Thermodynamic and Kinetic of Continuous Dye Removal Process	160
	4.4.3.1 Biosorption Thermodynamic Analysis	160
	4.4.3.2 Biosorption Kinetic Model Analysis	163
	4.4.3.3 Fixed-bed Breakthrough Model's Analysis	166
	4.4.3.4 Biodegradation Kinetic Model Analysis	169
4.5	Summary	171

CHAPTER 5	CONCLUSIONS AND RECOMMENDATIONS	173
5.1	Introduction	173
5.2	Summary of Research Findings	173
	5.2.1 Biocers Synthesis and Characterization	173
	5.2.2 Dye Removal Performances of Biocers in Batch Processes	174
	5.2.3 Dye Removal Performances of Biocers in Continuous Process	175
5.3	Recommendations for Future Researchers	176
	5.3.1 Biocers Synthesis and Characterization	176
	5.3.2 Dye Removal Performances of Biocers in Batch Process	177
	5.3.3 Dye Removal Performances of Biocers in Continuous Process	178
5.4	Concluding Remarks	178
REFERENCES		180
APPENDICES		200-255

LIST OF TABLES

TABLE NO.	TITLE	PAGE
Table 2.1	Applications of biocers and methods of preparations	13
Table 2.2	Examples of WRF incorporated with ceramic and their applications	27
Table 2.3	Examples of different WRF with other immobilization supports for pollutant removal purposes.	30
Table 2.4	Examples of pollutants removal by immobilized <i>T. versicolor</i>	33
Table 2.5	Dye removal by microorganism	40
Table 2.6	A summary of dye removal by fungi	41
Table 2.7	Comparison between biosorption, bioaccumulation and biodegradation process (Chojnacka, 2010).	42
Table 2.8	Biosorption isotherm models	44
Table 2.9	A summary of adsorption isotherms of dye removal process	45
Table 2.10	The common adsorption kinetic of diffusion-based models	47
Table 2.11	The common adsorption kinetic of chemical reaction-based models	48
Table 2.12	Summary of adsorption kinetics of dye removal process	49
Table 2.13	Continuous adsorption model.	51
Table 2.14	Various models for biodegradation	55
Table 2.15	Models for dye biodegradation	56
Table 3.1	Parameter studied in batch dye removal performance experiment	74
Table 3.2	Parameter studied in continuous dye removal performance experiment	77

Table 3.3	Linear forms of biosorption isotherms and kinetics	78
Table 4.1	BET surface area (A_s), total pore volume (V_p) and pore diameter (D_p) of the free silica (SS) and TV Biocers (TVB).	87
Table 4.2	BET surface area (A_s), total pore volume (V_p) and pore diameter (D_p) of the hSS and hTVB	94
Table 4.3	BET surface area (A_s), total pore volume (V_p) and pore diameter (D_p) of the TVB/AC and hTVB/AC	100
Table 4.4	Effect of pH on cell mass catalytic activity (U/g). Experimental conditions: initial concentration of MG, 0.2 mM; biocers loading, 30 % (w/v); temperature, 30 °C; and agitation, 150 rpm.	112
Table 4.5	MG dye removal performance and cell mass catalytic activity of TVB/AC and hTVB/AC at different pH, temperatures and initial dye concentrations. Experimental conditions: biocers loading, 30 % (w/v); and agitation, 150 rpm.	127
Table 4.6	Dimensionless constant separation factor, RL of SS, TVB, hSS, hTVB, TVB/AC, and hTVB/AC	131
Table 4.7	Isotherm parameters of MG dye biosorption onto SS, TVB, hSS, hTVB, TVB/AC, and hTVB/AC.	133
Table 4.8	Diffusion coefficients of MG dye biosorption onto biocers	140
Table 4.9	Reaction based kinetics model parameters of MG onto biocers	145
Table 4.10	Biodegradation kinetic parameters of MG dye on TVB, hTVB, TVB/AC and hTVB/AC in batch process	153
Table 4.11	Column data and parameters obtained at different bed heights, flow rates and initial MG dye concentration.	156
Table 4.12	Isotherm constants of MG dye biosorption in fixed-bed reactor.	161
Table 4.13	Kinetic constants of MG dye biosorption onto hSS and hTVB/AC in fixed-bed reactor.	165
Table 4.14	Thomas model parameters of MG dye biosorption onto hSS and hTVB/AC in fixed-bed reactor	166

Table 4.15	Adams-Bohart model parameters of MG dye biosorption onto hSS and hTVB/AC in fixed-bed reactor	168
Table 4.16	Yoon-Nelson model parameters of MG dye biosorption onto hSS and hTVB/AC in fixed-bed reactor	169
Table 4.17	Biodegradation kinetic parameters of MG dye on hTVB/AC in continuous process	171

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
Figure 2.1	Biocers preparation using sol-gel method, BC=biocomponents, living cells (Böttcher <i>et al.</i> , 2004).	15
Figure 2.2	Six types of isotherms and four types of hysteresis loop (Sing and Williams, 2005).	20
Figure 2.3	<i>Trametes versicolor</i> fruit bodies (left) and grown in pelleted morphology (right)	32
Figure 2.4	Examples of chromophoric and auxophoric groups present in organic dyes (International Agency for Research on Cancer, 2010).	37
Figure 2.5	Dye classification on the basis of dye chemical constitution and its application (Nadeem <i>et al.</i> , 2014).	38
Figure 3.1	Flow chart of research methodology	64
Figure 3.2	Design of experiment diagram	65
Figure 3.3	Illustrative diagram on TV Biocer preparation	69
Figure 3.4	Standard calibration curve for malachite green (MG), methylene blue (MB), methyl orange (MO) and reactive red (RR).	75
Figure 3.5	Schematic diagram of the continuous packed-bed reactor.	76
Figure 4.1	(a) Growth kinetics (dry mass) and cell mass catalytic activity of the free TV cells and (b) the growth images of TV cells in medium.	83
Figure 4.2	SEM images of (a) free silica (SS) and (b) TV Biocers (TVB). TEM images of (c) free silica (SS) and (d) TV Biocers (TVB).	86
Figure 4.3	(a) Nitrogen adsorption-desorption isotherm and (b) pore size distribution of free silica (SS) and TV Biocers (TVB).	88
Figure 4.4	FTIR spectra of the free silica (SS) and TV Biocers (TVB).	89
Figure 4.5	The cell mass catalytic activity of TV Biocers (TVB) over incubation time.	91

Figure 4.6	SEM images of (a) hybrid sol-gel silica and (b) hybrid TV Biocers.	92
Figure 4.7	(a) Nitrogen adsorption-desorption isotherm and (b) pore size distribution of free silica (SS) and TV Biocers (TVB)	93
Figure 4.8	FTIR spectra of hybrid sol-gel silica (hSS) and hybrid TV Biocers (hTVB)	95
Figure 4.9	Effect of (a) different biocers (i.e. TVB and hTVB) and (b) PVA loading in hTVB on the cell mass catalytic activity. Experimental conditions: Temperature, 30 °C; [Additive], 0.05 mg/mL; and dosage, 0.3 mg/mL.	97
Figure 4.10	SEM images of (a) acclimatized TV Biocers and (b) hybrid acclimatized TV Biocers	99
Figure 4.11	(a) Nitrogen adsorption-desorption isotherm and (b) pore size distribution of free silica (SS) and TV Biocers (TVB)	102
Figure 4.12	FTIR spectra of hybrid acclimatized TV Biocers (TVB/AC) and hybrid-acclimatized TV Biocers (hTVB/AC)	104
Figure 4.13	Acclimatization of TV in gradient concentration of MG (a) Dye removal performance (b) Catalytic activity.	105
Figure 4.14	The dye removal performance, η (mmol/g) of the methylene blue (MB) and malachite green (MG) by the TV Biocers (TVB), free silica (SS), and free TV cells.	107
Figure 4.15	Effect of pH on MG removal performance activity at the 5 th day of incubation. Experimental conditions: initial concentration of MG, 0.2 mM; biocers loading, 30 % (w/v); temperature, 30 °C; and agitation speed, 150 rpm.	110
Figure 4.16	Effect of temperature on (a) MG removal performance and (b) cell mass catalytic activity at 5 th day of incubation. Experimental conditions: initial concentration of MG, 0.2 mM; biocers loading, 30 % (w/v); agitation, 150 rpm; and pH, 6.0.	115
Figure 4.17	Effect of agitation speed on (a) MG removal performance and (b) cell mass catalytic activity. Experimental conditions: initial concentration of MG, 0.2 mM; temperature, 30 °C; biocers loading, 30 % (w/v); and pH, 6.0.	118

Figure 4.18	Effect of initial concentration of dye on MG removal performance. Experimental conditions: biocers loading, 30 % (w/v); temperature, 30 °C; agitation, 150 rpm; and pH, 6.0.	120
Figure 4.19	Effect of initial concentration of dye on cell mass catalytic activity. Experimental conditions: biocers loading, 30 % (w/v); temperature, 30 °C; agitation: 150 rpm; and pH, 6.0.	121
Figure 4.20	Effect of biocers loading on MG dye removal performance. Experimental conditions: initial concentration of MG, 0.2 mM; temperature, 30 °C; agitation: 150 rpm; and pH, 6.0.	122
Figure 4.21	Effect of various types of dye on (a) dye removal performance and (b) cell mass catalytic activity. Experimental conditions: initial concentration of dye, 100 mM; biocers loading, 30 % (w/v); temperature: 30 °C; agitation: 150 rpm; and pH, 6.0.	124
Figure 4.22	Effect of initial MG dye concentration biosorption onto SS, hSS, TVB, hTVB, TVB/AC and hTVB/AC	129
Figure 4.23	Biosorption isotherms of MG onto (a) SS, (b) hSS, (c) TVB, (d) hTVB, (e) TVB/AC and (f) hTVB/AC	134
Figure 4.24	Biosorption capacity of SS, hSS, TVB, hTVB, TVB/AC and hTVB/AC. Experimental conditions: initial concentration of MG, 0.2 mM; temperature, 30 °C; biocers loading, 30 % (w/v); and pH, 7.0.	136
Figure 4.25	Biosorption kinetics of MG dye onto SS, hSS, TVB, hTVB, TVB/AC and hTVB/AC	147
Figure 4.26	Biodegradation capacity of TVB, hTVB, TVB/AC and hTVB/AC. Experimental conditions: initial concentration of MG, 0.2 mM; temperature, 30 °C; biocers loading, 30 % (w/v); and pH, 7.0.	149
Figure 4.27	A comparison between experimental data and biodegradation kinetic model prediction for MG dye onto TVB, hTVB, TVB/AC and hTVB/AC in batch processes	152
Figure 4.28	Effect of initial concentration of dye on continuous MG removal performance. Experimental conditions: sample mass, 0.5 g, bed height, 1 cm and flow rate, 3 mL/min.	156

Figure 4.29	Effect of bed height on continuous MG removal performance. Experimental conditions: MG initial concentration, 0.1 mM and flow rate, 3 mL/min.	157
Figure 4.30	Effect of flow rate on continuous MG removal performance. Experimental conditions: MG initial concentration, 0.1 mM and bed height, 1 cm.	159
Figure 4.31	Experimental and isotherm models' prediction for MG dye removal onto (a) hTVB/AC and (b) hSS.	162
Figure 4.32	Experimental and kinetic models' prediction of MG dye biosorption onto (a) hTVB/AC and (b) hSS.	164
Figure 4.33	A comparison between experimental data and biodegradation kinetic model prediction for MG dye onto hTVB/AC in continuous process.	170

LIST OF ABBREVIATIONS

AB	-	Adam-Bohart
ABTS	-	Azino-bis (3-ethylbenzoline-6-sulfonic acid) diammonium
ATCC	-	American Tissue Culture Collection
AU	-	Activity Unit
BET	-	Brunauer-Emmett-Teller
BJH	-	Barrett-Joyner-Helenda
C	-	Carbon
DMP	-	Dimethoxyphenol
EDCs	-	Endocrine Disrupting Compounds
FTIR	-	Fourier Transform Infrared Spectroscopy
H ₂ O	-	Water
HCl	-	Hydrochloric Acid
hSS	-	Hybrid silica with PVA
hTVB	-	Hybrid TV Biocers with PVA
hTVB/AC	-	Hybrid acclimatized (in dye) TV Biocers with PVA
LiP	-	Lignin Peroxidase
MB	-	Methylene blue
MG	-	Malachite green
MO	-	Methyl orange
MnP	-	Manganese Peroxidase
N	-	Nitrogen
NaCl	-	Sodium Chloride
NAD	-	Nitrogen Adsorption/ Desorption
NaOH	-	Sodium Hydroxide
O	-	Oxygen
OTC	-	Oxytetracycline
PAHs	-	Polyaromatic Hydrocarbons
PCBs	-	Polychlorinated Biphenyls
PCPs	-	Personal Care Products
PFAs	-	Polyfluorinated Alkylated Substances
PFO	-	Pseudo-first order

