

PALM OIL EXTRACTION USING ENZYME MIXTURE TREATMENT

SONIA DILIP PATEL

A dissertation submitted in partial fulfilment of the
requirements for the award of the degree of
Master of Engineering

Faculty of Chemical and Energy Engineering
Universiti Teknologi Malaysia

JANUARY 2018

*Specially dedicated to my beloved family and friends for the continuous support,
encouragement and motivation*

ACKNOWLEDGEMENT

I am grateful to have given the strength and determination to complete this project, as part of requirement of Master Degree in Bioprocess Engineering. I could not have gone through this challenge without the support and guidance from the very talented and helpful lecturers from the Department of Bioprocess and Polymer Engineering, Faculty of Chemical and Energy Engineering, Universiti Teknologi Malaysia.

I would like to express my highest appreciation to two very caring supervisors, Dr. Syed Anuar Faua'ad bin Syed Muhammad and En. Nik Azmi bin Nik Mahmood for their encouragement, guidance, critics, advises and motivation. I would also like to extend my appreciation to En. Ya'akob Sabuddin and staffs in Department of Bioprocess and Polymer Engineering, who gave me the space as well as helped me to conduct my experiments.

I am also grateful to my family who have shown the support through challenging days. Lastly, I would like to extend my thank you to my course mates who have helped directly and indirectly. All these people have made the best out of me. Your kindness and cooperation is highly appreciated. Words alone are not sufficient to illustrate how much I owe you all.

ABSTRACT

In this study, application of aqueous enzymatic process to enhance the recovery of palm oil was studied. Experiments were conducted to study the effect of different combinations of enzyme mixture towards the percentage of oil extracted with respect to total oil content in palm mesocarp. The optimum combination of enzymes comprising of cellulase (X1), pectinase (X2), and xylanase (X3) for Aqueous Enzymatic Oil Extraction Process were determined using Simple Lattice mixture design (Design of Experiments). Maximum oil recovery of 85.95% was achieved with ratio of enzyme at 0.67:0.17:0.17 (X1: X2: X3), at enzyme loading of 30 mg protein/10 g substrate, substrate loading of 50% w/v, pH 4.8 and 2.0 hours of incubation at 50 °C. The concentration of reducing sugars at corresponding experimental runs was measured to evaluate the degree of hydrolysis and oil extracted. Concentration of reducing sugar trend was found not to be similar of the trend of oil extraction Analysis using Design Expert software for optimum condition showed a cellulase to xylanase ratio of 0.78:0.22 for 84.79% of oil recovery. A confirmation run performed produced 83.8% palm oil recovery.

ABSTRAK

Dalam kajian ini, penggunaan enzim untuk meningkatkan pengekstrakan minyak sawit telah dikaji. Eksperimen telah dijalankan untuk mengkaji kesan gabungan campuran enzim yang berlainan ke atas peratusan minyak yang diekstrak serta jumlah kandungan minyak dalam mesokarp sawit. Gabungan optimum enzim yang terdiri daripada selulase (X1), peptinase (X2), dan xylanase (X3) untuk Proses Pengekstrakan Minyak Enzimatik Aqueous telah ditentukan menggunakan reka bentuk campuran Simple Lattice (Reka Bentuk Eksperimen). Pengekstrakan minyak maksimum sebanyak 85.95% dicapai dengan nisbah enzim pada 0.67: 0.17: 0.17 (X1: X2: X3), pada pembebanan enzim 30 mg protein / 10 g substrate, pembebanan substrat sebanyak 50% w / v, pH 4.8 dan 2.0 jam inkubasi pada 50 °C. Kandungan gula dalam mesokarp diukur untuk menilai hubungkait antara tahap hidrolisis serta minyak yang diekstrak. Kepekatan mengurangkan trend gula didapati tidak sama dengan trend pengekstrakan minyak. Analisis menggunakan perisian Design Expert untuk mengetahui keadaan optimum menunjukkan nisbah selulase kepada xylanase 0.78: 0.22 akan menghasilkan pengekstrakan minyak sebanyak 84.79%. Ujikaji pengesahan yang dilakukan menghasilkan pengekstrakan minyak sawit sebanyak 83.8%

