

MULTIWALL CARBON NANOTUBE-G-(AGAROSE-G-POLYMETHYL
METHACRYLATE) AND (AGAROSE-G-POLYMETHYL METHACRYLATE)
SORBENTS FOR EXTRACTION OF PESTICIDES AND HEAVY METAL IONS
FROM AQUEOUS MATRICES

NEDA POURMAND

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (Chemistry)

Faculty of Science
Universiti Teknologi Malaysia

MAY 2016

In the name of Allah, Most Gracious, Most Merciful

*To my beloved husband Mohammad Reza Isvandzibaei, who are provided me hope
with support, help and encouragement that greatly contributed to the successful
completion of my studies,*

*To my mother, who are praying for me and who has brought, great motivation and
bright inspiration into my life*

ACKNOWLEDGEMENT

Alhamdulillah, all praise is due to Allah, the lord of the worlds, keep us on the right path, the most beneficent and the most merciful.

First and foremost, I wish to express special thanks, appreciation and deep gratitude to my main supervisor, Prof. Dr. Mohd Marsin Sanagi, who has provide continuous guidance, advice, support and helping me to complete this research. His remarkable unique ways and professionalism of handling my weaknesses has turned my simplistic mind to see think in more rational and critical view.

Special thanks also to Dr. Ahmedy Abu Naim my past co-supervisor, for his continuous guidance, committed support and invaluable advice throughout my study and Prof. Dr. Wan Aini Wan Ibrahim for giving me suggestions and critical comments during SepSTec group meeting.

I appreciate and acknowledge the significant role university for the entire study period. I wish to thank the Universiti Teknologi Malaysia (UTM) for all supports and facilities especially for Ministry of Science, Technology and Innovation Malaysia (MOSTI) during studies of my Ph.D programme.

Finally, I would like to thanks my lovely spouse Mohammad Reza and my parents and my friends for their support and encouragement throughout in the completion of this research. Without their endless sacrifices, constant love and steadfast support, I can not be where I am today.

ABSTRACT

Rapid developments in agriculture and industrial sectors have increased the levels of toxic heavy metals and pesticide residues in the environment. Many sample pretreatment methods utilized in the determination of heavy metals and pesticide residues are often time consuming, labor intensive and require substantial amounts of organic solvents. Thus, new approaches in miniaturized sample preparation are imperative. In this study, new modified agarose sorbents were prepared, characterized and applied to the extraction and pre-concentration of pesticides and heavy metal ions from aqueous matrices. Poly(methyl methacrylate)-grafted-agarose (Agarose-g-PMMA) was successfully synthesized using microwave radiation and ceric ammonium nitrate (CAN) as the radical initiator. The optimum condition for graft copolymerization was found to be at 9 g monomer, 0.4 g CAN, 60 s microwave irradiation time under 700W microwave radiation for 1 g of agarose. The graft copolymer was characterized by Fourier transform infrared (FTIR) spectroscopy, CHN analysis, field emission scanning electron microscopy (FESEM), differential scanning calorimetry (DSC) analysis, thermal gravimetric analysis (TGA) and gel permeation chromatography (GPC). A micro-solid phase extraction (μ -SPE) utilizing Agarose-g-PMMA as sorbent combined with gas chromatography-microelectron capture detection was developed for the determination of selected pesticides, namely diazenon, chlorpyrifos, hexaconazole and azaconazole in water samples. Under optimized conditions, low limits of detection (LODs) ($0.004 - 0.024 \text{ ng mL}^{-1}$) were obtained with good recoveries (82.23 - 103.58%). Agarose-g-PMMA was also employed as sorbent in dispersive micro-solid phase extraction (D- μ -SPE) combined with inductively coupled plasma-mass spectrometry for the analysis of heavy metals namely cadmium, nickel, copper and zinc in vegetables and natural water samples. Under the optimum conditions, the developed method showed excellent LODs ($0.6 - 1.8 \text{ ng L}^{-1}$) and good relative recoveries (92.0 - 103.0%) for the analytes. The Agarose-g-PMMA was covalently-modified with multi-walled carbon nanotubes (MWCNTs) and the composite formed (MWCNTs-g-Agarose-g-PMMA) was characterized by FTIR, TGA and transmission electron microscopy (TEM). A SPE method incorporating MWCNTs-g-Agarose-g-PMMA as sorbent was developed and combined with flame atomic absorption spectrometry for the determination of lead ions in natural water samples. The method provided fast analysis and showed good sensitivity and excellent precision and suitable for extraction and pre-concentration of pesticide residues and trace metal ions in water and vegetable samples.

ABSTRAK

Pembangunan pesat dalam sektor pertanian dan perindustrian telah meningkatkan tahap logam berat toksik dan sisa racun perosak dalam persekitaran. Banyak kaedah pra-rawatan sampel yang digunakan dalam penentuan logam berat dan sisa racun perosak biasanya memakan masa yang panjang, intensif buruh dan memerlukan sejumlah pelarut organik yang banyak. Justeru, pendekatan baru dalam penyediaan sampel bersaiz kecil adalah sangat penting. Dalam kajian ini, bahan pengerap agarosa terubahsuai baru telah disediakan, dicirikan dan digunakan bagi pengekstrakan dan pra-pemekatan racun perosak dan ion logam berat di dalam matriks akueus. Poli(metil metakrilat)-cangkuk-agarosa (Agarose-g-PMMA) telah berjaya disintesis menggunakan sinaran gelombang mikro dan serik ammonium nitrat (CAN) sebagai pemula radikal. Keadaan optimum bagi ko-pempolimeran cangkuk telah didapati sebagai 9 g monomer, 0.4 g CAN, 60 s masa penyinaran gelombang mikro di bawah sinaran gelombang mikro 700 W bagi 1 g agarosa. Polimer cangkuk telah dicirikan dengan spektroskopi inframerah transformasi Fourier (FTIR), analisis CHN, mikroskopi elektron imbasan pancaran medan (FESEM), analisis kalorimetri pengimbasan pembezaan (DSC), analisis gravimetri terma (TGA) dan kromatografi penelapan gel (GPC). Pengekstrakan fasa pepejal mikro menggunakan Agarose-g-PMMA sebagai bahan pengerap digabungkan dengan kromatografi gas-pengesan penangkapan mikroelektron telah dibangunkan bagi penentuan racun perosak terpilih, iaitu diazenon, klorpirifos, heksakonazola dan azakonazola di dalam sampel air. Di bawah keadaan yang optimum, had pengesanan (LOD) yang rendah ($0.004 - 0.024 \text{ ng mL}^{-1}$) telah diperolehi dengan perolehan balik yang baik (82.23 - 103.58%). Agarose-g-PMMA juga telah digunakan sebagai bahan pengerap dalam pengekstrakan fasa pepejal mikro (D- μ -SPE) digabungkan dengan spektrometri jisim-plasma ganding aruhan bagi analisis logam berat iaitu kadmium, nikel, kuprum dan zink di dalam sampel sayur-sayuran dan air semula jadi. Di bawah keadaan optimum, kaedah yang dibangunkan itu menunjukkan LOD yang cemerlang ($0.6 - 1.8 \text{ ng L}^{-1}$) dan perolehan balik relatif yang baik (92.0 - 103.0%) bagi analit tersebut. Agarose-g-PMMA telah diubahsuai secara kovalen dengan nanotub karbon dinding berbilang (MWCNT) dan komposit yang terbentuk (MWCNTs-g-Agarose-g-PMMA) telah dicirikan dengan FTIR, TGA dan mikroskopi elektron penghantaran (TEM). Kaedah SPE yang mengandungi MWCNTs-g-Agarose-g-PMMA sebagai bahan pengerap telah dibangunkan dan digabungkan dengan spektrometri penyerapan atom nyala bagi penentuan ion plumbum di dalam sampel air semula jadi. Kaedah tersebut memberikan analisis yang cepat, menunjukkan kepekaan yang baik dan kepersisan yang cemerlang, serta sesuai bagi pengekstrakan dan pra-pemekatan sisa racun perosak dan ion logam surih di dalam sampel air dan sayur-sayuran.

TABLES OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xiv
	LIST OF FIGURES	xvi
	LIST OF ABBREVIATIONS	xxi
	LIST OF SYMBOLS	xxiii
	LIST OF APPENDICES	xxiv
1	INTRODUCTION	1
	1.1 Research Background	1
	1.2 Problem Statement	4
	1.3 Objectives of the Study	5
	1.4 Scope of the Study	6
	1.5 Significance of the Research	7
2	LITERATURE REVIEW	8
	2.1 Modifications of Polysaccharide	8
	2.2 Graft Copolymerization onto Polysaccharides	9
	2.2.1 Conventional Grafting Procedures	10

2.2.2	Grafting Procedures with Microwaves	11
2.2.3	Grafting with Other Methods	13
2.3	Mechanism of Grafting on Polysaccharides under Microwaves	13
2.3.1	Microwaves Grafting in Aqueous Solution	14
2.3.1.1	Microwave Assisted Grafting	14
2.3.1.2	Microwave Initiated Grafting	16
2.3.2	Microwaves Grafting on Solid Support	18
2.4	Properties and Limitation of Carbon nanotubes	19
2.5	CNTs Functionalization with Polymers	20
2.6	Preparation Methods for MWCNTs Nanocomposites	22
2.6.1	Melt Mixing	23
2.6.2	Solution Blending	23
2.6.3	In-situ Polymerization	24
2.6.4	Polymer Grafting	24
2.7	Traditional Extraction Techniques for Chemical Analysis	26
2.7.1	Liquid-liquid Extraction	26
2.7.2	Solid Phase Extraction	27
2.8	Recent Microextraction Techniques for Chemical Analysis	28
2.8.1	Solid Phase Microextraction	28
2.8.2	Stir Bar Sorptive Extraction (SBSE)	29
2.8.3	Micro Solid Phase Extraction	29
2.8.4	Dispersive Solid Phase Extraction	31
2.9	Properties of Agarose	32
2.10	Application of Poly(methyl methacrylate) in Graft Copolymerization of Polysaccharides	33

3	PREPARATION, OPTIMIZATION AND CHARACTERIZATION OF AGAROSE GRAFTED POLY (METHYL METHACRYLATE) USING MICROWAVE- ASSISTED METHOD	35
3.1	Introduction	35
3.2	Experimental	36
3.2.1	Chemicals and Reagents	36

3.2.2	Equipments/Instruments	36
3.2.3	Optimization Procedure of Agarose-g-PMMA	37
3.2.3.1	General Procedure for Graft Copolymerization	37
3.2.3.2	Effect of Amount of Monomer	38
3.2.3.3	Effect of Amount of Initiator	38
3.2.3.4	Effect of Microwave Radiation Duration	38
3.2.4	Purification of the Graft Copolymer by Solvent Extraction Method	39
3.2.5	Hydrolysis of Graft Copolymers with Hydrochloric Acid	39
3.3	Characterization	39
3.3.1	Optimized Conditions of Graft Copolymer	39
3.3.2	Elemental Analysis	40
3.3.3	FTIR Spectroscopy	41
3.3.4	Field Emission Scanning Electron Microscopy	41
3.3.5	Differential Scanning Calorimetry Analysis	41
3.3.6	Thermogravimetric Analysis	42
3.3.7	Gel Permeation Chromatography	42
3.4	Results and discussion	42
3.4.1	Agarose-g-PMMA Synthesis by Microwave Assisted Method	42
3.4.1.1	Effect of Amount of Monomer	44
3.4.1.2	Effect of Amount of Initiator	45
3.4.1.3	Effect of Microwave Irradiation Duration	46
3.4.2	Elemental Analysis	47
3.4.3	FTIR Spectroscopy of Unmodified Agarose and Prepared Agarose-g-PMMA Copolymer	48
3.4.4	Field Emission Scanning Electron Microscopy	49
3.4.5	Differential Scanning Calorimetry	50
3.4.6	Thermogravimetric Analysis	51
3.4.7	Gel Permeation Chromatography	53
3.5	Conclusion	55

