

SYNTHESIS AND CHARACTERIZATION OF SPIONS-BROMELAIN-FOLIC
ACID ON FOLIC ACID RECEPTOR POSITIVE CANCER MODEL

ROZITA NASIRI

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (Bioprocess Engineering)

Faculty of Chemical and Energy Engineering
Universiti Teknologi Malaysia

JUNE 2016

Dedicated to

*My ever-supportive Mama (Ashraf Razavi), Papa (Sohrab Nasiri), my sisters, my
brothers and my colleague (Javad Hamzahalipour Almaki).*

*Specially dedicated to my beloved mother (Ashraf Razavi) and sisters (Setareh and
Mahtab) who are everything in my life.*

Thank you for being the best thing that ever happened in my life.

Love you all by very fabric of my being.

ACKNOWLEDGEMENT

I would like to express my sincere and utmost gratitude to my main supervisor Prof. Dr. Ani Binti Idris for her supervision, guidance and great support throughout the duration that I undertook to complete this project successfully. My sincere appreciation also extends to my co-supervisor Prof. Dr. Fadzilah Adibah Bin Abdul Majid for her kindness and guidance throughout the entire research. Last but not least, I would like to express my utmost appreciation to my lovely family who have given me all that I have. Indeed they are the best in my life.

ABSTRACT

Engineering of a physiologically compatible, stable and targetable delivery vehicle superparamagnetic iron oxide nanoparticles-Bromelain-folic acid (SPIONs-Br-FA) was reported. Initially, the synthesized bare SPIONs were coated with citric acid (CA) in order to increase biocompatibility, stability and solubility of the SPIONs. Moreover, through CA coating, carboxyl functional groups for further reactions were produced. Br (as an anti-cancer agent) and FA (as a targeting agent to the folic acid receptor positive (FAR+) cancer cells) were conjugated to the synthesized nanocarrier through 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride/ N-hydroxysuccinimide (EDC/NHS) click chemistry. Subsequently, characterization and physico-chemical analyses were carried out through methods such as Fourier transform infrared spectroscopy, atomic absorption spectroscopy (AAS), dynamic light scattering, vibrating sample magnetometer, x-ray diffraction, transmission electron microscopy (TEM) and field emission scanning electron microscopy. The *in vitro* tetrazolium dye (MTT) assay and blood compatibility tests were performed to confirm the biocompatibility of the engineered nano delivery system. High level of SPIONs-FA binding to FAR+ cell lines (HeLa, MDA-MB-231 and 4 T1) compared to folic acid receptor negative (FAR-) cell lines (HSF 1184 and MDA-MB-468) was assured via qualitative and quantitative *in vitro* binding studies (Prussian blue assay and AAS analysis). The reason may be higher transport of SPIONs-FA through the mechanism of receptor endocytosis pathway into FAR+ cells in comparison with the mechanism of passive diffusion of SPIONs into the FAR- cells. Cytotoxicity studies carried out in human cell lines (HSF 1184, MDA-MB-468, MDA-MB-231 and HeLa) and mouse breast cancer cells (4 T1) showed significant dose advantage with SPIONs-Br-FA in reducing the half maximal inhibitory concentration (IC₅₀) values compared with neat Br. Through morphological observation studies by inverted microscope and acridine orange/ethidium bromide fluorescent staining method, it was disclosed that the cells had undergone apoptosis since the shrinkage as well as the apoptotic bodies were obviously seen. The results showed that SPIONs-Br-FA was a rewarding candidate to suppress the migration of the FAR+ cancer cells as well as to inhibit colony formation of the FAR+ cancer cells compared to neat Br. The percentage of apoptotic cells (apoptotic index) with more condensed and fragmented chromatin increased sharply in SPIONs-Br-FA treated cells compared to the neat Br. Overall, the SPIONs-Br-FA induced higher percentage of apoptotic cells than the neat Br. Moreover, after treatment protocol performance on 4 T1 tumor bearing mice, the qualitative and quantitative biodistribution study were carried out in vital organs and tumor using colorimetric method (AAS) and TEM method which indicate significant tumor targetability of SPIONs-FA. Finally, the tumor volume and inhibition growth rate were measured in 4 T1 tumor bearing mice treated with different SPIONs formulations to investigate the effectiveness of SPIONs-Br-FA. Administration of SPIONs-Br-FA through tail vein (three times a week) during the four-week treatment period reduced the tumor burden of tumor bearing mice and also increased their life-span when compared with SPIONs-Br and neat Br at same concentration of bromelain. In conclusion, the current results indicated the dual-functional synthesized SPIONs-Br-FA is a promising tool in the field of biomedicine, chiefly cancer therapy.

ABSTRAK

Kejuruteraan yang serasi secara fisiologi, stabil dan boleh menyasarkan sarana pemasukan partikel nano ferum oksida super paramagnet-*Bromelain*-asid folik (SPIONs-Br-FA) telah dilaporkan. SPIONs telah disaluti dengan asid sitrik (CA) untuk meningkatkan bioserasian, kestabilan dan keterlarutan SPIONs tersebut. Selain itu, melalui penyalutan CA, kumpulan-kumpulan karboksil berfungsi untuk tindak balas lanjutan telah dihasilkan. Br (sebagai agen anti-kanser) dan FA (sebagai agen penyasaran terhadap sel-sel kanser positif reseptor asid folik (FAR+)) telah dikongjugasi pada pembawa-nano yang disintesis melalui kimia klik 1-etil-3-(3-dimetilaminopropil)karbodiimid hidroklorida/*N*-hidriksisusinimida (*EDC/NHS*). Seterusnya, pencirian dan analisis kimia-fizik telah dijalankan melalui kaedah tertentu seperti spektroskopi inframerah transformasi Fourier, spektroskopi penyerapan atom (AAS), penyerakan cahaya dinamik, magnetometer sampel bergetar, pembelauan sinar-x, mikroskopi pancaran elektron (TEM) dan mikroskop elektron pengimbas pancaran medan. Asai *in vitro* pewarna tetrazolium (MTT) dan ujian serasian darah telah dijalankan untuk mengesahkan bioserasian sistem pemasukan nano yang dibina. Tahap pengikatan SPIONs-FA yang tinggi terhadap titisan-titisan sel FAR+ (He La, MDA-MB-231 dan 4 T1) berbanding dengan titisan-titisan sel negatif reseptor asid folik (FAR-) (HSF 1184 dan MDA-MB-468) telah dikenal pasti melalui kajian pengikatan *in vitro* kualitatif dan kuantitatif (asai biru Prusia dan analisis AAS). Ini mungkin disebabkan oleh pengangkutan yang lebih tinggi SPIONs-FA menerusi mekanisme tapak jalan endositosis reseptor ke dalam sel-sel FAR+ berbanding dengan mekanisme peresapan pasif SPIONs ke dalam sel-sel FAR-. Kajian kesitoksikan yang dijalankan pada titisan-titisan sel manusia (HSF 1184, MDA-MB-468, MDA-MB-231 dan HeLa) dan sel-sel kanser payudara mencit (4 T1) telah menunjukkan kelebihan dos yang signifikan dengan SPIONs-Br-FA dalam mengurangkan nilai-nilai separuh maksimum kepekatan yang melarang (IC_{50}) berbanding dengan Br tulen. Melalui kajian pemerhatian morfologi oleh mikroskop songsang dan kaedah pewarnaan pendarfluor akridina jingga/etidium bromida, di dapati bahawa sel-sel telah menjalani apoptosis kerana pengecutan sel dan jasad-jasad apoptosis jelas kelihatan. Hasil kajian menunjukkan bahawa SPIONs-Br-FA merupakan sel yang sesuai untuk menyekat migrasi sel-sel kanser FAR+ serta merencat pembentukan koloni sel-sel kanser FAR+ berbanding dengan Br tulen. Peratusan sel apoptosis (indeks apoptosis) dengan kromatin mampat dan tersepih meningkat secara mendadak dalam sel-sel terawat SPIONs-Br-FA berbanding dengan Br tulen. Secara keseluruhan, SPIONs-Br-FA mengaruh peratusan sel-sel apoptosis yang lebih tinggi berbanding dengan Br tulen. Malahan, selepas protokol rawatan pada mencit terkandung sel-sel kanser 4 T1, kajian bio-pengedaran kualitatif dan kuantitatif telah dijalankan pada organ-organ penting dan tumor menggunakan kaedah kolorimetri (AAS) dan TEM yang menunjukkan kebolehan penyasaran tumor yang signifikan oleh SPIONs-FA. Akhir sekali, isipadu tumor dan kadar perencatan tumbesaran diukur pada mencit terkandung sel-sel kanser 4 T1 setelah dirawat dengan pelbagai formulasi SPIONs untuk menyelidik keberkesanan SPIONs-Br-FA. Pemberian SPIONs-Br-FA melalui vena ekor (tiga kali seminggu) semasa tempoh rawatan empat minggu telah mengurangkan bebanan tumor pada mencit dan juga meningkatkan jangka hayat mereka semasa perbandingan SPIONs-Br dengan Br tulen pada kepekatan bromelain yang sama. Sebagai kesimpulan, hasil kajian semasa menunjukkan bahawa SPIONs-Br-FA berdwi-fungsi yang disintesis merupakan alat berpotensi dalam bidang bioperubatan, terutamanya terapi kanser.

TABLE OF CONTENTS

| CHAPTER | TITLE | PAGE |
|----------|---|-------|
| | DECLARATION | ii |
| | DEDICATION | iii |
| | ACKNOWLEDGEMENT | iv |
| | ABSTRACT | v |
| | ABSTRAK | vi |
| | TABLE OF CONTENTS | vii |
| | LIST OF TABLES | xi |
| | LIST OF FIGURES | xii |
| | LIST OF ABBREVIATIONS | xvi |
| | LIST OF APPENDICES | xviii |
| | | |
| 1 | INTRODUCTION | 1 |
| | 1.1 Background of Study | 1 |
| | 1.2 Problem Statement | 7 |
| | 1.3 Research Objectives | 8 |
| | 1.4 Scope of Research | 8 |
| | 1.5 Significance of Study | 9 |
| | 1.6 Thesis Organization | 10 |
| | | |
| 2 | LITERATURE REVIEW | 12 |
| | 2.1 Introduction | 12 |
| | 2.2 Overview on Cancer | 13 |
| | 2.3 Barriers to Conventional Cancer Treatment | 17 |
| | 2.4 Nanotechnology | 21 |
| | 2.5 Tumor Vasculature | 23 |

| | | |
|----------|--|-----------|
| 2.6 | Enhanced Permeability and Retention (EPR) effect | 25 |
| 2.7 | Nano Delivery System | 26 |
| 2.7.1 | Types of Nano Delivery System | 28 |
| 2.8 | Magnetic Nano platforms as Drug Carriers | 31 |
| 2.8.1 | Superparamagnetic Nanoparticles (SPIONs) as Drug Carriers | 33 |
| 2.9 | Characterization of SPIONs in Drug Delivery Applications | 34 |
| 2.10 | Biocompatibility Evaluation of SPIONs | 34 |
| 2.11 | Bromelain (Br) | 36 |
| 2.12 | Cell Death, Apoptosis and Necrosis | 38 |
| 2.13 | Concepts of Passive and Active Targeting | 41 |
| 2.14 | Receptor Mediated Cell Uptake | 45 |
| 2.15 | Active Targeting by Folic Acid (FA) | 45 |
| 3 | RESEARCH METHODOLOGY | 48 |
| 3.1 | Introduction | 48 |
| 3.2 | Materials | 48 |
| 3.3 | Experimental Design | 50 |
| 3.4 | Initial Ferrofluids (γ -Fe ₂ O ₃) Synthesis | 50 |
| 3.5 | Citrate Coating of SPIONs | 50 |
| 3.6 | Bromelain Conjugation | 52 |
| 3.7 | Determination of Bromelain Loading Content | 52 |
| 3.8 | Folic Acid Conjugation | 52 |
| 3.9 | Determination of Folic Acid Loading Content using HPLC | 53 |
| 3.10 | Structure Analysis | 54 |
| 3.11 | Determination of Hydrodynamic Diameter | 54 |
| 3.12 | Iron Concentration of Samples | 54 |
| 3.13 | Assessment of Magnetic Properties | 55 |
| 3.14 | Crystallinity Structure | 55 |
| 3.15 | Assessing the Size of Samples | 55 |
| 3.16 | Cell Line | 56 |

| | | |
|----------|--|-----------|
| 3.17 | <i>In vitro</i> RBC, WBC and PRP aggregation and Haemolysis Studies | 56 |
| 3.18 | Coagulation Factors Assay (PT, TT, APTT and FB) and Hard Clotting Test | 57 |
| 3.19 | Cell Compatibility Study | 58 |
| 3.20 | Quantitative Binding Study | 59 |
| 3.21 | Qualitative Binding Study | 60 |
| 3.22 | Morphological Assessment by Phase Contrast Inverted Microscopy | 60 |
| 3.23 | Apoptosis Detection by AO/EB Staining | 60 |
| 3.24 | Scratch Motility Assay | 61 |
| 3.25 | Clonogenic Inhibition Assay | 62 |
| 3.26 | Animal Handling and Establish Tumor Bearing Mice | 62 |
| 3.27 | Quantitative Biodistribution Study in Tumor Bearing Mice | 63 |
| 3.28 | Qualitative Biodistribution Study in Tumor Bearing Mice | 64 |
| 3.29 | <i>In Vivo</i> Anti-Cancer Efficacy | 64 |
| 3.30 | Statistics Analysis | 66 |
| 4 | RESULTS AND DISCUSSION | 67 |
| 4.1 | Introduction | 67 |
| 4.2 | Synthesis and Characterization | 67 |
| 4.3 | Binding Study | 80 |
| 4.4 | <i>In Vitro</i> Coagulation Factors and Hard Clotting Time Assay | 85 |
| 4.5 | Anti-Proliferation and Cytotoxicity Assay | 89 |
| 4.6 | Morphological Assessment by Phase Contrast Inverted Microscopy | 93 |
| 4.7 | Apoptosis Detection by AO/EB Staining | 95 |
| 4.8 | Scratch Motility Assay | 98 |
| 4.9 | Clonogenic Inhibition Assay | 104 |
| 4.10 | Establish Mice Bearing 4 T1 Tumor Xenografts | 105 |

| | | |
|----------|---|---------|
| 4.11 | Biodistribution Study of Tumor Bearing Mice | 106 |
| 4.12 | <i>In Vivo</i> Anti-Cancer Efficacy | 110 |
| 5 | CONCLUSION AND RECOMMENDATIONS | 115 |
| 5.1 | Conclusion | 115 |
| 5.2 | Suggestions for future work | 116 |
| | REFERENCES | 118 |
| | Appendices A-C | 153-155 |

LIST OF TABLES

| TABLE NO. | TITLE | PAGE |
|------------------|--|-------------|
| 2.1 | List of nano delivery systems. | 28 |
| 2.2 | Summary of some of the <i>in vitro</i> and <i>in vivo</i> studies on effects of bromelain on different cell lines and animal models. | 38 |
| 2.3 | Examples of targets for different types of cancer. | 41 |
| 3.1 | List of materials used in the research. | 45 |
| 3.2 | First group of <i>in vivo</i> study. | 62 |
| 3.3 | Second group of <i>in vivo</i> study. | 62 |
| 4.1 | Percentage of haemolysis of bare and modified SPIONs at three different concentrations. | 82 |
| 4.2 | Inhibitory effect (IC ₅₀ values) of SPIONs in different formulations, neat Br and Cisplatin against cells after 24 h. | 89 |