TABLE OF CONTENT

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iv
	ACKNOWLEDGEMENT	v
	ABSTRACT	vi
	ABSTRAK	vii
	TABLE OF CONTENT	viii
	LIST OF TABLES	xi
	LIST OF FIGURES	xii
	LIST OF ABBREVIATIONS	xiv
	LIST OF APPENDICES	xvi
1	INTRODUCTION	
	1.1 Background	1
	1.2 Problem Statement	3
	1.3 Objective	4
	1.4 Scope	5
2	LITERATURE REVIEW	
	2.1 Introduction	6
	2.2 Market Review	8
	2.3 Importance of Palm Oil as Hydrocarbon Production System	12
	2.4 Uses of Palm Oil	14
	2.4.1 Food Products	15
	2.4.2 Non-Food Products	16
	2.4.3 Biocomposites	16
	2.4.4 Nutritional, Nutraceutical and Pharmaceutical	17
	2.5 Industrial Processing	18
	2.5.1 Reception, Transfer and Storage of Fresh Fruit	18

	Bunches	
	2.5.2 Sterilization	18
	2.5.3 Stripping	19
	2.5.4 Digestion	19
	2.5.5 Crude Palm Oil Extraction	20
	2.5.6 Depericarping and Nut Fibre Separation	20
	2.5.7 Nut Cracking	21
	2.5.8 Palm Kernel Separation and Drying	21
2.6	Enzymes	22
	2.6.1 Xylanase	22
	2.6.2 Peptinase	23
	2.6.3 Cellulase	24
2.7	Solvent Extraction Strategy	24
2.8	Usage of Enzyme in Oil Extraction	27
2.9	Component of Plant Cell Wall	33
2.10	Design Expert and Simplex Lattice Mixture	36
3	METHODOLOGY	
	3.1 Experimental Flow Chart	37
	3.2 Materials	38
	3.3 Enzyme Protein concentration determination	39
	3.4 Sample Preparation	40
	3.5 Organic solvent extraction of palm oil	41
	3.6 Preparation of Buffer	42
	3.7 Aqueous enzymatic treatment	42
	3.8 Reducing sugar concentration determination	44
	3.9 Experimental Design	46
4	RESULTS AND DISCUSSION	
	4.1 Organic Solvent Extraction of Palm Oil	47
	4.2 Experimental Runs Results and Analysis	48

5	CONCLUSION AND RECOMMENDATION	
5.1	Conclusion	56
5.2	Recommendation	57
	REFERENCES	58
	APPENDICES	67

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Major centres of Oil Palm Cultivation	8
2.2	Comparison of plantation area in hectare	10
2.3	Palm oil production in two most producer of oil in the world over the years	13
2.4	Effect of enzyme type on oil recovery from moringa oleifera seed	31
2.5	Enzymatic extraction for different oil bearing material.	32
2.6	Effect of enzymatic treatment on oil yield	32
2.7	Minimum and maximum range for enzyme mixture involved in the study	36
3.1	Commercial enzymes used in this study and manufacturer	38
3.2	Dilution solutions for standards for standard protein curve determination	40
3.3	Dilution for glucose standard curve determination	45
3.4	Composition of enzyme mixture in simple lattice mixture design	46
4.1	Results for oil and reducing sugar	48

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Structure of palm oil fruit	7
2.2	Oil efficiency vs other major oil crops	9
2.3	Top five production regions	10
2.4	World major production of vegetable oil in 2012	11
2.5	Comparison of prices of major vegetable oils between 2001 to 2013 (USD per tonne)	11
2.6	Global Palm Oil use	15
2.7	Plant plasma membrane and cell-wall structure	34
3.1	Experimental work flow chart	37
3.2	Fresh oil palm fruit	38
3.3	Reaction schematic for the Coomassie Plus	39
3.4	Method of protein concentration determination.	39
3.5	Initial stages of sample preparation	41
3.6	Set up of Soxhlet apparatus	41
3.7	Left- Samples in conical flask in water bath at 50 °C for 2 hours. Right – the layers of residual solid and oil formed after serial centrifugation	43
3.8	Sample of oil-hexane before rotary evaporated at 70 °C	44
3.9	Conversion of DNS compound to 3-amino-5-nitro salicylic acid	44
4.1	Oil extraction yield % and reducing sugar yield (mg/mL) for aqueous enzymatic process	49
4.2	Structure of palm oil fruit	50

4.3	Result analysis using Design Expert® software.	52
4.4	Optimum ratio of enzymes	53
4.5	A mixture surface plot	54
4.6	Contour plot of oil recovered percentage	55

