PRODUCTION OF NATURAL VANILLIN FROM Cymbopogon citratus USING Phanerochate chrysosporium

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SPECIALLY DEDICATED TO MY FAMILY & FRIENDS

"WORDS CAN NEVER EXPRESS MY GRATITUTES"

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ABSTRACT

Malaysia as one of the important agricultural countries in the world producing 70 million tonnes of lignocellulosic biomass. One of the abundant agricultural wastes which has high lignocellulose content is lemongrass leaves (Cymbopogon citratus). A total of 8,000 tonnes of dry leaves are produced annually from 1,150 hectares of C. citratus plantation in Malaysia. Parts of the leave wastes are burned for electric generation while the rest are left in the fields to decompose naturally. The use of lemongrass leaves as ruminant feedstock is, however, not favoured due to animal rejection against its aroma. C. citratus leaves have 58% hemicellulose and lignin content and are also rich in ferulic acid. Ferulic acid is the precursor for vanillin production. The C. citratus leave wastes that contain ferulic acid could be potentially used for vanillin production via microbial approach, which is currently less investigated. The main purpose of this research was to investigate the potential of a one-step natural vanillin production from C. citratus leaves hydrolysate by Phanerochaete chrysosporium, the white-rot fungi of basidiomycetes. The research work focused on the recovery of ferulic acid as well as the optimization of natural vanillin production from C. citratus leaves. Leaves within size of 125-249 µm appeared to be suitable for ferulic acid extraction, with 1.12g/L total recovery of ferulic acid by 55 minutes of boiling. A total of 27 strains of fungi were screened for natural vanillin and vanillic acid production. Among these fungi, Phanerochaete chrysosporium was chosen due to its high natural vanillin productivity. The effect of different nitrogen source in vanillin production was investigated using the General Factorial Design. A combination of ammonium chloride with yeast extract (ratio: 75:4) increased the vanillin production to 15-fold. To further optimize the process using 2-Level Factorial Design, the aforementioned nitrogen sources and 5 others independent variables (incubation temperature, pH, incubation time, agitation and inoculum size) were studied. The ammonium chloride concentration, temperature and pH were found to be the key factors for natural vanillin production and these factors were further examined using the Central Composite Design (CCD). The maximum vanillin production was observed with media composition of 1.0 g/L ferulic acid from hydrolysate of C. citratus leaves, 4.43 g/L ammonium chloride (inorganic nitrogen), 0.25 g/L yeast extract (organic nitrogen), 0.2 g/L KH₂PO₄, 0.013 g/L CaCl₂2.H₂O, 0.5 g/L MgSO₄.7H₂O and 0.0025 g/L thiamin hydrochloride, at 36.3 °C and pH 6.84 under shaking condition at 150 rpm with inoculum size of 7 % (w/v). The generated model fitted well to the data set with R^2 of 0.9059. The actual optimum conditions yielded 0.20 g/L vanillin (17-fold higher than non-optimised condition). The actual experimental data showed 2-fold higher yield as compared to previous reported studies. Artificial Neural Network model were also found to provide accurate validation and prediction result based on feed-forward back propagation as similar to Response Surface Methodology (RSM). A network with one hidden layer and 25 neurons were found to be the most optimum model with the lowest root mean squared error (RMSE; 0.0002415), absolute average deviation (AAD; 0.0004358) and the highest R^2 (0.9024). In this study, it was confirmed that C. citratus leaves could serve as a promising alternative source for vanillin production with potential to scale-up for commercial production in Malaysia.

Malaysia merupakan salah satu negara pertanian yang penting di dunia, telah menghasilkan 70 juta ton biojisim lignoselulotik. Sejumlah 8,000 ton daun kering dihasilkan pada setiap tahun daripada 1,150 hektar ladang C. citratus di Malaysia. Daun serai (Cymbopogon citratus) adalah hasil sisa pertanian yang mengandungi kandungan lignoselulosa yang tinggi. Sebahagian daripada sisa daun dibakar bagi menghasilkan tenaga elektrik sementara bakinya dibiarkan berdegradasi secara semulajadi. Penggunaan daun C. citratus sebagai sumber makanan ternakan tidak diterima oleh haiwan kerana aromanya. Daun C. citratus mempunyai 58% kandungan hemiselulosa dan lignin dan juga kaya dengan asid ferulik. Asid ferulik merupakan prekursor bagi penghasilan vanilin. Sisa daun C. citratus yang mengandungi asid ferulik berpotensi digunakan untuk penghasilan vanilin melalui pendekatan mikrob, di mana hingga kini kurang kajian dilakukan. Tujuan projek penyelidikan ini adalah untuk mengkaji potensi penghasilan vanilin semulajadi menggunakan satu langkah daripada hidrolisat daun C. citratus oleh Phanerochaete chrysosporium, kulat putih Basidiomycetes. Projek penyelidikan ini menfokuskan kepada pengestrakan asid ferulik dan juga pengoptimuman penghasilan vanilin semulajadi daripada C. *citratus*. Kajian mendapati, daun di dalam lingkungan saiz 125-249 um sesuai bagi pengekstrakan asid ferulik dengan jumlah keseluruhan pengestrakan asid ferulik 1.12g/L pada minit 55 prarawatan pendidihan. Sejumlah 27 strain kulat telah dikaji bagi penghasilan vanillin semulajadi dan asid vanilik. Di antara kulat-kulat ini, Phanerochaete chrysosporium telah dipilih berdasarkan kepada produktiviti penghasilan vanilin semulajadi yang tinggi. Pengaruh kepada perbezaan sumber nitrogen terhadap penghasilan vanilin semulajadi telah dikaji menggunakan Rekabentuk Faktorial Umum. Kombinasi ammonium klorida dan extrak yis pada nisbah 75:4 telah menunjukkan peningkatan penghasilan vanilin semulajadi sebanyak 15 kali ganda. Untuk pengoptimuman selanjutnya dengan menggunakan Rekabentuk Faktorial Tahap-2, sumber nitrogen yang dinyatakan di atas dan 5 lagi pembolehubah tak bersandar (suhu eraman, pH, tempoh masa eraman, pengadukan dan saiz inokulasi) telah dikaji. Kepekatan amonium klorida, suhu eraman dan pH telah didapati sebagai faktor kunci terhadap penghasilan vanilin semulajadi dan faktor-faktor ini telah dikaji selanjutnya menggunakan Rekabentuk Komposit Berpusat (RKB). Parameter-paramter yang ideal yang menghasilkan vanilin yang maksimum berlaku apabila 1.0 g/L asid ferulik daripada hidrolisat daun C. citratus, 4.43 g/L amonium klorida (nitrogen organik), 0.25 g/L ekstrak vis (nitrogen inorganik), 0.2 g/L KH₂PO₄, 0.0132 g/L CaCl₂2.H₂O, 0.5 g/L MgSO₄.7H₂O dan 0.0025 g/L tiamin hidroklorik digunakan pada 36.3 °C dan nilai pH 6.84 di bawah keadaan pengadukan 150 rpm dengan saiz inokulat 7 % (w/v). Model yang terhasil sangat sesuai dengan set data iaitu R² 0.9059. Keadaan optima pada keadaan sebenar telah menghasilkan 0.20 g/L vanilin semulajadi (17 kali ganda lebih tinggi berbanding keadaan tanpa pengoptimuman). Data eksperimen pada keadaan sebenar menunjukkan 2 kali ganda lebih tinggi hasil berbanding kajian yang dilaporkan terdahulu. Model Rangkaian Neural Buatan memberikan keputusan ramalan dan pengesahan yang tepat berdasarkan propogasi berbalik suapan ke hadapan yang sama seperti Rekabentuk Faktorial Umum (RFU). Satu jaringan dengan satu lapisan tersembunyi dan 25 neuron merupakan model paling optimum iaitu pada keadaan ralat kuadrat purata asas (RKPA: 0.0002415), perbezaan ralat mutlak (PRM; 0.0004358) yang paling rendah dan pada R² (0.9024) paling tinggi. Dalam kajian ini, telah disahkan bahawa daun C. citratus mampu menyediakan satu sumber yang menjanjikan kaedah alternatif bagi penghasilan yanilin dan berpotensi untuk digunakan secara komersil pada skala-besar di Malaysia.

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LIST OF ABBREVIATION

2-LFD	-	Two-factorial design
ANOVA	-	Analysis of Variance
BSA	-	Bovine Serum Albumin
CCD	-	Central Composite Design
СМС	-	Carboxymethyl cellulose
CMCase	-	Carboxymethyl cellulase
DNS	-	Dinitrosalicyclic acid
FESEM	-	Field Emission Scanning Electron Microscope
FID	-	Flame Ionized Detector
FPase	-	Filter Paper culture enzyme
FTIR	-	Fourier Transform Infrared Spectroscopy
g	-	Gram
GFD	-	General Factorial Design
H_2SO_4	-	Sulphuric acid
HCl	-	Hydrochloric acid
H_2O_2	-	Hydrogen Peroxides
HNO ₃	-	Nitric acid
HPLC	-	High Performance Liquid Chromatography
kDa	-	Kilo Dalton
L	-	Liter
min	-	Minute
mL	-	Milliliter
mm	-	Millimeter
MW	-	Molecular Weight

NaOH	-	Sodium hydroxide
N/A	-	Not applicable
nm	-	Nanometer
°C	-	Degree Celsius
OPT	-	Oil Palm Trunk
PAGE	-	Polyacrylamide Gel Electrophoresis
PDA	-	Potato Dextrose Agar
pNPG	-	p-nitrophenyl β -D-glucoside
RID	-	Refractive Index Detector
RSM	-	Response Surface Methodology
SDS	-	Sodium Dodecyl Sulfate
U/g	-	Unit of enzyme per gram
v/v	-	Volume per volume
w/v	-	Weight per volume
μL	-	Micro liter
μm	-	Micro meter

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CHAPTER 1

INTRODUCTION

1.1 Background of problem

Over 200 billion million tonnes of agricultural wastes were produced globally and they increased rapidly throughout the years (Figoli *et al.*, 2016; Agamuthu, 2009). In Malaysia, 15% (1.2 million tonnes) of agricultural waste has been disposed either to landfills or by burning activity that could lead to environmental problems (Agamuthu, 2015; Goralski *et al.*, 2015). Thus, the conversion of the lignocellulosic agriculture wastes into various value-added products can be a great idea to overcome this problem since they are considered as a promising natural, abundant and renewable feedstock. In addition, their physical and chemical properties made them potential biomass feedstock for secondary product in biofuel, macromolecules and aromatic compound production (Saini *et al.*, 2015; Ragauskas *et al.*, 2014; Zamzuri and Abd-Aziz, 2013; Chandra *et al.*, 2012).

As one of the world's largest producers and exporter of palm oil, cocoa and rubber, Malaysia creates a substantial amount of lignocellulosic agriculture wastes (Agamuthu, 2015). An estimation of 33 million tons of wastes are generated from the empty fruit bunch, fiber and shell itself (Abdullah and Sulaiman, 2013). Apart from the main plantation resources, herbs plants do contribute into Malaysia's economy

(Zakaria, 2015; International Trade Statistics Database, 2015). One of the most abundant and major herbs plant in the row is *Cymbobogon citratus* leaves (Agrofood Statistics and Department of Statistics, 2015; Ragauskas *et al.*, 2014; Nambiar and Matela, 2012).

The total production of Cymbopogon. citratus leaves in Malaysia was 12, 562 tonnes. 70% of the production of the C. citratus leaves, which is about 8800 tonnes of C. citratus leaves, was discarded as waste (Ministry of Agriculture and Agro-Based Industry report, 2015). The waste of *C. citratus* are partially dried in the fields and a fraction is burned to generate steam for stripping while the rest is left in the fields to naturally degrade (Syed et al., 2016; Machado et al., 2015; Amenaghawon et al., 2014). The usage as ruminant feedstock is however not a favor due to animal rejection against harsh or brittle texture of the leaves that can injure an animals' mouth (Kaur and Dutt, 2013; Guretzky et al., 2011; Ball et al., 2001). Cymbopogon citratus leaves consisted of 28.5-47% (w/v) of hemicelluloses, 29.9-35% (w/v) cellulose and 7-11% (w/v) lignin (Joshua et al., 2012; Nambiar and Matela 2012; Rolz, 1986). C. citratus leaves have approximately 58% of hemicellulose and lignin contents which related to rich of ferulic acid source. Although the utilization of C. citratus has been widely used in various fields, C. citratus leaves is an underexploited with ferulic acid as precursor for natural vanillin production. The idea of conducting the project is to generate sideincome to farmers and promote Bioeconomic National Transformation Programme agenda for high-income nation by 2020. The renewable product and their conversion by available lignocellulosic resource into food, feed stock, and healthcare wellness products via innovative and efficient of green, eco-friendly technologies should be emphasised.

Vanillin is the characteristic aroma component of the vanilla pod and frequently used for the production of flavors for foods and fragrances for perfume. The compounds practically prolonged the food life (Vandamme, 2003). At present, "artificial" or "nature-identical" vanillin is mostly produced from petrochemicals like guaiacol and from lignin. Due to the increasing demand for healthy and natural food, there is a growing interest to produce vanillin from natural raw materials by

biotransformation, which can then be regarded as a "natural" aroma chemical. This process is possible through microbial transformation when consortium of microorganisms and fungi are used to induce the production of these flavour compounds into optimum final yield (Lomascolo *et al.*, 1999).

