SUCCINIC ACID PRODUCTION VIA SIMULTANEOUS SACCHARIFICATION AND FERMENTATION OF OIL PALM EMPTY FRUIT BUNCH

JUNAID AKHTAR

A thesis submitted in fulfilment of the requirements for the award of the degree of Doctor of Philosophy (Bioprocess Engineering)

Faculty of Chemical and Energy Engineering Universiti Teknologi Malaysia

JANUARY 2017

To my beloved father and mother, Muhammad Israr & Shahida Nasreen, To my lovely wife, Nur Fasihah Abdul Kadir, My cute princess, Nur Ruhi Junaid Akhtar.

ACKNOWLEDGEMENT

First of all, all praises to Allah the Almighty for his kindness that give me strength to complete my PhD thesis. I would like to express my sincere gratitude and appreciation to my main supervisor Prof. Dr. Ani Idris for her guidance, advice, motivation, support and careful review during the course of my study. I benefited professionally and intellectually while working with her. More importantly, I appreciate her care, broad range of experience, technical insights, vision, patience and dedication in constructively criticizing my research work and thesis as well. I would like to express my genuine appreciation to Prof. Ramlan Abd Aziz for his inspiration and enthusiasm throughout this research work. It was a great pleasure for me to conduct research under his constructive comments.

My special acknowledgment to the Dean, Faculty of Chemical and Energy Engineering for their continuous support on postgraduate affairs. The greatest appreciation goes to all the technicians, especially Mr. Yakub, Mr Noordin, Mr. Muhammad, Mr. Ismail and Mrs. Fasha for their cooperation. Also to Mr. Affindi and Mr. Zainul of the Felda Semenchu Kota Tinggi, Johor for their warm cooperation and assistance in providing me the EFB samples.

Thanks to all my friends for the fun and laughter they bring to my life. I am very thankful to Universiti Teknologi Malaysia for the financial assistance and giving me the opportunity to learn how to learn. I thank all those who are not mentioned here, but have been a great help during my period of study.

ABSTRACT

Oil palm empty fruit bunch (EFB), a plentiful agricultural waste in Malaysia has never been utilized for the production of succinic acid via simultaneous saccharification and fermentation (SSF). The presence of lignin and hemicellulose makes the biomass difficult to be hydrolysed by enzymes and microbes. Hence, effective pretreatment method is required to release cellulose from the crystalline complex structure of lignocellulose. The novelty of this study is the production of succinic acid via SSF from EFB by a rumen bacteria Actinobaccilus succinogenes ATCC 55618. The effect of three different methods; autoclave/alkali (AA), dilute acid (DA) and sequential dilute acid microwave/alkali (DA-MwA) pretreatment on the physical and chemical properties of EFB were analysed and their influence on enzymatic hydrolysis and SSF process were also assessed. Results revealed that maximum amount of cellulose (86.8 g/100g) was achieved for DA-MwA as compared with AA (53.3 g/100g) and DA (46.7 g/100g). The highest glucose concentration among all pretreated EFB was DA-MwA (20.3 gL⁻¹) method using cellulase enzyme. The effect of different cellulase: cellobiase ratios on enzymatic hydrolysis of DA-MwA pretreated EFB showed that ratio of 7:1 produced 34.45 % higher glucose as compared when only cellulase was used. Succinic acid concentration via SSF from DA-MwA (33.4 gL⁻¹) was the highest followed by AA (20.9 gL^{-1}) and DA (14.4 gL^{-1}) . The SSF media for the succinic acid production was optimized using Full Factorial Design (FFD) by varying the EFB loading (10-70 gL⁻¹), yeast extract (0-20 gL⁻¹) and corn steep liquor (0-20 gL⁻¹) followed by Face Central Composite Design. The best concentration of succinic acid (39.14 gL⁻¹) was obtained when the values of EFB, yeast extract and corn steep liquor were set at 70 gL^{-1} , 30 gL^{-1} and 10 gL^{-1} , respectively. The influence of three independent SSF process variables: enzyme loading (10-70 FPU/g), temperature (36-40 °C) and pH (5-8) were investigated for succinic acid production using FFD. When the enzyme loading was set at 40 FPU/g, temperature 36 °C and pH 5; the experimental values were in good agreement with the predicted Response Surface Methodology model where the best succinic acid production of 42.5 gL⁻¹ was achieved. The present study revealed that using DA-MwA pretreated EFB, cellulose was utilized by cellulase and cellobiase enzymes via optimized SSF conditions resulting in optimum production of succinic acid.

ABSTRAK

Tandan kosong buah kelapa sawit (EFB), adalah sisa pertanian yang banyak di Malaysia dan tidak pernah lagi digunakan bagi penghasilan asid suksinik melalui penapaian dan pensakaridaan serentak (SSF). Kehadiran lignin dan hemiselulosa menjadikan bahan biojisim sukar untuk dihidrolsis oleh enzim dan mikrob. Oleh itu, kaedah pra-rawatan adalah diperlukan untuk membebaskon selulosa daripada struktur kompleks kristal lignoselulosa. Penemuan baru bagi kajian ini adalah penghasilan asid suksinik melalui SSF daripada EFB oleh bakteria rumen Actinobaccilus succinogenes ATCC 55618. Kesan daripada tiga kaedah yang berbeza; autoklaf/alkali (AA), asid cair (DA), dan siri asid cair gelombang mikro/ alkali (DA-MwA) ke atas sifat-sifat fizik dan kimia bagi EFB telah dikaji dan pengaruh mereka terhadap proses SSF dan hidrolisis enzim juga telah dinilai. Hasil kajian menunjukkan bahawa kuantiti maksima selulosa (86.8 g/100g) diperolehi melalui kaedah DA-MwA berbanding AA (53.3 g/100g) dan DA (46.7 g/100g). Kepekatan glukosa yang maksima dicapai antara pra-rawatan EFB adalah melalui kaedah DA-MwA (20.3 gL⁻¹) menggunakan enzim selulase. Kesan dari purata berbeza selulase dan selobiase bagi pra-rawatan EFB kaedah DA-MwA memberi hasil glukosa yang paling tinggi 34.45 % pada nisbah 7:1 berbanding penggunaan selulase sahaja. Kepekatan asid suksinik melalui SSF iaitu daripada DA-MwA (33.4 gL⁻¹) adalah yang tertinggi diikuti oleh AA (20.9 gL⁻¹) dan DA (14.4 gL⁻¹). Media SSF bagi penghasilan asid suksinik telah dioptimum menggunakan Reka bentuk Faktor Penuh (FFD) dengan mengubah kuantiti muatan EFB (10-70 gL⁻¹), ekstrak yis $(0-20 \text{ gL}^{-1})$ dan likuor tusuk jagong $(0-20 \text{ gL}^{-1})$, diikuti oleh Reka bentuk Komposit Muka Pusat. Kepekatan asid suksinik yang paling optimum (39.14 gL^{-1}) telah diperoleh apabila kuantiti EFB, ekstrak yis dan likuor tusuk jagong ditetapkan masing - masing pada 70 gL⁻¹, 30 gL⁻¹ dan 10 gL⁻¹. Pengaruh pembolehubah bebas: muatan enzim (10-70 FPU/g), suhu (36-40 °C) dan pH (5-8) telah dikaji untuk penghasilan asid suksinik menggunakan FFD. Apabila muatan enzim ditetapkan pada 40 FPU/g, suhu 36 °C dan pH 5; asid suksinik mencatatkan hasil pada 42.5 gL⁻¹ melalui model ramalan Kaedah Gerak Balas Permukaan. Kajian ini mendedahkan bahawa pra-rawatan melalui kaedah DA-MwA, selulosa telah digunakan dengan baik oleh enzim selulase dan selobiase melalui pengoptimuman keadaan SSF yang menghasilkan pengeluaran asid suksinik yang optimum.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	V
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xii
	LIST OF FIGURES	xiv
	LIST OF SYMBOLS	xviii
	LIST OF ABBREVIATIONS	XX
	LIST OF APPENDICES	xxiii
1	INTRODUCTION	1
	1.1 Overview	1
	1.2 Problem Statement	6
	1.3 Objective	7
	1.4 Scope of the Study	7
	1.5 Significance of the Study	8
	1.6 Organization of the Thesis	9
2	LITERATURE REVIEW	10
	2.1 Introduction	10
	2.2 Economics Potential of Succinic Acid	11
	2.3 Biomass in Malaysia	13

2.4	Composition of Lignocellulosic Biomass	14
2.5	Pretreatment of Lignocellulosic Materials	19
	2.5.1 Physical Methods	21
	2.5.2 Chemical Methods	22
	2.5.3 Biological Pretreatment	22
2.6	Pretreatment Methods in Current Study	23
	2.6.1 Alkaline Autoclave Pretreatment	23
	2.6.2 Dilute Acid Pretreatment	24
	2.6.3 Microwave Alkali Pretreatment	24
	2.6.4 Microwave versus Conventional	
	Heating	26
2.7	Saccharification of Lignocellulosic Biomass	27
	2.7.1 The Effect of <i>Cellulase</i> and <i>Cellobiase</i>	
	on Saccharification	28
	2.7.2 The Effect of Enzymes ratio on	
	Saccharification	31
	2.7.3 Kinetics of Hydrolysis	31
2.8	Production of Succinic Acid by SHF from	
	Refined Sugars	33
2.9	Production of Succinic Acid by SHF from	
	Lignocelluosic Biomass	36
2.10	Succinic Acid Production by Simultaneous	
	Saccharification and Fermentation (SSF))	40
2.11	Anaerobic Fermentation	41
2.12	Production of Succinic Acid using Different	
	Microorganisms	42
2.13	Process Parameters in Succinic Acid Production	44
	2.13.1 Effect of SSF Medium Composition	45
	2.13.2 Effect of Temperature	46
	2.13.3 Effect of pH	47
2.14	Design of Experiment (DoE)	48

3 METHODOLOGY

viii

50

3.1	Materia	als	50
3.2	Pretrea	tment of Empty Fruit Bunch	51
	3.2.1	Autoclave/ alkali (AA) Pretreatment	51
	3.2.2	Dilute acid (DA) Pretreatment	52
	3.2.3	Sequential Dilute Acid	
		Microwave/Alkali (DA-MwA)	
		Pretreatment	52
3.3	Morph	ology Analysis	53
	3.3.1	Analysis of Functional Group using	
		FT-IR	53
	3.3.2	Surface Analysis by SEM and EDX	53
	3.3.3	X-ray Diffraction (XRD)	53
3.4	Analys	is of EFB Composition	54
	3.4.1	Total Solid Contents	54
	3.4.2	Moisture Content	54
	3.4.3	Determination of Holocellulose	55
	3.4.4	Determination of α -Cellulose	56
	3.4.5	Determination of Lignin	56
	3.4.6	Determination of Silica	57
3.5	Enzym	atic Hydrolysis of EFB Pretreated with	
	Differe	ent Pretreatment Methods	57
	3.5.1	Enzyme Activity Assays	58
	3.5.2	Enzymatic Kinetics Study	58
3.6	SSF of	Differently Pretreated EFB	59
3.7	Effect	of Media Composition on SSF	60
	3.7.1	Full Factorial Experimental Design for	
		SSF Media Composition	61
	3.7.2	Central Composite Experimental Design	
		for SSF Media Composition	61
	3.7.3	Effect of Temperature, pH and Enzyme	
		Loading in SSF of Succinic Acid	63