LIST OF ABBREVIATION

α	-	alpha
β	-	beta
$^{\circ}\text{C}$	-	degree Celcius
μ	-	micro
%	-	percentage
AEOE	-	aqueous enzymatic oil extraction
AEOEP	-	aqueous enzymatic oil extraction process
AEP	-	aqueous extraction process
BSA	-	bovine serum albumin
BOD	-	biochemical oxygen demand
C	-	carbon
CBHII	-	Cellobiohydrolase II gene
COD	-	chemical oxygen demand
CPO	-	crude palm oil
CSTR	-	continuous stirred tank reactor.
DNS	-	dinitrosalicylic acid
EBB	-	empty fruit branches
ETP	-	effluent treatment plant.
FFA	-	free fatty acid
FFB	-	fresh fruit branches
G	-	gram
Ha	-	hectare
LDL	-	low density lipoprotein
Kg	-	kilogram
M	-	meter
M	-	Molar
MDF	-	medium density fibreboard

mg/L	- milligram per Litre
Nm	- Nano
OD	- optical density
pI	- isoelectric point
POME	- palm oil mill effluent
ppm	- parts per million
RBDO	- refined, bleached and deodorized oil
Rpm	- revolution per minute
T	- tonne
TAG	- triacylglycerol
UASB	- up-flow anaerobic sludge blanket
UASFF	- up-flow anaerobic sludge fixed-film
USD	- United States Dollar
USDA	- United States Department of Agriculture.
v/w	- volume by weight
w/w	- weight by weight

LIST OF APPENDICES

Appendix NO.	TITLE	PAGE
I	Standard Curve for Protein Concentration Determination	72
II	Protein Concentration Determination	74
III	Reducing Sugar Concentration Determination	76

CHAPTER 1

INTRODUCTION

1.1 Background

Palm oil is one of the 17 major oils and fats produced and traded worldwide (Jaafar and Sukaimi, 2001). It is widely used as edible product and its importance is increasing as dietary component for over one billion people. Edible fats are used as vegetable oil to produce margarine, shortenings and functional food. The dietary trend today is to replace animal fats with vegetable origin fats. Although oil palm diet can lead to higher cholesterol as compared to corn, soyabean, safflower seed and sunflower oil, intake of palm oil leads to endogenous cholesterol level to drop (Edem, D.O, 2002). It is believed that this is assisted by the presence of tocotrienols and peculiar isomeric position of fatty acids. Palm oil benefits to health include reduction in risk of arterial thrombosis and atherosclerosis, aggregation of platelet, inhibition of endogenous cholesterol biosynthesis and reduction in blood pressure (Edem, D.O, 2002).

The oil palm was introduced from West Africa to the Bogor Botanical Gardens, Indonesia in 1848, arrived on Malaysian shores in 1871 and known an ornamental or decorative plant at that time. In less than 100 years oil palm has moved from being a relatively minor subsistence crop in West and Central Africa to one of the world's major agricultural commodities (Wicke *et al*, 2011). Malaysia and Indonesia are two largest producers accounting for approximately 85% of world's oil palm production (Sulaiman *et al* 2011).

The oil palm was commercially exploited as an oil crop only from 1911 when the first oil palm estate was established (Basiron *et al*, 2000). The fruits produce two main products – crude palm oil from the palm fruit oil (outer mesocarp) and palm kernel oil from the kernel within the fruit. In December 2015, Malaysia has produced 1.4 million tonnes of crude palm oil and 200 thousand tonnes of kernel oil (MPOB 2001). This shows that we are the major key player in palm oil industry and contributing to economic growth.

The tree grows up to 20 to 30 meters high, has an economic life span of 25 to 30 years. The female bunch bears about 2500-3000 fruits borne on 100-120 spikelet attached to a peduncle from the axil of a frond and weighs as much as 30-40 kg (MPOB 2001). Palm oil is extracted from highly perishable oil palm fruit through a series of processing which involves harvesting, sterilization, stripping, digestion, clarification, purification, vacuum drying and nut recovery (Basiron *et al* 2000).

There have been a significant number of researches on enzymatic oil extraction from plant seeds such as rapeseed, soybean, coconut, avacoda, sunflower and peanut. Oil extracted from this seeds has been promising, yielding about 60-90%, mainly depending on enzyme used and other contributing factors such as oilseeds size, pH, time, temperature, solid water ratio, moisture, number of extraction stages and agitation degree (Cater C M *et al.*, 1974). Research by Rosenthal A, *et al*, 1996 showed that palm oil has very high oil content, of about 97.7%, followed by coconut 80-90%, soybean, 86% and avocado, 75%. This research would be

highlighting on the extraction of oil from palm oil by using different combination of enzymes as this has a potential to recover 80-90% of oil content in the palm oil. To further support this, there have been researches conducted on palm oil extraction using mixture of enzymes with good output yield of about 80-90% (Silvamany, H. & Jahim, J.M. 2015). This research is intended to study the different combination of three enzymes as compared to previous research and to see how different combination enzymes affects the oil output.

