

**SYNTHESIS AND CHARACTERIZATION OF HYPERTHERMIA INDUCED  
SPIONS-PEG-HER IN TARGETED BREAST CANCER**

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A thesis submitted in fulfillment of the  
requirements for the award of the degree of  
Doctor of Philosophy (Bioprocess Engineering)

Faculty of Chemical and Energy Engineering  
Universiti Teknologi Malaysia

MAY 2017

*Dedicated to*

*My beloved parents and siblings, my main supportive supervisor Prof. Dr. Ani Idris  
and also my helpful co-supervisor Prof. Dr. Fadzilah Adibah Abdul Majid*

*Thank you very much for being supportive, helpful and understanding*

## ACKNOWLEDGEMENT

The completion of this undertaking could not have been possible without the participation and assistance of so many people whose names may not all be enumerated. Their contributions are sincerely appreciated and gratefully acknowledged. However I would like to express my deep appreciation and indebtedness particularly to the following:

Prof. Ani Idris for her endless support, kind and understanding spirit as my main supervisor.

Prof. Fadzilah Adibah Abdul Majid for her invaluable assistance and understanding as my co-supervisor.

Prof. Mojtaba Salouti, my technical co-supervisor from Islamic Azad University, Zanjan branch, Iran, whose valuable guidance has always been there for me throughout my entire research.

Dr. Rozita Nasiri for her unforgettable assistance throughout all the experiments as my superb colleague.

To all relatives, friends and others who in one way or another shared their support morally, financially or physically, thank you.

Above all, to the Great Almighty, the author of knowledge and wisdom, for his countless love.

I thank you.

## ABSTRACT

Hyperthermia has opened up key avenues in cancer therapy. Nevertheless, engineering a smart and efficient tumor targeting superparamagnetic fluid agent capable of elevating the temperature of targeted sites as well as exposing a safe level of biocompatibility remain remarkably demanding and challenging. In this study, a novel core-shell tumor-targeting superparamagnetic iron oxide nanoparticle-polyethylene glycol-Herceptin (SPIONs-PEG-HER) was developed and evaluated for an efficient hyperthermia treatment of HER2+ breast cancer using an alternating magnetic field (AMF). Both *in vitro* and *in vivo* treatment models using four different cell lines and 7, 12 dimethylbenz ( $\alpha$ ) anthracene (DMBA)-induced balb/c mice were developed, respectively. SPIONs modification was carried out by PEGylation to provide biocompatibility and conjugation of Herceptin to add the tumor-targeting features to the SPIONs. The morphological characterization and physico-chemical analyses of SPIONs-PEG-HER, were carried out using transmission electron microscopy (TEM), field emission scanning electron microscopy, dynamic light scattering (DLS), x-ray diffraction (XRD), Fourier transform infrared spectroscopy (FT-IR), vibrating sample magnetometer (VSM) and specific absorption rate (SAR). The *in vitro* biocompatibility confirmation of SPIONs-PEG-HER was performed using tetrazolium dye (MTT) assay, Trypan blue staining method and blood compatibility tests. The ability of SPIONs-PEG-HER binding to HER2+ cell line (SK-BR-3) was measured comparatively to HER2- cell lines (HSF 1184, MDA-MB-231, MDA-MB-468) via *in vitro* binding studies. The qualitative and quantitative data were obtained using Prussian blue staining, TEM assay and atomic absorption spectroscopy (AAS) analysis. The comparison on the changes in growth inhibition rates in HSF 1184, MDA-MB-231, MDA-MB-468 and SK-BR-3 cell lines at different temperatures (40 °C, 42 °C, and 45 °C) were studied using a post-hyperthermia MTT assay. Then, post-hyperthermia morphological investigations were performed via inverted microscope and acridine orange/ethidium bromide (AO/EB) staining method. The *in vivo* model using DMBA-induced balb/c mice were injected with SPIONs-PEG-HER via tail vein. The biodistribution of SPIONs-PEG-HER at tumor site as well as in vital organs were qualitatively and quantitatively measured using colorimetric methods (AAS and TEM). Finally, the survival rate of the DMBA-induced balb/c mice injected with SPIONs-PEG-HER was measured in the presence and absence of AMF and the daily monitoring of the weight of the treated DMBA-induced balb/c mice during the treatment period was carried out. The morphological characterization and physico-chemical analyses revealed that the synthesized SPIONs-PEG-HER had a size of almost 17 nm and possessed a nearly spherical appearance as well as greater hydrodynamic diameter (~84 nm) and a wider distribution compared to the bare SPIONs. Moreover, XRD and FT-IR analyses confirmed that the processes of PEGylation and conjugation were successfully accomplished while the VSM and SAR analyses showed that the SPIONs-PEG-HER possess efficient magnetic properties to be used as a hyperthermia fluid agent. SPIONs-PEG-HER showed relatively low levels of toxicity even at the extremely high concentration of 1000  $\mu\text{g/ml}$ . The results of the binding studies indicated that the SPIONs-PEG-HER could selectively bind to the HER2+ cancer cells. The results obtained from the post-hyperthermia MTT assay indicated that exposing the HER2+ cells to the temperature of 45 °C for 20 minutes inhibited the growth of the cells by 90% where they did not regain their normal proliferation like the HER2- cell lines which corroborates the results obtained through post-hyperthermia morphological analyses where it was shown that the temperature of 45 °C induced significant apoptosis compared to the other temperatures. Through morphological alteration studies by inverted microscope and AO/EB staining method, it was disclosed that the SK-BR-3 cells had undergone apoptosis since apoptotic signs such as shrunk cells as well as apoptotic bodies were obviously seen. The results of biodistribution studies showed significantly higher accumulation of the SPIONs-PEG-HER in the tumor site compared to SPIONs-PEG. Based on the results obtained through hyperthermia treatment of DMBA-induced balb/c mice, it was revealed that the survival rate in the experimental group treated with SPIONs-PEG-HER in the presence of AMF was much higher than other experimental groups where 50% of the DMBA-induced balb/c mice survived and maintained their average body weight. Findings in this study illustrated that SPIONs-PEG-HER-mediated hyperthermia is a potent breast cancer treatment.

## ABSTRAK

Hipertermia telah membuka ruang utama dalam terapi kanser. Walau bagaimanapun, pembentukan tumor yang pantas dan berkesan menyasarkan ejen cecair super paramagnet yang mampu meningkatkan suhu asas yang disasarkan serta mendedahkan tahap selamat kesesuaian-bio agar kekal sebagai sesuatu yang amat mencabar. Dalam kajian ini, rangkaian teras-kelompok tumor mensasarkan partikel nano ferum oksida super paramagnet-polietilena-*Herceptin* (SPIONs-PEG-HER) telah dibangunkan dan dinilai untuk rawatan *hipertermia* HER2+ kanser payudara dengan menggunakan medan magnet selang-seli (AMF). Kedua-dua *in vitro* dan model rawatan *in vivo* menggunakan 4 baris sel yang berbeza dan 7, 12 dimetilbenz ( $\alpha$ ) antrasena (DMBA) yang disebabkan tetikus *balb/c* telah dibangunkan. Pengubahsuaian SPIONs telah dijalankan oleh *PEGylation* untuk menyediakan kesesuaian-bio dan hubungan antara HER untuk menambah ciri-ciri tumor yang disasarkan ke atas SPIONs. Pencirian morfologi dan analisis fiziko-kimia SPIONs-PEG-HER telah dijalankan dengan menggunakan mikroskopi pancaran elektron (TEM), mikroskop elektron pengimbas pancaran medan, penyerakan cahaya dinamik (DLS), pembelauan sinar-x (XRD), spektroskopi inframerah transformasi Fourier (FT-IR), magnetometer sampel bergetar (VSM) dan kadar penyerapan tertentu (SAR). Pengesahan kesesuaian-bio *in vitro* daripada SPIONs-PEG-HER dilakukan dengan menggunakan asai pewarna tetrazolium (MTT), kaedah pewarnaan biru *Trypan* dan ujian keserasian darah. Keupayaan SPIONs-PEG-HER mengikat sel garisan HER2+ (SK-BR-3) telah diukur secara perbandingan dengan rangkaian sel HER2- (HSF 1184, MDA-MB-231, MDA-MB-468) melalui kajian ikatan *in vitro*. Data kualitatif dan kuantitatif telah diperolehi dengan menggunakan pewarnaan biru *Prussian*, asai TEM dan analisis spektroskopi penyerapan atom (AAS). Perbandingan terhadap perubahan dalam kadar pertumbuhan perencatan dalam HSF 1184, MDA-MB-231, MDA-MB-468 dan SK-BR-3 baris sel pada suhu yang berbeza (40 °C, 42 °C dan 45 °C) dikaji dengan menggunakan asai MTT pasca hipertermia. Setelah itu, kajian morfologi pasca hipertermia telah dijalankan melalui mikroskop songsang dan kaedah pewarnaan oren akridina/etidium bromida (AO/EB). Model *in vivo* menggunakan DMBA yang disebabkan oleh tetikus *balb/c* telah disuntik dengan SPIONs-PEG-HER melalui hujung salur darah. Taburan-bio daripada SPIONs-PEG-HER ke atas tumor asas dan juga di organ-organ penting telah diukur secara kualitatif dan kuantitatif menggunakan kaedah kolorimetri (AAS dan TEM). Akhir sekali, kadar hayat DMBA yang disebabkan oleh tetikus *balb/c* disuntik dengan SPIONs-PEG-HER telah diukur dengan kehadiran dan ketiadaan AMF dan pemantauan harian ke atas berat DMBA yang disebabkan oleh tetikus *balb/c* dirawat semasa tempoh rawatan telah dijalankan. Analisis pencirian morfologi dan fiziko-kimia mendedahkan bahawa SPIONs-PEG-HER yang telah disintesis mempunyai saiz hampir 17 nm dan mempunyai penampilan yang hampir setara serta diameter hidrodinamik yang lebih besar (~ 84 nm) dan pengedaran yang lebih luas berbanding dengan pendedahan SPIONs. Selain itu, analisis XRD dan FT-IR mengesahkan bahawa proses *PEGylation* dan hubungan antaranya telah berjaya dicapai manakala analisis VSM dan SAR menunjukkan bahawa SPIONs-PEG-HER memiliki sifat-sifat magnet berkesan untuk digunakan sebagai ejen cecair hipertermia. SPIONs-PEG-HER menunjukkan tahap ketoksikan yang rendah walaupun pada kepekatan yang sangat tinggi 1000  $\mu\text{g/ml}$ . Keputusan kajian menunjukkan bahawa SPIONs-PEG-HER terpilih boleh mengikat kepada HER2+ sel-sel kanser. Keputusan yang diperolehi dari asai MTT pasca hipertermia menunjukkan bahawa pendedahan HER2+ sel-sel dengan suhu 45 °C selama 20 minit menghalang pertumbuhan sel-sel sebanyak 90% di mana mereka tidak mendapatkan semula percambahan normal mereka seperti bahagian-bahagian sel HER2- yang menguatkan keputusan yang diperolehi melalui analisis pasca hipertermia morfologi di mana ia telah menunjukkan bahawa suhu 45 °C secara signifikan amat ketara berbanding dengan suhu lain. Melalui kajian perubahan morfologi oleh mikroskop songsang dan kaedah pewarnaan AO/EB, telah dinyatakan bahawa sel-sel SK-BR-3 telah menjalani apoptosis sejak tanda-tanda apoptotik seperti kemerosotan sel-sel serta badan-badan apoptotik telah jelas dilihat. Dapatan kajian taburan-bio mendedahkan bahawa pengumpulan SPIONs-PEG-HER di dalam laman tumor jauh lebih tinggi berbanding SPIONs-PEG. Berdasarkan keputusan yang diperolehi melalui rawatan hipertermia daripada DMBA yang disebabkan oleh tetikus *balb/c*, telah mendedahkan bahawa kadar hayat dalam kumpulan eksperimen yang dirawat dengan SPIONs-PEG-HER di hadapan AMF adalah lebih tinggi daripada kumpulan eksperimen lain di mana 50% daripada DMBA yang disebabkan oleh tetikus *balb/c* terselamat dan mengekalkan berat badan purata mereka. Penemuan dalam kajian ini menggambarkan bahawa hipertermia SPIONs-PEG-HER-pengantara adalah rawatan kanser payudara yang mujarab..