3.8	Analys	is of the Collected Samples	63
	3.8.1	Succinic Acid Analysis	64
	3.8.2	Total Sugar Analysis using DNS	
		Method	64
SAC	CHARIF CHES FI	MENT AND ENZYMATIC ICATION OF EMPTY FRUIT IBER of Different Pretreatments on EFB	65
	Compo	osition	65
4.2	Effect	of Microwave Parameters on Chemical	
	Compo	osition of EFB	68
	4.2.1	Effect of Microwave Power on EFB	
		Composition	68
	4.2.2	Effect of Time on EFB Composition	70
	4.2.3	Effect of Temperature on EFB	
		Composition	71
4.3	Physic	al Appearance of the Pretreated EFB	72
	4.3.1	Morphology Analysis by Field Emission	
		Scanning Electron Microscopy	
		(FESEM)	73
	4.3.2	Spectroscopy Measurement by Fourier	
		Transform Infrared (FTIR)	78
	4.3.3	Crystallinity Index Measurement of EFB	79
4.4	Enzym	atic Saccharification of Pretreated EFB	81
	4.4.1	Effect of Different Pretreatment on	
		Enzymatic Hydrolysis	81
	4.4.2	Effect of Different Enzymes Ratios on	
		Enzymatic Hydrolysis	82
4.5	Kinetic	c of Enzymatic Hydrolysis	85

4

5 SIMULTANEOUS SACCHARIFICATION AND FERMENTATION OF THE PRETREATED OIL

	PAL	M EMPT	Y FRUIT BUNCHES BY A.	
	SUC	SUCCINOGENES		
	5.1	SSF of	EFB using Different Pretreatment	
		Method	ls	87
	5.2	Effect	of Media Composition	94
		5.2.1	Full Factorial Experimental Design for	
			SSF Media Composition	95
		5.2.2	Central Composite Experimental Design	
			for SSF Media Composition	102
		5.2.3	Optimization	110
	5.3	Effect	of Temperature, pH and Enzyme Loading	
		in SSF	of Succinic Acid	111
	5.4	Valida	tion of RSM Optimization Points	124
6	CON	CLUSIO	NS AND RECOMMENDATIONS	126
	6.1	Conclu	sion	126
	6.2	Recom	mendations	128
REFEREN	CES			130

A 1' A TT	140,170
Appendices A - H	148-158

LIST OF TABLES

TABLE NO.	TITLE	PAGE
1.1	The amount of biomass waste supply from 2001 to	
	2020 (tons per year of dry weight) in Malaysia	3
2.1	Composition of various lignocelluloses material	16
2.2	Component of cellulase system using 10% cheese	
	whey and 1% Avicel	29
2.3	Succinic acid concentration, productivity and yield	
	using glucose/glycerol, galactose as substrate by SHF	34
2.4	Succinic acid concentration, productivity and yield	
	using different lignocellulosic substrate, microbe and	
	fermentation systems using SHF	38
2.5	Succinic acid production via SSF and its	
	concentration, productivity and yield using different	
	substrate, microbe and fermentation systems	40
3.1	Chemicals used in the experiment	50
3.2	Full factorial design arrangement and responses	61
3.3	Central composite design arrangement and responses	62
3.4	Full factorial design arrangement and responses	63
4.1	Chemical composition of raw and the differently	
	treated EFB biomass (g/100 g biomass)	65
4.2	Composition of silica of raw and differently	
	pretreated EFB biomass	67
4.3	Effect of cellulase and cellobiase on enzymatic	
	hydrolysis	83
5.1	Succinic acid production via SSF and its	
	concentration and yield using different substrate in	93

anaerobic flasks

5.2	Full factorial design arrangement and responses	95
5.3	Analysis of variance (ANOVA) for succinic acid	
	production	96
5.4	Analysis of variance (ANOVA) for glucose	
	accumulation	96
5.5	Regression equation obtained for succinic acid	
	production and glucose accumulation	97
5.6	Central composite design arrangement and responses	103
5.7	Analysis of variance (ANOVA) for succinic acid	
	production	104
5.8	Analysis of variance (ANOVA) for glucose	
	accumulation	104
5.9	Regression equation obtained for succinic acid	
	production and glucose accumulation	105
5.10	Confirmation run at optimum RSM conditions	111
5.11	Full factorial design arrangement and responses	111
5.12	Analysis of variance (ANOVA) for succinic acid	
	production	112
5.13	Analysis of variance (ANOVA) for glucose	
	accumulation	113
5.14	Regression equation obtained for succinic acid	114
	production and glucose accumulation	
5.15	Confirmation run at optimum RSM conditions	125

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
1.1	Schematic diagram for SHF and SSF of succinic	
	acid production from EFB	5
2.1	Map of routes to various products formed from	
	succinic acid	12
2.2	Structure of cellulose, hemicellulose and lignin	15
2.3	Simplified form of pretreatment of lignocellulosic	
	biomass	20
2.4	Microwave and conventional heating mechanism	27
2.5	Mechanism of degradation of cellulose into glucose	
	by enzymes	30
3.1	Flow chart of the overall process in the present study	60
4.1	Effect of the microwave power on the composition	
	of the empty fruit bunch (EFB) with 2.5 M NaOH, at	
	80°C temperature within 60 minutes.	69
4.2	Effect of the time on the composition of the empty	
	fruit bunch (EFB) with 2.5M NaOH, at 80°C	
	temperature and 900 watts microwave power	70
4.3	Effect of the temperature on the composition of the	
	empty fruit bunch (EFB) with 2.5M NaOH, 900	
	watts microwave power within 80 min duration	71
4.4	Morphologies of the EFB samples	73
4.5	FESEM images of raw EFB fiber	74
4.6	FESEM images of autoclave/ alkali (AA) pretreated	
	EFB fiber	75
4.7	FESEM images of dilute acid pretreated EFB fiber	76

microwave/alkali (DA-MwA) pretreated EFB fiber774.9FTIR chromatograms: 4000–370 cm ⁻¹ of (a) the untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave-Alkali (MW-A) pretreated EFB784.10XRD diagram within 20 scale ranging from 5 to 40° images of (a) untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave- Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid production via SSF with untreated EFB855.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.6Normal probability plot of residuals for a) succinic92	4.8	FESEM images of sequential dilute acid	
untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave-Alkali (MW-A) pretreated EFB784.10XRD diagram within 20 scale ranging from 5 to 40° images of (a) untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave- Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB91		microwave/alkali (DA-MwA) pretreated EFB fiber	77
pretreated EFB; (c) dilute acid microwave-Alkali (MW-A) pretreated EFB784.10XRD diagram within 20 scale ranging from 5 to 40° images of (a) untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave- Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB91	4.9	FTIR chromatograms: $4000-370 \text{ cm}^{-1}$ of (a) the	
(MW-A) pretreated EFB784.10XRD diagram within 20 scale ranging from 5 to 40° images of (a) untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave- Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid production via SSF with untreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		untreated EFB; (b) conventional dilute alkali	
4.10XRD diagram within 20 scale ranging from 5 to 40° images of (a) untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave- Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB91		pretreated EFB; (c) dilute acid microwave-Alkali	
images of (a) untreated EFB; (b) conventional dilute alkali pretreated EFB; (c) dilute acid microwave- Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB885.2Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		(MW-A) pretreated EFB	78
alkali pretreated EFB; (c) dilute acid microwave- Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB885.2Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	4.10	XRD diagram within 2Θ scale ranging from 5 to 40°	
Alkali (MW-A) pretreated EFB804.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		images of (a) untreated EFB; (b) conventional dilute	
4.11Enzymatic hydrolysis of differently pretreated EFB with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		alkali pretreated EFB; (c) dilute acid microwave-	
with cellulase enzyme. The values stated are averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		Alkali (MW-A) pretreated EFB	80
averages from duplicate experiments824.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	4.11	Enzymatic hydrolysis of differently pretreated EFB	
4.12Enzymatic hydrolysis of Sequential DA-MwA EFF with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		with cellulase enzyme. The values stated are	
with cellulase and a supplementation of a small amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		averages from duplicate experiments	82
amount of cellobiase enzyme844.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	4.12	Enzymatic hydrolysis of Sequential DA-MwA EFF	
4.13Effect of substrate concentration on initial hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		with cellulase and a supplementation of a small	
hydrolysis rate of dilute acid microwave/alkali pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		amount of <i>cellobiase</i> enzyme	84
pretreated EFB854.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	4.13	Effect of substrate concentration on initial	
4.14Lineweaver Burk plot of enzymatic hydrolysis865.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		hydrolysis rate of dilute acid microwave/alkali	
5.1Time courses of succinic acid production via SSF with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92		pretreated EFB	85
with untreated EFB885.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	4.14	Lineweaver Burk plot of enzymatic hydrolysis	86
5.2Time courses of succinic acid production via SSF with autoclave/ alkali (AA) pretreated EFB895.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	5.1	Time courses of succinic acid production via SSF	
 with autoclave/ alkali (AA) pretreated EFB 89 5.3 Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB 90 5.4 Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB 91 5.5 Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB 92 		with untreated EFB	88
5.3Time courses of succinic acid production via SSF with dilute acid (DA) pretreated EFB905.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	5.2	Time courses of succinic acid production via SSF	
 with dilute acid (DA) pretreated EFB 90 5.4 Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB 91 5.5 Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB 92 		with autoclave/ alkali (AA) pretreated EFB	89
5.4Time courses of succinic acid production via SSF with Sequential DA-MwA pretreated EFB915.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	5.3	Time courses of succinic acid production via SSF	
 with Sequential DA-MwA pretreated EFB 91 5.5 Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB 92 		with dilute acid (DA) pretreated EFB	90
5.5Time courses of comparative analysis of succinic acid production via SSF different pretreated EFB92	5.4	Time courses of succinic acid production via SSF	
acid production via SSF different pretreated EFB 92		with Sequential DA-MwA pretreated EFB	91
	5.5	Time courses of comparative analysis of succinic	
5.6 Normal probability plot of residuals for a) succinic		acid production via SSF different pretreated EFB	92
	5.6	Normal probability plot of residuals for a) succinic	
acid production, b) glucose accumulation 98		acid production, b) glucose accumulation	98
5.7 Plot of residuals vs. predicted for a) succinic acid	5.7	Plot of residuals vs. predicted for a) succinic acid	

	production, b) glucose accumulation	98
5.8	Effect of substrate concentration on a) succinic acid	
	production, b) glucose accumulation	99
5.9	Effect of yeast extract concentration on a) succinic	
	acid production, b) glucose accumulation	99
5.10	Effect of corn steep liquor concentration on a)	
	succinic acid production, b) glucose accumulation	100
5.11	Response surfaces (a and b) and counter (c and d)	
	plots for succinic acid production	101
5.12	Response surfaces (a and b) and counter (c and d)	
	plots for glucose accumulation	102
5.13	Normal probability plot of residuals for a) succinic	
	acid production, b) glucose accumulation via CCD	106
5.14	Plot of residuals vs. predicted for a) succinic acid	
	production, b) glucose accumulation via CCD	107
5.15	Effect of substrate concentration on a) succinic acid	
	production, b) glucose accumulation via CCD	108
5.16	Effect of yeast extract concentration on a) succinic	
	acid production, b) glucose accumulation via CCD	108
5.17	Response surface (a) and counter plot (b) for	
	succinic acid production via CCD	109
5.18	Response surface (a) and counter plot (b) for glucose	
	accumulation via CCD	110
5.19	Normal probability plot of residuals for a) succinic	
	acid production, b) glucose accumulation via FFD	115
5.20	Plot of residuals vs. predicted for a) succinic acid	
	production, b) glucose accumulationvia FFD	115
5.21	Effect of enzyme loading on a) succinic acid	
	production, b) glucose accumulation	116
5.22	Effect of pH on a) succinic acid production, b)	
	glucose accumulation	117
5.23	Effect of temperature on a) succinic acid production,	
	b) glucose accumulation	118