LIST OF FIGURES

| FIGURE NO. | TITLE | PAGE |
|------------|---|------|
| 1.1 | Ten most frequent cancers, all residence, Malaysia 2007 | 2 |
| 1.2 | Ten most frequent cancers, female, Malaysia 2007 | 2 |
| 1.3 | Nano drug vs. classic drug distribution in body. | 47 |
| 2.1 | Difference between normal and cancer cells. | 15 |
| 2.2 | Most common cancer sites worldwide by sex 2012. | 16 |
| 2.3 | The side effects of chemotherapy on the body. | 19 |
| 2.4 | Side effects of surgery, chemotherapy and radiation therapy. | 20 |
| 2.5 | The existence of gaps in the cancer cells' chaotic vasculature architecture allowing the unspecific macromolecules and nano-scale materials transport into tumor tissue. | 24 |
| 2.6 | EPR Effect a) in normal cells b) in tumor cells. | 26 |
| 2.7 | Schematic representation of the steps and mechanisms of apoptosis versus necrosis. | 40 |
| 2.8 | Schematic diagram of the folate receptor-mediated endocytosis pathway. | 47 |
| 3.1 | Research methodology flowchart. (1) Engineering, (2) Assessment. | 51 |
| 4.1 | Schematic presentation of SPIONs-Br-FA binding to the FA receptors on the cancer cells and administration of nanoparticles with different formulation into 4 T1 breast tumor-bearing mice leading to inhibition of cancer growth. | 68 |
| 4.2 | Step by step functionalization of dual-functional SPIONs-Br-FA. | 70 |
| 4.3 | FT-IR spectra of synthesized and functionalized nanoparticles including (a) bare SPIONs, (b) SPIONs, (c) SPIONs-Br and (d) SPIONs-Br-FA. | 71 |

| | | |
|------|---|----|
| 4.4 | The stability characteristics of bare SPIONs, SPIONs-Br and SPIONs-Br-FA in terms of mean hydrodynamic diameter (nm). | 73 |
| 4.5 | Microscopic observation of the SPIONs-Br-FA, a) in the absence of the magnetic field and b) in the presence of the magnetic field. | 75 |
| 4.6 | Magnetization curves of synthesized nanoparticles. | 76 |
| 4.7 | XRD patterns of non-coated and coated SPIONs. | 77 |
| 4.8 | a) TEM bare SPIONs, b) TEM functionalized SPIONs, c) HRTEM bare SPIONs, d) HRTEM functionalized SPIONs, e) SAED patterns of bare SPIONs, f) intensity histogram of bare SPIONs | 79 |
| 4.9 | a) FESEM image of bare SPIONs, b) FESEM image of functionalized SPIONs, c) 3D image of bare SPIONs, d) 3D image of functionalized SPIONs, e) Size distribution of bare SPIONs, f) Size distribution of functionalized SPIONs. | 80 |
| 4.10 | Quantitative binding study. | 82 |
| 4.11 | Competitive binding assay. | 83 |
| 4.12 | Prussian blue stained images. | 84 |
| 4.13 | Blood aggregation studies. | 87 |
| 4.14 | Coagulation factors assay and hard clotting time test after blood incubation with different concentrations of nanoparticles. | 88 |
| 4.15 | MTT assay. | 90 |
| 4.16 | Growth inhibitory effects. | 94 |
| 4.17 | Detection of apoptosis by AO/EB staining. | 96 |
| 4.18 | Percentages of live, apoptotic, and necrotic cells at different cell lines treated with neat Br and SPIONs-Br-FA. | 97 |
| 4.19 | Inhibition of HSF 1184 cell migration after treatment with neat Br and SPIONs-Br-FA. | 98 |
| 4.20 | Inhibition of MDA-MB-468 cell migration after treatment with neat Br and SPIONs-Br-FA. | 99 |
| 4.21 | Inhibition of HeLa cell migration after treatment with neat Br and SPIONs-Br-FA. | 99 |

| | | |
|------|--|-----|
| 4.22 | Inhibition of MDA-MB-231 cell migration after treatment with neat Br and SPIONs-Br-FA. | 100 |
| 4.23 | Inhibition of 4 T1 cell migration after treatment with neat Br and SPIONs-Br-FA. | 100 |
| 4.24 | Quantitative analysis of migration inhibition rate of neat Br and SPIONs-Br-FA. | 102 |
| 4.25 | Qualitative analysis of colony forming inhibition potential of neat Br and-Br-FA. | 103 |
| 4.26 | Quantitative analysis of colony forming inhibition potential. | 105 |
| 4.27 | Image of mice bearing 4 T1 breast tumor in situ and a histological slice of tumor. | 106 |
| 4.28 | Quantitative biodistribution study. | 108 |
| 4.29 | Qualitative biodistribution study (TEM staining) of vital organs. | 109 |
| 4.30 | Tumor volume after <i>in vivo</i> treatment (Group 1). | 112 |
| 4.31 | Survival rate after <i>in vivo</i> treatment (Group 1). | 112 |
| 4.32 | Tumor volume after <i>in vivo</i> treatment (Group 2). | 113 |
| 4.33 | Extracted tumor volume after <i>in vivo</i> treatment (Group 2). | 113 |

LIST OF ABBREVIATIONS

| | | |
|-------------------|---|---|
| AAS | - | Atomic Absorption Spectroscopy |
| APTT | - | Activated partial thromboplastin time |
| AR | - | Androgen receptor |
| Br | - | Bromelain |
| CA | - | Trisodium citrate dihydrate |
| CI | - | Confidence interval |
| DLS | - | Dynamic Light Scattering |
| EGFR | - | Epidermal growth factor receptor |
| EPR | - | Enhanced permeability and retention |
| FA | - | Folic acid |
| FAR | - | Folic acid receptor |
| FAR- | - | Negative folic acid receptor |
| FAR+ | - | Positive folic acid receptor |
| FB | - | Fibrin formation |
| FESEM | - | Field emission scanning electron microscopy |
| FR α (FAR) | - | Folic acid receptors |
| FT-IR | - | Fourier transform Infrared |
| FWHM | - | Full width at half maximum |
| HB | - | Hemoglobin |
| HCT | - | Hematocrit |
| HCT | - | Hard clotting time |
| HER2 | - | Human epidermal growth factor receptor 2 |
| HER3 | - | Human epidermal growth factor receptor 3 |
| HPLC | - | High performance liquid chromatography |

| | | |
|--------------|---|--|
| ID | - | Injected dose |
| IGF-IR | - | Insulin-like growth factor receptor |
| MCHC | - | Mean corpuscular hemoglobin concentration |
| MCV | - | Average red blood cell size |
| MRI | - | Magnetic resonance imaging |
| MTT | - | Thiazolyl Blue Tetrazolium Bromide |
| NSCLC | - | Non-Small Cell Lung Cancer |
| PAMAM | - | Polyamidoamine |
| PARP | - | Poly(ADP-ribose) polymerase |
| PCV | - | Packed cell volume |
| PEG | - | Polyethylene glycol |
| PEI | - | Polyethylenimine |
| PRP | - | Platelets |
| PSMA | - | Prostate specific membrane antigen |
| PT | - | Prothrombin time |
| RBC | - | Red blood cells |
| RES | - | Reticuloendothelial system |
| Bare SPIONs | - | Superparamagnetic Iron oxide nanoparticles |
| SPIONs-Br | - | Bromelain conjugated citrate SPIONs |
| SPIONs-Br-FA | - | Bromelain and folate conjugated citrate SPIONs |
| SPIONs | - | Iron oxide nanoparticles coated with CA |
| SPIONs-FA | - | Folate conjugated citrate SPIONs |
| TEM | - | Transmission Electron Microscopy |
| TT | - | Thrombin time |
| VEGF-A | - | Vascular endothelial growth factor A |
| VEGFR | - | Vascular endothelial growth factor |
| VSM | - | Vibrating sample magnetometer |
| WBC | - | White blood cells |
| XRD | - | X-ray Diffraction |

LIST OF APPENDICES

| APPENDIX | TITLE | PAGE |
|-----------------|----------------------|-------------|
| A | List of Publications | 153 |
| B | HPLC Chromatogram | 154 |
| C | Ethical Endorsement | 155 |

CHAPTER 1

INTRODUCTION

1.1 Background of study

One of the most life threatening diseases is cancer where the number of new cases is growing increasingly (Boyle and Levin, 2008). According to the last report, breast cancer was the most common cancer in females and also the first most common cancer among population regardless of sex in Malaysia. There were 3,242 female breast cancer cases diagnosed and reported to NCR (National Cancer Registry) in 2007 which accounted for 18.1% of all cancer cases reported and 32.1% are all female cases. The age pattern in 2007 showed a peak ASR (age-standardised rate) at the 50-59 age groups. The incidence of breast cancer was highest among Chinese where the ASR was 38.1 per 100,000 population followed by Indian and Malay with the ASR of 33.7 per 100,000 population and 25.4 per 100,000 populations, respectively. The percentage of breast cancer detected at stage I and II was 58%. Cancer of the cervix was the third most common cancer among women and fifth most common cancer in the entire general population. There were a total of 847 cases diagnosed in 2007 registered at NCR. Cervical cancer incidence rate increased after 30 years old and peaks at ages 65-69 years. Compared among the major races, Indian women had the highest incidence for cervical cancer followed by Chinese and Malay. The ASR for Indian females was 10.3 per 100,000 populations. The percentage of breast cancer detected at stage I and II was 55% (Omar and Tamin, 2011). Figure 1.1 shows ten most frequent cancers in all residence and Figure 1.2 presents ten most frequent cancers among female in Malaysia in 2007.

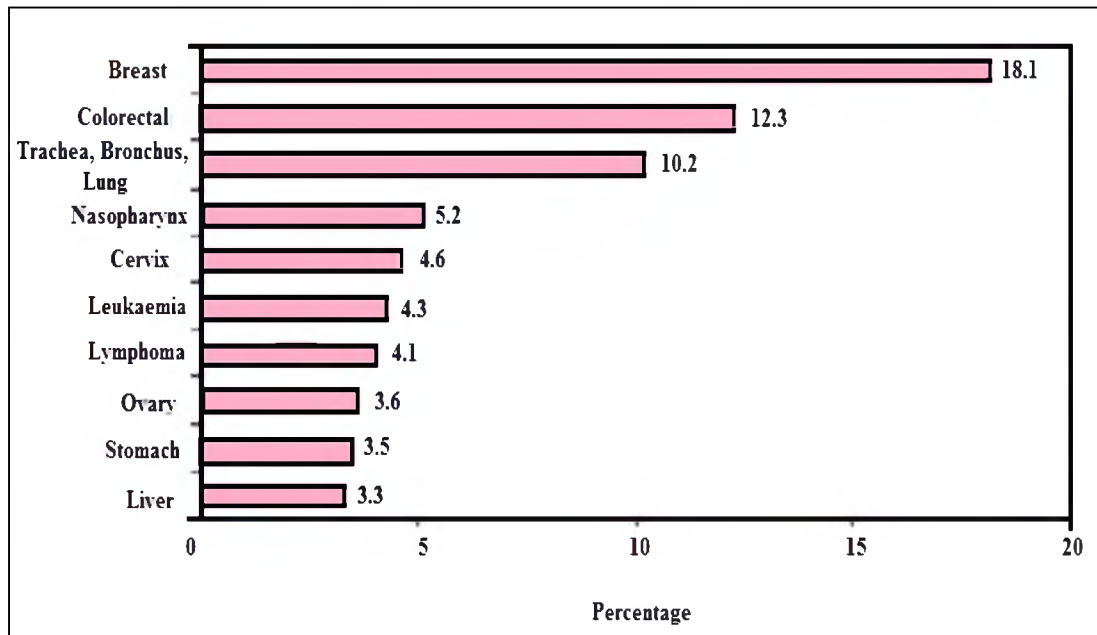


Figure 1.1 Ten most frequent cancers, all residence, Malaysia 2007 (Omar and Tamin, 2011).

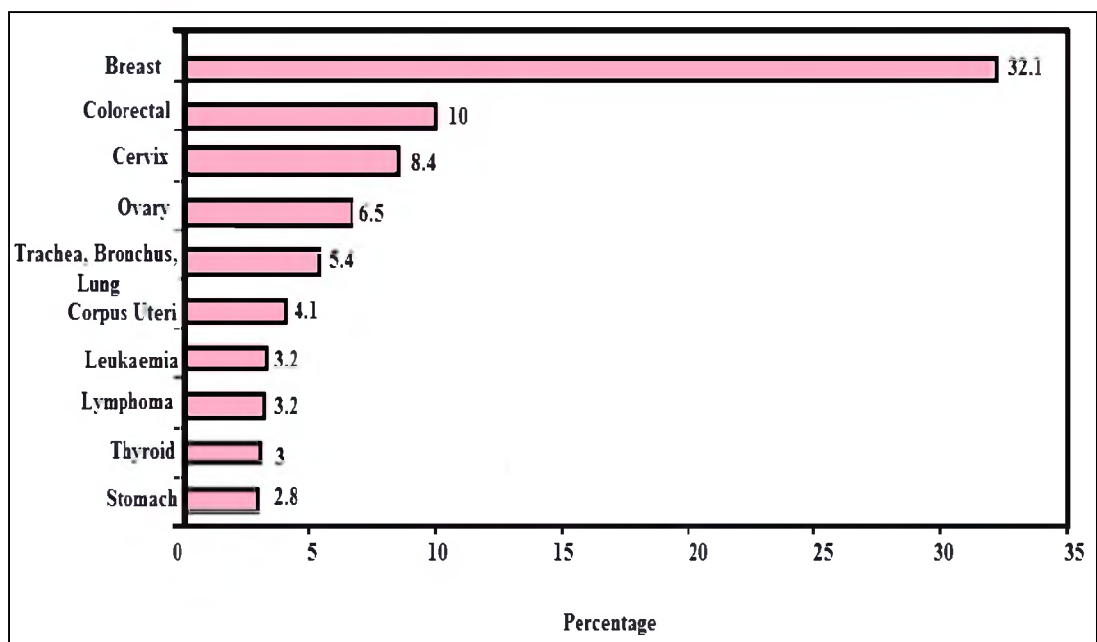


Figure 1.2 Ten most frequent cancers, female, Malaysia 2007 (Omar and Tamin, 2011).

In spite of the accelerated progress of diagnostics and treatments, substantial

improvements over the survival rate of patients have not yet been seen over the course of past few decades (Jemal *et al.*, 2010). Developing a novel approach to incur the detection of cancer in its early-stages as well as developing targetable therapies is a remarkable need.

Chemotherapy is the most practiced cancer treatment method in the world over the years. Nonspecific conventional chemotherapy normally leads to extreme side effects and is compromised because of its dose-limiting toxicity. Nanomaterials advances have made passive and active targeting strategies possible to boost up concentration of drugs inside tumor. Moreover, limiting the unwanted drug toxicity to healthy tissue is whereby achieved (Maeda, 2001; Allen, 2002; Torchilin, 2006). The targetable drug delivery is expected to eliminate troubles in conventional neat anticancer agents, including insolubility, accelerated clearance, unselective binding ability that leads to nonspecific toxicity towards healthy cells and decreases the drug dose delivered to the cancer cells (Ashley *et al.*, 2011). Since nano drug carriers offer longer half-lives in blood circulatory system compared to free drugs, they unfold a key potential to target the cancer cells. Increased amount of delivery to the cancer cells is highly dependent on the lowered total body clearance of the nano drug carriers. Additionally, due to the presence of poor lymphatic drainage and leaky blood vessels in the tumor site, retention and permeation of the nano drug carriers to the tumor site are highly enhanced. Since the conjugates find their way into the cancer cells through endocytosis, active drug molecules are released via either acid or intracellular enzymatic hydrolysis. Hence, drug internalization into the cancer cells is boosted up via raising the binding extent of conjugates to the cancer cells. This route, selective endocytosis, has been investigated by the attachment of targeting ligands to the nano carriers (Chau *et al.*, 2004).

Figure 1.3 illustrates the different localization of drug in targeted strategy by nano drugs compared with systemic treatment by classic drugs. For oral intake or intravenous injection of the classical drug, the bioactive component is distributed throughout the body without any distinctions between healthy and inflamed tissue. In targeting strategy, nano drugs are attached to the targeting agents whose ligands are overexpressed in interested areas. The nano drugs accumulate and the drug is

released in the specific area.