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**LIST OF ABBREVIATIONS**

AAS	-	Atomic Absorption Spectroscopy
AC	-	Alternating Current
AMF	-	Alternating Magnetic Field
AO	-	Acridine Orange
APTT	-	Activated Partial Thromboplastin Time
AR	-	Androgen Receptor
ASR	-	Age-Standardised Rate
DLS	-	Dynamic Light Scattering Analysis
DMBA	-	7,12-Dimethylbenz[a]anthracene
DNA	-	deoxyribonucleic acid
EAC	-	Ehrlich Ascites Carcinoma
EB	-	Ethidium Bromide
EDC	-	(1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride)
EDTA	-	Ethylenediaminetetraacetic Acid
EGFR	-	Epidermal Growth Factor Receptor
EPR	-	Enhanced Permeability and Retention
FA	-	Folic Acid
FB	-	Fibrin
FESEM	-	Field Emission Scanning Electron Microscope
FR $\alpha$	-	Folic Acid Receptor
FT-IR	-	Fourier Transform Infrared Spectroscopy
Hc	-	Coercive force
HER	-	Herceptin
HER2	-	Human Epidermal Growth Factor Receptor 2
HER3	-	Human Epidermal Growth Factor Receptor 3
HRTEM	-	High-Resolution Transmission Electron Microscopy

IGF-IR	-	Insulin-like Growth Factor Receptor
IR	-	Infrared Radiation
MFH	-	Magnetic Fluid Hyperthermia
MNP	-	Magnetic Nanoparticle
MOH	-	Ministry of Health
Mr	-	Remanent Magnetization
MRI	-	Magnetic Resonance Imaging
Ms	-	Saturation Magnetization
MTT	-	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium Bromide
NCR	-	National Cancer Registry
NHS	-	N-Hydroxysuccinimide
NPs	-	Nanoparticles
NSCLC	-	Non-Small Cell Lung Cancer
PARP	-	Poly(ADP-Ribose)Polymerase
PBS	-	Phosphate-buffered saline
PDI	-	Polydispersity Index
PEG	-	Polyethylene Glycol
PSMA	-	Prostate Specific Membrane Antigen
PT	-	Prothrombin Time
RBC	-	Red Blood Cell
S.E.M	-	Standard Error of the Mean
SAED	-	Selected Area Electron Diffraction Pattern
SAR	-	Specific Absorption Rate
SPIONs	-	Superparamagnetic Iron Oxide Nanoparticles
TEM	-	Transmission Electron Microscopy
TT	-	Thrombin Time
UKMAEC	-	Universiti Kebangsaan Malaysia Animal Ethics Committee
VEGF-A	-	Vascular Endothelial Growth Factor A
VEGFR	-	Vascular Endothelial Growth Factor Receptor
WHO	-	World Health Organization
XRD	-	X-Ray Diffraction

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## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of Study**

Breast cancer is the most commonly diagnosed cancer among American women and it is the second leading cause of cancer death. In 2012, an estimated 226,870 new cases of invasive breast cancer and 39,510 breast cancer deaths were expected to occur among American females. Breast cancer rates vary largely by race/ethnicity and geographic region. Historically, breast cancer was known as a disease in western countries. However, over the past two decades, breast cancer incidence and mortality rates have been growing up rapidly in economically less developed regions too. According to 2012 GLOBOCAN estimates, 52.6% of the new worldwide breast cancer cases (882,900) and 62.1 % of the breast cancer deaths in women (324,300) occurred in developing countries [1].

Among Malaysians in Peninsular Malaysia, approximately, a total 18,219 new cancer cases were diagnosed in 2007 according to a report published in February 2011 by National Cancer Registry (NCR), Malaysia. Breast the first most common cancer among population regardless of sex in Malaysia. There were 3,242 female breast cancer cases diagnosed in 2007 and reported to NCR, accounted for 18.1% of all cancer cases reported and 32.1% of all female cases [2].



Nowadays, cancer rate is increasing and it is predicted that the number of cancer patients will reach to just over 15 million until the year 2020 [3]. Breast cancer is known as the most frequently diagnosed cancer among women all over the world. Different common treatment methods including partial or radical mastectomy, chemotherapy and radiotherapy are not completely successful and induce unwanted side effects on healthy tissues in the body. So, novel treatment methods are highly required to be invented and designed [4].

“To do no harm” has always been the physician’s faith. Although, the harmful side effects of cancer treatments is known as a great paradox since chemicals or radioactive agents used in cancer treatments has the potential to adversely affect a patient’s overall health. The cancer treatment complication is due to the alterations and deviations in the function of the genes and the pathways controlling the cell cycles. It is clear to the world that acute and chronic side effect are always associated with the current cancer treatments. In addition, over the last thirty years, the rate of successful therapies for the majority of human cancers has marginally improved. Therefore, it is so clear that advances in cancer therapies are required especially the advances in the treatments during which not only a patient’s health is highly improved, but also the adverse side effects are minimized. Targeted cancer treatment has been exclusively developed to specifically affect the tumors and cancerous cells while the other surrounding tissues are spared throughout the chemotherapy procedure. Achieving this goal, the characteristics of the cancer cells must be distinguished amongst trillions of normal cells in the body so that the therapeutic agent can be specifically delivered to the cancer cells and destroy them while the surrounding tissues are remained unaffected. The idea of targeted cancer treatment has been actively followed up for many years aiming to improve the survival rate of the cancer patients. To implement such a complicated task, many advances are needed for the detection and the treatment of the cancer; and nanotechnology is considered as a key knowledge to resolve the challenging and problem of tracking and treating the cancer cell-by-cell [5].

Surgery, systemic therapy and radiation therapy are the most practiced breast cancer treatment methods. Since the natural history of breast cancer and the

understanding of the molecular biology have improved, an evolution in the breast cancer surgery has been seen over the past few decades. However, historically, aggressive nature of breast cancer surgery has always been a key issue since the breast and the surrounding tissues undergo removal [6].

Radiation therapy is practiced subsequent to the modern breast-conserving surgical treatments in order to control the cancer recurrences in the breast. Radiation is mostly practiced subsequent to the tumor removal (mastectomy) once the tumor size exceeds 5 cm or when the cancer is seen in lymph nodes. In under-developed regions, radiation therapy is mostly utilized to control the cancer symptoms and not used to cure the cancer since the majority of the patients present in the hospitals with a metastatic disease at its advanced stages. Radiation is exclusively effective to control the painful symptoms resulted from bone metastases [6].

Hormone therapy, chemotherapy and targeted biological therapies fall under category of systemic therapies. The advantages of chemotherapy depend on multiple factors: cancer size, the number of lymph nodes involved, the existence of hormone receptors, and the amount of human epidermal growth receptor 2 (HER2) protein overexpressed on the cancer cells. According to the availability of the resources, different agents are utilized. Hormone therapy like tamoxifen can be given to the patients with HER2+ breast cancer. However tamoxifen is known as an affordable treatment, sufficient pathology services are not accessible in order to monitor status of the hormone receptors in the lower-resource settings. Moreover, carrying out an appropriate tissue handling and processing is inevitable for valid hormone test results. The utilization of the HER2-targeted monoclonal antibody-based treatment herceptin together with chemotherapy has shown high effectiveness in curing HER2+ cancer, but is cost-prohibitive in most of the world. Herceptin has been considered for inclusion in the World Health Organization (WHO) Essential Medicine list, sparking a debate about how health care systems can and should balance high cost against proven curative benefits [6].

## 1.2 Problem Statement

Conventional cancer therapies including surgery, radiotherapy, and chemotherapy are not so effective in the treatment of certain cancers such as breast cancer since normal tissues surrounding the cancerous tissues are also adversely affected. Thus, a novel and effective method of treatment is required. Hyperthermia is a promising therapy for cancers such as breast cancer. Hyperthermia therapy is based on the fact that cancer cells are much more sensitive to heat than normal cells. However, the difficulty of delivering the necessary selective heating remains an important technical problem to be resolved [7].