5.24	Interaction effect for succinic acid production a)	
	enzyme – pH b) enzyme - temp c) pH - temp	119
5.25	Interaction effect for glucose accumulation a)	
	enzyme – pH b) enzyme - temp c) pH-temp	120
5.26	Response surface (a,c) enzyme - temp and contour	
	plot (b,d) pH - temp for succinic acid production	121
5.27	Response surface a) enzyme – pH b) enzyme - temp	
	c) pH - temp and contour plot d) enzyme – pH e)	
	enzyme - temp f) pH - temp for glucose	122
	accumulation	

LIST OF SYMBOLS

Α	-	Weight of lignin in grams			
a_3	-	Weight. of crucible with oven dry cellulose			
MT	-	Metric ton			
α	-	Alfa			
β	-	Beta			
€/MT	-	Euro per metric ton			
^o C	-	Degree Celsius			
%	-	Percentage			
FPU/g	-	Filter paper unit per gram			
CBU/g	-	Cellobiase unit per gram			
ст	-	Centimeter			
cm^{-1}	-	Reciprocal centimeters			
\$	-	Dollar sign			
USD	-	United States dollar			
MPa	-	Megapascal			
mg	-	Milligram			
mgL^{-1}	-	Milligram per liter			
L	-	Liter			
g	-	Gram			
h	-	Hour			
М	-	Molar			
gL^{-1}	-	Gram per liter			
kg^{-1}	-	Per kilogram			
тM	-	Millimolar			
mL	-	Milliliter			
min	-	Minute			

gg^{-1}	-	Gram per gram			
$gL^{-1}h^{-1}$	-	Gram per liter per hour			
rpm	-	Rounds per minute			
pH	-	Measure of hydrogen ion concentration			
V _{max}	-	Maximum rate of reaction at infinite substrate			
		concentration			
V _{emax}	-	Maximum velocity			
[S]	-	Substrate concentration			
R^2	-	Regression coefficient			
$[E]_o$	-	Initial enzyme concentration			
[E]	-	Enzyme concentration			
K_m	-	Michaelis-Menten constant			
K_e	-	Half saturation constant of substrate			
v	-	Initial hydrolysis velocity			
<i>w/v</i>	-	Weight per volume			
W	-	Watt			
W	-	Oven-dry weight of test specimen in grams			
W_1	-	Weight of sample			
W_2	-	Weight of dish			
W_3	-	Weight of dried sample plus dish			
W_3X	-	Weight. Of oven dried sample (thanmble)			
Z_3	-	Weight of cellulose which can be calculated by (air			
		dry holocellulose with crucible-weight of crucible)			
(<i>f</i>)	-	Function			
µmol	-	Micromoles			
μm	-	Micrometer			
θ	-	Theta			

LIST OF ABBREVIATIONS

AA	-	Autoclave alkali
ANOVA	-	Analysis of variance
ARP	-	Ammonia recycle percolation
ATCC	-	American Type Culture Collection
CCD	-	Central composite design
CFH	-	Corn fiber hydrlyzate
CMCase	-	Endoglucanase
CO_2	-	Carbon dioxide
CO3 ²⁻	-	Carbonate ion
CSH	-	Corn straw hydrolysate
CSL	-	Corn steep liquor
CaCl ₂	-	Calcium chloride
CaCO ₃	-	Calcium carbonate
Ca(OH) ₂	-	Calcium hydroxide
$C_4H_6O_4$	-	Succinic acid
DA	-	Dilute acid
DA-MwA	-	Dilute acid-microwave alkali
DNS	-	3, 5-dinitrosalicylic acid
DOE	-	Department of Energy
DoE	-	Design of expert
EFB	-	Empty fruit bunch
EDX	-	Energy Dispersive X-ray analysis
etc.	-	Et cetera
et al.	-	And others
FESEM	-	Field Emission Scanning Electron Microscope
FFB	-	Fresh fruit bunch

FT-IR	_	Fourier Transform Infrared spectroscopy
Fpase	_	Cellobiohydrolase
HCl	_	Hydrochloric acid
HCO ₃ ⁻	_	Hydrogen carbonate
HPLC	_	High Performance Liquid Chromatography
HILE H ₂ O	_	Water, ice or steam
H ₂ O ₂ -	_	Hydrogen peroxide
H_2O_2 H_2SO_4	_	Sulfuric acid
IU IU		International unit
КОН	-	Potassium hydroxide
	-	•
K_2 HPO ₄	-	Dipotassium phosphate
KH ₂ OH ₄	-	Potassium dihydrogen phosphate
LAPS	-	Laboratory testing analytical procedures
LOF	-	Lack of fit
MgCO ₃	-	Magnesium carbonate
MgCl ₂	-	Magnesium chloride
MgSO ₄ .7H ₂ 0	-	Magnesium sulphate heltahydrate
Mg(OH) ₂	-	Magnesium hydroxide
MwA	-	Microwave alkali
NREL	-	National Renewable Energy Laboratory
NaCl	-	Sodium chloride
NaOH	-	Sodium hydroxide
NaH ₂ PO ₄	-	Sodium dihydrogen phosphate
OPF	-	Oil palm frond
OPT	-	Oil palm trunk
O_2	-	Oxygen
PEP	-	Phosphoenolpyruvate
PEPCK	-	Phosphoenolpyruvate carboxykinase
PKS	-	Palm kernel shell
рКа	-	Acid dissociation constant
RM	_	Malaysian Ringgit
RSM	_	Response surface methodology
SAA	_	Aqueous ammonia
JAA	-	riqueous annionia

SEM	-	Scanning electron microscope			
SHF	-	Separate enzymatic hydrolysis and cultivation			
SP	-	Steam pretreatment			
SSF	-	Simultaneous saccharification and cultivation			
		or Simultaneous saccharification and			
		fermentation			
US	-	United States			
USA	-	United States of America			
XRD	-	X-ray powder diffraction			
YE	-	Yeast extract			
YSH	-	Yeast cell hydrolyzate			
ZnSO ₄ .7H ₂ 0	-	Zinc sulphate heltahydrate			
A	-	Actinobacillus			
M	-	Mannheimia			
3D	-	Three dimentional			

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Publications	148
В	Succinic Acid Concentration Standard Curve	150
С	Glucose Concentration Standard Curve	151
D	Formic Acid Concentration Standard Curve	152
E	Acetic Acid Concentration Standard Curve	153
F	Experimental Data for SSF process	154
G	Enzyme Activity Conversion	156
н	Experimental Data for Enzymatic	
п	Saccharification Process	158

CHAPTER 1

INTRODUCTION

1.1 Overview

Succinic acid ($C_4H_6O_4$), a dicarboxylic acid, was isolated for the first time from microbial fermentation in 1546. Its traditional name is amber acid, but it is also known as butanedioic acid. It is formed by plants, animals and microorganisms, but its maximum production is obtained through anaerobic fermentation by microbes. Succinic acid originates from carbohydrate fermentation and is extensively used in chemical markets and industries which are producing food, green solvents and biodegradable plastics and ingredients used for the stimulation of plant growth (Zeikus *et al.*, 1999). Succinate is the feedstock for several industrial products including tetrahydrofuran, adipic acid, 1,4-butanediol, and aliphatic esters (Willke and Vorlop, 2004).

Succinic acids were previously produced from refined sugar, maleic anhydride and petrochemicals. The production cost was high because of the expensive substrate, so a low price substrate lignocellulosic biomass was used to produce a high yield, concentration and productivity of organic acids such as lactic acid, succinic acid, acetic acid, citric acid and bioethanol (Fitzpatrick *et al.*, 2010). The maleic anhydride price and the environment is adversely impacted, therefore, the need has been felt in developing alternative resources and environmentally friendly technologies. This lead to the development of alternative cheaper biochemical processes which can complement and replace the conventional chemical methods (Pinazo *et al.*, 2015).

Lignocellulosic production is about 180 million tons, from terrestrial plants 1.3×10^{10} metric tons and coal is 7 x 10⁹ metric tons per year which are equal to fulfill two- thirds of world energy requirements (Demain *et al.*, 2005). Lignocellulosic biomass such EFB, oil palm trunks (OPT), oil palm fronds (OPF), rice straw, wheat straw, corn stem, corn husk, corn cobs, palm etc. contain high amounts of cellulose components. It has been calculated that cellulose production from biomass was around 1.5 trillion tons per year. High value-added and environmentally friendly product such as succinic acid can be produced from these low prices and inexhaustible source of lignocellulosic raw materials (Kim *et al.*, 2006). In order for the raw material to be considered a good substrate for the production of organic acids it must have the following characteristics: available abundantly throughout the year, renewable, cheap, produce less amounts or no by-products formation, stereospecific, high production rate, low level of contaminants and no competing food value (John *et al.*, 2006b).

Research on lignocellulosic biomass has gained interest because it can replace existing expensive raw materials and are abundantly available, renewable and cheap (Zhang *et al.*, 2007). Another reason that has made the re-use of lignocellulosic more attractive is the environmental issue. Before this lignocellulosic biomass especially EFB was burned and produced ash which consists of 30% potassium. Due to environmental pollution issue, these open combustion was forbidden to minimize emission of greenhouse gases (Suhaimi and Ong, 2001). Hence to achieve zero emission of greenhouse gases and maintain a healthy environment, production of high value-added products from this lignocellulosic biomass such as EFB are also aggressively considered. The utilization of lignocellulosic biomass opens a new field for the researchers to utilize it for the production of industrially important compounds such as succinic acid.