1.2 Problem Statement

The ideal composition of palm fruit bunch is usually as such – kernel per fruit: 5-8%, mesocarp per fruit: 85-92%, oil per mesocarp: 20-25%, oil per bunch: 23-25%. In the palm oil industry, the complete process of extraction of edible oil from oil palm involves mechanical pressing at temperature ranging from 90 °C to 140 °C. Generally, fresh fruit bunches undergo sterilization process at 140 °C for about 75 to 90 minutes to deactivate hydrolytic enzyme responsible for the breakdown of oil to free fatty acid (FFA) and also to loosen the fruits on the bunch to facilitate stripping (Mohammad N.E. *et al.*, 2015).

Separated fruits are then heated in a digester aided with rotating paddle impeller at a temperature of 85 to 90 °C to mash the fruit which results in release of 20 to 30 % of free oil from fruit mesocarp. The crude palm oil is extracted with a screw press under high pressure and then clarified to remove dirt, fibres or gums. The crude palm oil is further processed to obtain refined, bleached and deodorized oil (RBDO). The oil that was not extracted remains in solid residue and end up as waste oil. However, aqueous enzymatic oil extraction can be employed in our palm oil industry due to its potential as an environmentally cleaner alternative technology for oil extraction and produce significant increase in oil yield. The release of oil facilitated by cell wall degrading enzyme is able to exhibit greater than 90% oil

extraction efficiency (Rosenthal, A. *et al.*, 1996). Hence, enzymatic oil extraction is a promising area of study as compared to other oil extraction methods and is chosen as part of research for this study. Design expert® software with process variable of Simplex lattice is utilised in this study to analyse the outcome of the research. This class of design is chosen since a simplex lattice analyses mixture variable only provided all components have same range and no constrains on design space.

Aqueous enzymatic oil extraction from plant material is said to increase oil yield. The combination of enzymes which favours the oil extraction from mesocarp might not be at its optimum. Suitable combination is not exactly known. Since no single enzyme is adequate for the efficient maceration and extraction of oil, the best combination ratio of cellulose, xylanase and pectinase enzymes can give the highest oil extraction (Faveri D.D. *et al.*, 2008). Duration of reaction and optimum temperature will be fixed in order to study the relationship between percentage of oil extraction and reducing sugar concentration. The concentration of reducing sugar may give an indication of the extent of breakdown of cell wall in the palm oil mesocarp. A higher reducing sugar concentration indicates a higher degree of breakdown of cellulose to simpler forms of sugar, thus increasing the oil extraction percentage. Hence, the correlation of trend of reducing sugar concentration and percentage of oil extraction is not known and will be studied in this research. This is vital to further understand the behaviour of enzymatic reactions.

1.3 Objective

The objectives of this research are:

1. To formulate best enzyme mixture for aqueous enzymatic oil extraction by using Design Expert® version 10.

2. To study the reducing sugar concentration formation in an aqueous oil extraction process. This may provide a degree of knowledge on enzymatic degradation of cell wall.

1.4 Scope

The scopes of study include:

1. Pre-treatment of palm oil mesocarp.
2. Enzyme protein concentration determination.
3. Formulation of best enzyme mixture, based on different combination of enzyme mixture.
4. Aqueous oil extraction by organic solvent in a Soxhlet apparatus.
5. Analysis of reducing sugar concentration by using Dinitrosalicylic acid (DNS) method.

REFERENCES

- Abdulkarim, S.M., Lai, O.M., Muhammad, S. K. S., Long, K., and Ghazali, H.M. (2006). Use of enzymes to enhance oil recovery during aqueous extraction of *Moringa Oleifera* Seed oil. *Journal of Food Lipids*. 13(2), 113-130.
- Adler-Nissen, J. (1986). Enzyme hydrolysis of food proteins. Elsevier Applied Science Publishers, London and New York. pp. 427.
- Aliyu, S. and Zahangir, A. (2012). Palm oil mill Effluent: A waste or a raw material *Journal of Applied Sciences Research*, 8(1), 466-473.
- Aparna, S., Khare, S.K. and Gupta, M. N. (2002). Enzyme-assisted aqueous extraction of peanut oil. *J. Am. Oil Chem. Soc.* 79, 215-218.
- Athel, Cornish-Bowden. (2013). The Origins of Enzyme Kinetics. *FEBS Letters*. 587, 2725-2730.
- Badr, F. and Sitothy, M. Z. (1992). Optimizing conditions for enzymatic extraction of sunflower oil. *Grasas y Aceites* 43, 281-283.
- Bair, C.W, and Snyder, H.E. (1980). Electron microscopy of soybean lipid bodies. *J Am Oil Chem Soc*, 279-82.
- Basiron, Y, Jalani, B.S., and Chan K.W. (2000). Advances in oil palm research. Volume I and Volume II, MPOB Bangi. P.1-782, 783-1526.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology* 109. Wiley InterScience, Weinheim, pp 289-295.
- Be Miller, J. N. (1986). An Introduction to pectins: Structure and properties. In: Fishman, M.L., Jem, J. J. (Eds.), *Chemistry and Functions of Pectins*, ACS Symposium Series 310. American Chemistry Society, Washington DC.
- Borja, R. and Banks, C.J. (1994a). Anaerobic digestion of palm oil mill effluent using an up-flow anaerobic sludge blanket reactor. *Biomass and Bioenergy* 6, 381–389.
- Borja, R. and Banks, C. J. (1994b). Treatment of palm oil mill effluent by upflow anaerobic filtration. *Journal of Chemical Technology and Biotechnology* 61, 103–109.