Over the period of 2014 to 2019, flavors market will contribute a 5.5 billion dollar turnover and 4.86% growth rate which covers about one fourth of the global food additives market, and it is believed that this will continuously increase in proportion to the world demand of flavours (Sarethy and Pan, 2017). Flavouring compounds are known to be produced naturally from vanilla pod, chemically synthesized vanillin derived from lignin or fossil hydrocarbon and recently through bioconversion process, which is more eco-friendly towards environment (Leffingwell and Leffingwell, 2015; Gallage and Moller, 2015; Dal Bello, 2013; Havkin-Frenkel and Belanger, 2010; Brochado *et al.*, 2010).

1.2 Objectives

The main aim of the research is to develop environmental friendly bioconversion with significant yield of a one-step conversion of natural vanillin from *Cymbopogon citratus* leaves using *Phanerochaete chrysosporium*. In order to achieve the aim, the following studies have been conducted on;

- 1. Effect of chemical, physical, physicochemical and enzymatic pretreatments of *Cymbopogon citratus* leaves on the recovery of ferulic acid and reducing sugar
- Isolation of fungi for a one-step conversion of natural vanillin production in batch culture

 Screening and optimizing the significant factors of natural vanillin production using two level factorial design and central composite design (CCD) in batch culture

1.3 Limitation of consideration

In particular, this PhD work focused on the one step bioconversion of C. citratus leaves hydrolysate (containing ferulic acid) to natural vanillin production by selected fungi in which finally the *Phanerochaete chrysosporium* was employed. Recovery of ferulic acid made from Cymbopogon citratus leaves. Eventhough two common types of Cymbopogon available in Malaysia namely, Cymbopogon citratus and Cymbopogon nardus, yet Cymbopogon citratus leaves were chosen. This is because Cymbopogon nardus stalk and leaves are extracted for its essential oils in ingredient of therapeutic remedy. Cymbopogon citratus on the other hands, are abundance and feasible in terms of the production. The stalk of *Cymbopogon citratus* is mainly for cooking and flavouring, yet the leaves are discarded. There is eleven physical and chemical factors that involved in the production of natural vanillin. Due to time limitation, only seven major parameters were selected from previous studies for optimum production of natural vanillin. The selection parameters are inoculum size, pH value, supplement medium composition of inorganic and organic nitrogen source, incubation temperature, inoculum as well as incubation duration time. There are also many statistical approaches that can be used for statistical analysis. Plackett-Burman design is one of the choice but require more than 20 factors to fit a design. The onefactor-at-a-time (OFAT) is normally give accurate results but rather irrelevant with unnecessary experiment design that consume a lot of time and cost. The Response Surface Methodology (RSM) statistical approach of Design Expert ® version 6.04 is chosen. This is because RSM has better approach in identifying the ideal combination of factors with any levels that can affect the process and indicates the optimal value of the selected parameters. Artificial Neural Network (ANN) is chosen or further validation and prediction because it does not need any mathematical model. One of

advantages of an ANN is that it learns from examples and recognizes patterns in a series of input and output data without any prior assumptions about their nature and interrelations. There are many types of learning algorithms in the literature which can be used for training of the network which enable the network to model nonlinear and complex functions. However, feed-forward neural network is chosen as it could provide an accurate prediction even with a single hidden layer. The appropriate selection of propagation learning algorithm can be adopted for the training of all the ANN with high rate of successful data modelling. Due to time constraints, Genetic Algorithms could not be performed due to time limitation and require selection of larger class of evolutionary algorithms.

1.4 Organization of research

The research is initiated with screening of selected fungi through quantitative and qualitative analyses to identify the isolate that was capable to transform ferulic acid to natural vanillin through liquid-state fermentation (C. citratus leaves hydroysate), *Phanerochaete chrysosporium* was finally selected into further steps of optimization. The wild type strain is able to use ferulic acid as carbon source (Karode et al., 2013; Barbosa et al., 2008). The experimental activities focused on the recovery of ferulic acid as well as optimization of natural vanillin production. The aim was thus develop environmentally friendly bioconversion protocols based on the to combination of enzymatic and physicochemical (thermal/boiling) pretreatment of C. *citratus* leaves. The methods were used in order to obtain a high recovery of ferulic acid and sugar in hydrolysate. There is evidence that purification of ferulic acid from the carbohydrate occurring in the hydrolysate might be necessary to avoid increase in oxido-reductive enzymes activity that converts natural vanillin into side-products, thus reducing the natural vanillin molar yield (Dal Bello, 2013). The natural vanillin production from *Phanerochaete chrysosporium* was optimized using statistical approaches. The best nitrogenous supplement in the basal medium was determined

using general factorial design (GFD). The two-level factorial design (2LFD) was used to select the most significant parameters that influenced the natural vanillin production, and lastly CCD was used to determine their optimal values. Then followed by Artificial Neural Network (ANN) using a multi-layer perceptron (MLP) based feed-forward back propagation learning algorithm to validate datasets obtained during the natural vanillin production and predict value of sample as well. The thesis comprises of 7 chapters (including the present chapter), contents of which are briefly outlined below:

In chapter 2, a comprehensive review of literature on lignocellulosic biomass and agriculture waste residue as well as several important aspects of classification, type and processing methods of vanilla or natural vanillin. This including the physical and chemical factors that influence the natural vanillin production. The major findings of significant parameters in obtaining optimum natural vanillin production are also emphasised.

In chapter 3, the general research methodology repeatedly use throughout the study is presented in great depth.

In chapter 4, the biomass of lignocellulosic composition, phenolic compounds profile and reducing sugar analysis in raw *C. citratus* leaves were highlighted for substrate selection suitability. Then describe the chemical, physical, physicochemical and enzymatic pretreatment of the *C. citratus* leaves by measuring ferulic acid and phenolic compounds recovery, physical modification and total reducing sugar analysis as well.

Chapter 5, the isolation and screening of potential fungi amongst the local isolates and commercial strains that capable in a one-step conversion of natural vanillin production is presented.

Chapter 6, the statistical optimization of medium and physical factors including incubation temperature, inoculums sizes, pH and ratio of selected supplement medium composition that had strongly influenced the submerged liquid fermentation of vanillin production is described.

Chapter 7 summarizes the major finding of the thesis and gives some suggestions for further research.

- Abbas, A., Koc, H., Liu, F. and Tien, M. (2005). Fungal degradation of wood: initial proteomic analysis of extracellular proteins of *Phanerochaete chrysosporium* grown on oak substrate. *Current genetics*, 47(1), pp.49-56.
- Abdul Khalil, H.P.S., Siti Alwani, M., Ridzuan, R., Kamarudin, H. and Khairul, A. (2008). Chemical composition, morphological characteristics, and cell wall structure of Malaysian oil palm fibers. *Polymer-Plastics Technology and Engineering*, 47(3), pp.273-280.
- Abdullah, N. and Sulaiman, F. (2013). The oil palm wastes in Malaysia. *Biomass Now-Sustainable Growth and Use*, 1(3), pp.75-93.
- Adapa, P., Tabil, L. and Schoenau, G. (2009). Compaction characteristics of barley, canola, oat and wheat straw. *Biosystems engineering*, *104*(3), pp.335-344.
- Aftab, K., Ali, M.D., Aijaz, P., Beena, N., Gulzar, H.J., Sheikh, K., Sofia, Q. and Tahir Abbas, S. (2011). Determination of different trace and essential element in lemongrass samples by x-ray fluorescence spectroscopy technique. *International food research journal*, 18(1).
- Afzal, H., Shazad, S. and Nisa, S.O.U. (2013). Morphological identification of Aspergillus species from the soil of Larkana district (Sindh, Pakistan). *Asian J AgriBiol*, *1*, pp.105-117.
- Agbor, V.B., Cicek, N., Sparling, R., Berlin, A. and Levin, D.B. (2011). Biomass pretreatment: fundamentals toward application. *Biotechnology advances*, 29(6), pp.675-685.
- Agrofood Statistics and Department of Statistics (2015). http://www.moa.gov.my/documents/10157/010b6921-c643-421d-aef1-29ae379f6f85
- Agamuthu, P. (2015). Circular Economic Utilization of Agriculture and Biomass Waste : A Potential Opportunity for Asia and the Pacific. In *Inaugural Meeting of 6th Regional 3R Forum in Asia. Male,* Maldives.
- Agamuthu, P. (2009) Challenges and opportunities in agro-waste management: An Asian perspective. In *Inaugural Meeting of First Regional 3R Forum in Asia. Tokyo, Japan.*
- Agatonovic-Kustrin, S. and Beresford, R. (2000). Basic concepts of artificial neural network (ANN) modeling and its application in pharmaceutical research. *Journal of pharmaceutical and biomedical analysis*, 22(5), pp.717-727.
- Albores, L.C.O., Baños, S.B., Herrera, J.M.I., Necha, L.B., Lopez, M.O.H.A. and Hern&andez, A.E.C. (2014). Morphological and molecular characterization of pathogenic isolates of *Fusarium spp.* obtained from gladiolus corms and their sensitivity to Jatropha curcas L. oil. *African Journal of Microbiology Research*, 8(8), pp.724-733.
- Amaike, S. and Keller, N.P. (2011). Aspergillus flavus. Annual review of phytopathology, 49, pp.107-133.

- Amenaghawon, N.A., Okhueleigbe, K.E., Ogbeide, S.E. and Okieimen, C.O. (2014). Modelling the Kinetics of Steam Distillation of Essential Oils from Lemon Grass (*Cymbopogon spp.*). *International Journal of Applied Science and Engineering*, *12*(2),107-115.
- Ander, P., Hatakka, A. and Eriksson, K.E. (1980). Vanillic acid metabolism by the white-rot fungus *Sporotrichum pulverulentum*. *Archives of Microbiology*, *125*(3), pp.189-202.
- Anderson, M., Kraber, S., Hansel, H., Klick, S., Beckenbach, R. and Cianca-Betancourt, H., Design Expert Software (2002), Version 6 Users Guide. *MN: Statease Inc.*
- Anderson, M. and Whitcomb, P. (2005). RSM simplified: optimizing processes using Response Surface Methods for Design of Experiments. Productivity Press Florence, KY.
- Anderson, M. J. and Whitcomb, P. J. (2000). DOE simplified: practical tools for effective experimentation. Productivity OR.
- Andreoni, V., Bernasconi, S. and Bestetti, G. (1995). Biotransformation of ferulic acid and related compounds by mutant strains of *Pseudomonas fluorescens*. Applied microbiology and biotechnology, 42(6), pp.830-835.
- Ang, S., Shaza, E., Adibah, Y., Suraini, A. and Madihah, M. (2013). Production of cellulases and xylanase by *Aspergillus fumigatus* SK1 using untreated oil palm trunk through solid state fermentation. *Process Biochemistry*, 1293-1302.
- Antony, J., Chou, T. and Ghosh, S. (2003). Training for design of experiments. *Work study*, 52, 341-346.
- Artursson, V., Finlay, R.D. and Jansson, J.K. (2006). Interactions between arbuscular mycorrhizal fungi and bacteria and their potential for stimulating plant growth. *Environmental Microbiology*, 8(1), pp.1-10.
- Asaolu, M.F., Oyeyemi, O.A. and Olanlokun, J.O. (2009). Chemical compositions, phytochemical constituents and in vitro biological activity of various extracts of *Cymbopogon citratus*. *Pakistan Journal of Nutrition*, 8(12), pp.1920-1922.
- Ashengroph, M., Nahvi, I. and Amini, J. (2013). Application of Taguchi design and Response surface methodology for improving conversion of isoeugenol into vanillin by resting cells of *Psychrobacter sp. CSW4. Iranian Journal of Pharmaceutical Research*, *12*(3), pp.411-421.
- Ashengroph, M., Nahvi, I., Zarkesh-Esfahani, H. and Momenbeik, F. (2012). Conversion of isoeugenol to vanillin by *Psychrobacter sp.* strain CSW4. *Applied biochemistry and biotechnology*, 166(1), pp.1-12.
- Ashengroph, M., Nahvi, I., Zarkesh-Esfahani, H. and Momenbeik, F. (2011). Candida galli strain PGO6: a novel isolated yeast strain capable of transformation of isoeugenol into vanillin and vanillic acid. Current Microbiology, 62(3), pp.990-998.
- Ashokkumar, R. and Ramaswamy, M. (2014). Phytochemical screening by FTIR spectroscopic analysis of leaf extracts of selected Indian medicinal plants. *International Joural Current Microbiol Applied Science*, *3*, pp.395-6.