Malaysia is the world second largest producer of oil palm with approximately 59 million tons/year of the total lignocellulosic biomass agricultural waste. The oil

palm biomass comprised of empty fruit bunches (EFBs), oil palm fronds (OPFs), palm kernel shells (PKSs) and oil palm trunks (OPTs). These biomass wastes constitute 26. 2 million tons of oil palm fronds and 23% of empty fruit bunch (EFB) per ton of fresh fruit bunch (FFB), 7.0 million tons of oil palm trunks. In 2010 EFB that was processed were about 88.74 MT while palm biomass that was extracted from trunks and fronds were approximately 87 MT. There are many studies that suggest on the use of lignocellulosic biomass especially empty fruit bunches for the production of industrially important organic acids and biofuels (Hamzah *et al.*, 2009). The amount of biomass waste supply in Malaysia from 2001-2020 (tones per year, dry weight) is depicted in Table 1.1.

Table 1.1: The amount of biomass waste supply from 2001 to 2020 (tons peryear of dry weight) in Malaysia (Hassan *et al.*, 1997)

Year								
Biomass Supply	2001- 2003	2004- 2006	2007- 2010	2011- 2013	2014- 2016	2017- 2020		
Empty Fruit Bunches	2,870,148	2,860,194	2,823,695	2,830,331	2,906,647	2,863,512		
Oil Palm Fronds	7,412,074	7,025,525	6,890,223	6,803,260	7,044,853	7,141,490		
Oil Palm Trunk	3,933,442	4,020,852	3,234,164	4,283,082	3,583,803	2,971,934		

Currently, many researchers are working to synthesize succinic acid from lignocellulosic biomass to replace expensive pure sugar because of succinic acid demand in the world market. Production of succinic acid from corn fiber, corn stalk hydrolysate, rapeseed and cane molasses etc. has been studied in detail. The purpose of this research is to produce high value added product succinic acid from EFB, instead of burning, to solve environmental pollution issue.

Production of biosuccinate from renewable resources with the optimized condition is a focus point for the last two decades. There are several problems faced in biosuccinic acid production such as substrate requirements, auxotrophy and complex medium conditions and low production rate (Beauprez *et al.*, 2010). For

commercial and economic point of view, biosuccinic process must compete with a chemical process. The price of biobased succinic acid is 85 to $1040 \notin /MT$, while that produced from maleic anhydride is ~2550 \notin/MT . Biobased succinic acid price is even lower than maleic anhydride itself which cost $1059 \notin/MT$. Chemical and oil industry were highly interlaced with each other in the previous century and this price will be tripled in the coming years because of the increasing oil price. Therefore, the need will be for the cheaper biobased production of succinic acid from renewable sources that will replace the chemical method (Pinazo *et al.*, 2015)

Lignocelluloses conversion into the cellulose fraction and the production of succinic acid can be done by two methods, separate hydrolysis and fermentation (SHF) or simultaneous saccharification and fermentation (SSF). SHF refers to the process in which lignocellulosic bioconversion occurs in two steps; enzyme hydrolysis followed by fermentation in two different reactors. SSF is a one step process where enzymatic hydrolysis and fermentation occur within the same bioreactor (Lynd *et al.*, 2005, Xu *et al.*, 2009). Several studies (Mckinlay and Vieille, 2008) revealed that production of succinic acid from cellulose can be done more effectively by combining the two steps: enzymatic hydrolysis and microbial fermentation into a single step known as SSF.

SSF of lignocellulosic biomass are a novel technique, a time and cost effective process and can replace the two step fermentation process SHF, because it can reduce costs by replacing high amount of biomass consumption and also achieve high productivity by controlling the release of sugar. Process efficiency can be enhanced and brings it to the level equal to *cellulase* enzyme by the use of thermotolerant organisms. Also, the effect of glucose inhibition of enzymes is minimized (John *et al.*, 2009). Figure 1.1 clearly shows the process for succinic acid production through simultaneous saccharafication and fermentation (SSF) and the two-step process (SHF).

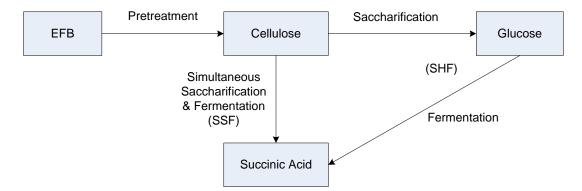


Figure 1.1Schematic diagram for SHF and SSF of succinic acid production fromEFB.

In SSF, cellulose was hydrolyzed to produce glucose and the glucose formed is simultaneously converted to succinic acid. The SSF process eliminates saccharafication of fermentable sugars step before fermentation. Hence, it is capable to substantially decrease the utilization of the enzyme loading. From the industrial view point this is important, as faster saccharification rate can be achieved in a reduced reactor volume. Besides, the SSF process also eliminates the use of different reaction vessels for both of the process. In SHF, the glucose produced was capable to competitively inhibit the *cellulase* and resulted in a low hydrolysis rate. Findings of studies revealed that the saccharification rate in SSF is faster than single saccharification process. Consequently, the higher saccharification rate increases productivity and reduce reactor volume and capital cost (Zhang *et al.*, 2007). Hence, the SSF process is seen to be a more comprehensive yet a simple process for utilization of lignocellulosic material.

However, it is also observed in SSF process the concentration and productivity achieved tend to be lower than SHF. The concentration of succinic acid attained using corn fiber as a substrate by *A. Succinogenes* via SHF process was 70.3 gL⁻¹. However, when the same substrate and bacteria in SSF process was used, the succinic acid concentration reduced to 47.4 gL⁻¹. However the metabolic engineering of the strains and fermentation conditions can solve this problem and increase the product concentration in SSF (Chen *et al.*, 2011d).

1.2 Problem Statement

Typical lignocellulosic biomasses contain 40% - 60% cellulose, 20% - 40% hemicelluloses and 10% - 25% lignin. (Mosier *et al.*, 2005). The potential of using the lignocellulosic material in this case of EFB as a substrate for organic acid production is due to the cellulose content that can be degraded into glucose subunits through hydrolysis process. However, due to the presence of lignin and hemicellulose which are entwined and closely attached to each other hindered the enzymatic hydrolysis and fermentation process. Hence, different pretreatment processes need to be performed to disrupt the lignocellulosic structure so as to remove maximum hemicellulose and lignin, thus exposing cellulose for enzymatic digestion.

Several pretreatment processes such as chemical, physical or both were performed for lignocellulosic materials and the choice for the process is very dependent on the type of lignocellulosic biomass used. Each pretreatment process has its advantages and disadvantages; steam pretreatment is effective for hardwoods but not for the softwood lignocellulose. Strong acid can produce high sugar yields but corrosive and has several disadvantages such as production of furfural (Ramos, 2003). Thus there is a need to find a suitable pretreatment method for EFB so that a high yield of succinic acid is achieved.

Succinic acid can be produced by two step process separate enzymatic hydrolysis and fermentation (SHF) and single step process simultaneous saccharification and fermentation (SSF). SSF process is quite complex and different substrates require different media composition, temperature and pH. Many studies suggest that enzymatic hydrolysis is favorable in low pH <5 and high temperature > 45° C while, fermentation is favorable at high pH > 6 and temperature below <40 °C (Chen *et al.*, 2011d, Li *et al.*, 2011, Li *et al.*, 2010, Li *et al.*, 2010b, Liu *et al.*, 2008a, Zheng *et al.*, 2009). Thus there is a need to determine the suitable process conditions when performing SSF for new substrates such as EFB.

1.3 Objective

- To investigate the effect of different pretreatment methods on the morphology and chemical composition of EFB.
- 2) To study the influence of differently pretreated EFB on the enzymatic hydrolysis in terms of glucose produced.
- 3) To determine the effect of different pretreated EFB on succinic acid production via SSF.
- 4) To optimize SSF conditions such as media composition, temperature, pH, enzyme and substrate loadings using Design of Experiment.

1.4 Scope of the Study

The research was conducted within the following limits:

- The influence of three different pretreatment methods: i) dilute acid (DA) pretreatment, using 8% Sulphuric acid, ii) autoclave alkali (AA) pretreatment using 20% NaOH, iii) sequential DA-MwA pretreatment, DA pretreatment followed by microwave alkali (MwA) pretreatment on composition of EFB were investigated. The morphological tests such as FESEM, XRD and FT-IR analysis of differently pretreated EFB samples were thoroughly examined.
- 2) The effect of differently pretreated EFB on enzymatic hydrolysis was investigated using *Cellulase* enzyme 25 FPU/g. The best pretreated method was selected to study different ratios of *cellulase* and *cellobiase* (10 CBU/g) (1:0, 1:1, 1:2, 2:1, 5:1, 7:1 and 10:1) to evaluate the comparatively glucose formation. The enzymatic hydrolysis were carried out in 150ml flasks having 50 ml of citrate buffer solution at 50°C, pH 4.8, 100 rpm in a water bath with 70 gL⁻¹ of the substrate loading.

- Succinic acid production via SSF was performed using rumen bacteria *A*. Succinogenes in the 150ml flasks containing 50ml of the fermentation medium for 48 h. Cellulase and cellubiase (Novozyme 188) were added into the fermentation together with *A. succinogenes*. Samples were taken at 6, 12, 24, 36 and 48 h for analysis of the products by HPLC.
- 4) SSF media composition were optimized by varying substrate loading, yeast extract and corn steep liquor using full factorial design followed by central composite design. SSF process conditions were also optimized based on three independent variables like enzyme loading, pH and temperature using full factorial design: All factors were statistically judged by analysis of variance (ANOVA). The optimal conditions of RSM for succinic acid production were validated by confirmation experiments.

1.5 Significance of the Study

Oil palm EFB was used for the first time to produce succinic acid via SSF. Succinic acid is largely produced from refined sugars, petrochemical and maleic anhydride which is quite expensive. EFB an agriculture waste is a suitable substrate for the production of succinic acid because it is easily available in the local milling area, low price, non-starchy and is rich in cellulose content.

The optimal temperature and pH to achieve a high succinic acid production in the one step SSF process using rumen bacteria *A succinogenes* were determined and this contribute to scientific research and advancement of data, as no work was performed to produce succinic acid from EFB via SSF. The effect of different pretreatment methods on the morphology and chemical composition of EFB and subsequently a succinic acid yield were disclosed. Similarly, various parameters of microwave/alkali (MW-A) pretreatment of EFB will be studied in detail to attain high amount of cellulose and remove maximum hemicellulose and lignin.

1.6 Organization of the Thesis

Chapter 1 gives a general overview of bioconversion of lignocellulose EFB to succinic acid. The chapter also focused on the SSF process, problem statement, objectives, scope and significance of the study. Chapter 2 gives a general overview of literature related to work done by the previous researchers on conversion of lignocellulose to succinic acid via SHF and SSF. The chapter also describes importance of succinic acid factor affecting SSF and optimization of SSF. Chapter 3 describes the methodology used in the study which includes pretreatment, enzymatic saccharification, SSF and optimization of SSF conditions. Chapter 4 explains the effect of pretreatment on the morphology and chemical composition. The effect of enzymatic saccharification of EFB was studied to attain maximum glucose accumulation and effect on enzyme kinetics. Chapter 5 briefly explains about SSF process, optimization of SSF media, factors affecting SSF. Chapter 6 reveals the conclusion of the present study and suggests recommendations for improvement in future studies.

