

PRODUCTION OF VIOLACEIN NANOPARTICLES VIA SONICATION  
TECHNIQUE WITH THE AID OF SURFACTANTS AS STABILIZER

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## DEDICATION

*This thesis is dedicated to my beloved father and mother, Mohd Hamzah bin Hassan and Norimah binti Salatin who have been very supportive through thick and thin for this meaningful two years. Thanks for all the prayers, advices and guidance.*

*Special thanks to all my wonderful friends who brighten my life with all the joys, sadness, smiles and laughters. This friendship has been very great experience in my life.*

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## ABSTRACT

Violacein, a violet pigment produced from *Chromobacterium violaceum* UTM5, has gained interest due to its biodegradability and pharmacological properties. However, its high production cost and limited solubility in water have become the major stumbling blocks for the pigment to be applied in different industries. In this study, liquid pineapple waste was used as an alternative inexpensive growth medium for bacteria cultivation instead of expensive synthetic nutrient broth, thus reducing the production cost of this pigment. The cultivation of *C. violaceum* in 50 L bioreactor gave a crude yield of  $11846 \pm 925 \text{ mg L}^{-1}$ , which was comparable to the yield obtained using commercial growth medium. The crude pigment was successfully extracted using ethyl acetate. The presence of violacein, the major active compound of the crude pigment, was confirmed using high performance liquid chromatography (HPLC), Fourier transform infrared spectroscopy (FTIR) and ultraviolet-visible spectrophotometry (UV-Vis). Thermal gravimetric analysis was used to determine crystallinity and thermal degradation while Zetasizer analyzer was used to identify the isoelectric point, stability at various pHs, and particle size of violacein. Violacein nanoparticles were produced via sonication technique, with the aid of surfactants (Tween 80, Triton X-100, sodium dodecyl sulfate and dodecyltrimethylammonium bromide) as solubilizing and stabilizing agent, to address the violacein's poor solubility in water. The violacein nanoparticles were characterized using UV-Vis spectrophotometry, FTIR, thermal analysis and Zetasizer analysis. Water soluble violacein nanoparticles were produced at surfactant concentration greater than its critical micelle concentration, as indicated by FTIR. Zetasizer analysis showed the smallest violacein nanoparticle, which was  $131.5 \pm 2.001 \text{ nm}$ , with polydispersity index (PDI) of  $0.180 \pm 0.018$ , which indicated a monodispersed violacein nanoparticle distribution. The thermal analysis showed that violacein nanoparticles were in amorphous state and stable upon dispersion in water, with a zeta potential of  $-49.8 \pm 3.49 \text{ mV}$ . The violacein nanoparticles have better solubility than the crude violacein pigment. The solubilized violacein nanoparticles remained well-dispersed upon storage in 28 days at different temperatures. In addition, the violet color of the violacein nanoparticles was maintained at pH range of 3 to 11, temperatures of up to  $60^\circ\text{C}$ , and under dark condition, despite its nanoscale size. Higher degradation rate was observed at high temperature and upon light illumination, with  $k = 6.51 \times 10^{-3} \text{ h}^{-1}$ ,  $t_{1/2} = 148 \text{ h}$  and  $k = 6.75 \times 10^{-4} \text{ h}^{-1}$ ,  $t_{1/2} = 1027 \text{ h}$ , respectively, following the first-order kinetics. In conclusion, this study confirmed the feasibility of using liquid pineapple waste as cheap growth medium for cultivation of *C. violaceum* UTM5 in pilot scale (50-L bioreactor) while production of water-soluble violacein nanoparticles via sonication method with the aid of surfactants as stabilizers would increase its usefulness, especially in pharmaceutical industry.

## ABSTRAK

Violasin, pigmen ungu yang terhasil daripada *Chromobacterium violaceum* UTM5, telah menarik banyak perhatian kerana sifat keterbiodegradan dan farmakologinya. Walau bagaimanapun, kos penghasilan yang tinggi dan keterlarutannya dalam air yang terhad menjadi penghalang utama untuk pigmen ini digunakan dalam pelbagai industri. Dalam kajian ini, sisa nenas cecair telah digunakan sebagai medium pertumbuhan alternatif yang murah bagi pemeliharaan bakteria untuk menggantikan kaldu nutrien sintetik yang mahal, seterusnya mengurangkan kos pengeluaran pigmen ini. Pemeliharaan *C. violaceum* di dalam bioreaktor bersaiz 50 L menghasilkan pigmen mentah sebanyak  $11846 \pm 925 \text{ mg L}^{-1}$ , yang setanding dengan hasil yang diperolehi menggunakan medium pertumbuhan komersial. Pigmen mentah telah berjaya diekstrak menggunakan etil asetat. Kehadiran violasin, komponen aktif utama di dalam pigmen mentah, telah disahkan menggunakan kromatografi cecair berprestasi tinggi (HPLC), spektroskopi inframerah transformasi Fourier (FTIR) dan spektrofotometri ultra ungu-cahaya nampak (UV-Vis). Analisis gravimetri terma telah digunakan untuk menentukan kehabluran dan degradasi haba manakala penganalisis Zetasizer telah digunakan untuk mengenalpasti titik isoelektrik, kestabilan pada pelbagai pH, dan saiz zarah violasin. Nanopartikel violasin telah dihasilkan menggunakan teknik sonikasi, dengan bantuan beberapa surfaktan (Tween 80, Triton X-100, sodium dodekil sulfat dan dodekiltrimetilammonium bromida) sebagai agen pelarut dan penstabil untuk menangani keterlarutan violasin yang rendah dalam air. Nanopartikel violasin telah dicirikan menggunakan spektrofotometri UV-vis, FTIR, analisis terma dan analisis Zetasizer. Nanopartikel violasin yang mudah larut dalam air telah berjaya dihasilkan pada kepekatan surfaktan melebihi kepekatan kritikal misel, seperti yang ditunjukkan oleh FTIR. Analisis Zetasizer menunjukkan saiz nanopartikel violasin yang terkecil iaitu  $131.5 \pm 2.001 \text{ nm}$ , dengan indeks kepoliserakan (PDI)  $0.180 \pm 0.018$ , yang menunjukkan taburan nanopartikel violasin yang sekata. Analisis terma menunjukkan nanopartikel violasin berada dalam keadaan amorfus dan stabil apabila terserak di dalam air dengan potensi zeta  $-49.8 \pm 3.49 \text{ mV}$ . Nanopartikel violasin mempunyai keterlarutan yang lebih baik berbanding pigmen violasin mentah. Nanopartikel violasin yang larut ini kekal terserak apabila disimpan selama 28 hari pada suhu yang berbeza. Tambahan pula, warna ungu nanopartikel violasin masih kekal pada julat pH antara 3 dengan 11, suhu sehingga  $60^\circ\text{C}$ , dan dalam keadaan gelap walaupun saiznya berskala nano. Kadar degradasi violasin yang lebih tinggi telah dilihat pada suhu tinggi dan apabila terdedah pada cahaya, masing-masing dengan  $k = 6.51 \times 10^{-3} \text{ h}^{-1}$ ,  $t_{1/2} = 148 \text{ h}$  dan  $k = 6.75 \times 10^{-4} \text{ h}^{-1}$ ,  $t_{1/2} = 1027 \text{ h}$ , mengikut kinetik tertib pertama. Kesimpulannya, kajian ini mengesahkan kebolehlaksanaan dalam menggunakan sisa nenas cecair sebagai medium pertumbuhan yang murah untuk pembiakan *C. violaceum* UTM5 pada skala perintis (50-L bioreaktor) manakala penghasilan nanopartikel violasin yang mudah larut dalam air menggunakan teknik sonikasi dengan bantuan surfaktan sebagai penstabil akan meningkatkan kegunaannya terutamanya dalam industri farmasi.