PSO	-	Pseudo-second order
RR	-	Reactive red
SEM	-	Scanning Electron Microscope
SiO ₂	-	Silica Oxide
Si-OH	-	Silanol group
SOLAC	-	Sol-gel Laccase
SS	-	Free silica
STP	-	Standard Temperature Pressure
TEA	-	Triethylamine
TEM	-	Transmission Electron Microscope
TEOS	-	Tetraethoxysilane
TV	-	<i>Trametes versicolor</i>
TVB/AC	-	Acclimatized (in dye) TV Biocers
TVB	-	TV Biocers
UTM	-	Universiti Teknologi Malaysia
UV	-	Ultraviolet
WRF	-	White Rot Fungi
YN	-	Yoon-Nelson

LIST OF SYMBOLS

μ	-	micro
A_t	-	Temkin constant (L/g)
β_L	-	External mass transfer coefficient in liquid phase (cm/s),
b_t	-	Temkin isotherm constant (J/mol)
C_{AO}	-	Initial concentration of biosorbents (mmol/ml)
C_A	-	Concentration of biosorbents (mmol/ml)
C_e	-	Concentration at equilibrium (mmol/L)
C_t	-	Concentration at time t (mmol/L)
C_0	-	Initial concentration (mmol/L)
D_{eff}	-	Effective liquid film diffusion coefficient (cm/h)
D_f	-	Film diffusion coefficient (m ² /h)
D_p	-	Particles diameter (nm)
F	-	Ratio of the amount of dye adsorbed at time t
F	-	Flow rates
g	-	gram
k_1	-	rate constant of the pseudo-first order kinetic model
k_2	-	rate constant of the pseudo-second order kinetic model
k_a	-	the sorption equilibrium constant
K_F	-	Freundlich constant related to the biosorption capacity (L/mol)
K_L	-	Langmuir constant related to the biosorption capacity (L/mol)
K_{id}	-	Intraparticle diffusion constant
N_T	-	External mass transfer
τ	-	Time required to achieve 50% breakthrough
η	-	Dye removal performance
q_{exp}	-	Experimental data biosorption capacity
q_{cal}	-	Model calculated biosorption capacity
Q_e	-	Equilibrium biosorption capacity (mmol/g),
Q_{max}	-	Maximum biosorption capacity
Q_t	-	Biosorption capacity at time t
r	-	radial position
R	-	Universal gas constant (8.314 J/mol.K)

R_L	-	Dimensionless constant separation factor
r^2	-	Linear regression coefficient for isotherm and kinetic models
T	-	Absolute temperature (298 K)
U	-	Laccase activity
ΔG^0	-	Gibb free energy (kJ/mol)
ΔH^0	-	Enthalpy change (kJ/mol)
ΔS^0	-	Entropy change (kJ/mol)
V_{pore}	-	Pore volume (cm ³ /g)
1/n	-	Biosorption intensity of the biosorbents
δ	-	The thickness of the film layer (cm)
φ	-	Associate parameter of water (2.60)
M_B	-	Molecular weight of water (18 g/mol)
μ_f	-	Viscosity of water (0.89)
V_A	-	liquid molar volume at normal boiling temperature
mL	-	milliliters
mg	-	milligrams
L	-	Liters
°C	-	Degree Celcius
cm	-	centimeters
%	-	percents
x^2	-	Linear regression coefficient for isotherm and kinetic models
ε	-	Absorptivity of ABTS
Z	-	Bed height

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Characterization of Biocers Data	201
B	Batch Dye Removal Performance Data	224
C	Batch Performance Modelling Fitting	231
D	Continuous Dye Removal Performance Data	246
E	Continuous Performance Modelling Fitting	252

CHAPTER 1

INTRODUCTION

1.1 Background of Research

Dye is a coloured substance that has an affinity to the substrate to which it is being applied. The dye is generally applied in an aqueous solution, and may require a mordant to improve the fastness of the dye on the fibre (Booth *et al.*, 2000). There are quite a few ways to classify dyes. For example, they may be classified based on charge characteristics, such as cationic, anionic and non-ionic dyes. The organic dyes can also be classified according to their chemical structure into groups such as azo compounds, anthraquinones, triarylmethanes and phthalocyanines. Dye can also be classified according to their solubility and chemical properties, for instance acid dyes, disperse dyes, direct dyes, reactive dyes, mordant dyes, vat dyes and sulphur dyes. The use of different dyes is dependent on the characteristics of the fibre, the specific colour to be applied and the desired outcome required on the fibre.

Over the past century, dyes play a significant role in the economy of many countries. Dyes have been widely used in many industries such as textiles, foods, plastics, leathers and pulps. Dyes are one of the most abundant organic pollutants that are produced from various industries and can be found in our wastewater treatment. The release of coloured wastewater from fast growing industries such as textile, paper and plastic may present eco-toxic hazard and may eventually lead to a high environmental impact if not treated effectively beforehand. For instance, malachite green is a most commonly used for dyeing of cotton, silk, paper, leather and also in manufacturing of paints and printing inks. Most of the dyes, including malachite green, are toxic and must be removed before discharge into receiving streams (Srivastava *et al.*, 2004). These important sectors use large amounts of process water and produce great amounts of polluted discharge. Water is used for cleaning the raw material and for many flushing steps during the whole production.

Produced wastewater should be cleaned from fat, oil, colour and other chemicals, which were used during the several production steps.

The discharge of coloured effluents from various industries without decolourization procedure may cause serious problems in the receiving environments. Depending on the exposure time and dye concentration, dyes may have acute and/or chronic effects on exposed organisms. Dyes may absorb and reflect sunlight that entering the water, affecting the growth of organisms thus interferes with the food chain. Both public and authorities are very aware with a slight abnormal colouration of surface waters (Shah and Patel, 2014). Moreover, the presence of even a small fraction of dyes in water is highly visible due to high colour or tinctorial value of dyes. This will have an effect on the aesthetic value of the water resources. Common waste treatment procedure unable to remove dye due to their complex aromatic structure that is resistant to light, biological activity, ozone and other degradative environments (Joshi *et al.*, 2004). The possible long-term effects of a few dyes and dye degradation products are becoming of increasing concern. Some dye effluents are found mutagenic, carcinogenic and/or allergenic effects (Alves de Lima *et al.*, 2007). The highest rates of toxicity were found amongst basic and diazo dyes (Yahagi *et al.*, 1975). Furthermore, dyes have also an effect on photosynthetic activity in aquatic life by reducing light penetration (Lavanya, 2014) and may also be toxic to certain forms of aquatic life due to presence of metals and chlorides in them (Khayat-zadeh and Abbasi, 2010). Dyes have also been known to interfere with certain municipal wastewater treatment operations such as UV disinfection (Ramakrishna and Viraraghavan, 1996).

Due to these possible drawbacks of dyes, many practices for wastewater decolourization treatment had been applied including physical, chemical and biological treatment. Several techniques for dye removal have been developed, and some have been widely employed. However, the conventional single methods are not efficient enough, thus, there is always a need for designing new materials that can adsorb and degrade dye with high removal performance capacity and selectivity as well as cost efficient.

1.2 Problem Statement

Immobilization is an attractive way for biological components physiological capabilities enhancement. Biocomponents such as enzymes or cells are connected to a surface by self-adhesion or chemical bonding or entrapped in the interstices of fibrous or porous materials or physically linked within or by solid or porous matrices. This may lead to increased resistance on changes in environment such as pH or temperature. However, an efficient system to avoid the problem of fixing biocomponents or biomolecules firmly without altering their original conformations and activities is still challenging for the utilization of biochemical functions of active biocatalyst. A recent trend in the field of bio-engineered materials is the research of nanocomposites, where bioactive compounds are embedded or immobilized contained by inorganic nanostructured oxide matrices (Soltmann *et al.*, 2003).

Mixing a ceramic-like oxide matrix with biological system (“biocers”) presents several returns. The main advantage of biocers is the mutual influence of the mechanical, chemical, thermal, and photochemical stability of the inorganic host matrix with the high variability of the sol-gel method (e.g., chemical modifications, tailored porosity) and the extensive range of sol-gel derived materials (e.g., coatings, granules, shaped-bulk products) enhance the technical applications of immobilization (Soltmann and Böttcher, 2008). Silica-derived ceramic matrices employment can ensure the viability of the encapsulated living cells. Silica is toxicologically and biologically inert and usually not a food source of microorganism and thus makes it suitable to be incorporated with biological components such as cell. It was demonstrated that biocers is practical to be used in many applications such as biosensors, bioremediation, structured material templates, at least at laboratory scale (Flickinger *et al.*, 2009). Moreover, the Biocers that employ sol gel matrix protects the biocatalyst and prevents cell lysis, thus lead to activity and stability preservation and improvement. It has been reported that the fine porosity of sol-gel allows nutrients to reach the cell and by-products to escape so that the physiological capabilities of the cells can be enhanced (Guan *et al.*, 2008). The favourable surface/mass ration, being inertness, mild operative conditions and mechanical strength or organically modified sol-gel materials are always a purpose why sol-gel method is

selected. On the contrary to organic polymeric matrices, silica matrices are inert and more resistant to microbial attack and contamination. Furthermore, the sol-gel process involving mild reaction conditions, because the chemical reactivity of precursor compounds accomplishes the free energy requirement leading to Si-O-Si network as a stable reaction product (Carturan *et al.*, 2004).

Despite all benefits of using biocers in several applications, there is no report of synthesis of biocers comprised of *Trametes versicolor* fungus using sol-gel method. Many studies of biocers using one of the most extensively used enzyme; laccase is widely reported (Mohidem and Mat, 2009a, 2009b, 2010a, 2010b, and 2011; Zhang *et al.*, 2013; Galliker *et al.*, 2010; Manna and Amutha, 2017). However, these studies only used the derived sub-unit components from the fungus (enzyme), not the fungus itself. The usage of fungus as whole is suggested as to give more benefits rather than used a single enzyme. The loss of biological activity of an enzyme during immobilization or while it is in use is possible and thus using whole cell can counter this problem. The immobilized cells possess regeneration capability and are particularly suitable for multiple enzymatic reactions. The advantage of using cell instead of enzyme include the need to purify is not needed, the cell system is less sensitive to operating conditions changes such as pH, and higher loading support could also be investigated. Using fungus, particularly gives several advantages especially based on its principal mechanism that comprise of biosorption, bioaccumulation and biodegradation. The three mechanism are hugely beneficial as to give extra credits to ceramic that solely make use of biosorption only.

Sometimes, immobilization draws issues like mass transfer limitations due to changes in structure. Thus, the incorporation of additive to the biocers was proposed to overcome this issue. Additive able to protect the cell by preparing a shield-like between the protein and its environment. Additives are believed to provide additional sites for hydrogen bonding with the enzyme surface, decreasing dehydration and provides barrier for enzymes from unfolding by covering the interface (Villalonga *et al.*, 2000). The addition of additive process is simple, fast and economic for enhancement of enzyme and cell stability. The immobilized cell activity can also be enhanced in the presence of additive. Several chemical reagents used as an additive

include surfactants, polyhydric alcohols, methyl esters, and metal ions. Some additive for examples polyvinyl alcohol (PVA) and polyethylene glycol (PEG) were proven to protects enzymes from denaturing effects without affecting their reaction rates (Soares *et al.*, 2002). The PVA and PEG have higher molecular weight, thus able to cover a large superficial area. This will help to improve the cell activity due to conformational and structural changes with the decrease in surface coverage (Wehtje *et al.*, 1993). In general, the addition of additive during immobilization may improve operational stability (Zhang *et al.*, 2014).

