

IN VITRO AND *IN SILICO* ANTICANCER ACTIVITIES OF *ANNONA MURICATA* LINN LEAVES EXTRACTS ON LUNG CANCER CELLS

MOHAMAD NORISHAM BIN MOHAMAD ROSDI

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (*Bioprocess Engineering*)

School of Chemical and Energy Engineering
Faculty of Engineering
Universiti Teknologi Malaysia

OCTOBER 2019

DEDICATION

A little gift.

ACKNOWLEDGEMENT

This thesis is a reflection of my journey over the past few years. It contains joy and misery; it represents my sweat and tears. In completing this thesis, I owe a great debt of appreciation to countless number of individuals. I would like to thank the many people who have contributed to the production of this thesis. First and foremost I thank Dr Harisun Ya'akob for her extensive help and guidance. I would also like to thank Dr Rauzaden Mohamed Zulkifli for his valuable support. I frequently consulted my scientific colleagues which have without exception been inspiring. Conversation with them were indispensable. In fact, the determination and tenacity of some individuals had inspired me in certain way. I also thank the staff in Institute of Bioproduct Development (IBD), University Industry Research Institute (UURL), School of Chemical and Energy Engineering, and School of Biosciences. I could not have written this thesis without their excellent support. I've had great support from family and friends and I hope they'll forgive me for not mentioning them all by name. Lastly, I hope that this thesis will assist the interested reader to get information of anticancer study of *Annona muricata* Linn.

ABSTRACT

Lung cancer specifically non-small cell lung cancer (NSCLC) is one of the most devastating cancers. Despite having diverse treatment methods such as surgery, chemotherapy, radiation and targeted therapies, the overall 5-year survival rate for NSCLC is accounted for only 18.2%. The high mortality rates for NSCLC are partially due to the lack of effective prognostic factors such as biomarkers. Nowadays, plant-derived bioactive substances have been making way as potential anticancer agents. One such plant is *Annona muricata* Linn, which is also known as soursop or graviola. This plant has been widely reported to contain valuable phytochemical substances that could be developed as chemopreventive agents. The antiproliferative and anticancer activities of this tropical plant have been demonstrated in *in vitro* cell culture studies as well as in *in vivo* studies of animals. It has been discovered that *A. muricata* L. extract exerts inhibition against various number of cancer cells, involving multiple mechanism of actions. Nonetheless, the mode of action and the molecular interactions of the plant have not yet been unveiled for most of these mechanisms. In the current study, response surface methodology was utilized to optimize the ultrasonic-assisted extraction (UAE) and maceration extraction (ME) of *A. muricata* L. leaves. Central composite design was applied to optimize the antioxidant activity of both extracts. It has been found that ME extract showed high antioxidant activity as compared to UAE extract with 83.3% and 31.6% respectively. However, through cytotoxicity study that was done by using MTT assay, UAE extract exhibited significant cytotoxicity effects in NSCLC cell line (HLFa) with IC_{50} of 139.6 $\mu\text{g/mL}$ as compared to ME extract with IC_{30} of 108.4 $\mu\text{g/mL}$. The effect of the extracts on nitric oxide (NO) production in HLFa cells was also evaluated using Griess reagent system assay. Both extracts reduced the release of nitrite in the cell supernatant which indicated the reduction in NO production. Caspase 3/7 apoptosis assay was used to detect the presence of apoptotic machinery in HLFa cells after incubation. The mRNA expression of several genes namely *HMGB1*, *BCL2* and *BAX* were quantified. The mechanistic evaluation of the results showed the possibility of involvement of these genes in *A. muricata* L. anticancer effects. At the end of this study, *in silico* molecular docking interaction of several phytoconstituents of *A. muricata* L. was analysed against Bcl-2 antiapoptotic proteins namely Bcl-2, Bcl-w and Mcl-1. The stability of complexes formed was evaluated using molecular dynamic simulation. Anonaine was also detected in UAE and ME extracts through HPLC screening process with 10.6 ppm and 10.7 ppm, respectively.

ABSTRAK

Kanser paru-paru terutamanya kanser paru-paru sel bukan kecil (NSCLC) merupakan salah satu kanser yang merbahaya. Walaupun terdapat kaedah-kaedah rawatan yang pelbagai seperti pembedahan, kemoterapi, terapi radiasi bersasar, namun kadar kemandirian keseluruhan dalam 5 tahun hanya mencecah sebanyak 18.2 %. Kadar kematian pesakit NSCLC yang tinggi sebahagiannya adalah disebabkan kekurangan cara yang efektif untuk mengenalpasti penyakit contohnya melalui penanda-penanda bio. Masa kini, bahan-bahan bioaktif bersumberkan tumbuh-tumbuhan berpotensi sebagai agen antikanser. Salah satu tumbuhan berpotensi tersebut ialah *Annona muricata* Linn ataupun juga dikenali sebagai durian belanda atau graviola. Tumbuhan ini telah dilaporkan dengan meluas mempunyai bahan fitokimia yang bernilai yang berkemungkinan dapat dibangunkan sebagai agen penghalang perkembangan kanser. Tumbuhan tropika ini telah menunjukkan aktiviti antiproliferatif dan antikanser melalui kajian kultur sel secara *in vitro* dan kajian terhadap binatang secara *in vivo*. Ekstrak *A. muricata* L. dilihat dapat memberi kesan perencatan terhadap pelbagai jenis sel kanser, yang meliputi pelbagai jenis mekanisme tindakbalas. Walaubagaimapun, cara tindakbalas dan interaksi molekul yang ditunjukkan oleh tumbuhan ini masih belum dapat dirungkai sepenuhnya. Dalam kajian ini, kaedah sambutan permukaan telah digunakan untuk mengoptimumkan proses pengestrakan daun *A. muricata* L. menggunakan kaedah ultrasonik (UAE) dan kaedah penyusutan (ME). Rekabentuk komposit berpusat telah digunakan untuk mengoptimumkan aktiviti antioksidasi bagi kedua-dua ekstrak. ME telah menunjukkan aktiviti antioksidasi yang tinggi dengan berbanding UAE dengan masing-masing merekodkan bacaan 83.3 % dan 31.6 %. Namun begitu, melalui kajian tahap toksik terhadap sel yang telah dilakukan menggunakan teknik MTT, UAE menunjukkan kesan toksik yang lebih ketara terhadap sel NSCLC (HLFa) dengan IC_{50} 139.6 $\mu\text{g/mL}$ berbanding dengan ekstrak ME dengan IC_{30} 108.4 $\mu\text{g/mL}$. Kesan kedua-dua ekstrak terhadap pengeluaran nitrik oksida (NO) dalam sel-sel HLFa juga telah dinilai menggunakan *Griess reagent system*. Ekstrak-ekstrak ini dilihat mengurangkan pelepasan nitrit di dalam media yang dapat disimpulkan sebagai pengurangan pengeluaran NO. Teknik apoptosis Caspase 3/7 telah digunakan untuk mengesan sel-sel HLFa yang mengalami apoptosis selepas inkubasi. Ekspresi gen mRNA seperti *HMGB1*, *BCL2* dan *BAX* telah dinilai. Penilaian keputusan ekspresi gen telah menunjukkan kemungkinan penglibatan gen-gen ini di dalam tindakbalas antikanser *A. muricata* L. Di akhir kajian ini, interaksi dok molekul *in silico* antara beberapa komponen fitokimia dari *A. muricata* L. dengan protein-protein seperti Bcl-2, Bcl-w dan Mcl-1 telah dianalisa. Kompleks yang terhasil dari interaksi tersebut telah dinilai kestabilannya menggunakan proses pemilihan simulasi molekul dinamik. Kewujudan anonaine di dalam UAE dan ME telah disahkan melalui teknik HPLC masing-masing dengan kepekatan 10.6 ppm dan 10.7 ppm.

TABLE OF CONTENTS

	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xii
	LIST OF FIGURES	xiv
	LIST OF ABBREVIATIONS	xvii
	LIST OF SYMBOLS	xx
	LIST OF APPENDICES	xxi
CHAPTER 1	INTRODUCTION	1
1.1	Research Background	1
1.2	Problem Statement	4
1.3	Research Objectives	5
1.4	Scope of the Study	6
1.5	Significances of the Study	6
1.6	Thesis Structure and Organization	7
CHAPTER 2	LITERATURE REVIEW	9
2.1	Plant-derived Anticancer Substances	9
2.2	<i>A. muricata</i> L.	12
2.2.1	<i>A. muricata</i> L.: An Anticancer Viewpoint	13
2.2.2	Anticancer Research of <i>A. muricata</i> L. Extract in Lung Cancer	14
2.3	Annonaceous Acetogenins	16
2.4	Free Radicals and Antioxidants Compounds	17

2.5	Non Small Cell Lung Cancer	18
2.5.1	Risk Factors	18
2.5.1.1	Cigarette Smoking	19
2.5.1.2	Radon Gas	20
2.5.1.3	Asbestos	21
2.5.1.4	Genetics	21
2.5.2	The Prevalence of Lung Cancer	21
2.6	Inflammation and Lung Cancer	22
2.7	Nitric Oxide	23
2.7.1	The Role of Nitric Oxide in Cancer	24
2.7.2	Nitric Oxide Synthase Expression in Cancers	24
2.7.3	Nitric Oxide and Lung Cancer	25
2.7.4	Nitric Oxide as a Novel Cancer Therapeutic Agent	25
2.8	High Mobility Group Box 1	26
2.8.1	High Mobility Group Box 1 and Cancer	27
2.8.2	High Mobility Group Box 1 on the Prognostic Features of Non-Small Cell Lung Cancer Cells	29
2.9	Cell Death	30
2.9.1	Apoptosis	30
2.9.1.1	Bcl-2 Family Protein and Mitochondrial Membrane Permeabilization	31
2.9.1.2	Caspase Cascade	33
2.9.1.3	Bcl-2 Antiapoptotic Proteins as Therapeutic Target for Cancer Treatment	34
2.9.1.4	Small Molecule Inhibitors	35
2.9.2	Necrosis	36
2.9.3	Necroptosis	38
2.9.3.1	Necroptosis in Cancer Therapy	40
2.10	Annonaceous Acetogenins Inhibit Mitochondrial Complex I	41
2.11	Extraction	43

2.11.1	Pre-extraction Preparation of the Plant Samples	43
2.11.2	Extraction Methods	43
2.11.2.1	Maceration	44
2.11.2.2	Ultrasonic-assisted Extraction (UAE) or sonication extraction	44
2.11.2.3	Optimization and Response Surface Methodology	45
2.12	Molecular Docking for Virtual Screening of Natural Product for Drug Discovery	46
2.12.1	AutoDock Software	47
CHAPTER 3	RESEARCH METHODOLOGY	49
3.1	Overview of the Experimental Design	49
3.2	Preparation of the Plant Materials	50
3.3	Optimization and Preparation of the Extracts	51
3.3.1	DPPH Free Radical Scavenging Assay	51
3.3.2	Optimization of UAE	52
3.3.3	Optimization of ME	54
3.4	Annonaceous Acetogenins Qualitative Identification using HPLC	56
3.5	HLFa Cell Line	56
3.5.1	HSF1184 Cell Line	56
3.5.2	Thawing and Cells Propagation	57
3.5.3	Subculturing	57
3.5.4	Cell Counting	58
3.5.5	Cryopreservation	59
3.6	Cell Viability Assay (MTT)	59
3.7	Nitric Oxide Production Assay	60
3.8	Apoptosis Detection Assay	60
3.9	Isolation of Total RNA and gDNA Removal	61
3.9.1	DNase Treatment and RNA Purification	62
3.9.2	Reverse Transcription (cDNA Synthesis)	62

3.9.3	Real-time Polymerase Chain Reaction (RT-PCR)	63
3.10	Statistical Analysis	64
3.11	Molecular Docking Study	64
3.11.1	Ligand Preparation	65
3.11.2	Target Preparation	66
3.11.3	Docking Protocol	66
3.12	Protein Model Development	66
3.12.1	Protein Model Validation	67
3.12.2	Molecular Dynamic (MD) Simulation and Analysis	67
CHAPTER 4	RESULT AND DISCUSSION	69
4.1	Overview	69
4.2	Statistical Analysis and Model Fitting of UAE	70
4.2.1	Response Surface Analysis of Antioxidant Effect	72
1.1.1	Effect of Interactions among Variables on Antioxidant Effect in UAE	75
4.3	Statistical Analysis and Model Fitting of ME	78
4.3.1	Response Surface Analysis of Antioxidant Effect	80
4.3.2	Effect of Interaction among Variables on Antioxidant Effect in ME	82
4.4	Comparison Analysis between UAE and ME Techniques	85
4.5	Antiproliferation Activity of UAE and ME	86
4.6	Effect of the Extracts on Nitric Oxide Production	90
4.7	Caspase Activation Induced by UAE and ME	91
4.8	Regulation of Bax, Bcl-2 and HMGB1 at Gene Expression Level	93
4.8.1	Analysis on UAE and ME Effects on High Mobility Group Box 1	96
4.9	Molecular Docking Analysis	98
4.9.1	Binding Interactions with Bcl-2	99
4.9.2	Binding Interactions with Bcl-w	102

4.9.3	Binding Interactions with Mcl-1	104
4.10	Protein Model Validation	106
4.10.1	MD Simulation of Bcl-2/obatoclax and Bcl-2/anonaine Complexes	106
4.11	Potential Bcl-2 Antiapoptotic Protein Inhibitors	110
4.11.1	Bcl-2/anonaine Complex Stability Analysis using MD Simulation	112
4.12	HPLC Screening	114
CHAPTER 5	CONCLUSION AND RECOMMENDATIONS	117
5.1	Conclusion	117
5.2	Recommendations	118
REFERENCES		119
APPENDICES A-D		147-151