4	AGAROSE GRAFTED POLY(METHYL METHACRYLATE) MICRO-SOLID PHASE EXTRACTION COMBINED WITH GAS-CHROMATOGRAPHY FOR THE DETERMINATION OF SELECTED PESTICIDES	56
4.1	Introduction	56
4.2	Experimental	57
4.2.1	Reagents and Materials	57
4.2.2	Synthesis and characterization of Agarose-g-PMMA	58
4.2.3	Chromatographic Conditions	59
4.2.4	μ -SPE Device Preparation	59
4.2.5	Optimization Procedure of Agarose-g-PMMA- μ -SPE	60
4.2.5.1	Effect of Mass of Sorbent	61
4.2.5.2	Effect of Desorption Solvent	62
4.2.5.3	Effect of Desorption Time	62
4.2.5.4	Effect of Extraction Time	62
4.2.5.5	Effect of Sample Volume	63
4.2.6	Solid Phase Extraction	63
4.2.7	Sample Preparation	64
4.2.8	Method Validation	64
4.3	Results and Discussions	65
4.3.1	Characterization of Agarose-g-PMMA	65
4.3.2	Peak Identification and Chromatographic Calibration	65
4.3.3	Optimization of Agarose-g-PMMA- μ -SPE	68
4.3.3.1	Mass of Sorbent	68
4.3.3.2	Effect of Desorption Solvent	69
4.3.3.3	Desorption Time	70
4.3.3.4	Extraction Time	71
4.3.3.5	Sample Volume	72
4.3.4	Optimization of C ₁₈ -SPE	73
4.3.5	Method Validation	74
4.3.6	Real Sample Analysis	77
4.6	Conclusion	77

5	DISPERSIVE MICRO-SOLID PHASE EXTRACTION COMBINED WITH INDUCTIVELY COUPLED PLASMA-MASS SPECTROMETRY FOR THE DETERMINATION OF HEAVY METALS IN VEGETABLES AND NATURAL WATER SAMPLES	80
5.1	Introduction	80
5.2	Experimental	81
5.2.1	Chemicals and Reagents	81
5.2.2	Instrumentation	82
5.2.3	Synthesis and Characterization of Agarose -g-PMMA	82
5.2.4	Optimization Procedure of Agarose-g-PMMA -D- μ -SPE	83
5.2.4.1	Effect of Mass of Sorbent	84
5.2.4.2	Effect of pH	84
5.2.4.3	Effect of Extraction Time	84
5.2.4.4	Effect of Type of Eluent	85
5.2.4.5	Effect of Concentrations of Eluent	85
5.2.4.6	Effect of Desorption Time	85
5.2.5	Validation of Agarose-g-PMMA-D- μ -SPE	86
5.2.6	Sample Preparation	86
5.3	Results and Discussion	87
5.3.1	Optimization of Agarose-g-PMMA-D- μ -SPE	87
5.3.2	Effect of Co-existing Substances	91
5.3.3	Effect Adsorption Capacity	92
5.3.4	Method Validation	92
5.3.5	Analysis of Real Sample	94
5.3.6	Comparison of Agarose-g-PMMA-D- μ -SPE with Other Reported Methods	94
5.4	Conclusion	97

6	PREPARATION , CHARACTERIZATION AND APPLICATION OF MWCNTs-G-AGAROSE-G-PMMA FOR SOLID-PHASE EXTRACTION OF Pb(II) IN ENVIRONMENTAL SAMPLES	99
6.1	Introduction	99
6.2	Experimental	100
6.2.1	Chemicals and Reagents	100
6.2.2	Preparation of MWCNTs-g-Agarose-g-PMMA	101
6.2.2.1	Synthesis and Characterization of Agarose-g-PMMA	101
6.2.2.2	Preparation of MWCNTs Functionalized with Carboxylic Acid Groups	101
6.2.2.3	Preparation of MWCNTs Functionalized with Formyl Chloride	102
6.2.2.4	Synthesis of MWCNTs-g-Agarose-g-PMMA	102
6.2.3	Characterization of MWCNTs-g-Agarose-g-PMMA	102
6.2.4	SPE Procedures	103
6.2.4.1	Effect of pH	103
6.2.4.2	Effect of Type of Eluent	103
6.2.4.3	Effect of Concentrations of Eluent	104
6.2.4.4	Effect of Volume of Eluent	104
6.2.4.5	Effect of Sample Volume	104
6.2.5	Method Validation	105
6.2.6	Sample Analysis	105
6.3	Results and Discussion	106
6.3.1	Optimization of MWCNTs-g-Agarose-g-PMMA Process Parameter	106
6.3.2	Characterization of MWCNTs-g-Agarose-g-PMMA Sorbent	107
6.3.2.1	Fourier Transform Infrared Analysis	107
6.3.2.2	Transmission Electron Microscopy (TEM)	109
6.3.2.3	Thermogravimetric Analysis (TGA)	110
6.3.3	Optimization of Extraction Parameters	112

6.3.4	Effect of Interfering Ions	117
6.3.5	Adsorption Capacity	118
6.3.6	Method Validation	119
6.3.7	Analysis of Real Samples	120
6.3.8	Comparison of the Nanocomposite with Other Materials for Solid Phase Extraction of Pb(II)	121
6.4	Conclusion	122
7	CONCLUSION AND FUTURE WORKS	124
7.1	Conclusion	124
7.2	Suggestion for Future works	126
	REFERENCES	128
	Appendices A-B	150-151

LIST OF TABLES

TABLE NO.	TITLE	PAGE
3.1	Effect of amount of monomer on the GE (%), G (%) and C (%) of samples polymerized at 60 s and CAN (0.2 g)	44
3.2	Effect of amount of initiator on the GE (%), G (%) and C (%) of samples polymerized	45
3.3	Effect of microwave irradiation duration on the GE (%), G (%) and C (%) of samples polymerized	46
3.4	Elemental analysis results	47
4.1	Pesticides studied and their structures, molecular weights, log octanol-water partition coefficient ($\log K_{ow}$) values and water solubilities.	58
4.2	Comparison of linear range, correlation coefficients (R), limit of detection (LOD) limit of quantification (LOQ) of the agarose-g-PMMA- μ -SPE and C ₁₈ -SPE of selected pesticides spiked in water ($n = 3$).	75
4.3	Precision studies (RSD) of the four pesticides using agarose-g-PMMA- μ -SPE.	76
4.4	Recovery (%) and method precisions (RSD %, $n = 3$) at two different concentrations for the extraction of the pesticides in tap water, lake water and river water samples.	79

5.1	Operating conditions for ICP-MS.	82
5.2	Effect of the concentration and time of eluent for desorption of target analytes from Agarose-g-PMMA. (Conditions: concentration of sample $4 \mu\text{g L}^{-1}$, solution volume; 50 mL, amount of sorbent: 80 mg and pH = 6; eluent, HNO_3).	91
5.3	Effects of the interferences ions on the recoveries.	92
5.4	Agarose-g-PMMA-D- μ -SPE validation data.	93
5.5	Analysis of Cd(II), Ni(II), Cu(II) and Zn(II) in environmental water samples. (Mean \pm SD, n = 5).	95
5.6	Analysis of Cd(II), Ni(II), Cu(II) and Zn(II) in vegetal samples. (Concentration \pm SD, n = 5).	96
5.7	Comparisons among the analytical performances of the Agarose-g-PMMA and other reported sorbents	98
6.1	Effects of the interference ions on the recoveries.	117
6.2	Precisions and relative recoveries of MWCNTs-g-Agarose-g-PMMA-SPE for lead in spiked tap water.	119
6.3	Analysis of Pb(II) in environmental water samples. (Mean \pm SD, n = 3).	121
6.4	Comparisons between the analytical performances of the MWCNTs-g-Agarose-g-PMMA nanocomposite with different nano absorbents used in solid phase extraction.	123

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Diagram of comparison between conventional and microwave methods for polysaccharides (Santagada <i>et al.</i> , 2004).	12
2.2	Schematic of the synthesis of graft copolymers using microwave-initiated and microwave-assisted method.	18
2.3	Modificaton of CNTs with amines or alcohols derivatization reaction.	22
2.4	Basic procedures in SPE (a) Conditioning of sorbent; (b) Sample loading; (c) Washing of unwanted compounds and (d) Elution of target analytes.	28
2.5	Schematic of micro solid phase extraction (μ -SPE).	30
2.6	Chemical structure of agarose.	32
3.1	FTIR spectra of (a) Agarose, and (b) Agarose-g-PMMA.	48
3.2	Infrared spectrum of linear poly(methyl methacrylate).	49
3.3	FESEM micrograph of (a) Agarose at 1,000 magnifications, and (b) Agarose at 5,000 magnifications.	49

3.4	FESEM micrograph of (a) Agarose-g-PMMA at 500 magnifications, and (b) Agarose-g-PMMA at 1,000 magnifications.	50
3.5	DSC thermogram of (a) Agarose and (b) Agarose-g-PMMA.	51
3.6	TGA and DTA thermograms of (a) Agarose and (b) Agarose-g-PMMA.	52
3.7	FTIR spectrum of poly(methyl methacrylate) from hydrolyzed agarose-g-PMMA copolymer.	54
3.8	Gpc curves of poly(methyl methacrylate) from hydrolyzed agarose-g-PMMA copolymer.	54
4.1	Schematic of a μ -SPE device preparation.	60
4.2	Schematics of (a) μ -SPE system (b) image of extraction device.	61
4.3	GC chromatograms of pesticides (a, b, c and d) for each pesticide and the mixture of pesticides (e), at $100 \mu\text{g L}^{-1}$ (each). Peaks: (1) diazinon, (2) chlorpyrifos, (3) hexaconazole and (4) azaconazole.	67
4.4	Effect of mass of agarose-g-PMMA on agarose-g-PMMA- μ -SPE of selected pesticides spiked distilled water. Agarose-g-PMMA- μ -SPE conditions: $1 \mu\text{g L}^{-1}$ of spiked solution; desorption solvent, $500 \mu\text{L}$ of methanol; extraction time, 30 min; desorption time, 20 min; sample volume, 10 mL.	69
4.5	Effect of desorption solvent on agarose-g-PMMA- μ -SPE of selected pesticides from spiked distilled water. Legends, agarose-g-PMMA- μ -SPE conditions are as in Figure 4.4 with 15 mg of agarose-g-PMMA as sorbent.	70