The synthesized biocers is studied for removal of dye as model emerging organic pollutants. Although selected dye stimulates biological reactions at low concentrations, most of dye are considered toxic to microorganisms at moderate concentrations and can cause inhibitory effects on the biological processes. Therefore, it is important that microorganisms were first acclimatized to dye before immobilization and used for further treatment process. The acclimatization of fungi in dye may influence the further removal dye studies. During acclimatization, the fungi was adjusted to a gradual change it is environment to allow them to a maintain performance and adhering changes across a range of environmental conditions. The interference from the functional group from the dye pollutant that may altered the nutrient dependency of the fungi thus affect the metabolic capacity and behavior of the fungus (Rajeswari *et al.*, 2013).

1.3 Objectives and Scopes of Study

The objectives and scopes of this research are:

- i. To synthesize and characterize the TV Biocers

The *Trametes versicolor* (TV) strain ATCC 42530 was obtained from the American Type Culture Collection. The free silica (SS) and TV Biocers (TVB) will be prepared using tetraethyl orthosilicate (TEOS) as a silica precursor using sol-gel method. Sol is a stable suspension of colloidal solid particles in a liquid while gel is a

porous, three-dimensional and continuous solid network surrounding a continuous liquid phase. The sol-gel process involves conversion of monomers into colloidal solution (sol) that acts as the precursor for an integrated network (gel) and comprises of five steps namely hydrolysis, condensation, gelation, ageing and drying. Hydrolysis occurs when TEOS and water are mixed and produces an intermediate silanols (Si-OH). In the subsequent step, condensation takes place to form bridging oxygen or a siloxane group Si-O-Si. As the sol aggregates the viscosity will increase until a gel is formed. Biological component was added at pre-gelation process. The sol-gel transition (gelation) is reached when a continuous network is formed and as the viscosity rapidly increase, the solvent is trapped inside the gel (ageing). The drying process was then taking place in order to remove remaining liquid (solvent) phase. During this step, a significant amount of shrinkage and densitification occur. The porosity of the gel was strongly determined by the rate of which the solvent can be removed.

The hybrid free silica (hSS) and hybrid TV Biocers (hTVB) were prepared using TEOS as a silica precursor and additive to further improve the catalytic activity and stability of sol-gel silica. The effect of additives types and loadings was investigated. Two different additives (PVA and PEG) were introduced at pre-gelation process in order to further protect the cell and enzyme from aggregation and unfolding effects. The TV cell was also acclimatized in dye before immobilized in sol-gel silica (TVB/AC). The dye used for acclimatization was the same as the dye tested for dye removal performance, which is malachite green (MG). The MG concentration was increased gradually from 10 mg/L until 50 mg/L, each for 7 days of incubation period. After each time interval, the TV cells were harvested and used as inoculation for subsequent dye concentration. The procedures were similar to the cell entrapment in objective number 1, except that TV cell was mixed with additives at prior gelation.

The synthesized SS, hSS, TVB, hTVB, and TVB/AC were characterized by a scanning electron microscope (SEM), transmission electron microscope (TEM), Fourier transform infrared (FTIR) spectrophotometer, nitrogen adsorption/desorption (NAD analyzer and cell mass catalytic activity measurement for further

understanding of physical and biological properties of synthesized TV Biocers. The cell mass catalytic activity was determined by using 2,2'-azino-bis (3-ethylbenzoline-6-sulfonic acid) diammonium salt (ABTS) as a substrate. A comparison study of free and immobilized cell catalytic activity was also conducted.

ii. To evaluate the batch dye removal performance of TV Biocers

The performance of SS, hSS, TVB, hTVB and TVB/AC were carried out in Erlenmeyer flask as batch reactor using malachite green (MG) dyes as emerging organic micropollutants model. Dye removal performance was investigated at various initial pH, temperature, initial dye concentrations, agitation speed, substrate/medium ratio, and other type of dye such as reactive red (reactive-azo dye), methylene blue (cationic heteropolyaromatic dye) and methyl orange (anionic-azo dye). The removal of dye was studied in terms of biosorption and degradation. Removal of dyes was analyzed using adsorption isotherm (Langmuir, Freundlich and Temkin) and kinetic (pseudo-first order, pseudo-second order, Elovich and diffusion models) for biosorption process and biodegradation kinetic model namely Haldane, Aiba and Edward kinetic model for biodegradation process.

iii. To evaluate the continuous dye removal performance of TV Biocers

The continuous dye removal performance was evaluated using fixed-bed adsorber. The performance was described through the concept of breakthrough concentration curves. The effect of bed height, flow rate and initial dye concentration was investigated. The cumulative biosorption data was analyzed using isotherm and kinetic models. The existing isotherm models such as Langmuir, Freundlich and Temkin isotherm models were used to analyse biosorption isotherm data. The pseudo-first order, pseudo-second order and Elovich kinetic models were used for the biosorption kinetic data analysis. The performance of the fixed-bed adsorber was analyzed using Thomas, Adam-Bohart and Yoon-Nelson models. The dye removal data were also analyzed using existing biodegradation kinetic model namely Haldane, Aiba and Edward kinetic model.

1.4 Thesis Outline

This thesis contains five chapters. Chapter 1 presents research introduction, problem background, objectives and scopes of this study, thesis outline and chapter summary. Literature reviews on micropollutants, biodegradation, immobilization, biocers, white-rot fungi *Trametes versicolor* and sol-gel technology are presented in Chapter 2. Chapter 3 discusses about research methodology which comprises research materials and experimental procedures including TV Biocers preparation and characterization and biodegradation testing evaluation. The results and discussions of TV Biocers synthesis and characterizations, evaluation of dye removal process and performances were presented in Chapter 4. Chapter 5 presents the conclusions of the study and recommendations for future work. This is followed by the list of references cited in the thesis.

1.5 Summary

Colour has always played a significant part in our lives, including presenting different cultures of human being all over the world, influencing the clothes we wear and the furnishings that we used. However, the existence of dye in environment especially in water can cause impacts in human health as well as to the environment. This motivated research and development scholars all over the world to explore new method to minimize the dye environmental impacts. The removal of dye through various physical, chemical and biological processes has been applied in removing dye. However, the effectiveness, pros and cons are varies depending on the types of dye, cost, procedures and environment condition. Combining physical and biological process shows promising result in removing dye. In this study, *Trametes versicolor* (known as laccase enzyme producing fungi) cells are immobilized in silica using sol-gel method to remove dye at environmentally relevant concentrations.

The immobilization of *Trametes versicolor* in inorganic matrices (biocers) using sol-gel method incorporated with additive has so far not been reported. Growth condition, cell morphology and physiology, physical and chemical properties,

enzyme activity and chemical stability of cell immobilization in biocer was studied. The dye removal performance by TV Biocers has also not been reported which it will thus be the subject of the present studies.

REFERENCES

- Abdel-Jabbar, N., Al-asheh, S. and Jordan, I. (2009) 'Factorial Design for the Analysis of Packed-bed Sorption of Copper using Eggshell as a Biosorbent', *J. Environ. Prot. Sci.*, 3, 133–139.
- Acevedo, F., Pizzul, L., Castillo, M., González, M. E., Cea, M., Gianfreda, L. and Diez, M. C. (2010) 'Degradation of polycyclic aromatic hydrocarbons by free and nanoclay-immobilized manganese peroxidase from *Anthracophyllum discolor*', *Chemosphere*, 80(3), 271–278.
- Addleman, K., Dumonceaux, T., Paice, M. G., Bourbonnais, R. and Archibald, F. S. (1995) 'Production and characterization of *Trametes versicolor* mutants unable to bleach hardwood kraft pulp', *Appl. Environ. Microbiol.*, 61(10), 3687–3694.
- Aiba, S., Shoda, M. and Nagatani, M. (1968) 'Kinetics of product inhibition in alcohol fermentation', *Biotechnol. Bioeng.*, 10(6), 845–864.
- Akar, S. T., Akar, T. and Çabuk, A. (2009) 'Decolorization of a textile dye , reactive red 198 (RR198), by *Aspergillus parasiticus* fungal biosorbent', *Braz. J. Chem. Eng.*, 26(2), 399–405.
- Aksu, Z. and Gönen, F. (2004) 'Biosorption of phenol by immobilized activated sludge in a continuous packed bed: Prediction of breakthrough curves', *Process Biochem.*, 39(5), 599–613.
- Ali, H. (2010) 'Biodegradation of synthetic dyes - A review', *Water, Air, and Soil Pollution*, 213 (1–4) 251–273.
- Allothman, Z. A. (2012) 'A review: Fundamental aspects of silicate mesoporous materials', *Materials*, 5 (12) 2874–2902.
- Alves de Lima, R. O., Bazo, A. P., Salvadori, D. M. F., Rech, C. M., de Palma Oliveira, D. and de Aragão Umbuzeiro, G. (2007) 'Mutagenic and carcinogenic potential of a textile azo dye processing plant effluent that impacts a drinking water source', *Mutat. Res. Genet. Toxicol. Environ. Mutagen*, 626(1–2), 53–60.
- Amoura, M., Nassif, N., Livage, J. and Coradin, T. (2007) 'Sol-gel encapsulation of cells is not limited to silica : bacteria long- term viability in alumina matrices', *RSC*, 1–4.

- Archibald, F., Bourbonnais, R., Jurasek, L., Paice, M. and Reid, I. (1997) 'Kraft pulp bleaching and delignification by *Trametes versicolor*', *J. Biotechnol.*, 53(2–3), 215–236.
- Aretxaga, A., Romero, S., Sarra, M. and Vicent, T. (2001) 'Adsorption Step in the Biological Degradation of a Textile Dye', *Biotechnol. Prog.*, 17(4), 664–668.
- Arica, M. Y., Kaçar, Y. and Genç, Ö. (2001) 'Entrapment of white-rot fungus *Trametes versicolor* in Ca-alginate beads: Preparation and biosorption kinetic analysis for cadmium removal from an aqueous solution', *Bioresour. Technol.*, 80(2), 121–129.
- Arica, M. Y., Arpa, Ç., Kaya, B., Bektaş, S., Denizli, A. and Genç, Ö. (2003) 'Comparative biosorption of mercuric ions from aquatic systems by immobilized live and heat-inactivated *Trametes versicolor* and *Pleurotus sajur-caju*', *Bioresour. Technol.*, 89(2), 145–154.
- Arica, M. Y., Kaçar, Y. and Genç, Ö. (2001) 'Entrapment of white-rot fungus *Trametes versicolor* in Ca-alginate beads: preparation and biosorption kinetic analysis for cadmium removal from an aqueous solution', *Bioresour. Technol.*, 80(2), 121–129.
- Bagchi, M. and Ray, L. (2015) 'Adsorption behavior of Reactive Blue 4, a tri-azine dye on dry cells of *Rhizopus oryzae* in a batch system', *Chem. Speciat. Bioavailab.*, 27(3), 112–120.
- Bajpai, P., Mehna, A. and Bajpai, P. K. (1993) 'Decolorization of kraft bleach plant effluent with the white rot fungus *Trametes versicolor*', *Process Biochem.*, 28(6), 377–384.
- Balarak, D., Mostafapour, F. K. and Azarpira, H. (2016) 'Biosorption of reactive blue 19 dye using *Lemna minor*: Equilibrium, kinetic and thermodynamic studies'. *Environ. Commun. Biosci. Biotech. Res. Comm. Thomson Reuters ISI ESC Crossref Index. J. NAAS J. Score.*, 9(3), 558–566.
- Bayramoğlu, G., Bektaş, S. and Arica, M. Y. (2003) 'Biosorption of heavy metal ions on immobilized white-rot fungus *Trametes versicolor*', *J. Hazard. Mater.*, 101(3), 285–300.
- Benderdouche, N., Bestani, B. and Hamzaoui, M. (2018) 'The use of linear and nonlinear methods for adsorption isotherm optimization of basic green 4-dye onto sawdust-based activated carbon'. *J. Mater. Environ. Sci.*, 9(4), 1110–1118.