- Borja, R., Banks, C.J. and Sánchez, E. (1996). Anaerobic treatment of palm oil mill effluent in a two-stage up-flow anaerobic sludge blanket (UASB) reactor. *Journal of Biotechnology* 45, 125–135.
- Boyer, R. (2006). Biochemistry laboratory: modern theory and techniques, Benjamin Cummings, Pearson Prentice Hall. New York.
- Buenrostro, M, and Lopez-Munguia, C.A. (1986). Enzymatic extraction of avocado oil. *Biotechnology Letters*. 8, 505-506.
- Cater, C. M., Rhee, K. C., Hagenmaier, R. D., & Maltil, K. F. (1974). Aqueous extraction- An alternative oilseed milling, *J Am Oil Chem Soc*, 51, 137-141.
- Chan, K. W. (2002). Oil Palm sequestration and carbon accounting : Our global strength. Proc. Malaysian Palm Oil Association Seminar on R&D for Competitive Edge in the Malaysian Palm Oil Industry. 19-20 March 2002. In pres. pp 17.
- Cheah, S. C., Augustin, M. A., and Ooi, L. C. L. (1990). Enzymatic extraction of palm oil. *Palm Oil Res. Bull. Malaysia* 20, 30-36.
- Choo, Y. M. and Nesaretnam, K. (2014). Research advancements in palm oil nutrition. *European Journal of Lipid Science and Technology*. 116(10), 1301-1315.
- Corley, R. H. V. and Tinker, P. B. (2003). The Oil Palm. Fourth edition, Blackwell, Oxford, United Kingdom.
- Deng, Y., Pyle, D. L. and Niranjana, K. (1992). Studies of aqueous enzymatic extraction of oil from rapeseed. In: Agricultural Engineering and Rural Development I Conference Proceedings. Beijing, China. Vol 1: 190-5.
- Dominguez, H., Hunez, M. J., and Lema, J. M. (1994). Enzymatic pretreatment to enhance oil extraction from fruits and oilseeds: A review. *Food Chemistry* 49, 271-286.
- Dominguez, H., Hunez, M. J., and Lema, J. M. (1995). Aqueous processing of sunflower kernels with enzymatic technology. *Food Chemistry*. 53(4): 427-434.
- Dominguez, H., Nunez, M. J., and Lema, J. M. (1994). Enzymatic pretreatment to enhance oil extraction from fruits and oilseeds-a review. *Food Chemistry*, 49(3), 271-286.

- Dominguez, H., Nunez, M.J. and Lema, J.M. (1994). Enzymatic pretreatment to enhance oil extraction from fruits and oilseeds: A review. *Food Chemistry*. 49, 271-286.
- Dourado, F., Barros, A., M., Coimbra, M. A., and Gama, F. M. (2004). Anatomy and cell wall polysaccharides of almond (*Prunus dulcis* D.A. Webb) seeds. *Journal of Agricultural and Food Chemistry*, 52(5), 1364-1370.
- Dourado, F., Vasco, P., Barros, A., Mota, M., Coimbra, M. A., and Gama, F. M. (2003). Characterisation of Chilean hazelnut (*Gevuina avellana*) tissues: Light microscopy and cellwall polysaccharides, *Journal of the Science of Food and Agriculture*, 83(3): 158-165.
- Dourado, F., Vasco, P., Gama, F. M., Coimbra, M. A., & Mota, M. (2000). Characterisation of Rosa Mosqueta seeds: Cell wall polysaccharide composition and light microscopy observations, *Journal of the Science of Food and Agriculture*, 80(13), 1859-1865.
- Dusterhoft, E. M., Bonte, A. W., Venekamp, J. C. and Voragen, A. G. J. (1993) The role of fungal polysaccharidases in the hydrolysis of cell wall materials from sunflower and palm-kernel meals. *World Journal of Microbiology and Biotechnology* 9, 544-554.
- Eapen, K. E., Kalbag, S. S., and Subrahmanyam, V. (1966). Operations in the wetrendering of peanut for the separation of protein, oil and starch. *Journal of the American Oil Chemists Society*, 43, 585-9.
- Edem, D. O. (2002). Palm Oil: Biochemical, physiological, nutritional, hematological and toxicological aspects: A review. *Plant Foods for Human Nutrition*. 57, 319-341.
- Food and Agricultural Organisation. (2013). World Agriculture trends: towards 2015-2013. An FAO Perspective [http://www.fao.org/docrep/005/y4252e/y4252e05d.htm].
- Food and Agricultural Organisation. (2016). FAOSTAT database collections, Food and Agricultural Organisation of the United Nations. Rome. Data average 2011-2013.
- Faveri, D. De., Aliakbarian B., Avogadro. M., Perego P., and Converti A. (2008). Improvement of olive oil phenolics content by means of enzyme formulation: effect of different enzymatic activities and concentrations. *Biochemical Engineering*. 41, 149-156.