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**LIST OF SYMBOLS AND ABBREVIATIONS**

ATR-FTIR	: Attenuated Total Reflectance - Fourier Transform Infrared Spectroscopy
CMC	: Critical micelle concentration
HPLC	: High-performance liquid chromatography
LPW	: Liquid pineapple waste
MHA	: Muller-Hinton agar
MHB	: Muller-Hinton broth
NA	: Nutrient agar
NB	: Nutrient broth
PDI	: Polydispersity index
TLC	: Thin layer chromatography
UV-vis	: UV-visible spectroscopy
mg	: Milligram
mg L <sup>-1</sup>	: Milligram per litre
°C	: Degree celcius
% w/v	: Percentage of weight in 100 mL of solvent/solution
% v/v	: Percentage of volume in 100 mL of total solvent/solution

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## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of the Study

In 2010, tartrazine, quinoline yellow and carmoisine are the synthetic colorants that are banned in the United Kingdom and European Union as they trigger hyperactive behavior amongst children (Fusaro, 2010). Increase of awareness in regard to the danger of artificial (synthetic) colorants to human safety and environment leads to the increase in the use of natural colorants, known as biological colorants. These pigment, extracted from flora and fauna are found to be non-toxic, non-carcinogenic, and biodegradable (Venil *et al.*, 2013). On top of that, pharmacological properties exhibited by natural colorants have better advantages over synthetic pigments. For example, chlorophylls found in green plants exhibit anticancer properties as they can bind with cancer-causing chemicals to form complex structure, thus minimizing the absorption of potential carcinogens via gastrointestinal tract (İnanç, 2011).

Besides extracting the pigments from animals and plants, microbial pigments are also chosen due to their wide strain selection, shorter fermentation period, gene manipulability, lesser downstream processing (involves simple liquid-liquid extraction step) and cheaper growth medium availability (Venil *et al.*, 2013; Tuli *et al.*, 2015). Also, microorganisms can produce unique pigments such as violacein, prodigiosin and flexirubin, which are non-synthesizable by animals and plants. For example, bacterial strain of genus *Chromobacterium* is known for its ability to produce violacein pigment, with pharmacological properties as an antioxidant, and serves as

antimicrobial, antiprotozoal, and antipyretic compound (Durán *et al.*, 2016). The multi-resistant *S. aureus* (MRSA) poses a challenge in dealing with antibiotics due to multidrug resistance behavior. The use of violacein to treat MRSA has been reported by Aruldass *et al.* (2015). As a colorant, the intense violet exhibited by violacein, even at low concentration, is useful to formulate solvent-based ink in plastic application (Venil *et al.*, 2017; Durán *et al.*, 2007). In addition, violacein has been tested and the findings show its application as food colorant in yogurt and jelly (Venil *et al.*, 2015).

However, the challenge in manufacturing bacterial pigments is the need to produce the pigments in a large quantity at low cost (Malik *et al.*, 2012). Nutrient-rich agricultural waste medium obtained from brown sugar, rice bran, pineapple and sugar cane are increasingly popular due to its availability, low cost, and renewability (Ahmad *et al.*, 2012). In addition, the use of agricultural waste residues in bioprocess helps to reduce environmental pollution. Pineapple waste is the choice for growth medium in this study due to its high glucose content and other nutrients such as esters, ketones, alcohols, aldehydes, and acids, which are required for bacterial growth (Hemalatha and Anbuselvi, 2013). The use of pineapple waste as growth medium for the production of *Lactobacilli* sp. has been demonstrated by Pyar *et al.* (2014).

Furthermore, like other natural pigments, violacein has poor solubility in water, thus limits its usage in industrial application. The common organic solvents for violacein are dimethyl sulfoxide (DMSO), acetone, methanol and ethyl acetate, which are harmful to health and environment upon emission. Thus, particle size reduction which includes mechanical nanosization can improve solubility and dissolution rate of violacein in water due to the increase of surface area to volume ratio (Khadka *et al.*, 2014). Besides, violacein nanoparticles allow better membrane penetration and increase its pharmacological activities in drug delivery. To achieve nanosization, sonication is one of the effective top-down particle size reduction approaches. Sonication employs non-interaction vibration energy to disagglomerate and overcome bonding forces in dispersing the nanomaterials.