- Audrey A. K., Daniel, T. S., Kanae, M., Joanna, L., William, O. W., David, M. U., Zongli X., Danica, D., Jack, A. T., David, M. DeM., Catherine, B. K.(2007). Antimutagenicity of cinnamaldehyde and vanillin in human cells: Global gene expression and possible role of DNA damage and repair, *Mutation Research/Fundamental and Molecular Mechanisms* of Mutagenesis, 616 (1–2), 60-69.
- Auxenfans, T., Crônier, D., Chabbert, B. and Paës, G. (2017). Understanding the structural and chemical changes of plant biomass following steam explosion pretreatment. *Biotechnology for biofuels*, *10*(1), p.36.
- Ayyachamy, M., Cliffe, F.E., Coyne, J.M., Collier, J. and Tuohy, M.G. (2013). Lignin: untapped biopolymers in biomass conversion technologies. *Biomass Conversion and Biorefinery*, 3(3), pp.255-269.
- Bahrin, E., Ibrahim, M., Abd Razak, M., Abd-Aziz, S., Shah, U. M., Alitheen, N. and Salleh, M. M. (2012). Improved cellulase production by *Botryosphaeria rhodina* from OPEFB at low level moisture condition through statistical optimization. *Preparative Biochemistry and Biotechnology*, 42, 155-170.
- Balasundram, N., K. Sundram, and S. Samman. (2006). Phenolic compounds in plants and agriindustrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chemistry*, 99, 191-203
- Ball, D.M., Collins, M., Lacefield, G.D., Martin, N.P., Mertens, D.A., Olson, K.E., Putnam, D.H., Undersander, D.J. and Wolf, M.W. (2001). Understanding forage quality. *American Farm Bureau Federation Publication*, 1(01).
- Barbosa, E.S., Perrone, D., Amaral Vendramini, A.L., and Ferreira Leite, S.G. (2008). Vanillin Production by *Phanerochaete chrysosporium* grown on green coconut agro-industrial husk in solid state fermentation. *BioResources*, 3, 1042-1050.
- Barghini, P., Di Gioia, D., Fava, F. and Ruzzi, M. (2007). Vanillin production using metabolically engineered Escherichia coli under non-growing conditions. *Microbial Cell Factories*, 6(1), p.13.
- Barghini, P., Montebove, F., Ruzzi, M. and Schiesser, A. (1998). Optimal conditions for bioconversion of ferulic acid into vanillic acid by *Pseudomonas fluorescens* BF13 cells. *Applied microbiology and biotechnology*, 49(3), pp.309-314.
- Baş, D., Dudak, F.C. and Boyacı, I.H. (2007). Modeling and optimization III: Reaction rate estimation using artificial neural network (ANN) without a kinetic model. *Journal of food engineering*, 79(2), pp.622-628.
- Basheer, I.A. and Hajmeer, M. (2000). Artificial neural networks: fundamentals, computing, design, and application. *Journal of microbiological methods*, 43(1), pp.3-31.
- Basri, M., Rahman, R.N.Z.R.A., Ebrahimpour, A., Salleh, A.B., Gunawan, E.R. and Rahman, M.B.A. (2007). Comparison of estimation capabilities of response surface methodology (RSM) with artificial neural network (ANN) in lipase-catalyzed synthesis of palm-based wax ester. *BMC biotechnology*, 7(1), p.53.

- Bauer, K., Garbe, D. and Surburg, H. (2008). *Common fragrance and flavor materials: preparation, properties and uses.* John Wiley & Sons.
- Belmessikh, A., Boukhalfa, H., Mechakra-Maza, A., Gheribi-Aoulmi, Z. and Amrane, A. (2013). Statistical optimization of culture medium for neutral protease production by *Aspergillus* oryzae. Comparative study between solid and submerged fermentations on tomato pomace. Journal of the Taiwan Institute of Chemical Engineers, 44(3), pp.377-385.
- Bensah, E.C. and Mensah, M. (2013). Chemical pretreatment methods for the production of cellulosic ethanol: technologies and innovations. *International Journal of Chemical Engineering*, 2013.
- Berger, R. G. (2015). Biotechnology as a source of natural volatile flavours. *Current Opinion in Food Science*, 1: 38-43.
- Berger, R.G. (2009). Biotechnology of flavours-the next generation. *Biotechnology letters*, *31*(11), 1651-1659.
- Berger, R.G. ed. (2007). *Flavours and fragrances: chemistry, bioprocessing and sustainability*. Springer Science & Business Media.
- Bernard, O., Bastin, G., Stentelaire, C., Lesage-Meessen, L. and Asther, M. (1999). Mass balance modeling of vanillin production from vanillic acid by cultures of the fungus *Pycnoporus cinnabarinus* in bioreactors. *Biotechnology and Bioengineering*, 65(5), 558-571.
- Betiku, E. and Taiwo, A.E. (2015). Modeling and optimization of bioethanol production from breadfruit starch hydrolyzate vis-à-vis response surface methodology and artificial neural network. *Renewable Energy*, *74*, pp.87-94.
- Bicas, J.L., Silva, J.C., Dionísio, A.P. and Pastore, G.M. (2010). Biotechnological production of bioflavors and functional sugars. *Food Science and Technology (Campinas)*, 30(1), 07-18.
- Bloem, A., Bertrand, A., Lonvaud-Funel, A. and De Revel, G. (2007). Vanillin production from simple phenols by wine-associated lactic acid bacteria. *Letters in Applied Microbiology*, 44(1), pp.62-67.
- Bodirlau, R. and Teaca, C.A. (2009). Fourier transform infrared spectroscopy and thermal analysis of lignocellulose fillers treated with organic anhydrides. *Rom Journal Physics*, 54(1-2), pp.93-104.
- Bonnin, E., Lesage-Meessen, L., Asther, M. and Thibault, J.-F. (1999). Enhanced bioconversion of vanillic acid into vanillin by the use of natural cellobiose. *Journal. Science. Food Agriculture*. 79: 484–486.
- Bonnin, E., Grange, H., Lesage-Meessen, L., Asther, M., and Thibaults, J.F. (2000). Enzymatic release of cellobiose from sugar beet pulp and its use to favour vanillin production in *Pycnoporus cinnabarinus* from vanillic acid. *Carbohydrate Polymer*, 41, 143-151.

- Bonnin, E., Saulnier, L., Brunel, M., Marot, C., Lesage-Meessen, L., Asther, M. and Thibault, J. F. (2002). Release of ferulic acid from agroindustrial by-products by the cell wall-degrading enzymes produced by Aspergillus niger I-1472. *Enzyme and Microbial Technology*, 31(7), 1000-1005.
- Boukhatem, M.N., Ferhat, M.A., Kameli, A., Saidi, F. and Kebir, H.T. (2014). Lemon grass (*Cymbopogon citratus*) essential oil as a potent anti-inflammatory and antifungal drugs. *Libyan Journal of Medicine*, 9(1).
- Brandt, A., Gräsvik, J., Hallett, J.P. and Welton, T. (2013). Deconstruction of lignocellulosic biomass with ionic liquids. *Green Chemistry*, *15*(3), pp.550-583.
- Bringans, S., Eriksen, S., Kendrick, T., Gopalakrishnakone, P., Livk, A., Lock, R. and Lipscombe, R. (2008). Proteomic analysis of the venom of *Heterometrus longimanus* (Asian black scorpion). *Proteomics*, 8(5), pp.1081-1096.
- Brochado, A.R., Matos, C., Møller, B.L., Hansen, J., Mortensen, U.H. and Patil, K.R. (2010). Improved vanillin production in baker's yeast through in silico design. *Microbial cell factories*, 9(1), p.84.
- Brodeur, G., Yau, E., Badal, K., Collier, J., Ramachandran, K.B. and Ramakrishnan, S. (2011). Chemical and physicochemical pretreatment of lignocellulosic biomass: a review. *Enzyme Research*, 2011.
- Brunati, M., Marinelli, F., Bertolini, C., Gandolfi, R., Daffonchio, D. and Molinari, F. (2004). Biotransformations of cinnamic and ferulic acid with actinomycetes. *Enzyme and Microbial Technology*, 34, 3-9.
- Brunk, E., Mih, N., Monk, J., Zhang, Z., O'Brien, E.J., Bliven, S.E., Chen, K., Chang, R.L., Bourne, P.E. and Palsson, B.O. (2016). Systems biology of the structural proteome. *BMC* systems biology, 10(1), p.10-26.
- Budavari, S., Smith, A. and Heckelman, P. E. (1996). Chemical summary for vanillin. Kinneary J.F.(eds). 12th Eds. Merck & Co., Inc., Whitehouse Station. pp. 1-7.
- Buranov, A.U. and Mazza, G. (2008). Lignin in straw of herbaceous crops. *Industrial crops and products*, 28(3), pp.237-259.
- Burri, J., Graf, M., Lambelet, P. and Löliger, J. (1989). Vanillin: more than a flavouring agent a potent antioxidant. *Journal of the Science of Food and Agriculture*, 48(1), pp.49-56
- Cadoche, L. and López, G.D. (1989). Assessment of size reduction as a preliminary step in the production of ethanol from lignocellulosic wastes. *Biological Wastes*, *30*(2), pp.153-157.
- Caldwell, D.R. and Bryant, M.P. (1966). Medium without rumen fluid for nonselective enumeration and isolation of rumen bacteria. *Applied microbiology*, *14*(5), pp.794-801.
- Canilha, L., Chandel, A.K., Suzane dos Santos Milessi, T., Antunes, F.A.F., Luiz da Costa Freitas,
 W., das Graças Almeida Felipe, M. and da Silva, S.S. (2012). Bioconversion of sugarcane
 biomass into ethanol: an overview about composition, pretreatment methods,

detoxification of hydrolysates, enzymatic saccharification, and ethanol fermentation. *BioMedical Research International*, 2012.

- Carvalheiro, F., Duarte, L.C. and Gírio, F.M. (2008). Hemicellulose biorefineries: a review on biomass pretreatments. *Journal of Scientific & Industrial Research*, pp.849-864.
- Casey, J. and Dobb, R. (1992). Microbial routes to aromatic aldehydes. *Enzyme and microbial technology*, *14*(9), pp.739-747.
- Castro-Concha, L.A., Tuyub-Che, J., Moo-Mukul, A., Vazquez-Flota, F.A. and Miranda-Ham, M.L. (2014). Antioxidant capacity and total phenolic content in fruit tissues from accessions of *Capsicum chinense Jacq*.(Habanero Pepper) at different stages of ripening. *The Scientific World Journal*, 2014.
- Cazetta, M.L., Celligoi, M.A.P.C., Buzato, J.B. and Scarmino, I.S. (2007). Fermentation of molasses by Zymomonas mobilis: Effects of temperature and sugar concentration on ethanol production. *Bioresource technology*, 98(15), pp.2824-2828.
- Cerrutti, P., Alzamora, S.M. and Vidales, S.L. (1997). Vanillin as an antimicrobial for producing shelf-stable strawberry puree. *Journal of Food Science*, *62*(3), 608-610.
- Ciftci, T., Constantinides, A. and Wang, S.S. (1983). Optimization of conditions and cell feeding procedures for alcohol fermentation. *Biotechnology and bioengineering*, *25*(8), pp.2007-2023.
- Chandel, A.K., Da Silva, S.S. and Singh, O.V. (2013). Detoxification of lignocellulose hydrolysates: biochemical and metabolic engineering toward white biotechnology. *BioEnergy Research*, 6(1), pp.388-401.
- Chandra, R., Takeuchi, H. and Hasegawa, T. (2012). Methane production from lignocellulosic agricultural crop wastes: A review in context to second generation of biofuel production. *Renewable and Sustainable Energy Reviews*, *16*(3), pp.1462-1476.
- Chen, P., Yan, L., Wu, Z., Li, S., Bai, Z., Yan, X. Wang, N. and Li, H. (2016). A microbial transformation using *Bacillus subtilis* B7-S to produce natural vanillin from ferulic acid. *Scientific Reports*, 6, 1-10.
- ^aChen, H. (2013). Modern solid state fermentation: theory and practice. Springer.
- ^bChen, S., Wang, D. and Xu, Y. (2013). Characterization of odor-active compounds in sweet-type Chinese rice wine by aroma extract dilution analysis with special emphasis on sotolon. *Journal of agricultural and food chemistry*, *61*(40), pp.9712-9718.
- ^aCheng, J., Li, Q.S. and Xiao, R.C. (2008). A new artificial neural network-based response surface method for structural reliability analysis. *Probabilistic Engineering Mechanics*, 23(1), pp.51-63.
- ^bCheng, Z.S., Tang, W.C., Su, Z.J., Cai, Y., Sun, S.F., Chen, Q.J., Wang, F.H., Lin, Y.C., She, Z.G. and Vrijmoed, L.L.P. (2008). Identification of mangrove endophytic fungus 1403 (*Fusarium proliferatum*) based on morphological and molecular evidence. *Journal of Forestry Research*, 19(3), pp.219-224.

Choi, Y.W., Hyde, K.D. and Ho, W.H. (1999). Single spore isolation of fungi. Fungal diversity.