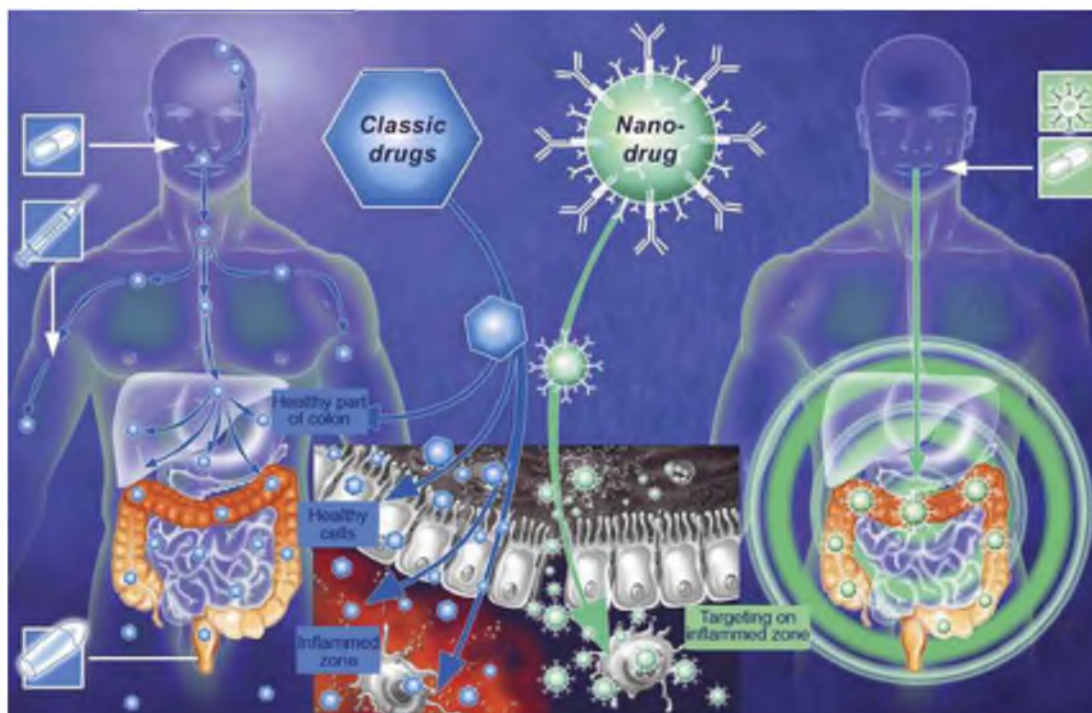


Figure 1.3 Nano drug vs. classic drug distribution in body (Laroui *et al.*, 2011).

Recent nanotechnological advances offer platforms to fabricate ultrasmall probes like superparamagnetic iron oxide nanoparticles (SPIONs). SPIONs are well known for their invaluable function in biomedical applications like magnetic resonance imaging (MRI), intracellular magnetic hyperthermia, targeted drug delivery, cell tracking and labelling, localized therapy, etc. (Laurent *et al.*, 2008; Fang and Zhang, 2009). SPIONs are in preclinical studies as well as in early stage clinical trials (Laurent *et al.*, 2008; McCarthy and Weissleder, 2008). A variety of methods have been reported to synthesize SPIONs like micro emulsion, sonochemical synthesis, thermal decomposition, hydrothermal synthesis and co-precipitation. (Woo *et al.*, 2004; Wu *et al.*, 2009). Co-precipitation is a neat and suitable method for synthesis of SPIONs smaller than 20 nm in diameter (Li *et al.*, 2013). Desired SPIONs for various biomedical applications are between 10 nm and 100 nm in diameter (Wahajuddin, 2012). Recent investigations have reflected the fact that SPIONs are highly favorable drug targeting platforms because of their rather poor toxic effects (Neuberger *et al.*, 2005) and high magnetic saturation magnitudes

(Bean and Livingston, 1959).

However, due to the hydrophobic nature of SPIONs, they are instable and prefer to aggregate in physiological condition (Jain *et al.*, 2005; Mahmoudi *et al.*, 2010). Moreover, the large surface area to volume ratio of SPIONs compels the tendency to aggregate thus limiting their naturally high level of surface energy (Vayssieres *et al.*, 1998). Therefore, organic or inorganic materials are used to coat the SPIONs surface to barricade agglomeration and ensure biocompatibility. Coating not only stabilizes the SPIONs, but also promotes the attachment of biological moieties to them; the particles are targeted to cells by attaching functional groups to the SPIONs. Citric acid ($C_6H_8O_7$) has been extensively used as a biocompatible and short-chained tri-carboxylic acid to stabilize SPIONs for different biomedical applications (Liu and Huang, 1999; Nigam *et al.*, 2011; Lapresta-Fernández *et al.*, 2011)

Since high level of targeting is not offered by SPIONs due to their physiochemical profiles, active biomolecules are attached to the surface of the SPIONs to heighten the targeting specificity of nanoparticles (Lee *et al.*, 2007; McCarthy and Weissleder, 2008; Goya *et al.*, 2008). Clinical utility of the SPIONs is significantly increased after being bonded to the contrast agents allowing the SPIONs to accumulate in the sites of interest (Artemov, 2003; Choi *et al.*, 2004; Leuschner *et al.*, 2006). Additionally, various studies have pointed out diverse approaches for active targeting of SPIONs by protein structures, nutrients and therapeutics. Internalization of structures attached to the SPIONs are inhibited due to the bulky and immunogenic nature of antibodies (Zhang *et al.*, 2002). Since nutrient pathways increase the uptake of SPIONs because of their direct linkage to cell proliferation process, most tumor types provide signals more excellently. Tumor cells are dependent to folic acid (FA) as it is one of the essential precursors in synthesis of DNA base (Weitman *et al.*, 1992; Garin-Chesa *et al.*, 1993; Ross *et al.*, 1994), , In normal cells, folate receptors are slightly expressed (Weitman *et al.*, 1992) and it assists the nanoparticles to conjugate with FA to be internalized to the cancer cells simultaneously expressing folate receptors (FAR+) through receptor-mediated endocytosis pathway due to high levels of penetration and affinity (Barz *et al.*, 2010).

In most of the studies (Müller *et al.*, 2008; Razjouyan *et al.*, 2015), mentioned, folic acid was used in combination with nanostructures other than citrate-coated maghemite. But, in this study, folic acid was conjugated to the SPIONs via the help of citric acid (CA) resulting in synthesis of a novel biomaterial with a monodisperse nature and desired characteristics offering targeting capabilities to track and attach to the FAR+ cancer cells while being highly blood compatible and remarkably reduced cytotoxicity.

Nowadays, the use of bromelain (Br) as an anticancer agent is fast becoming attractive. Several studies, both animal and human, indicate bromelain have antimetastatic activities (Pillai *et al.*, 2013). Bromelain due to its anti-inflammatory, mucolytic, antithrombotic, wound debridement and anticancer properties has undergone investigations as a cysteine proteinase extracted from pineapple (*Ananascomosus*). Bromelain also offers anti-tumorigenic properties so that it enhances chemotherapy effect in both *in vitro* and *in vivo* trials particularly in breast and pancreas cancers. Proteolytic component of Bromelain may be chiefly liable to its anti-tumor activity according to a recent review (Bala *et al.*, 2012). It is evident that glycosylated moieties providing cellular oncogenic survival pathways may be influenced since bromelain hydrolyses linkages of glycosides in glycoproteins. Moreover, there are merits to the disruption of the glycosidic linkages in the secreted mucin via proteolytic action of bromelain because it may disrupt the mucinous barrier and offers a more efficient passage for cytotoxic drugs (Pillai *et al.*, 2013).

In this research, to make the surface of the synthesized bare SPIONs (γ -Fe₂O₃) hydrophilic, functional groups for further surface functionalization were provided, nanoparticles agglomeration was prevented and absorption of CA onto the surface of nanoparticles was carried out leaving a carboxylic acid exposed on the surface. The final product was engineered by conjugation of bromelain (Br) and folic acid (FA) to the SPIONs (Citrate coated iron oxide nanoparticles). Briefly, in the study reported herein an attempt has been made to synthesize the SPIONs-Br-FA as a novel engineered delivery of bromelain to the FAR+ cancer cells.

1.2 Problem statement

Chemotherapy, mastectomy, and radiotherapy are conventional cancer treatment methods which are not completely successful and they induce many side effects on healthy tissues. Currently, immense numbers of different chemotherapeutic anticancer agents are available, but the problem is drugs that are more effective tend to be more toxic. For example, one of the most effective and widely used anticancer agent cisplatin, is reported to cause adverse effects including nausea, vomiting, diarrhea, hair loss, loss in ability to taste food, hiccups, dry mouth, dark urine, decreased sweating, dry skin, and other signs of dehydration which considerably limit its applicability (Santabarbara *et al.*, 2016). So, there is a need of a drug carrier system to minimize systemic side effects compared to chemotherapy by actively targeting the anticancer agent to the cancer cells.

Drugs used in classic chemotherapy are incapable of detecting cancer cells thus they influence both cancer cells as well as the healthy ones. Therefore patients tend to suffer from such classic conventional treatment. But, nano-drugs possess the capability to target the cancer cells actively since their surface can be functionalized via ligands that can specifically attach to the cancer cells.

On the other hand, metastasis treatment of tumors is unsuccessful by conventional methods. Metastasis is the secondary malignant growth at a distance from a primary site of cancer. Tumor itself can be treated by mastectomy or other treatment methods, but metastasis does not appear during the first stage of disease thus that it could not be detected and treated easily.

In the proposed study, folic acid will detect metastasis wherever it is and the complex will bind to the FAR+ cancer cell receptors to maximize the anticancer effect of bromelain on targeted cancer tissue (tumor site) and to minimize toxicity to the normal tissues (Wang *et al.*, 2011).

1.3 Research objectives

The objectives of study are as follows.

- I. To synthesize and characterize SPIONs-Br-FA.
- II. To evaluate *in vitro* and *in vivo* binding affinity of SPIONs-FA.
- III. To investigate *in vitro* and *in vivo* cancer inhibition efficacy of SPIONs-Br-FA and to compare it with the efficacy of neat Br.

1.4 Scope of research

In order to achieve the objectives, the scope of the study was as follows:

- i) The bare SPIONs ($\gamma\text{Fe}_2\text{O}_3$) were synthesized using co-precipitation method and coated with citric acid (CA). The prepared bare SPIONs and coated SPIONs were then analyzed using FT-IR. In addition, iron oxide concentration was determined by atomic absorption spectroscopy (AAS).
- ii) Then, bromelain (Br) as an anticancer agent and folic acid (FA) as a targeting agent were conjugated using 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC)/N-hydroxysuccinimide (NHS) click chemistry method to the coated SPIONs. The loading efficiencies of bromelain and folic acid were determined using Bradford assay and HPLC, respectively.

- iii) The functionalized SPIONs were characterized by FT-IR, AAS, VSM, DLS, XRD, FESEM and TEM equipment.
- iv) The biocompatibility of synthesized delivery system in each synthesis step was determined using MTT, haemolysis, blood aggregation and blood clotting time assays.
- v) Binding ability of the developed coated SPIONs and SPIONs-FA to HSF 1184, MDA-MB-231, MDA-MB-468, HeLa and 4 T1 cells was investigated using qualitative (Prussian Blue Assay) and quantitative (AAS) methods.
- vi) The MTT assay was carried out on HSF 1184, MDA-MB-231, MDA-MB-468, HeLa and 4 T1 cells to find the cytotoxicity effect of synthesized formulation in each step.
- vii) The biodistribution study was carried out in all important organs using colorimetric (AAS) and TEM methods in established 4 T1 tumor bearing mice model.
- viii) In the last step, after treatment protocol performance on 4 T1 tumor bearing mice model, the tumor volume and survival rate was measured to investigate the effectiveness of the proposed formulation on tumor bearing mice.

In this study, HeLa, 4 T1 and MDA-MB-231 cancer cell lines were used as (FAR+) targeted cells while HSF 1184 and MDA-MB-468 was used as (FAR-) non-targeted cells.

1.5 Significance of study

The synthesized SPIONs-Br-FA is a novel and safe delivery system to

minimize the side effect of anticancer agents to the normal tissues and to maximize their toxic effect on tumor site. In this study, a novel nano delivery system (SPIONs-Br-FA) was developed for the first time based on our hypothesis that the effectiveness of SPIONs-Br-FA on cancer cell was improved compared to neat Br treatment. Targeted delivery of bromelain as an anticancer agent to the cancer cells leads to the possibility of metastasis treatment and minor systemic side effects compared to the neat Br treatment. Another significance of this study includes the enhancement of bromelain delivery in the non-invasive and cost-effective manner utilizing minimal dosage of drug to achieve maximum potency.

1.6 Thesis Organization

The thesis is divided into five chapters. The first chapter describes the research background, problem statement, research objectives, scope of research and significance of study.

The second chapter consists of comprehensive literature review based on the research topics. In this chapter, the description on the related literatures such as barrier to conventional cancer treatments, tumor vasculature, drug delivery, nanotechnology advances in drug delivery, concept of passive and active targeting, active targeting by folic acid and recent work on bromelain are reviewed.

The third chapter explains the methodology used in this study. Initially, it describes the details and procedures to synthesize and characterize SPIONs-Br-FA and finally explains *in vitro* and *in vivo* assessments of synthesized formulations. This chapter also includes the list of materials used in this research.

Forth chapter exhibits and discusses the results obtained. This chapter has three main sections: (i) development and characterization of SPIONs-Br-FA, (ii) *in vitro* tests including biocompatibility studies, binding studies, cytotoxicity studies, morphological studies of cells, scratch motility and clonogenic assays, and (iii) *in*

REFERENCES

- Abdel-Mottaleb, M. M. A., Neumann, D. and Lamprecht, A. (2011). Lipid nanocapsules for dermal application: a comparative study of lipid-based versus polymer-based nanocarriers. *European Journal of Pharmaceutics and Biopharmaceutics*, 79(1), 36–42.
- Adamietz, I. A., Kurfürst, F., Müller, U., Renner, K. and Rimpler, M. (1989). Growth acceleration of Ehrlich ascites tumor cells treated by proteinase in vitro. *European Journal of Cancer and Clinical Oncology*, 25(12), 1837–1841.
- Agarwal, A., Shao, X., Rajian, J. R., Zhang, H., Chamberland, D. L., Kotov, N. A. and Wang, X. (2011). Dual-mode imaging with radiolabeled gold nanorods. *Journal of Biomedical Optics*, 16(5), 051307-051307.
- Alexiou, C., Arnold, W., Klein, R. J., Parak, F. G., Hulin, P., Bergemann, C., Erhardt, W., Wagenpfeil, S. and Luebbe, A. S. (2000). Locoregional cancer treatment with magnetic drug targeting. *Cancer Research*, 60(23), 6641–6648.
- Alexiou, C., Schmid, R. J., Jurgons, R., Kremer, M., Wanner, G., Bergemann, C., Huenges, E., Nawroth, T., Arnold, W. and Parak, F. G. (2006). Targeting cancer cells: magnetic nanoparticles as drug carriers. *European Biophysics Journal*, 35(5), 446–450.
- Alexis, F., Pridgen, E., Molnar, L. K. and Farokhzad, O. C. (2008). Reviews Factors Affecting the Clearance and Biodistribution of Polymeric Nanoparticles. *Molecular pharmaceutics*, 5(4), 505–515.
- Ali, A. S. G., Reza, M. A., Eshghi, H., Sazgarnia, A. and Montazerabadi, A. R. (2010). Cancerous Cells Targeting and Destruction Using Folate Conjugated Gold Nanoparticles. *Dyn Biochem Process Biotechnol Mol Biol*, 4(1), 06-12.
- Allen, T. M. (2002). Ligand-targeted therapeutics in anticancer therapy. *Nature Reviews Cancer*, 2(10), 750–763.
- Allen, T. M. and Cullis, P. R. (2004). Drug delivery systems: entering the mainstream. *Science*, 303(5665), 1818–1822.
- Almaki, J. H., Nasiri, R., Idris, A., Majid, F. A. A., Salouti, M., Wong, T. S., Dabagh, S., Marvibaigi, M. and Amini, N. (2016a). Synthesis, characterization and in vitro evaluation of exquisite targeting SPIONs–PEG–HER in HER2+

- human breast cancer cells. *Nanotechnology*, 27(10), 105601.
- Alphandery, E., Faure, S., Seksek, O., Guyot, F. and Chebbi, I. (2011). Chains of magnetosomes extracted from AMB-1 magnetotactic bacteria for application in alternative magnetic field cancer therapy. *ACS nano*, 5(8), 6279-6296.
- Amini, A., Ehteda, A., Masoumi, S., Akhter, M. J., Pillai, K. and Morris, D. L. (2013). Cytotoxic effects of bromelain in human gastrointestinal carcinoma cell lines (MKN45, KATO-III, HT29-5F12, and HT29-5M21). *OncoTargets & Therapy*, 6, 403-408.
- Amini, N., Abdul Majid, F. A., Marvibaigi, M., Supriyanto, E., Jaganathan, S. K., Tet Soon, W., Nasiri, R. and Hamzehalipour, J. (2016). CervicareTM induces apoptosis in HeLa and CaSki cells through ROS production and loss of mitochondrial membrane potential. *RSC Adv*, 6(29), 24391–24417.
- Ando, M., Yonemori, K., Katsumata, N., Shimizu, C., Hirata, T., Yamamoto, H., Hashimoto, K., Yunokawa, M., Tamura, K. and Fujiwara, Y. (2012). Phase I and pharmacokinetic study of nab-paclitaxel, nanoparticle albumin-bound paclitaxel, administered weekly to Japanese patients with solid tumors and metastatic breast cancer. *Cancer Chemotherapy and Pharmacology*, 69(2), 457–465.
- Arbab, A. S., Wilson, L. B., Ashari, P., Jordan, E. K., Lewis, B. K. and Frank, J. A. (2005). A model of lysosomal metabolism of dextran coated superparamagnetic iron oxide (SPIO) nanoparticles: implications for cellular magnetic resonance imaging. *NMR in Biomedicine*, 18(6), 383-389.
- Arruebo, M., Fernández-Pacheco, R., Ibarra, M. R. and Santamaría, J. (2007). Magnetic nanoparticles for drug delivery. *Nano Today*, 2(3), 22–32.
- Artemov, D. (2003). Molecular magnetic resonance imaging with targeted contrast agents. *Journal of Cellular Biochemistry*, 90(3), 518–524.
- Arvizo, R., Bhattacharya, R. and Mukherjee, P. (2010). Gold nanoparticles: opportunities and challenges in nanomedicine. *Expert Opinion on Drug Delivery*, 7(6), 753–763.
- Arvizo, R. R., De, M. and Rotello, V. M. (2007). *Proteins and Nanoparticles: Covalent and Noncovalent Conjugates*, in *Nanobiotechnology II: More Concepts and Applications*. Weinheim, Germany: Wiley-VCH Verlag GmbH

& Co. KGaA.