Nevertheless, agglomeration is a common issue in nanoparticle production. Agglomeration occurs when substances prefer to interact with the same molecules instead of interacting with solvent molecules to lower the kinetic energy and achieve a more stable structure (Mohd Hamzah *et al.*, 2017). Thus, the presence of surfactants as stabilizing agent is important to prevent agglomeration by providing steric or electrostatic repulsion. For example, baicalein nanocrystals which have potent antioxidant, antitumor, and anticancer properties were stable in water with the aid of a surfactant (Zhang *et al.*, 2011). Besides, surfactants can act as solubilizing agent by increasing the solubility in both organic and aqueous solvents. Tehrani-Bagha, Singh and Holmberg (2013) reported the increase of synthetic pigments' solubility when added with surfactant above its critical micelle concentration (CMC).

In this study, crude violacein extracted from *Chromobacterium violaceum* UTM5 was downsized to nanoparticles via sonication, with surfactants as stabilizing agent. The aim was to improve the violacein solubility in aqueous system with small particle size, narrower particle size distribution (low polydispersity index) and high zeta potential (high stability), besides retaining its violet color at different pH, temperature, time and light illumination.

## 1.2 Problem Statement

The major challenge to commercialize microbial pigments is to achieve high yield with cost-effective production (Malik *et al.*, 2012). The synthetic growth medium is expensive, thus hamper the production of the pigments at industrial scale, leading to its low usage in any applications where people prefer to use synthetic pigments. As for violacein, although exhibiting many pharmacological activities, it has not been utilized in commercial applications. Besides the high cost of production, another issue is its limited solubility in water, but can dissolve in methanol and DMSO (Durán *et al.*, 2007). The organic solvents are toxic even at low dosage (Galvao *et al.*, 2014). To reduce the production cost of violacein, liquid pineapple waste was used as a cheaper alternative growth medium. On the other hand, the solubility of violacein in

water can be increased by reducing its size into nanoscale, due to the increase of surface area to volume ratio. The high surface-to-volume ratio thus increases particle solubility in water system. This study focused on the use of sonication technique to produce the violacein nanoparticles. However, common issues with nanoparticles are poor solubility and dispersibility, this leads to aggregation and sedimentation process, which results in the loss of the bacterial pigment biological activities and reduces the pigment quality to be used as ink (Wu *et al.*, 2011). Thus, surface modification of the violacein nanoparticles with stabilizer molecules such as surfactants imparts the nanoparticles stability. The presence of surfactant as stabilizer acts as barrier, preventing agglomeration via two protection mechanisms, which are steric repulsion and electrostatic repulsion. In short, this research aims to produce a low cost and stable violacein nanoparticles with high dispersibility/solubility in aqueous system.

### **1.3 Objectives**

- 1) To produce, extract and characterize crude violacein pigment from *C. violaceum* UTM5 using liquid pineapple waste as the growth medium.
- 2) To produce and characterize violacein nanoparticles via sonication technique with the aid of surfactants as stabilizers.
- 3) To test the solubility, stability and color performance of the violacein nanoparticles.

### **1.4 Scope of Study**

The crude violacein used in this study was extracted from *C. violaceum* UTM5 strain and used without further purification. Liquid pineapple waste (LPW) and nutrient broth were used as growth medium for *C. violaceum* UTM5. The production of crude violacein was compared from using nutrient broth (NB), LPW (with and without L-tryptophan) as growth media. The bacteria was grown using continuous

shaking condition and extracted via liquid-liquid extraction using ethyl acetate and acetone as the solvents.

The production of violacein nanoparticles was done using water as the medium. The effectiveness of the sonication technique was first compared with mechanical stirring method. By focusing on sonication technique, several parameters including surfactant concentration and violacein concentration, pulse and sonication time were further investigated. Several common industrial surfactants were used in this study such as Tween 80 and Triton X-100 (nonionic surfactant), sodium dodecyl sulfate (anionic surfactant) and dodecyl trimethylammonium bromide (cationic surfactant). The performance of each surfactant was analyzed in terms of particle size, polydispersity index, zeta potential value, and solubilizing power. Sonication parameters such as sonication time and pulse time was optimized to produce stable violacein nanoparticles. The violacein nanoparticles were characterized using attenuated total reflectance Fourier transform infrared, thermal and Zetasizer analyzer.

The dispersion stability of the violacein nanoparticles were tested as a function of time (28 days) and at different temperatures. The dispersion stability was measured via violet color intensity using UV-Vis spectroscopy for upper and lower portion of the violacein solution. The color stability of the violacein nanoparticles was tested as a function of time, pH and temperature and under light illumination using UV-Vis spectrophotometer and Colorflex color meter.

## **1.5 Significance of Study**

The use of liquid pineapple waste as an alternative growth medium reduces the production cost of violacein. Besides that, the use of fruit wastes for the production of microbial pigments leads to lower waste generation, better waste management and fulfills the waste-to-wealth initiative as described in RMK-11. The development of violacein nanoparticles will improve the solubility of violacein in water, thus making them useful for various applications especially in pharmaceutical industry. The use of