REFERENCES

- Agarwal, L., Isar, J., Dutt, K. and Saxena, R. K. (2007). Statistical optimization for succinic acid production from E. coli in a cost-effective medium. *Applied Biochemistry and Biotechnology*, 142, 158-167.
- Alvira, P., Tomás-Pejó, E., Ballesteros, M. and Negro, M. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: a review. *Bioresource Technology*, 101, 4851-4861.
- Andersson, C., Helmerius, J., Hodge, D., Berglund, K. A. and Rova, U. (2009). Inhibition of succinic acid production in metabolically engineered Escherichia Coli by neutralizing agent, organic acids, and osmolarity. *Biotechnology Progress*, 25, 116-123.
- Bailey, C. J. (1989). Enzyme kinetics of cellulose hydrolysis. *Biochemical Journal*, 262, 1001.
- Beauprez, J. J., De Mey, M. and Soetaert, W. K. (2010). Microbial succinic acid production: Natural versus metabolic engineered producers. *Process Biochemistry*, 45, 1103-1114.
- Bell, J. (1984). Nutrients and toxicants in rapeseed meal: a review. *Journal of Animal Science*, 58, 996-1010.
- Beszédes, S., Ábel, M., Szabó, G., Hodúr, C. and László, Z. (2011). Enhanced enzymatic saccharification of agrifood solid wastes by microwave pretreatment. *Annals of the Faculty of Engineering Hunedoara*, 9, 453.
- Beszédes, S., Tachon, A., Lemmer, B., Ábel, M., Szabó, G. and Hodúr, C. (2012). Bio-fuels from cellulose by microwave irradiation. *Annals of the Faculty of Engineering Hunedoara*, 10, 43.
- Binod, P., Satyanagalakshmi, K., Sindhu, R., Janu, K. U., Sukumaran, R. K. and Pandey, A. (2012). Short duration microwave assisted pretreatment enhances the enzymatic saccharification and fermentable sugar yield from sugarcane bagasse. *Renewable Energy*, 37, 109-116.

- Bommarius, A. S., Katona, A., Cheben, S. E., Patel, A. S., Ragauskas, A. J., Knudson, K. and Pu, Y. (2008). Cellulase kinetics as a function of cellulose pretreatment. *Metabolic Engineeering*, 10, 370-381.
- Borges, E. R. and Pereira Jr, N. (2011). Succinic acid production from sugarcane bagasse hemicellulose hydrolysate by Actinobacillus succinogenes. *Journal of Industrial Microbiology and Biotechnology*, 38, 1001-1011.
- Bridgeman, T. G., Jones, J. M., Shield, I. and Williams, P. T. (2008). Torrefaction of reed canary grass, wheat straw and willow to enhance solid fuel qualities and combustion properties. *Fuel*, 87, 844-856.
- Cao, Y. and Tan, H. (2002). Effects of cellulase on the modification of cellulose. *Carbohydrate Research*, 337, 1291-1296.
- Carrillo, F., Lis, M., Colom, X., López-Mesas, M. and Valldeperas, J. (2005). Effect of alkali pretreatment on cellulase hydrolysis of wheat straw: Kinetic study. *Process Biochemistry*, 40, 3360-3364.
- Chandra, R. P., Bura, R., Mabee, W. E., Berlin, A., Pan, X. and Saddler, J. N. (2007). Substrate Pretreatment: The Key to Effective Enzymatic Hydrolysis of Lignocellulosics. *In:* Olsson, L. (ed.) *Biofuels*. Springer Berlin Heidelberg.
- Chang, V. S. and Holtzapple, M. T. (Year) Published. Fundamental factors affecting biomass enzymatic reactivity. Twenty-First Symposium on Biotechnology for Fuels and Chemicals, 2000. Springer, 5-37.
- Chatterjee, R., Millard, C. S., Champion, K., Clark, D. P. and Donnelly, M. I. (2001). Mutation of the ptsG Gene Results in Increased Production of Succinate in Fermentation of Glucose by Escherichia coli. *Applied and environmental microbiology*, 67, 148-154.
- Chaturvedi, V. and Verma, P. (2013). An overview of key pretreatment processes employed for bioconversion of lignocellulosic biomass into biofuels and value added products. 3 *Biotech*, 3, 415-431.
- Chayabutra, C., Wu, J. and Ju, L. K. (2001). Succinic acid production with reduced by-product formation in the fermentation of *Anaerobiospirillum* succiniciproducens using glycerol as a carbon source. *Biotechnology and Bioengineering*, 72, 41-48.
- Chen, J., Fales, S. L., Varga, G. A. and Royse, D. J. (1995). Biodegradation of cell wall components of maize stover colonized by white-rot fungi and resulting

impact on in-vitro digestibility. Journal of the Science of Food and Agriculture, 68, 91-98.

- Chen, K.-Q., Li, J., Ma, J.-F., Jiang, M., Wei, P., Liu, Z.-M. and Ying, H.-J. (2011a). Succinic acid production by *Actinobacillus succinogenes* using hydrolysates of spent yeast cells and corn fiber. *Bioresource Technology*, 102, 1704-8.
- Chen, K., Jiang, M., Wei, P., Yao, J. and Wu, H. (2010a). Succinic acid production from acid hydrolysate of corn fiber by Actinobacillus succinogenes. *Applied biochemistry and biotechnology*, 160, 477-485.
- Chen, K., Zhang, H., Miao, Y., Jiang, M. and Chen, J. (2010b). Succinic acid production from enzymatic hydrolysate of sake lees using *Actinobacillus* succinogenes 130Z. Enzyme and Microbial Technology, 47, 236-240.
- Chen, K., Zhang, H., Miao, Y., Wei, P. and Chen, J. (2011b). Simultaneous saccharification and fermentation of acid-pretreated rapeseed meal for succinic acid production using Actinobacillus succinogenes. *Enzyme and Microbial Technology*, 48, 339-344.
- Chen, W. H., Tu, Y. J. and Sheen, H. K. (2010c). Impact of dilute acid pretreatment on the structure of bagasse for bioethanol production. *International Journal of Energy Research*, 34, 265-274.
- Cheng, K. K., Zhao, X. B., Zeng, J. and Zhang, J. A. (2012). Biotechnological production of succinic acid: current state and perspectives. *Biofuels*, *Bioproducts and Biorefining*, 6, 302-318.
- Chiang, W.-D., Chang, S.-W. and Shieh, C.-J. (2003). Studies on the optimized lipase-catalyzed biosynthesis of cis-3-hexen-1-yl acetate in n-hexane. *Process Biochemistry*, 38, 1193-1199.
- Chrastil, J. (1988). Enzymic product formation curves with the normal or diffusion limited reaction mechanism and in the presence of substrate receptors. *International Journal of Biochemistry*, 20, 683-693.
- Christia, A., Setiowati, A. D., Millati, R., Karimi, K., Cahyanto, M. N., Niklasson, C. and Taherzadeh, M. J. (2016). Ethanol production from alkali-pretreated oil palm empty fruit bunch by simultaneous saccharification and fermentation with mucor indicus. *International Journal of Green Energy*, 13, 566-572.
- Datta, R., Glassner, D. A., Jain, M. K. and Roy, J. R. V. 1992. Fermentation and purification process for succinic acid. Google Patents.

- De La Hoz, A., Diaz-Ortiz, A. and Moreno, A. (2005). Microwaves in organic synthesis. Thermal and non-thermal microwave effects. *Chemical Society Reviews*, 34, 164-178.
- Deepak, V., Kalishwaralal, K., Ramkumarpandian, S., Babu, S. V., Senthilkumar, S. and Sangiliyandi, G. (2008). Optimization of media composition for Nattokinase production by Bacillus subtilis using response surface methodology. *Bioresource Technology*, 99, 8170-8174.
- Deguchi, S., Tsujii, K. and Horikoshi, K. (2008). Crystalline-to-amorphous transformation of cellulose in hot and compressed water and its implications for hydrothermal conversion. *Green Chemistry*, 10, 191-196.
- Demain, A. L., Newcomb, M. and Wu, J. D. (2005). Cellulase, clostridia, and ethanol. *Microbiology and Molecular Biology Reviews*, 69, 124-154.
- Dijkerman, R., Bhansing, D. C. P., Op Den Camp, H. J. M., Van Der Drift, C. and Vogels, G. D. (1997). Degradation of structural polysaccharides by the plant cell-wall degrading enzyme system from anaerobic fungi: An application study. *Enzyme and Microbial Technology*, 21, 130-136.
- Du, C., Lin, S. K. C., Koutinas, A., Wang, R. and Webb, C. (2007). Succinic acid production from wheat using a biorefining strategy. *Applied Microbiology* and Biotechnology, 76, 1263-1270.
- Efe, T., Van Der Wielen, L. a. M. and Straathof, A. J. J. (2013). Techno-economic analysis of succinic acid production using adsorption from fermentation medium. *Biomass and Bioenergy*, 56, 479-492.
- Elibol, M. and Ozer, D. (2002). Response surface analysis of lipase production by freely suspended *Rhizopus arrhizus*. *Process Biochemistry*, 38, 367-372.
- Fitzpatrick, M., Champagne, P., Cunningham, M. F. and Whitney, R. A. (2010). A biorefinery processing perspective: treatment of lignocellulosic materials for the production of value-added products. *Bioresource Technology*, 101, 8915-8922.
- Galbe, M. and Zacchi, G. (2007). Pretreatment of Lignocellulosic Materials for Efficient Bioethanol Production. Advances in Biochemichal Engineering/Biotechnology. 108, 41-65.
- Gan, Q., Allen, S. and Taylor, G. (2003). Kinetic dynamics in heterogeneous enzymatic hydrolysis of cellulose: an overview, an experimental study and mathematical modelling. *Process Biochemistry*, 38, 1003-1018.