LIST OF TABLES

TABLE NO.	TITLE	PAGE
Table 2.1	Anticancer studies on <i>A. muricata</i> L. Adapted from [58] with slight modification	15
Table 2.2	Comparison of apoptosis, necrosis and necroptosis	39
Table 3.1	Level of the variables tested in the RSM	52
Table 3.2	Non-coded and coded levels of independent variables of experimental design	52
Table 3.3	Design of factorial with CCD	53
Table 3.4	Level of the variables tested in the RSM	54
Table 3.5	Non-coded and coded levels of independent variables of experimental design	54
Table 3.6	Design of factorial with CCD	55
Table 3.7	Components for cDNA synthesis	63
Table 3.8	Gene ID for real-time PCR	63
Table 3.9	List of ligands used in molecular docking study	65
Table 4.1	CCD and observed experimental antioxidant effect	70
Table 4.2	Analysis of variance for the extraction of antioxidant effect	71
Table 4.3	CCD and observed experimental antioxidant effect	78
Table 4.4	Analysis of variance for the extraction of antioxidant effect	79
Table 4.5	Binding strength of investigated ligands with the antiapoptotic proteins	98
Table 4.6	The anonaine interacting residues of the three Bcl-2 antiapoptotic proteins are summarized with the number of hydrophobic interactions and the number of hydrogen bonds.	100
Table 4.7	The coreximine interacting residues of the three Bcl-2 antiapoptotic proteins are summarized with the number of hydrophobic interactions and the number of hydrogen bonds.	101

Table 4.8	The obatoclax interacting residues of the three Bcl-2 antiapoptotic proteins are summarized with the number of hydrophobic interactions and the number of hydrogen bonds.	104
Table 4.9	Summary of model validation	106

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
Figure 2.1	Mechanism of action by anticancer agents in hallmarks of cancer [55]	11
Figure 2.2	<i>A. muricata</i> L. leaves. The picture was taken at Institute of Bioproduct Development, Universiti Teknologi Malaysia	12
Figure 2.3	(A) Whole tree; (B) leaves (C) flowers and (D) fruits [58]	13
Figure 2.4	Chemical structure of 12, 15-cis-squamostatin-A [81]	16
Figure 2.5	Bak/Bax-lipid pore model. Illustration is adapted from Chipuk <i>et al.</i> , 2006 [200]	32
Figure 2.6	Illustration for caspase-catalyzed features of apoptosis [206]	34
Figure 2.7	Potential ACGs inhibitory mechanism of action	42
Figure 2.8	Molecular Docking Steps. (A) Protein files (.pdb) were retrieved from the pdb data bank (https://www.rcsb.org/); (B) Ligands that were retrieved from databases such as PubChem (https://pubchem.ncbi.nlm.nih.gov) and ChemSpider (http://www.chemspider.com) were docked into the protein active sites using AutoDock 4.2 software; (C) Docked complex after the docking process	48
Figure 3.1	The flow of the present study; (1) Optimization of UAE and ME extracts, (2) Annonaceous identification using HPLC, (3-6) Biological assays, (7) <i>In silico</i> analysis through Molecular docking and MD simulation	50
Figure 4.1	Scatterplot of predicted and observed (actual) value of antioxidant effect	73
Figure 4.2	Response surface plot showing the combine effect of time (A) and amplitude (B) on the antioxidant effect	76
Figure 4.3	Response surface plot showing the combine effect of time (A) and ratio* (C) on the antioxidant effect. *(10%=1:10 and so on)	77
Figure 4.4	Response surface plot showing the combine effect of amplitude (B) and ratio* (C) on the antioxidant effect. *(10%=1:10 and so on)	77
Figure 4.5	Scatterplot of predicted and observed (actual) value of antioxidant effect	81

Figure 4.6	Response surface plot showing the combine effect of time (X) and solvent-to-material ratio* (Y) on the antioxidant effect. *(10%=1:10 and so on)	83
Figure 4.7	Response surface plot showing the combine effect of time (X) and speed (Z) on the antioxidant effect	84
Figure 4.8	Response surface plot showing the combine effect of ratio* (Y) and speed (Z) on the antioxidant effect. *(10%=1:10 and so on)	84
Figure 4.9	HLFa cancer cells viability after 48 h treatment with UAE and ME at different concentrations. Data are mean \pm SEM; n = 3 experiments. * $P < 0.05$, ** $P < 0.01$, # $P < 0.001$ against control	87
Figure 4.10	HLFa cancer cells viability after 48 h treatment with cisplatin at different concentrations. Data are mean \pm SEM; n = 3 experiments. * $P < 0.01$, ** $P < 0.001$ against control	87
Figure 4.11	HSF1184 normal cells viability after 48 h treatment with UAE and ME at different concentrations. Data are mean \pm SEM; n = 3 experiments. Data shown have no significant differences with the control	88
Figure 4.12	NO concentration in HLFa cancer cells after 48 h treatment with UAE and ME at different concentrations. Data are mean \pm SEM; n = 3 experiments. * $P < 0.1$ against control (0 $\mu\text{g/mL}$)	91
Figure 4.13	Caspase 3/7 activity in HLFa cells after treatment with the extracts for 48 h. Data are mean \pm SEM; n = 3 experiments. ** $P < 0.01$ against control	92
Figure 4.14	<i>BAX</i> and <i>BCL2</i> mRNA gene expressions in HLFa cells after 48 h treatment with 50 $\mu\text{g/mL}$ of UAE and ME. Data are mean \pm SEM; n = 3 experiments. * $P < 0.01$, ** $P < 0.05$, *** $P < 0.001$ against control. (a) $P < 0.01$, (b) $P < 0.05$	94
Figure 4.15	Caspase-3 activation via the intrinsic and extrinsic apoptotic pathways	95
Figure 4.16	<i>BAX</i> , <i>BCL2</i> and <i>HMGB1</i> mRNA gene expressions in HLFa cells after 48 h treatment with 10 nM of cisplatin. Data are mean \pm SEM; n = 3 experiments. * $P < 0.01$, ** $P < 0.05$ against control	96
Figure 4.17	<i>HMGB1</i> mRNA gene expressions in HLFa cells after 48 h treatment with 50 $\mu\text{g/mL}$ of UAE and ME. Data are mean \pm SEM; n = 3 experiments. * $P < 0.01$ against control. (a) $P < 0.01$	97
Figure 4.18	Binding interactions of anonaine and coreximine in Bcl-2. (a) Representation illustration of anonaine interactions in	

	the hydrophobic pocket of Bcl-2. (b) Hydrophobic interactions of anonaine with residues in the hydrophobic pocket of Bcl-2. (c) Representation illustration of coreximine interactions in the hydrophobic pocket of Bcl-2. (d) Hydrophobic interactions of coreximine with residues in the hydrophobic pocket of Bcl-2	100
Figure 4.19	Binding interactions of anonaine and coreximine in Bcl-w. (a) Representation illustration of anonaine interactions in the hydrophobic pocket of Bcl-w. (b) Hydrophobic interactions of anonaine with residues in the hydrophobic pocket of Bcl-w. (c) Representation illustration of coreximine interactions in the hydrophobic pocket of Bcl-w. (d) Hydrophobic interactions of coreximine with residues in the hydrophobic pocket of Bcl-w	103
Figure 4.20	Binding interactions of anonaine and coreximine in Mcl-1. (a) Representation illustration of anonaine interactions in the hydrophobic pocket of Mcl-1. (b) Hydrophobic interactions of anonaine with residues in the hydrophobic pocket of Mcl-1. (c) Representation illustration of coreximine interactions in the hydrophobic pocket of Mcl-1. (d) Hydrophobic interactions of coreximine with residues in the hydrophobic pocket of Mcl-1	105
Figure 4.21	The superimposition of the generated Bcl-2 protein (blue) and 4MAN protein model template (red) shows high similarity in terms of the protein structure	107
Figure 4.22	(a) The structure of Bcl-2/obatoclax protein complex. Red colour structure is the ligand (obatoclax) and the close-up view of the ligand. (b) The structure of Bcl-2/anonaine protein complex. Blue colour structure is the ligand (anonaine) and the close-up view of the ligand	108
Figure 4.23	MD analysis of protein-ligand complex (anonaine and obatoclax) trajectories generated by GROMACS at 310.15 K for 20 ns (a) RMSD; (b) RMSF (c) Rg; (d) SASA. (Anonaine is shown in blue, obatoclax in red)	109
Figure 4.24	HPLC chromatograms of ACGs compound representative illustration; (i) Anonaine, (ii) UAE and (iii) ME; (A) Anonaine peak	116

LIST OF ABBREVIATIONS

ACGs	-	Annonaceous acetogenins
ADC	-	Adenocarcinoma
AIF	-	Apoptosis inducing factor
ANT	-	Adenine nucleotide transporter
APAF-1	-	Apoptotic inducing factor-1
C.V.	-	Coefficient of variation
CCD	-	Central composite design
CICD	-	Caspase-independent cell death
COPD	-	Chronic obstructive pulmonary disease
DAMP	-	Damage-associated molecular pattern
DISC	-	Death-inducing signal complex
DMEM	-	Dulbecco's Modified Essential Medium
DMSO	-	Dimethyl sulfoxide
DPPH	-	2, 2, diphenyl-2-picryl-hydrazyl
EndoG	-	Endonuclease G
eNOS	-	Endothelial NOS
FAD	-	Flavin adenine dinucleotide
FBS	-	Fetal bovine serum
FDA	-	Food and Drug Administration
FMN	-	Flavin mononucleotide
GAPDH	-	Glyceraldehyde-3-phosphate dehydrogenase
HMGB1	-	High Mobility Group Box 1 protein
HPLC	-	High Performance Liquid Chromatography
HTS	-	High-throughput screening
IAPs	-	Inhibitor of apoptosis protein
IFN- γ	-	Interferon- γ
IL-1 α	-	Interleukin-1 α
IL-1 β	-	Interleukin-1 β
IMS	-	Inter membrane space
iNOS	-	Inducible NOS

IUPAC	-	International Union of Pure and Applied Chemistry
LDH	-	Lactate dehydrogenase
LGA	-	Lamarckian Genetic Algorithm
LINCS	-	Linear Constraint
LPS	-	Lipopolysaccharide
MAE	-	Microwave-assisted extraction
ME	-	Maceration extraction
MD	-	Molecular Dynamics
MOMP	-	Mitochondrial outer membrane permeabilization
mPTP	-	Mitochondrial permeability transition pore
MTT	-	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NCR	-	National Cancer Registry
NMR	-	Nuclear magnetic resonance
nNOS	-	Neuronal NOS
NO	-	Nitric oxide
NOS	-	Nitric oxide synthase
NOS1	-	nNOS
NOS2	-	iNOS
NOS3	-	eNOS
NSCLC	-	Non-small cell lung cancer
OMM	-	Outer mitochondrial membrane
PAMP	-	Pathogen-associated molecular pattern
PARP	-	Poly (ADP-ribose) polymerase
PBC	-	Periodic boundary condition
PBS	-	Phosphate buffer saline
PDB	-	Protein Data Bank
PME	-	Particle Mesh Ewald
PS	-	Phosphatidylserine
RAGE	-	Receptor for advanced glycation end products
RAMPAGE	-	Ramachandran Plot Assessment
R _g	-	Radius of gyration
RMSD	-	Root mean square deviation
RMSF	-	Root mean square

ROS	-	Reactive oxygen species
RSM	-	Response Surface Methodology
RT-PCR	-	Real-time polymerase chain reaction
SASA	-	Solvent Accessible Surface Area
SAVES	-	Structural Analysis and Verification Server
SCC	-	Squamous cell carcinoma
SCLC	-	Small cell lung cancer
SEM	-	Standard error means
SMI	-	Small molecule inhibitor
SPC	-	Single Point Charge
SPE	-	Superficial fluid extraction
TNF- α	-	Tumor necrosis factor- α
UAE	-	Ultrasonic-assisted extraction
VEGF	-	Vascular endothelial growth factor
WHO	-	World Health Organization

LIST OF SYMBOLS

%	-	percent
°C	-	degree celcius
g	-	gram
g	-	relative centrifugal force
g/mol	-	gram per mol
h	-	hour
L	-	liter
mg/mL	-	milligram per mililiter
min	-	minute
mL	-	mili liter
mM	-	mili molar
nM	-	nano molar
rpm	-	revolution per minute
µg/mL	-	microgram per mililiter
µL	-	micro liter

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
Appendix A	HLFa Cells 5×and 10× Resolution	147
Appendix B	HSF1184 10× Resolution	148
Appendix C	IUPAC Names of the Compounds	149
Appendix D	Crystal Structure of the Ligands	151

CHAPTER 1

INTRODUCTION

1.1 Research Background

Lung cancer is a major healthcare problem in Malaysia and across the globe. This disease is one of the most prominent cause of cancer-related mortality; about 40% cases was reported in developing countries like Malaysia [1]. In 2012, approximately 1.8 million cases were recorded with more than 80% of the proportion succumb to this fatal disorder. In Malaysia, lung cancer is the leading cause of cancer-death among males population and the fifth cause among females with about 13.8% and 3.8% cases respectively [2,3]. Lung cancer is among the cancers that constitute more than 58% of the projected global cancer burden [4]. In 2030, the leading cancer locations include lung, prostate and melanoma for men, and breast, thyroid and uterine for women [5]. There are three major types of lung cancers: non-small cell lung cancer (NSCLC), small cell lung cancer (SCLC) and lung carcinoid tumor. Despite numerous efforts – ranging from cancer immunotherapy to natural-derived anticancer agents – have been taken in the recent years in pursuit to puzzle out the way to combat this incurable malady, the ‘war’ against cancers is getting more perplexing and gripping. The hard-truth reality is that some part of cancer mechanism – up to the present time – is obscure; no individual in scientific community could authoritatively explain the ‘actual’ causes of cancers and how to prevent it let alone to cure it. However, in the recent years, natural-derived products such as plant-based products have gained wide interest. Reflection of this circumstances can be seen from the report of World Health Organization (WHO) which stated that more than 80% of the global population relies on plant-based traditional medicine for their primary healthcare; this has raised up the possibility of plant-derived substances to be the potential remedy everyone is looking for [6,7]. For the record, there are more than 3000 plant species being applied in the treatments of cancer-like diseases including swelling, abscesses, calluses, warts and polyps prompting the birth of about 60% of the currently available anticancer drugs

[8,9]. The application of plant-derived substances as anticancer drugs such Taxol is beneficial to consumers as these substances exert fewer adverse effects, and they are cost-effective alternative [10,11].