## REFERENCES

- Aberoumand, A., 2011. A Review Article on Edible Pigments Properties and Sources as Natural Biocolorants in Foodstuff and Food Industry. *World Journal of Dairy and Food Sciences*, 6(1), pp.71–78.
- Adekunte, A.O., Tiwari, B.K., Cullen, P.J., Scannell, A.G.M. and O'Donnell, C.P., 2010. Effect of Sonication on Colour, Ascorbic Acid and Yeast Inactivation in Tomato Juice. *Food Chemistry*, 122(3), pp.500–507.
- Adeoye, M.D., Obi-Egbedi, N.O. and Iweibo, I., 2017. Solvent Effect and Photo-Physical Properties of 2,3-Diphenylcyclopropenone. *Arabian Journal of Chemistry*, 10, pp.134–140.
- Adesina, S.K., Ezeonyebuchi, U. and Akala, E.O., 2015. The Effect of Formulation Variables on Drug Loading of Antitubercular Drugs in Nanoparticle Formulations. *Materials Research Express*, 2(9), p.95403.
- Aggarwal, S. and Goel, A., 2012. Solubility and Its Enhancement Techniques of Poorly Soluble Drugs. *International Journal of Universal Pharmacy and Life Sciences*, 2(1), pp.65–82.
- Ahmad, W.A., Wan Ahmad, W.Y., Zakaria, Z.A. and Yusof, N.Z., 2012. Isolation of Pigment-Producing Bacteria and Characterization of the Extracted Pigments. In: *Application of Bacterial Pigments as Colorant The Malaysian Perspective*. Springer, Berlin, Heidelberg, pp. 25–44.
- Alshatwi, A.A., Subash-Babu, P. and Antonisamy, P., 2015. Violacein Induces Apoptosis in Human Breast Cancer Cells through Up Regulation of BAX, p53 and Down Regulation of MDM2. *Experimental and Toxicologic Pathology*, 68(1), pp.89–97.
- Andrighetti-Fröhner, C.R., Antonio, R. V., Creczynski-Pasa, T.B., Barardi, C.R.M. and Simões, C.M.O., 2003. Cytotoxicity and Potential Antiviral Evaluation of Violacein Produced by *Chromobacterium violaceum*. *Memorias do Instituto Oswaldo Cruz*, 98(6), pp.843–848.

- Antônio, R.V. and Creczynski-Pasa, T.B., 2004. Genetic Analysis of Violacein Biosynthesis by *Chromobacterium violaceum*. *Genetics and Molecular Research : GMR*, 3(1), pp.85–91.
- Antonisamy, P. and Ignacimuthu, S., 2010. Immunomodulatory, Analgesic and Antipyretic Effects of Violacein Isolated from *Chromobacterium violaceum*. *Phytomedicine*, 17(3–4), pp.300–304.
- Antonisamy, P., Kannan, P. and Ignacimuthu, S., 2009. Anti-Diarrhoeal and Ulcer-Protective Effects of Violacein Isolated from *Chromobacterium violaceum* in Wistar Rats. *Fundamental and Clinical Pharmacology*, 23(4), pp.483–490.
- Arif, S., Batool, A., Khalid, N., Ahmed, I. and Janjua, H.A., 2017. Comparative Analysis of Stability and Biological Activities of Violacein and Starch Capped Silver Nanoparticles. *RSC Adv.*, 7(8), pp.4468–4478.
- Aruldass, C.A., 2016. *Violet Pigment from Chromobacterium violaceum UTM5 Grown in Liquid Pineapple Waste and Its Antibacterial and Cytotoxicity Activities*. Universiti Teknologi Malaysia.
- Aruldass, C.A., Masalamany, S.R.L., Venil, C.K. and Ahmad, W.A., 2017. Antibacterial Mode of Action of Violacein from *Chromobacterium violaceum* UTM5 against *Staphylococcus aureus* and methicillin-resistant *Staphylococcus aureus* (MRSA). *Environmental Science and Pollution Research*, pp.1–17.
- Aruldass, C.A., Rubiyatno, R., Venil, C.K. and Ahmad, W.A., 2015. Violet Pigment Production from Liquid Pineapple Waste by *Chromobacterium violaceum* UTM5 and Evaluation of Its Bioactivity. *RSC Adv.*, 5(64), pp.51524–51536.
- Asencio, G., Lavin, P., Alegría, K., Domínguez, M., Bello, H., González-Rocha, G. and González-Aravena, M., 2014. Antibacterial Activity of the Antarctic Bacterium *Janthinobacterium* sp. SMN 33.6 against Multi-Resistant Gram-Negative Bacteria. *Electronic Journal of Biotechnology*, 17(1), pp.1–5.
- August, P.R., Grossman, T.H., Minor, C., Draper, M.P., MacNeil, I. a, Pemberton, J.M., Call, K.M., Holt, D. and Osburne, M.S., 2000. Sequence Analysis and Functional Characterization of The Violacein Biosynthetic Pathway from *Chromobacterium violaceum*. *Journal of Molecular Microbiology and Biotechnology*, 2(4), pp.513–519.
- Babitha, S., 2009. Microbial Pigments. In: *Biotechnology for Agro-Industrial Residues Utilisation*. Springer Netherlands, Dordrecht, pp. 147–162.
- Berde, C. V and Berde, V.B., 2015. Vegetable Waste as Alternative Microbiological

- Media for Laboratory and Industry. *World Journal Of Pharmacy And Pharmaceutical Sciences*, 4 (5), 1488, 4(5), pp.1488 – 1494.
- Berni, E., Marcato, P.D., Nakazato, G., Kobayashi, R.K.T., Vacchi, F.I., Umbuzeiro, G.A. and Durán, N., 2013. Violacein/Poly( $\epsilon$ -Caprolactone)/Chitosan Nanoparticles Against Bovine Mastitis: Antibacterial and Ecotoxicity Evaluation. *Journal of Physics: Conference Series*, 429(1).
- Bhadoriya, S.S., Madoriya, N., Madoriya, N., Shukla, K. and MS, P., 2013. Biosurfactants: A New Pharmaceutical Additive for Solubility Enhancement and Pharmaceutical Development. *Biochemistry & Pharmacology: Open Access*, 02(02).
- Biswas, S., Vaze, O.S., Movassaghian, S. and Torchilin, V.P., 2013. Polymeric Micelles for the Delivery of Poorly Soluble Drugs. In: *Drug Delivery Strategies for Poorly Water-Soluble Drugs*. John Wiley & Sons Ltd, Oxford, UK, pp. 411–476.
- Bohrey, S., Chourasiya, V. and Pandey, A., 2016. Polymeric Nanoparticles Containing Diazepam: Preparation, Optimization, Characterization, In-Vitro Drug Release and Release Kinetic Study. *Nano Convergence*, 3(1), p.3.
- Borowitzka, M.A., 2013. High-Value Products from Microalgae-Their Development and Commercialisation. *Journal of Applied Phycology*, 25(3), pp.743–756.
- Boverhof, D.R., Bramante, C.M., Butala, J.H., Clancy, S.F., Lafranconi, W.M., West, J. and Gordon, S.C., 2015. Comparative assessment of nanomaterial definitions and safety evaluation considerations. *Regulatory Toxicology and Pharmacology*, 73(1), pp.137–150.
- Bromberg, N., Dreyfuss, J.L., Regatieri, C. V., Palladino, M. V., Durán, N., Nader, H.B., Haun, M. and Justo, G.Z., 2010. Growth Inhibition and Pro-Apoptotic Activity of Violacein in Ehrlich Ascites Tumor. *Chemico-Biological Interactions*, 186(1), pp.43–52.
- Buckow, R., Kastell, A., Terefe, N.S. and Versteeg, C., 2010. Pressure and Temperature Effects on Degradation Kinetics and Storage Stability of Total Anthocyanins in Blueberry Juice. *Journal of Agricultural and Food Chemistry*, 58(18), pp.10076–10084.
- Cazoto, L.L., Martins, D., Ribeiro, M.G., Durán, N. and Nakazato, G., 2011. Antibacterial Activity of Violacein against *Staphylococcus aureus* Isolated from Bovine Mastitis. *The Journal of Antibiotics*, 64(5), pp.395–397.