4.6	Effect of desorption time on agarose-g-PMMA- μ -SPE of selected pesticides from spiked distilled water. Legends and agarose-g-PMMA- μ -SPE conditions are as in Figure 4.4 with 15 mg of agarose-g-PMMA as sorbent.	71
4.7	Effect of extraction time on agarose-g-PMMA- μ -SPE of selected pesticides from spiked distilled water. Legends and agarose-g-PMMA- μ -SPE conditions are as in Figure 4.4 with 15 mg of agarose-g-PMMA as sorbent and 15 min desorption time.	72
4.8	Effect of sample volume on agarose-g-PMMA- μ -SPE of selected pesticides from spiked distilled water. Legends and agarose-g-PMMA- μ -SPE conditions are as in Figure 4.4 with 15 mg of agarose-g-PMMA as sorbent, 15 min desorption time and 40 min extraction time.	73
4.9	GC- μ ECD chromatograms of tap water extracts using the optimum of agarose-g-PMMA- μ -SPE procedure. Chromatogram identities: (a) Blank tap water after extraction; (b) tap water spiked with selected pesticides at $1 \mu\text{g L}^{-1}$.	78
5.1	Schematic of Agarose-g-PMMA-D- μ -SPE analytical procedure.	83
5.2	Two Effects of sorbent mass on the preconcentration of analytes (other conditions: sample volume, 50.0 mL; eluent, 0.5 mol L^{-1} nitric acid; pH 7; n = 5).	87
5.3	Effects of pH on the recovery of analytes (conditions: sample volume, 50.0 mL; eluent: 1 mL of 0.5 mol L^{-1} nitric acid; n = 5).	88
5.4	Effects of extraction time on the recovery of analytes (conditions: sample volume, 50.0 mL; pH 6, eluent: 1mL of 0.5 mol L^{-1} nitric acid; n = 5).	89

5.5	Effects of type of eluen on the recovery of analytes (conditions: sample volume, 50.0 mL; pH 6, extraction time: 4 min; eluent: 1mL of 0.5 mol L ⁻¹ nitric acid; n= 5).	90
5.6	Recovery percentage of heavy metals with number of cycles using MWCNTs-g-Agarose-g-PMMA-D-μ-SP.	93
6.1	Effect of different ratio composition (MWCNTs:Agarose-g-PMMA) of the synthesized graft composite MWCNTs-g-Agarose-g-PMMA on the extraction efficiency of Pb. Extraction parameters: 50 mL sample volume, 5 mL of HNO ₃ 1M as elution condition in pH 7.	106
6.2	FTIR spectra of (a) MWCNTs, (b) MWCNTs- COOH (c) MWCNTs-COCl, (d) Agarose, (e) Agarose-g-PMMA and (f) MWCNTs-g-Agarose-g-PMMA	108
6.3	TEM images of (a) MWCNTs-COOH, (b) Agarose-g-PMMA, (c) MWCNTs-g-Agarose-g-PMMA, (d) MWCNTs-Agarose -g-PMMA	109
6.4	TGA and DTAcures of (a) MWCNTs-COOH, (b) Agarose-g- PMMA, (c) Agarose, (d) MWCNTs-g-Agarose-g-PMMA.	111
6.5	Effects of pH on the preconcentration of 500 μg L ⁻¹ Pb on MWCNTs, Agarose, Agarose-g-PMMA and MWCNTs-g-Agarose- g-PMMA (other conditions: sample volume, 50.0 mL; flow rate,1 mL min ⁻¹ ; eluent, 1.0 mol L ⁻¹ HNO ₃ ; n = 3).	112
6.6	Effects of type of acid on the preconcentration of 500 μg L ⁻¹ Pb on MWCNTs-g-Agarose-g-PMMA (other conditions: sample volume, 50.0 mL; flow rate,1 mL min ⁻¹ ; eluent, 1.0 mol L ⁻¹ HNO ₃ ; n = 3).	114

- 6.7 Effects of concentrations of acid on the preconcentration of 500 $\mu\text{g L}^{-1}$ Pb on MWCNTs-g-Agarose-g-PMMA (other conditions: sample volume, 50.0 mL; flow rate, 1 mL min^{-1} ; eluent, 5 mL, HNO_3 ; $n = 3$). 115
- 6.8 Effects of volume of acid on the preconcentration of 500 $\mu\text{g L}^{-1}$ Pb on MWCNTs-g-Agarose-g-PMMA (other conditions: sample volume, 50.0 mL; flow rate, 1 mL min^{-1} ; eluent, 0.5 mol L^{-1} , HNO_3 ; $n = 3$). 115
- 6.9 Effects of volume of sample on the preconcentration of 500 $\mu\text{g L}^{-1}$ Pb on MWCNTs-g-Agarose-g-PMMA (other conditions setting was same as Figure 6.8). 116
- 6.10 Effect of initial concentration (C_0) of Pb(II) on the adsorption quantity(Q) of MWCNTs-g-Agarose-g-PMMA. pH 5.0; sample volume, 50.0 mL; flow rate, 1 mL min^{-1} . 118
- 6.11 Recovery percentage of lead with number of cycles using MWCNTs-g-Agarose-g-PMMA-SPE. 120

LIST OF ABBREVIATIONS

AA	-	Acrylic acid
APS	-	Ammonium peroxydisulfate
ATRP	-	Atom Transfer Radical Polymerization
CAN	-	Ceric ammonium nitrate
CHN	-	Carbon hydrogen and nitrogen analysis
CNTs	-	Carbon nanotubes
DMF	-	Dimethyl formamide
DSC	-	Differential scanning calorimetry
D- μ -SPE	-	Dispersive micro-solid phase extraction
EF	-	Enrichment factor
FESEM	-	Field emission scanning electron microscope
FTIR	-	Fourier Transform Infrared Spectroscopy
G (%)	-	Percentage grafting
GC	-	Gas chromatography
GPC	-	Gel permeation chromatography
HPLC	-	High performance liquid chromatography
KPS	-	Potassium persulfate
LLE	-	Liquid-liquid extraction
LOD	-	Limit of detection
LOQ	-	Limit of quantification
MMA	-	Methyl methacrylate
M_n	-	Number average molecular weight
M_w	-	Mass average molecular weight
MWCNTs	-	Multiwall carbon nanotubes
PANI	-	Polyaniline
PBMA	-	Poly(butyl methacrylate)

PDI	-	Polydispersity index
PDMS	-	Polydimethylsiloxane
PEMA	-	Poly(ethyl methacrylate)
PHMA	-	Poly(hexyl methacrylate)
PMMA	-	Poly (methyl methacrylate)
PNVI	-	Poly(N-vinyl imidazole)
PS	-	Polystyrene
PVP	-	Polyvinylpyrrolidone
r.p.m	-	Rotation per minute
R ²	-	Coefficients of determination
RR	-	Relative recovery
RSD	-	Relative standard deviation
SBSE	-	Stir bar sorptive extraction
SPE	-	Solid phase extraction
SPME	-	Solid phase microextraction
T _g	-	Glass transition temperature
UV	-	Ultraviolet
μ-SPE	-	Micro solid phase extraction

LIST OF SYMBOLS

%	-	Percent
°C	-	Degree celcius
cm	-	Centimeter
g	-	Gram
g/mol	-	Gram per mol
kg	-	Kilogram
$K_{o/w}$	-	Octanol-water distribution coefficient
L	-	Liter
M	-	Molarity
mg	-	Milligram
min	-	Minutes
mL	-	Milliliter
mL/min	-	Milliliter per minute
mm	-	Millimeter
ng	-	Nanogram
nm	-	Nanometer
ppb	-	Part per billion
ppb	-	Part per billion
s	-	Second
v/v	-	Volume per volume
µg	-	Microgram
µg/L	-	Microgram per liter
µg/mL	-	Microgram per mililiter
µL	-	Microliter

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	List of Publications Related to this Study	150
B	List of Presentations Related to this Study	151

CHAPTER 1

INTRODUCTION

1.1 Research Background

Rapid development of industrial and agriculture are resulting in incrementing levels of heavy metals and pesticide residues in biological and environmental samples, and this pollutants have toxic effects on human health and growth of plant. These toxic heavy metals and pesticide residues can gradually pile up in the human being through the food chain. When the amounts are very high, the metals can be hazardous to human health. Biological and environmental sample matrices are complex and often contain interfering elements and organic compounds that can mask or interfere with the compounds of interest, thus, direct analysis may not be possible. The sample preparation step before chromatographic analysis is considerable intricate, and expensive.

Many routine pretreatment methods utilized in the determination of heavy metal are often time period consuming, labor intensive and substantial amount of organic solvent wasting. Sample preparations are very commonly carried out using conventional extraction methods such as liquid-liquid extraction (LLE) and solid phase extraction (SPE) to isolate, extract and concentrate target analytes. However, LLE are time consuming, labour-intensive and multi-stage operations. SPE has significant improvement over LLE by minimizing the consumption of chemicals and disposal cost of organic wastes.

Researchers have been devoted to the development of miniaturized sample preparation which resulted in significant solvent and sample savings. Apparently, the introduction and development of new materials/sorbents for microextraction have promising alternative to overcome some problems that occurred in analytical chemistry field. This microextraction is called sorbent/material-based microextraction. These relatively new techniques include such as solid phase microextraction (SPME) (Cavaliere *et al.*, 2012), stir bar sorptive extraction (SBSE) (Maggi *et al.*, 2008), dispersive solid phase extraction (DSPE) (Guan *et al.*, 2013) and micro solid phase extraction (μ -SPE). Microextraction is one of the solventless microextraction techniques that dramatically simplifies the sample preparation procedures by combining the integration of sampling, isolation and enrichment in one step (Pawliszyn, 1997).

Carbon nanotubes (CNTs) have attracted special interest in multidisciplinary study and have shown potential applications in many research areas since their discovery by Iijima in 1991 (Iijima, 1991). Their nanometer size, high aspect ratios, and more importantly their excellent mechanical, electrical and conducting properties make them highly suitable in the preconcentration of metal ions and organic pollutants in the treatment of polluted biological and environmental samples.

CNTs are generally insoluble and astringently aggregate due to Van der Waals magnetization among the nanotubes, engendering a quandary when homogenous dispersion is desirable. To surmount this barrier, sundry physical and chemical approaches have been explored including direct suspension of CNTs in polymer solution via sonication, in situ polymerization in the presence of CNTs, and the chemical modification of CNTs to enhance solubilization. Overall, the grafting of CNTs to polymer through covalent functionalization appears to be one of the most attractive and efficacious ways to achieve homogenous dispersion of CNTs in polymer matrices in order to make high quality nanocomposites. This technique has been used to make nanocomposites of CNT-polymers, mostly utilizing synthetic based polymers. However, information on the utilization of natural biopolymer grafted to carbon nanotube is currently scarce and thus of interest in this study.

Generally, chemical modification is the inter-conversion or replacement of functional groups or atoms (including hydrogen) already present on the main chain (backbone) (Ebdon, 1991). This can also be described as the alteration of chemical structure of group of atoms or molecules by introducing another group of atoms or molecules via chemical means which can provide hybrid materials with improved chemical and physical properties. Chemical modification of polymer is a process which polymers/monomers are transformed into distinct copolymers via chemical means, having improved chemical and physical properties. Chemical modification of high polymers is an important aspect of polymer science which continues to receive considerable attention. One of the important aspects in the chemical modification is in the reaction of polymers leading to block and graft copolymers. An example of the formation of graft and block copolymers from synthetic polymers has been reported (Stevens, 1999). Recently, more research works have been introduced to prepare graft copolymers from natural polymers. For example, cellulose has been grafted with several monomers to improve its properties. These include homogenous graft copolymerization of styrene onto cellulose in sulphur dioxide-diethylamine-dimethyl sulphoxide solvent (Tsuzuki *et al.*, 1980), acrylonitrile and methyl methacrylate monomers grafted onto cellulose in dimethyl sulphoxide-paraformaldehyde solvent (Nishioka & Kosai, 1981), methyl acrylate onto cellulose (Nishioka *et al.*, 1983), hydroxyethyl methacrylate graft copolymerization onto cellulose (Nishioka *et al.*, 1986), grafting of styrene monomer onto starch (Kaewtatip and Tanrattanakul, 2008).