- Chopada, G.B., Singh, P. and Chandulal, K. (2015). Cultural and morphological variability among *Fusarium oxysporum f. sp. lycopersici* causing wilt of tomato in south Gujarat region. *Archives of Phytopathology and Plant Protection*, 48(2), pp.104-110.
- Coates, J. (2000). Interpretation of infrared spectra, a practical approach. *Encyclopedia of* analytical chemistry.
- Converti, A., Aliakbarian, B., Domínguez, J.M., Vázquez, G.B. and Perego, P. (2010). Microbial production of biovanillin. *Brazilian Journal of Microbiology*, *41*(3), 519-530.
- Czitrom, V. (1999). One-factor-at-a-time versus designed experiments. *The American Statistician*, 53, 126-131.
- Dal Bello, E. (2013). Vanillin production from ferulic acid with *Pseudomonas fluorescens BF13-1p4*, 1-128.
- Daniel, T. S., Roel, M. S., David, M. U., David, M.. DeM. (2006). Inhibition of spontaneous mutagenesis by vanillin and cinnamaldehyde in *Escherichia coli*: Dependence on recombinational repair, *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 602 (1–2), 54-64.
- Daniel, T. S., Woodrow, R. S., David, M. DeM. (2001). The antimutagenic effect of vanillin and cinnamaldehyde on spontaneous mutation in *Salmonella TA104* is due to a reduction in mutations at GC but not AT sites, *Mutation Research/Fundamental and Molecular Mechanisms of Mutagenesis*, 480–481, 55-69.
- Das, L., Raychaudhuri, U. and Chakraborty, R. (2012). Effect of baking conditions on the physical properties of herbal bread using RSM. *International Journal of Food, Agriculture and Veterinary Sciences*, 2(2), pp.106-114.
- del Carmen Vazquez-Briones, M., Hernández, L.R. and Guerrero-Beltrán, J.Á. (2015). Physicochemical and antioxidant properties of *Cymbopogon citratus* essential oil. *Journal* of Food Research, 4(3), p.36.
- De Faveri, D., Torre, P., Aliakbarian, B., Domínguez, J. M., Perego, P., & Converti, A. (2007). Response surface modeling of vanillin production by *Escherichia coli* JM109pBB1. *Biochemical Engineering Journal*, *36*(3), 268-275.
- de Oliveira, J.M.P.F. and de Graaff, L.H. (2011). Proteomics of industrial fungi: trends and insights for biotechnology. *Applied microbiology and biotechnology*, 89(2), pp.225-237.
- da Silva, E.B., Zabkova, M., Araújo, J.D., Cateto, C.A., Barreiro, M.F., Belgacem, M.N. and Rodrigues, A.E. (2009). An integrated process to produce vanillin and lignin-based polyurethanes from Kraft lignin. *Chemical Engineering Research and Design*, 87(9), 1276-1292.

- Diba, K., Kordbacheh, P., Mirhendi, S.H., Rezaie, S. and Mahmoudi, M. (2007). Identification of Aspergillus species using morphological characteristics. *Pakistan Journal of Medical Sciences*, 23(6), p.867.
- Dignum, M.J., Kerler, J. and Verpoorte, R. (2001). Vanilla production: technological, chemical, and biosynthetic aspects. *Food Reviews International*, *17*(2), pp.119-120.
- Diaz-Godinez, G., Soriano-Santos, J., Augur, C. and Viniegra-González, G. (2001). Exopectinases produced by Aspergillus niger in solid-state and submerged fermentation: a comparative study. *Journal of industrial microbiology & biotechnology*, 26(5), pp.271-275.
- Di Blasi, C., Signorelli, G. and Portoricco, G. (1999). Countercurrent fixed-bed gasification of biomass at laboratory scale. *Industrial & engineering chemistry research*, *38*(7), pp.2571-2581.
- Di Gioia, D., Luziatelli, F., Negroni, A., Ficca, A.G., Fava, F. and Ruzzi M. (2011). Metabolic engineering of *Pseudomonas fluorescens* for the production of vanillin from ferulic acid. *J. Biotechnol.*, 156(4):309-16
- Di Gioia, D., Sciubba, L., Ruzzi, M., Setti, L. and Fava, F. (2009). Production of vanillin from wheat bran hydrolyzates via microbial bioconversion. *Journal of Chemical Technology and Biotechnology*, 84, 1441-1448.
- Di Gioia, D., Sciubba, L., Setti, L., Luziatelli, F., Ruzzi, M., Zanichelli, D. and Fava, F. (2007). Production of vanillin from wheat bran. *Journal of Enzyme and Microbial Technology*, 41, 498-505.
- Douglas, A. B. (2012). Food Flavouring Compounds, Vanillin in : *Kirk-Othmer* Food and Feed Technology. *John Wiley & Sons, Incorporated*, 2, 523-538.
- Ecott, T. (2004). Vanilla: Travels in Search of the Luscious Substance. Penguin, London.
- Ewansiha, J.U., Garba, S.A., Mawak, J.D. and Oyewole, O.A. (2012). Antimicrobial Activity of Cymbopogon citratus (Lemon Grass) and It's Phytochemical Properties.
- Falconnier, B., C. Lapierre, L. Lesage-Meessen, G. Yonnet, P. Brunerie, B. Colonna-Ceccaldi, G. Corrieu, and M. Asther. (1994). Vanillin as a product of ferulic acid biotransformation by the white-rot fungus *Pycnoporus cinnabarinus I-937*: Identification of metabolic pathways. *Journal of Biotechnology*, 37, 123-132.
- FAO-STAT (2014) FAO Statistical Database. Available; <u>http://www.fao.org</u>. Accessed 18 February 2014.
- Farnet, A.M., Criquet, S., Cigna, M., Gil, G. and Ferré, E. (2004). Purification of a laccase from *Marasmius quercophilus* induced with ferulic acid: reactivity towards natural and xenobiotic aromatic compounds. *Enzyme and Microbial Technology*, 34(6), pp.549-554.
- Faulds, C.B. and Williamson, G.(1994). Purification and characterization of a ferulic acid esterase (FAE-III) from Aspergillus niger: specificity for the phenolic moiety and binding to microcrystalline cellulose. *Microbiology*, 140(4), pp.779-787.

- Ferrochio, L., Cendoya, E., Farnochi, M.C., Massad, W. and Ramirez, M.L. (2013). Evaluation of ability of ferulic acid to control growth and fumonisin production of Fusarium verticillioides and *Fusarium proliferatum* on maize based media. *International journal of* food microbiology, 167(2), pp.215-220.
- Firdaus, M.Y.N., Osman, H., Metselaar, H.S. and Rozyanty, A.R. (2016). Preparation and Characterization of Active SiO2 from *Cymbopogon citratus* Ash Calcined at Different Temperature. *BioResources*, 11(1), pp.2839-2849.
- Figoli, A., Cassano, A. and Basile, A. (2016). Membrane Technologies for Biorefining. *Woodhead Publishing*, pp. 1-483.
- Flachowsky, G., Peyker, W., Schneider, A. and Henkel, K. (1993). Fibre analyses and in sacco degradability of plant fractions of two corn varieties harvested at various times. *Animal Feed Science and Technology*, 43(1-2), pp.41-50.
- Fleige, C., Hansen, G., Kroll, J., & Steinbüchel, A. (2013). Investigation of the Amycolatopsis sp. Strain ATCC 39116 Vanillin Dehydrogenase and Its Impact on the Biotechnical Production of Vanillin. *Applied and Environmental Microbiology*, 79(1), 81-90.
- Francisco, V., Figueirinha, A., Costa, G., Liberal, J., Lopes, M.C., García-Rodríguez, C., Geraldes, C.F., Cruz, M.T. and Batista, M.T. (2014). Chemical characterization and antiinflammatory activity of luteolin glycosides isolated from lemongrass. *Journal of functional foods*, 10, pp.436-443.
- Frost, J.W., Board of Directors operating Michigan State University. (2002). Synthesis of vanillin from a carbon source. U.S. Patent, 6,372,461.
- Gallage N.J. and Møller B.L. (2015). Vanillin–Bioconversion and Bioengineering of the Most Popular Plant Flavor and Its *De Novo* Biosynthesis in the Vanilla Orchid. *Mol. Plant.* 8, 40–57.
- Gams, W., Bissett, J. and Malik, A.F. (1998). *Morphology and identification of Trichoderma* (pp. 3-34).
- Gama, R., Van Dyk, J.S., Burton, M.H. and Pletschke, B.I. (2017). Using an artificial neural network to predict the optimal conditions for enzymatic hydrolysis of apple pomace. *3 Biotech*, *7*(2), p.138.
- Gañán, P., Cruz, J., Garbizu, S., Arbelaiz, A. and Mondragon, I. (2004). Stem and bunch banana fibers from cultivation wastes: Effect of treatments on physico-chemical behavior. *Journal of Applied Polymer Science*, *94*(4), pp.1489-1495.
- Gangopadhyay, N., Hossain, M.B., Rai, D.K. and Brunton, N.P. (2015). A review of extraction and analysis of bioactives in oat and barley and scope for use of novel food processing technologies. *Molecules*, 20(6), pp.10884-10909.
- Garai, D. and Kumar, V. (2013). A Box–Behnken design approach for the production of xylanase by Aspergillus candidus under solid state fermentation and its application in

saccharification of agro residues and Parthenium hysterophorus L. Industrial Crops and Products, 44, pp.352-363.

- Gawande, P.V. and Kamat, M.Y. (1999). Production of xylanase (*Aspergillus*) by lignocellulosic waste fermentation and its application. *Journal of Applied Microbiology*, 87(4), pp.511-519.
- Ghaffari, A., Abdollahi, H., Khoshayand, M.R., Bozchalooi, I.S., Dadgar, A. and Rafiee-Tehrani, M. (2006). Performance comparison of neural network training algorithms in modeling of bimodal drug delivery. *International journal of pharmaceutics*, 327(1-2), pp.126-138.
- Ghasemzadeh, A., Jaafar, H.Z., Rahmat, A., Wahab, P.E.M. and Halim, M.R.A. (2010). Effect of different light intensities on total phenolics and flavonoids synthesis and anti-oxidant activities in young ginger varieties (Zingiber officinale Roscoe). *International journal of* molecular sciences, 11(10), pp.3885-3897.
- Ghanem, N.B., Yusef, H.H. and Mahrouse, H.K. (2000). Production of *Aspergillus terreus* xylanase in solid-state cultures: application of the Plackett–Burman experimental design to evaluate nutritional requirements. *Bioresource Technology*, 73(2), pp.113-121.
- Ghobadian, B., Rahimi, H., Nikbakht, A.M., Najafi, G. and Yusaf, T.F. (2009). Diesel engine performance and exhaust emission analysis using waste cooking biodiesel fuel with an artificial neural network. *Renewable energy*, *34*(4), pp.976-982.
- Ghosh, S., Sanchan, A. and Mitra, A. (2005). Degradation of ferulic acid by white rot fungus *Schizophyllum commune. Journal of Microbiology and Biotechnology Advances*, 21, 385-388.
- Goralski, M., Benoist, A., Baptiste, A., Boudjema, V., Galanos, T., Georget, M., Hévin, J.E., Lavergne, S., Eychenne, F., Liew, K.E. and Schwob, C. (2015). Sustainability of biojetfuel in Malaysia. CIRAD. pp. 1-36.
- Gottipati, R. and Mishra, S. (2010). Process optimization of adsorption of Cr (VI) on activated carbons prepared from plant precursors by a two-level full factorial design. *Chemical Engineering Journal*, *160*(1), pp.99-107.
- Gould, G. W. (1996). Industry perspectives on the use of natural antimicrobials and inhibitors for food applications. *Journal Food* Protect, 82-86.
- Grbić-Galić, D. and La Pat-Polasko, L. (1985). Enterobacter cloacae DG-6: a strain that transforms methoxylated aromatics under aerobic and anaerobic conditions. *Current Microbiology*, *12*(6), pp.321-324.
- Green, J.M. (1996). Peer reviewed: a practical guide to analytical method validation. *Analytical Chemistry*, 68(9), pp.305A-309A.
- Gupta, R.B. and Demirbas, A. (2010). *Gasoline, diesel, and ethanol biofuels from grasses and plants. Cambridge University Press.*

- Guretzky, J.A., Biermacher, J.T., Cook, B.J., Kering, M.K. and Mosali, J. (2011). Switchgrass for forage and bioenergy: harvest and nitrogen rate effects on biomass yields and nutrient composition. *Plant and Soil*, *339*(1-2), pp.69-81.
- Gustafson, D.L., Franz, H.R., Ueno, A.M., Smith, C.J., Doolittle, D.J., Waldren, C.A. (2001). Vanillin (3-methoxy-4-hydroxybenzaldehyde) inhibits mutation induced by hydrogen peroxide, N-methyl-N-nitrosoguanidine and mitomycin C but not (137) Cs gammaradiation at the CD59 locus in human-hamster hybrid A(L) cells. *Mutagenesis*, 15(3), 207-213
- Hafizi, R., Salleh, B. and Latiffah, Z. (2013). Morphological and molecular characterization of Fusarium. solani and F. oxysporum associated with crown disease of oil palm. Brazilian Journal of Microbiology, 44(3), pp.959-968.
- Haveri, M., Taponen, S., Vuopio-Varkila, J., Salmenlinna, S. and Pyörälä, S. (2005). Bacterial genotype affects the manifestation and persistence of bovine *Staphylococcus aureus* intramammary infection. *Journal of Clinical Microbiology*, *43*(2), pp.959-961.
- Havkin-Frenkel, D. and Belanger, F.C. (2010). Handbook of Vanilla Science and Technology. *Wiley-Blackwell Incorporated*, 1, 1-365.
- Heer, D. and Sauer, U. (2008). Identification of furfural as a key toxin in lignocellulosic hydrolysates and evolution of a tolerant yeast strain. *Microbial biotechnology*, *1*(6), pp.497-506.
- Howard, R. L., Abotsi, E., Jansen van Rensburg, E. L. and Joward, S. (2003). Lignocellulose biotechnology: issues of bioconversion and enzyme production. *African Journal of Biotechnology*, 2, 602-619
- Hua, D., Ma, C., Song, L., Lin, S., Zhang, Z., Deng, Z. and Xu, P. (2007). Enhanced vanillin production from ferulic acid using adsorbent resin. *Applied Microbiology and Biotechnology*, 74(4), 783-790.
- Huang, Z., Dostal, L. and Rosazza, J.P.N. (1993). Mechanisms of ferulic acid conversions to vanillic acid and guaiacol by *Rhodotorula rubra*. *Journal of Biological Chemistry*, 268, 23954-23958.
- Iekhsan, O., Jaya, V., Mustaffa, M.A. and Rais, M.M. (1998). Antitoxin compounds from the plant extracts of *Ipomea pes-caprae* (L.) R. Br. In 8th FAOBMB Congress. Kuala Lumpur (p. 20).