- Chatterjee, S., Salaün, F. and Campagne, C., 2014. The Influence of 1-Butanol and Trisodium Citrate Ion on Morphology and Chemical Properties of Chitosan-Based Microcapsules during Rigidification by Alkali Treatment. *Marine Drugs*, 12(12), pp.5801–16.
- Choi, S.Y., Yoon, K.H., Lee, J. Il and Mitchell, R.J., 2015. Violacein: Properties and Production of a Versatile Bacterial Pigment. *BioMed Research International*, 2015, pp.1–8.
- Cooper, D.L. and Harirforoosh, S., 2014. Effect of Formulation Variables on Preparation of Celecoxib Loaded Polylactide-Co-Glycolide Nanoparticles. *PLoS ONE*, 9(12), pp.1–22.
- Darshan, N. and Manonmani, H.K., 2015. Prodigiosin and Its Potential Applications. *Journal of Food Science and Technology*, 52(9), pp.5393–5407.
- Dhakar, R.C., 2012. From Formulation Variables To Drug Entrapment Efficiency of Microspheres: A Technical Review. *Journal of Drug Delivery and Therapeutics*, 2(6).
- Dhakar, R.C., Maurya, S.D., Sagar, B.P.S., Bhagat, S., Kumar, P.S. and Jain, C.P., 2010. Variables Influencing the Drug Entrapment Efficiency of Microspheres : A Pharmaceutical Review. *Der Pharmacia Lettre*, 2(5), pp.102–116.
- Dole, M.N., Patel, P.A., Sawant, S.D. and Shedpure, P.S., 2011. Advance Applications of Fourier Transform Infrared Spectroscopy. *International Journal of Pharmaceutical Sciences Review and Research*, 7(2), pp.159–166.
- Dufossé, L., 2017. Current Carotenoid Production Using Microorganisms. In: Om V. Singh, (ed.) *Bio-pigmentation and Biotechnological Implementations*. John Wiley & Sons, Inc., Hoboken, NJ, USA, pp. 87–106.
- Dufossé, L., 2006. Microbial Production of Food Grade Pigments. *Food Technology and Biotechnology*, 44(3), pp.313–321.
- Durán, M., Ponezi, A.N., Faljoni-Alario, A., Teixeira, M.F.S., Justo, G.Z. and Durán, N., 2012. Potential Applications of Violacein: A Microbial Pigment. *Medicinal Chemistry Research*, 21(7), pp.1524–1532.
- Durán, N., Justo, G.Z., Durán, M., Brocchi, M., Cordi, L., Tasic, L., Castro, G.R. and Nakazato, G., 2016. Advances in *Chromobacterium violaceum* and Properties of Violacein-Its Main Secondary Metabolite: A Review. *Biotechnology Advances*, 34(5), pp.1030–1045.
- Durán, N., Justo, G.Z., Ferreira, C. V., Melo, P.S., Cordi, L. and Martins, D., 2007.

- Violacein: Properties and Biological Activities. *Biotechnology and Applied Biochemistry*, 48(3), p.127.
- Fakhr, F.A., Khanafari, A., Baserisalehi, M., Yaghoobi, R. and Shahghasempour, S., 2012. An Investigation of Antileukemia Activity of Violacein-Loaded Dendrimer in Jurkat Cell Lines. *African Journal of Microbiology Research*, 6(33), pp.6235–6242.
- Fang, M.-Y., Zhang, C., Yang, S., Cui, J.-Y., Jiang, P.-X., Lou, K., Wachi, M. and Xing, X.-H., 2015. High Crude Violacein Production from Glucose by *Escherichia coli* Engineered with Interactive Control of Tryptophan Pathway and Violacein Biosynthetic Pathway. *Microbial Cell Factories*, 14(1), p.8.
- Fariya, M., Jain, A., Dhawan, V., Shah, S. and Nagarsenker, M.S., 2014. Bolaamphiphiles: A Pharmaceutical Review. *Advanced Pharmaceutical Bulletin*, 4(Suppl 2), pp.483–491.
- Füller, J.J., Röpke, R., Krausze, J., Rennhack, K.E., Daniel, N.P., Blankenfeldt, W., Schulz, S., Jahn, D. and Moser, J., 2016. Biosynthesis of Violacein, Structure and Function of L-Tryptophan Oxidase VioA from *Chromobacterium violaceum*. *Journal of Biological Chemistry*, 291(38), pp.20068–20084.
- Fusaro, D., 2010. When It Comes to Synthetic Food Colors: Beware the “Southampton Six.” *Food Processing*.
- Galvao, J., Davis, B., Tilley, M., Normando, E., Duchon, M.R. and Cordeiro, M.F., 2014. Unexpected Low-Dose Toxicity of the Universal Solvent DMSO. *FASEB Journal*, 28(3), pp.1317–1330.
- Gillis, M. and De Ley, J., 2006. The Genera *Chromobacterium* and *Janthinobacterium*. In: Dworkin, M., Falkow, S., Rosenberg, E., Schleifer, K.-H. and Stackebrandt, E., (eds.) *The Prokaryotes*. Springer New York, New York, NY, pp. 737–746.
- Gürses, A., Açıkyıldız, M., Güneş, K. and Gürses, M.S., 2016. Dyes and Pigments. In: *Dyes and Pigments*. Springer International Publishing, pp. 13–29.
- Hait, S.K. and Moulik, S.P., 2001. Determination of Critical Micelle Concentration (CMC) of Nonionic Surfactants by Donor-Acceptor Interaction with Iodine and Correlation of CMC with Hydrophile-Lipophile Balance and Other Parameters of The Surfactants. *Journal of Surfactants and Detergents*, 4(3), pp.303–309.
- Hajar, N., Zainal, S., Nadzirah, K.Z., Roha, A.M.S., Atikah, O. and Elida, T.Z.M.T., 2012. Physicochemical Properties Analysis of Three Indexes Pineapple (Ananas