- Benhassine, S., Kacem, C. N. and Destain, J. (2016) 'Production of laccase without inducer by *Chaetomium* species isolated from Chettaba forest situated in the East of Algeria', *African J. Biotechnol.*, 15(7), 207–213.
- Berrazoum, A., Marouf, R., Ouadjenia, F. and Schott, J. (2015) 'Bioadsorption of a reactive dye from aqueous solution by municipal solid waste', *Biotechnol. Reports*, 7, 44-50.
- Binupriya, A. R., Sathishkumar, M., Dhamodaran, K., Jayabalan, R., Swaminathan, K. and Yun, S. E. (2007) 'Liquid-phase separation of reactive dye by wood-rotting fungus: A biotechnological approach', *Biotechnol. J.*, 2(8), 1014–1025.
- Booth, G., Zollinger, H., McLaren, K., Sharples, W. G. and Westwell, A. (2000) 'Dyes, General Survey', *Int. Ullmann's Encyclopedia of Industrial Chemistry*, 8676–8694.
- Borchert, M. and Libra, J. A. (2001) 'Decolorization of reactive dyes by the white rot fungus *Trametes versicolor* in sequencing batch reactors', *Biotechnol. Bioeng.*, 75(3), 313–321.
- Böttcher, H., Soltmann, U., Mertig, M. and Pompe, W. (2004) 'Biocers: ceramics with incorporated microorganisms for biocatalytic, biosorptive and functional materials development', *Journal of Materials Chemistr.*, 14 (14) .2176-2193.
- Bumpus, J. A. and Aust, S. D. (1987) 'Biodegradation of environmental pollutants by the white rot fungus *Phanerochaete chrysosporium*: Involvement of the lignin degrading system', *BioEssays*, 6(4), 166–170.
- Burden, D. (2012) 'Guide to the Disruption of Biological Samples', *Random Prim.*, 25(12), 1–25.
- Cabuk, A., Unal, A. T. and Kolankaya, N. (2006) 'Biodegradation of cyanide by a white rot fungus, *Trametes versicolor*', *Biotechnol. Lett.*, 28(16), 1313–7.
- Cai, P.-J., Xiao, X., He, Y.-R., Li, W.-W., Chu, J., Wu, C., He, M.-X., Zhang, Z., Sheng, G.-P., Lam, M. H.-W., Xu, F. and Yu, H.-Q. (2012) 'Anaerobic biodecolorization mechanism of methyl orange by *Shewanella oneidensis* MR-1', *Appl. Microbiol. Biotechnol.*, 93(4), 1769–1776.
- Campostrini, R., Carturan, G., Caniato, R., Piovan, a., Filippini, R., Innocenti, G. and Cappelletti, E. M. (1996) 'Immobilization of plant cells in hybrid sol-gel materials', *J. Sol-Gel Sci. Technol.*, 7(1–2), 87–97.

- Carabajal, M., Perullini, M., Jobbágy, M., Ullrich, R., Hofrichter, M. and Levin, L. (2016) 'Removal of Phenol by Immobilization of *Trametes versicolor* in Silica-Alginate-Fungus Biocomposites and Loofa Sponge', *Clean - Soil, Air, Water*, 44(2), 180–188.
- Carturan, G., Dal Toso, R., Boninsegna, S. and Dal Monte, R. (2004a) 'Encapsulation of functional cells by sol-gel silica: actual progress and perspectives for cell therapy', *J. Mater. Chem.*, 14(14), 2087–2098.
- Carturan, G., Dal Toso, R., Boninsegna, S. and Dal Monte, R. (2004b) 'Encapsulation of functional cells by sol-gel silica: actual progress and perspectives for cell therapy', *Journal of Materials Chemistry*, 14 (14) 2087-2098.
- Cea, M., Jorquera, M., Rubilar, O., Langer, H., Tortella, G. and Diez, M. C. (2010) 'Bioremediation of soil contaminated with pentachlorophenol by *Anthracophyllum discolor* and its effect on soil microbial community', *J. Hazard. Mater.*, 181(1–3), 315–323.
- Çelekli, A., Yavuzatmaca, M. and Bozkurt, H. (2012) 'Binary Adsorption of Reactive Red 120 and Yellow 81 on *Spirogyra majuscula*. 13(i), 29–36.
- Chacko, J. T. and Subramaniam, K. (2011). Enzymatic Degradation of Azo Dyes – A Review', *Int. J. Environmetal Sci.*, 1(6), 1250–1260.
- Chakraborty, S., Basak, B., Dutta, S., Bhunia, B. and Dey, A. (2013) 'Decolorization and biodegradation of congo red dye by a novel white rot fungus *Alternaria alternata* CMERI F6', *Bioresour. Technol.*, 147, 662–666.
- Charumathi, D. and Das, N. (2012) 'Packed bed column studies for the removal of synthetic dyes from textile wastewater using immobilised dead *C. tropicalis*', *Desalination*. 285, 22–30.
- Chen, J. P. and Lin, W. S. (2003) 'Sol-gel powders and supported sol-gel polymers for immobilization of lipase in ester synthesis', *Enzyme Microb. Technol.*, 32(7), 801–811.
- Chivukula, M. and Renganathan, V. (1995) 'Phenolic azo dye oxidation by laccase from *Pyricularia oryzae*', *Appl. Environ. Microbiol.*, 61(12), 4374–4377.
- Chonde Sonal, G., Chonde Sachin, G., Bhosale, P. ., Nakade, D. B. and Raut, P. D. (2012) 'Studies on degradation of synthetic polymer Nylon 6 by fungus *Trametes versicolor* NCIM 1086', *Int. J. Environ. Sci.*, 2(3), 2435–2442.

- Chowdhury, S. and Saha, P. Das (2013) 'Adsorption of malachite green from aqueous solution by naoh-modified rice husk: Fixed-bed column studies', *Environ. Prog. Sustain. Energy.*, 32(3), 633–639.
- Çorman, M. E., Öztürk, N., Bereli, N., Akgöl, S. and Denizli, A. (2010) Preparation of nanoparticles which contains histidine for immobilization of *Trametes versicolor* laccase', *J. Mol. Catal. B Enzym.*, 63(1–2), 102–107.
- Crini, G. and Badot, P. M. (2008) 'Application of chitosan, a natural aminopolysaccharide, for dye removal from aqueous solutions by adsorption processes using batch studies: A review of recent literature'. *Prog. Polym. Sci.*, 33(4), 399–447.
- Daâssi, D., Mechichi, T., Nasri, M. and Rodriguez-Couto, S. (2013) Decolorization of the metal textile dye Lanaset Grey G by immobilized white-rot fungi', *J. Environ. Manage.*, 129, 324–332.
- Dada, A. O., Olalekan, A. P., Olatunya, A. M. and Dada (2012) 'Langmuir, Freundlich, Temkin and Dubinin-Radushkevich Isotherms Studies of Equilibrium Sorption of Zn 2+ Unto Phosphoric Acid Modified Rice Husk', *J. App. Chem.*, 3(1), 38-45.
- de Kreij, A., van den Burg, B., Venema, G., Vriend, G., Eijsink, V. G. H. and Nielsen, J. E. (2002) 'The Effects of Modifying the Surface Charge on the Catalytic Activity of a Thermolysin-like Protease', *J. Biol. Chem.*, 277(18),15432–15438.
- Dehghanifard, E., Jonidi Jafari, A., Rezaei Kalantary, R., Mahvi, A. H., Faramarzi, M. A. and Esrafil, A. (2013) 'Biodegradation of 2,4-dinitrophenol with laccase immobilized on nano-porous silica beads', *Iranian J. Environ. Health Sci. Eng.*, 10(1), 25-34.
- Dhakar, K. and Pandey, A. (2013) 'Laccase Production from a temperature and pH tolerant fungal strain of *Trametes hirsuta* (MTCC 11397)', *Enzyme Res.*, 2013, 1–9.
- Dickson, D. J. and Ely, R. L. (2011) 'Evaluation of encapsulation stress and the effect of additives on viability and photosynthetic activity of *Synechocystis* sp. PCC 6803 encapsulated in silica gel', *Appl. Microbiol. Biotechnol.*, 91(6), 1633–1646.
- Dorado, A. D., Gamisans, X., Valderrama, C., Solé, M. and Lao, C. (2014) 'Cr(III) removal from aqueous solutions: a straightforward model approaching of the

- adsorption in a fixed-bed column', *J. Environ. Sci. Health. A. Tox. Hazard. Subst. Environ. Eng.*, 49(2), 179–186.
- Dos Santos, A. B., Bisschops, I. A. E., Cervantes, F. J. and Van Lier, J. B. (2004) 'Effect of different redox mediators during thermophilic azo dye reduction by anaerobic granular sludge and comparative study between mesophilic (30°C) and thermophilic (55°C) treatments for decolourisation of textile wastewaters', *Chemosphere*, 55(9), 1149–1157.
- Dursun, A. Y. and Tepe, O. (2011) 'Removal of Chemazol Reactive Red 195 from aqueous solution by dehydrated beet pulp carbon', *J. Hazard. Mater.*, 194, 303–311.
- Ehlers, G. A. and Rose, P. D. (2005) 'Immobilized white-rot fungal biodegradation of phenol and chlorinated phenol in trickling packed-bed reactors by employing sequencing batch operation', *Bioresour. Technol.*, 96(11), 1264–1275.
- Einarsrud, M. A., Nilsen, E., Rigacci, A., Pajonk, G. M., Buathier, S., Valette, D., Durant, M., Chevalier, B., Nitz, P. and Ehrburger-Dolle, F. (2001) 'Strengthening of silica gels and aerogels by washing and aging processes', *J. Non. Cryst. Solids*, 285(1–3), 1–7.
- El-Enshasy, H. A. (2007) 'Filamentous Fungal Cultures-Process Characteristics, Products, and Applications', *New Technol. App.*, 225–261.
- El-Naas, M. H., Al-Muhtaseb, S. A and Makhlof, S. (2009) 'Biodegradation of phenol by *Pseudomonas putida* immobilized in polyvinyl alcohol (PVA) gel', *J. Hazard. Mater.*, 164(2–3), 720–725.
- Enayatizamir, N., Tabandeh, F., Rodríguez-Couto, S., Yakhchali, B., Alikhani, H. A. and Mohammadi, L. (2011) 'Biodegradation pathway and detoxification of the diazo dye Reactive Black 5 by *Phanerochaete chrysosporium*', *Bioresour. Technol.*, 102(22), 10359–10362.
- Erden, E., Kaymaz, Y. and Pazarlioglu, N. K. (2011a) 'Biosorption kinetics of a direct azo dye Sirius Blue K-CFN by *Trametes versicolor*', *Electron. J. Biotechnol.*, 14(2), 8-18.
- Erkurt, E. A., Ünyayar, A. and Kumbur, H. (2007) 'Decolorization of synthetic dyes by white rot fungi, involving laccase enzyme in the process', *Process Biochem.*, 42(10), 1429–1435.
- Evstatieva, Y., Yordanova, M., Chernev, G., Ruseva, Y. and Nikolova, D. (2014) 'Sol-gel immobilization as a suitable technique for enhancement of α -amylase

activity of *Aspergillus oryzae* PP', *Biotechnol. Equip.* 28(4), 728–732.

- Fernando Bautista, L., Morales, G. and Sanz, R. (2010) 'Immobilization strategies for laccase from *Trametes versicolor* on mesostructured silica materials and the application to the degradation of naphthalene', *Bioresour. Technol.*, 101(22), 8541–8548.
- Fiedler, D., Hager, U., Franke, H., Soltmann, U. and Bottcher, H. (2007) 'Algae biocers: astaxanthin formation in sol-gel immobilised living microalgae', *J. Mater. Chem.*, 17(3), 261–266.
- Fomina, M. and Gadd, G. M. (2014) 'Biosorption: Current perspectives on concept, definition and application', *Bioresour. Technol.*, 160, 3–14.
- Font, X., Caminal, G., Gabarrell, X., Romero, S. and Vicent, M. T. (2003) 'Black liquor detoxification by laccase of *Trametes versicolor* pellets', *J. Chem. Technol. Biotechnol.*, 78(5), 548–554.
- Font, X., Caminal, G., Gabarrell, X. and Vicent, T. (2006) 'Treatment of toxic industrial wastewater in fluidized and fixed-bed batch reactors with *Trametes versicolor*: influence of immobilisation', *Environ. Technol.*, 27(8), 845–854.
- Fourest, E. and Volesky, B. (1997) 'Alginate Properties and Heavy Metal Biosorption by Marine Algae', *Appl. Biochem. Biotechnol.*, 67(3), 215–226.
- Fragoheiro, S. and Magan, N. (2008) 'Impact of *Trametes versicolor* and *Phanerochaete chrysosporium* on differential breakdown of pesticide mixtures in soil microcosms at two water potentials and associated respiration and enzyme activity', *Int. Biodeterior. Biodegrad.*, 62(4), 376–383.
- Furusaki, S. (1988) 'Engineering aspects of immobilized biocatalysts', *J. Chem. Eng. Japan*, 21(3), 219–230.
- Galliker, P., Hommes, G., Schlosser, D., Corvini, P. and Shahgaldian, P. (2010) 'Laccase-modified silica nanoparticles efficiently catalyze the transformation of phenolic compounds', *J. Colloid Interface Sci.*, 349(1), 98–105.
- Gao, D., Du, L., Yang, J., Wu, W.-M. and Liang, H. (2010a) 'A critical review of the application of white rot fungus to environmental pollution control', *Crit. Rev. Biotechnol.*, 30(1), 70–77.
- Gao, S., Wang, Y., Diao, X., Luo, G. and Dai, Y. (2010b) 'Effect of pore diameter and cross-linking method on the immobilization efficiency of *Candida rugosa* lipase in SBA-15', *Bioresour. Technol.*, 101(11), 3830–3837.