Graft copolymerization is one of the interesting and promising techniques used to prepare a well-defined copolymer with novel architecture, composition and functionalities. As a result, graft copolymers have a variety of potential applications in many fields including textile industries, medical, pharmaceuticals and chromatography column (separation) technology. A graft copolymer is a macromolecular chain with one or more sort of block connected to the main chemical chain as side chain(s). Thus, grafted polymer has the general structure of polymer backbone or trunk polymer with different points along its size. An external factor is utilized to create free radical sites on this preformed polymer (polysaccharide in case of grafted polysaccharides) for starting graft copolymerization.

Graft-modification of polysaccharides is widely research in recent years to obtain macromolecular materials greater to the source polysaccharides, exhibiting more preponderant resistance to heat or abrasion, more preponderant oil/water repellent qualities, or medicament activity and higher mechanical vigor. In their native form polysaccharides are fairly resistant to degradation under shear and perform as subsidiary flocculants (Singh *et al.*, 2000), but have poor shelf life because of their susceptibility to biodegradation. On the other hand synthetic polymers can be easily modified, but poor shear resistant is their properties. Natural polysaccharides grafted to synthetic polymers can be changed into highly customizable matrices with hybrid properties opportune for different applications (Gref *et al.*, 2002, Ohya, 2001).

1.2 Problem Statement

In the last two decades, efficient and sensitive analytical instrumentation have been introduced for various applications. However, sample preparation today remains considered as the most crucial step in the whole analytical process. Numerous sample preparation techniques have been developed with the following main goals: to improve the selectivity in extraction, to minimize the initial sample sizes, to facilitate the automation and to reduce or eliminate the volume of organic solvent involved in the extraction (Curylo *et al.*, 2007; Smith, 2003).

Based on the aims of sample preparation techniques, traditional LLE method does not fulfill current requirements and it has been displaced by new extraction techniques such as SPE, D- μ -SPE, μ -SPE, magnetic solid phase extraction (MSPE) and solid phase membrane tip extraction (SPMTE). Several comprehensive reviews on the current advances in sample preparation field and emphasizing the importance of sample preparation in the analytical process have been published (Turiel and Martín-Esteban, 2010; Augusto *et al.*, 2010).

In recent years, adsorption of toxic compounds by polysaccharides and modified polysaccharides is increasing because these sorbents are low-cost (Górecki *et al.*, 1999). Polysaccharides have poor shelf life and bad resistant in their natural forms but they can be changed into hybrid properties under grafting for various applications. To the best of our knowledge, there has been no report on agarose (as polysaccharide) grafted with methyl methacrylate copolymer using ceric ammonium nitrate (CAN) as redox initiator with microwave assisted method were synthesized and characterized.

Raw multiwall carbon nanotube (MWCNTs) are suitable for adsorption of toxic compounds but they are generally insoluble and severely aggregated, due to Van der Waals attraction among the nanotubes, creating a problem when homogenous dispersion is desirable, thus modification of MWCNTs is important. Modifications of MWCNTs by special functional groups or polymers have been proven to be more selective than raw or oxidized MWCNTs because of their excellent properties. Thus in this study, agarose was used as new natural polymer for the modification of MWCNTs and subsequent separation of lead as heavy metal.

1.3 Objectives of the Study

This study embarks on the following objectives:

1. To prepare graft polymer based on agarose grafted with methyl methacrylate using microwave-assisted method and to characterize the properties of agarose and graft copolymer.
2. To graft the Agarose-g-PMMA with MWCNTs and to characterize the physical properties of the products obtained using FTIR, FESEM and TGA.

3. To develop Agarose-g-PMMA micro-solid phase extraction combined with gas-chromatography with micro-electron capture detector for the analysis of selected pesticides in environmental water samples.
4. To develop and apply Agarose-g-PMMA dispersive micro-solid phase extraction combined with inductively plasma-mass spectrometry for the extraction and determination of selected heavy metals in vegetables and natural waters.
5. To develop and apply MWCNTs-g-Agarose-g-PMMA-SPE combined with flame atomic absorption spectrometric detection to the determination of lead ions in vegetables and natural waters.

1.4 Scope of the Study

This study was designed to prepare agarose-g-PMMA using microwave-assisted method. Characterization of agarose-g-PMMA was by using Fourier transformed infrared (FTIR) analysis to ascertain the presence of methyl methacrylate on the agarose backbone. Glass transition temperatures were studied by means of differential scanning calorimetry (DSC), which measures endothermic and exothermic heat flow of the copolymer (in relation to their chemical structure and molecular weight) as a function of temperature/time. Thermal stability of the selected graft copolymer was studied by thermogravimetric analysis (TGA), which is based on the continuous measurement of change in weight of sample as a function of temperature. Molecular weight and molecular weight distribution of the hydrolyzed grafted poly methyl methacrylate (PMMA) were also evaluated using gel permeation chromatography (GPC). CHN analysis was also carried out to determine the percentage elemental composition of carbon, hydrogen and oxygen. Development and application of Agarose-g-PMMA sorbent based on microextraction methods were investigated. Several important extraction parameters were optimized comprehensively and analytical performances of the developed method were evaluated in the study. Agarose-g-PMMA- μ -SPE and Agarose-g-PMMA-D- μ -SPE methods were demonstrated to extract selected pesticides and heavy metals using

REFERENCES

- Adams, S., Elias, A.J., and Cohen, P.S (Eds.). (1983). Recent Advances in Food Irradiation. *Elsevier Biomedical Press, Amsterdam*, 149.
- Ahmadi, F., Shahsavari, A. A. and Rahimi-Nasrabadi, M. (2008). Automated extraction and preconcentration of multiresidue of pesticides on a micro-solid-phase extraction system based on polypyrrole as sorbent and off-line monitoring by gas chromatography-flame ionization detection. *Journal of Chromatography A*, 1193(1-2), 26-31.
- Ajayan, P. M., Stephan, O., Colliex, C. and Trauth, D. (1994). Aligned carbon nanotube arrays formed by cutting a polymer resin-nanotube composite. *Science*, 265, 1212-4.
- Allcock, H. R., Lampe, F. W. and Mark, J. E. (2003). *Contemporary Polymer Chemistry*. Upper Saddle River, N.J.: Pearson Education/Prentice Hall.
- ALothman, Z. A., Habila, M., Yilmaz, E. and Soylak, M. (2012). Solid phase extraction of Cd(II), Pb(II), Zn(II) and Ni(II) from food samples using multiwalled carbon nanotubes impregnated with 4-(2-thiazolylazo)resorcinol. *Microchemica Acta*. 177, 397-403.
- Anastassiades, M., Lehotay, S. J., Stajnbaher, D. and Schenck, F. J. (2003). Fast and easy multiresidue method employing acetonitrile extraction/partitioning and "dispersive solid-phase extraction" for the determination of pesticide residues in produce. *Journal of AOAC International*. 86(2), 412-431.
- Arthur, C. L. and Pawliszyn, J. (1990). Solid phase microextraction with thermal desorption using fused silica optical fibers. *Analytical Chemistry*, 82, 2145-2148.
- Athawale, V. D. and Lele, V. (1998). Graft copolymerization onto starch. II. Grafting of acrylic acid and preparation of it's hydrogels. *Carbohydrate Polymers*, 35(1-2), 21-27.

- Augusto, F., Carasek, E., Silva, R. G. C., Rivellino, S. R., Batista, A. D., and Martendal, E. (2010). New Sorbents for Extraction and Microextraction Techniques. *Journal of Chromatography A*, 1217, 2533–2542.
- Aydin, F. A. and Soylak, M. (2007). A novel multi-element coprecipitation technique for separation and enrichment of metal ions in environmental samples. *Talanta*. 73, 134-141.
- Baek, J. B., Lyons, C. B. and Tan, L. S. (2004). Grafting of vapor-grown carbon nanofibers via in situ polycondensation of 3-phenoxybenzoic acid in poly(phosphoric acid). *Macromolecules*, 37, 8278-85.
- Bailón-Pérez, M. I., García-Campaña, A. M., del Olmo-Iruela, M., Gámiz-Gracia, L. and Cruces-Blanco, C. (2009). Trace determination of 10 β -lactam antibiotics in environmental and food samples by capillary liquid chromatography. *Journal of Chromatography A*, 1216, 8355-8361.
- Baltussen, E., Sandra, P., David, F. and Cramers, C. (1999). Stir Bar Sorptive Extraction (SBSE), a Novel Extraction Technique for Aqueous Samples: Theory and Principles. *Journal of Microcolumn Separation*, 11, 737–747.
- Banci, L., Bertini, I., Cantini, F. and Ciofi-Baffoni, S. (2010). Cellular copper distribution: a mechanistic systems biology approach. *Cellular and Molecular Life Science*, 67, 2563-2589.
- Basheer, C., Alnedhary, A. A., Rao, B. S. M. and Lee, H. K. (2009). Determination of carbamate pesticides using micro-solid-phase extraction combined with high performance liquid chromatography. *Journal of Chromatography A*, 1216, 211-216.
- Basheer, C., Alnedhary, A. A., Rao, B. S. M., Valliyaveetil, S. and Lee, H. K. (2006). Development and application of porous membrane-protected carbon nanotube micro-solid phase extraction combined with gas chromatography/mass spectrometry. *Analytical Chemistry*, 78, 2853-2858.
- Basheer, C., Chong, H. G., Hii, T. M. and Lee, H. K. (2007). Application of porous membrane protected micro-solid-phase extraction combined with HPLC for the analysis of acidic drugs in wastewater. *Analytical Chemistry*, 79(17), 6845–6850.
- Basheer, C., Narasimhan, K., Yin, M., Zhao, C., Choolani, M. and Lee, H. K. (2008). Application of micro-solid-phase extraction for the determination of

- persistent organic pollutants in tissue samples. *Journal of Chromatography A*, 1186, 358-364.
- Basheer, C., Wong, W., Makahleh, A., Tameem, A. A., Salhin, A., Saad, B. and Lee, H.K. (2011). Hydrazone-based ligands for micro-solid phase extraction-high performance liquid chromatographic determination of biogenic amines in orange juice. *Journal of Chromatography A*, 1218, 4332-4339.
- Baskaran, D., Dunlap, JR., Mays, J. W. and Bratcher MS. (2005). Grafting efficiency of hydroxy-terminated poly(methyl methacrylate) with multiwalled carbon nanotubes. *Macromol Rapid Commun.* 26, 481-6.
- Baskaran, D., Mays, J. W. and Bratcher, M. S. (2004). Polymer-grafted multiwalled carbon nanotubes through surface-initiated polymerization. *Angewandte Chemie International Edition*, 43, 2138-42.
- Bati, B. and Cesur, H. (2002). Determination of Copper in Edible Oils by Atomic Absorption Spectrometry after Lead Piperazinedithiocarbamate Solid-Phase Extraction and Potassium Cyanide Back-Extraction. *Analytical Science*, 18, 1273-1274.
- Benhabib, K. and Town, R. M. (2012). Dynamic solid phase microextraction (SPME) of atrazine at PDMS and PA coated fibers. *Chemical Engineering Transation*,. 29, 151-156.
- Bubb, J. M. and Lester, J. N. (1991). The impact of heavy metals on lowland rivers and the implications for man and the environment. *Science of the Total Environment*, 100, 207-233.
- Cai, Y. Q., Jiang, G. B., Liu, J. F. and Zhou, Q. X. (2003). Multiwalled carbon nanotubes as a solid-phase extraction adsorbent for the determination of bisphenol A, 4-nonylphenol, and 4-tert-octylphenol. *Analytical Chemistry*, 75, 2517-2521.
- Camel, C. (2003). Review Solid Phase Extraction of Trace Elements, *Spectrochim. Acta B*. 58, 1177–1233.
- Carletto, J. S., Roux, K. C., Maltez, H. F., Martendal, E. and Carasek, E. (2008). Use of 8-hydroxyquinoline-chitosan chelating resin in an automated on-line preconcentration system for determination of zinc(II) by F AAS. *Journal Of Hazardous Materials*, 157, 88-93.
- Cavaliere, B., Monteleone, M., Naccarato, A., Sindona, G. and Jagarelli, A. (2012). A solid-phase microextraction-gas chromatographic approach combined with