- Comosus) Peel Extract Variety N36. *APCBEE Procedia*, 4, pp.115–121.
- Hemalatha, R. and Anbuselvi, S., 2013. Physicochemical Constituents of Pineapple Pulp and Waste. *Journal of Chemical and Pharmaceutical Research*, 5(2), pp.240–242.
- Hoshino, T., 2011. Violacein and Related Tryptophan Metabolites Produced by *Chromobacterium violaceum*: Biosynthetic Mechanism and Pathway for Construction of Violacein Core. *Applied Microbiology and Biotechnology*, 91(6), pp.1463–1475.
- Humayun, H.Y., Shaarani, M.N.N.M., Warrior, A., Abdullah, B. and Salam, M.A., 2016. The Effect of Co-solvent on the Solubility of a Sparingly Soluble Crystal of Benzoic Acid. *Procedia Engineering*, 148, pp.1320–1325.
- İnanç, A.L., 2011. Chlorophyll: Structural Properties, Health Benefits and Its Occurrence in Virgin Olive Oils. *Akademik Gıdatr (A.L. İnanç)*, 9(2), pp.90–344.
- Joana Gil-Chávez, G., Villa, J.A., Fernando Ayala-Zavala, J., Basilio Heredia, J., Sepulveda, D., Yahia, E.M. and González-Aguilar, G.A., 2013. Technologies for Extraction and Production of Bioactive Compounds to be Used as Nutraceuticals and Food Ingredients: An Overview. *Comprehensive Reviews in Food Science and Food Safety*, 12(1), pp.5–23.
- Jódar-Reyes, A.B., Martín-Rodríguez, A. and Ortega-Vinuesa, J.L., 2006. Effect of the Ionic Surfactant Concentration on the Stabilization/Destabilization of Polystyrene Colloidal Particles. *Journal of Colloid and Interface Science*, 298(1), pp.248–257.
- Kandisa, R.V., Saibaba KV, N., Shaik, K.B. and Gopinath, R., 2016. Dye Removal by Adsorption: A Review. *Journal of Bioremediation & Biodegradation*, 07(06).
- Khadka, P., Ro, J., Kim, H., Kim, I., Kim, J.T., Kim, H., Cho, J.M., Yun, G. and Lee, J., 2014. Pharmaceutical Particle Technologies: An Approach to Improve Drug Solubility, Dissolution and Bioavailability. *Asian Journal of Pharmaceutical Sciences*, 9(6), pp.304–316.
- Khan, M.I., 2016. Plant Betalains: Safety, Antioxidant Activity, Clinical Efficacy, and Bioavailability. *Comprehensive Reviews in Food Science and Food Safety*, 15(2), pp.316–330.
- Kirti, K., Amita, S., Priti, S., Kumar, A.M. and Jyoti, S., 2014. Colorful World of Microbes: Carotenoids and Their Applications. *Advances in Biology*, 2014(1), pp.1–13.

- Klaessig, F., Marrapese, M. and Abe, S., 2011. Current Perspectives in Nanotechnology Terminology and Nomenclature. In: Murashov, V. and Howard, J., (eds.) *Nanotechnology Standards*. Springer-Verlag New York, pp. 21–52.
- Konzen, M., De Marco, D., Cordova, C.A.S., Vieira, T.O., Antônio, R. V. and Creczynski-Pasa, T.B., 2006. Antioxidant Properties of Violacein: Possible Relation on Its Biological Function. *Bioorganic and Medicinal Chemistry*, 14(24), pp.8307–8313.
- Kumar, A., Vishwakarma, H.S., Singh, J. and Kumar, M., 2015. Microbial Pigments : Production and Their Applications in Various Industries. *International Journal of Pharmaceutical, Chemical and Biological Sciences*, 5(1), pp.203–212.
- Kumar, S. and Singh, P., 2016. Various Techniques for Solubility Enhancement : An Overview. *The Pharma Innovation Journal*, 5(1), pp.23–28.
- Li, X., Qin, Y., Liu, C., Jiang, S., Xiong, L. and Sun, Q., 2016. Size-Controlled Starch Nanoparticles Prepared by Self-Assembly with Different Green Surfactant: The Effect of Electrostatic Repulsion or Steric Hindrance. *Food Chemistry*, 199, pp.356–363.
- Liu, R., 2008. *Water-Insoluble Drug Formulation* 2nd ed., CRC Press.
- Lomax, S.Q. and Learner, T., 2006. A Review of the Classes, Structures, and Methods of Analysis of Synthetic Organic Pigments. *Journal of the American Institute for Conservation*, 45(2), pp.107–125.
- Mahmood, M.E. and Al-koofee, D. a F., 2013. Effect of Temperature Changes on Critical Micelle Concentration for Tween Series Surfactant. *Global Journal of Science Frontier Research Chemistry*, 13(4), pp.1–7.
- Malik, K., Tokkas, J. and Goyal, S., 2012. Microbial Pigments: A Review. *International Journal of Microbial Resource Technology Accepted*, 41(4), pp.361–365.
- Manzo, G., Carboni, M., Rinaldi, A.C., Casu, M. and Scorciapino, M.A., 2013. Characterization of Sodium Dodecylsulphate and Dodecylphosphocholine Mixed Micelles through NMR and Dynamic Light Scattering. *Magnetic Resonance in Chemistry*, 51(3), pp.176–183.
- Martins, D., Costa, F.T.M., Brocchi, M. and Durán, N., 2011. Evaluation of the Antibacterial Activity of Poly-(d,l-Lactide-co-Glycolide) Nanoparticles Containing Violacein. *Journal of Nanoparticle Research*, 13(1), pp.355–363.
- Martins, D., Frungillo, L., Anazzetti, M.C., Melo, P.S. and Durán, N., 2010.