- Food and Agricultural Policy Research (2010)b, Food and agricultural commodities production statistics: Indonesia and Production Indices: Indonesia accessible at: <http://faostat.fao.org>, accessed September 2010.
- Fullbrook, P. D. (1983). The use of enzymes in the processing of oilseeds. *Journal of the American Oil Chemists Society*, 60 (2):428A-430A.
- Fullbrook, P. D. (1983). The use of enzymes in the processing of oilseeds. *Journal of the American Oil Chemists Society*, 60(2), 476-478.
- Fullbrook, P. D. (1983). The use of enzymes in the processing of oilseeds. *Journal of the American Oil Chemists' Society* 60, 476-478.
- Gandhi, Y. S., Bankar, V. H., Vishwakarma, R. P., Satpute, S.R., and Upkare, M. M. 2017. Reducing sugar determination of jaggery classical lane and eynon method & 3,5-Dinitrosalicylic acid method. *Imperial Journal of Interdisciplinary research*. 3(6), 602.
- Gaur, R., Sharma, A., Khare, S.K., and Gupta, M.N. (2007). A Novel process for extraction of edible oils enzyme assisted three phase partitioning (EATPP). *Bioresource Technology* 98, 696-699.
- Glaeut, A. M. (1980). Fixation, dehydration and embedding of biology specimens, in practical methods in electron microscopy, A M Glaeut, ed (North-Holland, Amsterdam).
- Gomez, A. M., Lopez, C. P., and De la Ossa, E. M. (1996). Recovery of grape seed oil by liquid and supercritical carbon dioxide extraction: A comparison with conventional solvent extraction. *The Chemical Engineering Journal and the Biochemical Engineering Journal*, 61(3), 227-231.
- Guillaume, R., and Lionel, M. (2010). Influence of enzymes on the oil extraction process in aqueous media. *Journal of Technology-Innovation*, 17(6), 356-359.
- Gusakov, A. V., Kondratyeva E.G., and Sinitsyn, A.P. (2011). Comparison of two methods for assaying reducing sugars in the determination of carbohydrase activities. *International Journal of Analytical Chemistry*. Article ID 283658, 4 pages
- Hagenmaier, R D. (1974). Aqueous processing of full-fat sunflower seeds: yield of oil and protein, *Journal American Oil Chemists Society*, 51, 470-471.
- Hamada, I. S. and Marshall, W. E. (1989). Preparation and functional properties of enzymatically deamidated soy proteins. *Journal of Food Science*, 54, 598-635.