- Ashley, C. E., Carnes, E. C., Phillips, G. K., Padilla, D., Durfee, P. N., Brown, P. A., Hanna, T.N., Liu, J., Phillips, B. and Carter, M. B. (2011). The targeted delivery of multicomponent cargos to cancer cells by nanoporous particle-supported lipid bilayers. *Nature Materials*, 10(5), 389–397.
- Bae, K. H., Chung, H. J. and Park, T. G. (2011). Nanomaterials for cancer therapy and imaging. *Molecules and Cells*, 31(4), 295–302.
- Bae, Y. H. (2009). Drug targeting and tumor heterogeneity. *Journal of Controlled Release: Official Journal of the Controlled Release Society*, 133(1), 2.
- Báez, R., Lopes, M. T. P., Salas, C. E. and Hernández, M. (2007). In vivo antitumoral activity of stem pineapple (*Ananas comosus*) bromelain. *Planta Medica*, 73(13), 1377–1383.
- Bala, M., Ismail, N. A., Mel, M., Jami, M. S. and Jami, M. S. (2012). Bromelain Production : Current Trends and Perspective. *Archives Des Sciences*, 65(11), 369–399.
- Balogh, L., Nigavekar, S. S., Nair, B. M., Lesniak, W., Zhang, C., Sung, L. Y., Kariapper, M.S., El-Jawahri, A., Llanes, M., Bolton, B. and Mamou, F. (2007). Significant effect of size on the in vivo biodistribution of gold composite nanodevices in mouse tumor models. *Nanomedicine: Nanotechnology, Biology and Medicine*, 3(4), 281–296.
- Bander, N. H., Nanus, D. M., Milowsky, M. I., Kostakoglu, L., Vallabahajosula, S. and Goldsmith, S. J. (2003). Targeted systemic therapy of prostate cancer with a monoclonal antibody to prostate-specific membrane antigen. *Oncology*, 30(5), 667–676.
- Banerjee, R., Katsenovich, Y., Lagos, L., McIntosh, M., Zhang, X. and Li, C. Z. (2010). Nanomedicine: magnetic nanoparticles and their biomedical applications. *Current Medicinal Chemistry*, 17(27), 3120–3141.
- Barakat, N. S., Taleb, D. A. Bin and Salehi, A. S. Al. (2012). Target Nanoparticles : An Appealing Drug Delivery Platform, *Journal of Nanomedicine & Nanotechnology*, S4-009.
- Barz, M., Canal, F., Koynov, K., Zentel, R. and Vicent, M. J. (2010). Synthesis and in vitro evaluation of defined HPMA folate conjugates: influence of aggregation

- on folate receptor (FR) mediated cellular uptake. *Biomacromolecules*, 11(9), 2274–2282.
- Baumhackl, U., Kappos, L., Radue, E. W., Freitag, P., Guseo, A., Daumer, M. and Mertin, J. (2005). A randomized, double-blind, placebo-controlled study of oral hydrolytic enzymes in relapsing multiple sclerosis. *Multiple Sclerosis*, 11(2), 166–168.
- Bean, C. P. and Livingston, J. D. (1959). Superparamagnetism. *Journal of Applied Physics*, 30(4), S120–S129.
- Bereznicki, L. (2012). Factors affecting wound healing. *Australian Pharmacist*, 31(6), 484.
- Berry, C. C., Wells, S., Charles, S. and Curtis, A. S. G. (2003). Dextran and albumin derivatised iron oxide nanoparticles: influence on fibroblasts in vitro. *Biomaterials*, 24(25), 4551–4557.
- Beuth, J. and Braun, J. A. N. M. (2005). Modulation of murine tumor growth and colonization by bromelaine, an extract of the pineapple plant (*Ananas comosum* L.). *In Vivo*, 19(2), 483–485.
- Bharti, C., Nagaich, U., Pal, A. K. and Gulati, N. (2015). Mesoporous silica nanoparticles in target drug delivery system: a review. *International journal of pharmaceutical investigation*, 5(3), 124.
- Bhatnagar, P., Patnaik, S., Srivastava, A. K., Mudiam, M. K. R., Shukla, Y., Panda, A. K., Pant, A.B., Kumar, P. and Gupta, K. C. (2014). Anti-cancer activity of bromelain nanoparticles by oral administration. *Journal of Biomedical Nanotechnology*, 10(12), 3558–3575.
- Boisselier, E. and Astruc, D. (2009). Gold nanoparticles in nanomedicine: preparations, imaging, diagnostics, therapies and toxicity. *Chemical Society Reviews*, 38(6), 1759–1782.
- Bora, D. K. and Deb, P. (2008). Fatty Acid Binding Domain Mediated Conjugation of Ultrafine Magnetic Nanoparticles with Albumin Protein. *Nanoscale Research Letters*, 4(2), 138–143.
- Boyle, P. and Levin, B. (2008). *World cancer report 2008*. Lyon, France: IARC Press, International Agency for Research on Cancer.
- Bradford, M. M. (1976). A Rapid and Sensitive Method for the Quantitation of

- Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical and Bioanalytical Chemistry*, 72(1-2), 248–254.
- Brien, S., Lewith, G., Walker, A. F., Middleton, R., Prescott, P. and Bundy, R. (2006). Bromelain as an adjunctive treatment for moderate-to-severe osteoarthritis of the knee: a randomized placebo-controlled pilot study. *QJM*, 99(12), 841–850.
- Brien, S., Lewith, G., Walker, A., Hicks, S. M. and Middleton, D. (2004). Bromelain as a treatment for osteoarthritis: a review of clinical studies. *Evidence-Based Complementary and Alternative Medicine*, 1(3), 251–257.
- Briley-Saebo, K., Bjørnerud, A., Grant, D., Ahlstrom, H., Berg, T. and Kindberg, G. M. (2004). Hepatic cellular distribution and degradation of iron oxide nanoparticles following single intravenous injection in rats: implications for magnetic resonance imaging. *Cell and tissue research*, 316(3), 315–323.
- Brown, S. D., Nativo, P., Smith, J. A., Stirling, D., Edwards, P. R., Venugopal, B., Flint, D.J., Plumb, J.A., Graham, D. and Wheate, N. J. (2010). Gold nanoparticles for the improved anticancer drug delivery of the active component of oxaliplatin. *Journal of the American Chemical Society*, 132(13), 4678–4684.
- Bruchez, M., Moronne, M., Gin, P., Weiss, S. and Alivisatos, A. P. (1998). Semiconductor nanocrystals as fluorescent biological labels. *Science*, 281(5385), 2013–2016.
- Brusentsov, N. A., Gogosov, V. V, Brusentsova, T. N., Sergeev, A. V, Jurchenko, N. Y., Kuznetsov, A. A., Kuznetsov, O.A. and Shumakov, L. I. (2001). Evaluation of ferromagnetic fluids and suspensions for the site-specific radiofrequency-induced hyperthermia of MX11 sarcoma cells in vitro. *Journal of Magnetism and Magnetic Materials*, 225(1), 113–117.
- Bulte, J. W. M., Ben-Hur, T., Miller, B. R., Mizrahi-Kol, R., Einstein, O., Reinhartz, E., Zywicke, H.A., Douglas, T. and Frank, J. A. (2003). MR microscopy of magnetically labeled neurospheres transplanted into the Lewis EAE rat brain. *Magnetic Resonance in Medicine*, 50(1), 201–205.
- Bulte, J. W. M., Ma, L. D., Magin, R. L., Kamman, R. L., Hulstaert, C. E., Go, K. G., The, T.H. and De Leij, L. (1993). Selective MR imaging of labeled human peripheral blood mononuclear cells by liposome mediated incorporation of

- dextran magnetite particles. *Magnetic Resonance in Medicine*, 29(1), 32–37.
- Burda, C., Chen, X., Narayanan, R. and El-Sayed, M. A. (2005). Chemistry and properties of nanocrystals of different shapes. *Chemical Reviews*, 105(4), 1025–1102.
- Burns, A., Ow, H. and Wiesner, U. (2006). Fluorescent core–shell silica nanoparticles: towards “Lab on a Particle” architectures for nanobiotechnology. *Chemical Society Reviews*, 35(11), 1028–1042.
- Byrne, J. D., Betancourt, T. and Brannon-Peppas, L. (2008). Active targeting schemes for nanoparticle systems in cancer therapeutics. *Advanced Drug Delivery Reviews*, 60(15), 1615–1626.
- Cairns, R., Papandreou, I. and Denko, N. (2006). Overcoming physiologic barriers to cancer treatment by molecularly targeting the tumor microenvironment. *Molecular Cancer Research*, 4(2), 61–70.
- Campbell, I. G., Jones, T. A., Foulkes, W. D. and Trowsdale, J. (1991). Folate-binding protein is a marker for ovarian cancer. *Cancer Research*, 51(19), 5329–5338.
- Carmeliet, P. and Jain, R. K. (2000). Angiogenesis in cancer and other diseases. *Nature*, 407(6801), 249–257.
- Cassidy, J., Clarke, S., Díaz-Rubio, E., Scheithauer, W., Figer, A., Wong, R., Koski, S., Lichinitser, M., Yang, T.S., Rivera, F. and Rivera, F. (2008). Randomized phase III study of capecitabine plus oxaliplatin compared with fluorouracil/folinic acid plus oxaliplatin as first-line therapy for metastatic colorectal cancer. *Journal of Clinical Oncology*, 26(12), 2006–2012.
- Castell, J. V., Friedrich, G., Kuhn, C. S. and Poppe, G. E. (1997). Intestinal absorption of undegraded proteins in men: presence of bromelain in plasma after oral intake. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 273(1), G139–G146.
- Chandel, A. K. S., Kumar, C. U. and Jewrajka, S. K. (2016). Effect of Polyethylene Glycol on Properties and Drug Encapsulation–Release Performance of Biodegradable/Cytocompatible Agarose–Polyethylene Glycol–Polycaprolactone Amphiphilic Co-Network Gels. *ACS applied materials & interfaces*, 8(5), 3182–3192.

- Chandna, A., Batra, D., Kakar, S. and Singh, R. (2013). A review on target drug delivery: magnetic microspheres. *Journal of Acute Disease*, 2(3), 189-195.
- Chau, Y., Tan, F. E. and Langer, R. (2004). Synthesis and characterization of dextran-peptide-methotrexate conjugates for tumor targeting via mediation by matrix metalloproteinase II and matrix metalloproteinase IX. *Bioconjugate Chemistry*, 15(4), 931-41.
- Chen, B., Wu, W. and Wang, X. (2011). Magnetic iron oxide nanoparticles for tumor-targeted therapy. *Current Cancer Drug Targets*, 11(2), 184-189.
- Chen, L., Mooso, B. A., Jathal, M. K., Madhav, A., Johnson, S. D., Van Spyk, E., Mikhailova, M., Zierenberg-Ripoll, A., Xue, L., Vinall, R.L. and Vinall, R. L. (2011). Dual EGFR/HER2 inhibition sensitizes prostate cancer cells to androgen withdrawal by suppressing ErbB3. *Clinical Cancer Research*, 17(19), 6218-6228.
- Chen, L.-C., Wu, Y. H., Liu, I. H., Ho, C. L., Lee, W. C., Chang, C. H. and Shien, J. H. (2012). Pharmacokinetics, dosimetry and comparative efficacy of 188 Re-liposome and 5-FU in a CT26-luc lung-metastatic mice model. *Nuclear Medicine and Biology*, 39(1), 35-43.
- Chen, T., Cheng, T., Hung, Y., Lin, K., Liu, G. and Wang, Y. (2008). Targeted folic acid PEG nanoparticles for noninvasive imaging of folate receptor by MRI. *Journal of Biomedical Materials Research Part A*, 87(1), 165-175.
- Cho, K., Wang, X. U., Nie, S. and Shin, D. M. (2008). Therapeutic nanoparticles for drug delivery in cancer. *Clinical Cancer Research*, 14(5), 1310-1316.
- Chobotova, K., Vernallis, A. B. and Majid, F. A. A. (2010). Bromelain's activity and potential as an anti-cancer agent: current evidence and perspectives. *Cancer Letters*, 290(2), 148-156.
- Choi, H., Choi, S. R., Zhou, R., Kung, H. F. and Chen, I. W. (2004). Iron oxide nanoparticles as magnetic resonance contrast agent for tumor imaging via folate receptor-targeted delivery 1. *Academic Radiology*, 11(9), 996-1004.
- Cole, A. J., Yang, V. C. and David, A. E. (2011). Cancer theranostics: the rise of targeted magnetic nanoparticles. *Trends in Biotechnology*, 29(7), 323-332.
- Coney, L. R., Tomassetti, A., Carayannopoulos, L., Frasca, V., Kamen, B. A., Colnaghi, M. I. and Zurawski, V. R. (1991). Cloning of a tumor-associated

- antigen: MOv18 and MOv19 antibodies recognize a folate-binding protein. *Cancer Research*, 51(22), 6125–6132.
- Constine, L. S., Milano, M. T., Friedman, D., Morris, M., Williams, J. P., Rubin, P. and Okunie, P. (2008). *Late effects of cancer treatment on normal tissues. Principles and Practice of Radiation Oncology*. (5th Ed.). Newyork: Springer.
- Corot, C., Robert, P., Idée, J. M. and Port, M. (2006). Recent advances in iron oxide nanocrystal technology for medical imaging. *Advanced drug delivery reviews*, 58(14), 1471-1504.
- Danhier, F., Feron, O. and Pr at, V. (2010a). To exploit the tumor microenvironment: passive and active tumor targeting of nanocarriers for anti-cancer drug delivery. *Journal of Controlled Release*, 148(2), 135–146.
- Danhier, F., Feron, O. and Pr at, V. (2010b). To exploit the tumor microenvironment: Passive and active tumor targeting of nanocarriers for anti-cancer drug delivery. *Journal of Controlled Release*, 148(2), 135–46.
- De Bono, J. S., Logothetis, C. J., Molina, A., Fizazi, K., North, S., Chu, L. and Saad, F. (2011). Abiraterone and increased survival in metastatic prostate cancer. *New England Journal of Medicine*, 364(21), 1995–2005.
- De Cuyper, M. and Joniau, M. (1988). Magnetoliposomes. *European Biophysics Journal*, 15(5), 311–319.
- De Freitas, E. R. L., Soares, P. R. O., de Paula Santos, R., dos Santos, R. L., da Silva, J. R., Porfirio, E. P. and Moraes, P. C. (2008). In Vitro Biological Activities of Anionic-Fe₂O₃ Nanoparticles on Human Melanoma Cells. *Journal of Nanoscience and Nanotechnology*, 8(5), 2385–2391.
- De Souza, R., Zahedi, P., Allen, C. J. and Piquette-Miller, M. (2010). Polymeric drug delivery systems for localized cancer chemotherapy. *Drug Delivery*, 17(6), 365–375.
- Dhar, S., Reddy, E. M., Shiras, A., Pokharkar, V. and Prasad, B. E. E. (2008). Natural gum reduced/stabilized gold nanoparticles for drug delivery formulations. *Chemistry–A European Journal*, 14(33), 10244-10250.
- Desagher, S. and Martinou, J. C. (2000). Mitochondria as the central control point of apoptosis. *Trends in Cell Biology*, 10(9), 369–377.
- Differences between normal cell and tumor* 2009. Available from: <