- Antitumoral Activity of L-Ascorbic Acid-Poly- D,L-(Lactide-Co-Glycolide) Nanoparticles Containing Violacein. *International Journal of Nanomedicine*, 5, pp.77–85.
- Masilamani, K. and Ravichandiran, V., 2012. Effect of Formulation and Process Variables on Drug Content and Entrapment Efficiency of Aceclofenac Nanosuspension. *International Research Journal of Pharmacy*, 3(3), pp.315–318.
- Mata-Gómez, L.C., Montañez, J.C., Méndez-Zavala, A. and Aguilar, C.N., 2014. Biotechnological Production of Carotenoids by Yeasts: An Overview. *Microbial Cell Factories*, 13(1), pp.1–11.
- McLellan, M.R., Lind, L.R. and Kime, R.W., 1995. Hue Angle Determinations and Statistical Analysis for Multiquadrant Hunter L,a,b Data. *Journal of Food Quality*, 18, pp.235–240.
- Mishra, M., Muthuprasanna, P. and Prabha, K., 2009. Basics and Potential Applications of Surfactants—A Review. *Int J PharmTech Res*, 1(4), pp.1354–1365.
- Moawad, H., Abd El-Rahim, W.M. and Khalafallah, M., 2003. Evaluation of Biototoxicity of Textile Dyes Using Two Bioassays. *Journal of Basic Microbiology*, 43(3), pp.218–229.
- Mohajeri, E. and Noudeh, G.D., 2012. Effect of Temperature on the Critical Micelle Concentration and Micellization Thermodynamic of Nonionic Surfactants: Polyoxyethylene Sorbitan Fatty Acid Esters. *E-Journal of Chemistry*, 9(4), pp.2268–2274.
- Mohd Hamzah, M.A.A., Aruldass, C.A., Ahmad, W.A. and Setu, S.A., 2017. Effects of Surfactants on Antibacterial Drugs – A Brief Review. *Malaysian Journal of Fundamental and Applied Sciences*, 13(2), pp.118–123.
- MPIB, 2016. *Data Pengeluaran Nanas Malaysia Tahun 2016*, Johor Bahru, Johor.
- Muhammad Khan, A. and Shah, S.S., 2008. Determination of Critical Micelle Concentration (Cmc) of Sodium Dodecyl Sulfate (SDS) and the Effect of Low Concentration of Pyrene on its Cmc Using ORIGIN Software. *Journal of Chemistry Society Pakistan*, 30(2), pp.186–191.
- Nakamura, Y., Asada, C. and Sawada, T., 2003. Production of Antibacterial Violet Pigment by Psychrotropic Bacterium RT102 Strain. *Biotechnology and Bioprocess Engineering*, 8(1), pp.37–40.



- Namazkar, S., Garg, R., Ahmad, W.Z. and Nordin, N., 2013. Production and Characterization of Crude and Encapsulated Prodigiosin Pigment. *International Journal of Chemical Sciences and Applications*, 4(3), pp.2278–6015.
- Natalia, C.-O., Mayra-Alexandra, C.-C., Vanessa, C.-A. and Luis-Daniel, P.-S., 2017. Influence of Environmental Factors on the Production of Violacein Synthesized By *Janthinobacterium lividum*. *The International Journal of Engineering and Science*, 06(01), pp.76–83.
- Nayak, A.K. and Panigrahi, P.P., 2012. Solubility Enhancement of Etoricoxib by Cosolvency Approach. *ISRN Physical Chemistry*, 2012, pp.1–5.
- Ngamwonglumlert, L., Devahastin, S. and Chiewchan, N., 2017. Natural Colorants: Pigment Stability and Extraction Yield Enhancement via Utilization of Appropriate Pretreatment and Extraction Methods. *Critical Reviews in Food Science and Nutrition*, 57(15), pp.3243–3259.
- Nobbmann, U. and Morfesis, A., 2008. Characterization of Nanoparticles by Light Scattering. *MRS Proceedings*, 1074(April 2016), pp.1074-I10-45.
- Nogueira, D.R., Mitjans, M., Infante, M.R. and Vinardell, M.P., 2011. The Role of Counterions in the Membrane-Disruptive Properties of pH-Sensitive Lysine-Based Surfactants. *Acta Biomaterialia*, 7(7), pp.2846–2856.
- Novotný, Č., Dias, N., Kapanen, A., Malachová, K., Vándrovcová, M., Itävaara, M. and Lima, N., 2006. Comparative Use of Bacterial, Algal and Protozoan Tests to Study Toxicity of Azo- and Anthraquinone Dyes. *Chemosphere*, 63(9), pp.1436–1442.
- O'Neill, C., Hawkes, F.R., Hawkes, D.L., Lourenço, N.D., Pinheiro, H.M. and Delée, W., 1999. Colour in Textile Effluents - Sources, Measurement, Discharge Consents and Simulation: A Review. *Journal of Chemical Technology and Biotechnology*, 74(11), pp.1009–1018.
- Olorunsola, E.O. and Adedokun, M.O., 2014. Surface Activity as Basis for Pharmaceutical Applications of Hydrocolloids: A Review. *Journal of Applied Pharmaceutical Science*, 4(10), pp.110–116.
- Orna, M.V., 2013. Discovery of the Physics of Color. In: *The Chemical History of Color*. Springer-Verlag Berlin Heidelberg, pp. 11–28.
- Pereira, L. and Alves, M., 2012. Dyes-Environmental Impact and Remediation. In: *Environmental Protection Strategies for Sustainable Development*. Springer Netherlands, Dordrecht, pp. 111–162.