- Huang, A. H. C. (1994). Structure of plant seed oil bodies. *Curr Opin Structural Biol.* 4, 494-8.
- Jaafar, M., and Sukaimi. M. (2001). The future of palm oil in the new millennium in Malaysia. *Borotrop Bulletin*, 16, 10-13.
- Jarvis, D., Richmond, N. Phua, K. H., Pococok, N., Sovacool, B. K. and D'Agostino, A. (2010). Palm Oil in South-east Asia. *Asian Trends Monitoring Bulletin*, 2-12.
- Karlovic, D., Bocevska, M., Jakovlevic, J., and Turkulov, J. (1994). Corn germ oil extraction by a new enzymatic process. *Acta Alimentaria*, 23, 389-400.
- Kashyap, D. R., Vohra, P. K., Chopra, S., and Tewari, R. (2000). Applications of pectinases in the commercial sector: A review. *Bioresources Technology* 77, 215-227.
- Kilara, A., (1982). Enzymes and their uses in the processed apple industry: A review. *Process Biochemistry Journal.* 23, 35-41.
- Lamsal, B. P., Murphy, P. A., and Johnson, L. A. (2006). Flaking and extrusion as mechanical treatments for enzyme-assisted aqueous extraction of oil from soybeans. *Journal American Oil Chemists Society*, 83(11), 973-979.
- Lawhon, J. T., Manak, L. J., Rhee, K. C., & Lusas, E. W. (1981). Production of oil and protein food product from raw peanuts by aqueous extraction and ultrafiltration, *Journal of Food Science*, 46, 391-395.
- Lusas, E. W., Lawhon, J. T., & Rhee, K. C. (1982). Producing edible oil and protein from oilseeds by aqueous processing. *Oil Mill Gazette*, 4, 28-34.
- Mccarthy, J. F. (2010). Process of inclusion and adverse incorporation: Oil palm and agrarian change in Sumatra, Indonesia. *The Journal of Peasant Studies*, 37, 821-850.
- Mcglone, O. C., Lopez-Munguia, C. A., and Carter, J. V. (1986). Coconut oil extraction by a new enzyme process. *Journal of Food Science.* 51, 695-697.
- Mohammad, N. E., Kamonwan, P., Nuttawan, Y., and Jarupan, K. (2015). Enhanced oil extraction from palm fruit mesocarp using technical enzymes. *International Journal of Advances in Science Engineering and Technology*, ISSN: 2321-9009, 3(1).

- Motta, F. L., Andrade, C. C. P., and Santana, M. H. A. (2013). A Review of xylanase production by the fermentation of xylan: Classification, Characterization and Applications. Sustainable Degradation of Lignocellulosic Biomass-Techniques, Applications and Commercialization. Chapter 10.
- MPOB. (2001). Oil Palm Statistics. 21st edition, MPOB, Bangi. 131 pp.
- Murphy, D. J. (2010). Manipulation of oil crops for industrial applications. Industrial Crops and Uses (Singh, B P ed.). CABI Press, UK. p. 183-206.
- Murphy, D. J. (2014). The future of oil Palm As A Major Global Crop: Opportunities and Challenges. *Journal of Oil Palm Research*. 26(1), 1-24.
- Najafpour, G. D., Zinatizadeh, A. A. L., Mohamed, A. R., Hasnain, I. M., and Nasrollahzadeh, H. (2006). High-rate anaerobic digestion of palm oil mill effluent in an upflow anaerobic sludge-fixed film bioreactor. *Process Biochemistry* 41, 370–379.
- Oil World (2012). <http://www.oilworld.biz/app.php>.
- Ong, A. S. H. and Goh, S. H. (2002). Palm Oil: A healthful and cost-effective dietary component. *Food and Nutrition Bulletin*, 23(1): 11-22.
- Passos, C. P., Yilmaz, S., Silva, C. M. and Coimbra, M. A. (2009). Enhancement of grape seed oil extraction using a cell wall degrading enzyme cocktail. *Food Chemistry*. 115(1), 48-53.
- Perez, E. E., Fernandez, M. B., Nolasco, S. M. and Crapiste, G. H. (2013). Effect of pectinase on the oil solvent extraction from different genotypes of sunflower (*Helianthus annuus L.*) *Journal of Food Engineering*, 117, 393-398.
- Rathi, C. L., Pradhan, S., Javvadi, S. and Wani, A. (2012). An enzyme composition and process for extracting oil from palm oil fruits. WO2012011130 A3.
- Rosenthal, A, Pyle, D. L., Niranjana, K, Gilmour, S. and Trinca, L. (2001). Combined effect of operational variables and enzyme activity on aqueous enzymatic extraction of oil and protein from soybean. *Enzyme and Microbial Activity*. 28(6): 499-509.
- Rosenthal, A, Pyle, D. L., Niranjana, K. (1998). Mechanisms in the simultaneous aqueous extraction of oil and protein from soybean. Transactions of The Institute of Chemical Engineers. Part C. Food and Bioproducts. 76, 224-30.
- Rosenthal, A, Pyle, D. L, Niranjana, K. (1996). Aqueous and enzymatic processes for edible oil extraction (review). *Enzyme Microbial Technology*. 19(6), 204-40.