## 1.3 Research Objectives

The aim of this study is to develop a stable and biocompatible magnetic tumor-targeting hyperthermia complex “SPIONs-PEG-HER” and assess its effectiveness via *in vitro* and *in vivo* evaluations. Thus, the objectives of the study were phrased as follow:

1. To develop SPIONs-PEG-HER targeting complex which include stabilization and vectorization of iron oxide nanoparticles ( $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>).
2. To perform physico-chemical characterization of the synthesized SPIONs-PEG-HER.
3. To investigate the effectiveness of magnetic targeted hyperthermia *in vitro*.
4. To investigate the effectiveness of targeted hyperthermia *in vivo*.

## 1.4 Scope of Research

In order to achieve the goals of this study, the scope of the research is as follows.

- i) Superparamagnetic iron oxide nanoparticles/maghemite ( $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>) was stabilized by PEG (MW 2000) in order to increase the biocompatibility, solubility and stability in aqueous solution as well as in physiological saline.
- ii) Then, HER (monoclonal anti-her2 antibody/herceptin) was conjugated to the previously coated SPIONs through EDC/NHS click chemistry method for detection of HER2/neu antigen on HER2+ breast cancer cells.
- iii) Investigation of stability level of SPIONs-PEG-HER in the blood compartment was the next step during the experimental work to assure that the synthesized complex is stable and do not aggregate when exposed to the physiological conditions.
- iv) SK-BR-3 cell line (human breast cancer cell line that over expresses HER2) was used as targeted cells while HSF-1184 cell line (human skin fibroblast cell line), MDA-MB-231 cell line (human breast cancer cell line that does not over express HER2) and MDA-MB-468 cell line (human breast cancer cell line that does not over express HER2) were used as control cells.
- v) Attachment of SPIONs-PEG-HER to HER2+ cells were investigated prior to proceed to the *in vitro* evaluation. Then, magnetic hyperthermia using SPIONs-PEG-HER was performed to investigate the treatment effects of the proposed treatment method on the nominated cells.
- vi) Finally, treatment effects of the *in vivo* magnetic tumor-targeting hyperthermia on the tumor bearing balb/c mice were investigated.

In brief, successful synthesis of SPIONs-PEG-HER and magnetic tumor-targeting hyperthermia treatment effects using SPIONs-PEG-HER as the targeting agent on HER2+ cells and tumor bearing balb/c mice were focused in this study.

### **1.5 Significance of Study**

Advances in new technologies such as molecular biomarkers and nanoparticles are considered as highly qualified tools to diagnose and effectively treat the breast cancer. During the breast cancer improvement stages, genetic mutations occur and create certain molecular effects that can be used as biomarkers. Attaching the nanoparticles to the specific biomarkers of breast cancer has the potential to follow and limit the cancer cells with higher sensitivity and selectivity [8]. There are many advantages in using the targeted iron oxide nanoparticles combined with intracellular hyperthermia as targeted treatment in chemotherapy, radiotherapy, conventional hyperthermia, immunotherapy and mastectomy [9]. These advantages are explained as the matters of significance in this study.

Possibility of treatment of metastasis is the first advantage. Metastasis is the growth of secondary malignant cells at a distance from a primary site of cancer. One of the most important obstacles in treatment of cancerous tumors is the unsuccessful treatment of metastasis. Tumor itself can be treated by mastectomy or the other treatment methods, but metastasis does not appear at first or cannot be treated by the conventional methods. In the proposed targeted treatment with iron oxide nanoparticles, monoclonal antibody against a specific antigen is attached to the nanoparticles and detects the antigen wherever it is and attaches to it. Then, after the heat is induced by iron oxide nanoparticles using magnetic hyperthermia, cancer cells and metastasis are effectively treated [4],[9].

Minor systemic side effects compared to chemotherapy, radiotherapy and conventional hyperthermia is the second advantage. In the conventional methods, radiation dose absorbed by the skin, heat absorbed by the surrounding tissues and

organs on the way towards the tumor are the issues forcing the dosage of the treatment to be highly increased. But, in the targeted treatment of breast cancer via coated nanoparticles (biocompatible and far less toxic) attached to specific monoclonal antibodies, nonspecific heating of the surrounding tissues is eliminated and the dosage of the monoclonal antibody as a drug is significantly reduced which is so cost effective and less harmful. Moreover, iron oxide nanoparticles are only accumulated in the tumor area in its metastasis so that only these areas are under the influence of magnetic field. So, the surrounding tissues remain healthy and unaffected [4],[9].

Reduction of immune system response compared to unaccompanied immunotherapy is the third advantage of the proposed treatment. In some situations, immunotherapy is practiced to treat the cancer. In this case, not only super expensive cost of treatment is imposed to the patient, but also severe response of the immune system is faced due to the injection of large amounts of antibody. Normally immune system should be suppressed in this case but the amount of antibody used in the proposed treatment is highly reduced compared to the unaccompanied immunotherapy. So, this problem is solved by the proposed targeted treatment [4].

The proposed treatment method by SPIONs-PEG-HER by magnetic tumor targeting hyperthermia has never been studied anywhere else in the world. So, applying hyperthermia using magnetic coil where SPIONs-PEG-HER are injected intravenously to target the tumor cells and turn the magnetic energy to heat is introduced as the novelty of this study.

## **1.6 Thesis Organization**

This thesis includes five chapters. The first chapter describes the key information of the study: research background, problem statement, significance of study, research objectives, scope of research.

The second chapter comprehensively reviews the literature regarding the topic of the study. The introduction to the breast cancer treatments, the nature of SPIONs, the stabilization of SPIONs, the vectorization of the SPIONs and magnetic tumor-targeting hyperthermia are critically discussed and reviewed.

The third chapter describes the methodology in this study. This chapter elaborates the step-by-step synthesis of SPIONs-PEG-HER, physico-chemical characterization of the engineered system, *in vitro* and *in vivo* evaluation of the SPIONs-PEG-HER.

The fourth chapter of this thesis illustrates and discusses the obtained results of the experiments. It discusses on the properties of the developed SPIONs-PEG-HER and its effectiveness.

The entire findings of the research are concluded in chapter five. This chapter recommends the future works regarding this research.

## REFERENCES

- [1] J. Ma, A. Jemal, Breast Cancer Statistics, In: A. Ahmad (Ed.), *Breast Cancer Metastasis Drug Resist.* New York: Springer, 1-18; 2013.
- [2] Z.A. Omar, N.S. Ibrahim Tamin, National Cancer Registry Report, *NCR Report, 2007, Ministry of Health Malaysia.* 2011.
- [3] K. Kairemo, P. Erba, K. Bergström, E.K.J. Pauwels. Nanoparticles in cancer. *Curr. Radiopharm.*, 2008. 1:30-36.
- [4] S. Rasaneh. *Treatment and dosimetry of mouse breast cancer by magnetic nanoparticles conjugated with Herceptin and Lu177.* Ph.D. Thesis. Tarbiat Modares University of Tehran; 2010.
- [5] P. Cherukuri, E.S. Glazer, S. a Curley. Targeted hyperthermia using metal nanoparticles. *Adv. Drug Deliv. Rev.*, 2010. 62: 339–45.
- [6] M. Center, R. Siegel, A. Jemal. Global cancer facts & figures. *Atlanta Am. Cancer Soc.*, 2011. 1–52.
- [7] M. Yanase, M. Shinkai, H. Honda, T. Wakabayashi, J. Yoshida, T. Kobayashi. Intracellular hyperthermia for cancer using magnetite cationic liposomes: an in vivo study. *Jpn. J. Cancer Res.*, 1998. 89: 463–9.
- [8] J. Xing, J. Zeng, J. Yang, T. Kong, T. Xu, W. Roa, X. Wang, J. Chen. Gold-based nanoparticles for breast cancer diagnosis and treatment, *ISCAS.* 2007. IEEE Int. Symp.: IEEE. 2007.2882–2885.
- [9] E.B. Dickerson, E.C. Dreaden, X. Huang, I.H. El-Sayed, H. Chu, S. Pushpanketh, J.F. McDonald, M.A. El-Sayed. Gold nanorod assisted near-infrared plasmonic photothermal therapy (PPTT) of squamous cell carcinoma in mice. *Cancer Lett.*, 2008. 269: 57–66.
- [10] A. Jemal, F. Bray, M.M. Center, J. Ferlay, E. Ward, D. Forman. Global cancer statistics, *CA. Cancer J. Clin.*, 2011. 61: 69–90.
- [11] T.J. Wilt, R. MacDonald, I. Rutks, T.A. Shamliyan, B.C. Taylor, R.L. Kane.



- Systematic review: comparative effectiveness and harms of treatments for clinically localized prostate cancer. *Ann. Intern. Med.*, 2008. 148: 435–448.
- [12] R. Misra, S. Acharya, S.K. Sahoo. Cancer nanotechnology: application of nanotechnology in cancer therapy. *Drug Discov. Today.*, 2010. 15:842–850.
- [13] J.D. Martinez, M.T. Parker, K.E. Fultz, N.A. Ignatenko, E.W. Gerner. Burger's Medical Chemistry and Drug Discovery. *Molecular biology of cancer*. John Wiley and Sons, Inc. 2003.
- [14] R. Hergt, S. Dutz. Magnetic particle hyperthermia—biophysical limitations of a visionary tumour therapy. *J. Magn. Magn. Mater.*, 2007. 311: 187–192.
- [15] S. Dutz, R. Hergt, J. Mürbe, R. Müller, M. Zeisberger, W. Andrä, J. Töpfer, M.E. Bellemann. Hysteresis losses of magnetic nanoparticle powders in the single domain size range. *J. Magn. Magn. Mater.*, 2007. 308: 305–312.
- [16] J. Ferlay, I. Soerjomataram, M. Ervik, R. Dikshit, S. Eser, C. Mathers, M. Rebelo, D.M. Parkin, D. Forman, F. Bray. GLOBOCAN 2012, *Cancer Incid. Mortal. Worldw. IARC Cancer Base.*, 2013.
- [17] O.Z. Ariffin, I.T.N. Saleha. NCR report 2007. *Minist. Heal. Malaysia*. 2011.
- [18] A. Silverstein, V.B. Silverstein, L.S. Nunn. *Cells*. Twenty-First Century Books. 2009.
- [19] D.G. Hicks, S. Kulkarni. HER2+ breast cancer. *Am. J. Clin. Pathol.*, 2008. 129: 263–273.
- [20] U.S. Congress. Unconventional Cancer Treatment, In: OTA-H-405. Washington, DC: US Government Printing Office. 1990.
- [21] B.O. Anderson, E. Cazap, N.S. El Saghir, C.-H. Yip, H.M. Khaled, I. V Otero, C.A. Adebamowo, R.A. Badwe, J.B. Harford. Optimisation of breast cancer management in low-resource and middle-resource countries: executive summary of the Breast Health Global Initiative consensus. *Lancet Oncol.* 2011. 12: 387–398.
- [22] S. Akulapalli, History of cancer, ancient and modern treatment methods. *Journal of cancer science & therapy*, 2009. 1(2): 1.
- [23] F. Danhier, O. Feron, V. Préat. To exploit the tumor microenvironment: passive and active tumor targeting of nanocarriers for anti-cancer drug delivery. *J. Control. Release*, 2010. 148: 135–146.