- Garde, A., Jonsson, G., Schmidt, A. S. and Ahring, B. K. (2002). Lactic acid production from wheat straw hemicellulose hydrolysate by Lactobacillus pentosus and Lactobacillus brevis. *Bioresource Technology*, 81, 217-223.
- Glassner, D. A. and Datta, R. 1992. Process for the production and purification of succinic acid. Google Patents.
- Gokarn, R. R., Eiteman, M. A. and Altman, E. (1998). Expression of pyruvate carboxylase enhances succinate production in *Escherichia coli* without affecting glucose uptake. *Biotechnology Letters*, 20, 795-798.
- Goudar, C. T., Harris, S. K., McInerney, M. J. and Suflita, J. M. (2004). Progress Curve Analysis for Enzyme and Microbial Kinetic Reactions using Explicit Solutions Based on the Lambert W Function. *Journal of Microbiological Methods*, 59. 317-326.
- Gowen, A., Abu-Ghannam, N., Frias, J. and Oliveira, J. (2006). Optimisation of dehydration and rehydration properties of cooked chickpeas (Cicer arietinum L.) undergoing microwave-hot air combination drying. *Trends in Food Science & Technology*, 17, 177-183.
- Gubicza, K., Nieves, I. U., Sagues, W. J., Barta, Z., Shanmugam, K. and Ingram, L.
 O. (2016). Techno-economic analysis of ethanol production from sugarcane bagasse using a Liquefaction plus Simultaneous Saccharification and co-Fermentation process. *Bioresource Technology*, 208, 42-48.
- Guettler, M. V., Jain, M. K. and Rumler, D. 1996. Method for making succinic acid, bacterial variants for use in the process, and methods for obtaining variants. Google Patents.
- Guettler, M. V., Jain, M. K. and Soni, B. K. 1998. Process for making succinic acid, microorganisms for use in the process and methods of obtaining the microorganisms. Google Patents.
- Guettler, M. V., Rumler, D. and Jain, M. K. (1999). Actinobacillus succinogenes sp. nov., a novel succinic-acid-producing strain from the bovine rumen. International Journal of Systematic and Evolutionary Microbiology, 49, 207-216.
- Haki, G. and Rakshit, S. (2003). Developments in industrially important thermostable enzymes: a review. *Bioresource Technology*, 89, 17-34.

- Hamzah, F., Idris, A., Rashid, R. and Ming, S. (2009). Lactic acid production from microwave-alkali pre-treated empty fruit bunches fiber using *Rhizopus oryzae* pellet. *Journal of Applied Sciences*, 9, 3086-3091.
- Hamzah, F., Idris, A. and Shuan, T. K. (2011). Preliminary study on enzymatic hydrolysis of treated oil palm (Elaeis) empty fruit bunches fiber (EFB) by using combination of *cellulase* and β *1-4 glucosidase*. *Biomass and Bioenergy*, 35, 1055-1059.
- Hansen, S. (2007). Feasibility study of performing an life cycle assessment on crude palm oil production in Malaysia (9 pp). *The International Journal of Life Cycle Assessment*, 12, 50-58.
- Haque, M. A., Barman, D. N., Kim, M. K., Yun, H. D. and Cho, K. M. (2015). Cogon grass (Imperata cylindrica), a potential biomass candidate for bioethanol: cell wall structural changes enhancing hydrolysis in a mild alkali pretreatment regime. *Journal of the Science of Food and Agriculture*, 96: 1790–1797
- Hassan, K., Husin, M., Darus, A. and Jalani, B. (1997). *An estimated availability of oil palm biomass in Malaysia*, Palm Oil Research Institute of Malaysia.
- Hodge, D. B., Andersson, C., Berglund, K. A. and Rova, U. (2009). Detoxification requirements for bioconversion of softwood dilute acid hydrolyzates to succinic acid. *Enzyme and Microbial Technology*, 44, 309-316.
- Howard, R. L., Abotsi, E., Van Rensburg, E. L. J. and Howard, S. (2003). Lignocellulose biotechnology: Issues of bioconversion and enzyme production. *African Journal of Biotechnology*, 2, 702-733.
- Hu, Z., Wang, Y. and Wen, Z. (2008). Alkali (NaOH) pretreatment of switchgrass by radio frequency-based dielectric heating. *Applied biochemistry and biotechnology*, 148, 71-81.
- Hu, Z. and Wen, Z. (2008). Enhancing enzymatic digestibility of switchgrass by microwave-assisted alkali pretreatment. *Biochemical Engineering Journal*, 38, 369-378.
- Imai, M., Ikari, K. and Suzuki, I. (2004). High-performance hydrolysis of cellulose using mixed cellulase species and ultrasonication pretreatment. *Biochemical Engineering Journal*, 17, 79-83.
- Itoh, H., Wada, M., Honda, Y., Kuwahara, M. and Watanabe, T. (2003). Bioorganosolve pretreatments for simultaneous saccharification and

fermentation of beech wood by ethanolysis and white rot fungi. *Journal of Biotechnology*, 103, 273-280.

- Jaafar, M. Z., Kheng, W. H. and Kamaruddin, N. (2003). Greener energy solutions for a sustainable future: issues and challenges for Malaysia. *Energy policy*, 31, 1061-1072.
- Jahaan, F., Wasoh, H. and Abd-Aziz, S. (2014). Effect of different alkaline treatment on the release of ferulic acid from oil palm empty fruit bunch fibers. *Journal* of Oil Palm Research, 26, 321-331.
- Jantama, K., Zhang, X., Moore, J. C., Shanmugam, K. T., Svoronos, S. A. and Ingram, L. O. (2008). Eliminating side products and increasing succinate yields in engineered strains of *Escherichia coli* C. *Biotechnology and Bioengineering*, 101, 881-893.
- Jeffries, T. W. and Schartman, R. (Year) Published. Bioconversion of secondary fiber fines to ethanol using counter-current enzymatic saccharification and co-fermentation. Twentieth Symposium on Biotechnology for Fuels and Chemicals, 1999. Springer, 435-444.
- Jeoh, T., Ishizawa, C. I., Davis, M. F., Himmel, M. E., Adney, W. S. and Johnson, D. K. (2007). Cellulase digestibility of pretreated biomass is limited by cellulose accessibility. *Biotechnology and Bioengineering*, 98, 112-122.
- John, R. P., Nampoothiri, K. M. and Pandey, A. (2006a). Simultaneous saccharification and L-(+)-lactic acid fermentation of protease-treated wheat bran using mixed culture of *lactobacilli*. *Biotechnology Letters*, 28, 1823-1826.
- John, R. P., Nampoothiri, K. M. and Pandey, A. (2006b). Solid-state fermentation for L-lactic acid production from agro wastes using *Lactobacillus delbrueckii*. *Process Biochemistry*, 41, 759-763.
- John, R. P., Anisha, G., Nampoothiri, K. M. and Pandey, A. (2009). Direct lactic acid fermentation: focus on simultaneous saccharification and lactic acid production. *Biotechnology Advances*, 27, 145-152.
- Jönsson, L. J. and Martín, C. (2016). Pretreatment of lignocellulose: Formation of inhibitory by-products and strategies for minimizing their effects. *Bioresource Technology*, 199, 103-112.
- Kamm, B., Kamm, M., Schmidt, M., Starke, I. and Kleinpeter, E. (2006). Chemical and biochemical generation of carbohydrates from lignocellulose-feedstock

(Lupinus nootkatensis) - Quantification of glucose. *Chemosphere*, 62, 97-105.

- Kaur, K. and Phutela, U. G. (2016). Enhancement of paddy straw digestibility and biogas production by sodium hydroxide-microwave pretreatment. *Renewable Energy*, 92, 178-184.
- Kim, D. Y., Yim, S. C., Lee, P. C., Lee, W. G., Lee, S. Y. and Chang, H. N. (2004). Batch and continuous fermentation of succinic acid from wood hydrolysate by *Mannheimia succiniciproducens* MBEL55E. *Enzyme and Microbial Technology*, 35, 648-653.
- Kim, J., Yun, S. and Ounaies, Z. (2006). Discovery of cellulose as a smart material. *Macromolecules*, 39, 4202-4206.
- Kim, S., Park, J. M., Seo, J.-W. and Kim, C. H. (2012). Sequential acid-/alkalipretreatment of empty palm fruit bunch fiber. *Bioresource Technology*, 109, 229-233.
- Krishna, S. H., Sattur, A. and Karanth, N. (2001). Lipae-catalyzed synthesis of isoamyl isobutyrate—optimization using a central composite rotatable design. *Process Biochemistry*, 37, 9-16.
- Kuittinen, S., Rodriguez, Y. P., Yang, M., Keinänen, M., Pastinen, O., Siika-Aho,
 M., Turunen, O. and Pappinen, A. (2016). Effect of Microwave-Assisted
 Pretreatment Conditions on Hemicellulose Conversion and Enzymatic
 Hydrolysis of Norway Spruce. *BioEnergy Research*, 9, 344-354.
- Kumar, P., Barrett, D. M., Delwiche, M. J. and Stroeve, P. (2009). Methods for Pretreatment of Lignocellulosic Biomass for Efficient Hydrolysis and Biofuel Production. *Industrial & Engineering Chemistry Research*, 48, 3713-3729.
- Kumar, R., Singh, S. and Singh, O. V. (2008). Bioconversion of lignocellulosic biomass: Biochemical and molecular perspectives. *Journal of Industrial Microbiology and Biotechnology*, 35, 377-391.
- Lai, L. W., Idris, A. (2016). Comparison of steam-alkali-chemical and microwavealkali pretreatment for enhancing the enzymatic saccharification of oil palm trunk. *Renewable Energy* 99, 738–746
- Lai, L.-W. and Idris, A. (2013). Disruption of oil palm trunks and fronds by microwave-alkali pretreatment. *BioResources*, 8, 2792-2804.
- Lee, J. (1997). Biological conversion of lignocellulosic biomass to ethanol. *Journal* of Biotechnology, 56, 1-24.

- Lee, P., Lee, S., Hong, S. and Chang, H. (2002). Isolation and characterization of a new succinic acid-producing bacterium, *Mannheimia succiniciproducens* MBEL55E, from bovine rumen. *Applied Microbiology and Biotechnology*, 58, 663-668.
- Lee, P., Lee, S., Hong, S. and Chang, H. N. (2003a). Batch and continuous cultures of *Mannheimia succiniciproducens* MBEL55E for the production of succinic acid from whey and corn steep liquor. *Bioprocess and Biosystems Engineering*, 26, 63-67.
- Lee, P. C., Lee, W. G., Kwon, S., Lee, S. Y. and Chang, H. N. (1999). Succinic acid production by *Anaerobiospirillum succiniciproducens*: effects of the H₂/CO₂ supply and glucose concentration. *Enzyme and Microbial Technology*, 24, 549-554.
- Lee, P. C., Lee, W. G., Lee, S. Y. and Chang, H. N. (2001). Succinic acid production with reduced by-product formation in the fermentation of *Anaerobiospirillum* succiniciproducens using glycerol as a carbon source. *Biotechnology and Bioengineering*, 72, 41-48.
- Lee, P. C., Lee, S. Y., Hong, S. H. and Chang, H. N. (2003b). Batch and continuous cultures of Mannheimia succiniciproducens MBEL55E for the production of succinic acid from whey and corn steep liquor. *Bioprocess and Biosystems Engineering*, 26, 63-67.
- Lee, P. C., Lee, S. Y., Hong, S. H., Chang, H. N. and Park, S. C. (2003c). Biological conversion of wood hydrolysate to succinic acid by *Anaerobiospirillum* succiniciproducens. Biotechnology Letters, 25, 111-114.
- Lee, P. C., Lee, S. Y. and Chang, H. N. (2008). Succinic acid production by Anaerobiospirillum succiniciproducens ATCC 29305 growing on galactose, galactose/glucose, and galactose/lactose. *Journal of Microbiology and Biotechnology*, 18, 1792-1796.
- Lee, S. J., Song, H. and Lee, S. Y. (2006). Genome-based metabolic engineering of Mannheimia succiniciproducens for succinic acid production. *Applied and Environmental Microbiology*, 72, 1939-1948.
- Li, H., Kim, N.-J., Jiang, M., Kang, J. W. and Chang, H. N. (2009). Simultaneous saccharification and fermentation of lignocellulosic residues pretreated with phosphoric acid–acetone for bioethanol production. *Bioresource Technology*, 100, 3245-3251.