As one of the most arguably potential source of anticancer agent, *Annona muricata* Linn, has been the subject of extensive researches for a number of years. Groups of phytochemical compounds including flavonoids, alkaloids and annonaceous acetogenins (ACGs) and several types of extracts from *A. muricata* L. have been the candidates for anticancer drugs – widely manipulated to determine their characteristics and mode of actions. They have been reported to eliminate cancer cells and disrupt cancer cells development through numerous numbers of mechanisms in cell culture studies as well as in animal studies. For instance, *A. muricata* L. leaves ethyl acetate extract inhibited proliferation of lung cancer A549 cells via mitochondrial-mediated pathway with the involvement of NF- κ B, a cell signaling regulator [12]. Likewise, commercialized leaves water extract of *A. muricata* L. minimized pancreatic metastasis and tumorigenicity of implanted pancreatic tumors [13]. A synergistic study of major bioactive phytochemicals (flavonoids and ACGs) from *A. muricata* L. leaves showed that these substances inhibited proliferation activity of prostate cancer cells and the cells' clonogenic capacity [14]. On the other hand, (-)-anonaine, an alkaloid, demonstrated dose-dependent antiproliferation, DNA damage and antimigration in human lung carcinoma (H1299) cells [15]. Furthermore, it also stimulated cell cycle arrest in the similar cell line [15]. These indicate that more studies involving lung cancer such as NSCLC are welcomed, as there are room for manipulation and improvement.

NSCLC is the most common type of lung cancers accounting for about 80-85%, including adenocarcinoma (ADC) and squamous cell carcinoma (SCC) [16]. NSCLC is a type of aggressive cancer in which cancer cells form in the lung tissues. Risk factors for NSCLC include smoking, secondary smoking, radiation, environmental factors, genetic (family history) and HIV. Despite diverse treatment methods such as surgery, chemotherapy, radiation and targeted therapies, the overall 5-year survival rate for NSCLC is accounted for only 18.2% [17]. The high mortality rates for NSCLC are partially due to the lack of effective prognostic factors such as

biomarkers. NSCLC is associated with several symptoms including shortness of breath, chest pain, lost of appetite and tiredness. To date, there is no cure or preventive treatment for NSCLC; however, several factors – including the stage of cancer, type of mutations in cancer and patients' health – may affect the chance of recovery and treatment. A meta-analysis study involving 2651 NSCLC patients has detected the higher High Mobility Group Box 1 (HMGB1) expression in NSCLC cells than that in the healthy cells [16]. Thus, identifying the expression and behavior of novel prognostic factors like HMGB1, as biomarkers, may be a clinically useful tool for early detection of NSCLC and for potential cancer therapeutic target.

HMGB1, an extracellular damage-associated molecular pattern (DAMP), is a vital regulator of cell death and cell survival. It possesses several important functions in many diseases especially inflammatory diseases and cancers [18–20]. Its overexpression is linked to the hallmarks of cancer including angiogenesis capacity, apoptosis evasion, insensitivity to growth inhibitors, tissue invasion and metastasis, inflammation, self-sufficiency in growth signals and unlimited replicative potential [21]. The expression of vascular endothelial growth factor (VEGF), one of the main angiogenic factors, in ovarian carcinoma (SKOV3) cells was reduced with the interference of HMGB1 [22]. During cell death mechanisms, including apoptosis and necrosis, HMGB1 release was observed in pancreatic cancer (Panc-1) and cervical cancer (HeLa) cell lines [23]. Overexpression of HMGB1 in colorectal cancers had contributed significantly in tumor progression and tumors' ability to metastasize [24]. While in NSCLC, HMGB1 enhanced the increase in cancer cells migration ability through the activation of TLR4/NF- κ B signaling – inducing metastasis [25]. It was revealed that the protein level of HMGB1 in patients with NSCLC of TNM Stages III-IV was significantly higher as compared to TNM Stages I-II, indicating that HMGB1 plays a crucial role in the progression of NSCLC [26]. These also demonstrated that HMGB1 expression could be regarded as important prognostic biomarker in cancers development specifically for NSCLC diagnosis, thus making it particularly interesting as potential therapeutic targets for cancer drug discovery.

Many studies have examined the relationship HMGB1 expression and NSCLC. Even though most of the results remain unsettled, some study has shown significance

correlation [26]. Hence, the present study focuses to investigate the mechanism involved in the effect of *A. muricata* L. leaves extract on the expression level of HMGB1 and several other biomarkers involved in regulating apoptosis in NSCLC. This study would provide additional information to the present anticancer knowledge and future research.

1.2 Problem Statement

A. muricata L. has long been postulated to possess anticancer properties against various types of cancer. The plant bioactive constituents such as ACGs have been identified to be vital contributors to anticancer effects, leading to its characterization and isolation. Even though the researches involving ACGs have been carried out for few decades, these compounds have never been commercialized or developed as anticancer drug for clinical test. These circumstances prompt a wave of questions debating the idea with hope that the premise relating to the anticancer effect of *A. muricata* L. is not just another scientific blunder.

From a local perspective, there have been a growing number of products based on *A. muricata* L. such as health supplement capsule, pill, juice and ice cream. The common conception among the public in Malaysia is that by consuming these products, their detrimental health could be ameliorated thus preventing them from succumbing to cancer. Therefore, there is huge responsibility lies within scientific community to clear the air to avoid any further misunderstanding that is not beneficial for the society in large.

In hope that *A. muricata* L. might display significant effect against lung cancer cells at the end of this study, the current research adds fresh and deepening information to the current anticancer knowledge. Even though NSCLC is the leading contributor of cancer mortality, there is lack of *A. muricata* L. anticancer study on this type of lung cancer cell. It was reported that the high 5-year survival rate for NSCLC (18.2%) are due to the inadequacy of functional prognostic biomarkers and proper treatment [27]. This shortfall is due to difficulties in diagnosing NSCLC; patients are diagnosed at

advanced stages with distant and local metastases. The discovery of HMGB1 as potential effective markers for early diagnosis of NSCLC has allowed this biomarker to be the subject of thorough experimentations. *In silico* molecular docking and molecular dynamic (MD) simulation were also applied to investigate the intriguing possibility of *A. muricata* Linn's bioactive compounds namely annonaceous acetogenins (ACGs) to possess Bcl-2 antiapoptotic inhibitory properties. Through molecular docking approach, the affinity of the compounds of interest towards Bcl-2 antiapoptotic proteins would be determined and possible interaction between amino acids and compounds would be crucial information for development of anticancer drug using ACGs.

Thus, the understanding upon the underlying mechanism triggered by potential anticancer agent such as *A. muricata* L. extract in NSCLC would certainly ignite a slight hope in this long-running battle. This study is the first to evaluate the correlation between *A. muricata* L. effect and release of HMGB1 during cell death mechanism in NSCLC.

1.3 Research Objectives

The main objective of this study was to investigate the mechanistic effect induced by *A. muricata* L. optimized leaves extracts in NSCLC. The objective is further separated into several other objectives as listed below:

- (a) To optimize the extraction parameters for high yield of *A. muricata* L. extracts
- (b) To determine the antioxidant activity of the optimized extracts
- (c) To investigate the effect of the optimized extracts on HMGB1 gene expression in NSCLC cells
- (d) To characterize the *in silico* molecular interactions of the plant's bioactive compounds

1.4 Scope of the Study

The scope of this research are as listed below:

1. Antioxidant-response optimization of ultrasonic-assisted extraction parameters (time, amplitude, ratio) and maceration extraction parameters (time, ratio, speed) of *A. muricata* L. leaves by using response surface methodology.
2. Determination of the antioxidant activity of the extracts by using 2, 2-diphenyl-2-picryl-hydrazyl (DPPH) free radical scavenging assay.
3. Cytotoxicity and anti-proliferation activities determination of the extracts by using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) against human lung epidermoid carcinoma (HLFa) cells and human fibroblast cells (HSF1184).
4. Nitric oxide (NO) determination by using nitrite concentration assay.
5. Determination of apoptotic activity by using apoptosis detection assay.
6. Quantification of gene expression of *BCL2*, *BAX* and *HMGB1* by using real-time polymerase chain reaction (RT-PCR).
7. Characterization of binding interactions involving acetogenins of *A. muricata* L. and Bcl-2 antiapoptotic proteins, by using *in silico* molecular docking software, AutoDock 4.2 and molecular dynamics (MD) simulation.
8. Screening of bioactive compounds (ACGs) in the extracts by using High Performance Liquid Chromatography (HPLC).

1.5 Significances of the Study

The current study presents several novel contributions as well as additional knowledge in the field of *A. muricata* L anticancer research. The present study emphasizes the effect of extraction method on antioxidant effect response of *A. muricata* L and their correlation to antioxidant activities. This study is among a few

studies, which have been done to optimize the ultrasonic-assisted and maceration extractions, and comparison of anticancer effects between both optimized extracts provide new information to this particular field of research. This study also highlights the potential of *A. muricata* L. extracts to be further developed as anticancer agent against lung cancer especially NSCLC. Furthermore, as the effect of plant extracts on HMGB1 has never been explored before, this study would be the first to offer novel report upon *A. muricata* L. mechanistic activity against HMGB1 in NSCLC, in addition to other regulators such as Bcl-2 and Bax that could be a steppingstone to future anticancer studies as well as provides insight on the novel understanding of therapeutic potential of HMGB1 as prognostic biomarker.

1.6 Thesis Structure and Organization

This thesis comprises five chapters. The first chapter serves as a backbone of the whole thesis. It covers the underlying premise that led to the initiation of this study. The idea was presented in summarized language, of which consists of research background, problem statement, objective, scope and significances of the study.

Chapter 2 gives thorough reviews and discussions on newest literatures available. In addition, it discusses the arguable potential of *A. muricata* L. as candidate for anticancer agent. It also reviews the prevalence of lung cancer.

Chapter 3 elucidates the selected experimental-methodologies, of which were conducted to present the evidences for the hypothesis of the present study.

Chapter 4 puts forward the observed results together with the discussion of the findings that covers the anti-oxidative characteristics of *A. muricata* L. extract, the gene expression analysis of *BCL2*, *BAX* and *HMGB1* in lung cancer cells and the *in silico* virtual screening analysis.

Chapter 5 concludes the current study and summarizes the outcome. It also recommends suggestions for future researches.

REFERENCES

1. Siegel, R., Naishadham, D., Cancer statistics, 2013. *CA A Cancer J.* 2013, 63, 11–30.
2. Chye, G.L.C., Rampal, S., Yahaya, H., Cancer Incidence in Peninsular Malaysia 2003-2005. *Natl. Cancer Regist.* 2008, 53–57.
3. Department of Statistics, M., Population distribution and basic demographic Characteristic Report 2010, vol. 2010, 2011.
4. Bray, F., Jemal, A., Grey, N., Ferlay, J., Forman, D., Global cancer transitions according to the Human Development Index (2008-2030): a population-based study. *Lancet Oncol.* 2012, 13, 790–801.
5. Rahib, L., Smith, B.D., Aizenberg, R., Rosenzweig, A.B., et al., Projecting cancer incidence and deaths to 2030: The unexpected burden of thyroid, liver, and pancreas cancers in the united states. *Cancer Res.* 2014, 74, 2913–2921.
6. Surh, Y.-J., Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer* 2003, 3, 768–780.
7. Farnsworth, N.R., Akerele, O., Bingel, A.S., Soejarto, D.D., Guo, Z., Medicinal plant in therapy. *Bull. World Heal. Organ.* 1985, 63, 965–981.
8. Graham, J., Quinn, M., Plants used against cancer—an extension of the work of Jonathan Hartwell. *J. ...* 2000.
9. Cragg, G.M., Newman, D.J., Plants as a source of anti-cancer agents. *J. Ethnopharmacol.* 2005, 100, 72–79.
10. Veeresham, C., Natural products derived from plants as a source of drugs. *J. Adv. Pharm. Technol. Res.* 2014.
11. Newton, K., Matsumoto, M.L., Wertz, I.E., Kirkpatrick, D.S., et al., Ubiquitin chain editing revealed by polyubiquitin linkage-specific antibodies. *Cell* 2008, 134, 668–78.
12. Zorofchian Moghadamtousi, S., Abdul Kadir, H., Paydar, M., Rouhollahi, E., Karimian, H., *Annona muricata* leaves induced apoptosis in A549 cells through mitochondrial-mediated pathway and involvement of NF-kappaB. *BMC Complement. Altern. Med.* 2014, 14, 299.