- [24] R. De Souza, P. Zahedi, C.J. Allen, M. Piquette-Miller. Polymeric drug delivery systems for localized cancer chemotherapy. *Drug Deliv.*, 2010. 17: 365–375.
- [25] J. L. Arias. Drug targeting strategies in cancer treatment: an overview. *Mini Rev. Med. Chem.*, 2011. 11: 1–17.
- [26] G.A. Giovino, S.A. Mirza, J.M. Samet, P.C. Gupta, M.J. Jarvis, N. Bhala, R. Peto, W. Zatonski, J. Hsia, J. Morton. Tobacco use in 3 billion individuals from 16 countries: an analysis of nationally representative cross-sectional household surveys. *Lancet*, 2012. 380: 668–679.
- [27] N.L.S.T.R. Team. Reduced lung-cancer mortality with low-dose computed tomographic screening. *N Engl J Med.*, 2011. 365: 395–409.
- [28] S.L.V. Do. *Indoor air pollution in rural China; associated with lung function?*. Master's thesis. Universitetet i Agder ; University of Agder; 2013.
- [29] A.H. Aitkenhead. *The use of ultrasound to create tissue hyperthermia to support the treatment of cancer*. Ph.D. Thesis. Iniversity of Warwick; 2008.
- [30] M.R. Horsman, J. Overgaard. Hyperthermia: a potent enhancer of radiotherapy. *Clin. Oncol.*, 2007. 19: 418–426.
- [31] D. Sardari, N. Verga. *Cancer treatment with hyperthermia*. China: INTECH Open Access Publisher. 2011.
- [32] Y. Tang. *Cancer Therapy Combining Modalities of Hyperthermia and Chemotherapy: in vitro Cellular Response after Rapid Heat Accumulation in the Cancer Cell*. Master's Dissertation. Florida International University; 2010.
- [33] S. Laurent, D. Forge, M. Port, A. Roch, C. Robic, L. Vander Elst, R.N. Muller. Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications. *Chem. Rev.*, 2008. 108: 2064–110.
- [34] A. Jordan, P. Wust, H. Fähling, W. John, A. Hinz, R. Felix. Inductive heating of ferrimagnetic particles and magnetic fluids: physical evaluation of their potential for hyperthermia. *Int. J. Hyperth.*, 2009. 25: 499–511.
- [35] Q.A. Pankhurst, J. Connolly, S.K. Jones, J. Dobson. Applications of magnetic nanoparticles in biomedicine. *J. Phys. D. Appl. Phys.*, 2003. 36: R167.
- [36] D.-H. Kim, D.E. Nikles, D.T. Johnson, C.S. Brazel, Heat generation of aqueously dispersed  $\text{CoFe}_2\text{O}_4$  nanoparticles as heating agents for magnetically activated drug delivery and hyperthermia. *J. Magn. Magn. Mater.*, 2008. 320:

- 2390–2396.
- [37] R. Hergt, S. Dutz, M. Zeisberger. Validity limits of the Néel relaxation model of magnetic nanoparticles for hyperthermia. *Nanotechnology*, 2009. 21: 15706.
- [38] S. Chandra, K.C. Barick, D. Bahadur. Oxide and hybrid nanostructures for therapeutic applications. *Adv. Drug Deliv. Rev.*, 2001. 63: 1267–1281.
- [39] J.L. Phillips. A topical review of magnetic fluid hyperthermia. *J. Sci. Heal. Univ. Alabama*, 2005. 3:14–18.
- [40] R. Hergt, S. Dutz, R. Müller, M. Zeisberger. Magnetic particle hyperthermia: nanoparticle magnetism and materials development for cancer therapy. *J. Phys. Condens. Matter.*, 2006. 18: S2919.
- [41] S. Laurent, S. Dutz, U.O. Häfeli, M. Mahmoudi. Magnetic fluid hyperthermia: focus on superparamagnetic iron oxide nanoparticles. *Adv. Colloid Interface Sci.*, 2011. 166: 8–23.
- [42] Y. Shi. *Superparamagnetic nanoparticles for magnetic resonance imaging (MRI) diagnosis*. Ph.D. Thesis. The University of Adelaide; 2006.
- [43] G. Meier, S. Yuasa, A.T. Weddemann, J. Schmalhorst, S. See, C. Ruegg, T. Momoi, P. Sindzingre. Temperature-dependent investigation of domain wall depinning in nanowires by ballistic Hall micromagnetometr. *Annual General Meeting of the Magnetism Division*. Germany: Magnetism Division Fachverband Magnetismus (MA). 2009.
- [44] V.K. Varadan, L. Chen, J. Xie. *Nanomedicine: design and applications of magnetic nanomaterials, nanosensors and nanosystems*. John Wiley & Sons. 2008.
- [45] B.C. Saravanan, C. Sreekumar, G.C. Bansal, D. Ray, J.R. Rao, A.K. Mishra. A rapid MTT colorimetric assay to assess the proliferative index of two Indian strains of *Theileria annulata*. *Vet. Parasitol.*, 2003. 113: 211–216.
- [46] N. Amini, F.A. Abdul Majid, M. Marvibaigi, E. Supriyanto, S.K. Jaganathan, W. Tet Soon, R. Nasiri, J. Hamzehalipour. Cervicare<sup>TM</sup> induces apoptosis in HeLa and CaSki cells through ROS production and loss of mitochondrial membrane potential. *RSC Adv.*, 2016. 6: 24391–24417.
- [47] W. Strober. Trypan blue exclusion test of cell viability. *Curr. Protoc. Immunol.*, 2001. A3–B.
- [48] M. Mahmoudi, M.A. Sahraian, M.A. Shokrgozar, S. Laurent.

- Superparamagnetic iron oxide nanoparticles: promises for diagnosis and treatment of multiple sclerosis. *ACS Chem. Neurosci.*, 2011. 2: 118–140.
- [49] S. Sharifi, S. Behzadi, S. Laurent, M.L. Forrest, P. Stroeve, M. Mahmoudi. Toxicity of nanomaterials. *Chem. Soc. Rev.*, 2012. 41: 2323–2343.
- [50] M. Mahmoudi, A. Simchi, M. Imani, M.A. Shokrgozar, A.S. Milani, U.O. Häfeli, P. Stroeve. A new approach for the in vitro identification of the cytotoxicity of superparamagnetic iron oxide nanoparticles. *Colloids Surfaces B Biointerfaces*, 2010. 75: 300–309.
- [51] M. Mahmoudi, H. Hosseinkhani, M. Hosseinkhani, S. Boutry, A. Simchi, W.S. Journey, K. Subramani, S. Laurent. Magnetic resonance imaging tracking of stem cells in vivo using iron oxide nanoparticles as a tool for the advancement of clinical regenerative medicine. *Chem. Rev.* 2010. 111: 253–280.
- [52] M. Mahmoudi, S. Sant, B. Wang, S. Laurent, T. Sen. Superparamagnetic iron oxide nanoparticles (SPIONs): development, surface modification and applications in chemotherapy. *Adv. Drug Deliv. Rev.*, 2011. 63: 24–46.
- [53] V.I. Shubayev, T.R. Pisanic, S. Jin. Magnetic nanoparticles for theragnostics. *Adv. Drug Deliv. Rev.*, 2009. 61: 467–477.
- [54] H.M. Zareie, C. Boyer, V. Bulmus, E. Nateghi, T.P. Davis. Temperature-responsive self-assembled monolayers of oligo (ethylene glycol): control of biomolecular recognition. *ACS Nano*. 2008. 2: 757–765.
- [55] F.M. Veronese. Peptide and protein PEGylation: a review of problems and solutions. *Biomaterials*, 2001. 22: 405–417.
- [56] R. Gref, M. Lück, P. Quellec, M. Marchand, E. Dellacherie, S. Harnisch, T. Blunk, R.H. Müller. “Stealth” corona-core nanoparticles surface modified by polyethylene glycol (PEG): influences of the corona (PEG chain length and surface density) and of the core composition on phagocytic uptake and plasma protein adsorption. *Colloids Surfaces B Biointerfaces*, 2000. 18: 301–313.
- [57] B. Ballou, B.C. Lagerholm, L.A. Ernst, M.P. Bruchez, A.S. Waggoner. Noninvasive imaging of quantum dots in mice. *Bioconjug. Chem.* 2004. 15: 79–86.
- [58] L.E. van Vlerken, T.K. Vyas, M.M. Amiji. Poly (ethylene glycol)-modified nanocarriers for tumor-targeted and intracellular delivery. *Pharm. Res.* 2007. 24: 1405–1414.