- review.ca/R10-36-medical05b.htm>. [2009]
- Douillard, J. Y., Siena, S., Cassidy, J., Tabernero, J., Burkes, R., Barugel, M. and Jassem, J. (2010). Randomized, phase III trial of panitumumab with infusional fluorouracil, leucovorin, and oxaliplatin (FOLFOX4) versus FOLFOX4 alone as first-line treatment in patients with previously untreated metastatic colorectal cancer: the PRIME study. *Journal of Clinical Oncology*, JCO–2009.
- Dreaden, E. C., Austin, L. A., Mackey, M. A., and El-Sayed, M. A. (2012). Size matters: gold nanoparticles in targeted cancer drug delivery. *Therapeutic delivery*, 3(4), 457-478.
- Dreher, M. R., Liu, W., Michelich, C. R., Dewhirst, M. W., Yuan, F. and Chilkoti, A. (2006). Tumor vascular permeability, accumulation, and penetration of macromolecular drug carriers. *Journal of the National Cancer Institute*, 98(5), 335–344.
- Duncan, R. (2006). Polymer conjugates as anticancer nanomedicines. *Nature Reviews Cancer*, 6(9), 688–701.
- Durgadas, C. V, Sreenivasan, K. and Sharma, C. P. (2012). Bright blue emitting CuSe/ZnS/silica core/shell/shell quantum dots and their biocompatibility. *Biomaterials*, 33(27), 6420–6429.
- Eckert, K., Grabowska, E., Stange, R., Schneider, U., Eschmann, K. and Maurer, H. R. (1999). Effects of oral bromelain administration on the impaired immunocytotoxicity of mononuclear cells from mammary tumor patients. *Oncology Reports*, 6, 1191–1200.
- Engels, F. K., Mathot, R. A. A. and Verweij, J. (2007). Alternative drug formulations of docetaxel: a review. *Anti-Cancer Drugs*, 18(2), 95–103.
- Evan, G. I. and Vousden, K. H. (2001). Proliferation, cell cycle and apoptosis in cancer. *Nature*, 411(6835), 342–348.
- Fang, C. and Zhang, M. (2009). Multifunctional magnetic nanoparticles for medical imaging applications. *Journal of Materials Chemistry*, 19(35), 6258–6266.
- Fang, J., Nakamura, H. and Maeda, H. (2011). The EPR effect: unique features of tumor blood vessels for drug delivery, *factors involved, and limitations and augmentation of the effect*. *Advanced Drug Delivery Reviews*, 63(3), 136–151.
- Felici, A., Verweij, J. and Sparreboom, A. (2002). Dosing strategies for anticancer

- drugs: the good, the bad and body-surface area. *European Journal of Cancer*, 38(13), 1677–1684.
- Lodhia, J., Mandarano, G., Ferris, N. J., Eu, P. and Cowell, S. F. (2010). Development and use of iron oxide nanoparticles (Part 1): Synthesis of iron oxide nanoparticles for MRI. *Biomed Imaging Interv J*, 6(2), e12.
- Fink, S. L. and Cookson, B. T. (2005). Apoptosis, Pyroptosis, and Necrosis: Mechanistic Description of Dead and Dying Eukaryotic Cells. *Infection and Immunity*, 73(4), 1907–1916.
- Fisher, D. E. (1994). Apoptosis in cancer therapy: crossing the threshold. *Cell*, 78(4), 539–542.
- Fleming, M. T., Sonpavde, G., Kolodziej, M., Awasthi, S., Hutson, T. E., Martincic, D. and Galsky, M. D. (2012). Association of rash with outcomes in a randomized phase II trial evaluating cetuximab in combination with mitoxantrone plus prednisone after docetaxel for metastatic castration-resistant prostate cancer. *Clinical Genitourinary Cancer*, 10(1), 6–14.
- Fulda, S. and Debatin, K.M. (2004). Targeting Apoptosis Pathways in Cancer Therapy. *Current Cancer Drug Targets*, 4(7), 569–576.
- Fura, A., Harper, T. W., Zhang, H., Fung, L. and Shyu, W. C. (2003). Shift in pH of biological fluids during storage and processing: effect on bioanalysis. *Journal of Pharmaceutical and Biomedical Analysis*, 32(3), 513–522.
- Gabizon, A. and Martin, F. (1997). Polyethylene glycol-coated (pegylated) liposomal doxorubicin. *Drugs*, 54(4), 15–21.
- Gani, M. B. A., Nasiri, R., Almaki, J. H., Majid, F. A. A., Marvibaigi, M., Amini, N. and Mashudin, M. (2015). In Vitro Antiproliferative Activity of Fresh Pineapple Juices on Ovarian and Colon Cancer Cell Lines. *International Journal of Peptide Research and Therapeutics*, 21(3), 353–364.
- Gao, X., Cui, Y., Levenson, R. M., Chung, L. W. K. and Nie, S. (2004). In vivo cancer targeting and imaging with semiconductor quantum dots. *Nature Biotechnology*, 22(8), 969–976.
- Garin-Chesa, P., Campbell, I., Saigo, P. E., Lewis Jr, J. L., Old, L. J. and Rettig, W. J. (1993). Trophoblast and ovarian cancer antigen LK26. Sensitivity and specificity in immunopathology and molecular identification as a folate-binding

- protein. *The American Journal of Pathology*, 142(2), 557.
- Giri, S., Trewyn, B. G. and Lin, V. S. Y. (2007). Mesoporous silica nanomaterial-based biotechnological and biomedical delivery systems, *Future Medicine*, 2(1), 99-111.
- Giri, S., Trewyn, B. G., Stellmaker, M. P. and Lin, V. S. (2005). Stimuli-responsive controlled release delivery system based on mesoporous silica nanorods capped with magnetic nanoparticles. *Angewandte Chemie International Edition*, 44(32), 5038–5044.
- Gokce, G., Cital, M., Gunes, V. and Atalan, G. (2004). Effect of time delay and storage temperature on blood gas and acid–base values of bovine venous blood. *Research in Veterinary Science*, 76(2), 121–127.
- Goss, G. D., Arnold, A., Shepherd, F. A., Dediu, M., Ciuleanu, T. E., Fenton, D. and Vincent, M. D. (2010). Randomized, double-blind trial of carboplatin and paclitaxel with either daily oral cediranib or placebo in advanced non–small-cell lung cancer: NCIC Clinical Trials Group BR24 study. *Journal of Clinical Oncology*, 28(1), 49–55.
- Gottesman, M. M., Fojo, T. and Bates, S. E. (2002). Multidrug resistance in cancer: role of ATP–dependent transporters. *Nature Reviews Cancer*, 2(1), 48–58.
- Goya, G. F., Grazu, V. and Ibarra, M. R. (2008). Magnetic nanoparticles for cancer therapy. *Current Nanoscience*, 4(1), 1–16.
- Greco, F. and Vicent, M. J. (2009). Combination therapy: opportunities and challenges for polymer–drug conjugates as anticancer nanomedicines. *Advanced Drug Delivery Reviews*, 61(13), 1203–1213.
- Grüttner, C., Müller, K., Teller, J., Westphal, F., Foreman, A. and Ivkov, R. (2007). Synthesis and antibody conjugation of magnetic nanoparticles with improved specific power absorption rates for alternating magnetic field cancer therapy. *Journal of Magnetism and Magnetic Materials*, 311(1), 181–186.
- Guimaraes-Ferreira, C. A., Rodrigues, E. G., Mortara, R. A., Cabral, H., Serrano, F. A., Ribeiro-dos-Santos, R. and Travassos, L. R. (2007). Antitumor effects in vitro and in vivo and mechanisms of protection against melanoma B16F10-Nex2 cells by fastuosain, a cysteine proteinase from *Bromelia fastuosa*. *Neoplasia*, 9(9), 723–733.

- Gullotti, E. and Yeo, Y. (2009). Extracellularly activated nanocarriers: a new paradigm of tumor targeted drug delivery. *Molecular Pharmaceutics*, 6(4), 1041–1051.
- Guo, M., Yan, Y., Liu, X., Yan, H., Liu, K., Zhang, H. and Cao, Y. (2010). Multilayer nanoparticles with a magnetite core and a polycation inner shell as pH-responsive carriers for drug delivery. *Nanoscale*, 2(3), 434–441.
- Guo, R., Canter, P. H. and Ernst, E. (2006). Herbal medicines for the treatment of rhinosinusitis: a systematic review. *Otolaryngology-Head and Neck Surgery*, 135(4), 496–506.
- Gupta, A. K. and Gupta, M. (2005a). Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications. *Biomaterials*, 26(18), 3995–4021.
- Gupta, A. K. and Gupta, M. (2005b). Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications. *Biomaterials*, 26(18), 3995–4021.
- Guyton, A. C. and Hall, J. E. (2015). *Text book of medical physiology*. (12th ed.). USA: Elsevier.
- Hahn, M. A., Singh, A. K., Sharma, P., Brown, S. C. and Moudgil, B. M. (2011). Nanoparticles as contrast agents for in-vivo bioimaging: current status and future perspectives. *Analytical and Bioanalytical Chemistry*, 399(1), 3–27.
- Hale, L. P. (2004). Proteolytic activity and immunogenicity of oral bromelain within the gastrointestinal tract of mice. *International Immunopharmacology*, 4(2), 255–264.
- Hale, L. P., Greer, P. K., Trinh, C. T. and James, C. L. (2005). Proteinase activity and stability of natural bromelain preparations. *International Immunopharmacology*, 5(4), 783–793.
- Han, D. H., Luo, H. L. and Yang, Z. (1996). Remanent and anisotropic switching field distribution of platelike Ba-ferrite and acicular particulate recording media. *Journal of Magnetism and Magnetic Materials*, 161, 376–378.
- Hanahan, D. and Weinberg, R. A. (2000). The hallmarks of cancer. *Cell*, 100(1), 57–70.
- Hanahan, D. and Weinberg, R. A. (2011). Hallmarks of cancer: the next generation. *Cell*, 144(5), 646–674.
- Harris, L. A., Goff, J. D., Carmichael, A. Y., Riffle, J. S., Harburn, J. J., St. Pierre, T.

- G. and Saunders, M. (2003). Magnetite nanoparticle dispersions stabilized with triblock copolymers. *Chemistry of Materials*, 15(6), 1367–1377.
- Hashizume, H., Baluk, P., Morikawa, S., McLean, J. W., Thurston, G., Roberge, S. and McDonald, D. M. (2000). Openings between defective endothelial cells explain tumor vessel leakiness. *The American Journal of Pathology*, 156(4), 1363–1380.
- Haun, J. B., Yoon, T., Lee, H. and Weissleder, R. (2010). Magnetic nanoparticle biosensors. *Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology*, 2(3), 291–304.
- Heidari, Z., Sariri, R., and Salouti, M. (2014). Gold nanorods-bombesin conjugate as a potential targeted imaging agent for detection of breast cancer. *Journal of Photochemistry and Photobiology B: Biology*, 130, 40-46.
- Hennenfent, K. L. and Govindan, R. (2006). Novel formulations of taxanes: a review. Old wine in a new bottle. *Annals of Oncology*, 17(5), 735–749.
- Hickman, J. A. (1992). Apoptosis induced by anticancer drugs. *Cancer and Metastasis Reviews*, 11(2), 121–139.
- Hild, W. A., Breunig, M., and Göpferich, A. (2008). Quantum dots–nano-sized probes for the exploration of cellular and intracellular targeting. *European Journal of Pharmaceutics and Biopharmaceutics*, 68(2), 153-168.
- Hirsch, F. R., Varella-Garcia, M., Bunn, P. A., Di Maria, M. V, Veve, R., Bremnes, R. M. and Franklin, W. A. (2003). Epidermal growth factor receptor in non–small-cell lung carcinomas: correlation between gene copy number and protein expression and impact on prognosis. *Journal of Clinical Oncology*, 21(20), 3798–3807.
- Hirsch, L., Stafford, R. J., Bankson, J. A., Sershen, S. R., Rivera, B., Price, R. E. and West, J. L. (2003). Nanoshell-mediated near-infrared thermal therapy of tumors under magnetic resonance guidance. *Proceedings of the National Academy of Sciences*, 100(23), 13549–13554.
- Ho, K. S. (2012). *Targeted drug delivery to breast cancer using polymeric nanoparticle micelles*, PhD Thesis Page 8, University of Toronto, Canada.
- Hobbs, S. K., Monsky, W. L., Yuan, F., Roberts, W. G., Griffith, L., Torchilin, V. P. and Jain, R. K. (1998). Regulation of transport pathways in tumor vessels: role

- of tumor type and microenvironment. *Proceedings of the National Academy of Sciences*, 95(8), 4607–4612.
- Holm, J. A. N., Hansen, S. I., HØier-Madsen, M., SØndergaard, K. and Bzorek, M. (1994). Folate receptor of human mammary adenocarcinoma. *Apmis*, 102(1-6), 413–419.
- Huang, F. K., Chen, W. C., Lai, S. F., Liu, C. J., Wang, C. L., Wang, C. H. and Wu, M. K. (2009). Enhancement of irradiation effects on cancer cells by cross-linked dextran-coated iron oxide (CLIO) nanoparticles. *Physics in Medicine and Biology*, 55(2), 469.
- Huang, H. C., Barua, S., Sharma, G., Dey, S. K. and Rege, K. (2011). Inorganic nanoparticles for cancer imaging and therapy. *Journal of Controlled Release*, 155(3), 344–357.
- Huang, J. R., Wu, C. C., Hou, R. C. W. and Jeng, K. C. (2008). Bromelain inhibits lipopolysaccharide-induced cytokine production in human THP-1 monocytes via the removal of CD14. *Immunological Investigations*, 37(4), 263–277.
- Huang, X., El-Sayed, I. H., Qian, W. and El-Sayed, M. A. (2006). Cancer cell imaging and photothermal therapy in the near-infrared region by using gold nanorods. *Journal of the American Chemical Society*, 128(6), 2115–2120.
- Hudis, C. A. (2007). Trastuzumab-mechanism of action and use in clinical practice. *New England Journal of Medicine*, 357(1), 39–51.
- Hurwitz, H., Fehrenbacher, L., Novotny, W., Cartwright, T., Hainsworth, J., Heim, W. and Holmgren, E. (2004). Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *New England Journal of Medicine*, 350(23), 2335–2342.
- Idris, A., Hassan, N., Suriani, N., Ismail, M., Misran, E., Mohd, N. and Bee, A. (2010). Photocatalytic magnetic separable beads for chromium (VI) reduction. *Water Research*, 44(6), 1683–1688.
- Idris, A., Ismail, N. S. M., Hassan, N., Misran, E. and Ngomsik, A. F. (2012). Synthesis of magnetic alginate beads based on maghemite nanoparticles for Pb(II) removal in aqueous solution. *Journal of Industrial and Engineering Chemistry*, 18(5), 1582–1589.
- Illés, E., Szekeres, M., Kupcsik, E., Tóth, I. Y., Farkas, K., Jedlovszky-Hajdú, A. and

- Tombácz, E. (2014). PEGylation of surfacted magnetite core-shell nanoparticles for biomedical application. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 460, 429–440.
- Iyer, A. K., Khaled, G., Fang, J. and Maeda, H. (2006). Exploiting the enhanced permeability and retention effect for tumor targeting. *Drug Discovery Today*, 11(17), 812–818.
- Jain, K. K. (2012). Advances in use of functionalized carbon nanotubes for drug design and discovery. *Expert Opinion on Drug Discovery*, 7(11), 1029–1037.
- Jain, R. K. (2005). Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science*, 307(5706), 58–62.
- Jain, R. K. and Stylianopoulos, T. (2010). Delivering nanomedicine to solid tumors. *Nature Reviews Clinical Oncology*, 7(11), 653–664.
- Jain, T. K., Morales, M. A., Sahoo, S. K., Leslie-Pelecky, D. L. and Labhasetwar, V. (2005). Iron oxide nanoparticles for sustained delivery of anticancer agents. *Molecular Pharmaceutics*, 2(3), 194–205.
- Jangde, R. (2011). Magnetically modulated Drug Delivery Systems: An Overview. *Research Journal of Pharmacy and Technology*, 4(11), 1649–1657.
- Jayalekshmi, A. C., Victor, S. P. and Sharma, C. P. (2013). Magnetic and degradable polymer/bioactive glass composite nanoparticles for biomedical applications. *Colloids and Surfaces B: Biointerfaces*, 101, 196–204.
- Jemal, A., Siegel, R., Xu, J. and Ward, E. (2010). Cancer statistics, 2010. *CA: A Cancer Journal for Clinicians*, 60(5), 277–300.
- Jokerst, J. V and Gambhir, S. S. (2011). Molecular imaging with theranostic nanoparticles. *Accounts of Chemical Research*, 44(10), 1050–1060.
- Jones, A. and Harris, A. L. (1998). New developments in angiogenesis: a major mechanism for tumor growth and target for therapy. *The Cancer Journal from Scientific American*, 4(4), 209.
- Jordan, M. A. and Wilson, L. (2004). Microtubules as a target for anticancer drugs. *Nature Reviews Cancer*, 4(4), 253–265.
- Jung, J., Matsuzaki, T., Tatematsu, K., Okajima, T., Tanizawa, K. and Kuroda, S. (2008). Bio-nanocapsule conjugated with liposomes for in vivo pinpoint delivery of various materials. *Journal of Controlled Release*, 126(3), 255–264.