- triple quadrupole mass spectrometry for the assay of carbamate pesticides in water samples. *Journal of Chromatography A*, 1257, 149-157.
- Celik, M. (2006). Preparation and characterization of starch-gpolymethacrylamide copolymers. *Journal of Polymer Research*, 13, 427–32.
- Chao, C., Man, H. and Bin, H. (2011). Membrane solid phase microextraction with alumina hollow fiber on line coupled with ICP-OES for the determination of trace copper, manganese and nickel in environmental water samples. *Journal of Hazardous Materials*, 187, 379-385.
- Chen, D. H., Hu, B. and Huang, C. Z. (2009). Chitosan modified ordered mesoporous silica as micro-column packing materials for on-line flow injection-inductively coupled plasma optical emission spectrometry determination of trace heavy metals in environmental water samples. *Talanta*, 78, 491-497.
- Chen, J., Hamon, M. A., Hu, H., Chen, Y., Rao, AM., Eklund, P. C. and et al. (1998). Solution properties of single-walled carbon nanotubes. *Science*, 282, 95-98.
- Chen, J., Rao, A. M., Lyuksyutov, S., Itkis, M. E., Hamon, M. A., Hu, H. and et al. (2001). Dissolution of full-length single-walled carbon nanotubes. *Journal of Physical Chemistry B*, 105, 2525-2528.
- Ciucanu, I., Swallow, K. C. and Caprita, R. (2004). Micro-solid phase extraction with helical solid-sorbent in the presence of organic solvent for gas chromatography–mass spectrometry analysis of per-O-methylated mono- and disaccharides. *Analytica Chimica Acta*, 519(1), 93–101.
- Crescenzi, V., Dentini, M., Risica, D., Spadoni, S., Skjak-Brak, G., Capitani, D., Mannina, L. and Viel S. (2004). C(6)-oxidation followed by C(5)-epimerization of guar gum studied by high field NMR. *Biomacromolecules*, 5, 537–46.
- Cui, Y. M., Chang, X. J., Zhu, X. B., Jiang, N., Hu, Z. and Lian, N. (2007). Nanometer SiO₂ modified with 5-sulfosalicylic acid as selective solid-phase extractant for Fe(III) determination by ICP-AES from biological and natural water samples. *Microchemical Journal*. 86(1), 23-28.
- Curyło, J., Wardencki, W., and Namieśnik, J. (2007). Green Aspects of Sample Preparation – a Need for Solvent Reduction. *Polish Journal of Environmental Science*, 16, 5- 16.

- Dagnac, T., Garcia-Chao, M., Pulleiro, P., Garcia-Jares, C. and Llompart, M. (2009). Dispersive solid-phase extraction followed by liquid chromatography-tandem mass spectrometry for the multi-residue analysis of pesticides in raw bovine milk. *Journal of Chromatography A*, 1216, 3702-3709.
- Dallinger, D. and Kappe, CO. (2007). Microwave-assisted synthesis in water as solvent. *Chemical Reviews*, 107, 2563–91.
- Dean, J.R. (1998). *Extraction Methods for Environmental Analysis*, Wiley, New York.
- Ebdon, J. R. (1991). New methods for polymer synthesis. *Glasgow u.a: Blackie*, New York : Chapman and Hall, vol 2.
- Edgar, K. J., Buchanan, C. M., Debenham, J. S., Rundquist, P. A. and Seiler, B. D., Shelton, MC., Tindall, D. (2001). Advances in cellulose ester performance and application. *Progress in Polymer Science*, 26, 1605–88.
- El Tahlawy, K., and Hudson, S. M. (2003). Synthesis of a well-defined chitosan graft poly(methoxy polyethyleneglycol methacrylate) by atom transfer radical polymerization. *Journal of Applied Polymer Science*, 89(4), 901-912.
- El-Hefian, E. A., Nasef, M. M., Yahaya, A. H. and Khan, R. A. (2010). Preparation and Characterization of Chitosan/Agar Blends: Rheological and Thermal Studies. *Journal of Chilean Chemical Society*, 55, 130-136.
- El-Sheikh, A. H. (2008). Effect of oxidation of activated carbon on its enrichment efficiency of metal ions: comparison with oxidized and non-oxidized multi walled carbon nanotubes. *Talanta*. 75, 127-134.
- El-Sheikh, A. H., Insisi, A. A. and Sweileh, J. A. (2007a). Effect of oxidation and dimensions of multi-walled carbon nanotubes on solid phase extraction and enrichment of some pesticides from environmental waters prior to their simultaneous determination by high performance liquid chromatography. *Journal of Chromatography A*, 1164, 25-32.
- El-Sheikh, A. H., Sweileh, J. A. and Al-Degs, Y. S. (2007b). Effect of dimensions of multi-walled carbon nanotubes on its enrichment efficiency of metal ions from environmental waters. *Analytical Chemistry Acta*. 604, 119-126.
- Escudero, L. A., Martinez, L. D., Salonia, J. A. and Gasquez, J. A. (2010). Determination of Zn(II) in natural waters by ICP-OES with on-line preconcentration using a simple solid phase extraction system. *Microchemical Journal*, 95, 164-168.

- Ezoddin, M., Shemirani, F., Abdi, Kh., Saghezchi, M.K. and Jamali, M. R. (2010). Application of modified nano-alumina as a solid phase extraction sorbent for the preconcentration of Cd and Pb in water and herbal samples prior to flame atomic absorption spectrometry determination. *Journal of Hazardous Materials*. 178, 900-905.
- Fagerquist, C. K., Lightfield, A. R. and Lehotay, S. J. (2005). Confirmatory and quantitative analysis of beta-lactam antibiotics in bovine kidney tissue by dispersive solid-phase extraction and liquid chromatography-tandem mass spectrometry. *Analytical Chemistry*, 77(5), 1473-82.
- Fang, G. Z., He, J. X. and Wang, S. (2006). Multiwalled carbon nanotubes as sorbent for on-line coupling of solid-phase extraction to high-performance liquid chromatography for simultaneous determination of 10 sulfonamides in eggs and pork. *Journal of Chromatography A*, 1127, 12-17.
- Fares, M. M., El-faqeeh, A. S. and Osman, M. E. (2003). Graft copolymerization onto starch. I. Synthesis and optimization of starch grafted with Ntert-butylacrylamide copolymer and its hydrogels. *Journal of Polymer Research*, 10,119–25.
- Fernando, K. A. S., Lin, Y., Zhou, B., Grah, M., Joseph, R. and Allard, LF. (2005). Poly(ethylene-co-vinyl alcohol) functionalized single-walled carbon nanotubes and related nanocomposites. *Journal of Nanoscience and Nanotechnol.* 5, 1050-5.
- Fontana, A. R., Camargo, A., Martinez, L. D. and Altamirano, J. C. (2011). Dispersive solid-phase extraction as a simplified clean-up technique for biological sample extracts. Determination of polybrominated diphenyl ethers by gas chromatography-tandem mass spectrometry. *Journal of Chromatography A*, 1218, 2490- 2496.
- Galanos, C., Luderitz, O. and Himmelsbach, K. (2004). The partial acid hydrolysis of polysaccharides: a new method for obtaining oligosaccharides in high yield. *European Journal of Biochemistry*, 8, 332–6.
- Gilart, N., Cormack, P. A. G., Maarce, R. M., Borrull, S. and Fontanals, N. (2013). Preparation of a Polar Monolithic Coating for Stir bar Sorptive Extraction of Emerging Contaminants from Wastewaters. *Journal of Chromatography A*, 1295, 42-47.

- González-Curbelo, M. Á., Herrera-Herrera, A.V., Hernández-Borges, J. and Rodríguez-Delgado, M. Á. (2013). Analysis of pesticides residues in environmental water samples using multiwalled carbon nanotubes dispersive solid-phase extraction. *Journal of Separation Science*, 36, 556-563.
- Górecki, T., Yu, X. and Pawliszyn, J. (1999). Theory of analyte extraction by selected porous polymer SPME fibres, *Analyst*, 124, 643–649.
- Gref, R., Rodrigues, J. and Couvrceur, P. (2002). Polysaccharides grafted with polyesters: novel amphiphilic copolymers for biomedical applications. *Macromolecules*, 35, 9861–7.
- Guan, W., Li, Z., Zhang, H., Hong, H., Rebeyev, N., Ye, Y. and Ma, Y. (2013). Amine modified graphene as reversed-dispersive solid phase extraction materials combined with liquid chromatography-tandem mass spectrometry for pesticide multi-residue analysis in oil crops. *Journal of Chromatography A*, 1286, 1-8.
- Guilmeau, I., Esnouf, S., Betz, N., and Le , M.A. (1997). Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms, *Elsevier*, 131, 270–275.
- Gupta, B., Begum, Z. and Rajput G. (2007). Equilibrium and Kinetic Studies for the Adsorption of Mn(II) and Co(II) from Aqueous Media using Agar-agar as Sorbent. *Research Journal of chemistry and Environment*, 11, 2.
- Gupta, S., Sharma, P. and Soni, P. L. (2004). Carboxymethylation of Cassia occidentalis seed gum. *Journal of Applied Polymer Science*, 94, 1606–1611.
- Hemon, M. A., Chen, J., Hu, H., Chen, Y., Itkis, M. E., Rao, A. M. and et al. (1999). Dissolution of single-walled carbon nanotubes. *Advanced Materials*, 11, 834-40.
- Hu, N., Zhou, H., Dang, G., Rao, X., Chen, C. and Zhang, W. (2007). Efficient dispersion of multi-walled carbon nanotubes by in situ polymerization. *Polymere Internatinal*, 56, 655-9.
- Hu, Y., S. Zhou, and Wu, L. (2009). Surface mechanical properties of transparent poly(methyl methacrylate)/zirconia nanocomposites prepared by in situ bulk polymerization. *Polymer*, 50, 3609-3616.
- Huang, X. P., Chang, X. J., He, Q., Cui, Y. M., Zhai, Y. H. and Jiang, N. (2008). Tris (2-aminoethyl) amine functionalized silica gel for solid-phase extraction and

- preconcentration of Cr(III), Cd(II) and Pb(II) from waters. *Journal of Hazardous Materials*. 157(1), 154-160.
- Iijima, S. (1991). Helical microtubules of graphitic carbon, *Nature*, 354, 56-58.
- Ikhuria, E. U., Folayan, A. S., Okieimen, F. E. (2010). Studies in the graft copolymerization of acrylonitrile onto cassava starch by ceric ion induced initiation. *International Journal of Biotechnology And Molecular Biology Research*, 1, 10-14.
- Jia, Z., Wang, Z., Xu, C., Liang, J., Wei, B., Wu, D. and et al. (1999). Study on poly(methyl methacrylate)/carbon nanotube composites. *Material Science Engineering A*, 271(1-2), 395-400.
- Jiménez-Soto, J. M., Cárdenas, S. and Valcárcel, M. (2012). Evaluation of singlewalled carbon nanohorns as sorbent in dispersive micro solid-phase extraction. *Analytical Chemistry Acta*, 714, 76-81.
- Jin, Z., Sun, X., Xu, G., Goh, S. H. and Ji, W. (2000). Nonlinear optical properties of some polymer/multi-walled carbon nanotube composites. *Chemical Physics Letters*, 318, 505-10.
- Kaewtatip, K., and Tanrattanakul, V. (2008). Preparation of cassava starch grafted with polystyrene by suspension polymerization. *Carbohydrate Polymers*, 73(4), 647-655.
- Kang, X., Ma, W., Zhang, H-L., Xu, Z-G., Guo, Y. and Xiong, Y. (2008). Vinyl-Carbon Nanotubes for Composite Polymer Materials. *Journal of Applied Polymer Science*, 110, 1915-1920.
- Kanimozhi, S., Basheer, C., Narasimhan, K., Liu, L., Koh, S., Xue, F., Choolani, M. and Lee, H.K. (2011). Application of porous membrane protected microsolid-phase extraction combined with gas chromatography-mass spectrometry for the determination of estrogen in ovarian cyst fluid samples. *Analytica Chimica Acta*. 687, 56-60.
- Karimipour, G., Ghaedi, M., Sahraei, M., Daneshfar, R. and Biyareh, M.N. (2012). Modification of gold nanoparticle loaded on activated carbon with bis (4 methoxysalicylaldehyde) -1,2-phenylenediamine as new sorbent for enrichment of some metal ions. *Biological Trace Element Research*, 145, 109-117.
- Kataoka, H. (2003). New trends in sample preparation for clinical and pharmaceutical analysis. *Trends in Analytical Chemistry*, 22, 232-244.