- [59] T.J. Daou, L. Li, P. Reiss, V. Josserand, I. Texier, Effect of poly (ethylene glycol) length on the in vivo behavior of coated quantum dots. *Langmuir*, 2009. 25: 3040–3044.
- [60] B. Roberts. *Biographical research*. Buckingham: Open University Press. 2002.
- [61] C. Gao, D. Yan. Hyperbranched polymers: from synthesis to applications. *Prog. Polym. Sci.*, 2004. 29: 183–275.
- [62] L.X. Tiefenauer, A. Tschirky, G. Kühne, R.Y. Andres. In vivo evaluation of magnetite nanoparticles for use as a tumor contrast agent in MRI. *Magn. Reson. Imaging*, 1996. 14: 391–402.
- [63] K.G. Paul, T.B. Frigo, J.Y. Groman, E. V. Groman. Synthesis of ultrasmall superparamagnetic iron oxides using reduced polysaccharides. *Bioconjug. Chem.*, 2004. 15: 394–401.
- [64] A. Gabizon, D. Papahadjopoulos. The role of surface charge and hydrophilic groups on liposome clearance in vivo. *Biochim. Biophys. Acta (BBA)-Biomembranes*. 1992. 1103: 94–100.
- [65] D.D. Lasic, F.J. Martin, A. Gabizon, S.K. Huang, D. Papahadjopoulos. Sterically stabilized liposomes: a hypothesis on the molecular origin of the extended circulation times. *Biochim. Biophys. Acta (BBA)-Biomembranes*, 1991. 1070: 187–192.
- [66] V.P. Torchilin, V.G. Omelyanenko, M.I. Papisov, A.A. Bogdanov, V.S. Trubetskoy, J.N. Herron, C.A. Gentry, Poly (ethylene glycol) on the liposome surface: on the mechanism of polymer-coated liposome longevity. *Biochim. Biophys. Acta (BBA)-Biomembranes*. 1994. 1195: 11–20.
- [67] V.P. Torchilin, M.I. Papisov. Why do polyethylene glycol-coated liposomes circulate so long?: Molecular mechanism of liposome steric protection with polyethylene glycol: Role of polymer chain flexibility. *J. Liposome Res.*, 1994. 4: 725–739.
- [68] J.H. Senior. Fate and behavior of liposomes in vivo: a review of controlling factors. *Crit. Rev. Ther. Drug Carrier Syst.*, 1986. 3: 123–193.
- [69] A. Gabizon, F. Martin. Polyethylene glycol-coated (pegylated) liposomal doxorubicin. *Drugs*, 1997. 54: 15–21.
- [70] D. Needham, T.J. McIntosh, D.D. Lasic. Repulsive interactions and mechanical stability of polymer-grafted lipid membranes. *Biochim. Biophys.*

- Acta (BBA)-Biomembranes*. 1992. 1108: 40–48.
- [71] J. van der Zee. Heating the patient: a promising approach?. *Ann. Oncol.*, 2002. 13: 1173–1184.
- [72] E.C. Halperin, L.W. Brady, D.E. Wazer, C.A. Perez. *Perez & Brady's principles and practice of radiation oncology*. Lippincott Williams & Wilkins. 2013.
- [73] L.F.F. LG. Pathological effects of hyperthermia in normal tissues. *Cancer Res.*, 1984. 44: 4826–4835.
- [74] P. Sminia, J. Van Der Zee, J. Wondergem, J. Haveman. Effect of hyperthermia on the central nervous system: a review. *Int. J. Hyperth.* 1994. 10: 1–30.
- [75] J. Wondergem, J. Haveman, V. Rusman, P. Sminia, J.D.P. Van Dijk. Effects of local hyperthermia on the motor function of the rat sciatic nerve. *Int. J. Radiat. Biol.* 1988. 53: 429–438.
- [76] V.P. Torchilin. *Drug Delivery*. Berlin: Springer. 2010.
- [77] A. Jones, A.L. Harris. New developments in angiogenesis: a major mechanism for tumor growth and target for therapy. *Cancer J. Sci. Am.*, 1998. 4: 209.
- [78] S.K. Hobbs, W.L. Monsky, F. Yuan, W.G. Roberts, L. Griffith, V.P. Torchilin, R.K. Jain. Regulation of transport pathways in tumor vessels: role of tumor type and microenvironment. *Proc. Natl. Acad. Sci.*, 1998. 95: 4607–4612.
- [79] F. Yuan, M. Dellian, D. Fukumura, M. Leunig, D.A. Berk, V.P. Torchilin, R.K. Jain. Vascular permeability in a human tumor xenograft: molecular size dependence and cutoff size. *Cancer Res.*, 1995. 55: 3752–3756.
- [80] T.M. Allen, P.R. Cullis. Drug delivery systems: entering the mainstream. *Science*, 2004. 303: 1818–1822.
- [81] J.D. Byrne, T. Betancourt, L. Brannon-Peppas. Active targeting schemes for nanoparticle systems in cancer therapeutics. *Adv. Drug Deliv. Rev.*, 2008. 60: 1615–1626.
- [82] H. Maeda. Macromolecular therapeutics in cancer treatment: the EPR effect and beyond. *J. Control. Release*, 164: 138–144.
- [83] E. Miele, G.P. Spinelli, E. Miele, F. Tomao, S. Tomao. Albumin-bound formulation of paclitaxel (Abraxane® ABI-007) in the treatment of breast cancer. *International Journal of Nanomedicine*, 2009. 4: 99-105.

- [84] Y.H. Bae. Drug targeting and tumor heterogeneity. *J. Control. Release Off. J. Control. Release Soc.*, 2009. 133: 2-3.
- [85] D. Peer, J.M. Karp, S. Hong, O.C. Farokhzad, R. Margalit, R. Langer. Nanocarriers as an emerging platform for cancer therapy. *Nat. Nanotechnol.* 2007. 2: 751–760.
- [86] J. Shi, Z. Xiao, N. Kamaly, O.C. Farokhzad. Self-assembled targeted nanoparticles: evolution of technologies and bench to bedside translation. *Acc. Chem. Res.*, 2011. 44: 1123–1134.
- [87] U.B. Nielsen, G.P. Adams, L.M. Weiner, J.D. Marks. Targeting of bivalent anti-ErbB2 diabody antibody fragments to tumor cells is independent of the intrinsic antibody affinity. *Cancer Res.*, 2000. 60: 6434–6440.
- [88] G.P. Adams, R. Schier, K. Marshall, E.J. Wolf, A.M. McCall, J.D. Marks, L.M. Weiner. Increased affinity leads to improved selective tumor delivery of single-chain Fv antibodies. *Cancer Res.*, 1998. 58: 485–490.
- [89] Y. Stupp, T. Yoshida, W.E. Paul. Determination of antibody-hapten equilibrium constants by an ammonium sulfate precipitation technique. *J. Immunol.* 1969. 103: 625–627.
- [90] A. Gabizon, A.T. Horowitz, D. Goren, D. Tzemach, F. Mandelbaum-Shavit, M.M. Qazen, S. Zalipsky. Targeting folate receptor with folate linked to extremities of poly (ethylene glycol)-grafted liposomes: in vitro studies. *Bioconjug. Chem.*, 1999. 10: 289–298.
- [91] M. Mammen, S.-K. Choi, G.M. Whitesides. Polyvalent interactions in biological systems: implications for design and use of multivalent ligands and inhibitors, *Angew. Chemie Int. Ed.*, 1998. 37: 2754–2794.
- [92] H.T. Davis. *Statistical mechanics of phases, interfaces, and thin films.* Weinheim: Wiley-VCH; 1996.
- [93] K.S. Ho. *Targeted drug delivery to breast cancer using polymeric nanoparticle micelles.* Ph.D. Thesis. University of Toronto; 2012.
- [94] T. Seddiki, M. Ollivier-Bousquet. Temperature dependence of prolactin endocytosis and casein exocytosis in epithelial mammary cells. *Eur. J. Cell Biol.*, 1991. 55: 60–70.
- [95] J.H. Schiller, T. Larson, S.I. Ou, S.A. Limentani, A.B. Sandler, E.E. Vokes, S. Kim, K.F. Liau, P.W. Bycott, A.J. Olszanski. Efficacy and safety of axitinib (AG-013736; AG) in patients (pts) with advanced non-small cell lung cancer

- (NSCLC): a phase II trial. *ASCO Annu. Meet.*, Asco University: ASCO Annu. Meet. Proc. 2007. 7507.
- [96] G.D. Goss, A. Arnold, F.A. Shepherd, M. Dediu, T.-E. Ciuleanu, D. Fenton, M. Zukin, D. Walde, F. Laberge, M.D. Vincent. Randomized, double-blind trial of carboplatin and paclitaxel with either daily oral cediranib or placebo in advanced non-small-cell lung cancer: NCIC Clinical Trials Group BR24 study. *J. Clin. Oncol.*, 2010. 28: 49–55.
- [97] F.R. Hirsch, M. Varella-Garcia, P.A. Bunn, M. V Di Maria, R. Veve, R.M. Bremnes, A.E. Barón, C. Zeng, W.A. Franklin. Epidermal growth factor receptor in non-small-cell lung carcinomas: correlation between gene copy number and protein expression and impact on prognosis. *J. Clin. Oncol.*, 2003. 21: 3798–3807.
- [98] M.J. Moore, D. Goldstein, J. Hamm, A. Figer, J.R. Hecht, S. Gallinger, H.J. Au, P. Murawa, D. Walde, R.A. Wolff. Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *J. Clin. Oncol.*, 2007. 25: 1960–1966.
- [99] D.D. Karp, L.G. Paz-Ares, S. Novello, P. Haluska, L. Garland, F. Cardenal, L.J. Blakely, P.D. Eisenberg, C.J. Langer, G. Blumenschein. Phase II study of the anti-insulin-like growth factor type 1 receptor antibody CP-751,871 in combination with paclitaxel and carboplatin in previously untreated, locally advanced, or metastatic non-small-cell lung cancer. *J. Clin. Oncol.*, 2009. 27: 2516–2522.
- [100] L.B. Saltz, S. Clarke, E. Díaz-Rubio, W. Scheithauer, A. Figer, R. Wong, S. Koski, M. Lichinitser, T.-S. Yang, F. Rivera. Bevacizumab in combination with oxaliplatin-based chemotherapy as first-line therapy in metastatic colorectal cancer: a randomized phase III study. *J. Clin. Oncol.*, 2008. 26: 2013–2019.
- [101] H. Hurwitz, L. Fehrenbacher, W. Novotny, T. Cartwright, J. Hainsworth, W. Heim, J. Berlin, A. Baron, S. Griffing, E. Holmgren, Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N. Engl. J. Med.*, 2004. 350: 2335–2342.
- [102] J. Cassidy, S. Clarke, E. Díaz-Rubio, W. Scheithauer, A. Figer, R. Wong, S. Koski, M. Lichinitser, T.-S. Yang, F. Rivera. Randomized phase III study of