- Kainz, Q. M., Fernandes, S., Eichenseer, C. M., Besostri, F., Körner, H., Müller, R. and Reiser, O. (2015). Synthesis of functionalized, dispersible carbon-coated cobalt nanoparticles for potential biomedical applications. *Faraday Discuss*, 175, 27–40.
- Kamal Ahmadi, M. and Vossoughi, M. (2013). Immobilization of α -Chymotrypsin on the Surface of Magnetic/Gold Core/Shell Nanoparticles. *Journal of Nanotechnology*, 2013, 1–7.
- Karp, D. D., Paz-Ares, L. G., Novello, S., Haluska, P., Garland, L., Cardenal, F. and Blumenschein, G. (2009). Phase II study of the anti-insulin-like growth factor type 1 receptor antibody CP-751,871 in combination with paclitaxel and carboplatin in previously untreated, locally advanced, or metastatic non-small-cell lung cancer. *Journal of Clinical Oncology*, 27(15), 2516–2522.
- Kaufmann, S. H. and Earnshaw, W. C. (2000). Induction of apoptosis by cancer chemotherapy. *Experimental Cell Research*, 256(1), 42–49.
- Keating, G. M. (2012). Pertuzumab. *Drugs*, 72(3), 353–360.
- Kerbel, R. S. (2000). Tumor angiogenesis: past, present and the near future. *Carcinogenesis*, 21(3), 505–515.
- Kerbel, R. S., Cornil, I. and Theodorescu, D. (1991). Importance of orthotopic transplantation procedures in assessing the effects of transfected genes on human tumor growth and metastasis. *Cancer and Metastasis Reviews*, 10(3), 201–215.
- Kerr, J. F., Wyllie, A. H. and Currie, A. R. (1972). Apoptosis: A basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer*, 26(4), 239.
- Kievit, F. M., Wang, F. Y., Fang, C., Mok, H., Wang, K., Silber, J. R. and Zhang, M. (2011). Doxorubicin loaded iron oxide nanoparticles overcome multidrug resistance in cancer in vitro. *Journal of Controlled Release*, 152(1), 76–83.
- Killion, J. J., Radinsky, R. and Fidler, I. J. (1998). Orthotopic models are necessary to predict therapy of transplantable tumors in mice. *Cancer and Metastasis Reviews*, 17(3), 279–284.
- Klein, G., Kullich, W., Schnitker, J. and Schwann, H. (2006). Efficacy and tolerance of an oral enzyme combination in painful osteoarthritis of the hip. A double-

- blind, randomised study comparing oral enzymes with non-steroidal anti-inflammatory drugs. *Clinical and Experimental Rheumatology*, 24(1), 25.
- Knop, K., Hoogenboom, R., Fischer, D. and Schubert, U. S. (2010). Poly (ethylene glycol) in drug delivery: pros and cons as well as potential alternatives. *Angewandte Chemie International Edition*, 49(36), 6288–6308.
- Kramer, N., Walzl, A., Unger, C., Rosner, M., Krupitza, G., Hengstschläger, M., and Dolznig, H. (2013). In vitro cell migration and invasion assays. *Mutation Research/Reviews in Mutation Research*, 752(1), 10-24.
- Krop, I. E., Beeram, M., Modi, S., Jones, S. F., Holden, S. N., Yu, W. and Sliwkowski, M. X. (2010). Phase I study of trastuzumab-DM1, an HER2 antibody-drug conjugate, given every 3 weeks to patients with HER2-positive metastatic breast cancer. *Journal of Clinical Oncology*, 28(16), 2698–2704.
- Kukowska-Latallo, J. F., Candido, K. A., Cao, Z., Nigavekar, S. S., Majoros, I. J., Thomas, T. P. and Baker, J. R. (2005). Nanoparticle targeting of anticancer drug improves therapeutic response in animal model of human epithelial cancer. *Cancer Research*, 65(12), 5317–5324.
- Kumar, A. and Gupta, M. (2005). Synthesis and surface engineering of iron oxide nanoparticles for biomedical applications. *Biomaterials*, 26, 3995–4021.
- L Arias, J. (2011). Drug targeting strategies in cancer treatment: an overview. *Mini Reviews in Medicinal Chemistry*, 11(1), 1–17.
- Ladino, C. A., Chari, R. V. J., Bourret, L. A., Kedersha, N. L. and Goldmacher, V. S. (1997). Folate-maytansinoids: Target-selective drugs of low molecular weight. *International Journal of Cancer*, 73(6), 859–864.
- Lamanna, G., Kueny-Stotz, M., Mamlouk-Chaouachi, H., Ghobril, C., Basly, B., Bertin, A. and Begin-Colin, S. (2011). Dendronized iron oxide nanoparticles for multimodal imaging. *Biomaterials*, 32(33), 8562–8573.
- Lapresta-Fernández, A., Doussineau, T., Dutz, S., Steiniger, F., Moro, A. J. and Mohr, G. J. (2011). Magnetic and fluorescent core-shell nanoparticles for ratiometric pH sensing. *Nanotechnology*, 22(41), 415501.
- Laroui, H., Wilson, D. S., Dalmasso, G., Salaita, K., Murthy, N., Sitaraman, S. V. and Merlin, D. (2011). Nanomedicine in GI. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 300 (3), G371-G383.

- Lasic, D. D. (1997). Recent developments in medical applications of liposomes: sterically stabilized liposomes in cancer therapy and gene delivery in vivo. *Journal of controlled release*, 48(2), 203-222.
- Laurent, S., Dutz, S., Häfeli, U. O. and Mahmoudi, M. (2011). Magnetic fluid hyperthermia: focus on superparamagnetic iron oxide nanoparticles. *Advances in Colloid and Interface Science*, 166(1), 8–23.
- Laurent, S., Forge, D., Port, M., Roch, A., Robic, C., Vander Elst, L. and Muller, R. N. (2008). Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications. *Chemical Reviews*, 108(6), 2064–110.
- Lee, I.-H., Bulte, J. W. M., Schweinhardt, P., Douglas, T., Trifunovski, A., Hofstetter, C. and Spenger, C. (2004). In vivo magnetic resonance tracking of olfactory ensheathing glia grafted into the rat spinal cord. *Experimental Neurology*, 187(2), 509–516.
- Lee, J. H., Huh, Y. M., Jun, Y., Seo, J., Jang, J., Song, H. T., Suh, J. S. (2007). Artificially engineered magnetic nanoparticles for ultra-sensitive molecular imaging. *Nature Medicine*, 13(1), 95–99.
- Lee, S. and Pérez-Luna, V. H. (2005). Dextran-gold nanoparticle hybrid material for biomolecule immobilization and detection. *Analytical chemistry*, 77(22), 7204-7211.
- Leuschner, C., Kumar, C. S. S. R., Hansel, W., Soboyejo, W., Zhou, J. and Hormes, J. (2006). LHRH-conjugated magnetic iron oxide nanoparticles for detection of breast cancer metastases. *Breast Cancer Research and Treatment*, 99(2), 163–76.
- Levy, M., Luciani, N., Alloyeau, D., Elgrabli, D., Deveaux, V., Pechoux and C., Factor, C. (2011). Long term in vivo biotransformation of iron oxide nanoparticles. *Biomaterials*, 32(16), 3988-3999.
- Li, L., Mak, K. Y., Leung, C. W., Chan, K. Y., Chan, W. K., Zhong, W. and Pong, P. W. T. (2013). Effect of synthesis conditions on the properties of citric-acid coated iron oxide nanoparticles. *Microelectronic Engineering*, 110, 329–334.
- Libutti, S. K., Paciotti, G. F., Byrnes, A. A., Alexander, H. R., Gannon, W. E., Walker, M. and Tamarkin, L. (2010). Phase I and pharmacokinetic studies of

- CYT-6091, a novel PEGylated colloidal gold-rhTNF nanomedicine. *Clinical Cancer Research*, 16(24), 6139–6149.
- Liechty, W. B. and Peppas, N. A. (2012). Expert opinion: responsive polymer nanoparticles in cancer therapy. *European Journal of Pharmaceutics and Biopharmaceutics*, 80(2), 241–246.
- Lipka, J., Semmler-Behnke, M., Sperling, R. A., Wenk, A., Takenaka, S., Schleh, C. and Kreyling, W. G. (2010). Biodistribution of PEG-modified gold nanoparticles following intratracheal instillation and intravenous injection. *Biomaterials*, 31(25), 6574–6581.
- Lim, D. J., Sim, M., Oh, L., Lim, K. and Park, H. (2014). Carbon-based drug delivery carriers for cancer therapy. *Archives of pharmacal research*, 37(1), 43–52.
- Liu, C. and Huang, P. M. (1999). Atomic force microscopy and surface characteristics of iron oxides formed in citrate solutions. *Soil Science Society of America Journal*, 63(1), 65–72.
- Liu, Z., Robinson, J. T., Tabakman, S. M., Yang, K., and Dai, H. (2011). Carbon materials for drug delivery & cancer therapy. *Materials today*, 14(7), 316–323.
- Lotti, T., Mirone, V., Imbimbo, C., Corrado, F., Corrado, G., Garofalo, F. and Scaricabarozzi, I. (1993). Controlled clinical studies of nimesulide in the treatment of urogenital inflammation. *Drugs*, 46(1), 144–146.
- Lu, W., Melancon, M. P., Xiong, C., Huang, Q., Elliott, A., Song, S. and Wang, L. V. (2011). Effects of photoacoustic imaging and photothermal ablation therapy mediated by targeted hollow gold nanospheres in an orthotopic mouse xenograft model of glioma. *Cancer Research*, 71(19), 6116–6121.
- Lübbe, A. S., Alexiou, C. and Bergemann, C. (2001). Clinical applications of magnetic drug targeting. *Journal of Surgical Research*, 95(2), 200–206.
- Lübbe, A. S., Bergemann, C., Riess, H., Schriever, F., Reichardt, P., Possinger, K. and Gürtler, R. (1996). Clinical experiences with magnetic drug targeting: a phase I study with 4'-epidoxorubicin in 14 patients with advanced solid tumors. *Cancer Research*, 56(20), 4686–4693.
- Ma, J. and Jemal, A. (2013). *Breast Cancer Statistics*. New York: Springer.
- Madihah, B. A. G. (2011). *In vitro inhibition of human ovarian and colon cancer cell*

- lines using crude bromelain in ananas comosus juice*. M.Sc. Thesis Page 21, Universiti Teknologi Malaysia, Skudai.
- Maeda, H. (2001). The enhanced permeability and retention (EPR) effect in tumor vasculature: the key role of tumor-selective macromolecular drug targeting. *Advances in Enzyme Regulation*, 41(1), 189–207.
- Mahmoudi, M., Hosseinkhani, H., Hosseinkhani, M., Boutry, S., Simchi, A., Journeay, W. S. and Laurent, S. (2010). Magnetic resonance imaging tracking of stem cells in vivo using iron oxide nanoparticles as a tool for the advancement of clinical regenerative medicine. *Chemical Reviews*, 111(2), 253–280.
- Mahmoudi, M., Sahraian, M. A., Shokrgozar, M. A. and Laurent, S. (2011). Superparamagnetic iron oxide nanoparticles: promises for diagnosis and treatment of multiple sclerosis. *ACS Chemical Neuroscience*, 2(3), 118–140.
- Mahmoudi, M., Simchi, A. and Imani, M. (2010). Recent advances in surface engineering of superparamagnetic iron oxide nanoparticles for biomedical applications. *Journal of the Iranian Chemical Society*, 7(2), S1–S27.
- Mahmoudi, M., Simchi, A., Imani, M., Shokrgozar, M. A., Milani, A. S., Häfeli, U. O. and Stroeve, P. (2010). A new approach for the in vitro identification of the cytotoxicity of superparamagnetic iron oxide nanoparticles. *Colloids and Surfaces B: Biointerfaces*, 75(1), 300–309.
- Maingi, V., Kumar, M. V. S., and Maiti, P. K. (2012). PAMAM dendrimer–drug interactions: effect of pH on *the binding and release pattern*. *The Journal of Physical Chemistry B*, 116(14), 4370–4376.
- Makin, G. and Dive, C. (2001). Apoptosis and Cancer Chemotherapy. *Trends in Cell Biology*, 11(11), S22–S26.
- Mansoori, G. A., Brandenburg, K. S. and Shakeri-zadeh, A. (2010). A Comparative Study of Two Folate-Conjugated Gold Nanoparticles for Cancer Nanotechnology Applications. *Cancers*, 2(4), 1911–1928.
- Manuscript, A. (2012). Clearance Properties of Nano-sized Particles and Molecules as Imaging Agents: Considerations and Caveats. *Nanomedicine*, 3(5), 703–717.
- Marcucci, F. and Lefoulon, F. (2004). Active targeting with particulate drug carriers in tumor therapy: fundamentals and recent progress. *Drug Discovery Today*,

- 9(5), 219–228.
- Martinez, J. D., Parker, M. T., Fultz, K. E., Ignatenko, N. A. and Gerner, E. W. (2003). Molecular biology of cancer. *Burger's Medicinal Chemistry and Drug Discovery*, 1–50.
- Mashima, T. and Tsuruo, T. (2005). Defects of the Apoptotic Pathway as Therapeutic Target Against Cancer. *Drug Resistance Updates*, 8(6), 339–343.
- Massart, R. (1981). Preparation of aqueous magnetic liquids in alkaline and acidic media. *IEEE Transactions on Magnetics*, 17(2), 1247–1248.
- Matsumura, Y. and Maeda, H. (1986). A new concept for macromolecular therapeutics in cancer chemotherapy: mechanism of tumorotropic accumulation of proteins and the antitumor agent smancs. *Cancer Research*, 46(12 Part 1), 6387–6392.
- Maurer, H. R. (2001). Bromelain: biochemistry, pharmacology and medical use. *Cellular and Molecular Life Sciences CMLS*, 58(9), 1234–1245.
- McBain, S. C., Yiu, H. H., and Dobson, J. (2008). Magnetic nanoparticles for gene and drug delivery. *International journal of nanomedicine*, 3(2), 169.
- McCarthy, J. R. and Weissleder, R. (2008). Multifunctional magnetic nanoparticles for targeted imaging and therapy. *Advanced Drug Delivery Reviews*, 60(11), 1241–1251.
- Medintz, I. L., Uyeda, H. T., Goldman, E. R., Mattoussi, H. (2005). Quantum dot bioconjugates for imaging, labelling and sensing. *Nature Materials*, 4(6), 435–446.
- Melani, C., Figini, M., Nicosia, D., Luison, E., Ramakrishna, V., Casorati, G. and Colombo, M. P. (1998). Targeting of interleukin 2 to human ovarian carcinoma by fusion with a single-chain Fv of antifolate receptor antibody. *Cancer Research*, 58(18), 4146–4154.
- Mialovyts' k, O. A. (2002). Effect of phlogenzym in long-term treatment of patients with multiple sclerosis. *Likars' Ka sprava/Ministerstvo Okhorony Zdorov'ia Ukrainy*, (3-4), 109–113.
- Michalet, X., Pinaud, F. F., Bentolila, L. A., Tsay, J. M., Doose, S., Li, J. J. and Weiss, S. (2005). Quantum dots for live cells, in vivo imaging, and diagnostics. *Science*, 307(5709), 538–544.