13. Torres, M.P., Rachagani, S., Purohit, V., Pandey, P., et al., Graviola: A novel promising natural-derived drug that inhibits tumorigenicity and metastasis of pancreatic cancer cells in vitro and in vivo through altering cell metabolism. *Cancer Lett.* 2012, 323, 29–40.
14. Yang, C., Gundala, S.R., Mukkavilli, R., Vangala, S., et al., Synergistic interactions among flavonoids and acetogenins in Graviola (*Annona muricata*) leaves confer protection against prostate cancer. *Carcinogenesis* 2015, 36, 656–665.
15. Chen, B.-H., Chang, H.-W., Huang, H.-M., Chong, I.-W., et al., (-)-Anonaine induces DNA damage and inhibits growth and migration of human lung carcinoma h1299 cells. *J. Agric. Food Chem.* 2011, 59, 2284–2290.
16. Feng, A., Tu, Z., Yin, B., The effect of HMGB1 on the clinicopathological and prognostic features of non-small cell lung cancer. *Oncotarget* 2016, 7, 20507–19.
17. DeSantis, C.E., Lin, C.C., Mariotto, A.B., Siegel, R.L., et al., Cancer facts and Figures 2013 Annual Report. *CA Cancer J Clin* 2014, 64, 25–271.
18. Kang, R., Zhang, Q., Zeh, H.J., Lotze, M.T., Tang, D., HMGB1 in cancer: Good, bad, or both? *Clin. Cancer Res.* 2013, 19, 4046–4057.
19. Tang, D., Kang, R., Zeh, H.J., Lotze, M.T., High-mobility group box 1 and cancer. *Biochim. Biophys. Acta* 2010, 1799, 131–40.
20. Andersson, U., Tracey, K.J., HMGB1 is a therapeutic target for sterile inflammation and infection. *Annu Rev Immunol* 2011, 29, 139–162.
21. Pilzweger, C., Holdenrieder, S., Circulating HMGB1 and RAGE as Clinical Biomarkers in Malignant and Autoimmune Diseases. *Diagnostics* 2015, 5, 219–253.
22. Zhou, L., Shi, L., Xiao, Y., Changes of HMGB1 expression on angiogenesis of ovarian cancer and its mechanism. *J. Biol. Regul. Homeost. Agents* 2016, 30, 233–238.
23. Bell, C.W., Jiang, W., Reich Iii, C.F., Pisetsky, D.S., The extracellular release of HMGB1 during apoptotic cell death. *Am J Physiol Cell Physiol* 2006, 291, 1318–1325.
24. Süren, D., Yıldırım, M., Demirpençe, Ö., Kaya, V., et al., The role of high mobility group box 1 (HMGB1) in colorectal cancer. *Med. Sci. Monit.* 2014, 20, 530–537.

25. Zhu, J., Luo, J., Li, Y., Jia, M., et al., HMGB1 induces human non-small cell lung cancer cell motility by activating integrin $\alpha 5 \beta 1$ /FAK through TLR4/NF- κ B signaling pathway. *Biochem. Biophys. Res. Commun.* 2016, 480, 522–527.
26. Xia, Q., Xu, J., Chen, H., Gao, Y., et al., Association between an elevated level of HMGB1 and non-small-cell lung cancer: a meta-analysis and literature review. *Onco. Targets. Ther.* 2016, 9, 3917–3923.
27. Miller, K.D., Siegel, R.L., Lin, C.C., Mariotto, A.B., et al., Cancer treatment and survivorship statistics, 2016. *CA. Cancer J. Clin.* 2016, 66, 271–89.
28. Surh, Y., Cancer chemoprevention with dietary phytochemicals. *Nat. Rev. Cancer* 2003.
29. Lako, J., Trenerry, V.C., Wahlqvist, M., Wattanapenpaiboon, N., et al., Phytochemical flavonols, carotenoids and the antioxidant properties of a wide selection of Fijian fruit, vegetables and other readily available foods. *Food Chem.* 2007, 101, 1727–1741.
30. Russo, M., Spagnuolo, C., Tedesco, I., GL, R., Phytochemicals in Cancer Prevention and Therapy: Truth or Dare? *Toxins (Basel)*. 2010, 2, 517–551.
31. Jansen, R., Robinson, D., Stolzenberg-Solomon, R., Bamlet, W., et al., Fruit and vegetable consumption is inversely associated with having pancreatic cancer. *Cancer Causes Control* 2011, 22, 1613–1625.
32. Matsuoka, H., Furusawa, M., Tomoda, H., Seo, Y., Difference in cytotoxicity of paclitaxel against neoplastic and normal cells. *Anticancer Res.* 14, 163–7.
33. Schulz, W., Molecular biology of human cancers, 2005.
34. Wani, M.C., Taylor, H.L., Wall, M.E., Coggon, P., Mcphail, A.T., Plant Antitumor Agents.VI.The Isolation and Structure of Taxol, a Novel Antileukemic and Antitumor Agent from *Taxus brevifolia*2. *J. Am. Chem. Soc.* 1971, 93, 2325–2327.
35. Kingston, D.G.I., Recent Advances in the Chemistry of Taxol 1,2. *J. Nat. Prod.* 2000, 63, 726–734.
36. Kingston, D.G.I., Taxol, a molecule for all seasons. *Chem. Commun.* 2001, 867–880.
37. Malik, S., Cusidó, R.M., Mirjalili, M.H., Moyano, E., et al., Production of the anticancer drug taxol in *Taxus baccata* suspension cultures: A review. *Process Biochem.* 2011, 46, 23–34.

38. Xiao, H., Verdier-Pinard, P., Fernandez-Fuentes, N., Burd, B., et al., Insights into the mechanism of microtubule stabilization by Taxol. *Proc. Natl. Acad. Sci. U. S. A.* 2006, 103, 10166–73.
39. Hait, W.N., Rubin, E., Alli, E., Goodin, S., Tubulin Targeting Agents. *Update Cancer Ther.* 2007, 2, 1–18.
40. Prota, A.E., Bargsten, K., Zurwerra, D., Field, J.J., et al., Molecular mechanism of action of microtubule-stabilizing anticancer agents. *Science* 2013, 339, 587–90.
41. Bharadwaj, R., Yu, H., The spindle checkpoint, aneuploidy, and cancer. *Oncogene* 2004, 23, 2016–2027.
42. Ganguly, A., Yang, H., Cabral, F., Paclitaxel-dependent cell lines reveal a novel drug activity. *Mol. Cancer Ther.* 2010, 9, 2914–23.
43. Priyadarshini, K., U, K.A., Paclitaxel Against Cancer : A Short Review. *Med. Chem. (Los. Angeles)*. 2012, 2, 139–141.
44. George, J., Banik, N.L., Ray, S.K., Combination of taxol and Bcl-2 siRNA induces apoptosis in human glioblastoma cells and inhibits invasion, angiogenesis and tumour growth. *J. Cell. Mol. Med.* 2009, 13, 4205–4218.
45. Maráz, A., Furák, J., Pálföldi, R., Eller, J., et al., Roles of BCL-2 and MDR1 expression in the efficacy of paclitaxel-based lung cancer chemoradiation. *Anticancer Res.* 2011, 31, 1431–1436.
46. Sun, T.M., Du, J.Z., Yao, Y.D., Mao, C.Q., et al., Simultaneous delivery of siRNA and paclitaxel via a “two-in-one” micelleplex promotes synergistic tumor suppression. *ACS Nano* 2011, 5, 1483–1494.
47. Korbakis, D., Scorilas, A., Quantitative expression analysis of the apoptosis-related genes BCL2, BAX and BCL2L12 in gastric adenocarcinoma cells following treatment with the anticancer drugs cisplatin, etoposide and taxol. *Tumor Biol.* 2012, 33, 865–875.
48. Shajahan, A.N., Dobbin, Z.C., Hickman, F.E., Dakshanamurthy, S., Clarke, R., Tyrosine-phosphorylated caveolin-1 (Tyr-14) increases sensitivity to paclitaxel by inhibiting BCL2 and BCLxL proteins via c-Jun N-terminal Kinase (JNK). *J. Biol. Chem.* 2012, 287, 17682–17692.

49. Morales-Cano, D., Calviño, E., Rubio, V., Herráez, A., et al., Apoptosis induced by paclitaxel via Bcl-2, Bax and caspases 3 and 9 activation in NB4 human leukaemia cells is not modulated by ERK inhibition. *Exp. Toxicol. Pathol.* 2013, 65, 1101–1108.
50. Zhou, M., Liu, Z., Zhao, Y., Ding, Y., et al., MicroRNA-125b confers the resistance of breast cancer cells to paclitaxel through suppression of pro-apoptotic Bcl-2 antagonist killer 1 (Bak1) expression. *J. Biol. Chem.* 2010, 285, 21496–21507.
51. Suffness, M., Taxol: From Discovery to Therapeutic Use. *Annu. Rep. Med. Chem.* 1993, 28, 305–314.
52. Gordaliza, M., Natural products as leads to anticancer drugs. *Clin. Transl. Oncol.* 2007, 9, 767–776.
53. Douglas Kinghorn, a, Pharmacognosy in the 21st century. *J. Pharm. Pharmacol.* 2001, 53, 135–48.
54. Demain, A.L., Vaishnav, P., Natural products for cancer chemotherapy. *Microb. Biotechnol.* 2011, 4, 687–699.
55. Hanahan, D., Weinberg, R.A., Hallmarks of Cancer: The Next Generation. *Cell* 2011, 144, 646–674.
56. Wiart, C., Medicinal plants of Southeast Asia., 2002.
57. Ezirim AU, Okochi VI, James AB, Adebeshi OA, Ogunnowo SO, O.O., Induction of Apoptosis in Myelogenous Leukemic K562 Cells by Ethanolic Leaf Extract of *Annona Muricata* L. *Glob. J. Res. Med. Plants Indig. Med.* 2013, 2, 142–151.
58. Moghadamtousi, S.Z., Fadaeinasab, M., Nikzad, S., Mohan, G., et al., *Annona muricata* (Annonaceae): A review of its traditional uses, isolated acetogenins and biological activities. *Int. J. Mol. Sci.* 2015, 16, 15625–15658.
59. Wiart, C., Medicinal plants of Asia and the Pacific, 2006.
60. Taylor, L., Press, S., Technical Data Report for Graviola (*Annona muricata*), Austin, TX, 2005.
61. Arroyo, J., Prashad, M., Vásquez, Y., Li, E., Tomás, G., Actividad citotóxica in vitro de la mezcla de *Annona muricata* y *Krameria lappacea* sobre células cancerosas de glándula mamaria, pulmón y sistema nervioso central. *Med Exp Salud Publica* 2005, 22, 247–253.

62. Astirin, O.P., Artanti, A.N., Fitria, M.S., Perwitasari, E.A., Prayitno, A., *Annona muricata* Linn Leaf Induce Apoptosis in Cancer Cause Virus. *J. Cancer Ther.* 2013, 4, 1244–1250.
63. Gavamukulya, Y., Abou-Ellella, F., Wamunyokoli, F., AEl-Shemy, H., Phytochemical screening, anti-oxidant activity and in vitro anticancer potential of ethanolic and water leaves extracts of *Annona muricata* (Graviola). *Asian Pac. J. Trop. Med.* 2014, 7, S355–S363.
64. Cijo George, V., Naveen Kumar, D.R., Rajkumar, V., Suresh, P.K., Ashok Kumar, R., Quantitative assessment of the relative antineoplastic potential of the n-butanolic leaf extract of *Annona Muricata* Linn. in normal and immortalized human cell lines. *Asian Pacific J. Cancer Prev.* 2012, 13, 699–704.
65. Jaramillo, M.C., Arango, G.J., González, M.C., Robledo, S.M., Velez, I.D., Cytotoxicity and antileishmanial activity of *Annona muricata* pericarp. *Fitoterapia* 2000, 71, 183–186.
66. Zorofchian Moghadamtousi, S., Karimian, H., Rouhollahi, E., Paydar, M., et al., *Annona muricata* leaves induce G1 cell cycle arrest and apoptosis through mitochondria-mediated pathway in human HCT-116 and HT-29 colon cancer cells. *J. Ethnopharmacol.* 2014, 156, 277–289.
67. Zorofchian Moghadamtousi, S., *Annona muricata* leaves induce G(1) cell cycle arrest and apoptosis through mitochondria-mediated pathway in human HCT-116 and HT-29 colon cancer cells. *J Ethnopharmacol* 2014, 156.
68. Asare, G. a., Afriyie, D., Ngala, R. a., Abutiate, H., et al., Antiproliferative Activity of Aqueous Leaf Extract of *Annona muricata* L. on the Prostate, BPH-1 Cells, and Some Target Genes. *Integr. Cancer Ther.* 2014, 14, 65–74.
69. Minari, J.B., Okeke, U., Chemopreventive effect of *Annona muricata* on DMBA-induced cell proliferation in the breast tissues of female albino mice. *Egypt. J. Med. Hum. Genet.* 2014, 15, 327–334.
70. Hamizah, S., Roslida, A.H., Fezah, O., Tan, K.L., et al., Chemopreventive potential of *Annona muricata* L leaves on chemically-induced skin papillomagenesis in mice. *Asian Pacific J. Cancer Prev.* 2012, 13, 2533–2539.
71. Hansra, D.M., Silva, O., Mehta, A., Ahn, E., Patient with Metastatic Breast Cancer Achieves Stable Disease for 5 Years on Graviola & Xeloda after Progressing on Multiple Lines of Therapy. *Adv. breast cancer Res.* 2014, 3, 84

72. Dai, Y., Hogan, S., Schmelz, E.M., Ju, Y.H., et al., Selective growth inhibition of human breast cancer cells by graviola fruit extract in vitro and in vivo involving downregulation of EGFR expression. *Nutr Cancer* 2011, 63, 795–801.
73. Syed Najmuddin, S.U.F., Romli, M.F., Hamid, M., Alitheen, N.B., Nik Abd Rahman, N.M.A., Anti-cancer effect of *Annona Muricata* Linn Leaves Crude Extract (AMCE) on breast cancer cell line. *BMC Complement. Altern. Med.* 2016, 16, 311.
74. Ko, Y.-M., Wu, T.-Y., Wu, Y.-C., Chang, F.-R., et al., Annonacin induces cell cycle-dependent growth arrest and apoptosis in estrogen receptor- α -related pathways in MCF-7 cells. *J. Ethnopharmacol.* 2011, 137, 1283–1290.
75. Leboeuf, M., Cavé, A., Bhaumik, P.K., Mukherjee, B., Mukherjee, R., The phytochemistry of the annonaceae. *Phytochemistry* 1980, 21, 2783–2813.
76. Matsushige, A., Matsunami, K., Kotake, Y., Otsuka, H., Ohta, S., Three new megastigmanes from the leaves of *Annona muricata*. *J. Nat. Med.* 2012, 66, 284–291.
77. Nawwar, M., Ayoub, N., Hussein, S., Hashim, A., et al., A flavonol triglycoside and investigation of the antioxidant and cell stimulating activities of *Annona muricata* Linn. *Arch. Pharm. Res.* 2012, 35, 761–767.
78. Jiménez, V.M., Gruschwitz, M., Schweiggert, R.M., Carle, R., Esquivel, P., Identification of phenolic compounds in soursop (*Annona muricata*) pulp by high-performance liquid chromatography with diode array and electrospray ionization mass spectrometric detection. *Food Res. Int.* 2014, 65, 42–46.
79. Rupprecht, J.K., Hui, Y.H., McLaughlin, J.L., Annonaceous acetogenins: A review. *J. Nat. Prod.* 1990.
80. McLaughlin, J.L., Paw paw and cancer: Annonaceous acetogenins from discovery to commercial products. *J. Nat. Prod.* 2008, 71, 1311–1321.
81. Chen, Y., Xu, S.S., Chen, J.W., Wang, Y., et al., Anti-tumor activity of *Annona squamosa* seeds extract containing annonaceous acetogenin compounds. *J. Ethnopharmacol.* 2012, 142, 462–466.
82. Diplock, a T., Charleux, J.L., Crozier-Willi, G., Kok, F.J., et al., Functional food science and defence against reactive oxidative species. *Br. J. Nutr.* 1998.