- capecitabine plus oxaliplatin compared with fluorouracil/folinic acid plus oxaliplatin as first-line therapy for metastatic colorectal cancer. *J. Clin. Oncol.*, 2008. 26: 2006–2012.
- [103] E. Van Cutsem, C.-H. Köhne, I. Láng, G. Folprecht, M.P. Nowacki, S. Cascinu, I. Shchepotin, J. Maurel, D. Cunningham, S. Tejpar. Cetuximab plus irinotecan, fluorouracil, and leucovorin as first-line treatment for metastatic colorectal cancer: updated analysis of overall survival according to tumor KRAS and BRAF mutation status. *J. Clin. Oncol.*, 2011. JCO-2010.
- [104] J.-Y. Douillard, S. Siena, J. Cassidy, J. Tabernero, R. Burkes, M. Barugel, Y. Humblet, G. Bodoky, D. Cunningham, J. Jassem, Randomized, phase III trial of panitumumab with infusional fluorouracil, leucovorin, and oxaliplatin (FOLFOX4) versus FOLFOX4 alone as first-line treatment in patients with previously untreated metastatic colorectal cancer: the PRIME study. *J. Clin. Oncol.*, 2010. JCO-2009.
- [105] C.A. Hudis. Trastuzumab—mechanism of action and use in clinical practice. *N. Engl. J. Med.*, 2007. 357: 39–51.
- [106] J.H. Almaki, R. Nasiri, A. Idris, F.A.A. Majid, M. Salouti, T.S. Wong, S. Dabagh, M. Marvibaigi, N. Amini. Synthesis, characterization and in vitro evaluation of exquisite targeting SPIONs–PEG–HER in HER2+ human breast cancer cells. *Nanotechnology*, 2016. 27: 105601.
- [107] I.E. Krop, M. Beeram, S. Modi, S.F. Jones, S.N. Holden, W. Yu, S. Girish, J. Tibbitts, J.-H. Yi, M.X. Sliwkowski. Phase I study of trastuzumab-DM1, an HER2 antibody-drug conjugate, given every 3 weeks to patients with HER2-positive metastatic breast cancer. *J. Clin. Oncol.*, 2010. 28: 2698–2704.
- [108] G.M. Keating. Pertuzumab. *Drugs*, 2012. 72: 353–360.
- [109] R. Nasiri, J.H. Almaki, A.B. Idris, F.A.A. Majid, M. Nasiri, M. Salouti, M. Irfan, N. Amini, M. Marvibaigi. In vitro evaluation of actively targetable superparamagnetic nanoparticles to the folate receptor positive cancer cells. *Mater. Sci. Eng. C*, 2016. 69: 1147–1158.
- [110] A. Tutt, M. Robson, J.E. Garber, S.M. Domchek, M.W. Audeh, J.N. Weitzel, M. Friedlander, B. Arun, N. Loman, R.K. Schmutzler. Oral poly (ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and advanced breast cancer: a proof-of-concept trial. *Lancet*, 2010. 376: 235–244.

- [111] J.S. De Bono, C.J. Logothetis, A. Molina, K. Fizazi, S. North, L. Chu, K.N. Chi, R.J. Jones, O.B. Goodman Jr, F. Saad. Abiraterone and increased survival in metastatic prostate cancer. *N. Engl. J. Med.*, 2011. 364: 1995–2005.
- [112] H.I. Scher, T.M. Beer, C.S. Higano, A. Anand, M.-E. Taplin, E. Efstathiou, D. Rathkopf, J. Shelkey, Y.Y. Evan, J. Alumkal. Antitumour activity of MDV3100 in castration-resistant prostate cancer: a phase 1–2 study. *Lancet*, 2010. 375: 1437–1446.
- [113] M.T. Fleming, G. Sonpavde, M. Kolodziej, S. Awasthi, T.E. Hutson, D. Martincic, A. Rastogi, S.R. Rousey, R.E. Weinstein, M.D. Galsky. Association of rash with outcomes in a randomized phase II trial evaluating cetuximab in combination with mitoxantrone plus prednisone after docetaxel for metastatic castration-resistant prostate cancer. *Clin. Genitourin. Cancer.*, 2012. 10: 6–14.
- [114] L. Chen, B.A. Mooso, M.K. Jathal, A. Madhav, S.D. Johnson, E. Van Spyk, M. Mikhailova, A. Zierenberg-Ripoll, L. Xue, R.L. Vinall. Dual EGFR/HER2 inhibition sensitizes prostate cancer cells to androgen withdrawal by suppressing ErbB3. *Clin. Cancer Res.*, 2011, 17: 6218–6228.
- [115] B. Schoeberl, A.C. Faber, D. Li, M.-C. Liang, K. Crosby, M. Onsum, O. Burenkova, E. Pace, Z. Walton, L. Nie, An ErbB3 antibody, MM-121, is active in cancers with ligand-dependent activation. *Cancer Res.*, 2010. 70: 2485–2494.
- [116] N.H. Bander, D.M. Nanus, M.I. Milowsky, L. Kostakoglu, S. Vallabahajosula, S.J. Goldsmith. Targeted systemic therapy of prostate cancer with a monoclonal antibody to prostate-specific membrane antigen, in: *Semin. Oncol.*, 2003. 30(5): 667–676.
- [117] R.J. Lee, P.S. Low. Folate-mediated tumor cell targeting of liposome-entrapped doxorubicin in vitro, *Biochim. Biophys. Acta (BBA)-Biomembranes*, 1995. 1233: 134–144.
- [118] R.J. Lee, P.S. Low. Folate-targeted liposomes for drug delivery. *J. Liposome Res.*, 1997. 7: 455–466.
- [119] C.J. Roberts, P.M. Williams, J. Davies, A.C. Dawkes, J. Sefton, J.C. Edwards, A.G. Haymes, C. Bestwick, M.C. Davies, S.J.B. Tendler. Real-space differentiation of IgG and IgM antibodies deposited on microtiter wells by scanning force microscopy. *Langmuir*, 1995. 11: 1822–1826.

- [120] V. Kouloulis, G. Plataniotis, J. Kouvaris, C. Dardoufas, C. Gennatas, N. Uzunoglu, C. Papavasiliou, L. Vlahos. Chemoradiotherapy combined with intracavitary hyperthermia for anal cancer: feasibility and long-term results from a phase II randomized trial. *Am. J. Clin. Oncol.*, 2005. 28: 91–99.
- [121] M. Palazzi, S. Maluta, S. Dall'Oglio, M. Romano, The role of hyperthermia in the battle against cancer. *Tumori*, 2010. 96: 902.
- [122] J.L. Roti Roti. Cellular responses to hyperthermia (40–46 C): Cell killing and molecular events. *Int. J. Hyperth.*, 2008. 24: 3–15.
- [123] E.P. Armour, D. McEachern, Z. Wang, P.M. Corry, A. Martinez. Sensitivity of human cells to mild hyperthermia. *Cancer Res.*, 1993. 53: 2740–2744.
- [124] F. Sonvico, S. Mornet, S. Vasseur, C. Dubernet, D. Jaillard, J. Degrouard, J. Hoebeke, E. Duguet, P. Colombo, P. Couvreur. Folate-conjugated iron oxide nanoparticles for solid tumor targeting as potential specific magnetic hyperthermia mediators: synthesis, physicochemical characterization, and in vitro experiments. *Bioconjug. Chem.*, 2005. 16: 1181–1188.
- [125] M. Shinkai, M. Yanase, T. Wakabayashi, J. Yoshida, T. Kobayashi. Intracellular hyperthermia for cancer using magnetite cationic liposomes: in vitro study. *Jpn. J. Cancer Res.*, 1996. 87: 1179–1183.
- [126] H.S. Huang, J.F. Hainfeld. Intravenous magnetic nanoparticle cancer hyperthermia. *Int. J. Nanomedicine*. 2013. 8: 2521:2532.
- [127] S. Hamaguchi, I. Tohnai, A. Ito, K. Mitsudo, T. Shigetomi, M. Ito, H. Honda, T. Kobayashi, M. Ueda. Selective hyperthermia using magnetoliposomes to target cervical lymph node metastasis in a rabbit tongue tumor model. *Cancer Sci.*, 2003. 94: 834–839.
- [128] A.A.M. Elsherbini, M. Saber, M. Aggag, A. El-Shahawy, H.A.A. Shokier. Magnetic nanoparticle-induced hyperthermia treatment under magnetic resonance imaging. *Magn. Reson. Imaging*. 2011. 29: 272–280.
- [129] A. Espinosa, R. Di Corato, J. Kolosnjaj-Tabi, P. Flaud, T. Pellegrino, C. Wilhelm. Duality of Iron Oxide Nanoparticles in Cancer Therapy: Amplification of Heating Efficiency by Magnetic Hyperthermia and Photothermal Bimodal Treatment. *ACS Nano*, 2016. 10: 2436–2446.
- [130] S. Kossatz, R. Ludwig, H. Dähring, V. Ettelt, G. Rimkus, M. Marciello, G. Salas, V. Patel, F.J. Teran, I. Hilger. High therapeutic efficiency of magnetic hyperthermia in xenograft models achieved with moderate temperature