- Kidwai, M., Misra, P. and Bhushan, K. R. (2001). Alumina-supported synthesis of thiadiazolyl thiazolothiones. *Synthetic Communication*, 31, 817-22.
- Kim, S. W., Kim, T., Kim, Y. S., Choi, H. S., Lim, H. J., Yang, S. J. and Park, C. R. (2012). Surface modifications for the effective dispersion of carbon nanotubes in solvents and polymers. *Carbon*, 50, 3-33.
- Koh, Y. (2001). Zinc and disease of the brain. *Mol. Neurobiol.* 24, 99-106.
- Kong, H., Gao, C. and Yan, D. (2004a). Functionalization of multiwalled carbon nanotubes by atom transfer radical polymerization and defunctionalization of the products. *Macromolecules*, 37, 4022-30.
- Kong, H., Gao, C. and Yan, D. (2004b). Constructing amphiphilic polymer brushes on the convex surfaces of multi-walled carbon nanotubes by in situ atom transfer radical polymerization. *Journal of Material Chemistry*, 14, 1401-5.
- Kong, H., Gao, C. and Yan, D. (2004d) Controlled functionalization of multiwalled carbon nanotubes by in situ atom transfer radical polymerization. *Journal of American Chemistry Society*, 126(2), 412-3.
- Kong, H., Li, W., Gao, C., Yan, D., Jin, Y. and Walton, D. R. M., et al. (2004c). Poly(Nisopropylacrylamide)-coated carbon nanotubes: temperaturesensitive molecular nanohybrids in water. *Macromolecules*, 37, 6683-6.
- Kong, H., Luo, P., Gao, C. and Yan, D. (2005). Polyelectrolyte-functionalized multiwalled carbon nanotubes: preparation, characterization and layer-by-layer self-assembly. *Polymer*, 46, 2472-85.
- Kurita, K., Inoue, M. and Harata, M. (2002). Graft Copolymerization of Methyl Methacrylate onto Mercaptochitin and Some Properties of the Resulting Hybrid Materials. *Biomacromolecules*. 3, 147-152.
- Lagos, A., and Reyes, J. (1988). Grafting onto chitosan. I. Graft copolymerization of methyl methacrylate onto chitosan with fenton's reagent ($\text{Fe}^{2+} - \text{H}_2\text{O}_2$) as a redox initiator. *Journal of Polymer Science, Part A: Polymer Chemistry*, 26(4), 985-991.
- Lee, T. P., Saad, B., Khayoon, W. S. and Salleh, B. (2012). Molecularly imprinted polymer as sorbent in micro-solid phase extraction of ochratoxin A in coffee, grape juice and urine. *Talanta*, 88, 129–135.
- Li, Y. H., Wang, S., Luan, Z., Ding, J., Xu, C. and Wu, D. (2003). Adsorption of cadmium(II) from aqueous solution by surface oxidized carbon nanotubes, *Carbon*. 41, 1057-1062.

- Liang, P., Liu, Y. and Guo, L. (2005). Determination of trace rare earth elements by inductively coupled plasma atomic emission spectrometry after preconcentration with multiwalled carbon nanotubes. *Spectrochim. Acta B*, 60, 125-129.
- Lin, Y., Zhou, B., Fernando, K. A. S., Liu, P., Allard, L.F. and Sun, YP. Polymeric carbon nanocomposites from carbon nanotubes functionalized with matrix polymer. *Macromolecules*, 36, 7199-204.
- Liu, J, Rinzler, A. G., Dai, H., Hafner, J. H., Bradley, R. K., Boul, P. J. and et al. (1998). Fullerene pipes. *Science*, 280, 1253-6.
- Liu, X. Y., Ji, Y. S., Zhang, Y. H., Zhang, H. X. and Liu, M. C. (2007). Oxidized multiwalled carbon nanotubes as a novel solid-phase microextraction fiber for determination of phenols in aqueous samples. *Journal of Chromatography A*, 1165, 10-17.
- Liu, Y., Li, Y. and Yan, X. P. (2008). Preparation, characterization, and application of L-Cysteine functionalized multiwalled carbon nanotubes as a selective sorbent for separation and preconcentration of heavy metals. *Advanced Functional Materials*, 18, 1536-1543.
- Liu, Z., Wu, G., and Liu, Y. (2006). Graft copolymerization of methyl acrylate onto chitosan initiated by potassium diperiodatoargentate (III). *Journal of Applied Polymer Science*, 101, 799-804.
- Maggi, L., Zalacain, A. Mazzoleni, V., Alonso, G. L. and Salinas, M. R. (2008). Comparison of stir bar sorptive extraction and solid-phase microextraction to determine halophenols and halonitrobenzenes by gas chromatography-ion trap tandem mass spectrometry. *Talanta*, 75, 753-759.
- Maiti, S., Ranjit, S., Biswanath, S. (2010). Polysaccharide-based graft copolymers in controlled drug delivery. *International Journal of Pharmacy and Technology*, 2, 1350-8.
- Martin-Esteban, A. (2013). Molecular-Imprinted Polymers as a Versatile, Highly Selective Tool in Sample Preparation. *Trends Analytical Chemistry*, 45, 169-181.
- Matahwa, H., Ramiah, V., Jarrett, WL., Mcleary, JB. and Sanderson, RD. (2007). Microwave assisted graft copolymerization of n-isopropyl acrylamide and methyl acrylate on cellulose: solid state NMR analysis and CaCO₃ crystallization. *Macromolecular Symposia*, 255, 50-6.

- Matyjaszewski, K. and Gaynor, S. G. (2000). Applied Polymer Science 21st Century, Ed. C.C. Craver and C.E. Carraher, Jr., Elsevier, Amsterdam.,929-977.
- Mingos, D. and Baghurst, DR. (1991). Applications of microwave dielectric heating effects to synthetic problems in chemistry. *Chemical Society Reviews*, 20, 1–47.
- Mishra, S. and Sen, G. (2011). Microwave initiated synthesis of polymethylmethacrylate grafted guar (GG-g-PMMA), characterizations and applications. *International Journal of Biological Macromolecules*, 48, 688-694.
- Mishra, S., Sen, G., Rani, G. U., and Sinha, S. (2011). Microwave assisted synthesis of polyacrylamide grafted agar (Ag-g-PAM) and its application as flocculant for wastewater treatment. *International Journal of Biological Macromolecules*, 49(4), 591-598.
- Mishra, S., Sinha, S., Dey, K. P. and Sen, G. (2014). Synthesis, characterization and applications of polymethylmethacrylate grafted psyllium as flocculant. *Carbohydrate Polymers*, 99, 462-468.
- Mitra, S. (2003). *Sample Preparation Techniques in Analytical Chemistry*. New Jersey: John Wiley & Sons, Inc.
- Morel, F. M. M. and Price, N. M. (2003). The biogeochemical cycles of trace metals in the oceans. *Science*, 300, 944-947.
- Munoz, J., Gallego, M. and Valcarcel, M. (2005). Speciation of organometallic compounds in environmental samples by gas chromatography after flow preconcentration on fullerenes and nanotubes. *Analytical Chemistry*, 77, 5389-5395.
- Neas, E. D. and Collins, M. J. (1988). Microwave heating: theoretical concepts and equipment design. In: Kingston HM, Jassie LB, editors. Introduction to microwave sample preparation: theory and practice. Washington, DC: American Chemical Society, 7–22.
- Nishioka, N., and Kosai, K. (1981). Homogeneous graft copolymerization of vinyl monomers onto cellulose in a dimethyl sulfoxide-paraformaldehyde solvent system - 1. Acrylonitrile and methyl methacrylate. *Polymer Journal*, 13(12), 1125-1133.
- Nishioka, N., Matsumoto, K. and Kosai, K. (1983). *Homogeneous graft copolymerization of vinyl monomers onto cellulose in a dimethyl*

sulphoxide/paraformaldehyde solvent system. II. Characterization of graft copolymers.

- Nishioka, N., Matsumoto, Y., Yumen, T., Monmae, K., and Kosai, K. (1986). Homogeneous graft copolymerization of vinyl monomers onto cellulose in a dimethyl sulfoxide-paraformaldehyde solvent system. IV. 2-Hydroxyethyl methacrylate. *Polymer Journal, (Tokyo)*, 18, 323-330.
- Nishioka, N., Minami, K., and Kosai, K. (1983). Homogeneous graft copolymerization of vinyl monomers onto cellulose in a dimethyl sulfoxide-paraformaldehyde solvent system. III. *Methyl acrylate*. *Polymer Journal, (Tokyo)*, 15, 591-596.
- Niyogi, S., Hamon, M. A., Hu, H., Zhao, B., Bhowmik, P. and Sen, R. (2002). Chemistry of single-walled carbon nanotubes. *Accounts Chemical Research.*, 35, 1105-13.
- Ohya, Y., Maruhashi, S., Hirano, T. and Ouchi, T. (2001). Preparation of poly(lactic acid)-grafted polysaccharides as biodegradable amphiphilic materials. In: Chiellini E, Sunamoto J, Migliaresi C, Ottenbrite RM, Cohn D, editors. *Biomedical polymers and polymer therapeutics. New York: Kluwer Academic/Plenum publishers*, 139–48.
- Ozcan, S. G., Satiroglu, N. and Soylak, M. (2010). Column solid phase extraction of iron(III), copper(II), manganese(II) and lead(II) ions food and water samples on multi-walled carbon nanotubes. *Food and Chemical Toxicology*. 48, 2401-2406.
- Park, H., Zhao, J. and Lu, J. P. (2006). Effects of sidewall functionalization on conducting properties of single wall carbon nanotubes. *Nano Letters*, 6, 916-9.
- Parkm, S. J., Chom, M. S., Lim, S. T., Cho, H. J. and Jhon, M. S. (2003). Synthesis and dispersion characteristics of multi-walled carbon nanotube composites with poly(methyl methacrylate) prepared by in-situ bulk polymerization. *Macromol Rapid Commun*, 24, 1070-3.
- Pawliszyn, J. (1997). *Solid Phase Microextraction : Theory and Practise*. (1st ed.) New York : Wiley-VCH.
- Pawliszyn, J. and Pedersen-Bjergaard, S. (2006). Analytical Microextraction: Current status and future trends. *Journal of Chromatographic Science*, 44, 291-307.