83. Valko, M., Leibfritz, D., Moncol, J., Cronin, M.T.D., et al., Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* 2007.
84. Liao, J.-C., Deng, J.-S., Chiu, C.-S., Huang, S.-S., et al., Chemical compositions, anti-inflammatory, antiproliferative and radical-scavenging activities of *Actinidia callosa* var. *ephippioides*. *Am. J. Chin. Med.* 2012.
85. Chen, W., Weng, Y.-M., Tseng, C.-Y., Antioxidative and antimutagenic activities of healthy herbal drinks from Chinese medicinal herbs. *Am. J. Chin. Med.* 2003.
86. George, V.C., Kumar, D.R.N., Suresh, P.K., Kumar, R.A., Antioxidant, DNA protective efficacy and HPLC analysis of *Annona muricata* (soursop) extracts. *J. Food Sci. Technol.* 2015.
87. Baskar, R., Rajeswari, V., Kumar, T.S., In vitro antioxidant studies in leaves of annona species. *Indian J. Exp. Biol.* 2007, 45, 480–485.
88. Vijayameena, C., Subhashini, G., Loganayagi, M., Ramesh, B., Phytochemical screening and assessment of antibacterial activity for the bioactive compounds in *Annona muricata*. *Int. J. Curr. Microbiol. Appl. Sci.* 2013.
89. Ettinger, D.S., Akerley, W., Borghaei, H., Chang, A.C., et al., Non-small cell lung cancer. *J. Natl. Compr. Canc. Netw.* 2012, 10, 1236–71.
90. Benjamin, C.L., Ullrich, S.E., Kripke, M.L., Ananthaswamy, H.N., p53 tumor suppressor gene: a critical molecular target for UV induction and prevention of skin cancer. *Photochem. Photobiol.* 84, 55–62.
91. Assessment., U.S.C.O. of T., Assessment of Technologies for Determining Cancer Risks From the Environment 1981.
92. De Maria, N., Manno, M., Villa, E., Sex hormones and liver cancer. *Mol. Cell. Endocrinol.* 2002, 193, 59–63.
93. Ho, E., Zinc deficiency, DNA damage and cancer risk. *J. Nutr. Biochem.* 2004, 15, 572–8.
94. Frezza, C., Pollard, P.J., Gottlieb, E., Inborn and acquired metabolic defects in cancer. *J. Mol. Med. (Berl)*. 2011, 89, 213–20.
95. Herr, H.W., Percivall Pott, the environment and cancer. *BJU Int.* 2011, 108, 479–81.
96. Yokota, J., Tumor progression and metastasis. *Carcinogenesis* 2000, 21, 497–503.

97. Hanahan, D., Weinberg, R.A., The hallmarks of cancer. *Cell* 2000, 100, 57–70.
98. Hanahan, D., Weinberg, R.A., Hallmarks of cancer: The next generation. *Cell* 2011, 144, 646–674.
99. Secretan, B., Straif, K., Baan, R., Grosse, Y., et al., A review of human carcinogens—Part E: tobacco, areca nut, alcohol, coal smoke, and salted fish. *Lancet Oncol.* 2009, 10, 1033–1034.
100. Doll, R., Gray, R., Hafner, B., Peto, R., Mortality in relation to smoking: 22 years' observations on female British doctors. *BMJ* 1980, 280, 967–971.
101. United States Department of Health and Human Services, The Health Consequences of Smoking—50 Years of Progress A Report of the Surgeon General. *A Rep. Surg. Gen.* 2014, 1081.
102. Taylor, R., Najafi, F., Dobson, A., Meta-analysis of studies of passive smoking and lung cancer: Effects of study type and continent. *Int. J. Epidemiol.* 2007, 36, 1048–1059.
103. Hackshaw, A.K., Law, M.R., Wald, N.J., The accumulated evidence on lung cancer and environmental tobacco smoke. *BMJ* 1997, 315, 980–988.
104. Centers for Disease Control and Prevention (US), U.S. Department of Health and Human Services, The Health Consequences of Involuntary Exposure to Tobacco Smoke: A Report of the Surgeon General, 2006.
105. Wald, N.J., Nanchahal, K., Thompson, S.G., Cuckle, H.S., Does breathing other people's tobacco smoke cause lung cancer? *Br. Med. J. (Clin. Res. Ed).* 1986, 293, 1217–22.
106. National Cancer Registry, Malaysia, M. of H., Malaysia Cancer Statistics-Data and Figure 2007. *Natl. Cancer Regist. Rep.* 2011, 42–43.
107. Parkin, D.M., Bray, F., Ferlay, J., Pisani, P., Global Cancer Statistics, 2002. *CA. Cancer J. Clin.* 2005, 55, 74–108.
108. National Cancer Registry, Malaysia cancer statistics data and figure Peninsular Malaysia. *Minist. Heal. Malaysia* 2006, 1–137.
109. WHO, Globocan 2012 - Home. *Globocan 2012* 2012.
110. Siegel, R., Ma, J., Zou, Z., Jemal, A., Cancer statistics, 2014. *CA. Cancer J. Clin.* 2014, 64, 9–29.
111. Zainal, A.O., Nor Saleha, I.T., National Cancer Registry Report, 2011.
112. Grivennikov, S.I., Greten, F.R., Karin, M., Immunity, Inflammation, and Cancer. *Cell* 2010, 140, 883–899.

113. O'Byrne, K.J., Dalgleish, A.G., Chronic immune activation and inflammation as the cause of malignancy. *Br. J. Cancer* 2001.
114. Schmidt, a, Weber, O.F., In memoriam of Rudolf virchow: a historical retrospective including aspects of inflammation, infection and neoplasia. *Contrib. Microbiol.* 2006.
115. Takahashi, H., Ogata, H., Nishigaki, R., Broide, D.H., Karin, M., Tobacco smoke promotes lung tumorigenesis by triggering IKKbeta- and JNK1-dependent inflammation. *Cancer Cell* 2010.
116. Walser, T., Cui, X., Yanagawa, J., Lee, J.M., et al., Smoking and lung cancer: the role of inflammation. *Proc. Am. Thorac. Soc.* 2008.
117. Adcock, I.M., Caramori, G., Barnes, P.J., Chronic obstructive pulmonary disease and lung cancer: new molecular insights. *Respiration.* 2011.
118. Sekine, Y., Hata, A., Koh, E., Hiroshima, K., Lung carcinogenesis from chronic obstructive pulmonary disease: Characteristics of lung cancer from COPD and contribution of signal transducers and lung stem cells in the inflammatory microenvironment. *Gen. Thorac. Cardiovasc. Surg.* 2014.
119. Nomura, A., Stemmermann, G.N., Chyou, P.H., Marcus, E.B., Buist, A.S., Prospective study of pulmonary function and lung cancer. *Am Rev Respir Dis* 1991.
120. Skillrud, D.M., Offord, K.P., Miller, R.D., Higher risk of lung cancer in chronic obstructive pulmonary disease. A prospective, matched, controlled study. *Ann. Intern. Med.* 1986.
121. Saetta, M., Turato, G., Maestrelli, P., Mapp, C.E., Fabbri, L.M., Cellular and structural bases of chronic obstructive pulmonary disease. *Am. J. Respir. Crit. Care Med.* 2001.
122. Xu, W.M., Liu, L.Z., Nitric oxide: from a mysterious labile factor to the molecule of the Nobel Prize. Recent progress in nitric oxide research. *Cell Res.* 1998, 8, 251–258.
123. Moncada, S., Palmer, R.M.J., Higgs, E.A., Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol. Rev.* 1991, 43, 109–142.
124. Furchgott, R.F., Zawadzki, J. V., The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* 1980, 288, 373–376.

125. Ignarro, L.J., Buga, G.M., Wood, K.S., Byrns, R.E., Chaudhuri, G., Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proc. Natl. Acad. Sci. U. S. A.* 1987, 84, 9265–9.
126. Katsuki, S., Arnold, W., Mittal, C., Murad, F., Stimulation of guanylate cyclase by sodium nitroprusside, nitroglycerin and nitric oxide in various tissue preparations and comparison to the effects of sodium azide and hydroxylamine. *J. Cyclic Nucleotide Res.* 1977, 3, 23–35.
127. Palmer, R.M., Ferrige, A.G., Moncada, S., Nitric oxide release accounts for the biological activity of endothelium-derived relaxing factor. *Nature* 1987, 327, 524–526.
128. XU, W., LIU, L.Z., LOIZIDOU, M., AHMED, M., CHARLES, I.G., The role of nitric oxide in cancer. *Cell Res.* 2002, 12, 311–320.
129. MacMicking, J., Xie, Q.W., Nathan, C., Nitric oxide and macrophage function. *Annu. Rev. Immunol.* 1997, 15, 323–50.
130. Garban, H.J., Bonavida, B., Nitric oxide sensitizes ovarian tumor cells to Fas-induced apoptosis. *Gynecol. Oncol.* 1999, 73, 257–264.
131. Xu, L., Xie, K., Fidler, I.J., Therapy of human ovarian cancer by transfection with the murine interferon beta gene: role of macrophage-inducible nitric oxide synthase. *Hum Gene Ther* 1998, 9, 2699–2708.
132. Xie, K.P., Huang, S.Y., Dong, Z.Y., Juang, S.H., et al., Transfection with the Inducible Nitric-Oxide Synthase Gene Suppresses Tumorigenicity and Abrogates Metastasis by K-1735 Murine Melanoma-Cells. *J. Exp. Med.* 1995, 181, 1333–1343.
133. Juang, S.H., Xie, K., Xu, L., Shi, Q., et al., Suppression of tumorigenicity and metastasis of human renal carcinoma cells by infection with retroviral vectors harboring the murine inducible nitric oxide synthase gene. *Hum Gene Ther* 1998, 9, 845–854.
134. Hibbs, J., Taintor, R., Vavrin, Z., Macrophage cytotoxicity: role for L-arginine deiminase and imino nitrogen oxidation to nitrite. *Science* 1987, 235, 473–476.
135. Reveneau, S., Arnould, L., Jolimoy, G., Hilpert, S., et al., Nitric oxide synthase in human breast cancer is associated with tumor grade, proliferation rate, and expression of progesterone receptors. *Lab Invest* 1999, 79, 1215–1225.
136. Thomsen, L.L., Lawton, F.G., Knowles, R.G., Beesley, J.E., et al., Nitric oxide synthase activity in human gynecological cancer. *Cancer Res* 1994, 54, 1352

137. Taysi, S., Uslu, C., Akcay, F., Sutbeyaz, M.Y., Malondialdehyde and nitric oxide levels in the plasma of patients with advanced laryngeal cancer. *Surg. Today* 2003, 33, 651–654.
138. Cobbs, C.S., Brenman, J.E., Aldape, K.D., Bredt, D.S., Israel, M.A., Expression of Nitric Oxide Synthase in Human Central Nervous System Tumors. *Cancer Res.* 1995, 55, 727–730.
139. Rosbe, K.W., Prazma, J., Petrusz, P., Mims, W., et al., Immunohistochemical characterization of nitric oxide synthase activity in squamous cell carcinoma of the head and neck. *Otolaryngol. Head. Neck Surg.* 1995, 113, 541–549.
140. Scott, D.J., Hull, M. a, Cartwright, E.J., Lam, W.K., et al., Lack of inducible nitric oxide synthase promotes intestinal tumorigenesis in the Apc(Min/+) mouse. *Gastroenterology* 2001, 121, 889–899.
141. Thomsen, L.L., Miles, D.W., Happerfield, L., Bobrow, L.G., et al., Nitric oxide synthase activity in human breast cancer. *Br. J. Cancer* 1995, 72, 41–44.
142. Mulshine, J.L., Cuttitta, F., Tockman, M.S., De Luca, L.M., Lung cancer evolution to preinvasive management. *Clin. Chest Med.* 2002, 23, 37–48.
143. Bilello, K.S., Murin, S., Matthay, R.A., Epidemiology, etiology, and prevention of lung cancer. *Clin. Chest Med.* 2002, 23, 1–25.
144. Masri, F.A., Comhair, S.A.A., Koeck, T., Xu, W., et al., Abnormalities in nitric oxide and its derivatives in lung cancer. *Am. J. Respir. Crit. Care Med.* 2005, 172, 597–605.
145. Wei, X., Wang, Q., Gao, S., Sui, L., [Relationship between nitric oxide in cervical microenvironment and different HPV types and effect on cervical cancer cells]. *Zhonghua Fu Chan Ke Za Zhi* 2011, 46, 260–265.
146. Chen, G.G., Lee, T.W., Xu, H., Yip, J.H.Y., et al., Increased inducible nitric oxide synthase in lung carcinoma of smokers. *Cancer* 2008, 112, 372–381.
147. Puhakka, A.R.A., Harju, T.H., Pääkkö, P.K., Soini, Y.M., Kinnula, V.L., Nitric oxide synthases are associated with bronchial dysplasia. *Lung Cancer* 2006, 51, 275–282.
148. Ohshima, H., Bartsch, H., Chronic infections and inflammatory processes as cancer risk factors: possible role of nitric oxide in carcinogenesis. *Mutat. Res.* 1994, 305, 253–264.