- dosages in the tumor area. *Pharm. Res.*, 2014. 31: 3274–3288.
- [131] R.J. Wydra, A.M. Kruse, Y. Bae, K.W. Anderson, J.Z. Hilt. Synthesis and characterization of PEG-iron oxide core-shell composite nanoparticles for thermal therapy. *Mater. Sci. Eng. C*, 2013. 33: 4660–4666.
- [132] M. Shinkai, M. Yanase, H. Honda, T. Wakabayashi, J. Yoshida, T. Kobayashi. Intracellular hyperthermia for cancer using magnetite cationic liposomes: in vitro study. *Cancer Sci.*, 1996. 87: 1179–1183.
- [133] J. Motoyama, N. Yamashita, T. Morino, M. Tanaka, T. Kobayashi, H. Honda. Hyperthermic treatment of DMBA-induced rat mammary cancer using magnetic nanoparticles. *Biomagn. Res. Technol.*, 2008. 6(1): 1.
- [134] R. Massart. Preparation of aqueous magnetic liquids in alkaline and acidic media. *Magn. IEEE Trans.*, 1981. 17: 1247–1248.
- [135] A. Idris, N. Hassan, N.S. Mohd Ismail, E. Misran, N.M. Yusof, A.-F. Ngomsik, A. Bee. Photocatalytic magnetic separable beads for chromium (VI) reduction. *Water Res.*, 2010. 44: 1683–1688.
- [136] K. Hervé, L. Douziech-Eyrolles, E. Munnier, S. Cohen-Jonathan, M. Soucé, H. Marchais, P. Limelette, F. Warmont, M.L. Saboungi, P. Dubois, I. Chourpa. The development of stable aqueous suspensions of PEGylated SPIONs for biomedical applications. *Nanotechnology*, 2008. 19: 465608.
- [137] H. Wang, D.G. Castner, B.D. Ratner, S. Jiang. Probing the orientation of surface-immobilized immunoglobulin G by time-of-flight secondary ion mass spectrometry. *Langmuir*, 2004. 20: 1877–1887.
- [138] N. Patel, M.C. Davies, M. Hartshorne, R.J. Heaton, C.J. Roberts, S.J.B. Tandler, P.M. Williams. Immobilization of protein molecules onto homogeneous and mixed carboxylate-terminated self-assembled monolayers. *Langmuir*, 1997. 13: 6485–6490.
- [139] C. Grüttner, K. Müller, J. Teller, F. Westphal, A. Foreman, R. Ivkov. Synthesis and antibody conjugation of magnetic nanoparticles with improved specific power absorption rates for alternating magnetic field cancer therapy. *J. Magn. Magn. Mater.*, 2007. 311: 181–186.
- [140] M.M. Bradford. A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical Biochemistry*, 1976. 254: 248–254.
- [141] E. Munnier, S. Cohen-Jonathan, K. Hervé, C. Linossier, M. Soucé, P. Dubois,

- I. Chourpa. Doxorubicin delivered to MCF-7 cancer cells by superparamagnetic iron oxide nanoparticles: effects on subcellular distribution and cytotoxicity. *J. Nanoparticle Res.*, 2010. 13: 959–971.
- [142] J.-P. Fortin, C. Wilhelm, J. Servais, C. Ménager, J.-C. Bacri, F. Gazeau. Size-sorted anionic iron oxide nanomagnets as colloidal mediators for magnetic hyperthermia. *J. Am. Chem. Soc.*, 2007. 129: 2628–2635.
- [143] A. Ito, Y. Kuga, H. Honda, H. Kikkawa, A. Horiuchi, Y. Watanabe, T. Kobayashi. Magnetite nanoparticle-loaded anti-HER2 immunoliposomes for combination of antibody therapy with hyperthermia. *Cancer Lett.*, 2004. 212 : 167–75.
- [144] A. Saraswathy, S.S. Nazeer, N. Nimi, S. Arumugam, S.J. Shenoy, R.S. Jayasree. Synthesis and characterization of dextran stabilized superparamagnetic iron oxide nanoparticles for in vivo MR imaging of liver fibrosis. *Carbohydr. Polym.*, 2014. 101: 760–768.
- [145] N.H.A. Ngadiman, A. Idris, M. Irfan, D. Kurniawan, N.M. Yusof, R. Nasiri. Gamma-Fe<sub>2</sub>O<sub>3</sub> nanoparticles filled polyvinyl alcohol as potential biomaterial for tissue engineering scaffold. *J. Mech. Behav. Biomed. Mater.*, 2015. 49: 90–104.
- [146] T. Mosmann. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J. Immunol. Methods*, 1983. 65: 55–63.
- [147] M. Eghtedari, A. V Liopo, J.A. Copland, A.A. Oraevsky, M. Motamedi. Engineering of hetero-functional gold nanorods for the in vivo molecular targeting of breast cancer cells. *Nano Lett.*, 2008. 9: 287–291.
- [148] G.I. Salti, S. Grewal, R.R. Mehta, T.K. Das Gupta, A.W. Boddie Jr, A.I. Constantinou. Genistein induces apoptosis and topoisomerase II-mediated DNA breakage in colon cancer cells. *Eur. J. Cancer*, 2000. 36: 796–802.
- [149] N.A. Manaf, M.N.C. Aziz, D.S. Ridzuan, M.I.M. Salim, A.A. Wahab, K.W. Lai, Y.C. Hum, Feasibility of A-mode ultrasound attenuation as a monitoring method of local hyperthermia treatment. *Med. Biol. Eng. Comput.*, 2016. 54: 967–981.
- [150] L. Landini, R. Sarnelli. Evaluation of the attenuation coefficients in normal and pathological breast tissue. *Med. Biol. Eng. Comput.* 1986. 24: 243–247.
- [151] J. Lipka, M. Semmler-Behnke, R.A. Sperling, A. Wenk, S. Takenaka, C.

- Schleh, T. Kissel, W.J. Parak, W.G. Kreyling. Biodistribution of PEG-modified gold nanoparticles following intratracheal instillation and intravenous injection. *Biomaterials*, 2010. 31: 6574–6581.
- [152] X. Huang, I.H. El-Sayed, W. Qian, M.A. El-Sayed. Cancer cell imaging and photothermal therapy in the near-infrared region by using gold nanorods. *J. Am. Chem. Soc.*, 2006. 128: 2115–2120.
- [153] F.-K. Huang, W.-C. Chen, S.-F. Lai, C.-J. Liu, C.-L. Wang, C.-H. Wang, H.-H. Chen, T.-E. Hua, Y.-Y. Cheng, M.K. Wu. Enhancement of irradiation effects on cancer cells by cross-linked dextran-coated iron oxide (CLIO) nanoparticles. *Phys. Med. Biol.*, 2009. 55: 469.
- [154] D. Maiti, U. Manju, S. Velaga, P.S. Devi. Phase Evolution and Growth of Iron Oxide Nanoparticles: Effect of Hydrazine Addition During Sonication. *Crystal Growth and Design*. 2013. 13(8): 3637-3644.
- [155] W. Wu, X.H. Xiao, S.F. Zhang, T.C. Peng, J. Zhou, F. Ren, C.Z. Jiang. Synthesis and Magnetic Properties of Maghemite ( $\gamma\text{-Fe}_2\text{O}_3$ ) Short-Nanotubes. *Nanoscale Res. Lett.*, 2010. 5: 1474–1479.
- [156] E. Cheraghipour, a. M. Tamaddon, S. Javadpour, I.J. Bruce. PEG conjugated citrate-capped magnetite nanoparticles for biomedical applications. *J. Magn. Mater.*, 2013. 328: 91–95.
- [157] C. Liu, P.M. Huang. Atomic force microscopy and surface characteristics of iron oxides formed in citrate solutions. *Soil Sci. Soc. Am. J.*, 1999. 63: 65–72.
- [158] C. Sun, R. Sze, M. Zhang, Folic acid-PEG conjugated superparamagnetic nanoparticles for targeted cellular uptake and detection by MRI. *J. Biomed. Mater. Res. Part A*, 2006. 78: 550–557.
- [159] M. Lattuada, T.A. Hatton. Preparation and controlled self-assembly of Janus magnetic nanoparticles. *J. Am. Chem. Soc.*, 2007. 129: 12878–12889.
- [160] S. Nigam, K.C. Barick, D. Bahadur. Development of citrate-stabilized  $\text{Fe}_3\text{O}_4$  nanoparticles: Conjugation and release of doxorubicin for therapeutic applications. *J. Magn. Mater.*, 2010. 323: 237–243.
- [161] M.P. Patil, N.J. Gaikwad. Characterization of gliclazide-polyethylene glycol solid dispersion and its effect on dissolution. *Brazilian Journal of Pharmaceutical Sciences*, 2011. 47(1): 161-166.
- [162] A. Mukhopadhyay, N. Joshi, K. Chattopadhyay, G. De. A Facile Synthesis of PEG-Coated Magnetite ( $\text{Fe}_3\text{O}_4$ ) Nanoparticles and Their Prevention of the

- Reduction of Cytochrome C. *ACS applied materials & interfaces*, 2011. 4(1): 142-149.
- [163] F. Alexis, E. Pridgen, L.K. Molnar, O.C. Farokhzad. Factors Affecting the Clearance and Biodistribution of Polymeric Nanoparticles. *Molecular pharmaceuticals*, 2008. 5(4): 505–515.
- [164] M. Longmire, P.L. Choyke, H. Kobayashi. Clearance properties of nano-sized particles and molecules as imaging agents: considerations and caveats. *Nanomedicine*, 2008. 3(5): 703-717.
- [165] M. Răcuciu, D.E. Creangă, A. Airinei. Citric-acid-coated magnetite nanoparticles for biological applications. *Eur. Phys. J. E. Soft Matter.*, 2006. 21: 117–21.
- [166] T.-Y. Juang, S.-J. Kan, Y.-Y. Chen, Y.-L. Tsai, M.-G. Lin, L.-L. Lin. Surface-functionalized hyperbranched poly(amido acid) magnetic nanocarriers for covalent immobilization of a bacterial  $\gamma$ -glutamyltranspeptidase. *Molecules*, 2014. 19: 4997–5012.
- [167] K. Herve, L. Douziech-Eyrolles. The development of stable aqueous suspensions of PEGylated SPIONs for biomedical applications. *Nanotechnology*, 2008. 19(46): 465608.
- [168] N. Mansour, a Momeni, R. Karimzadeh, M. Amini. Surface effects on the luminescence properties of colloidal silicon nanocrystals in water. *Phys. Scr.*, 2013. 87: 35701.
- [169] R. Mehvar. Dextrans for targeted and sustained delivery of therapeutic and imaging agents. *Journal of controlled release*, 2000. 69: 1–25.
- [170] C. Flesch, M. Joubert, E. Bourgeat-Lami, S. Mornet, E. Duguet, C. Delaite, P. Dumas. Organosilane-modified maghemite nanoparticles and their use as co-initiator in the ring-opening polymerization of  $\epsilon$ -caprolactone, *Colloids Surfaces A Physicochem. Eng. Asp.*, 2005. 262: 150–157.
- [171] F. Hu, Q. Jia, Y. Li, M. Gao. Facile synthesis of ultrasmall PEGylated iron oxide nanoparticles for dual-contrast T1- and T2-weighted magnetic resonance imaging. *Nanotechnology*, 2011. 22: 245604.
- [172] M.S. Sadjadi, A. Sharafi, N. Farhadyar. Preparation of Surface Modified Fe<sub>3</sub>O<sub>4</sub> Nanostructures via Inverse Micelle Method and Study of their Magnetic Properties for Biological Applications. *J. Nano Res.*, 2013. 21: 37–42.