International Trade Statistics Database, http://comtrade.un.org/. Access on Jan 2015.

Iwaki, A., Ohnuki, S., Suga, Y., Izawa, S. and Ohya, Y. (2013). Vanillin inhibits translation and induces messenger ribonucleoprotein (mRNP) granule formation in *Saccharomyces cerevisiae*: application and validation of high-content, image-based profiling. *PloS one*, 8(4), p.e61748.

- Jabasingh, S.A. and Nachiyar, C.V. (2011). Utilization of pretreated bagasse for the sustainable bioproduction of cellulase by *Aspergillus nidulans* MTCC344 using response surface methodology. *Industrial Crops and Products*, 34(3), pp.1564-1571.
- Jancik, F., Homolka, P., Cermak, B. and Lád, F. (2008). Determination of indigestible neutral detergent fibre contents of grasses and its prediction from chemical composition. *Czech Journal of Animal Science-UZPI (Czech Republic)*.
- Jönsson, L.J. and Martín, C. (2016). Pretreatment of lignocellulose: formation of inhibitory byproducts and strategies for minimizing their effects. *Bioresource technology*, *199*, pp.103-112.
- Jorjani, E., Chelgani, S.C. and Mesroghli, S.H. (2008). Application of artificial neural networks to predict chemical desulfurization of Tabas coal. *Fuel*, 87(12), pp.2727-2734.
- Joshua, A.A., Usunomena, U., Lanre, A.B., Amenze, O. and Gabriel, O.A. (2012). Comparative studies on the chemical composition and antimicrobial activities of the ethanolic extracts of lemon grass leaves and stems. *Asian Journal of Medical Sciences*, 4(4), pp.145-148.
- Juhasz, T., Szengyel, Z., Reczey, K., Siika-Aho, M. and Viikari, L. (2005). Characterization of cellulases and hemicellulases produced by *Trichoderma reesei* on various carbon sources. *Process Biochemistry*, 40, 3519-3525.
- Jurková, M. and Wurst, M.(1993). Biodegradation of aromatic carboxylic acids by *Pseudomonas* mira. FEMS Microbiology Letters, 111(2-3), pp.245-250.
- Jüsten, P., Paul, G.C., Nienow, A.W. and Thomas, C.R. (1998). Dependence of *Penicillium chrysogenum* growth, morphology, vacuolation, and productivity in fed-batch fermentations on impeller type and agitation intensity. *Biotechnology and Bioengineering*, *59*(6), pp.762-775.
- Kalra, A., Kumar, S. and Waliya, S.S. (2016). ANN Training: A Survey of classical and Soft Computing Approaches. *International Journal of Control Theory and Applications*, 9(34), pp.715-736.
- Kameshwar, A.K.S. and Qin, W. (2017). Gene expression metadata analysis reveals molecular mechanisms employed by *Phanerochaete chrysosporium* during lignin degradation and detoxification of plant extractives. *Current Genetics*, pp.1-18.
- Kamisaka, S., Takeda, S., Takahashi, K. and Shibata, K. (1990). Diferulic and ferulic acid in the cell wall of Avena coleoptiles—Their relationships to mechanical properties of the cell wall. *Physiologia Plantarum*, 78(1), 1-7.
- ^aKaur, B. and Chakraborty, D. (2013). Biotechnological and molecular approaches for vanillin production: a review. *Applied Biochemistry and Biotechnology*, *169*(4), pp.1353-1372.
- ^bKaur, B. and Chakraborty, D. (2013). Statistical media and process optimization for biotransformation of rice bran to vanillin using *Pediococcus acidilactici*. *Indian Journal of Experimental Biology*. 51, 935-943.

- Kaur, H. and Dutt, D. (2013). Anatomical, morphological and chemical characterization of lignocellulose by-products of lemon and sofia grasses obtained after recuperation of essential oils by steam distillation. *Cellulose Chemistry and Technology*, 47(1-2), 83-94.
- Karode, B., Patil, U. and Jobanputra, A. (2013). Biotransformation of low cost lignocellulosic substrates into vanillin by white rot fungus, *Phanerochaete chrysosporium* NCIM 1197.
- Kim, H.J., Gao, W., Chung, C.H. and Lee, J.W. (2011). Statistical optimization for production of carboxymethylcellulase from rice hulls by a newly isolated marine microorganism *Bacillus licheniformis* LBH-52 using response surface method. *Journal of Life Science*, 21(8), pp.1083-1093.
- Kimbrough, R.A., Ruppert, G.P., Wiggins, W.D., Smith, R.R. and Kresch, D.L. (2006). Water resources data-Washington water year 2005 (No. WA-05-1). U. S. Geological Survey.
- Kimi, M., Yuliati, L. and Shamsuddin, M. (2015). Preparation of high activity Ga and Cu doped ZnS by hydrothermal method for hydrogen production under visible light irradiation. *Journal of Nanomaterials*, 16(1), p.200.
- Kirk, T.K. and Farrell, R.L. (1987). Enzymatic combustion: the microbial degradation of lignin. *Annual Reviews in Microbiology*, 41(1), pp.465-501.
- Kirk, T. K., Tien, M., Kersten, P. J., Mozuch, M. D. and Kalyanaraman, B. (1986). Ligninase of *Phanerochaete chrysosporium*. Mechanism of its degradation of the non-phenolic arylglycerol beta-aryl ether substructure of lignin. *Biochemical Journal*, 236, 279-287
- Kirwin, C.J., Galvin, J.B. (1993). Ethers. In: Clayton, G.D., Clayton, F.E. (Eds.), Patty's Industrial Hygiene and Toxicology, 4th edition. *John Wiley & Sons, New York*, Vol. (2), 445–525.
- Konstantinović, S.S., Konstantinović, B.V. and Jovanović, J.M. (2009). Synthesis and structure of vanillin azomethines. *Chemical Industry and Chemical Engineering Quarterly/CICEQ*, 15(4), pp.279-281.
- Krimitzas, A., Pyrri, I., Kouvelis, V.N., Kapsanaki-Gotsi, E. and Typas, M.A. (2013). A phylogenetic analysis of Greek isolates of *Aspergillus* species based on morphology and nuclear and mitochondrial gene sequences. *BioMed research international*, 2013.
- Krings, U., Pilawa, S., Theobald, C. and Berger, R.G. (2001). Phenyl propenoic side chain degradation of ferulic acid by *Pycnoporus cinnabarinus*—elucidation of metabolic pathways using [5-2 H]-ferulic acid. *Journal of Biotechnology*, 85(3), pp.305-314.
- Krings, U. and Berger, R. G. (1998). Biotechnological production of flavours and fragrances. *Journal of Applied Microbiology Biotechnology*, 49, 1-8.
- Kumar, N. and Pruthi, V. (2014). Potential applications of ferulic acid from natural sources. *Biotechnology Reports*, 4, pp.86-93.
- Kumar, G.S., Nayaka, H., Dharmesh, S.M. and Salimath, P.V. (2006). Free and bound phenolic antioxidants in amla (*Emblica officinalis*) and turmeric (*Curcuma longa*). Journal of Food Composition and Analysis, 19(5), pp.446-452.

- Kumar, R., Sharma, P.K. and Mishra, P.S. (2012). A review on the vanillin derivatives showing various biological activities. *International Journal of Pharmaceutical Technology Research*, 4(1), pp.266-279.
- Kumar, S. and Kaushik, N. (2013). Endophytic fungi isolated from oil-seed crop Jatropha curcas produces oil and exhibit antifungal activity. *PloS one*, 8(2), p.e56202.
- Kumar, S. and Pandey, A.K. (2013). Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*, 2013. Article ID 162750 pp.1-16
- Larone, D.H. and Larone, D.H. (1987). Medically important fungi: a guide to identification. *New York: Elsevier*. Vol. 189.
- Leffingwell, J. & Leffingwell, D. (2015). Flavours and fragrances: Recent advances in biotechnology. *Speciality Chemicals Magazine*, 35 (5), 32-34.
- Lenth, R.V. (2009). Response-Surface Methods in R, using RSM. Journal of Statistical Software, 32(7), 1-17.
- Lesage-Meessen, L., Lomascolo, A., Bonnin, E., Thibault, J. F., Buleon, A., Roller, M. and Asther, M. (2002). A biotechnological process involving filamentous fungi to produce natural crystalline vanillin from maize bran. *AppliedBbiochemistry and Biotechnology*, 102(1-6), 141-153.
- Lesage-Meessen, L., Stentelaire, C., Lomascolo, A., Couteau, D., Asther, M., Moukha, S., Record, E., Sigoillot, J.C. and Asther, M. (1999). Fungal transformation of ferulic acid from sugar beet pulp to natural vanillin. *Journal of the Science of Food and Agriculture*, 79(3), pp.487-490.
- Lesage-Meessen, L., Haon, M., Delattre, M., Thibault, J.F., Ceccaldi, B.C. and Asther, M. (1997). An attempt to channel the transformation of vanillic acid into vanillin by controlling methoxyhydroquinone formation in *Pycnoporus cinnabarinus* with cellobiose. *Applied microbiology and biotechnology*, 47(4), pp.393-397.
- Lesage-Meessen, L., Delattre, M., Haon, M., Thibault, J.F., Ceccaldi, B.C., Brunerie, P. and Asther, M. (1996). A two steps bioconversion process for vanillin production from ferulic acid combining *Aspergillus niger* and *Pycnoporus cinnabarinus*. *Journal of Biotechnology*, 50(2), pp.107-113.
- Lewinsohn, E., Dudai, N., Tadmor, Y., Katzir, I., Ravid, U.Z.I., Putievsky, E.L.I. and Joel, D.M. (1998). Histochemical localization of citral accumulation in lemongrass leaves (Cymbopogon citratus (DC.) Stapf., Poaceae). *Annals of Botany*, *81*(1), pp.35-39.
- Liew S. L., Osman H., Lee Y.H., Mustapha, W. A. W. and Maskat, M. Y. (2010). Synthesis of bio-vanillin from lignin of palm oil empty fruit bunch. *Directory of Research Projects of* the Department of Higher Education of the Ministry of Higher Education 2010.
- Li, K. and Frost, J.W. (1998). Synthesis of vanillin from glucose. *Journal of the American Chemical Society*, 120(40), pp.10545-10546.

- Li, T. and Rosazza, J.P. (2000). Biocatalytic synthesis of vanillin. *Applied and Environmental Microbiology*, 66(2), pp.684-687.
- Li, X., Yang, J., Li, X., Gu, W., Huang, J. and Zhang, K.Q. (2008). The metabolism of ferulic acid via 4-vinylguaiacol to vanillin by Enterobacter sp. Px6-4 isolated from Vanilla root. *Process Biochemistry*, 43(10), pp.1132-1137.
- Li, Y., Cui, F., Liu, Z., Xu, Y. and Zhao, H. (2007). Improvement of xylanase production by Penicillium oxalicum ZH-30 using response surface methodology. *Enzyme and Microbial Technology*, 40(5), pp.1381-1388.
- Liguori, R., Amore, A. and Faraco, V. (2013). Waste valorization by biotechnological conversion into added value products. *Applied Microbiology and Biotechnology*, 97(14), pp.6129-6147.
- Lin, L., Yan, R., Liu, Y. and Jiang, W. (2010). In-depth investigation of enzymatic hydrolysis of biomass wastes based on three major components: cellulose, hemicellulose and lignin. *Bioresource Technology*, 101, 8217-8223.
- Liu, Q., Wang, S., Zheng, Y., Luo, Z. and Cen, K. (2008). Mechanism study of wood lignin pyrolysis by using TG–FTIR analysis. *Journal of Analytical and Applied Pyrolysis*, 82(1), pp.170-177.
- Lomascolo, A., Stentelaire, C., Asther, M. and Lesage-Meessen. L. (1999). Basidiomycetes as new biotechnological tools to generate natural aromatic flavours for the food industry. *Trends in Biotechnology*, 17:282-289.
- Longo, M.A. and Sanromán, M.A. (2006). Production of food aroma compounds: microbial and enzymatic methodologies. *Food Technology and Biotechnology*, 44(3), pp.335-353.
- Lopez-Malo, A., Alzamora, S.M. and Argaiz, A. (1997). Effect of vanillin concentration, pH and incubation temperature on *Aspergillus flavus*, *Aspergillus niger*, *Aspergillus ochraceus* and *Aspergillus parasiticus* growth. *Food Microbiology*, *14*(2), pp.117-124.
- López-Malo, A., Alzamora, S.M. and Argaiz, A. (1995). Effect of natural vanillin on germination time and radial growth of moulds in fruit-based agar systems. *Food Microbiology*, 12, pp.213-219.
- Lun, O.K., Wai, T.B. and Ling, L.S. (2014). Pineapple cannery waste as a potential substrate for microbial biotranformation to produce vanillic acid and vanillin. *International Food Research Journal*, 21(3).
- Lupi, E., Christoph, H. and Patricia, S. (2013). The efficacy of repellents against Aedes, Anopheles, Culex and Ixodes spp.- A literature review. Travel medicine and infectious disease, 11(6), 374-411.
- Lupoi, J.S., Singh, S., Parthasarathi, R., Simmons, B.A. and Henry, R.J. (2015). Recent innovations in analytical methods for the qualitative and quantitative assessment of lignin. *Renewable and Sustainable Energy Reviews*, 49, pp.871-906.