149. Beckman, J.S., Ischiropoulos, H., Zhu, L., van der Woerd, M., et al., Kinetics of superoxide dismutase- and iron-catalyzed nitration of phenolics by peroxynitrite. *Arch. Biochem. Biophys.* 1992, 298, 438–445.
150. Haddad, I.Y., Pataki, G., Hu, P., Galliani, C., et al., Quantitation of nitrotyrosine levels in lung sections of patients and animals with acute lung injury. *J. Clin. Invest.* 1994, 94, 2407–2413.
151. Cobbs, C.S., Whisenhunt, T.R., Wesemann, D.R., Harkins, L.E., et al., Inactivation of Wild-Type p53 Protein Function by Reactive Oxygen and Nitrogen Species in Malignant Glioma Cells. *Cancer Res.* 2003, 63, 8670–8673.
152. Masri, F., Role of nitric oxide and its metabolites as potential markers in lung cancer. *Ann. Thorac. Med.* 2010, 5, 123–7.
153. Brucefy, W., William Murrell. *Clin. Cardiol.* 1995, 18, 426–427.
154. Tzeng, E., Yoneyama, T., Hatakeyama, K., Shears, L.L., Billiar, T.R., Vascular inducible nitric oxide synthase gene therapy: Requirement for guanosine triphosphate cyclohydrolase I. *Surgery* 1996, 120, 315–321.
155. Goodwin, G.H., Johns, E.W., The Isolation and Purification of the High Mobility Group (HMG) Nonhistone Chromosomal Proteins. *Methods Cell Biol.* 1977.
156. Lotze, M.T., Tracey, K.J., High-mobility group box 1 protein (HMGB1): Nuclear weapon in the immune arsenal. *Nat. Rev. Immunol.* 2005.
157. Müller, S., Scaffidi, P., Degryse, B., Bonaldi, T., et al., The double life of HMGB1 chromatin protein: Architectural factor and extracellular signal. *EMBO J.* 2001.
158. Dong, X.D.E., Ito, N., Lotze, M.T., Demarco, R. a, et al., High mobility group box I (HMGB1) release from tumor cells after treatment: implications for development of targeted chemoimmunotherapy. *J. Immunother.* 2007.
159. Ellerman, J.E., Brown, C.K., de Vera, M., Zeh, H.J., et al., Masquerader: high mobility group box-1 and cancer. *Clin Cancer Res* 2007.
160. Sims, G.P., Rowe, D.C., Rietdijk, S.T., Herbst, R., Coyle, A.J., HMGB1 and RAGE in Inflammation and Cancer. *Annu. Rev. Immunol.* 2010.
161. Andersson, U., Tracey, K.J., HMGB1 Is a Therapeutic Target for Sterile Inflammation and Infection. *Annu. Rev. Immunol.* 2011.

162. Jube, S., Rivera, Z.S., Bianchi, M.E., Powers, A., et al., Cancer cell secretion of the DAMP protein HMGB1 supports progression in malignant mesothelioma. *Cancer Res.* 2012.
163. Kang, R., Tang, D., Schapiro, N.E., Loux, T., et al., The HMGB1/RAGE inflammatory pathway promotes pancreatic tumor growth by regulating mitochondrial bioenergetics. *Oncogene* 2014, 33, 567–77.
164. Tang, D., Kang, R., Livesey, K.M., Kroemer, G., et al., High-mobility group box 1 is essential for mitochondrial quality control. *Cell Metab.* 2011.
165. Taguchi, A., Blood, D.C., Del Toro, G., Canet, A., et al., Blockade of RAGE-amphoterin signalling suppresses tumour growth and metastases. *Nature* 2000, 405, 354–360.
166. Huttunen, H.J., Fages, C., Kuja-Panula, J., Ridley, A.J., Rauvala, H., Receptor for advanced glycation end products-binding COOH-terminal motif of amphoterin inhibits invasive migration and metastasis. *Cancer Res.* 2002, 62, 4805–4811.
167. Tang, D., Kang, R., Zeh 3rd, H.J., Lotze, M.T., High-mobility group box 1, oxidative stress, and disease. *Antioxid Redox Signal* 2011.
168. Campana, L., Bosurgi, L., Rovere-Querini, P., HMGB1: a two-headed signal regulating tumor progression and immunity. *Curr. Opin. Immunol.* 2008.
169. Kusume, A., Sasahira, T., Luo, Y., Isobe, M., et al., Suppression of dendritic cells by HMGB1 is associated with lymph node metastasis of human colon cancer. *Pathobiology* 2009.
170. Liu, Z., Falo, L.D., You, Z., Knockdown of HMGB1 in Tumor Cells Attenuates Their Ability To Induce Regulatory T Cells and Uncovers Naturally Acquired CD8 T Cell-Dependent Antitumor Immunity. *J. Immunol.* 2011.
171. He, Y., Zha, J., Wang, Y., Liu, W., et al., Tissue damage-associated “danger signals” influence T-cell responses that promote the progression of preneoplasia to cancer. *Cancer Res.* 2013.
172. Yan, W., Chang, Y., Liang, X., Cardinal, J.S., et al., High-mobility group box 1 activates caspase-1 and promotes hepatocellular carcinoma invasiveness and metastases. *Hepatology* 2012.
173. Tafani, M., Schito, L., Pellegrini, L., Villanova, L., et al., Hypoxia-increased RAGE and P2X7R expression regulates tumor cell invasion through phosphorylation of Erk1/2 and Akt and nuclear translocation of NF-κB.

174. Van Beijnum, J.R., Nowak-Sliwinska, P., Van Den Boezem, E., Hautvast, P., et al., Tumor angiogenesis is enforced by autocrine regulation of high-mobility group box 1. *Oncogene* 2013.
175. Jiao, Y., Wang, H.C., Fan, S.J., Growth suppression and radiosensitivity increase by HMGB1 in breast cancer. *Acta Pharmacol. Sin.* 2007.
176. Giavara, S., Kosmidou, E., Hande, M.P., Bianchi, M.E., et al., Yeast Nhp6A/B and mammalian Hmgb1 facilitate the maintenance of genome stability. *Curr. Biol.* 2005.
177. Celona, B., Weiner, A., Di Felice, F., Mancuso, F.M., et al., Substantial Histone reduction modulates Genomewide nucleosomal occupancy and global transcriptional output. *PLoS Biol.* 2011.
178. Polanská, E., Dobšáková, Z., Dvořáčková, M., Fajkus, J., Štros, M., HMGB1 gene knockout in mouse embryonic fibroblasts results in reduced telomerase activity and telomere dysfunction. *Chromosoma* 2012.
179. Sasahira, T., Akama, Y., Fujii, K., Kuniyasu, H., Expression of receptor for advanced glycation end products and HMGB1/amphoterin in colorectal adenomas. *Virchows Arch.* 2005, 446, 411–5.
180. Kuniyasu, H., Oue, N., Wakikawa, A., Shigeishi, H., et al., Expression of receptors for advanced glycation end-products (RAGE) is closely associated with the invasive and metastatic activity of gastric cancer. *J. Pathol.* 2002, 196, 163–170.
181. Dumitriu, I.E., Baruah, P., Manfredi, A.A., Bianchi, M.E., Rovere-Querini, P., HMGB1: Guiding immunity from within. *Trends Immunol.* 2005, 26, 381–387.
182. Chang, Y.-H., Chen, C.-M., Chen, H.-Y., Yang, P.-C., Pathway-based gene signatures predicting clinical outcome of lung adenocarcinoma. *Sci. Rep.* 2015, 5, 10979.
183. Kerr, J., Wyllie, A., Currie, A., Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br. J. Cancer* 1972.
184. Fan, T.J., Han, L.H., Cong, R.S., Liang, J., Caspase family proteases and apoptosis. *Acta Biochim. Biophys. Sin. (Shanghai).* 2005, 37, 719–727.
185. Eimon, P.M., Ashkenazi, A., The zebrafish as a model organism for the study of apoptosis. *Apoptosis* 2010, 15, 331–349.
186. Dewson, G., Kluck, R.M., Bcl-2 family-regulated apoptosis in health and disease. *Cell Health Cytoskeleton.* 2010, 2, 9–22.

187. Cory, S., Huang, D.C.S., Adams, J.M., The Bcl-2 family: roles in cell survival and oncogenesis. *Oncogene* 2003, 22, 8590–607.
188. Sharpe, J.C., Arnoult, D., Youle, R.J., Control of mitochondrial permeability by Bcl-2 family members. *Biochim. Biophys. Acta* 2004, 1644, 107–13.
189. Adams, J.M., Cory, S., The Bcl-2 apoptotic switch in cancer development and therapy. *Oncogene* 2007, 26, 1324–37.
190. Kim, H., Rafiuddin-Shah, M., Tu, H.-C., Jeffers, J.R., et al., Hierarchical regulation of mitochondrion-dependent apoptosis by BCL-2 subfamilies. *Nat. Cell Biol.* 2006, 8, 1348–58.
191. García-Sáez, A.J., The secrets of the Bcl-2 family. *Cell Death Differ.* 2012, 19, 1733–1740.
192. Wei, M.C., Zong, W.X., Cheng, E.H., Lindsten, T., et al., Proapoptotic BAX and BAK: a requisite gateway to mitochondrial dysfunction and death. *Science* 2001, 292, 727–30.
193. Letai, A., Bassik, M.C., Walensky, L.D., Sorcinelli, M.D., et al., Distinct BH3 domains either sensitize or activate mitochondrial apoptosis, serving as prototype cancer therapeutics. *Cancer Cell* 2002, 2, 183–192.
194. Kuwana, T., Bouchier-Hayes, L., Chipuk, J.E., Bonzon, C., et al., BH3 domains of BH3-only proteins differentially regulate Bax-mediated mitochondrial membrane permeabilization both directly and indirectly. *Mol. Cell* 2005, 17, 525–535.
195. Certo, M., Moore, V.D.G., Nishino, M., Wei, G., et al., Mitochondria primed by death signals determine cellular addiction to antiapoptotic BCL-2 family members. *Cancer Cell* 2006, 9, 351–365.
196. Llambi, F., Moldoveanu, T., Tait, S.W.G., Bouchier-Hayes, L., et al., A Unified Model of Mammalian BCL-2 Protein Family Interactions at the Mitochondria. *Mol. Cell* 2011, 44, 517–531.
197. Saelens, X., Festjens, N., Vande Walle, L., van Gurp, M., et al., Toxic proteins released from mitochondria in cell death. *Oncogene* 2004, 23, 2861–74.
198. Von Ahsen, O., Waterhouse, N., Kuwana, T., Newmeyer, D., Green, D., The “harmless” release of cytochrome C. *Cell Death Differ.* 2000, 7, 1192–1199.
199. Ricci, J.E., Waterhouse, N., Green, D.R., Mitochondrial functions during cell death, a complex (I-V) dilemma. *Cell Death Differ.* 2003, 10, 488–492.

200. Chipuk, J.E., Bouchier-Hayes, L., Green, D.R., Mitochondrial outer membrane permeabilization during apoptosis: the innocent bystander scenario. *Cell Death Differ.* 2006, 13, 1396–1402.
201. Taylor, R.C., Cullen, S.P., Martin, S.J., Apoptosis: controlled demolition at the cellular level. *Nat. Rev. Mol. Cell Biol.* 2008, 9, 231–41.
202. Chowdhury, I., Tharakan, B., Bhat, G.K., Caspases - an update. *Comp. Biochem. Physiol. B. Biochem. Mol. Biol.* 2008, 151, 10–27.
203. Elmore, S., Apoptosis: a review of programmed cell death. *Toxicol. Pathol.* 2007, 35, 495–516.
204. Lakhani, S.A., Masud, A., Kuida, K., Porter, G.A., et al., Caspases 3 and 7: key mediators of mitochondrial events of apoptosis. *Science* 2006, 311, 847–51.
205. Delgado, M.E., Olsson, M., Lincoln, F.A., Zhivotovsky, B., Rehm, M., Determining the contributions of caspase-2, caspase-8 and effector caspases to intracellular VDVADase activities during apoptosis initiation and execution. *Biochim. Biophys. Acta - Mol. Cell Res.* 2013, 1833, 2279–2292.
206. Ravichandran, K.S., Find-me and eat-me signals in apoptotic cell clearance: progress and conundrums. *J. Exp. Med.* 2010.
207. Radha, G., Raghavan, S.C., BCL2: A promising cancer therapeutic target. *Biochim. Biophys. Acta - Rev. Cancer* 2017, 1868, 309–314.
208. Oltersdorf, T., Elmore, S.W., Shoemaker, A.R., Armstrong, R.C., et al., An inhibitor of Bcl-2 family proteins induces regression of solid tumours. *Nature* 2005, 435, 677–681.
209. Delbridge, A.R.D., Grabow, S., Strasser, A., Vaux, D.L., Thirty years of BCL-2: Translating cell death discoveries into novel cancer therapies. *Nat. Rev. Cancer* 2016, 16, 99–109.
210. Chang, J., Wang, Y., Shao, L., Laberge, R.M., et al., Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat. Med.* 2016, 22, 78–83.
211. Zerp, S.F., Stoter, R., Kuipers, G., Yang, D., et al., AT-101, a small molecule inhibitor of anti-apoptotic Bcl-2 family members, activates the SAPK/JNK pathway and enhances radiation-induced apoptosis. *Radiat. Oncol.* 2009, 4, 47.
212. Rippin, T.M., Bykov, V.J.N., Freund, S.M.V., Selivanova, G., et al., Characterization of the p53-rescue drug CP-31398 in vitro and in living cells. *Oncogene* 2002.