- García-Galán, M. J., Rodríguez-Rodríguez, C. E., Vicent, T., Caminal, G., Díaz-Cruz, M. S. and Barceló, D. (2011) 'Biodegradation of sulfamethazine by *Trametes versicolor*: Removal from sewage sludge and identification of intermediate products by UPLC-QqTOF-MS', *Sci. Total Environ.*, 409(24), 5505–5512.
- Ge, Y., Yan, L. and Qinge, K. (2004) 'Effect of environment factors on dye decolorization by *P. sordida* ATCC90872 in a aerated reactor', *Process Biochem.*, 39(11), 1401–1405.
- Gòdia, F. and Solà, C. (1995) 'Fluidized-Bed Bioreactors', *Biotechnol. Prog.*, 11(5), 479–497.
- Goudar, C. T., Ganji, S. H., Pujar, B. G. and Strevett, K. A. (2000) 'Substrate inhibition kinetics of phenol biodegradation'. *Water Environ. Res.*, 72(1), 50–55.
- Gramss, G., Voigt, K. D. and Kirsche, B. (1999) 'Degradation of polycyclic aromatic hydrocarbons with three to seven aromatic rings by higher fungi in sterile and unsterile soils', *Biodegradation*, 10(1), 51–62.
- Guan, C., Wang, G., Ji, J., Wang, J., Wang, H. and Tan, M. (2008) 'Bioencapsulation of living yeast (*Pichia pastoris*) with silica after transformation with lysozyme gene', *J. Sol-Gel Sci. Technol.*, 48, 369–377.
- Gunasekar, V., Gowdhaman, D. and Ponnusami, V. (2013) Biodegradation of reactive red M5B dye using *Bacillus subtilis*', *Int. J. ChemTech Res.*, 5(1), 131–135.
- Gupta, V. K. and Suhas (2009) 'Application of low-cost adsorbents for dye removal - A review', *J. Environ. Manage.*, 90(8), 2313–2342.
- Haldane, J. B. S. (1965) *Enzymes*. Cambridge, MA: MIT Press.
- Hameed, B. H. and El-Khaiary, M. I. (2008) 'Malachite green adsorption by rattan sawdust: Isotherm, kinetic and mechanism modeling', *J. Hazard. Mater.*, 159(2–3), 574–579.
- Han, K. and Levenspiel, O. (1988) 'Extended monod kinetics for substrate, product, and cell inhibition', *Biotechnol. Bioeng.*, 32(4), 430–447.
- Han, M.-J., Choi, H.-T. and Song, H.-G. (2004) 'Degradation of phenanthrene by *Trametes versicolor* and its laccase', *J. Microbiol.*, 42(2), 94–98.
- Harazono, K., Watanabe, Y. and Nakamura, K. (2003) 'Decolorization of azo dye by the white-rot basidiomycete *Phanerochaete sordida* and by its manganese peroxidase', *J. Biosci. Bioeng.*, 95, 455–459.

- He, M., Ichinose, T., Liu, B., Song, Y., Yoshida, Y., Kobayashi, F., Maki, T., Yoshida, S., Nishikawa, M., Takano, H. and Sun, G. (2016) 'Silica-Carrying Particulate Matter Enhances *Bjerkandera adusta*-Induced Murine Lung Eosinophilia', *Environ. Toxicol.*, 31(1), 93–105.
- Heinfling, A., Ruiz-Dueñas, F. J., Martínez, M. J., Bergbauer, M., Szewzyk, U. and Martínez, A. T. (1998) 'A study on reducing substrates of manganese-oxidizing peroxidases from *Pleurotus eryngii* and *Bjerkandera adusta*', *FEBS Lett.*, 428(3), 141–146.
- Hestbjerg, H., Willumsen, P. A., Christensen, M., Andersen, O. and Jacobsen, C. S. (2003) 'Bioaugmentation of tar-contaminated soils under field conditions using *Pleurotus ostreatus* refuse from commercial mushroom production', *Environ. Toxicol. Chem.*, 22(4), 692–698.
- Hites, R. a. (2006) 'Persistent Organic Pollutants in the Great Lakes: An Overview', *Handb. Environ. Chem.*, 5, 1–12.
- Ho, C. W., Tan, W. S., Yap, W. B., Ling, T. C. and Tey, B. T. (2008) 'Comparative evaluation of different cell disruption methods for the release of recombinant hepatitis B core antigen from *Escherichia coli*', *Biotechnol. Bioprocess Eng.*, 13(5), 577–583.
- Hu, X., Wang, P. and Hwang, H. (2009) 'Oxidation of anthracene by immobilized laccase from *Trametes versicolor*', *Bioresour. Technol.*, 100(21), 4963–4968.
- Huang, J., Fu, Y. and Liu, Y. (2014) 'Comparison of Alkali-Tolerant Fungus *Myrothecium* Sp. IMER1 and White-Rot Fungi for Decolorization of Textile Dyes and Dye Effluents', *J. Bioremediation Biodegrad.*, 5(3), 221-226.
- Isroi, Millati, R., Syamsiah, S., Niklasson, C., Cahyanto, M. N., Lundquist, K. and Taherzadeh, M. J. (2011) 'Biological pretreatment of lignocelluloses with white-rot fungi and its applications: A review', *BioResources*, 6(4), 5224–5259.
- Jamil, N., Ahsan, N., Munwar, M. A., Anwar, J. and Shafiq, U. (2011) 'Removal of Toxic Dichlorophenol from Water by Sorption with Chemically Activated Carbon of Almond Shells - A Green Approach', *J. Chem. Soc. Pakistan*, 33(5), 640–645.
- Jaroch, D., Mclamore, E., Zhang, W., Shi, J., Garland, J., Banks, M. K., Porterfield, D. M. and Rickus, J. L. (2011) 'Cell-mediated deposition of porous silica on bacterial biofilms', *Biotechnol. Bioeng.*, 108(10), 2249–2260.

- Jiang, G. X., Niu, J. F., Zhang, S. P., Zhang, Z. Y. and Xie, B. (2008) 'Prediction of biodegradation rate constants of hydroxylated polychlorinated biphenyls by fungal laccases from *Trametes versicolor* and *Pleurotus ostreatus*', *Bull. Environ. Contam. Toxicol.*, 81(1), 1–6.
- Jin, W. and Brennan, J. D. (2002) 'Properties and applications of proteins encapsulated within sol–gel derived materials', *Anal. Chim. Acta.*, 461(1), 1–36.
- Jo, W.-S., Kang, M.-J., Choi, S.-Y., Yoo, Y.-B., Seok, S.-J. and Jung, H.-Y. (2010) 'Culture Conditions for Mycelial Growth of *Coriolus versicolor*', *Mycobiology*, 38(3), 195–202.
- Jolivalt, C., Brenon, S., Caminade, E., Mougin, C. and Pontié, M. (2000) 'Immobilization of laccase from *Trametes versicolor* on a modified PVDF microfiltration membrane: Characterization of the grafted support and application in removing a phenylurea pesticide in wastewater', *J. Memb. Sci.*, 180(1), 103–113.
- Joshi, M., Bansal, R. and Purwar, R. (2004) 'Colour removal from textile effluents', *Indian J. Fibre Text. Res.*, 29(2), 239–259.
- Kalme, S. D., Parshetti, G. K., Jadhav, S. U. and Govindwar, S. P. (2007) 'Biodegradation of benzidine based dye Direct Blue-6 by *Pseudomonas desmolyticum* NCIM 2112', *Bioresour. Technol.*, 98(7), 1405–1410.
- Kalpana, D., Velmurugan, N., Shim, J. H., Oh, B. T., Senthil, K. and Lee, Y. S. (2012) 'Biodecolorization and biodegradation of reactive Levafix Blue E-RA granulate dye by the white rot fungus *Irpex lacteus*', *J. Environ. Manage.*, 111, 142–149.
- Kanagaraj, J., Senthilvelan, T. and Panda, R. C. (2015) 'Biodegradation of azo dyes in industrial effluent: An eco-friendly way toward green technology', *Clean Technol. Environ. Policy.*, 17(2), 331–341.
- Kapdan, I. K., Kargia, F., McMullan, G. and Marchant, R. (2000) 'Effect of environmental conditions on biological decolorization of textile dyestuff by *C. versicolor*', *Enzyme Microb. Technol.*, 26(5), 381–387.
- Kaushik, P. and Malik, A. (2009) 'Fungal dye decolourization: Recent advances and future potential', *Environ. Int.*, 35(1), 127–141.
- Keeling-Tucker, T., Rakic, M., Spong, C. and Brennan, J. D. (2000) 'Controlling the material properties and biological activity of lipase within sol-gel derived bioglasses via organosilane and polymer doping', *Chem. Mater.*, 12(12), 3695–

3704.

- Khayat-zadeh, J. and Abbasi, E. (2010) 'The effects of heavy metals on aquatic animals', *1st Int. Appl. Geol. Congr. Dep. Geol. Islam. Azad Univ. Branch, Iran.* 1(April), 26–28.
- Khongkhaem, P., Intasiri, A. and Luepromchai, E. (2011) 'Silica-immobilized *Methylobacterium* sp. NP3 and *Acinetobacter* sp. PK1 degrade high concentrations of phenol', *Lett. Appl. Microbiol.*, 52(5), 448–455.
- Kim, T. H., Lee, Y., Yang, J., Lee, B., Park, C. and Kim, S. (2004) 'Decolorization of dye solutions by a membrane bioreactor (MBR) using white-rot fungi', *Desalination*, 168(1–3), 287–293.
- Kitching, M., Ramani, M. and Marsili, E. (2015) 'Fungal biosynthesis of gold nanoparticles: Mechanism and scale up. *Microb. Biotechnol.*, 8(6), 904–917.
- Knapp, J. S., Newby, P. S. and Reece, L. P. (1995) 'Decolorization of dyes by wood-rotting basidiomycete fungi', *Enzyme Microb. Technol.*, 17(7), 664–668.
- de Kreijl, A., van den Burg, B., Venema, G., Vriend, G., Eijsink, V. G. H. and Nielsen, J. E. (2002) 'The Effects of Modifying the Surface Charge on the Catalytic Activity of a Thermolysin-like Protease', *J. Biol. Chem.*, . 277(18), 15432–15438.
- Kryst, K. and Karamanev, D. G. (2001) 'Aerobic Phenol Biodegradation in an Inverse Fluidized-Bed Biofilm Reactor', *Ind. Eng. Chem. Res.*, 40, 5436–5439.
- Kurniawati, S. and Nicell, J. A. (2008) 'Characterization of *Trametes versicolor* laccase for the transformation of aqueous phenol', *Bioresour. Technol.*, 99(16), 7825–7834.
- Kuśmierk, K. and Wiatkowski, A. (2015) 'The influence of different agitation techniques on the adsorption kinetics of 4-chlorophenol on granular activated carbon', *React. Kinet. Mech. Catal.*, 116(1), 261–271.
- Lakshmanaperumalsamy, P., Karthikeyan, K. and Nanthakumar, K. (2009) 'Kinetic and Equilibrium Studies on In-Situ Biosorption of Reactive Blue 140 Dye by Live Biomass Preparation of *Aspergillus niger* HM11', *Glob. J. Environ. Res.*, 3(3), 264–273.
- Lara, M. A., Rodríguez-Malaver, A. J., Rojas, O. J., Holmquist, O., González, A. M., Bullón, J., Peñaloza, N. and Araujo, E. (2003) 'Black liquor lignin biodegradation by *Trametes elegans*', *Int. Biodeterior. Biodegrad.*, 52(3), 167–173.