- Li, H., Qu, Y., Yang, Y., Chang, S. and Xu, J. (2016). Microwave irradiation–A green and efficient way to pretreat biomass. *Bioresource Technology*, 199, 34-41.
- Li, J., Zheng, X. Y., Fang, X. J., Liu, S. W., Chen, K. Q., Jiang, M., Wei, P. and Ouyang, P. K. (2011). A complete industrial system for economical succinic acid production by *Actinobacillus succinogenes*. *Bioresource Technology*, 102, 6147-6152.
- Li, Q., Siles, J. A. and Thompson, I. P. (2010a). Succinic acid production from orange peel and wheat straw by batch fermentations of Fibrobacter succinogenes S85. *Appl Microbiol Biotechnol*, 88, 671-678.
- Li, Q., Yang, M., Wang, D., Li, W., Wu, Y., Zhang, Y., Xing, J. and Su, Z. (2010b). Efficient conversion of crop stalk wastes into succinic acid production by Actinobacillus succinogenes. *Bioresource Technology*, 101, 3292-3294.
- Lim, S. and Teong, L. K. (2010). Recent trends, opportunities and challenges of biodiesel in Malaysia: an overview. *Renewable and Sustainable Energy Reviews*, 14, 938-954.
- Lin, H., Bennett, G. N. and San, K.-Y. (2005). Metabolic engineering of aerobic succinate production systems in Escherichia coli to improve process productivity and achieve the maximum theoretical succinate yield. *Metabolic engineering*, 7, 116-127.
- Lindsey, T. C. (2010). Conversion of Existing Dry-Mill Ethanol Operations to Biorefineries. *Biofuels from Agricultural Wastes and Byproducts*, 161.
- Liu, J., Wang, Q., Wang, S., Zou, D. and Sonomoto, K. (2012). Utilisation of microwave-NaOH pretreatment technology to improve performance and Llactic acid yield from vinasse. *Biosystems Engineering*, 112, 6-13.
- Liu, R.-S. and Tang, Y.-J. (2010). Tuber melanosporum fermentation medium optimization by Plackett–Burman design coupled with Draper–Lin small composite design and desirability function. *Bioresour Technol*, 101, 3139-3146.
- Liu, Y.-P., Zheng, P., Sun, Z.-H., Ni, Y., Dong, J.-J. and Zhu, L.-L. (2008a). Economical succinic acid production from cane molasses by Actinobacillus succinogenes. *Bioresour Technol*, 99, 1736-1742.
- Liu, Y. P., Zheng, P., Sun, Z. H., Ni, Y., Dong, J. J. and Wei, P. (2008b). Strategies of pH control and glucose-fed batch fermentation for production of succinic

acid by Actinobacillus succinogenes CGMCC1593. Journal of chemical Technology and Biotechnology, 83, 722-729.

- Liu, Z. and Fei, B. (2013). Characteristics of Moso Bamboo with Chemical Pretreatment. *Edited by Anuj K. Chandel and Silvio Silvério da Silva*, 1.
- Loow, Y.-L., Wu, T. Y., Jahim, J. M., Mohammad, A. W. and Teoh, W. H. (2016).Typical conversion of lignocellulosic biomass into reducing sugars using dilute acid hydrolysis and alkaline pretreatment. *Cellulose*, 23, 1491-1520.
- Luo, L., Van Der Voet, E. and Huppes, G. (2010). Biorefining of lignocellulosic feedstock – Technical, economic and environmental considerations. *Bioresource Technology*, 101, 5023-5032.
- Lynd, L. R., Van Zyl, W. H., Mcbride, J. E. and Laser, M. (2005). Consolidated bioprocessing of cellulosic biomass: an update. *Curr Opin Biotechnol*, 16, 577-583.
- Ma, J. F., Jiang, M., Chen, K. Q., Xu, B., Liu, S. W., Wei, P., Ying, H. J., Chang, H. N. and Ouyang, P. K. (2011). Strategies for efficient repetitive production of succinate using metabolically engineered Escherichia coli. *Bioprocess and Biosystems Engineering*, 34, 411-418.
- Markowitz, V. M., Ivanova, N. N., Szeto, E., Palaniappan, K., Chu, K., Dalevi, D., Chen, I.-M. A., Grechkin, Y., Dubchak, I. and Anderson, I. (2008). IMG/M: a data management and analysis system for metagenomes. *Nucleic acids research*, 36, D534-D538.
- Mckinlay, J. B., Zeikus, J. G. and Vieille, C. (2005). Insights into Actinobacillus succinogenes fermentative metabolism in a chemically defined growth medium. *Applied Environmental Microbiology*, 71, 6651-6656.
- Mckinlay, J. B., Vieille, C. and Zeikus, J. G. (2007). Prospects for a bio-based succinate industry. *Applied Microbiology and Biotechnology*, 76, 727-740.
- Mckinlay, J. B. and Vieille, C. (2008). 13 C-metabolic flux analysis of Actinobacillus succinogenes fermentative metabolism at different NaHCO 3 and H 2 concentrations. *Metabolic Engineering*, 10, 55-68.
- Merino, S. T. and Cherry, J. (2007). Progress and challenges in enzyme development for biomass utilization. *Biofuels*. Springer.
- Michael, L. S. and Kargi, F. 1992. Bioprocess engineering: basic concepts. Prentice-Hall, Englewood Cliffs, NJ.

- Millati, R., Syamsiah, S., Niklasson, C., Nur Cahyanto, M., Lundquist, K. and Taherzadeh, M. J. (2011). Biological pretreatment of lignocelluloses with white-rot fungi and its applications: a review. *BioResources*, 6.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical chemistry*, 31, 426-428.
- Miyamoto, K. (1997). *Renewable biological systems for alternative sustainable energy production*, Food & Agriculture Org.
- Montgomery, D. C. (2008). Design and analysis of experiments, John Wiley & Sons.
- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y., Holtzapple, M. and Ladisch,
 M. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology*, 96, 673-686.
- Myers, R. H. (1999). Response surface methodology--current status and future directions. *Journal of Quality Technology*, 31, 30.
- Nghiem, N. P. (1997). Production of succinic acid by anaerobiospirillum succiniciproducens. *Applied Biochemistry and Biotechnology Part A Enzyme Engineering and Biotechnology*, 63-65, 565-576.
- Nghiem, N. P., Davison, B. H., Suttle, B. E. and Richardson, G. R. (1997). Production of succinic acid by Anaerobiospirillum succiniciproducens. *Applied biochemistry and biotechnology*, 63, 565-576.
- Nidetzky, B., Steiner, W., Hayn, M. and Claeyssens, M. (1994). Cellulose hydrolysis by the cellulases from Trichoderma reesei: a new model for synergistic interaction. *Biochemical Journal*, 298, 705-710.
- Oh, I. J., Lee, H. W., Park, C. H., Lee, S. Y. and Lee, J. (2008). Succinic acid production by continuous fermentation process using Mannheimia succiniciproducens LPK7.
- Oh, I. J., Kim, D. H., Oh, E. K., Lee, S. Y. and Lee, J. (2009). Optimization and scale-up of succinic acid production by Mannheimia succiniciproducens LPK7.
- Okino, S., Inui, M. and Yukawa, H. (2005). Production of organic acids by Corynebacterium glutamicum under oxygen deprivation. *Applied Microbiology and Biotechnology*, 68, 475-480.
- Okino, S., Noburyu, R., Suda, M., Jojima, T., Inui, M. and Yukawa, H. (2008a). An efficient succinic acid production process in a metabolically engineered

Corynebacterium glutamicum strain. *Applied Microbiology and Biotechnology*, 81, 459-464.

- Okino, S., Noburyu, R., Suda, M., Jojima, T., Inui, M. and Yukawa, H. (2008b). An efficient succinic acid production process in a metabolically engineered Corynebacterium glutamicum strain. *Applied Microbiology and Biotechnology*, 81, 459-464.
- Oosterveer, P. (2015). Promoting sustainable palm oil: viewed from a global networks and flows perspective. *Journal of Cleaner Production*, 107, 146-153.
- Ortega, N., Busto, D. and Perez-Mateos, M. (2001). Kinetics of cellulose saccharification by Trichoderma reesei cellulases. *International biodeterioration & biodegradation*, 47, 7-14.
- Paiva, M., Ammar, I., Campos, A., Cheikh, R. B. and Cunha, A. (2007). Alfa fibers: Mechanical, morphological and interfacial characterization. *Composites science and technology*, 67, 1132-1138.
- Pang, F., Xue, S., Yu, S., Zhang, C., Li, B. and Kang, Y. (2012). Effects of microwave power and microwave irradiation time on pretreatment efficiency and characteristics of corn stover using combination of steam explosion and microwave irradiation (SE–MI) pretreatment. *Bioresource technology*, 118, 111-119.
- Park, D., Laivenieks, M., Guettler, M., Jain, M. and Zeikus, J. (1999). Microbial utilization of electrically reduced neutral red as the sole electron donor for growth and metabolite production. *Applied Environmental Microbiology*, 65, 2912-2917.
- Park, E. Y., Anh, P. N. and Okuda, N. (2004). Bioconversion of waste office paper to 1 (+)-lactic acid by the filamentous fungus< i> Rhizopus oryzae</i>. *Bioresource technology*, 93, 77-83.
- Perlack, R. D., Wright, L. L., Turhollow, A. F., Graham, R. L., Stokes, B. J. and Erbach, D. C. 2005. Biomass as feedstock for a bioenergy and bioproducts industry: the technical feasibility of a billion-ton annual supply. DTIC Document.
- Pinazo, J. M., Domine, M. E., Parvulescu, V. and Petru, F. (2015). Sustainability metrics for succinic acid production: A comparison between biomass-based and petrochemical routes. *Catalysis Today*, 239, 17-24.