213. Philchenkov, A., Miura, K., The IAP Protein Family, SMAC Mimetics and Cancer Treatment. *Crit. Rev. Oncog.* 2016.
214. Wang, Z., Song, W., Aboukameel, A., Mohammad, M., et al., TW-37, a small-molecule inhibitor of Bcl-2, inhibits cell growth and invasion in pancreatic cancer. *Int. J. Cancer* 2008.
215. Jiang, J., Slivova, V., Jedinak, A., Sliva, D., Gossypol inhibits growth, invasiveness, and angiogenesis in human prostate cancer cells by modulating NF- κ B/AP-1 dependent- and independent-signaling. *Clin. Exp. Metastasis* 2012.
216. Murphy, E., Steenbergen, C., Mechanisms underlying acute protection from cardiac ischemia-reperfusion injury. *Physiol. Rev.* 2008.
217. Bianchi, L., Gerstbrein, B., The neurotoxic MEC-4 (d) DEG/ENaC sodium channel conducts calcium: implications for necrosis initiation. *Nat. ...* 2004.
218. Galluzzi, L., Kroemer, G., Necroptosis: a specialized pathway of programmed necrosis. *Cell* 2008.
219. Kokoszka, J., Waymire, K., Levy, S., Sligh, J., The ADP/ATP translocator is not essential for the mitochondrial permeability transition pore. *Nature* 2004.
220. Baines, C.P., Kaiser, R.A., Sheiko, T., Craigen, W.J., Molkentin, J.D., Voltage-dependent anion channels are dispensable for mitochondrial-dependent cell death. *Nat. Cell Biol.* 2007, 9, 550–5.
221. Baines, C., Kaiser, R., Purcell, N., Blair, N., Loss of cyclophilin D reveals a critical role for mitochondrial permeability transition in cell death. *Nature* 2005.
222. Kim, I., Rodriguez-Enriquez, S., Lemasters, J.J., Selective degradation of mitochondria by mitophagy. *Arch. Biochem. Biophys.* 2007, 462, 245–53.
223. Soldani, C., Scovassi, A.I., Poly(ADP-ribose) polymerase-1 cleavage during apoptosis: An update. *Apoptosis* 2002, 7, 321–328.
224. Krysko, D., Berghe, T., Parthoens, E., Methods for distinguishing apoptotic from necrotic cells and measuring their clearance. *Methods ...* 2008.
225. Liu, G., Wang, J., Park, Y., High mobility group protein-1 inhibits phagocytosis of apoptotic neutrophils through binding to phosphatidylserine. *J. ...* 2008.
226. Degterev, A., Huang, Z., Boyce, M., Li, Y., et al., Chemical inhibitor of nonapoptotic cell death with therapeutic potential for ischemic brain injury. *Nat. Chem. Biol.* 2005, 1, 112–9.

227. Kaczmarek, A., Vandenabeele, P., Krysko, D. V., Necroptosis: The Release of Damage-Associated Molecular Patterns and Its Physiological Relevance. *Immunity* 2013, 38, 209–223.
228. Welz, P.-S., Wullaert, A., Vlantis, K., Kondylis, V., et al., FADD prevents RIP3-mediated epithelial cell necrosis and chronic intestinal inflammation. *Nature* 2011, 477, 330–334.
229. Bonnet, M.C., Preukschat, D., Welz, P.S., Van Loo, G., et al., The Adaptor Protein FADD Protects Epidermal Keratinocytes from Necroptosis In Vivo and Prevents Skin Inflammation. *Immunity* 2011, 35, 572–582.
230. Günther, C., Martini, E., Wittkopf, N., Amann, K., et al., Caspase-8 regulates TNF- α -induced epithelial necroptosis and terminal ileitis. *Nature* 2011, 477, 335–339.
231. Vercammen, D., Beyaert, R., Denecker, G., Goossens, V., et al., Inhibition of caspases increases the sensitivity of L929 cells to necrosis mediated by tumor necrosis factor. *J. Exp. Med.* 1998, 187, 1477–1485.
232. Tesniere, A., Schlemmer, F., Boige, V., Kepp, O., et al., Immunogenic death of colon cancer cells treated with oxaliplatin. *Oncogene* 2010, 29, 482–491.
233. Vakkila, J., Lotze, M.T., Inflammation and necrosis promote tumour growth. *Nat. Rev. Immunol.* 2004, 4, 641–648.
234. Li, L., Thomas, R.M., Suzuki, H., De Brabander, J.K., et al., A small molecule Smac mimic potentiates TRAIL- and TNF α -mediated cell death. *Science* 2004, 305, 1471–4.
235. Hu, X., Xuan, Y., Bypassing cancer drug resistance by activating multiple death pathways--a proposal from the study of circumventing cancer drug resistance by induction of necroptosis. *Cancer Lett.* 2008, 259, 127–37.
236. Han, W., Li, L., Qiu, S., Lu, Q., et al., Shikonin circumvents cancer drug resistance by induction of a necroptotic death. *Mol. Cancer* ... 2007.
237. Horita, H., Frankel, A.E., Thorburn, A., Acute myeloid leukemia-targeted toxin activates both apoptotic and necroptotic death mechanisms. *PLoS One* 2008, 3, e3909.
238. Bonapace, L., Bornhauser, B.C., Schmitz, M., Cario, G., et al., Induction of autophagy-dependent necroptosis is required for childhood acute lymphoblastic leukemia cells to overcome glucocorticoid resistance. *J. Clin. Invest.* 2010, 120, 1310–23.

239. Zafra-Polo, M.C., González, M.C., Estornell, E., Sahpaz, S., Cortes, D., Acetogenins from Annonaceae, inhibitors of mitochondrial complex I. *Phytochemistry* 1996, 42, 253–271.
240. Chahboune, N., Barrachina, I., Royo, I., Romero, V., et al., Guanaconetins, new antitumoral acetogenins, mitochondrial complex I and tumor cell growth inhibitors. *Bioorganic Med. Chem.* 2006, 14, 1089–1094.
241. Degli Esposti, M., Ghelli, A., Ratta, M., Cortes, D., Estornell, E., Natural substances (acetogenins) from the family Annonaceae are powerful inhibitors of mitochondrial NADH dehydrogenase (Complex I). *Biochem J* 1994, 301, 161–167.
242. De Pedro, N., Cautain, B., Melguizo, A., Vicente, F., et al., Mitochondrial complex I inhibitors, acetogenins, induce HepG2 cell death through the induction of the complete apoptotic mitochondrial pathway. *J. Bioenerg. Biomembr.* 2013, 45, 153–164.
243. Li, N., Ragheb, K., Lawler, G., Sturgis, J., et al., Mitochondrial complex I inhibitor rotenone induces apoptosis through enhancing mitochondrial reactive oxygen species production. *J. Biol. Chem.* 2003, 278, 8516–8525.
244. Radad, K., Rausch, W.D., Gille, G., Rotenone induces cell death in primary dopaminergic culture by increasing ROS production and inhibiting mitochondrial respiration. *Neurochem. Int.* 2006, 49, 379–386.
245. Sukhdev Swami Handa, Extraction Technologies for Medicinal and Aromatic Plants. *Uma ética para quantos?* 2014, XXXIII, 81–87.
246. Yung, O.H., Maskat, M.Y., Wan Mustapha, W.A., Kesan pengekstrakan terhadap kandungan polifenol, aktiviti antipengoksida dan pH ekstrak Pegaga (*Centella asiatica*). *Sains Malaysiana* 2010, 39, 747–752.
247. Vilku, K., Mawson, R., Simons, L., Bates, D., Applications and opportunities for ultrasound assisted extraction in the food industry - A review. *Innov. Food Sci. Emerg. Technol.* 2008.
248. Vinatoru, M., in: *Ultrason. Sonochem.*, vol. 8, 2001, pp. 303–313.
249. Chen, L., Jin, H., Ding, L., Zhang, H., et al., Dynamic microwave-assisted extraction of flavonoids from *Herba Epimedii*. *Sep. Purif. Technol.* 2008.
250. Wang, L., Weller, C.L., Recent advances in extraction of nutraceuticals from plants. *Trends Food Sci. Technol.* 2006.

251. Luque-García, J.L., Luque De Castro, M.D., Ultrasound: A powerful tool for leaching. *TrAC - Trends Anal. Chem.* 2003.
252. Wu, J., Lin, L., Chau, F.T., Ultrasound-assisted extraction of ginseng saponins from ginseng roots and cultured ginseng cells. *Ultrason. Sonochem.* 2001.
253. Dhanani, T., Shah, S., Gajbhiye, N.A., Kumar, S., Effect of extraction methods on yield, phytochemical constituents and antioxidant activity of *Withania somnifera*. *Arab. J. Chem.* 2013, 10, S1193–S1199.
254. Kaufmann, B., Christen, P., Recent extraction techniques for natural products: Microwave-assisted extraction and pressurised solvent extraction. *Phytochem. Anal.* 2002, 13, 105–113.
255. Araujo, P.W., Brereton, R.G., Experimental design I. Screening. *TrAC - Trends Anal. Chem.* 1996.
256. Lundstedt, T., Seifert, E., Abramo, L., Thelin, B., et al., Experimental design and optimization. *Chemom. Intell. Lab. Syst.* 1998.
257. Neto, B.B., Statistical Design – Chemometrics, 2005.
258. Abagyan, R., Totrov, M., High-throughput docking for lead generation. *Curr. Opin. Chem. Biol.* 2001, 5, 375–382.
259. Shoichet, B.K., McGovern, S.L., Wei, B., Irwin, J.J., Lead discovery using molecular docking. *Curr. Opin. Chem. Biol.* 2002, 6, 439–446.
260. McInnes, C., Virtual screening strategies in drug discovery. *Curr. Opin. Chem. Biol.* 2007, 11, 494–502.
261. Kitchen, D.B., Decornez, H., Furr, J.R., Bajorath, J., Docking and scoring in virtual screening for drug discovery: methods and applications. *Nat. Rev. Drug Discov.* 2004, 3, 935–49.
262. Ghosh, S., Nie, A., An, J., Huang, Z., Structure-based virtual screening of chemical libraries for drug discovery. *Curr. Opin. Chem. Biol.* 2006, 10, 194–202.
263. Cavasotto, C.N., Orry, A.J.W., Ligand docking and structure-based virtual screening in drug discovery. *Curr. Top. Med. Chem.* 2007, 7, 1006–1014.
264. Bernstein, F.C., Koetzle, T.F., Williams, G.J.B., Meyer, E.F., et al., The protein data bank: A computer-based archival file for macromolecular structures. *Arch. Biochem. Biophys.* 1978, 185, 584–591.
265. Pagadala, N.S., Syed, K., Tuszynski, J., Software for molecular docking: a review. *Biophys. Rev.* 2017, 9, 91–102.

266. Paulinus, O.N., Kinsley, A., Ikechi, E.G., Protective effect of ethanolic leaf extract of *Annona muricata* Linn. on some early events in cycas induced colorectal carcinogenesis in rats. *J. Pharm. Sci. Innov.* 2013, 2, 14–21.
267. Freshney, R.I., Culture of Animal Cells, a Manual of Basic Technique, John Wiley & Sons, 2005.
268. Mosmann, T., Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *J. Immunol. Methods* 1983, 65, 55–63.
269. Ragasa, C.Y., Soriano, G., Torres, O.B., Don, M.-J., Shen, C.-C., Acetogenins from *Annona muricata*. *Pharmacogn. J.* 2012, 4, 32–37.
270. Chang, F.R., Liaw, C.C., Lin, C.Y., Chou, C.J., et al., New adjacent bis-tetrahydrofuran annonaceous acetogenins from *Annona muricata*. *Planta Med.* 2003, 69, 241–246.
271. Sun, S., Liu, J., Kadouh, H., Sun, X., Zhou, K., Three new anti-proliferative Annonaceous acetogenins with mono-tetrahydrofuran ring from graviola fruit (*Annona muricata*). *Bioorg. Med. Chem. Lett.* 2014, 24, 2773–2776.
272. Coria-Télez, A. V., Montalvo-González, E., Yahia, E.M., Obledo-Vázquez, E.N., *Annona muricata*: A comprehensive review on its traditional medicinal uses, phytochemicals, pharmacological activities, mechanisms of action and toxicity. *Arab. J. Chem.* 2015.
273. Rizvi, S.M.D., Shakil, S., Haneef, M., A simple click by click protocol to perform docking: Autodock 4.2 made easy for non-bioinformaticians. *EXCLI J.* 2013, 12, 830–857.
274. Kelly, L.A., Mezulis, S., Yates, C., Wass, M., Sternberg, M., The Phyre2 web portal for protein modelling, prediction, and analysis. *Nat. Protoc.* 2015, 10, 845–858.
275. Colovos, C., Yeates, T.O., Verification of protein structures: Patterns of nonbonded atomic interactions. *Protein Sci.* 1993, 2, 1511–1519.
276. Bowie, J., Luthy, R., Eisenberg, D., A method to identify protein sequences that fold into a known three-dimensional structure. *Science (80-.)*. 1991, 253, 164–170.
277. Lüthy, R., Bowie, J.U., Eisenberg, D., Assessment of protein models with three-dimensional profiles. *Nature* 1992, 356, 83–85.