- Larsson, S., Cassland, P. and Jönsson, L. J. (2001) 'Development of a *Saccharomyces cerevisiae* strain with enhanced resistance to phenolic fermentation inhibitors in lignocellulose hydrolysates by heterologous expression of laccase', *Appl. Environ. Microbiol.*, 67(3), 1163–1170.
- Lavanya, C. (2014) 'Review Article Degradation of Toxic Dyes : A Review', *Int. J. Curr. Microbiol. Applied Sci.*, 3(6), 189–199.
- Levin, L., Carabajal, M., Hofrichter, M. and Ullrich, R. (2016) 'Degradation of 4-nitrophenol by the white-rot polypore *Trametes versicolor*', *Int. Biodeterior. Biodegrad.*, 107, 174–179.
- Leyva-Ramos, R., Ocampo-Perez, R. and Mendoza-Barron, J. (2012) 'External mass transfer and hindered diffusion of organic compounds in the adsorption on activated carbon cloth', *Chem. Eng. J.*, 183, 141–151.
- Liu, J., Li, E., You, X., Hu, C. and Huang, Q. (2016) 'Adsorption of methylene blue on an agro-waste oiltea shell with and without fungal treatment', *Sci. Rep.*, 6(1), 1-10.
- Livage, J. and Coradin, T. (2017). *Encapsulation of Enzymes, Antibodies, and Bacteria BT - Handbook of Sol-Gel Science and Technology*. Cham: Springer International Publishing.
- Livernoche, D., Jurasek, L., Desrochers, M., Dorica, J. and Veliky, I. A. (1983) 'Removal of color from kraft mill wastewaters with cultures of white-rot fungi and with immobilized mycelium of *Coriolus versicolor*', *Biotechnol. Bioeng.*, 25(8), 2055–2065.
- Loomis, A. K., Childress, A. M., Daigle, D. and Bennett, J. W. (1997) 'Alginate encapsulation of the white rot fungus *Phanerochaete chrysosporium*', *Curr. Microbiol.*, 34(2), 127–130.
- Da Luz, J. M. R., Paes, S. A., Ribeiro, K. V. G., Mendes, I. R. and Kasuya, M. C. M. (2015) 'Degradation of green polyethylene by *Pleurotus ostreatus*', *PLoS One*, 10(6), 1-12.
- Shin, M., Nguyen, T., and Ramsay, J. (2002) 'Evaluation of support materials for the surface immobilization and decoloration of amaranth by *Trametes versicolor*', *Appl. Microbiol. Biotechnol.*, 60(1–2), 218–223.
- Malik, A. (2004) 'Metal bioremediation through growing cells', *Environ. Int.*, 30(2), 261–278.

- Manna, A. and Amutha, C. (2017) 'Laccase–silica nanoparticle conjugates can efficiently reduce the early maturation risk due to BPA in female *Oreochromis mossambicus* and its toxic load from the contaminated effluent', *Environ. Sci. Nano.*, 4(7), 1553–1568.
- Mansoori, G. A., Data, P. P. and Cited, R. (2013) (12) *United States Patent*. 2(12).
- Mansor, A. F., Mohidem, N. A., Wan Mohd Zawawi, W. N. I., Othman, N. S., Endud, S. and Mat, H. (2015) 'Preparation and characterization of in situ entrapment of laccase in silica microparticles via an ambient drying procedure', *J. Sol-Gel Sci. Technol.*, 75(2), 323–335.
- Mansur, H. S., Sadahira, C. M., Souza, A. N. and Mansur, A. A. P. (2008) 'FTIR spectroscopy characterization of poly (vinyl alcohol) hydrogel with different hydrolysis degree and chemically crosslinked with glutaraldehyde', *Mater. Sci. Eng. C.*, 28(4), 539–548.
- Manzanares, P., Fajardo, S. and Martin, C. (1995) 'Production of ligninolytic activities when treating paper pulp effluents by *Trametes versicolor*', *J. Biotechnol.*, 43(2), 125–132.
- Merchuk, J. C. and Asenjo, J. A. (1995) 'The Monod equation and mass transfer. *Biotechnol. Bioeng.*, 45(1), 91–94.
- Meunier, C. F., Dandoy, P. and Su, B.-L. (2010) 'Encapsulation of cells within silica matrixes: Towards a new advance in the conception of living hybrid materials', *J. Colloid Interface Sci.*, 342(2), 211–224.
- Michaels, J. D. and Papoutsakis, E. T. (1991) 'Polyvinyl alcohol and polyethylene glycol as protectants against fluid-mechanical injury of freely-suspended animal cells (CRL 8018)', *J. Biotechnol.*, 19(2–3), 241–257.
- Minussi, R. C., Miranda, M. A., Silva, J. A., Ferreira, C. V, Aoyama, H., Rotilio, D., Pastore, G. M. and Durán, N. (2007) 'Purification , characterization and application of laccase from *Trametes versicolor* for colour and phenolic removal of olive mill wastewater in the presence of 1- hydroxybenzotriazole', *African J. Biotechnol.*, 6(10), 1248–1254.
- Mishra, A. and Malik, A. (2013) 'Recent Advances in Microbial Metal Bioaccumulation', *Crit. Rev. Environ. Sci. Technol.*, 43(11), 1162–1222.
- Mohidem, N. A. and Mat, H. (2009) 'The catalytic activity of laccase immobilized in sol-gel silica', *J. App. Sci.*, 9(17), 3141–3145.

- Mohidem, N. A. and Mat, H. (2012a) 'Catalytic activity and stability of laccase entrapped in sol-gel silica with additives', *J. Sol-Gel Sci. Technol.* 61(1), 96–103.
- Mohidem, N. A. and Mat, H. Bin (2012b). The catalytic activity enhancement and biodegradation potential of free laccase and novel sol-gel laccase in non-conventional solvents. *Bioresour. Technol.* 114, 472–477.
- Monod, J. (1949) 'The Growth of Bacterial Cultures', *Annu. Rev. Microbiol.*, 3(1), 371–394.
- Muñoz, C., Guillén, F., Martínez, A. T. and Martínez, M. J. (1997) 'Induction and characterization of laccase in the ligninolytic fungus *Pleurotus eryngii*', *Curr. Microbiol.*, 34(1), 1–5.
- Murugesan, K. and Kalaichelvan, P. T. (2003) 'Synthetic dye decolourization by white rot fungi', *Indian J. Exp. Biol.*, 41(9), 1076–1087.
- Nadeem, M. A., Ajmal, A., Malik, R. N., Majeed, I. and Idriss, H. (2014) 'Principles and mechanisms of photocatalytic dye degradation on TiO₂ based photocatalysts: A comparative overview', *RSC Adv.* (2005), 37003–37026.
- Necochea, R., Valderrama, B., Díaz-Sandoval, S., Folch-Mallol, J. L., Vázquez-Duhalt, R. and Iturriaga, G. (2005) 'Phylogenetic and biochemical characterisation of a recombinant laccase from *Trametes versicolor*', *FEMS Microbiol. Lett.*, 244(2), 235–241.
- Novotný, Č., Erbanová, P., Šašek, V., Kubátová, A., Cajthaml, T., Lang, E., Krahel, J. and Zadražil, F. (1999) 'Extracellular oxidative enzyme production and PAH removal in soil by exploratory mycelium of white rot fungi', *Biodegradation*, 10(3), 159–168.
- Ocampo-Pérez, R., Rivera-Utrilla, J., Gómez-Pacheco, C., Sánchez-Polo, M. and López-Peñalver, J. J. (2012) 'Kinetic study of tetracycline adsorption on sludge-derived adsorbents in aqueous phase', *Chem. Eng. J.*, 213, 88–96.
- Ogugbue, C. J. and Sawidis, T. (2011) 'Bioremediation and detoxification of synthetic wastewater containing triarylmethane dyes by *Aeromonas hydrophila* isolated from industrial effluent', *Biotechnol. Res. Int.*, 2011, 1-11.
- Ogura, M. (2008) 'Towards realization of a micro- and mesoporous composite silicate catalyst', *Catal. Surv. from Asia*, 12(1), 16–27.

- Ohkuma, M., Maeda, Y., Johjima, T. and Kudo, T. (2001) 'Lignin degradation and roles of white rot fungi: Study on an efficient symbiotic system in fungus-growing termites and its application to bioremediation', *Microbiology*, 42(42), 39–42.
- Okamoto, K., Uchii, A., Kanawaku, R. and Yanase, H. (2014) 'Bioconversion of xylose, hexoses and biomass to ethanol by a new isolate of the white rot basidiomycete *Trametes versicolor*', *Springerplus*, 3, (121), 1-9.
- Okpokwasili, G. C. and Nweke, C. O. (2005) 'Microbial growth and substrate utilization kinetics', *African J. Biotechnol.*, 5(4), 305–317.
- Osma, J. F., Saravia, V., Herrera, J. L. T. and Couto, S. R. (2007) 'Mandarin peelings: The best carbon source to produce laccase by static cultures of *Trametes pubescens*', *Chemosphere*, 67(8), 1677–1680.
- Pallerla, S. and Chambers, R. P. (1997) 'Characterization of a Ca-alginate-immobilized *Trametes versicolor* bioreactor for decolorization and AOX reduction of paper mill effluents', *Bioresour. Technol.*, 60(1), 1–8.
- Pallerla, S. and Chambers, R. P. (1998) 'Reactor development for biodegradation of pentachlorophenol', *Catal. Today*, 40(1), 103–111.
- Pannier, A., Lehrer, T., Vogel, M., Soltmann, U., Böttcher, H., Tarre, S., Green, M., Raff, J. and Pollmann, K. (2014) 'Long-term activity of biohybrid coatings of atrazine-degrading bacteria *Pseudomonas* sp. ADP', *RSC Adv.*, 4(38), 19970–19979.
- Papinutti, L. and Forchiassin, F. (2010) 'Adsorption and decolorization of dyes using solid residues from *Pleurotus ostreatus* mushroom production', *Biotechnol. Bioprocess Eng.*, 15(6), 1102–1109.
- Park, = Donghee, Yun, Y.-S. and Park, J. M. (2010) 'The past, present, and future trends of biosorption', *Biotechnol. Bioprocess Eng.*, 15, 86–102.
- Pavko, A. (2011) 'Fungal decolourization and degradation of synthetic dyes some chemical engineering aspects', *Waste Water- Treat. Reutil.*, 65–88.
- Pazarlioğlu, N. K., Sarişik, M. and Telefoncu, A. (2005) 'Laccase: Production by *Trametes versicolor* and application to denim washing', *Process Biochem.*, 40(5), 1673–1678.
- Pearce, C. I., Lloyd, J. R. and Guthrie, J. T. (2003) 'The removal of colour from textile wastewater using whole bacterial cells: A review', *Dye Pigment.*, 58(3), 179–196.