- Prasad, S., Singh, A. and Joshi, H. (2007). Ethanol as an alternative fuel from agricultural, industrial and urban residues. *Resources, Conservation and Recycling*, 50, 1-39.
- Raab, A. M., Gebhardt, G., Bolotina, N., Weuster-Botz, D. and Lang, C. (2010). Metabolic engineering of Saccharomyces cerevisiae for the biotechnological production of succinic acid. *Metabolic Engineering*, 12, 518-525.
- Ramos, L., Breuil, C. and Saddler, J. (1992). Comparison of steam pretreatment of eucalyptus, aspen, and spruce wood chips and their enzymatic hydrolysis. *Applied biochemistry and biotechnology*, 34, 37-48.
- Ramos, L. P. (2003). The chemistry involved in the steam treatment of lignocellulosic materials. *Química Nova*, 26, 863-871.
- Ranaldi, F., Vanni, P. and Giachetti, E. (1999). What Students Must Know About the Determination of Enzyme Kinetics Parameters. *Biochemical Education*, 27, 87-91.
- Rivas, B., Moldes, A. B., DomíNguez, J. M. and Parajó, J. C. (2004). Lactic acid production from corn cobs by simultaneous saccharification and fermentation: a mathematical interpretation. *Enzyme and Microbial Technology*, 34, 627-634.
- Rodríguez-Zúñiga, U. F., Cannella, D., De Campos Giordano, R., Giordano, R. D. L. C., Jørgensen, H. and Felby, C. (2015). Lignocellulose pretreatment technologies affect the level of enzymatic cellulose oxidation by LPMO. *Green Chemistry*, 17, 2896-2903.
- Samuelov, N. S., Datta, R., Jain, M. K. and Zeikus, J. G. (1999a). Whey fermentation by Anaerobiospirillum succiniciproducens for production of a succinatebased animal feed additive. *Applied Environmental Microbiology*, 65, 2260-2263.
- Sánchez, A. M., Bennett, G. N. and San, K.-Y. (2005). Novel pathway engineering design of the anaerobic central metabolic pathway in Escherichia coli to increase succinate yield and productivity. *Metabolic Engineering*, 7, 229-239.
- Saritha, M. and Arora, A. (2012). Biological pretreatment of lignocellulosic substrates for enhanced delignification and enzymatic digestibility. *Indian journal of microbiology*, 52, 122-130.
- Shen, Y., Zhang, Y., Ma, T., Bao, X., Du, F., Zhuang, G. and Qu, Y. (2008). Simultaneous saccharification and fermentation of acid-pretreated corncobs

with a recombinant Saccharomyces cerevisiae expressing β -glucosidase. *Bioresource Technology*, 99, 5099-5103.

- Shujun, W., Jinglin, Y., Jiugao, Y., Haixia, C. and Jiping, P. (2007). The effect of acid hydrolysis on morphological and crystalline properties of Rhizoma Dioscorea starch. *Food Hydrocolloids*, 21, 1217-1222.
- Sluiter, J. B., Ruiz, R. O., Scarlata, C. J., Sluiter, A. D. and Templeton, D. W. (2010). Compositional analysis of lignocellulosic feedstocks. 1. Review and description of methods. *Journal of Agricultural and Food Chemistry*, 58, 9043-9053.
- Song, H. and Lee, S. Y. (2006). Production of succinic acid by bacterial fermentation. *Enzyme and Microbial Technology*, 39, 352-361.
- Song, H., Huh, Y. S., Lee, S. Y., Hong, W. H. and Hong, Y. K. (2007). Recovery of succinic acid produced by fermentation of a metabolically engineered Mannheimia succiniciproducens strain. *Journal of Biotechnology*, 132, 445-452.
- Stols, L. and Donnelly, M. I. (1997). Production of succinic acid through overexpression of NAD (+)-dependent malic enzyme in an Escherichia coli mutant. *Applied and environmental microbiology*, 63, 2695-2701.
- Suhaimi, M. and Ong, H. (2001). Composting empty fruit bunches of oil palm. *Extension Bulletin-Food & Fertilizer Technology Center*, 1-8.
- Sun, Y. and Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource Technology*, 83, 1-11.
- Tan, H. T. and Lee, K. T. (2012). Understanding the impact of ionic liquid pretreatment on biomass and enzymatic hydrolysis. *Chemical Engineering Journal*, 183, 448-458.
- Thostenson, E. and Chou, T.-W. (1999). Microwave processing: fundamentals and applications. *Composites Part A: Applied Science and Manufacturing*, 30, 1055-1071.
- Tokuhiro, K., Ishida, N., Kondo, A. and Takahashi, H. (2008). Lactic fermentation of *cellobiase* by a yeast strain displaying β-glucosidase on the cell surface. *Applied Microbiology and Biotechnology*, 79, 481-488.
- Van Der Werf, M. J., Guettler, M. V., Jain, M. K. and Zeikus, J. G. (1997). Environmental and physiological factors affecting the succinate product ratio

during carbohydrate fermentation by Actinobacillus sp. 130Z. Archives of Microbiology, 167, 332-342.

- Van Dyk, J. and Pletschke, B. (2012). A review of lignocellulose bioconversion using enzymatic hydrolysis and synergistic cooperation between enzymes factors affecting enzymes, conversion and synergy. *Biotechnology Advances*, 30, 1458-1480.
- Van Gylswyk, N. (1995). Succiniclasticum ruminis gen. nov., sp. nov., a ruminal bacterium converting succinate to propionate as the sole energy-yielding mechanism. *International Journal of Systematic and Evolutionary Microbiology*, 45, 297-300.
- Vemuri, G., Eiteman, M. and Altman, E. (2002a). Succinate production in dual-phase Escherichia coli fermentations depends on the time of transition from aerobic to anaerobic conditions. *Journal of Industrial Microbiology and Biotechnology*, 28, 325-332.
- Vemuri, G., Eiteman, M. and Altman, E. (2002b). Effects of growth mode and pyruvate carboxylase on succinic acid production by metabolically engineered strains of Escherichia coli. *Applied and Environmental Microbiology*, 68, 1715-1727.
- Wang, D., Li, Q., Yang, M., Zhang, Y., Su, Z. and Xing, J. (2011). Efficient production of succinic acid from corn stalk hydrolysates by a recombinant Escherichia coli with ptsG mutation. *Process Biochemistry*, 46, 365-371.
- Wang, L., Han, G. and Zhang, Y. (2007). Comparative study of composition, structure and properties of Apocynum venetum fibers under different pretreatments. *Carbohydrate Polymers*, 69, 391-397.
- Wee, Y.-J., Kim, J.-N. and Ryu, H.-W. (2006). Biotechnological production of lactic acid and its recent applications. *Food Technology and Biotechnology*, 44, 163-172.
- Weerasai, K., Suriyachai, N., Poonsrisawat, A., Arnthong, J., Unrean, P., Laosiripojana, N. and Champreda, V. (2014). Sequential acid and alkaline pretreatment of rice straw for bioethanol fermentation. *BioResources*, 9, 5988-6001.
- Willke, T. and Vorlop, K.-D. (2004). Industrial bioconversion of renewable resources as an alternative to conventional chemistry. *Applied Microbiology* and Biotechnology, 66, 131-142.

- Wu, H., Li, Z.-M., Zhou, L. and Ye, Q. (2007). Improved succinic acid production in the anaerobic culture of an Escherichia coli pflB ldhA double mutant as a result of enhanced anaplerotic activities in the preceding aerobic culture. *Appilied and Environmental Microbiology*, 73, 7837-7843.
- Wu, T. Y., Mohammad, A. W., Jahim, J. M. and Anuar, N. (2010). Pollution control technologies for the treatment of palm oil mill effluent (POME) through endof-pipe processes. *Journal of Environmental Management*, 91, 1467-1490.
- Xu, Q., Singh, A. and Himmel, M. E. (2009). Perspectives and new directions for the production of bioethanol using consolidated bioprocessing of lignocellulose. *Current Opinion in Biotechnology*, 20, 364-371.
- Xu, Z., Wang, Q., Jiang, Z., Yang, X.-X. and Ji, Y. (2007). Enzymatic hydrolysis of pretreated soybean straw. *Biomass and Bioenergy*, 31, 162-167.
- Yang, B., Dai, Z., Ding, S.-Y. and Wyman, C. E. (2011). Enzymatic hydrolysis of cellulosic biomass. *Biofuels*, 2, 421-449.
- Yusof, B. and Yew, F. (2009). Potential of palm oil for developing countries and role in the food and fuel debate. *Global Oils Fats Bus Mag (Pullout)*, 6, 1-8.
- Zakaria, M. R., Hirata, S. and Hassan, M. A. (2015). Hydrothermal pretreatment enhanced enzymatic hydrolysis and glucose production from oil palm biomass. *Bioresource Technology*, 176, 142-148.
- Zeikus, J., Jain, M. and Elankovan, P. (1999). Biotechnology of succinic acid production and markets for derived industrial products. *Applied Microbiology* and Biotechnology, 51, 545-552.
- Zeitsch, K. J. (2000). The chemistry and technology of furfural and its many byproducts, Elsevier.
- Zhang, Z. Y., Jin, B. and Kelly, J. M. (2007). Production of lactic acid from renewable materials by Rhizopus fungi. *Biochemical Engineering Journal*, 35, 251-263.
- Zhao, C., Shao, Q., Ma, Z., Li, B. and Zhao, X. (2016). Physical and chemical characterizations of corn stalk resulting from hydrogen peroxide presoaking prior to ammonia fiber expansion pretreatment. *Industrial Crops and Products*, 83, 86-93.
- Zheng, P., Dong, J.-J., Sun, Z.-H., Ni, Y. and Fang, L. (2009). Fermentative production of succinic acid from straw hydrolysate by *Actinobacillus succinogenes*. *Bioresource Technology*, 100, 2425-2429.

- Zheng, P., Fang, L., Xu, Y., Dong, J.-J., Ni, Y. and Sun, Z.-H. (2010). Succinic acid production from corn stover by simultaneous saccharification and fermentation using *Actinobacillus succinogenes*. *Bioresource Technology*, 101, 7889-7894.
- Zhu, L.-W., Wang, C.-C., Liu, R.-S., Li, H.-M., Wan, D.-J. and Tang, Y.-J. (2012). Actinobacillus succinogenes ATCC 55618 fermentation medium optimization for the production of succinic acid by response surface methodology. *BioMed Research International*, 2012.
- Zhu, S., Wu, Y., Yu, Z., Liao, J. and Zhang, Y. (2005). Pretreatment by microwave/alkali of rice straw and its enzymic hydrolysis. *Process Biochemistry*, 40, 3082-3086.
- Zhu, S., Wu, Y., Yu, Z., Wang, C., Yu, F., Jin, S., Ding, Y., Chi, R., Liao, J. and Zhang, Y. (2006). Comparison of three microwave/chemical pretreatment processes for enzymatic hydrolysis of rice straw. *Biosystems Engineering*, 93, 279-283.
- Zhu, Z., Simister, R., Bird, S., Mcqueen-Mason, S. J., Gomez, L. D. and Macquarrie,D. J. (2015). Microwave assisted acid and alkali pretreatment of Miscanthus biomass for biorefineries. *matrix*, 10, 16.
- Zhu, Z., Rezende, C. A., Simister, R., Mcqueen-Mason, S. J., Macquarrie, D. J., Polikarpov, I. and Gomez, L. D. (2016). Efficient sugar production from sugarcane bagasse by microwave assisted acid and alkali pretreatment. *Biomass and Bioenergy*, 93, 269-278.
- Zou, W., Zhu, L.-W., Li, H.-M. and Tang, Y.-J. (2011). Significance of CO₂ donor on the production of succinic acid by *Actinobacillus succinogenes* ATCC 55618. *Microbial Cell Factories*, 10, 1.