- Miele, E., Spinelli, G. P., Miele, E., Tomao, F. and Tomao, S. (2009). Albumin-bound formulation of paclitaxel (Abraxane® ABI-007) in the treatment of breast cancer. *International Journal of Nanomedicine*, 4, PMC2720743.
- Minchinton, A. I. and Tannock, I. F. (2006). Drug penetration in solid tumours. *Nature Reviews Cancer*, 6(8), 583–592.
- Mishra, S., Webster, P. and Davis, M. E. (2004). PEGylation significantly affects cellular uptake and intracellular trafficking of non-viral gene delivery particles. *European Journal of Cell Biology*, 83(3), 97–111.
- Mody, V. V., Cox, A., Shah, S., Singh, A., Bevins, W., and Parihar, H. (2014). Magnetic nanoparticle drug delivery systems for targeting tumor. *Applied Nanoscience*, 4(4), 385-392.
- Mollarazi, E., Jalilian, A. R., Johari-daha, F. and Atyabi, F. (2015). Development of ¹⁵³Sm-folate-polyethyleneimine-conjugated chitosan nanoparticles for targeted therapy. *Journal of Labelled Compounds and Radiopharmaceuticals*, 58(8), 327–335.
- Moore, M. J., Goldstein, D., Hamm, J., Figer, A., Hecht, J. R., Gallinger and S., Wolff, R. A. (2007). Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: a phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *Journal of Clinical Oncology*, 25(15), 1960–1966.
- Morikawa, S., Baluk, P., Kaidoh, T., Haskell, A., Jain, R. K. and McDonald, D. M. (2002). Abnormalities in pericytes on blood vessels and endothelial sprouts in tumors. *The American Journal of Pathology*, 160(3), 985–1000.
- Mosmann, T. (1983). Rapid Colorimetric Assay for Cellular Growth and Survival : Application to Proliferation and Cytotoxicity Assays. *Journal of immunological methods*, 65(1-2), 55–63.
- Mulder, W. J. M., Strijkers, G. J., van Tilborg, G. A. F., Griffioen, A. W. and Nicolay, K. (2006). Lipid-based nanoparticles for contrast-enhanced MRI and molecular imaging. *NMR in Biomedicine*, 19(1), 142–164.
- Müller, C., Forrer, F., Schibli, R., Krenning, E. P. and de Jong, M. (2008). SPECT study of folate receptor-positive malignant and normal tissues in mice using a novel ^{99m}Tc-radiofolate. *Journal of Nuclear Medicine : Official Publication*,

Society of Nuclear Medicine, 49(2), 310–7.

- Munnier, E., Cohen-Jonathan, S., Hervé, K., Linassier, C., Soucé, M., Dubois, P. and Chourpa, I. (2010). Doxorubicin delivered to MCF-7 cancer cells by superparamagnetic iron oxide nanoparticles: effects on subcellular distribution and cytotoxicity. *Journal of Nanoparticle Research*, 13(3), 959–971.
- Nasiri, R., Almaki, J. H., Idris, A. B., Majid, F. A. A., Nasiri, M., Salouti, M. and Marvibaigi, M. (2016). In vitro evaluation of actively targetable superparamagnetic nanoparticles to the folate receptor positive cancer cells. *Materials Science and Engineering: C*, 69, 1147–1158.
- Nateghian, N., Goodarzi, N., Amini, M., Atyabi, F., Khorramizadeh, M. R. and Dinarvand, R. (2015). Biotin/Folate-decorated Human Serum Albumin Nanoparticles of Docetaxel: Comparison of Chemically Conjugated Nanostructures and Physically Loaded Nanoparticles for Targeting of Breast Cancer. *Chemical Biology & Drug Design*, 87(1), 69-82.
- Neuberger, T., Schöpf, B., Hofmann, H., Hofmann, M. and Von Rechenberg, B. (2005). Superparamagnetic nanoparticles for biomedical applications: possibilities and limitations of a new drug delivery system. *Journal of Magnetism and Magnetic Materials*, 293(1), 483–496.
- Nevozhay, D., Kańska, U., Budzyńska, R. and Boratyński, J. (2006). Current status of research on conjugates and related drug delivery systems in the treatment of cancer and other diseases. *Postepy Higieny I Medycyny Doswiadczalnej*, 61, 350–360.
- Ngadiman, N. H. A., Idris, A., Irfan, M., Kurniawan, D., Yusof, N. M. and Nasiri, R. (2015). γ -Fe₂O₃ nanoparticles filled polyvinyl alcohol as potential biomaterial for tissue engineering scaffold. *Journal of the Mechanical Behavior of Biomedical Materials*, 49, 90–104.
- Nicolas, J., Mura, S., Brambilla, D., Mackiewicz, N. and Couvreur, P. (2013). Design, functionalization strategies and biomedical applications of targeted biodegradable/biocompatible polymer-based nanocarriers for drug delivery. *Chemical Society Reviews*, 42(3), 1147–1235.
- Niemeyer, C. M. (2001). Nanoparticles, proteins, and nucleic acids: biotechnology meets materials science. *Angewandte Chemie International Edition*, 40(22),

4128–4158.

- Nigam, S., Barick, K. C. and Bahadur, D. (2010). Development of citrate-stabilized Fe₃O₄ nanoparticles : Conjugation and release of doxorubicin for therapeutic applications. *Journal of Magnetism and Magnetic Materials*, 323(2), 237–243.
- Nigam, S., Barick, K. C. and Bahadur, D. (2011a). Development of citrate-stabilized Fe₃O₄ nanoparticles: conjugation and release of doxorubicin for therapeutic applications. *Journal of Magnetism and Magnetic Materials*, 323(2), 237–243.
- Nigam, S., Barick, K. C. and Bahadur, D. (2011b). Development of citrate-stabilized Fe₃O₄ nanoparticles: Conjugation and release of doxorubicin for therapeutic applications. *Journal of Magnetism and Magnetic Materials*, 323(2), 237–243.
- Okon, E., Pouliquen, D., Okon, P., Kovaleva, Z. V., Stepanova, T. P., Lavit, S. G. and Jallet, P. (1994). Biodegradation of magnetite dextran nanoparticles in the rat. A histologic and biophysical study. *Laboratory investigation; a journal of technical methods and pathology*, 71(6), 895-903.
- Omar, Z. A. and Ibrahim Tamin, N. S. (2011). *NCR Report 2007*. Ministry of Health: Malaysia.
- Orsini, R.A. Plastic Surgery Educational Foundation Technology Assessment Committee. (2006). *Bromelain: Plastic and Reconstructive Surgery*, 118(7), 1640-1644.
- Pan, L., Chai, H. B. and Kinghorn, A. D. (2012). Discovery of New Anticancer Agents from Higher Plants. *Frontiers in Bioscience*, 4, 142–56.
- Pankhurst, Q. A., Connolly, J., Jones, S. K. and Dobson, J. J. (2003). Applications of magnetic nanoparticles in biomedicine. *Journal of Physics D: Applied Physics*, 36(13), R167.
- Panyam, J. and Labhasetwar, V. (2003). Biodegradable nanoparticles for drug and gene delivery to cells and tissue. *Advanced Drug Delivery Reviews*, 55(3), 329–347.
- Panyam, J. and Labhasetwar, V. (2004). Targeting intracellular targets. *Current Drug Delivery*, 1(3), 235–247.
- Park, J., Dvoracek, C., Lee, K. H., Galloway, J. F., Bhang, H. C., Pomper, M. G. and Searson, P. C. (2011). CuInSe/ZnS core/shell NIR quantum dots for biomedical imaging. *Small*, 7(22), 3148–3152.

- Park, K. S., Tae, J., Choi, B., Kim, Y. S., Moon, C., Kim, S. H. and Park, J. (2010). Characterization, in vitro cytotoxicity assessment, and in vivo visualization of multimodal, RITC-labeled, silica-coated magnetic nanoparticles for labeling human cord blood-derived mesenchymal stem cells. *Nanomedicine: Nanotechnology, Biology and Medicine*, 6(2), 263–276.
- Parodi, A., Haddix, S. G., Taghipour, N., Scaria, S., Taraballi, F., Cevenini, A. and Khaled, S. Z. (2014). Bromelain surface modification increases the diffusion of silica nanoparticles in the tumor extracellular matrix. *ACS Nano*, 8(10), 9874–9883.
- Pastan, I., Hassan, R., FitzGerald, D. J. and Kreitman, R. J. (2006). Immunotoxin therapy of cancer. *Nature Reviews Cancer*, 6(7), 559–565.
- Patel, N., Davies, M. C., Hartshorne, M., Heaton, R. J., Roberts, C. J., Tendler, S. J. B. and Williams, P. M. (1997). Immobilization of protein molecules onto homogeneous and mixed carboxylate-terminated self-assembled monolayers. *Langmuir*, 13(24), 6485–6490.
- Patel, B. K., Parikh, R. H., and Aboti, P. S. (2013). Development of oral sustained release rifampicin loaded chitosan nanoparticles by design of experiment. *Journal of drug delivery*, 2013, 1-10.
- Paulo, C. S. O., Pires das Neves, R. and Ferreira, L. S. (2011). Nanoparticles for intracellular-targeted drug delivery. *Nanotechnology*, 22(49), 494002.
- Peer, D., Karp, J. M., Hong, S., Farokhzad, O. C., Margalit, R. and Langer, R. (2007). Nanocarriers as an emerging platform for cancer therapy. *Nature Nanotechnology*, 2(12), 751–760.
- Petryayeva, E., Algar, W. R. and Medintz, I. L. (2013). Quantum dots in bioanalysis: a review of applications across various platforms for fluorescence spectroscopy and imaging. *Applied Spectroscopy*, 67(3), 215–252.
- Pillai, K., Akhter, J., Ehteda, A., Badar, S., Chua, T. C. and Morris, D. L. (2013). Glycobiology Anti-Tumour and Chemosensitising Effect of a Combination of Bromelain + N-Acetyl Cysteine with Cisplatin or 5-Fu on Malignant Peritoneal Mesothelioma Cells. *Journal of Glycobiology*, S1-005.
- Prato, M., Kostarelos, K. and Bianco, A. (2007). Functionalized carbon nanotubes in drug design and discovery. *Accounts of Chemical Research*, 41(1), 60–68.

- Purushotham, S. and Ramanujan, R. V. (2010). Thermoresponsive magnetic composite nanomaterials for multimodal cancer therapy. *Acta Biomaterialia*, 6(2), 502–10.
- Qi, L. and Gao, X. (2008). Emerging application of quantum dots for drug delivery and therapy. *Expert Opinion on Drug Delivery*, 5(3), 263–267.
- Răcuciu, M., Creangă, D.E. and Airinei, A. (2006). Citric-acid-coated magnetite nanoparticles for biological applications. *The European Physical Journal E*, 21(2), 117-121.
- Radwan, S. H. and Azzazy, H. M. E. (2009). Gold nanoparticles for molecular diagnostics. *Expert Review of Molecular Diagnostics*, 9(5), 511–524.
- Razjouyan, J., Zolata, H., Khayat, O., Nowshiravan, F., Shadanpour, N. and Mohammadnia, M. (2015). Synthesis and evaluation of radiolabeled, folic acid-PEG conjugated, amino silane coated magnetic nanoparticles in tumor bearing Balb/C mice. *Nukleonika*, 60(3), 479–502.
- Rettig, W. J., Garin-Chesa, P., Beresford, H. R., Oettgen, H. F., Melamed, M. R. and Old, L. J. (1988). Cell-surface glycoproteins of human sarcomas: differential expression in normal and malignant tissues and cultured cells. *Proceedings of the National Academy of Sciences*, 85(9), 3110–3114.
- Ross, J. F., Chaudhuri, P. K. and Ratnam, M. (1994). Differential regulation of folate receptor isoforms in normal and malignant tissues in vivo and in established cell lines. Physiologic and clinical implications. *Cancer*, 73(9), 2432–2443.
- Ross, J. S. and Fletcher, J. A. (1998). The HER-2/neu Oncogene in Breast Cancer: Prognostic Factor, Predictive Factor, and Target for Therapy. *Stem Cells*, 16(6), 413–428.
- Safarzadeh, E., Shotorbani, S. S. and Baradaran, B. (2014). Herbal Medicine as Inducers of Apoptosis in Cancer Treatment. *Advanced Pharmaceutical Bulletin*, 4(1), 421–427.
- Saltz, L. B., Clarke, S., Díaz-Rubio, E., Scheithauer, W., Figer, A., Wong, R. and Rivera, F. (2008). Bevacizumab in combination with oxaliplatin-based chemotherapy as first-line therapy in metastatic colorectal cancer: a randomized phase III study. *Journal of Clinical Oncology*, 26(12), 2013–2019.
- Salvatorelli, E., Menna, P., Cascegnà, S., Liberi, G., Calafiore, A. M., Gianni, L. and

- Minotti, G. (2006). Paclitaxel and docetaxel stimulation of doxorubicinol formation in the human heart: implications for cardiotoxicity of doxorubicin-taxane chemotherapies. *Journal of Pharmacology and Experimental Therapeutics*, 318(1), 424–433.
- Santabarbara, G., Maione, P., Rossi, A. and Gridelli, C. (2016). Pharmacotherapeutic options for treating adverse effects of Cisplatin chemotherapy. *Expert Opinion on Pharmacotherapy*, 17(4), 561–570.
- Santra, S., Tapeç, R., Theodoropoulou, N., Dobson, J., Hebard, A. and Tan, W. (2001). Synthesis and characterization of silica-coated iron oxide nanoparticles in microemulsion: the effect of nonionic surfactants. *Langmuir*, 17(10), 2900–2906.
- Saraswathy, A., Nazeer, S. S., Nimi, N., Arumugam, S., Shenoy, S. J. and Jayasree, R. S. (2014). Synthesis and characterization of dextran stabilized superparamagnetic iron oxide nanoparticles for in vivo MR imaging of liver fibrosis. *Carbohydrate Polymers*, 101, 760–768.
- Saravanan, B. C., Sreekumar, C., Bansal, G. C., Ray, D., Rao, J. R. and Mishra, A. K. (2003). A rapid MTT colorimetric assay to assess the proliferative index of two Indian strains of *Theileria annulata*. *Veterinary Parasitology*, 113(3), 211–216.
- Saul, J. M., Annapragada, A., Natarajan, J. V. and Bellamkonda, R. V. (2003). Controlled targeting of liposomal doxorubicin via the folate receptor in vitro. *Journal of Controlled Release*, 92(1-2), 49–67.
- Scher, H. I., Beer, T. M., Higano, C. S., Anand, A., Taplin, M.-E., Efstathiou, E. and Alumkal, J. (2010). Antitumour activity of MDV3100 in castration-resistant prostate cancer: a phase 1–2 study. *The Lancet*, 375(9724), 1437–1446.
- Schiller, J. H., Larson, T., Ou, S. I., Limentani, S. A., Sandler, A. B., Vokes, E. E. and Olszanski, A. J. (2007). Efficacy and safety of axitinib (AG-013736; AG) in patients (pts) with advanced non-small cell lung cancer (NSCLC): a phase II trial. *ASCO Annual Meeting Proceedings*, 25(18), 7507.
- Schoeberl, B., Faber, A. C., Li, D., Liang, M. C., Crosby, K., Onsum, M. and Nie, L. (2010). An ErbB3 antibody, MM-121, is active in cancers with ligand-dependent activation. *Cancer Research*, 70(6), 2485–2494.