278. Lovell, S.C., Davis, I.W., Arendall, W.B., de Bakker, P.I.W., et al., Structure validation by Calpha geometry: phi,psi and Cbeta deviation. *Proteins* 2003, 50, 437–450.
279. Pettersen, E.F., Goddard, T.D., Huang, C.C., Couch, G.S., et al., UCSF Chimera—A Visualization System for Exploratory Research and Analysis. *J Comput Chem* 2004, 25, 1605–1612.
280. Abraham, M.J., Murtola, T., Schulz, R., Páll, S., et al., Gromacs: High performance molecular simulations through multi-level parallelism from laptops to supercomputers. *SoftwareX* 2015, 1–2, 19–25.
281. Schmid, N., Eichenberger, A.P., Choutko, A., Riniker, S., et al., Definition and testing of the GROMOS force-field versions 54A7 and 54B7. *Eur. Biophys. J.* 2011, 40, 843–856.
282. Schüttelkopf, A.W., Van Aalten, D.M.F., PRODRG: A tool for high-throughput crystallography of protein-ligand complexes. *Acta Crystallogr. Sect. D Biol. Crystallogr.* 2004, 60, 1355–1363.
283. Essmann, U., Perera, L., Berkowitz, M.L., Darden, T., et al., A smooth particle mesh Ewald method. *J. Chem. Phys.* 1995, 103, 8577–8593.
284. Hess, B., Bekker, H., Berendsen, H.J.C., Fraaije, J.G.E.M., LINCS: A linear constraint solver for molecular simulations. *J. Comput. Chem.* 1997, 18, 1463–1472.
285. Li, Y., Fabiano-Tixier, A.S., Tomao, V., Cravotto, G., Chemat, F., Green ultrasound-assisted extraction of carotenoids based on the bio-refinery concept using sunflower oil as an alternative solvent. *Ultrason. Sonochem.* 2013, 20, 12–18.
286. Erbay, Z., Icier, F., Optimization of hot air drying of olive leaves using response surface methodology. *J. Food Eng.* 2009, 91, 533–541.
287. Samavati, V., Polysaccharide extraction from *Abelmoschus esculentus*: Optimization by response surface methodology. *Carbohydr. Polym.* 2013, 95, 588–597.
288. Jin, X., Ning, Y., Extraction optimization and bioactivity of polysaccharides from *Aspergillus fumigatus* AF1. *Carbohydr. Polym.* 2013, 96, 411–416.
289. Prakash Maran, J., Mekala, V., Manikandan, S., Modeling and optimization of ultrasound-assisted extraction of polysaccharide from *Cucurbita moschata*. *Carbohydr. Polym.* 2013, 92, 2018–2026.

290. Hemwimol, S., Pavasant, P., Shotipruk, A., Ultrasound-assisted extraction of anthraquinones from roots of *Morinda citrifolia*. *Ultrason. Sonochem.* 2006, 13, 543–548.
291. Chemat, F., Rombaut, N., Sicaire, A.G., Meullemiestre, A., et al., Ultrasound assisted extraction of food and natural products. Mechanisms, techniques, combinations, protocols and applications. A review. *Ultrason. Sonochem.* 2017.
292. Lou, Z., Wang, H., Zhang, M., Wang, Z., Improved extraction of oil from chickpea under ultrasound in a dynamic system. *J. Food Eng.* 2010, 98, 13–18.
293. Zhang, Q.W., Lin, L.G., Ye, W.C., Techniques for extraction and isolation of natural products: A comprehensive review. *Chinese Med. (United Kingdom)* 2018.
294. Li, W., Liu, Z., Wang, Z., Chen, L., et al., Application of accelerated solvent extraction to the investigation of saikosaponins from the roots of *Bupleurum falcatum*. *J. Sep. Sci.* 2010.
295. Romdhane, M., Gourdon, C., Investigation in solid-liquid extraction: Influence of ultrasound. *Chem. Eng. J.* 2002.
296. Albu, S., Joyce, E., Paniwnyk, L., Lorimer, J.P., Mason, T.J., in: *Ultrason. Sonochem.*, 2004.
297. Rostagno, M.A., Palma, M., Barroso, C.G., Ultrasound-assisted extraction of soy isoflavones. *J. Chromatogr. A* 2003.
298. Zeng, L., Ye, Q., Oberlies, N.H., Shi, G., et al., Recent advances in annonaceous acetogenins. *Nat. Prod. Rep.* 2004.
299. Dasari, S., Bernard Tchounwou, P., Cisplatin in cancer therapy: Molecular mechanisms of action. *Eur. J. Pharmacol.* 2014, 740, 364–378.
300. Cavé, A., Figadère, B., Laurens, A., Cortes, D., Acetogenins from Annonaceae. *Fortschritte der Chemie ...* 1997.
301. McLaughlin, J.L., Paw Paw and Cancer: Annonaceous Acetogenins from Discovery to Commercial Products I. *J. Nat. Prod.* 2008, 71, 1311–1321.
302. Liu, M., Li, X.Q., Weber, C., Lee, C.Y., et al., Antioxidant and antiproliferative activities of raspberries. *J. Agric. Food Chem.* 2002.
303. Wolfe, K.L., Kang, X., He, X., Dong, M., et al., Cellular antioxidant activity of common fruits. *J. Agric. Food Chem.* 2008.

304. Hirano, K., Budiyo, E., Swastika, N., Fujii, K., Population dynamics of the whitefly, *Bemisia tabaci* (Gennadius) (Homoptera: Aleyrodidae), in Java, Indonesia, with special reference to spatio-temporal changes in the quantity of food resources. *Ecol. Res.* 1995.
305. Aboul-Enein, A.M., Abu El-Ela, F., Shalaby, E.A., El-Shemy, H.A., Traditional medicinal plants research in Egypt: Studies of antioxidant and anticancer activities. *J. Med. Plants Res.* 2012.
306. Ezirim, A., Okochi, V., James, A., Adebashi, O., et al., Induction of apoptosis in myelogenous leukemic K562 cells by ethanolic leaf extract of *Annona muricata*. *Indian J. Drugs ...* 2013.
307. Rieser, M.J., Fang, X.P., Rupprecht, J.K., Hui, Y.H., et al., Bioactive single-ring acetogenins from seed extracts of *Annona muricata*. *Planta Med.* 1993.
308. Wu, F.E., Gu, Z.M., Zeng, L., Zhao, G.X., et al., Two new cytotoxic monotetrahydrofuran annonaceous acetogenins, anomuricins A and B, from the leaves of *Annona muricata*. *J. Nat. Prod.* 1995.
309. Kojima, N., Tanaka, T., Medicinal chemistry of annonaceous acetogenins: Design, synthesis, and biological evaluation of novel analogues. *Molecules* 2009.
310. Alali, F.Q., Liu, X.X., McLaughlin, J.L., Annonaceous acetogenins: Recent progress. *J. Nat. Prod.* 1999, 62, 504–540.
311. Dzoyem, J.P., Donfack, A.R.N., Tane, P., McGaw, L.J., Eloff, J.N., Inhibition of nitric oxide production in LPS-stimulated RAW264.7 macrophages and 15-LOX activity by anthraquinones from *Pentas schimperii*. *Planta Med.* 2016, 82, 1246–1251.
312. Ignarro, L.J., Nitric Oxide: Biology and Pathobiology, vol. 1, 2009.
313. Lala, P.K., Chakraborty, C., Role of nitric oxide in carcinogenesis and tumour progression. *Lancet Oncol.* 2001, 2, 149–156.
314. Mocellin, S., Bronte, V., Nitti, D., Nitric oxide, a double edged sword in cancer biology: Searching for therapeutic opportunities. *Med. Res. Rev.* 2007, 27, 317–352.
315. Reuter, S., Gupta, S.C., Chaturvedi, M.M., Aggarwal, B.B., Oxidative stress, inflammation, and cancer: How are they linked? *Free Radic. Biol. Med.* 2010, 49, 1603–1616.

316. Choudhari, S.K., Nitric oxide and cancer: a review. *World J Surg Oncol* 2013, 11.
317. Papapetropoulos, A., Nitric oxide production contributes to the angiogenic properties of vascular endothelial growth factor in human endothelial cells. *J Clin Invest* 1997, 100.
318. Coneski, P.N., Schoenfisch, M.H., Nitric oxide release: Part III. Measurement and reporting. *Chem. Soc. Rev.* 2012, 41, 3753.
319. Jagetia, G.C., Baliga, M.S., The Evaluation of Nitric Oxide Scavenging Activity of Certain Indian Medicinal Plants In Vitro: A Preliminary Study. *J. Med. Food* 2004, 7, 343–348.
320. Mfotie Njoya, E., Munvera, A.M., Mkounga, P., Nkengfack, A.E., McGaw, L.J., Phytochemical analysis with free radical scavenging, nitric oxide inhibition and antiproliferative activity of *Sarcocephalus pobeguini* extracts. *BMC Complement. Altern. Med.* 2017, 17.
321. Kumar, S., Kashyap, P., Antiproliferative activity and nitric oxide production of a methanolic extract of *Fraxinus micrantha* on Michigan Cancer Foundation-7 mammalian breast carcinoma cell line. *J. Intercult. Ethnopharmacol.* 2015, 4, 109.
322. Kim, G.-T., Tran, N.K.S., Choi, E.-H., Song, Y.-J., et al., Immunomodulatory Efficacy of Standardized *Annona muricata* (Graviola) Leaf Extract via Activation of Mitogen-Activated Protein Kinase Pathways in RAW 264.7 Macrophages. *Evidence-Based Complement. Altern. Med.* 2016, 2016, 1–10.
323. Su, H., Bidère, N., Zheng, L., Cubre, A., et al., Requirement for caspase-8 in NF-kappaB activation by antigen receptor. *Science* 2005.
324. Schwerk, C., Schulze-Osthoff, K., in: *Biochem. Pharmacol.*, 2003.
325. Amarante-Mendes, G.P., Finucane, D.M., Martin, S.J., Cotter, T.G., et al., Anti-apoptotic oncogenes prevent caspase-dependent and independent commitment for cell death. *Cell Death Differ.* 1998.
326. Zamzami, N., Kroemer, G., The mitochondrion in apoptosis: How Pandora's box opens. *Nat. Rev. Mol. Cell Biol.* 2001, 2, 67–71.
327. Shimizu, S., Eguchi, Y., Kamiike, W., Itoh, Y., et al., Induction of apoptosis as well as necrosis by hypoxia and predominant prevention of apoptosis by Bcl-2 and Bcl-XL. *Cancer Res.* 1996, 56, 2161–2166.

328. Hockenbery, D.M., Oltvai, Z.N., Yin, X.M., Milliman, C.L., Korsmeyer, S.J., Bcl-2 functions in an antioxidant pathway to prevent apoptosis. *Cell* 1993, 75, 241–251.
329. Nakashima, T., Miura, M., Hara, M., Tetrocarcin A inhibits mitochondrial functions of Bcl-2 and suppresses its anti-apoptotic activity. *Cancer Res.* 2000, 60, 1229–1235.
330. Huang, Z., Bcl-2 family proteins as targets for anticancer drug design. *Oncogene* 2000, 19, 6627–6631.
331. Antony, P., Vijayan, R., Acetogenins from *Annona muricata* as potential inhibitors of antiapoptotic proteins: A molecular modeling study. *Drug Des. Devel. Ther.* 2016, 10, 1399–1410.
332. Petros, A., Olejniczak, E., Fesik, S., Structural biology of the Bcl-2 family of proteins. *Biochim. Biophys. Acta (BBA)- ...* 2004.
333. Souers, A.J., Levenson, J.D., Boghaert, E.R., Ackler, S.L., et al., ABT-199, a potent and selective BCL-2 inhibitor, achieves antitumor activity while sparing platelets. *Nat. Med.* 2013, 19, 202–208.
334. Chen, C.Y., Liu, T.Z., Tseng, W.C., Lu, F.J., et al., (-)-Anonaine induces apoptosis through Bax- and caspase-dependent pathways in human cervical cancer (HeLa) cells. *Food Chem. Toxicol.* 2008, 46, 2694–2702.
335. Mohamed, S.M., Hassan, E.M., Ibrahim, N. a, Cytotoxic and antiviral activities of aporphine alkaloids of *Magnolia grandiflora* L. *Nat. Prod. Res.* 2010, 24, 1395–402.
336. Meng, X.-Y., Zhang, H.-X., Mezei, M., Cui, M., Molecular docking: a powerful approach for structure-based drug discovery. *Curr. Comput. Aided. Drug Des.* 2011, 7, 146–57.
337. Plewczynski, D., Łaźniewski, M., Augustyniak, R., Ginalski, K., Can we trust docking results? Evaluation of seven commonly used programs on PDBbind database. *J. Comput. Chem.* 2011, 32, 742–755.
338. Chaitanya, M., Babajan, B., Anuradha, C.M., Naveen, M., et al., Exploring the molecular basis for selective binding of *Mycobacterium tuberculosis* Asp kinase toward its natural substrates and feedback inhibitors: A docking and molecular dynamics study. *J. Mol. Model.* 2010, 16, 1357–1367.
339. Shamriz, S., Ofoghi, H., Design, structure prediction and molecular dynamics simulation of a fusion construct containing malaria pre-erythrocytic vaccine

- candidate, PfCelTOS, and human interleukin 2 as adjuvant. *BMC Bioinformatics* 2016, 17, 71.
340. Esmaili, E., Shahlaei, M., Analysis of the flexibility and stability of the structure of magainin in a bilayer, and in aqueous and nonaqueous solutions using molecular dynamics simulations. *J. Mol. Model.* 2015, 21, 1–15.
341. Lobanov, M.I., Bogatyreva, N.S., Galzitskaia, O. V, Radius of gyration is indicator of compactness of protein structure. *Mol. Biol. (Mosk)*. 2008, 42, 701–706.