- Rosenthal, A., Pyle, D. L. and Niranjana, K. (1996). Aqueous and enzymatic process for edible oil extraction. *Enzyme Microbial Technology*, 19(6), 402-420.
- Rosenthal, A., Pyle, D.L., and Niranjana, K. (1996). Aqueous and enzymatic processes for edible oil extraction. *Enzyme and Microbiology Technology*, 19, 402-420.
- Rosenthal, A., Pyle, D.L., Niranjana, K., (1996). Aqueous enzymatic processes for edible oil extraction. *Enzyme and Microbiology Technology*, 19, 402-420.
- Rosenthal, A., Pyle, D.L., Niranjana, K., Gilmour, S. and Trinca, L., (2001). Combined effect of operational variables and enzyme activity in aqueous enzymatic extraction of oil and protein from soybean. *Enzyme and Microbiology Technology*, 28, 499-509.,
- Rosillo-Calle, Pelkmans, L. and Walter A. (2007). A Global Overview of Vegetable Oils with reference to Biodiesel. A report for the IEA Bioenergy Task 40, International Energy Agency.
- Roy, I. and Gupta, M. N. Current trends in affinity-based separation of proteins/enzymes, *Current Science*, 78, 587-591.
- Sarker, B. C., Singh, B. P. N., Agrawal, Y. C. and Gupta, D. K. (1998). Optimization of enzyme pretreatment of rapeseed for enhanced oil recovery. *Journal of Food Science and Technology*. 35, 183-186.
- Saupe, S.G. (2011). College of St. Benedict/ St. John's University. Biology Department; Collegeville, MN 56321; (320) 363 – 2782.
- Silvamany, H. and Jahim, J. M. (2015). Enhancement of palm oil extraction using cell wall degrading enzyme Formulation. *Malaysian Journal of Analytical Sciences*. Vol 19(1), 77-87.
- Silverstein, R. A, Y. Chen, R. R., Sharma-Shivappa, M. D. Boyette, and Osborne, J. (2007). A comparison of chemical pretreatment methods for improving saccharification of cotton stalks. *Bioresource Technology*, 98 : 3000-3011.
- Sime Darby Plantation, Palm Oil Fact and Figures. [www.simedarby.com/upload/Palm_Oil_Facts_and_Figures.pdf]
- Sineiro, J., Dominguez, H., and Lema, J. M. (1998). Optimization of the enzymatic treatment during aqueous oil extraction from sunflower seeds. *Food chemistry*. 61(4): 467-474.

- Sineiro, J., Dominguez, H., Nunez, M. J. and Lema, J. M. (1997). Optimization of the enzymatic treatment during aqueous oil extraction from sunflower seed. *Journal of Food Chemistry*, 61 (4): 467-474.
- Singh, A. (1999). Engineering enzyme properties. *Indian Journal of Microbiology*, vol. 39, no. 2, pp. 65–77.
- Sluiter, A., Hames, B., Ruiz, R., Scarlata, C., Sluiter, J., Templeton, D. and Crocker, D. (2001). Determination of Structural carbohydrates and lignin in Biomass. Golden, CO:U.S. Department of Energy National Renewable Energy Laboratory.
- Sosulski, K., Sosulski, F. W., and Coxworth, E. (1988). Carbohydrase hydrolysis of canola to enhance oil extraction with hexane. *Journal American Oil Chemists Society*, 65(3), 357-61.
- Subramanian, N. (1959). Integrated processing of peanut for the separation of major constituents. *J Am Oil Chem Soc* 36, 66-70.
- Sukumaran, R. K., Singhania, R. R., and Pandey A. (2005). Microbial cellulases-production, applications and challenges. *Journal of Scientific and Industrial Research*, vol. 64, no. 11, pp. 832–844.
- Sulaiman, F., Abdullah, N., Gerhauser, H. and Shariff, A. (2011). An Outlook of Malaysian energy, oil palm industry and its utilization of wastes as useful resources. *Biomass Bioenergy* 35, 3775-3786.
- Tano-Debrah, K., Yoshimura, Y. and Ohta, Y. (1996) Enzyme-assisted extraction of shea fat:evidence from light microscopy on the degradation effects of enzyme treatment on cells of shea kernel meal. *Journal of the American Oil Chemists' Society* 73, 449-453.
- USDA. (2012). Malaysia: stagnating palm oil yields impede growth. Commodity Intelligence Report (December 2012). <http://www.pecad.fas.usda.gov/highlights/2012/12/Malaysia/>
- Wicke, B., Sikkema, R., Dornburg, V. and Faaij, A. (2011). Exploring land use changes and the role of palm oil production in Indonesia and Malaysia. *Land Use Policy* 28: 193–206.
- Winkler, E, Foidl, N., Gubitz, G.M., Staubman, R. and Steiner, W. (1997). Enzyme supported oil extraction from *Jatropha curcas* seeds, *Applied Biochemistry Biotechnology*, 65, 449-156.

Yusof, B. and Chan, K. W. (2004). The palm oil and its sustainability. *Journal of Oil palm Research*. 16(01), 1-10.