- Ma, X. K., and Daugulis, A. J. (2014). Effect of bioconversion conditions on vanillin production by *Amycolatopsis sp.* ATCC 39116 through an analysis of competing by-product formation. *Bioprocess and Biosystems Engineering*, 37(5), 891-899.
- Machado, M.S.S., Silva, H.B.F., Rios, R., de Oliveira, A.P., Carneiro, N.V.Q., Costa, R.S., Alves, W.S., Souza, F.L.M., Velozo, E.S., de Souza, S.A. and Silva, T.M.S. (2015). The anti-allergic activity of *Cymbopogon citratus* is mediated via inhibition of nuclear factor kappa B (Nf-Kb) activation. *BMC Complementary and Alternative Medicine*, *15*(1), p.168.
- Mais, U., Esteghlalian, A.R., Saddler, J.N. and Mansfield, S.D. (2002). Enhancing the enzymatic hydrolysis of cellulosic materials using simultaneous ball milling. In *Biotechnology for Fuels and Chemicals* (pp. 815-832). Humana Press, Totowa, NJ.
- Maity, A., Pal, R.K., Chandra, R. and Singh, N.V. (2014). *Penicillium pinophilum*—A novel microorganism for nutrient management in pomegranate (*Punica granatum L.*). *Scientia Horticulturae*, 169, pp.111-117.
- Mandal, S., Sivaprasad, P.V., Venugopal, S. and Murthy, K.P.N. (2009). Artificial neural network modeling to evaluate and predict the deformation behavior of stainless steel type AISI 304L during hot torsion. *Applied Soft Computing*, 9(1), pp.237-244.
- Mani, S., Tabil, L.G. and Sokhansanj, S. (2006). Effects of compressive force, particle size and moisture content on mechanical properties of biomass pellets from grasses. *Biomass and Bioenergy*, 30(7), pp.648-654.
- Marini, F. (2009). Artificial neural networks in foodstuff analyses: Trends and perspectives A review. *Analytica Chimica Acta*, 635(2), pp.121-131.
- Markus, P.H, Peters, A.L.J., Roos, R. (1992). Process for the preparation of phenylaldehydes. *European Patent Application*, EP 0 542 348 A2.
- Martinez, D., Larrondo, L.F., Putnam, N., Gelpke, M.D.S., Huang, K., Chapman, J., Helfenbein, K.G., Ramaiya, P., Detter, J.C., Larimer, F. and Coutinho, P.M. (2004). Genome sequence of the lignocellulose degrading fungus *Phanerochaete chrysosporium* strain RP78. *Nature Biotechnology*, 22(6), pp.695-700.
- Marzouk, B. (2013). Characterization of bioactive compounds in Tunisian bitter orange (*Citrus aurantium L.*) peel and juice and determination of their antioxidant activities. *BioMedical Research international*, 2013.
- Mathew, S. and Abraham, T.E. (2006) Bioconversions of ferulic acid, an hydroxycinnamic acid. *Critical Reviews in Microbiology*, *32*(3), pp.115-125.
- Mathew, S., and Abraham, T. E. (2004). Ferulic acid: an antioxidant found naturally in plant cell walls and feruloyl esterases involved in its release and their applications. *Critical Reviews in Biotechnology*, 24, 59-83.
- Matsuura, S. and Miyazima, S. (1994). Colony morphology of the fungus *Aspergillus oryzae*. In *Fractals in Biology and Medicine*. pp. 274-282. Birkhäuser Basel.

- McGinnis, M.R. and Tyring, S.K., (1996). Introduction to mycology. *Medical Microbiology*. *University of Texas Medical Branch at Galveston, Galveston, TX, https://www. ncbi. nlm. nih. gov/books/NBK8471/McKone, TE*, pp.165-181.
- McLean, K.J., Leys, D. and Munro, A.W. (2015). Microbial cytochromes P450. In *Cytochrome* P450 (pp. 261-407). Springer International Publishing.
- Mechichi, T., Labat, M., Garcia, J.L., Thomas, P. and Patel, B.K. (1999). Sporobacterium olearium gen. nov., sp. nov., a new methanethiol-producing bacterium that degrades aromatic compounds, isolated from an olive mill wastewater treatment digester. *International Journal of Systematic and Evolutionary Microbiology*, *49*(4), pp.1741-1748.
- Mélida, H., Álvarez, J., Acebes, J.L., Encina, A. and Fry, S.C. (2011). Changes in cinnamic acid derivatives associated with the habituation of maize cells to dichlobenil. *Molecular Plant*, 4(5), pp.869-878.
- Mihali, C.V., Buruiana, A., Turcus, V., Covaci, A. and Ardelean, A. (2011). Morphological aspects of fruiting bodies in *Microsporum gypseum* on Saboureauds dextrose agar medium *Annmedium als of the Romanian Society for Cell Biology*, *16*(2).
- Ministry of Agriculture and Agro-based Industry (2015). National Agro-food Policy (2011-2020). Ministry of Agriculture and Agro-food Industry. Kuala Lumpur.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, 31, 426-428.
- Ministry of Agriculture and Agro-Based Industry Malaysia Report: Agrofood & Crops Statistic (2015). http://www.moa.gov.my/documents/10157/bcf8ee39-938b-4748-93a8-363e74529bcb
- Mitchell, D.A., Berovič, M. and Krieger, N. (2006). Solid-state fermentation bioreactor fundamentals: introduction and overview. In *Solid-State Fermentation Bioreactors* (pp. 1-12). Springer, Berlin, Heidelberg.
- Moghaddam, M.G., Ahmad, F.B.H., Basri, M. and Rahman, M.B.A. (2010). Artificial neural network modeling studies to predict the yield of enzymatic synthesis of betulinic acid ester. *Electronic Journal of Biotechnology*, *13*(3), pp.3-4.
- Mohamad Yusof, N.N. and Kobayashi, T. (2013). Efficient separation on vanillin operated with permeability performance of hollow fiber membranes embedded vanillin imprinted polymer particles. *Industrial & Engineering Chemistry Research*, 52(47), pp.16951-16957.

Montgomery, D.C. (1984). Design and analysis of experiments. New York: Wiley.

Motedayen, N., Ismail, M.B. and Nazarpour, F. (2013). Bioconversion of ferulic acid to vanillin by combined action of *Aspergillus niger K8* and *Phanerochaete crysosporium ATCC 24725*. *African Journal of Biotechnology*, *12*(47), pp.6618-6624.

- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y.Y., Holtzapple, M. and Ladisch, M. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology*, 96(6), pp.673-686.
- Mtui, G.Y. (2009). Recent advances in pretreatment of lignocellulosic wastes and production of value added products. *African Journal of Biotechnology*,8(8).
- Muchuweti, M., Kativu, E., Mupure, C.H., Chidewe, C., Ndhlala, A.R. and Benhura, M.A.N. (2007). Phenolic composition and antioxidant properties of some spices. *American Journal of Food Technology*, 2(5), pp.414-420.
- Müller, B., Reinhardt, J. and Strickland, M.T. (2012). *Neural networks: an introduction*. Springer Science & Business Media.
- Muheim, A., and Lerch, K. (1999). Towards a high-yield bioconversion of ferulic acid to vanillin. *Applied Microbiology and Biotechnology*, 51, 456-461.
- Mussatto, S. I., Dragone, G. and Roberto, I. C. (2007). Ferulic and p-coumaric acids extraction by alkaline hydrolysis of brewer's spent grain. *Industrial Crops and Products*, 25, 231-237.
- Myers, R.H. and Montgomery, D.C. (2002). Response Surface Methodology, John Wiley & Sons. *Inc.*, USA.
- Nambiar, V.S. and Matela, H. (2012). Potential functions of lemongrass (*Cymbopogon citratus*) in health and disease. *International Journal of Pharmaceutical and Biological Archives*, *3*(5), pp.1035-1043.
- Nampoothiri, S.V., Esakkidurai, T. and Pitchumani, K. (2015). Identification and quantification of phenolic compounds in Alpinia galanga and Alpinia calcarata and its relation to free radical quenching properties: a comparative study. *Journal of Herbs, Spices & Medicinal Plants*, 21(2), pp.140-147.
- Nandy, S., Sarkar, P.P. and Das, A. (2012). Training a feed-forward neural network with artificial bee colony based backpropagation method. *arXiv preprint arXiv:1209.2548*.
- Narbad, A., and Gasson, M. J. (1998). Metabolism of ferulic acid to vanillin using a novel CoA dependent pathway in a newly isolated strain *Pseudomonas fluorescens*. *Microbiology*, 144, 1397-1405.
- Nardini, M., Cirillo, E., Natella, F., Mencarelli, D., Comisso, A. and Scaccini, C. (2002). Detection of bound phenolic acids: prevention by ascorbic acid and ethylenediaminetetraacetic acid of degradation of phenolic acids during alkaline hydrolysis. *Food Chemistry*, 79(1), pp.119-124.
- Nazareth, S. and Mavinkurve, S. (1986). Degradation of ferulic acid via 4-vinylguaiacol by Fusarium solani (Mart.) Sacc. *Canadian Journal of Microbiology*, *32*(6), pp.494-497.
- Negrelle, R.R.B. and Gomes, E.C. (2007). *Cymbopogon citratus* (DC.) Stapf: chemical composition and biological activities. *Rev Bras Pl Med*, 9(1), pp.80-92.

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- Nelson, P.E., Toussoun, T.A. and Marasas, W.F.O. (1983). *Fusarium* species: an illustrated manual for identification.
- Nickell, L.G. (1976). Vanillin as ripener for sugarcane. United States Patent, 1-6.
- Nigam, P. S. and Pandey, A. (2009). Biotechnology for agro-industrial residues utilisation: utilisation of agro-residues. Springer.
- Noor Hasyierah, M.S., Mohamed Zulkali, M.D. and Ku Syahidah Ku, I. (2008). Ferulic acid from lignocellulosic biomass: review. *Proceeding of MUCET2008* p. 1-8
- Ofori-Boateng, C. and Lee, K.T. (2014). Ultrasonic-assisted simultaneous saccharification and fermentation of pretreated oil palm fronds for sustainable bioethanol production. *Fuel*, *119*, pp.285-291.
- Oloyede, O.I. (2009). Chemical profile and antimicrobial activity of Cymbopogon citratus leaves. *Journal of Natural Products (India)*, 2, pp.98-103.
- Ou, S.Y., Luo, Y.L., Huang, C.H. and Jackson, M. (2009). Production of coumaric acid from sugarcane bagasse. *Innovative Food Science & Emerging Technologies*, 10(2), pp.253-259.
- Ou, S., Luo, Y., Xue, F., Huang, C., Zhang, N. and Liu, Z. (2007). Separation and purification of ferulic acid in alkaline-hydrolysate from sugarcane bagasse by activated charcoal adsorption/anion macroporous resin exchange chromatography. *Journal of Food Engineering*, 78(4), pp.1298-1304.
- Overhage, J., Steinbüchel, A. and Priefert, H. (2003). Highly efficient biotransformation of eugenol to ferulic acid and further conversion to vanillin in recombinant strains of *Escherichia coli*. *Applied and Environmental Microbiology*, 69(11), pp.6569-6576.
- Overhage, J., Steinbüchel, A. and Priefert, H. (2002). Biotransformation of eugenol to ferulic acid by a recombinant strain of Ralstonia eutropha H16. *Applied and Environmental Microbiology*, 68(9), pp.4315-4321.
- Overhage, J., Priefert, H., Rabenhorst, J. and Steinbüchel, A. (1999). Biotransformation of eugenol to vanillin by a mutant of *Pseudomonas sp. strain HR199* constructed by disruption of the vanillin dehydrogenase (vdh) gene. *Applied Microbiology and Niotechnology*, 52(6), pp.820-828.
- Özgen, S. and Yildiz, A. (2010). Application of Box–Behnken design to modeling the effect of smectite content on swelling to hydrocyclone processing of bentonites with various geologic properties. *Clays and Clay Minerals*, *58*(3), pp.431-448.
- Pandey, A. and Mann, M. (2000). Proteomics to study genes and genomes. *Nature*, 405(6788), pp.837-846.
- Pandey, K.K. and Pitman, A.J. (2003). FTIR studies of the changes in wood chemistry following decay by brown-rot and white-rot fungi. *International biodeterioration & biodegradation*, *52*(3), pp.151-160.