- Peralta-Pérez, M. R., Martínez-Trujillo, M. a., Nevárez-Moorillón, G. V., Pérez-Bedolla, R. and García-Rivero, M. (2010) 'Immobilization of *Aspergillus niger* sp. in sol gel and its potential for production of xylanases', *J. Sol-Gel Sci. Technol.*, 57(1), 6–11.
- Pérez-Quintanilla, D., Hierro, I. del, Fajardo, M. and Sierra, I. (2006) '2-Mercaptothiazoline modified mesoporous silica for mercury removal from aqueous media', *J. Hazard. Mater.*, 134(1–3), 245–256.
- Peterson, K. P., Peterson, C. M. and Pope, E. J. (1998) 'Silica sol-gel encapsulation of pancreatic islets', *Proc. Soc. Exp. Biol. Med.*, 218(4), 365–9.
- Pezzella, C. Russo, M. E., Marzocchella, A., Salatino, P. and Sannia, G. (2014) 'Immobilization of a *Pleurotus ostreatus* laccase mixture on perlite and its application to dye decolourisation', *BioMed Research Int.*, 1-11
- Pierre, A. C. and Rigacci, A. (2011) *Aerogels Handbook*. 1. London: Springer.
- Plagemann, R., Jonas, L. and Kragl, U. (2011) 'Ceramic honeycomb as support for covalent immobilization of laccase from *Trametes versicolor* and transformation of nuclear fast red', *Appl. Microbiol. Biotechnol.*, 90(1), 313–320.
- Pramanik, S. and Chaudhuri, S. (2018) 'Laccase activity and azo dye decolorization potential of *Podoscypha elegans*', *Mycobiology*, 46(1),79–83.
- Priddy, S. A. and Hanley, T. R. (2003) 'Effect of agitation on removal of acetic acid from pretreated hydrolysate by activated carbon', *Applied Biochem. Biotechnol.*, 2003, 353–364.
- Punt, P. J., Van Biezen, N., Conesa, A., Albers, A., Mangnus, J. and Van Den Hondel, C. (2002) 'Filamentous fungi as cell factories for heterologous protein production', *Trends in Biotechnol.*, 20(5), 200–206.
- Puvaneswari, N., Muthukrishnan, J. and Gunasekaran, P. (2006) 'Toxicity assessment and microbial degradation of azo dyes', *Indian J. Exp. Biol.* 44(8), 618–626.
- Raff, J., Soltmann, U., Matys, S. and Bo, H. (2003) 'Biosorption of uranium and copper by Biocers', *Chem. Mater.*, 15(1), 240–244.
- Rajaguru, P., Kalaiselvi, K., Palanivel, M. and Subburam, V. (2000) 'Biodegradation of azo dyes in a sequential anaerobic-aerobic system', *Appl. Microbiol. Biotechnol.*, 54(2), 268–273.
- Rajasulochana, P. and Preethy, V. (2016) 'Comparison on efficiency of various techniques in treatment of waste and sewage water – A comprehensive review',

- Resour. Technol.*, 2(4), 175–184.
- Rajeshkannan, R., Rajasimman, M. and Rajamohan, N. (2012) 'Packed bed column studies for the removal of dyes using novel sorbent', *Chem. Ind. Chem. Eng. Q.*, 19(4), 461–470.
- Rajeshkannan, R., Rajasimman, M. and Rajamohan, N. (2013) 'Packed bed column studies for the removal of dyes using novel sorbent', *Ind. Chem. Eng.*, 19(4), 461-470.
- Rajeswari, K., Subashkumar, R. and Vijayaraman, K. (2013) 'Decolorization and degradation of textile dyes by *Stenotrophomonas maltophilia* RSV-2', *Int. J. Environ. Bioremediation Biodegrad.*, 1(2), 60–65.
- Ramakrishna, K. and Viraraghavan, T. (1996) 'Dye removal using peat', *Am. Dyest. Report.*, 85(10), 28–34.
- Ramsay, J. A., Mok, W. H. W., Luu, Y. S. and Savage, M. (2005) 'Decoloration of textile dyes by alginate-immobilized *Trametes versicolor*', *Chemosphere*, 61(7), 956–964.
- Rao, K. S., Anand, S. and Venkateswarlu, P. (2011) 'Modeling the kinetics of Cd(II) adsorption on *Syzygium cumini* L leaf powder in a fixed bed mini column', *J. Ind. Eng. Chem.*, 17(2), 174–181.
- Rawat, V., Rai, P., Gautam, R. K. and Chattopadhyaya, M. C. (2013). 'Kinetic and equilibrium isotherm studies for the adsorptive removal of Brilliant Green Dye from aqueous solution by *Oplismenus frumentaceus* husk', *J. Indian Chem. Soc.*, 90(5), 577–583.
- Reichenauer, G. (2004) 'Thermal aging of silica gels in water', *J. Non-Crystalline Solids*, 2004, 189–195.
- Rica, C., Girona, J. and Park, T. (2015) 'Biodegradation of Polybrominated Diphenyl Ethers in Liquid Media and Sewage Sludge by *Trametes versicolor*', *Int. J. Environ. Res.*, 9(1), 273–280.
- Rocha, J. M. S., Gil, M. H. and Garcia, F. A. P. (1998) 'Effects of additives on the activity of a covalently immobilised lipase in organic media', *J. Biotechnol.*, 66(1), 61–67.
- Rodríguez-Rodríguez, C. E., García-Galán, M. a J., Blánquez, P., Díaz-Cruz, M. S., Barceló, D., Caminal, G. and Vicent, T. (2012) 'Continuous degradation of a mixture of sulfonamides by *Trametes versicolor* and identification of metabolites from sulfapyridine and sulfathiazole', *J. Hazard. Mater.*, 213–214,

- Rodríguez Couto, S. (2009) 'Dye removal by immobilised fungi', *Biotechnol. Adv.*, 27(3), 227–235.
- Roig, M. G., Bello, J. F., Fernando, G., Celis, C. D. D. E. and Juan, M. (1987) 'Applications of immobilized enzymes', *Biochem. Educ.*, 15(4), 198–208.
- Rozada, F., Otero, M., García, A. I. and Morán, A. (2007) 'Application in fixed-bed systems of adsorbents obtained from sewage sludge and discarded tyres', *Dye. Pigment.*, 72(1), 47–56.
- Russo, M. E., Di Natale, F., Prigione, V., Tigini, V., Marzocchella, A. and Varese, G. C. (2010) 'Adsorption of acid dyes on fungal biomass: Equilibrium and kinetics characterization', *Chem. Eng. J.*, 162(2), 537–545.
- Rybczyńska-Tkaczyk, K. and Korniewicz-Kowalska, T. (2016) 'Biosorption optimization and equilibrium isotherm of industrial dye compounds in novel strains of microscopic fungi', *Int. J. Environ. Sci. Technol.*, 13(12), 2837–2846.
- Saman, N., Johari, K., Song, S. T., Kong, H., Cheu, S. C. and Mat, H. (2016) 'High removal efficiency of Hg(II) and MeHg(II) from aqueous solution by coconut pith - Equilibrium, kinetic and mechanism analyses', *J. Environ. Chem. Eng.*, 4(2), 2487–2499.
- Sampedro, I., Cajthaml, T., Marinari, S., Stazi, S. R., Grego, S., Petruccioli, M., Federici, F. and D'Annibale, A. (2009) 'Immobilized inocula of white-rot fungi accelerate both detoxification and organic matter transformation in two-phase dry olive-mill residue', *J. Agric. Food Chem.*, 57(12), 5452–5460.
- Santhi, T., Manonmani, S., Vasantha, V. S. and Chang, Y. T. (2011) 'A new alternative adsorbent for the removal of cationic dyes from aqueous solution', *Arab. J. Chem.*, 9(1), 466-474.
- Saravanan, N., Kannadasan, T., Basha, C. A. and Manivasagan, V. (2013) 'Biosorption of textile dye using immobilized bacterial (*Pseudomonas aeruginosa*) and fungal (*Phanerochate chrysosporium*) cells', *Am. J. Environ. Sci. Publ. Online*, 9(94), 377–387.
- Sasaki, T., Kajino, T., Li, B., Sugiyama, H. and Takahashi, H. (2001) 'New Pulp Biobleaching System Involving Manganese Peroxidase Immobilized in a Silica Support with Controlled Pore Sizes', *Appl. Environ. Microbiol.*, 67(5), 2208–2212.

- Sato, S. H. I. M. I. O. and Suzuki, T. (1990) 'Control of pore size distribution of silica gel through sol-gel process using water soluble polymers as additives', *J. Mater. Sci.*, 25, 4880–4885.
- Schwarze F. , Engels J., and C. M. (2000) '*Fungal Strategies of Wood Decay in Trees*. Berlin, Heidelberg: Springer
- Sedarati, M. R., Keshavarz, T., Leontievsky, A. A. and Evans, C. S. (2003) 'Transformation of high concentrations of chlorophenols by the white-rot basidiomycete *Trametes versicolor* immobilized on nylon mesh', *Electron. J. Biotechnol.*, 6(2), 104–114.
- Shah, M. and Patel, K. (2014) 'Decolorization of Remazol Black-B by Three Bacterial Isolates', *Int. J. Environ. Bioremediation Biodegrad.*, 2(1), 44–49.
- Shah, V. and Nerud, F. (2002) 'Lignin degrading system of white-rot fungi and its exploitation for dye decolorization', *Can. J. Microbiol.*, 48(10), 857–870.
- Sharma, P., Singh, L. and Dilbaghi, N. (2009) 'Biodegradation of Orange II dye by *Phanerochaete chrysosporium* in simulated wastewater', *J. Sci. Ind. Res.*, 68(2), 157–161.
- Shraddha, Shekher, R., Sehgal, S., Kamthania, M. and Kumar, A. (2011) 'Laccase: microbial sources, production, purification, and potential biotechnological applications', *Enzyme Res.*, 2011, 217861-217872.
- Shuler, M. L. and Kargi, F. (2002) *Bioprocess engineering: Basic concepts*. 3rd Edition. America: Prentice Hall International.
- Sing, K. S. W. and Williams, R. T. (2005) 'Physisorption Hysteresis Loops and the Characterization of Nanoporous Materials', *Adsorpt. Sci. Technol.*, 22(10), 773–782.
- Singh, N. and Balomajumder, C. (2016) 'Simultaneous biosorption and bioaccumulation of phenol and cyanide using coconut shell activated carbon immobilized *Pseudomonas putida* (MTCC 1194)', *J. Environ. Chem. Eng.*, 4(2), 1604–1614.
- Singh, R. S. and Sooch, B. S. (2009). 'Review Paper High cell density reactors in production of fruits wine with special reference to cider – An overview', *Nat. Prod. Rad.*, 8(4), 323–333.
- Singh, S. K., Katoria, D., Mehta, D. and Sehgal, D. (2015) 'Fixed Bed Column Study and adsorption modelling on the adsorption of malachite green dye from wastewater using acid activated sawdust', *Int. J. Adv. Res.*, 3(7), 521–529.

- Singha, B. and Das, S. K. (2011) 'Biosorption of Cr(VI) ions from aqueous solutions: Kinetics, equilibrium, thermodynamics and desorption studies', *Colloids Surfaces B Biointerfaces*, 84(1), 221–232.
- Siripong, P., Oraphin, B., Sanro, T. and Duanporn, P. (2009) 'Screening of Fungi from natural sources in Thailand for degradation of polychlorinated hydrocarbons', *Am. J. Agric Environ. Sci.*, 5(4), 466–472.
- Smith, F. W. (1970) 'The behavior of partially hydrolyzed polyacrylamide solutions in porous media', *J. Pet. Technol.*, 22(2), 148–156.
- Soares, C. M. F., De Castro, H. F., Santana, M. H. a and Zanin, G. M. (2002) 'Intensification of lipase performance for long-term operation by immobilization on controlled pore silica in presence of polyethylene glycol', *Appl. Biochem. Biotechnol.*, 98(100), 863–874.
- Solids, F. (1997) 'Chemical Modification of Silica Gels', *J. Sol-Gel Sci Technol.*, 8(1-3), 499–505.
- Soltmann, U. (2010) 'Algae-Silica Hybrid Materials for Biosorption of Heavy Metals', *J. Water Resour. Prot.*, 2(2), 115–122.
- Soltmann, U. and Böttcher, H. (2008) 'Utilization of sol-gel ceramics for the immobilization of living microorganisms', *J. Sol-Gel Sci. Technol.*, 48, 66–72.
- Soltmann, U., Böttcher, H., Koch, D. and Grathwohl, G. (2003) 'Freeze gelation: a new option for the production of biological ceramic composites (biocers)', *Mater. Lett.*, 57(19), 2861–2865.
- Song, J., Zou, W., Bian, Y., Su, F. and Han, R. (2011) 'Adsorption characteristics of methylene blue by peanut husk in batch and column modes', *Desalination*, 265(1–3), 119–125.
- Soni, M., K.Sharma, A., K.Srivastava, J. and Yadav, J. S. (2012) 'Adsorptive removal of methylene blue dye from an aqueous solution using water hyacinth root powder as a low cost adsorbent', *Int. J. Chem. Sci. Appl.*, 3(3), 338–345.
- Souza, R. L., Faria, E. L. P., Figueiredo, R. T., Fricks, A. T., Zanin, G. M., Santos, O. a. a., Lima, Á. S. and Soares, C. M. F. (2014) 'Use of polyethylene glycol in the process of sol–gel encapsulation of *Burkholderia cepacia* lipase', *J. Therm. Anal. Calorim.*, 117(1), 301–306.
- Spahn, C. and Minter, S. D. (2008) 'Enzyme Immobilization in Biotechnology', *Recent Patents Eng.*, 2(3), 195–200.