- Shah, D. A., Kwon, S. J., Bale, S. S., Banerjee, A., Dordick, J. S. and Kane, R. S. (2011). Regulation of stem cell signaling by nanoparticle-mediated intracellular protein delivery. *Biomaterials*, 32(12), 3210–3219.
- Shahidi, S. K., Kundra, N. H. T. M., Shanbag, P. and Schiess, G. V. D. W. (2002). Efficacy and Safety of Phlogenzym-A Protease Formulation, in Sepsis in Children. *Journal of the Association of Physicians of India*, 50, 527–531.
- Sharifi, S., Behzadi, S., Laurent, S., Forrest, M. L., Stroeve, P. and Mahmoudi, M. (2012). Toxicity of nanomaterials. *Chemical Society Reviews*, 41(6), 2323–2343.
- Shi, J., Xiao, Z., Kamaly, N. and Farokhzad, O. C. (2011). Self-assembled targeted nanoparticles: evolution of technologies and bench to bedside translation. *Accounts of Chemical Research*, 44(10), 1123–1134.
- Skotland, T., Iversen, T. G. and Sandvig, K. (2010). New metal-based nanoparticles for intravenous use: requirements for clinical success with focus on medical imaging. *Nanomedicine: Nanotechnology, Biology and Medicine*, 6(6), 730–737.
- Singh, A. and Sahoo, S. K. (2014). Magnetic nanoparticles: a novel platform for cancer theranostics. *Drug Discovery Today*, 19(4), 474–481.
- Sisson, T. A. T. A. T. on R. S. J. C., Freitas, J., McDougall, I. R., Dauer, L. T., Hurley, J. R., Brierley, J. D. and Wexler, J. A. (2011). Radiation safety in the treatment of patients with thyroid diseases by radioiodine 131I: practice recommendations of the American Thyroid Association. *Thyroid*, 21(4), 335–346.
- Skouras, A., Mourtas, S., Markoutsas, E., De Goltstein, M.-C., Wallon, C., Catoen, S. and Antimisiaris, S. G. (2011). Magnetoliposomes with high USPIO entrapping efficiency, stability and magnetic properties. *Nanomedicine: Nanotechnology, Biology and Medicine*, 7(5), 572–579.
- Slowing, I. I., Vivero-Escoto, J. L., Wu, C. W. and Lin, V. S. Y. (2008). Mesoporous silica nanoparticles as controlled release drug delivery and gene transfection carriers. *Advanced drug delivery reviews*, 60(11), 1278–1288.
- Song, H. T., Choi, J., Huh, Y. M., Kim, S., Jun, Y., Suh, J. S. and Cheon, J. (2005). Surface modulation of magnetic nanocrystals in the development of highly

- efficient magnetic resonance probes for intracellular labeling. *Journal of the American Chemical Society*, 127(28), 9992–9993.
- Steeg, P. S. (2006). Tumor metastasis: mechanistic insights and clinical challenges. *Nature Medicine*, 12(8), 895–904.
- Steitz, B., Hofmann, H., Kamau, S. W., Hassa, P. O., Hottiger, M. O., von Rechenberg, B. and Petri-Fink, A. (2007). Characterization of PEI-coated superparamagnetic iron oxide nanoparticles for transfection: Size distribution, colloidal properties and DNA interaction. *Journal of Magnetism and Magnetic Materials*, 311(1), 300–305.
- Stewart, B. W. (1994). Mechanisms of apoptosis: integration of genetic, biochemical, and cellular indicators. *Journal of the National Cancer Institute*, 86(17), 1286–1296.
- Strable, E., Bulte, J. W. M., Moskowitz, B., Vivekanandan, K., Allen, M. and Douglas, T. (2001). Synthesis and characterization of soluble iron oxide-dendrimer composites. *Chemistry of Materials*, 13(6), 2201–2209.
- Strober, W. (2001). Trypan blue exclusion test of cell viability. *Current Protocols in Immunology*, A3–B.
- Sudhakar, A. (2009). History of cancer, ancient and modern treatment methods. *Journal of cancer science & therapy*, 1(2), 1.
- Sudimack, J. and Lee, R. J. (2000). Targeted drug delivery via the folate receptor. *Advanced Drug Delivery Reviews*, 41(2), 147–162.
- Sun, C., Sze, R. and Zhang, M. (2006). Folic acid-PEG conjugated superparamagnetic nanoparticles for targeted cellular uptake and detection by MRI. *Journal of Biomedical Materials Research Part A*, 78(3), 550–557.
- Suri, S. S., Fenniri, H. and Singh, B. (2007). Nanotechnology-based drug delivery systems. *Journal of Occupational Medicine and Toxicology*, 2(1), 1.
- Szekeres, M., Tóth, I. Y., Illés, E., Hajdú, A. and Zupkó, I. (2013). Chemical and Colloidal Stability of Carboxylated Core-Shell Magnetite Nanoparticles Designed for Biomedical Applications. *International journal of molecular sciences*, 14(7), 14550–14574.
- Taussig, S. J. and Batkin, S. (1988). Bromelain, the enzyme complex of pineapple (*Ananas comosus*) and its clinical application. An update. *Journal of*

- Ethnopharmacology*, 22(2), 191–203.
- Taussig, S. J., Szekerczes, J. and Batkin, S. (1985). Inhibition of tumour growth in vitro by bromelain, an extract of the pineapple plant (*Ananas comosus*). *Planta Medica*, 51(06), 538–539.
- The side effects of chemotherapy* 2009. Available from: <<http://www.lapcsg.org/articles--blog/category/support%20tools>>. [9 April 2014]
- Timm, D. M., Chen, J., Sing, D., Gage, J. A., Haisler, W. L., Neeley, S. K. and Tseng, H. (2013). A high-throughput three-dimensional cell migration assay for toxicity screening with mobile device-based macroscopic image analysis. *Scientific reports*, 3, 3000.
- Tomitaka, A., Koshi, T., Hatsugai, S., Yamada, T. and Takemura, Y. (2011). Magnetic characterization of surface-coated magnetic nanoparticles for biomedical application. *Journal of Magnetism and Magnetic Materials*, 323(10), 1398–1403.
- Tomlinson, I. D., Gussin, H. A., Little, D. M., Warnement, M. R., Qian, H., Pepperberg, D. R. and Rosenthal, S. J. (2008). Imaging GABAC receptors with ligand-conjugated quantum dots. *BioMed Research International*, 2007.
- Torchilin, V. P. (2000). Drug targeting. *European Journal of Pharmaceutical Sciences*, 11, S81–S91.
- Torchilin, V. P. (2006). *Nanoparticulates as drug carriers*. London: Imperial college press.
- Torchilin, V. P. (2010). Passive and active drug targeting: drug delivery to tumors as an example. *Handbook of Experimental Pharmacology*. (pp. 5–53). London :Springer.
- Torre, L. A., Bray, F., Siegel, R. L., Ferlay, J., Lortet-Tieulent, J. and Jemal, A. (2015). Global cancer statistics, 2012. *CA: A Cancer Journal for Clinicians*, 65(2), 87–108.
- Tutt, A., Robson, M., Garber, J. E., Domchek, S. M., Audeh, M. W., Weitzel, J. N. and Schmutzler, R. K. (2010). Oral poly (ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and advanced breast cancer: a proof-of-concept trial. *The Lancet*, 376(9737), 235–244.

- Tysnes, B. B., Maurer, H. R., Porwol, T., Probst, B., Bjerkvig, R. and Hoover, F. (2001). Bromelain reversibly inhibits invasive properties of glioma cells. *Neoplasia*, 3(6), 469–79.
- Van Blaaderen, A. and Vrij, A. (1992). Synthesis and characterization of colloidal dispersions of fluorescent, monodisperse silica spheres. *Langmuir*, 8(12), 2921–2931.
- Van, C. S. and Van, D. B. (2002). Morphological and Biochemical Aspects of Apoptosis, Oncosis and Necrosis. *Anatomia, Histologia, Embryologia*, 31(4), 214–223.
- Van Cutsem, E., Köhne, C.-H., Láng, I., Folprecht, G., Nowacki, M. P., Cascinu, S. and Tejpar, S. (2011). Cetuximab plus irinotecan, fluorouracil, and leucovorin as first-line treatment for metastatic colorectal cancer: updated analysis of overall survival according to tumor KRAS and BRAF mutation status. *Journal of Clinical Oncology*, JCO–2010.
- Van De Ven, A. L., Kim, P., Fakhoury, J. R., Adriani, G., Schmulen, J., Moloney, P. and Yun, S. H. (2012). Rapid tumoritropic accumulation of systemically injected plateloid particles and their biodistribution. *Journal of Controlled Release*, 158(1), 148–155.
- van Zijl, F., Krupitza, G. and Mikulits, W. (2011). Initial steps of metastasis: cell invasion and endothelial transmigration. *Mutation Research/Reviews in Mutation Research*, 728(1), 23–34.
- Varadan, V. K., Chen, L. and Xie, J. (2008). *Nanomedicine: design and applications of magnetic nanomaterials, nanosensors and nanosystems*. United Kingdom: John Wiley & Sons.
- Varshosaz, J., Ghasemi, S. and Behdadfar, B. (2013). Use of Magnetic Folate-Dextran-Retinoic Acid Micelles for Dual Targeting of Doxorubicin in Breast Cancer. *BioMed Research International*, 2013, 1–16.
- Vayssieres, L., Chanéac, C., Tronc, E. and Jolivet, J. P. (1998). Size tailoring of magnetite particles formed by aqueous precipitation: An example of thermodynamic stability of nanometric oxide particles. *Journal of Colloid and Interface Science*, 205(2), 205–212.
- Veiseh, O., Gunn, J. W. and Zhang, M. (2010). Design and fabrication of magnetic

- nanoparticles for targeted drug delivery and imaging. *Advanced Drug Delivery Reviews*, 62(3), 284–304.
- Vigderman, L., and Zubarev, E. R. (2013). Therapeutic platforms based on gold nanoparticles and their covalent conjugates with drug molecules. *Advanced drug delivery reviews*, 65(5), 663-676.
- Wahajuddin, S. A. (2012). Superparamagnetic iron oxide nanoparticles: magnetic nanoplatforms as drug carriers. *International Journal of Nanomedicine*, 7, 3445.
- Walter, G. A., Cahill, K. S., Huard, J., Feng, H., Douglas, T., Sweeney, H. L. and Bulte, J. W. M. (2004). Noninvasive monitoring of stem cell transfer for muscle disorders. *Magnetic Resonance in Medicine*, 51(2), 273–277.
- Wang, H., Castner, D. G., Ratner, B. D. and Jiang, S. (2004). Probing the orientation of surface-immobilized immunoglobulin G by time-of-flight secondary ion mass spectrometry. *Langmuir*, 20(5), 1877–1887.
- Wang, J., Chen, B., Chen, J., Cai, X., Xia, G., Liu, R. and Wang, X. (2011). Synthesis and antitumor efficacy of daunorubicin-loaded magnetic nanoparticles. *International Journal of Nanomedicine*, 6, 203–11.
- Wang, J., Chen, B., Cheng, J., Cai, X., Xia, G., Liu, R. and Wang, X. (2011). Apoptotic mechanism of human leukemia K562/A02 cells induced by magnetic iron oxide nanoparticles co-loaded with daunorubicin and 5-bromotetrandrin. *International Journal of Nanomedicine*, 6, 1027–34.
- Wang, S.J., Zheng, C.J., Peng, C., Zhang, H., Jiang, Y.P., Han, T. and Qin, L.P. (2013). Plants and Cervical Cancer: an Overview. *Expert Opinion on Investigational Drugs*, 22(9), 1133–56.
- Wang, T., Wu, H., Wang, W., Lin, F., Lou, P., Shieh, M. and Young, T. (2007). The development of magnetic degradable DP-Bioglass for hyperthermia cancer therapy. *Journal of Biomedical Materials Research Part A*, 83(3), 828–837.
- Wang, Y., Bansal, V., Zelikin, A. N. and Caruso, F. (2008). Templated synthesis of single-component polymer capsules and their application in drug delivery. *Nano Letters*, 8(6), 1741–1745.
- Wang, Y., Zhao, Q., Han, N., Bai, L., Li, J., Liu, J. and Wang, S. (2015). Mesoporous silica nanoparticles in drug delivery and biomedical applications. *Nanomedicine: Nanotechnology, Biology and Medicine*, 11(2),

313-327.

- Wang, Z., Zhou, C., Xia, J., Via, B., Xia, Y., Zhang and F., Xia, L. (2013). Colloids and Surfaces B: Biointerfaces Fabrication and characterization of a triple functionalization of graphene oxide with Fe₃O₄, folic acid and doxorubicin as dual-targeted drug nanocarrier. *Colloids and Surfaces B: Biointerfaces*, 106, 60–65.
- Weitman, S. D., Lark, R. H., Coney, L. R., Fort, D. W., Frasca, V., Zurawski, V. R. and Kamen, B. A. (1992). Distribution of the folate receptor GP38 in normal and malignant cell lines and tissues. *Cancer Research*, 52(12), 3396–3401.
- Wilt, T. J., MacDonald, R., Rutks, I., Shamliyan, T. A., Taylor, B. C. and Kane, R. L. (2008). Systematic review: comparative effectiveness and harms of treatments for clinically localized prostate cancer. *Annals of Internal Medicine*, 148(6), 435–448.
- Woo, K., Hong, J., Choi, S., Lee, H. W., Ahn, J. P., Kim, C. S. and Lee, S. W. (2004). Easy synthesis and magnetic properties of iron oxide nanoparticles. *Chemistry of Materials*, 16(14), 2814–2818.
- Wooster, R. and Weber, B. L. (2003). Breast and ovarian cancer. *New England Journal of Medicine*, 348(23), 2339–2347.
- Wu, W., He, Q. and Jiang, C. (2009). Magnetic iron oxide nanoparticles: synthesis and surface functionalization strategies. *Nanoscale Research Letters*, 3(11), 397.
- Xie, J., Huang, J., Li, X., Sun, S. and Chen, X. (2009). Iron oxide nanoparticle platform for biomedical applications. *Current Medicinal Chemistry*, 16(10), 1278–1294.
- Xiong, X. B., Huang, Y., LU, W. L., Zhang, X., Zhang, H., Nagai and T., Zhang, Q. (2005). Intracellular delivery of doxorubicin with RGD-modified sterically stabilized liposomes for an improved antitumor efficacy: In vitro and in vivo. *Journal of pharmaceutical sciences*, 94(8), 1782-1793.
- Xu, C., Xu, K., Gu, H., Zhong, X., Guo, Z., Zheng, R. and Xu, B. (2004). Nitriiotriacetic acid-modified magnetic nanoparticles as a general agent to bind histidine-tagged proteins. *Journal of the American Chemical Society*, 126(11), 3392–3393.

- Yabbarov, N. G., Posypanova, G. A., Vorontsov, E. A., Popova, O. N. and Severin, E. S. (2013). Targeted delivery of doxorubicin: drug delivery system based on PAMAM dendrimers. *Biochemistry (Moscow)*, 78(8), 884-894.
- Yang, J., Lee, T. I., Lee, J., Lim, E. K., Hyung, W., Lee, C.H. and Huh, Y. M. (2007). Synthesis of ultrasensitive magnetic resonance contrast agents for cancer imaging using PEG-fatty acid. *Chemistry of Materials*, 19(16), 3870–3876.
- Yellepeddi, V. K., Kumar, A., and Palakurthi, S. (2009). Biotinylated poly (amido) amine (PAMAM) dendrimers as carriers for drug delivery to ovarian cancer cells in vitro. *Anticancer research*, 29(8), 2933-2943.
- Yoo, M. K., Park, I. K., Lim, H. T., Lee, S. J., Jiang, H. L., Kim, Y. K. and Cho, C. S. (2012). Folate-PEG-superparamagnetic iron oxide nanoparticles for lung cancer imaging. *Acta Biomaterialia*, 8(8), 3005–3013.
- Yu, B., Mao, Y., Bai, L. Y., Herman, S. E. M., Wang, X., Ramanunni, A. and Chan, K. K. (2013). Targeted nanoparticle delivery overcomes off-target immunostimulatory effects of oligonucleotides and improves therapeutic efficacy in chronic lymphocytic leukemia. *Blood*, 121(1), 136–147.
- Yuan, F., Dellian, M., Fukumura, D., Leunig, M., Berk, D. A., Torchilin, V. P. and Jain, R. K. (1995). Vascular permeability in a human tumor xenograft: molecular size dependence and cutoff size. *Cancer Research*, 55(17), 3752–3756.
- Zänker, K. S. (2001). The use of systemic enzyme therapy in oncology. *Cancer Chemotherapy and Pharmacology*, 47(7), S1–S3.
- Zavadova, E., Desser, L. and Mohr, T. (1995). Stimulation of reactive oxygen species production and cytotoxicity in human neutrophils in vitro and after oral administration of a polyenzyme preparation. *Cancer Biotherapy & Radiopharmaceuticals*, 10(2), 147–152.
- Zhang, Y., Kohler, N. and Zhang, M. (2002). Surface modification of superparamagnetic magnetite nanoparticles and their intracellular uptake. *Biomaterials*, 23(7), 1553–1561.
- Zhao, Jiangqi, Canhui L., Xu H., Xiaofang Z., Wei Z., and Ximu Z. (2015). Polyethylenimine-grafted cellulose nanofibril aerogels as versatile vehicles for