- [173] S.K. Shukla, A.K. Mishra, B.B. Mamba, O.A. Arotiba. Amperometric and Photometric Responses of in Situ Coupled Glucose Oxidase-Poly ( Propylene Imine ) Dendrimer Based Glucose Biosensor. *Int. J. Electrochem. Sci.*, 2013. 8: 11711–11722.
- [174] E.F. dos Reis, F.S. Campos, A.P. Lage, R.C. Leite, L.G. Heneine, W.L. Vasconcelos, Z.I.P. Lobato, H.S. Mansur. Synthesis and characterization of poly (vinyl alcohol) hydrogels and hybrids for rMPB70 protein adsorption. *Mater. Res.*, 2006. 9: 185–191.
- [175] Z. Yan-feng, G.A.O. Zhi-xian, S.U.N. Hong-wen, D.A.I. Shu-gui. Characterization of Hapten-Protein Conjugates for Immunoassay of Polycyclic Aromatic Hydrocarbons ( PAHs ). 2008. 24: 697–700.
- [176] T. Wang, H. Wu, W. Wang, F. Lin, P. Lou, M. Shieh, T. Young. The development of magnetic degradable DP-Bioglass for hyperthermia cancer therapy. *J. Biomed. Mater. Res. Part A*, 2007. 83: 828–837.
- [177] T. Theivasanthi, M. Alagar. X-ray diffraction studies of copper nanopowder. *arXiv Prepr. arXiv1003.6068*. 2010.
- [178] N.A. Brusentsov, V. V Gogosov, T.N. Brusentsova, A. V Sergeev, N.Y. Jurchenko, A.A. Kuznetsov, O.A. Kuznetsov, L.I. Shumakov. Evaluation of ferromagnetic fluids and suspensions for the site-specific radiofrequency-induced hyperthermia of MX11 sarcoma cells in vitro. *J. Magn. Magn. Mater.*, 2001. 225: 113–117.
- [179] C. Xu, K. Xu, H. Gu, X. Zhong, Z. Guo, R. Zheng, X. Zhang, B. Xu. Nitrotri-acetic acid-modified magnetic nanoparticles as a general agent to bind histidine-tagged proteins. *J. Am. Chem. Soc.*, 2004. 126: 3392–3393.
- [180] A. Idris, N.S.M. Ismail, N. Hassan, E. Misran, A.-F. Ngomsik. Synthesis of magnetic alginate beads based on maghemite nanoparticles for Pb(II) removal in aqueous solution. *J. Ind. Eng. Chem.*, 2012. 18: 1582–1589.
- [181] J.-H. Lee, J. Jang, J. Choi, S.H. Moon, S. Noh, J. Kim, J.-G. Kim, I.-S. Kim, K.I. Park, J. Cheon. Exchange-coupled magnetic nanoparticles for efficient heat induction. *Nat. Nanotechnol.*, 2011. 6: 418–422.
- [182] P. Guardia, R. Di Corato, L. Lartigue, C. Wilhelm, A. Espinosa, M. Garcia-Hernandez, F. Gazeau, L. Manna, T. Pellegrino. Water-soluble iron oxide nanocubes with high values of specific absorption rate for cancer cell hyperthermia treatment. *ACS Nano*, 2012. 6: 3080–3091.



- [183] G. Gokce, M. Cital, V. Gunes, G. Atalan. Effect of time delay and storage temperature on blood gas and acid–base values of bovine venous blood. *Res. Vet. Sci.*, 2004. 76: 121–127.
- [184] A. Fura, T.W. Harper, H. Zhang, L. Fung, W.C. Shyu. Shift in pH of biological fluids during storage and processing: effect on bioanalysis. *J. Pharm. Biomed. Anal.*, 2003. 32: 513–522.
- [185] A.C. Jayalekshmi, S.P. Victor, C.P. Sharma. Magnetic and degradable polymer/bioactive glass composite nanoparticles for biomedical applications. *Colloids Surfaces B Biointerfaces*, 2013. 101: 196–204.
- [186] C. V Durgadas, K. Sreenivasan, C.P. Sharma. Bright blue emitting CuSe/ZnS/silica core/shell/shell quantum dots and their biocompatibility. *Biomaterials.*, 2012. 33: 6420–6429.
- [187] M. Yu, S. Huang, K.J. Yu, A.M. Clyne. Dextran and Polymer Polyethylene Glycol (PEG) Coating Reduce Both 5 and 30 nm Iron Oxide Nanoparticle Cytotoxicity in 2D and 3D Cell Culture. *Int. J. Mol. Sci.*, 2012. 13: 5554–70.
- [188] M. Eghtedari, A. V Liopo, J.A. Copland, A.A. Oraevsky, M. Motamedi. Engineering of Hetero-Functional Gold Nanorods for the in vivo Molecular Targeting of Breast Cancer Cells. *Nanoletters*, 2009. 9(1): 287-291.
- [189] C.C. Berry, S. Wells, S. Charles, A.S.G. Curtis. Dextran and albumin derivatised iron oxide nanoparticles: influence on fibroblasts in vitro. *Biomaterials*, 2003. 24: 4551–4557.
- [190] E.R.L. de Freitas, P.R.O. Soares, R. de Paula Santos, R.L. dos Santos, J.R. da Silva, E.P. Porfirio, S.N. Bão, L. de Oliveira, E. Celma, P.C. Morais. In Vitro Biological Activities of Anionic-Fe<sub>2</sub>O<sub>3</sub> Nanoparticles on Human Melanoma Cells, *J. Nanosci. Nanotechnol*, 2008. 8: 2385–2391.
- [191] L.L. Rouhana, J.A. Jaber, J.B. Schlenoff. Aggregation-resistant water-soluble gold nanoparticles. *Langmuir*, 2007. 23: 12799–12801.
- [192] A. Ito, M. Shinkai, H. Honda, T. Wakabayashi, J. Yoshida, T. Kobayashi. Augmentation of MHC class I antigen presentation via heat shock protein expression by hyperthermia. *Cancer Immunol. Immunother.*, 2001. 50: 515–522.
- [193] M. Hedayati, O. Thomas, B. Abubaker-Sharif, H. Zhou, C. Cornejo, Y. Zhang, M. Wabler, J. Mihalic, C. Gruettner, F. Westphal. The effect of cell cluster size on intracellular nanoparticle-mediated hyperthermia: is it possible to treat

- microscopic tumors?. *Nanomedicine*, 2013. 8: 29–41.
- [194] B. V Harmon, A.M. Corder, R.J. Collins, G.C. Gobe, J. Allen, D.J. Allan, J.F.R. Kerr. Cell death induced in a murine mastocytoma by 42–47 C heating in vitro: evidence that the form of death changes from apoptosis to necrosis above a critical heat load. *Int. J. Radiat. Biol.*, 1990. 58: 845–858.
- [195] K.R. Bhayani, J.M. Rajwade, K.M. Paknikar. Radio frequency induced hyperthermia mediated by dextran stabilized LSMO nanoparticles: in vitro evaluation of heat shock protein response. *Nanotechnology*, 2012. 24: 15102.
- [196] M. Etrati Khosroshahi, L. Ghazanfari, Z. Hasan-Nejad. Preliminary results of treating cancerous cells of lung (QU-DB) by hyperthermia using diode laser and gold coated Fe<sub>3</sub>O<sub>4</sub>/SiO<sub>2</sub> nano-shells: an in-vitro assay. *Iran. J. Med. Phys.*, 2013. 9: 253–263.
- [197] J. Nakayama, T. Kageshita, M. Nakashima, M. Tsujisaki, K. Imai, Y. Hori. Increase in Shedding of Intercellular Adhesion Molecule-1 in Human Malignant Melanoma Cell Lines Treated With Hyperthermia In Vitro. *Pigment Cell Res.*, 1996. 9: 154–158.
- [198] V. Milani, B. Frankenberger, O. Heinz, A. Brandl, S. Ruhland, R.D. Issels, E. Noessner. Melanoma-associated antigen tyrosinase but not Melan-A/MART-1 expression and presentation dissociate during the heat shock response. *Int. Immunol.*, 2005. 17: 257–268.
- [199] D.E. Fisher. Apoptosis in cancer therapy: crossing the threshold. *Cell*, 1994. 78: 539–542.
- [200] M. Nasiri, S.A.H. Tabrizi, J.H. Almaki, R. Nasiri, A. Idris, S. Dabagh. Synthesis, functionalization, characterization, and in vitro evaluation of robust pH-sensitive CFNs–PA–CaCO<sub>3</sub>. *RSC Adv.*, 2016. 6: 84217–84230.
- [201] R. Sgonc, J. Gruber, Apoptosis detection: an overview. *Exp. Gerontol.*, 1998. 33: 525–533.
- [202] H. Jin, X. Xie, B. Hu, F. Gao, J. Zhou, Y. Zhang, L. Du, X. Wang, L. Zhao, X. Zhang. Hyperthermia inhibits the proliferation and invasive ability of mouse malignant melanoma through TGF- $\beta$ 1. *Oncol. Rep.*, 2013. 29: 725–734.
- [203] J. Razjouyan, H. Zolata, O. Khayat, F. Nowshiravan, N. Shadanpour, M. Mohammadnia. Synthesis and evaluation of radiolabeled, folic acid-PEG conjugated, amino silane coated magnetic nanoparticles in tumor bearing Balb/C mice. *Nukleonika*. 2015. 60: 497–502.

- [204] J. Mohammadnejad, M.J. Rasaei, M.H. Babaei, M. Paknejad, Z.M. Hasan, M. Salouti, M. Gandomkar, K. Sadri. Radioimmunotherapy of MCF7 breast cancer cell line with  $^{131}\text{I}$ -PR81 monoclonal antibody against MUC1: comparison of direct and indirect radioiodination methods. *Hum. Antibodies*, 19 (2010) 15–25.
- [205] L. Hosta-Rigau, I. Olmedo, J. Arbiol, L.J. Cruz, M.J. Kogan, F. Albericio. Multifunctionalized gold nanoparticles with peptides targeted to gastrin-releasing peptide receptor of a tumor cell line. *Bioconjug. Chem.*, 2010. 21: 1070–1078.