- Panthapulakkal, S., Zereshkian, A. and Sain, M. (2006). Preparation and characterization of wheat straw fibers for reinforcing application in injection molded thermoplastic composites. *Bioresource technology*, *97*(2), pp.265-272.
- Papagianni, M. and Mattey, M. (2006). Morphological development of *Aspergillus niger* in submerged citric acid fermentation as a function of the spore inoculum level. Application of neural network and cluster analysis for characterization of mycelial morphology. *Microbial Cell Factories*, 5(1), p.1.
- Papagianni, M. (2004). Fungal morphology and metabolite production in submerged mycelial processes. *Biotechnology advances*, 22(3), pp.189-259.
- Paterson, D. (2010). Vanilla: natural or not. Chemistry Education in New Zealand, pp.1-5.
- Pérez-Rodríguez, N., de Souza Oliveira, R.P., Agrasar, A.M.T. and Domínguez, J.M. (2016). Ferulic acid transformation into the main vanilla aroma compounds by Amycolatopsis sp. ATCC 39116. *Applied microbiology and biotechnology*, *100*(4), pp.1677-1689.
- Phillips, T. (2008). Genetically modified organisms (GMOs): Transgenic crops and recombinant DNA technology. *Nature Education*, 1(1), p.213.
- Pinjare, S.L. and Kumar, A. (2012). Implementation of neural network back propagation training algorithm on FPGA. *International Journal of Computer Applications*, 52(6).
- Pino-García, O. (2004). Influence of Admixtures on Crystal Nucleation of Vanillin. *Doctoral dissertation. Stockholm: Kemiteknik.*
- Pinto, P.C.R., da Silva, E.A.B. and Rodrigues, A.E. (2012). Lignin as source of fine chemicals: vanillin and syringaldehyde. In *Biomass Conversion, Springer Berlin Heidelberg*, pp. 381-420.
- Plaggenborg, R., Overhage, J., Loos, A., Archer, J.A., Lessard, P., Sinskey, A.J., Steinbüchel, A. and Priefert, H. (2006). Potential of Rhodococcus strains for biotechnological vanillin production from ferulic acid and eugenol. *Applied microbiology and biotechnology*, 72(4), p.745.
- Plaggenborg, R., Steinbüchel, A. and Priefert, H. (2001). The coenzyme A-dependent, non-βoxidation pathway and not direct deacetylation is the major route for ferulic acid degradation in Delftia acidovorans. *FEMS microbiology letters*, 205(1), pp.9-16.
- Prasad, S., Singh, A. and Joshi. H. C. (2007). Ethanol as an alternative fuel from agricultural, industrial and urban residues. *Resources, Conservation and Recycling*, 50, 1-39.
- Prasertsan, P., H-kittikul, A. and Chitmanee, B. (1992). Isolation and selection of cellulolytic fungi from palm oil mill effluent. *World Journal of Microbiology and Biotechnology*, 8, 614-617.
- Priefert, H., Rabenhorst, J., and Steinbuchel, A. (2001). Biotechnology Production of Vanillin. *Applied Microbiology Biotechnology*, 56, 296-314.

- Pua, F.L., Zakaria, S., Chia, C.H., Fan, S.P., ROSEnAU, T., POTThAST, A. and LIEBnER, F.A.L.K. (2013). Solvolytic liquefaction of oil palm empty fruit bunch (EFB) fibres: analysis of product fractions using FTIR and pyrolysis-GCMS. Sains Malaysiana, 42(6), pp.793-799.
- Puviarasan, N., Arjunan, V. and Mohan, S. (2002). FT-IR and FT-Raman studies on 3aminophthalhydrazide and N-aminophthalimide. *Turkish Journal of Chemistry*, 26(3), pp.323-334.
- Rabenhorst, J., Hopp, R. (1991) Process for the preparation of vanillin. *Patent application* EP0405197
- Rabenhorst, J. and Hopp, R. (2000). Process for the preparation of vanillin and microorganisms suitable therefor. *U.S. patent* 6,133,003.
- Ragauskas, A.J., Beckham, G.T., Biddy, M.J., Chandra, R., Chen, F., Davis, M.F., Davison, B.H., Dixon, R.A., Gilna, P., Keller, M. and Langan, P. (2014). Lignin valorization: improving lignin processing in the biorefinery. *Science*, 344(6185), p.1246843.
- Ralph, J., Quideau, S., Grabber, J. H. and Hatfield, R. D. (1994). Identification and synthesis of new ferulic acid dehydrodimers present in grass cell walls. *Journal of the Chemical Society, Perkin Transactions*, 1, 3485-3498.
- Rao, S.R. and Ravishankar, G.A. (2000). Vanilla flavour: production by conventional and biotechnological routes. *Journal of the Science of Food and Agriculture*, 80(3), 289-304.
- Ravindran, R. and Jaiswal, A.K. (2016). A comprehensive review on pre-treatment strategy for lignocellulosic food industry waste: challenges and opportunities. *Bioresource Technology*, 199, pp.92-102.
- Razavi, S.M.A., Mousavi, S.M. and Mortazavi, S.A. (2003). Dynamic prediction of milk ultrafiltration performance: A neural network approach. *Chemical Engineering Science*, 58(18), pp.4185-4195.
- Rinaldi, R., Jastrzebski, R., Clough, M.T., Ralph, J., Kennema, M., Bruijnincx, P.C. and Weckhuysen, B.M. (2016). Paving the way for lignin valorisation: recent advances in bioengineering, biorefining and catalysis. *Angewandte Chemie International Edition*, 55(29), pp.8164-8215.
- Ristolainen, M., Alén, R., Malkavaara, P. and Pere, J. (2002). Reflectance FTIR microspectroscopy for studying effect of xylan removal on unbleached and bleached birch kraft pulps. *Holzforschung*, *56*(5), pp.513-521.
- Rolz, C., De Leon, R., De Arriola, M.C. and De Cabrera, S. (1986). Biodelignification of lemon grass and citronella bagasse by white-rot fungi. *Applied and environmental microbiology*, 52(4), pp.607-611.
- Rosazza, J.P.N., Huang, Z., Dostal, L., Volm, T. and Rousseau, B. (1995). Review: biocatalytic transformations of ferulic acid: an abundant aromatic natural product. *Journal of Industrial Microbiology*, *15*(6), pp.457-471.

- Rose, D.J., Inglett, G.E. and Liu, S.X. (2010). Utilisation of corn (Zea mays) bran and corn fiber in the production of food components. *Journal of the Science of Food and Agriculture*, 90(6), pp.915-924.
- Sabiha-Hanim, S., Noor, M.A.M. and Rosma, A. (2011). Effect of autohydrolysis and enzymatic treatment on oil palm (*Elaeis guineensis Jacq.*) frond fibres for xylose and xylooligosaccharides production. *Bioresource Technology*, *102*(2), pp.1234-1239.
- Sacchetti, G., Maietti, S., Muzzoli, M., Scaglianti, M., Manfredini, S., Radice, M. and Bruni, R. (2005). Comparative evaluation of 11 essential oils of different origin as functional antioxidants, antiradicals and antimicrobials in foods. *Food Chemistry*, 91(4), pp.621-632.
- Sah, S.Y., Sia, C.M., Chang, S.K., Ang, Y.K. and Yim, H.S. (2012). Antioxidant capacity and total phenolic content of lemongrass (*Cymbopogon citratus*) leave. Annals Food Science and Technology, 13(2), pp.150-155.
- Saini, J.K., Saini, R. and Tewari, L. (2015). Lignocellulosic agriculture wastes as biomass feedstocks for second-generation bioethanol production: concepts and recent developments. *3 Biotech*, 5(4), pp.337-353.
- Sajjadi, S.E., Shokoohinia, Y. and Moayedi, N.S. (2012). Isolation and identification of ferulic acid from aerial parts of *Kelussia odoratissima Mozaff. Jundishapur Journal of Natural Pharmaceutical Products*, 7(4), pp.159-162.
- Sangnark, A. and Noomhorm, A. (2004). Effect of dietary fiber from sugarcane bagasse and sucrose ester on dough and bread properties. *LWT-Food Science and Technology*, *37*(7), pp.697-704.
- Sarangi, P.K. and Sahoo, H.P. (2010). Ferulic acid production from wheat bran using *Staphylococcus aureus. New York Science Journal*, *3*, pp.79-81.
- Sarethy, I.P. and Pan, S., 2017. Designer Foods: Scope for Enrichment With Microbe-Sourced Antioxidants. In *Microbial Production of Food Ingredients and Additives* pp. 423-449.
- Saulnier, L., Vigouroux, J. and Thibault, J.F. (1995). Isolation and partial characterization of feruloylated oligosaccharides from maize bran. *Carbohydrate Research*, 272(2), pp.241-253.
- Serra, S., Fuganti, C. and Brenna, E. (2005). Biocatalytic preparation of natural flavours and fragrances. *TRENDS in Biotechnology*, 23(4), 193-198.
- Shah, G., Shri, R., Panchal, V., Sharma, N., Singh, B. and Mann, A.S. (2011). Scientific basis for the therapeutic use of *Cymbopogon citratus*, stapf (Lemon grass). *Journal of advanced pharmaceutical technology & research*, 2(1), p.3.
- Sharma, K.K. and Singh, U.S. (2014). Cultural and morphological characterization of rhizospheric isolates of fungal antagonist *Trichoderma*. *Journal of Applied and Natural Science*, *1*(6), pp.451-456.

- Shin, H.D., McClendon, S., Le, T., Taylor, F. and Chen, R.R. (2006). A complete enzymatic recoveryof ferulic acid from corn residue with extracellular enzymes from *Neosartorya* spinosa NRRL185. Biotechnology Bioengineering, 95, 1108-1115
- Singh, P., Shera, S.S., Banik, J. and Banik, R.M. (2013). Optimization of cultural conditions using response surface methodology versus artificial neural network and modeling of L-glutaminase production by *Bacillus cereus* MTCC 1305. *Bioresource technology*, *137*, pp.261-269.
- Smith, M.M. and Hartley, R.D. (1983). Occurrence and nature of ferulic acid substitution of cellwall polysaccharides in graminaceous plants. *Carbohydrate Research*, *118*, pp.65-80.
- Soares, M., Christen, P., Pandey, A. and Soccol, C.R. (2000). Fruity flavour production by Ceratocystis fimbriata grown on coffee husk in solid-state fermentation. *Process Biochemistry*, 35(8), pp.857-861.
- Srivastava, S., Luqman, S., Khan, F., Chanotiya, C.S. and Darokar, M.P. (2010). Metabolic pathway reconstruction of eugenol to vanillin bioconversion in Aspergillus niger. *Bioinformation*, 4(7), 320-325.
- Srivastava, N., Hinton, G., Krizhevsky, A., Sutskever, I. and Salakhutdinov, R. (2014). Dropout: a simple way to prevent neural networks from overfitting. *The Journal of Machine Learning Research*, 15(1), pp.1929-1958
- Stentelaire, C., Lesage-Meessen, L., Oddou, J., Bernard, O., Bastin, G., Ceccaldi, B. C. and Asther, M. (2000). Design of a fungal bioprocess for vanillin production from vanillic acid at scalable level by *Pycnoporus cinnabarinus*. *Journal of Bioscience and Bioengineering* 89:223-230.
- Sun, J., Yin, Y., Sheng, G.H., Yang, Z.B. and Zhu, H.L. (2013). Synthesis, molecular modeling and structural characterization of vanillin derivatives as antimicrobial agents. *Journal of Molecular Structure*, 1039, pp.214-218.
- Su, Y., Zhang, X., Hou, Z., Zhu, X., Guo, X. and Ling, P. (2011). Improvement of xylanase production by thermophilic fungus *Thermomyces lanuginosus* SDYKY-1 using response surface methodology. *New biotechnology*, 28(1), pp.40-46.
- Sun, R. and Tomkinson, J. (2001). Fractional separation and physico-chemical analysis of lignins from the black liquor of oil palm trunk fibre pulping. *Separation and Purification Technology*, 24(3), pp.529-539.
- Sun, R.C., Sun, X.F. and Zhang, S.H. (2001). Quantitative determination of hydroxycinnamic acids in wheat, rice, rye, and barley straws, maize stems, oil palm frond fiber, and fastgrowing poplar wood. *Journal of Agricultural and Food Chemistry*, 49(11), pp.5122-5129.
- Sun, Y. and Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource Technology*, 83(1), pp.1-11.