- Plössl, F., Giera, M. and Bracher, F. (2006). Multiresidue analytical method using dispersive solid-phase extraction and gas chromatography/ion trap mass spectrometry to determine pharmaceuticals in whole blood. *Journal of Chromatography A*, 1135(1), 19-26.
- Polshettiwar, V. and Varma, R. S. (2008). Microwave-assisted organic synthesis and transformations using benign reaction media. *Acc Chem Res*, 41, 629-39.
- Poole, C. F. (2003). New Trends in Solid-Phase Extraction. *Trends Analytical Chemistry*, 22, 362-373.
- Poole, C. F., Gunatilleka, A. D., and Sethuraman., R. (2000). Review – Contributions of Theory to Method Development in Solid-Phase Extraction, *Journal of Chromatography A*, 885, 17-39.
- Poole, C.F. (2003). New trends in solid-phase, extraction. *Trends Analytical Chemistry*, 22, 362-373.
- Posyniak, A., Zmudzki, J. and Mitrowska, K. (2005). Dispersive solid-phase extraction for the determination of sulfonamides in chicken muscle by liquid chromatography. *Journal of Chromatography A*, 1087(1-2), 259-64.
- Pourreza, N. and Ghanemi, K. (2009). Determination of mercury in water and fish samples by cold vapor atomic absorption spectrometry after solid phase extraction on agar modified with 2-mercaptobenzimidazole. *Journal of Hazardous Materials*, 161, 982–987.
- Pourreza, N., Mouradzadegan, A., and Mohammadi, S. (2011). Solid Phase Extraction of Zirconium as Arsenazo(III) Complex on Agar and Spectrophotometric Determination. *Journal of Iranian Chemical Society*, 8(4), 951-957.
- Prasad, K., Meena, R. and Siddhanta, A. K. (2006a). Microwave-Induced Rapid One-Pot Synthesis of κ -Carrageenan-g-PMMA Copolymer by Potassium Persulphate Initiating System, *Journal of Applied Polymer Science*, 101, 161–166.
- Prasad, K., Mehta, G., Meena, R. and Siddhanta, AK. (2006b). Hydrogel-forming agar-graft PVP and κ carrageenan graft-PVP blends: rapid synthesis and characterization. *Journal of Applied Polymer Science*, 102, 3654-63.
- Prieto, A., Basauri, O., Rodil, R., Usobiaga, A., Fernández, L. A., Etxebarria, N., and Zuloaga, O. (2010). Stir-Bar Sorptive Extraction: A View on Method

- Optimisation, Novel Applications, Limitations and Potential Solutions. *Journal of Chromatography A*, 1217, 2642–2666.
- Pyrzyska, K. (1999). Functionalized cellulose sorbents for on-line preconcentration of trace metals for environmental analysis, *Critical Reviews Analytical Chemistry*, 29, 313-321.
- Qin, S., Qin, D., Ford, W. T., Herrera, J. E. and Resasco, D. E. (2004). Grafting of poly(4- vinylpyridine) to single-walled carbon nanotubes and assembly of multilayer films. *Macromolecules*, 37, 9963-7.
- Qin, S., Qin, D., Ford, W., Resasco, D. E. and Herrera, J. E. (2004). Polymer brushes on single-walled carbon nanotubes by atom transfer radical polymerization of n-butyl methacrylate. *Journal of the American Chemical Society*, 126(1), 170-176.
- Qu, L., Veca, L. M., Lin, Y., Kitaygorodskiy, A., Chen, B., McCall, A. M. and et al. (2005). Soluble nylon-functionalized carbon nanotubes from anionic ringopening polymerization from nanotube surface. *Macromolecules*, 38, 10328-31.
- Racamonde, I., Rodil, R., Quintana, J. B. and Cela, R. (2013). In-sample derivatization-solid-phase microextraction of amphetamines and ecstasy related stimulants from water and urine. *Analytical Chemistry Acta*. 770, 75-84.
- Ramaprasad, A.T., Rao, V., Sanjeev, G., Ramanani, S.P. and Sabharwal, S. (2009). Grafting of polyaniline onto the radiation crosslinked chitosan. *Synthetic Metterials* ,159, 1983–90.
- Rani, P., Mishra, S. and Sen, G. (2013). Microwave based synthesis of polymethyl methacrylate grafted sodium alginate: Its application as flocculant. *Carbohydrate Polymers*, 91(2), 686-692.
- Richard, C., Balavoine, F., Schultz, P., Ebbesen, T. W. and Mioskowski, C. (2003). Supramolecular self-assembly of lipid derivatives on carbon nanotubes. *Science*, 300, 775-778.
- Riggs, J. E., Guo, Z., Carroll, D. L. and Sun, Y. P. (2000). Strong luminescence of solubilized carbon nanotubes. *Journal of the American Chemical Society*, 122, 5879-80.

- Ruan, S. L., Gaob, P., Yangb, X. G. and Yu, T. X. (2003). Toughening high performance ultrahigh molecular weight polyethylene using multiwalled carbon nanotubes. *Polymer*, 44(19), 5643-5654.
- Salgueiro, M. J., Zubillaga, M., Lysionek, A., Sarabia, M. I., Caro, R., De Paoli, T., Hager, A. and Weill, R. (2005). Zinc as an essential micro nutrient: a review, *Nutr. Res. Journal of Boccio*, 20, 737-755.
- Samiey, B. and Ashoori, F. (2012). Adsorptive removal of methylene blue by agar: effects of NaCl and ethanol. *Chemistry Central Journal*, 6, 14.
- Sand, A., Yadav, M., Mishra, D. K. and Behari, K. (2010). Modification of alginate by grafting of N-vinyl-2-pyrrolidone and studies of physicochemical properties in terms of swelling capacity, metal-ion uptake and flocculation. *Carbohydrat Polymer*, 80, 1147-54.
- Sanghi, R. and Bhattacharya, B. (2005). Comparative evaluation of natural polyelectrolytes psyllium and chitosan for decolorisation of dye solutions. *Water Quality Research Journal*, 40, 97-101.
- Sanghi, R., Bhattacharya, B. and Singh, V. (2002). Cassia angustifolia seed gum as an effective natural coagulant for decolourisation of dye solutions. *Green Chemistry*, 4, 252-254.
- Santagada, V., Frecentese, F., Perissutti, E., Favretto, L., and Caliendo, G. (2004). The application of microwaves in combinatorial and high-throughput synthesis as new synthetic procedure in drug discovery. *QSAR and Combinatorial Science*, 23, 919-44.
- Saracoglu, S., Soylak, M. and Elic, L. (2003). Determination of trace amounts of copper in natural water samples by flame atomic absorption spectrometry coupled with flow injection on-line solid phase extraction on amborsorb 563 adsorption resin. *Chemia Analityczna*. 48, 77-85.
- Saraydin, D., Unver-Saraydin, S., Karada, E., Koptagel, E., Guven, O. (2004). In vivo biocompatibility of radiation crosslinked acrylamide copolymers. *Nuclear Instruments and Methods in Physics Research Section B*, 217, 281-92.
- Schummer, C., Tuduri, L., Briand, O., Appenzeller, B. M. and Millet, M. (2012). Application of XAD-2 resin-based passive samplers and SPME-GC-MS/MS analysis for the monitoring of spatial and temporal variations of atmospheric pesticides in Luxembourg. *Environmental Pollution*, 170, 88-94.

- See, H. H., Sanagi, M. M., Ibrahim, W. A.W. and Naim, A. A. (2010). Determination of triazine herbicides using membrane tip extraction prior to micro-liquid chromatography. *Journal of Chromatography A*, 1217, 1767-1772.
- Sen, G., Singh, RP. and Pal, S. (2010). Microwave-initiated synthesis of polyacrylamide grafted sodium alginate: synthesis and characterization. *Journal of Applied Polymer Science*, 115, 63–71.
- Sergi, M., Battista, N., Montesano, C., Curini, R., Maccarrone, M. and Compagnone, D. (2013). Determination of the two major endocannabinoids in human plasma by μ -SPE followed by HPLC-MS/MS. *Analytical and Bioanalytical Chemistry*, 405, 785-793.
- Shams, E. and Torabi, R. (2006). Determination of nanomolar concentrations of cadmium by anodic-stripping voltammetry at a carbon paste electrode modified with zirconium phosphated amorphous silica. *Sensors and Actuators B*, 117, 86-92.
- Sharma, B. R., Kumar, V. and Soni, P. L. (2003). Ce(IV)-ion initiated graft copolymerization of methyl methacrylate onto guar gum. *Journal of Macromolecul Science A*, 40, 49–60.
- Silva, E. L., Roldan, P. S. and Gine, M. F. (2009). Simultaneous preconcentration of copper, zinc, cadmium, and nickel in water samples by cloud point extraction using 4-(2- pyridylazo)-resorcinol and their determination by inductively coupled plasma optic emission spectrometry. *Journal of Hazardous Materials*, 171, 1133-1138.
- Singh, R. P., Tripathy, T., Karmakar, G. P., Rath, S. K., Karmakar, N. C., Pandey, S. R., Kannan, K., Jain, S. K. and Lan, N. T. (2000). Novel biodegradable flocculants based on polysaccharides. *Current Science*, 78, 798-803.
- Singh, V. and Tripathi, DN. (2006). Microwave promoted grafting of acrylonitrile onto Cassia siamea seed gum. *Journal of Applied Polymer Science*, 101, 2384-2389.
- Singh, V., Kumari, PL., Tiwari, A. and Pandey, S. (2010). Alumina-supported microwave synthesis of Cassia marginata seed gum-graftpolyacrylamide. *J Journal of Applied Polymer Science*, 117, 3630-8.
- Singh, V., Sharma, A. K. and Maurya, S. (2009). Efficient cadmium(II) removal from aqueous solution using microwave synthesized guar-graftpoly(ethylacrylate). *Industrial & Engineering Chemistry Research*, 48, 4688-96.

- Singh, V., Sharma, A.K., Kumari, P. and Tiwari, S. (2008). Efficient chromium(VI) adsorption by *Cassia marginata* seed gum functionalized with poly(methylmethacrylate) using microwave irradiation. *Industrial & Engineering Chemistry Research*, 47, 5267-76.
- Singh, V., Tiwari, A., Pandey, S. and Singh, S.K. (2006b). Microwave-accelerated synthesis and characterization of potato starch-g-poly(acrylamide). *Starch/Starke*, 58, 536-43.
- Singh, V., Tiwari, A., Tripathi, D. N. and Sanghi, R. (2004). Microwave assisted synthesis of guar-g-polyacrylamide. *Carbohydrate Polymer*, 58,1-6.
- Singh, V., Tiwari, A., Tripathi, D.N. and Sanghi, R. (2005). Microwave promoted synthesis of chitosan-graft-poly(acrylonitrile). *Journal of Applied Polymer Science*, 95, 820-5.
- Singh, V., Tiwari, A., Tripathi, D.N. and Sanghi, R. (2006a). Microwave enhanced synthesis of chitosan-graft-polyacrylamide. *Polymer*, 47, 254-60.
- Singh, V., Tripathi, D.N., Tiwari, A. and Sanghi, R. (2006c). Microwave synthesized chitosan-graft-poly(methylmethacrylate): an efficient Zn^{2+} ion binder. *Carbohydrate Polymer*, ;65, 35-41.
- Smith, R. M. (2003). Review Before the Injection—Modern Methods of Sample Preparation for Separation Techniques. *J. Chromatogr. A*. 1000, 3-27.
- Soylak, M. and Elic, L. (2000). Solid phase extraction of trace metal ions in drinking water samples from Kayseri – Turkey. *Journal of Trace and Microprobe Techniques*. 18, 397-403.
- Soylak, M. and Tuzan, M. (2008). Coprecipitation of gold(III), palladium(II) and lead(II) for their flame atomic absorption spectrometric determinations. *Journal of Hazardous Materials*. 152, 656-661.
- Spietelun, A., Marcinkowski, L., Guardia, M. and Namiesnik, J. (2014). Green Aspects, Developments and Perspectives of Liquid Phase Microextraction Techniques. *Talanta*. 119, 34-45.
- Stafiej, A. and Pyrzynska, K. (2008). Solid phase extraction of metal ions using carbon nanotubes, *Microchemistry Journal*, 89, 29-33.
- Stevens, M. P. (1999). *Polymer chemistry: An Introduction* (3rd Ed ed.). New York: Oxford University Press.
- Streit, B. and Winter, S. (1993). Cadmium uptake and compartmental time characteristics in the freshwater mussel. *Chemosphere*, 26, 1479-1490.