- Spinelli, D., Fatarella, E., Di Michele, A. and Pogni, R. (2013) 'Immobilization of fungal (*Trametes versicolor*) laccase onto Amberlite IR-120 H beads: Optimization and characterization', *Process Biochem.*, 48(2), 218–223.
- Srivastava, S., Sinha, R. and Roy, D. (2004) 'Toxicological effects of malachite green', *Aquatic Toxicology*, 66(3),319–329.
- Stoilova, I., Krastanov, A. and Stanchev, V. (2010) 'Properties of crude laccase from *Trametes versicolor* produced by solid-substrate fermentation', *Adv. Biosci. Biotechnol.*, 1(3), 208–215.
- Strøm, R. A., Masmoudi, Y., Rigacci, A., Petermann, G., Gullberg, L., Chevalier, B. and Einarsrud, M. A. (2007) 'Strengthening and aging of wet silica gels for up-scaling of aerogel preparation', *J. Sol-Gel Sci. Technol.*, 41(3), 291–298.
- Suk Choi, H., Sup Kim, D., Thapa, L., Lee, S.-J., Kim, S. B., Cho, J., Park, C. and Wook Kim, S. (2016) 'Production and characterization of cellobiose dehydrogenase from *Phanerochaete chrysosporium* KCCM 60256 and its application for an enzymatic fuel cell', *Korean J. Chem. Eng.*,33(12), 3434-3441.
- Taylor, A. P., Finnie, K. S., Bartlett, J. R. and Holden, P. J. (2004) 'Encapsulation of viable aerobic microorganisms in silica gels', *J. Sol-Gel Sci. and Technol.*, 2004, 223–228.
- Theodore, L. (2012) 'Continuous Stirred Tank Reactors', *Chem. React. Anal. Appl. Pract. Eng.*, 181–207.
- Tortolini, C., Rea, S., Carota, E., Cannistraro, S. and Mazzei, F. (2012). Influence of the immobilization procedures on the electroanalytical performances of *Trametes versicolor* laccase based bioelectrode. *Microchem. J.* 100(1), 8–13.
- Trgo, M., Medvidović, N. V. and Perić, J. (2011) 'Application of mathematical empirical models to dynamic removal of lead on natural zeolite clinoptilolite in a fixed bed column', *Indian J. Chem. Technol.*, 18(2), 123–131.
- Triantafyllou, A Ö., Wehtje, E., Adlercreutz, P. and Mattiasson, B. (1997) 'How do additives affect enzyme activity and stability in nonaqueous media?', *Biotechnol. Bioeng.*, 54(1), 67–76.
- Tsekova, K. V, Chernev, G. E. and Hristov, A. E. (2013) 'Phenol Biodegradation by Fungal Cells Immobilized in Sol-Gel Hybrids', *J. Biosci.*, 68(1-2):53-59
- Tseng, M. M. and Wayman, M. (2010) 'Kinetics of yeast growth: inhibition-threshold substrate concentrations', *Can. J. Microbiol.*, 21(7), 994–1003.

- Tuomela, M., Lyytikäinen, M., Oivanen, P. and Hatakka, A. (1998). Mineralization and conversion of pentachlorophenol (PCP) in soil inoculated with the white-rot fungus *Trametes versicolor*. *Soil Biol. Biochem.* 31(1), 65–74.
- Uhnáková, B., Petricková, a, Biedermann, D., Homolka, L., Vejvoda, V., Bednár, P., Papoušková, B., Sulc, M. and Martínková, L. (2009) 'Biodegradation of brominated aromatics by cultures and laccase of *Trametes versicolor*', *Chemosphere*, 76(6), 826–832.
- Umoren, S. A., Etim, U. J. and Israel, A. U. (2013) 'Adsorption of methylene blue from industrial effluent using poly (vinyl alcohol)', *J. Mater. Environ. Sci.*, 4(1), 75–86.
- Ünyayar, A., Mazmanci, M. a., Ataçağ, H., Erkurt, E. a. and Coral, G. (2005) 'A Drimaren Blue X3LR dye decolorizing enzyme from *Funalia trogii*: One step isolation and identification', *Enzyme Microb. Technol.*, 36(1), 10–16.
- Vadivelan, V. and Vasanth Kumar, K. (2005) 'Equilibrium, kinetics, mechanism, and process design for the sorption of methylene blue onto rice husk', *J. Colloid Interface Sci.*, 286(1), 90–100.
- Vaithanomsat, P., Apiwatanapiwat, W., Petchoy, O. and Chedchant, J. (2010a) 'Decolorization of Reactive Dye by White-Rot Fungus *Datronia* sp', *Natural Resources*, 890, 879–890.
- Vaithanomsat, P., Apiwatanapiwat, W., Petchoy, O. and Chedchant, J. (2010b). Production of ligninolytic enzymes by white-rot fungus *Datronia* sp. KAPI0039 and their application for reactive dye removal. *Int. J. Chem. Eng.*, 2010, 1-6.
- Verbelen, P. J., De Schutter, D. P., Delvaux, F., Verstrepen, K. J. and Delvaux, F. R. (2006) 'Immobilized yeast cell systems for continuous fermentation applications', *Biotechnology Letters*. 28 (19) p.1515–1525.
- Vijayaraghavan, K. and Yun, Y. S. (2008) 'Bacterial biosorbents and biosorption', *Biotechnol. Adv.*, 26(3), 266–291.
- Villalonga, R., Villalonga, M. and Gómez, L. (2000) 'Preparation and functional properties of trypsin modified by carboxymethylcellulose', *J. Mol. Catal. B Enzym.*, 10, 483–490.
- Viswanath, B., Chandra, M. S., Pallavi, H. and Reddy, B. R. (2008) 'Screening and assessment of laccase producing fungi isolated from different environmental samples', *African J. Biotechnol.*, 7(April), 1129–1133.
- Volesky, B. (2007) 'Biosorption and me', *Water Research*, 41(18), 4017–4029.

- Walker, G. M. and Weatherley, L. R. (2000) 'Biodegradation and biosorption of acid anthraquinone dye', *Environ. Pollut.*, 108(2), 219–223.
- Walker, G. M. and Weatherley, L. R. (1998). Fixed bed adsorption of acid dyes onto activated carbon. *Environ. Pollut.* 99(1), 133–136.
- Wan Mohd Zawawi, W. N. I., Mansor, A. F., Othman, N. S., Mohidem, N. A., Nik Malek, N. A. N. and Mat, H. (2015) 'Synthesis and characterization of immobilized white-rot fungus *Trametes versicolor* in sol–gel ceramics', *J. Sol-Gel Sci. Technol.*, 77(1), 28-38.
- Wang, B. e. and Hu, Y. you (2007) 'Comparison of four supports for adsorption of reactive dyes by immobilized *Aspergillus fumigatus* beads', *J. Environ. Sci.* 19(4), 451–457.
- Wang, F., Guo, C. and Liu, C. Z. (2013) 'Immobilization of *Trametes versicolor* cultures for improving laccase production in bubble column reactor intensified by sonication', *J. Ind. Microbiol. Biotechnol.*, 40(1), 141–150.
- Wang, J., Gao, F., Liu, Z., Qiao, M., Niu, X., Zhang, K.-Q. and Huang, X. (2012) 'Pathway and molecular mechanisms for malachite green biodegradation in *Exiguobacterium* sp. MG2', *PLoS One*, 7(12), 1-10.
- Ward, K. R., Adams, G. D., Alpar, H. O. and Irwin, W. J. (1999) 'Protection of the enzyme L-asparaginase during lyophilization-a molecular modeling approach to predict required level of lyoprotectant', *Int. J. Pharm.*, 187(2), 153-162.
- Warren, J. C. and Cheatum, S. G. (1966) 'Effect of Neutral Salts on Enzyme Activity and Structure', *Biochemistry*, 5(5), 1702–1707.
- Webb, J. L. (1963) *Enzyme and metabolic inhibitors*. Boston, USA: Academic Press.
- Webb, P. A. and Orr, C. (1997). *Analytical Methods in Fine Particle Technology*. Cornell University, New York: Micromeritics Instrument Corporation.
- Wehtje, E., Adlercreutz, P. and Mattiasson, B. (1993) 'Improved activity retention of enzymes deposited on solid supports', *Biotechnol. Bioeng.*, 41(2), 171–178.
- Werther, J. (2007) 'Fluidized-Bed Reactors', *Ullmann's Encyclopedia of Industrial Chemistry*, New York: Wiley-VCH Verlag GmbH & Co.
- Wesenberg, D., Kyriakides, I. and Agathos, S. N. (2003) 'White-rot fungi and their enzymes for the treatment of industrial dye effluents', *Biotechnology Advances*, 2003, 161–187.

- Wu, J. and Yu, H. Q. (2007) 'Biosorption of 2,4-dichlorophenol by immobilized white-rot fungus *Phanerochaete chrysosporium* from aqueous solutions', *Bioresour. Technol.*, 98(2), 253–259.
- Xavier, A. M. R. B., Tavares, A. P. M., Agapito, M. S. M. and Evtuguin, D. V. (2008) 'Sequential batch reactor for eucalypt kraft pulp effluent treatment with *Trametes versicolor*', *J. Chem. Technol. Biotechnol.*, 83(12), 1602–1608.
- Xu, L., Ke, C., Huang, Y. and Yan, Y. (2016) 'Immobilized *Aspergillus niger* Lipase with SiO₂ Nanoparticles in Sol-Gel Materials', *Catalysts*, 6(10), 149–161.
- Xu, Z., Cai, J.-G. and Pan, B.-C. (2013) 'Mathematically modeling fixed-bed adsorption in aqueous systems', *Applied Phys. Eng.*, 14(3), 155–176.
- Yahagi, T., Degawa, M., Seino, Y., Matsushima, T., Nagao, M., Sugimura, T. and Hashimoto, Y. (1975) 'Mutagenicity of carcinogenic azo dyes and their derivatives', *Cancer Lett.*, 1(1), 91–96.
- Yaithongkum, J., Kooptarnond, K., Sikong, L. and Kantachote, D. (2011) 'Photocatalytic Activity Against *Penicillium expansum* of Ag-doped TiO₂/SnO₂/SiO₂', *Adv. Key Eng. Mater.*, 214, 212–217.
- Yalçınkaya, Y., Soysal, L., Denizli, A., Arıca, M. ., Bektaş, S. and Genç, Ö. (2002) 'Biosorption of cadmium from aquatic systems by carboxymethylcellulose and immobilized *Trametes versicolor*', *Hydrometallurgy*, 63(1), 31–40.
- Yang, F. C. and Yu, J. T. (1996) 'Development of a bioreactor system using an immobilized white rot fungus for decolorization', *Bioprocess Eng.*, 16(1), 9–11.
- Yano, K. and Ishihara, S. (2006) 'Differential geometry in tangent bundle', *Kodai Math. Semin. Reports*, 18(4), 271–292.
- Yano, T. and Koga, S. (1969). Dynamic behavior of the chemostat subject to substrate inhibition. *Biotechnol. Bioeng.* 11(2), 139–153.
- Yi, Y., Neufeld, R. and Kermasha, S. (2007) 'Controlling sol-gel properties enhancing entrapped membrane protein activity through doping additives', *J. Sol-Gel Sci. Technol.*, 43(2), 161–170.
- Yordanova, M., Evstatieva, Y., Chernev, G., Ilieva, S., Denkova, R. and Nikolova, D. (2013) 'Enhancement of xylanase production by sol-gel immobilization of *Aspergillus awamori* K-1', *Bulg. J. Agric. Sci.*, 19(2), 117–119.
- Yum, K.-J. and Peirce, J. J. (1998) 'Biodegradation kinetics of chlorophenols in immobilized-cell reactors using a white-rot fungus on wood chips', *Water Environ. Res.*, 70(2), 205–213.

- Zhang, W. W., Wang, N., Zhang, L., Wu, W. X., Hu, C. L. and Yu, X. Q. (2014) 'Effects of additives on lipase immobilization in microemulsion-based organogels', *Appl. Biochem. Biotechnol.*, 172(6), 3128–3140.
- Zhao, X., Hardin, I. R. and Hwang, H. M. (2006) 'Biodegradation of a model azo disperse dye by the white rot fungus *Pleurotus ostreatus*', *Int. Biodeterior. Biodegrad.*, 57(1), 1–6.
- Zhu, Y., Kaskel, S., Shi, J., Wage, T. and van Pée, K.-H. (2007) 'Immobilization of *Trametes versicolor* Laccase on Magnetically Separable Mesoporous Silica Spheres', *Chem. Mater.*, 19(26), 6408–6413.
- Zuo, K. H., Zeng, Y.-P. and Jiang, D. (2010) 'Effect of polyvinyl alcohol additive on the pore structure and morphology of the freeze-cast hydroxyapatite ceramics', *Mater. Sci. Eng. C. Mater. Biol. Appl.*, 30(2), 283—287.