- Suzuki, H., MacDonald, J., Syed, K., Salamov, A., Hori, C., Aerts, A., Henrissat, B., Wiebenga, A., Barry, K., Lindquist, E. and LaButti, K. (2012). Comparative genomics of the whiterot fungi, *Phanerochaete carnosa* and *P. chrysosporium*, to elucidate the genetic basis of the distinct wood types they colonize. *BMC Genomics*, 13(1), p.444.
- Syed, N.F.N., Zakaria, M.H. and Bujang, J.S. (2016). Fiber Characteristics and papermaking of seagrass using hand-beaten and blended pulp. *BioResources*, *11*(2), pp.5358-5380
- Szwajgier, D. and Jakubczyk, A. (2010). Biotransformation of ferulic acid by *Lactobacillus* acidophilus KI and selected Bifidobacterium strains. ACTA Scientiarum Polonorum Technologia Alimentaria, 9(1), pp.45-59.
- Taherzadeh, M.J. and Karimi, K. (2008). Pretreatment of lignocellulosic wastes to improve ethanol and biogas production: a review. *International journal of molecular sciences*, 9(9), pp.1621-1651.
- Talebian-Kiakalaieh, A., Amin, N.A.S., Zarei, A. and Noshadi, I. (2013). Transesterification of waste cooking oil by heteropoly acid (HPA) catalyst: optimization and kinetic model. *Applied energy*, 102, pp.283-292.
- Tan, M.C., Liew, S.L., Maskat, M.Y., Wan Aida, W.M. and Osman, H. (2015). Optimization of vanillin production using isoeugenol as substrate by Aspergillus niger I-1472. International Food Research Journal, 22(4), pp.1651-1656.
- Tang, H., Krishnakumar, V., Bidwell, S., Rosen, B., Chan, A., Zhou, S., Gentzbittel, L., Childs, K.L., Yandell, M., Gundlach, H. and Mayer, K.F. (2014). An improved genome release (version Mt4. 0) for the model legume *Medicago truncatula*. *BMC Genomics*, 15(1), p.312.
- Tanruean, K., Chandet, N. and Rakariyatham, N. (2013). Bioconversion of Ferulic Acid into High Value Metabolites by White Rot Fungi Isolated from Fruiting-Body of the Polypore Mushroom. *Journal of Medical and Bioengineering Vol*, 2(3).
- Tengerdy, R.P. and Szakacs, G. (2003). Bioconversion of lignocellulose in solid substrate fermentation. *Biochemical Engineering Journal*, *13*(2), pp.169-179.
- Thibault, J., Micard, V., Renard, C., Asther, M., Delattre, M., Lesage-Meessen, L., Faulds, C., Kroon, P., Williamson, G., Duarte, J., Duarte, J. C., Ceccaldi, B. C., Tuohy, M., Couteau, D., Van Hulle, S. and Heldt-Hansen, H. P. (1998). Fungal Bioconversion of Agricultural By-Products to Vanillin. *Lebensmittel-Wissenschaft und-Technologie-Food Science Technology*, 31, 530-536.
- Thygesen, A., Thomsen, A.B., Schmidt, A.S., Jørgensen, H., Ahring, B.K. and Olsson, L. (2003). Production of cellulose and hemicellulose-degrading enzymes by filamentous fungi cultivated on wet-oxidised wheat straw. *Enzyme and microbial technology*, 32(5), pp.606-615.

- Tilay, A., Bule, M. and Annapure, U. (2010). Production of Biovanillin by One-Step Biotransformation Using Fungus *Pycnoporous cinnabarinus*. *Journal of Agricultural and Food Chemistry*, 58, 4401-4405.
- Tiwari, K.L., Jadhav, S.K. and Kumar, A. (2011). Morphological and molecular study of different *Penicillium* species. *Middle-East J Sci Res*, 7(1), pp.203-10.
- Topakas, E., Kalogeris, E., Kekos, D., Macris, B.J. and Christakopoulos, P. (2003). Bioconversion of ferulic acid into vanillic acid by the thermophilic fungus *Sporotrichum thermophile*. *LWT-Food Science and Technology*, *36*(6), pp.561-565.
- Tomlinson 2nd, G.H. and Hibbert, H. (1936). Studies on Lignin and Related Compounds. XXIV. The Formation of Vanillin from Waste Sulfite Liquor1. *Journal of the American Chemical Society*, 58(2), 345-348.
- Tomlinson, G.H. and Ibbert, H. (1936). Studies on lignin and related compounds. XXIV. The formation of vanillin from waste sulfite liquor. *J. Am. Chem. Soc.* 58, 345-348.
- Torres, B.R., Aliakbarian, B., Torre, P., Perego, P., Domínguez, J.M., Zilli, M. and Converti, A. (2009). Vanillin bioproduction from alkaline hydrolyzate of corn cob by Escherichia coli JM109/pBB1. Enzyme and Microbial Technology, 44(3), pp.154-158.
- Torre, P., Aliakbarian, B., Rivas, B., Domínguez, J.M. and Converti, A. (2008). Release of ferulic acid from corn cobs by alkaline hydrolysis. *Biochemical Engineering Journal*, 40(3), pp.500-506.
- Torre, P., De Faveri, D., Perego, P., Converti, A., Barghini, P., Ruzzi, M., Faria, F.P. (2004). Selection of co-substrate and aeration conditions for vanillin production by *E. coli* JM109/pBB1. *Food Technology Biotechnology*. 42, 193-196.
- Traynard, P., Ayroud, A.M. and Eymery, A. (1953). Existence dune liaison lignine-hydrates de carbone dans la bois. *Assoc Tech Ind Papetiere Bull*, 2, pp.45-52.
- Umikalsom, M.S., Ariff, A.B., Zulkifli, H.S., Tong, C.C., Hassan, M.A. and Karim, M.I.A. (1997). The treatment of oil palm empty fruit bunch fibre for subsequent use as substrate for cellulase production by *Chaetomium globosum Kunze*. *Bioresource Technology*, 62(1-2), pp.1-9.
- Vallejos, M.E., Felissia, F.E., Cruvelo, A.A., Zambon, M.D., Ramos, L. and Area, M.C. (2011). Chemical and physico-chemical characterization of lignins obtained from ethanol-water fractionation of bagasse. *BioResources*, 6(2), pp.1158-1171.
- Vandamme, E. J. (2003). Bioflavours and Fragrances via Fungi and Their Enzymes. *Fungal Diversity*, 13, 153-166.
- Vaithanomsat, P. and Apiwatanapiwat, W. (2009). Feasibility study on vanillin production from *Jatropha curcas* stem using steam explosion as a pretreatment. *Inter J Chem Biolo Engr*, 2(4), pp.211-4.

Vaughn, N. (2007). Design-Expert Software. Minneapolis, MN, Stat-Ease, Inc.

- Venkitasubramanian, P., Daniels, L., Das, S., Lamm, A.S. and Rosazza, J.P. (2008). Aldehyde oxidoreductase as a biocatalyst: reductions of vanillic acid. *Enzyme and microbial technology*, 42(2), pp.130-137.
- Vidal, J.P. (2007). Vanillin: in Kirk-Othmer Encyclopedia of Chemical Technology, John Wiley & Sons, Incorporated, 30-69.
- Vidal, J.P. (2006) Vanillin. Kirk-Othmer Encyclopedia of Chemical Technology. John Wiley & Sons, Incorporated, 1-10.
- Vaithanomsat, P. and Apiwatanapiwat, W. (2009). Feasibility study on vanillin production from Jatropha curcas stem using steam explosion as a pretreatment. International Journal of Chemistry. Biomolecule. Engineering, 2(4), 211-214.
- Wahid, Z. and Nadir, N. (2013). Improvement of one factor at a time through design of experiments. *World Appl. Sci. J*, 21, pp.56-61.
- Walton, N. J., Mayer, M. J. and Narbad, A. (2003). Vanillin. Phytochemistry, 63, 505-515.
- Walton, N. J., Narbad, A., Faulds, C. and Williamson, G. (2000). Novel approaches to the biosynthesis of vanillin. *Current Opinion in Biotechnology*, 11, 490-496.
- Wang, B., Yu, Y. and Wang, L. (2014). *Penicillium fusisporum* and *P. zhuangii*, two new monoverticillate species with apical-swelling stipes of section *Aspergilloides* isolated from plant leaves in China. *PloS One*, 9(7), p.e101454.
- Wang, L., Yang, B., Wang, R. and Du, X. (2008). Extraction of pepsin-soluble collagen from grass carp (Ctenopharyngodon idella) skin using an artificial neural network. *Food Chemistry*, 111(3), pp.683-686.
- Washisu, Y., Tetsushi, A., Hashimoto, N. and Kanisawa, T. (1993). Manufacture of vanillin and related compounds with *Pseudomonas*. *Japanese patent*, 5.
- Weising, K., Nybom, H., Pfenninger, M., Wolff, K. and Meyer, W. (1994). DNA fingerprinting in plants and fungi. CRC press.
- Wymelenberg, A. V., Minges, P., Sabat, G., Martinez, D., Aerts, A., Salamov, A., Grigoriev, I., Shapiro, H., Putnam, N., Belinky, P., Dosoretz, C., Gaskell, J., Kersten, P., Cullen, D. (2006). Computational analysis of the *Phanerochaete chrysosporium* v2.0 genome database and mass spectrometry identification of peptides in ligninolytic cultures reveal complex mixtures of secreted proteins. *Fungal Genetics and Biology*. 43(5), 343-356.
- Wymelenberg, A.V., Gaskell, J., Mozuch, M., Kersten, P., Sabat, G., Martinez, D. and Cullen, D. (2009). Transcriptome and secretome analyses of *Phanerochaete chrysosporium* reveal complex patterns of gene expression. *Applied and Environmental Microbiology*, 75(12), pp.4058-4068.
- Wymelenberg A.V., Sabat, G., Martinez, D., Rajangam, A.S., Teeri, T.T., Gaskell, J., Kersten, P.J., Cullen, D. (2005) The *Phanerochaete chrysosporium* secretome: database predictions and initial mass spectrometry peptide identifications in cellulose-grown medium. *Journal of Biotechnology*, 118(1), 17-34.

- ^aWymelenberg, A.V., Sabat, G., Mozuch, M., Kersten, P.J., Cullen, D. and Blanchette, R.A. (2006). Structure, organization, and transcriptional regulation of a family of copper radical oxidase genes in the lignin-degrading basidiomycete *Phanerochaete chrysosporium. Applied and Environmental Microbiology*, *72*(7), pp.4871-4877.
- ^bWymelenberg, A.V., Minges, P., Sabat, G., Martinez, D., Aerts, A., Salamov, A., Grigoriev, I., Shapiro, H., Putnam, N., Belinky, P. and Dosoretz, C. (2006). Computational analysis of the *Phanerochaete chrysosporium* v2. 0 genome database and mass spectrometry identification of peptides in ligninolytic cultures reveal complex mixtures of secreted proteins. *Fungal Genetics and Biology*, 43(5), pp.343-356.
- Xu, S. and Chen, L. (2008). A novel approach for determining the optimal number of hidden layer neurons for FNN's and its application in data mining.
- Xu, F., Yu, J., Tesso, T., Dowell, F. and Wang, D. (2013). Qualitative and quantitative analysis of lignocellulosic biomass using infrared techniques: a mini-review. *Applied Energy*, 104, pp.801-809.
- Yadav, J.S. (1988). SSF of wheat straw with alcaliphilic Coprinus. *Biotechnology and bioengineering*, *31*(5), pp.414-417.
- Yan, L., Chen, P., Zhang, S., Li, S., Yan, X., Wang, N., Liang, N. and Li, H. (2016). Biotransformation of ferulic acid to vanillin in the packed bed-stirred fermentors. *Scientific reports*, 6, p.34644.
- Yang, W., Tang, H., Ni, J., Wu, Q., Hua, D., Tao, F. and Xu, P. (2013). Characterization of two Streptomyces enzymes that convert ferulic acid to vanillin. *PloS one*, 8(6), p.e67339
- Yun Ping, N., Azis, A., Chin-Ping, T. and Yew-Ai, T. (2008). Determination of oil palm fruit phenolic compounds and their antioxidant activities using spectrophotometric methods. *International Journal of Food Science and Technology*, 43, 1832-1837.
- Yusof, N.N.M., Tanioka, E. and Kobayashi, T. (2014). Molecularly imprinted polymer particles having coordinated hydrogen bonding in covalent-imprinting for efficient recognition towards vanillin. *Separation and Purification Technology*, 122, pp.341-349.
- Zakaria, M.H. (2015). Review of Policies and Issues in the Malaysian Herbal Industry. *Economic* and Technology Management Centre. MARDI.
- Zamzuri, N.A. and Abd-Aziz, S. (2013). Biovanillin from agro wastes as an alternative food flavour. *Journal of the Science of Food and Agriculture*, 93(3), 429-438.
- Zenk, M.H., Ulbrich, B., Busse, J. and Stöckigt, J. (1980). Procedure for the enzymatic synthesis and isolation of cinnamoyl-CoA thiolesters using a bacterial system. *Analytical Biochemistry*, 101(1), pp.182-187.
- Zhang, Y., Mo, L., Chen, F., Lu, M., Dong, W., Wang, Q., Xu, F. and Gu, F. (2014). Optimized production of vanillin from green vanilla pods by enzyme-assisted extraction combined with pre-freezing and thawing. *Molecules*,19(2), pp.2181-2198.

- Zhang, Y., Yu, Z., Lu, Y., Wang, Y., She, D., Song, M. and Wu, Y. (2007). Effect of the absence of lipoxygenase isoenzymes on the storage characteristics of rice grains. *Journal of Stored Products Research*, 43(1), pp.87-91.
- Zheng, L., Zheng, P., Sun, Z., Bai, Y., Wang, J. and Guo, X. (2007). Production of vanillin from waste residue of rice bran oil by Aspergillus niger and Pycnoporus cinnabarinus. Bioresource Technology, 98(5), pp.1115-1119.
- Zheng, Y., Pan, Z. and Zhang, R. (2009). Overview of biomass pretreatment for cellulosic ethanol production. *International Journal of Agricultural and Biological Engineering*, 2(3), pp.51-68.