- Su, S., Chen, B., He, M., Hu, B. and Xiao, Z. (2014). Determination of trace/ultra-trace rare earth elements in environmental samples by ICP-MS after magnetic solid phase extraction with Fe₃O₄@SiO₂@polyaniline-graphene oxide composite. *Talanta*, 119, 458-466.
- Suleiman, J. S., Hu, B., Peng, H. and Huang, Ch. (2009). Separation/preconcentration of trace amounts of Cr, Cu and Pb in environmental samples by magnetic solid-phase extraction with Bismuthiol-II-immobilized magnetic nanoparticles and their determination by ICP-OES. *Talanta*. 77, 1579-1583.
- Tasis, D., Tagmatarchis, N., Bianco, A. and Prato, M. (2006). Chemistry of carbon nanotubes. *Chemical Reviews*. 106,1105-36.
- Tiwari, A. and Singh, V. (2008). Microwave induced synthesis of gum-acacia graft-polyaniline. *Carbohydrate Polymer*, 74, 427-34.
- Trivedi, T. J., Rao, K. S. and Kumar, A. (2014). Facile preparation of agarose-chitosan hybrid materials and nanocomposite ionogels using an ionic liquid via dissolution, regeneration and sol-gel transition. *Green Chemistry*, 16, 320-330.
- Tsuzuki, M., Hagiwara, I., Shiraishi, N., and Yokota, T. (1980). Characterization of products prepared by homogeneous grafting of styrene onto cellulose in a sulfur dioxide-diethylamine-dimethyl sulfoxide medium. *Journal of Applied Polymer Science*, 25, 2909-2924.
- Tunali, S. and Akar, T. (2006). Zn(II) Biosorption Properties of Botrytis Cinerea Biomass. *Journal of hazardous materials*. 131(1-3), 137-145.
- Tuncel, D. (2011). Non-covalent interactions between carbon nanotubes and conjugated polymers. *Nanoscale*, 3, 3545-3554.
- Turiel, E., and Martín-Esteban, A. (2010). Molecularly Imprinted Polymers for Sample Preparation: A review. *Analytical Chemistry Acta*, 668, 87-99.
- Tuzan, M., Saygi, K. O. and Soylak, M. (2008). Solid phase extraction of heavy metal ions in environmental samples on multiwalled carbon nanotubes. *Journal of Hazardous Materials*. 152, 632-639.
- Tuzen, M., Saygi, K. M. and Soylak, M. (2008). Novel solid phase extraction procedure for gold(III) on Dowex M 4195 prior to its flame atomic absorption spectrometric determination. *Journal of Hazardous Materials*. 156, 591-595.

- Velasco-Santos, C., Martínez-Hernández, A. L., Fisher, F. T., Ruoff, R., and Castaño, V. M. (2003). Improvement of Thermal and Mechanical Properties of Carbon Nanotube Composites through Chemical Functionalization. *Chemistry of Materials*, 15, 4470-4475.
- Vellaichamy, S. and Palanivelu, K. (2011). Preconcentration and separation of copper, nickel and zinc in aqueous samples by flame atomic absorption spectrometry after column solid-phase extraction onto MWCNTs impregnated with D2EHPA-TOPO mixture. *Journal of Hazardous Materials*, 185, 1131-1139.
- Vijan, V., Kaity, S., Biswas, S., Isaac, J. and Ghosh, A. (2012). Microwave assisted synthesis and characterization of acrylamide grafted gellan, application in drug delivery. *Carbohydrate Polymers*, 90(1), 496-506.
- Wan Ibrahim, W. A., Wan Ismail, W. N., Abdul Keyon, A. S. and Sanagi, M. M. (2011b). Synthesis and Characterization of a New Sol-Gel Hybrid Based on Tetraethoxysilane-Polydimethylsiloxane as a Stir Bar Extraction Sorbent Materials. *Journal of Sol-Gel Science and Technology*, 58, 602-611.
- Wan Ibrahim, W.A., Abdul Keyon, A.S., Prastomo, N. and Matsuda, A. (2011a). Synthesis and Characterization of Polydimethylsiloxane-Cyanopropyltriethoxysilane-Derived Hybrid Coating for Stir Bar Sorptive Extraction. *Journal of Sol-Gel Science and Technology*, 59, 128-134.
- Wang, W. B. and Wang, A. Q. (2010). Preparation, swelling and water retention properties of crosslinked supersorbent hydrogels based on guar gum. *Advance Materials Research*, 96, 177-82.
- Wu, W., Zhang, S., Li, Y., Li, J., Liu, L., Qin, Y. and et al. (2003). PVK-modified singlewalled carbon nanotubes with effective photoinduced electron transfer. *Macromolecules*, 36, 6286-8.
- Yang, B., Gong, Q., Zhao, L., Sun, H., Ren, N., Qin, J., Xu, J. and Yang, H. (2011). Preconcentration and determination of lead and cadmium in water samples with a MnO₂ coated carbon nanotubes by using ETAAS. *Desalination*. 278. 65-69.
- Yang, K., and Xing, B. (2007). Desorption of Polycyclic Aromatic Hydrocarbons from Carbon Nanomaterials in Water. *Environmental Pollution*, 145, 529-537.

- Yang, L., Zhang, X., Ye, M., Jiang, J., Yang, R., Fu, T., Chen, Y., Wang, K., Liu, C. and Tan, W. (2011). Aptamer-conjugated nanomaterials and their applications. *Advanced Drug Delivery Reviews*, 63, 1361-1370.
- Yang, M., Gao, Y., Li, H. and Adronov, A. (2007). Functionalization of multiwalled carbon nanotubes with polyamide 6 by anionic ring-opening polymerization. *Carbon*, 45, 2327-33.
- Yao, Z., Braidy, N., Botton, G. A. and Adronov, A. (2003). Polymerization from the surface of single-walled carbon nanotubes-preparation and characterization of nanocomposites. *Journal of the American Chemical Society*, 125, 16015-24.
- Ying, L. S., Salleh, M. A. b.M., Yusoff, H. b. M., Rashid, S. B. A. and Razak, J. b. A. (2011). Continuous production of carbon nanotubes– a review. *Journal of Industrial and Engineering Chemistry*, 17, 367-376.
- Zhang, N. and Hu, B. (2012). Cadmium (II) imprinted 3-mercaptopropyltrimethoxysilane coated stir bar for selective extraction of trace cadmium from environmental water samples followed by inductively coupled plasma mass spectrometry detection. *Analytical Chemistry Acta*, 723, 54-60.
- Zhang, N., Peng, H. and Hu, B. (2012). Light-induced pH change and its application to solid phase extraction of trace heavy metals by high-magnetization Fe₃O₄@SiO₂@TiO₂ nanoparticles followed by inductively coupled plasma mass. *Talanta*, 94, 278-283.
- Zhang, W. D., Shen, L., Phang, I. Y. and Liu, T. (2004). Carbon nanotubes reinforced nylon-6 composite prepared by simple melt-compounding. *Macromolecules*, 37, 256-9.
- Zhang, X., Sreekumar, T. V., Liu, T. and Kumar, S. (2004). Properties and structure of nitric acid oxidized single wall carbon nanotube films. *Journal of Physical Chemistry B.*, 108, 16435-40.
- Zhao, B., Hu, H., Yu, A., Perea, D. and Haddon, R. C. (2005). Synthesis and characterization of water soluble single-walled carbon nanotube graft copolymers. *Journal of the American Chemical Society*, 127, 8197-203.
- Zhao, BX., Wang, P., Zheng, T., Chen, Cy. and Shu, J. (2006). Preparation and adsorption performance of a cellulosic-adsorbent resin for copper(II). *Journal of Applied Polymer Science*, 99, 2951–6.

- Zhao, J. J., Buldum, A., Han, J., and Lu, J. P. (2002). Gas Molecule Adsorption in Carbon Nanotubes and Nanotube Bundles. *Nanotechnology*, 13, 195-200.
- Zhao, P., Wang, L., Zhou, L., Zhang, F., Kang, S. and Pan, C. (2012). Multi-walled carbon nanotubes as alternative reversed-dispersive solid phase extraction materials in pesticide multi-residue analysis with QuEChERS method. *Journal of Chromatography A*, 1225, 17-25.
- Zhao, X. W., Song, N. Z. and Jia, Q. O. (2009). Determination of Cu, Zn, Mn and Pb by microcolumn packed with multiwalled carbon nanotubes on-line coupled with flame atomic absorption spectrometry. *Microchimica Acta*, 166, 329-335.
- Zheng, H., Chang, X., Lian, N., Wang, S., Cui, Y. and Zhai, Y. (2006). A pre-enrichment procedure using diethyldithiocarbamate-modified TiO₂ nanoparticles for the analysis of biological and natural water samples by ICP-AES. *International Journal of Environmental Analytical Chemistry*, 86, 431-441.
- Zheng, M. M., Ruan, G. D. and Feng, Y. Q. (2009). Hybrid organic-inorganic silica monolith with hydrophobic/strong cation-exchange functional groups as a sorbent for micro-solid phase extraction. *Journal of Chromatography A*, 1216(45), 7739-7746.
- Zheng, M., Jagota, A., Semke, E. D., Diner, B. A., McLean, R. S., Lustig, S. R., Richardson, R. E. and Tassi, N. G. (2003). DNA-assisted dispersion and separation of carbon nanotubes. *Natural Materials*, 2, 338-342.
- Zhou, Q. X., Wang, W. D. and Xiao, J. P. (2006). Preconcentration and determination of nicosulfuron, thifensulfuron-methyl and metsulfuron-methyl in water samples using carbon nanotubes packed cartridge in combination with high performance liquid chromatography. *Analytical Chemistry Acta*, 559, 200-206.
- Zhou, Q. X., Xiao, J. P. and Wang, W. D. (2006). Using multi-walled carbon nanotubes as solid phase extraction adsorbents to determine dichlorodiphenyltrichloroethane and its metabolites at trace level in water samples by high performance liquid chromatography with UV detection. *Journal of Chromatography A*, 1125, 152-158.
- Zorbas, V., Smith, A. L., Xie, H., Ortiz-Acevedo, A., Dalton, A. B., Dieckmann, G. R., Draper, R. K., Baughman, R. H. and Musselman, I. H. (2005). Importance