- Pongpeerapat, A., Itoh, K., Tozuka, Y., Moribe, K., Oguchi, T. and Yamamoto, K., 2004. Formation and Stability of Drug Nanoparticles Obtained from Drug/PVP/SDS Ternary Ground Mixture. *Journal of Drug Delivery Science and Technology*, 14(6), pp.441–447.
- Pyar, H., Liong, M.T. and Peh, K.K., 2014. Potentials of Pineapple Waste as Growth Medium for *Lactobacillus* species. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(1), pp.142–145.
- Rahman, F., 2015. The Treatment of Industrial Effluents for the Discharge of Textile Dyes Using by Techniques and Adsorbents. *Journal of Textile Science & Engineering*, 06(01), pp.1–9.
- Rahman, M., Bashar, A., Khalipha, R., Azad, A.K., Hossain, S. and Haque, S., 2014. Methods of Solubility and Dissolution Enhancement for Poorly Water Soluble Drugs : a Review. *World Journal of Pharmacy and Pharmaceutical Sciences*, 3(5), pp.107–130.
- Rajendran, R. and Selvi, B.T., 2014. Natural Dyeing of Cotton Fabrics with Pigment Extracted from *Roseomonas Fauriae*. *Universal Journal of Environmental Research and Technology*, 4(1), pp.54–59.
- Rathi, P.B., Kale, M., Soleymani, J. and Jouyban, A., 2018. Solubility of Etoricoxib in Aqueous Solutions of Glycerin, Methanol, Polyethylene Glycols 200, 400, 600, and Propylene Glycol at 298.2 K. *Journal of Chemical & Engineering Data*.
- Ratna, P.B.S., 2012. Pollution due to Synthetic Dyes Toxicity & Carcinogenicity Studies and Remediation. *International Journal of Environmental Sciences*, 3(3), pp.940–955.
- Reshmi, S.K., Aravindhan, K.M. and Devi, P.S., 2012. The Effect of Light, Temperature, pH on Stability of Betacyanin Pigments in *Basella Alba* Fruit. *ASian Journal of Phatmaceutical and Clinical Research*, 5(4), pp.5–8.
- Rettori, D. and Durán, N., 1998. Production, Extraction and Purification of Violacein: an Antibiotic Pigment Produced by *Chromobacterium violaceum*. *World Journal of Microbiology and Biotechnology*, 14, pp.685–689.
- Reynolds, D.M., 2014. The Principles of Fluorescence. In: Coble, P., Lead, J., Baker, A., Reynolds, D.M. and Spencer, R.G.M., (eds.) *Aquatic Organic Matter Fluorescence*. Cambridge University Press, Cambridge, pp. 3–34.
- Rodrigo-Baños, M., Garbayo, I., Vílchez, C., Bonete, M.J. and Martínez-Espinosa, R.M., 2015. Carotenoids from Haloarchaea and Their Potential in Biotechnology.

- Marine Drugs*, 13(9), pp.5508–5532.
- Rodrigues, A.L., Göcke, Y., Bolten, C., Brock, N.L., Dickschat, J.S. and Wittmann, C., 2012. Microbial Production of the Drugs Violacein and Deoxyviolacein: Analytical Development and Strain Comparison. *Biotechnology Letters*, 34(4), pp.717–720.
- Samani, B.H. and Lorigooini, Z., 2015. Effects of Ultrasonic on Microorganisms and Enzymes. *International Science and Investigation Journal*, 4(January), pp.106–113.
- Scheeren, L.E., Nogueira, D.R., Macedo, L.B., Vinardell, M.P., Mitjans, M., Infante, M.R. and Rolim, C.M.B., 2016. PEGylated and Poloxamer-Modified Chitosan Nanoparticles Incorporating a Lysine-Based Surfactant for pH-Triggered Doxorubicin Release. *Colloids and Surfaces B: Biointerfaces*, 138, pp.117–127.
- Seedher, N. and Agarwal, P., 2009. Various Solvent Systems for Solubility Enhancement of Enrofloxacin. *Indian Journal of Pharmaceutical Sciences*, 71(1), pp.82–7.
- Seedher, N. and Kanojia, M., 2009. Co-Solvent Solubilization of Some Poorly-Soluble Antidiabetic Drugs Solubilization Antidiabetic Drugs. *Pharmaceutical Development and Technology*, 14(2), pp.185–192.
- Sekhon, B.S., 2013. Surfactants: Pharmaceutical and Medicinal Aspects. *Journal of Pharmaceutical Technology, Research and Management*, 1(1), pp.43–68.
- Seyedin, A., Hatamian-zarmi, A., Rasekh, B. and Mir-derikvand, M., 2015. Natural Pigment Production by *Monascus purpureus*: Bioreactor Yield Improvement through Statistical Analysis. *Applied Food Biotechnology*, 2(2), pp.23–30.
- Shah, M., Agrawal, Y.K., Garala, K. and Ramkishan, A., 2012. Solid Lipid Nanoparticles of a Water Soluble Drug, Ciprofloxacin Hydrochloride. *Indian Journal of Pharmaceutical Sciences*, 74(5), pp.434–42.
- Sharma, D., Maheshwari, D., Philip, G., Rana, R., Bhatia, S., Singh, M., Gabrani, R., Sharma, S.K., Ali, J., Sharma, R.K. and Dang, S., 2014. Formulation and Optimization of Polymeric Nanoparticles for Intranasal Delivery of Lorazepam Using Box-Behnken Design: In Vitro and In Vivo Evaluation. *BioMed Research International*, 2014.
- Sharma, N., Madan, P. and Lin, S., 2016. Effect of Process and Formulation Variables on the Preparation of Parenteral Paclitaxel-Loaded Biodegradable Polymeric Nanoparticles: A Co-Surfactant Study. *Asian Journal of Pharmaceutical*

- Sciences*, 11(3), pp.404–416.
- Shid, R.L., Dhole, S.N., Kulkarni, N. and Shid, S.L., 2014. Formulation and Evaluation of Nanosuspension Formulation for Drug Delivery of Simvastatin. *Int J Pharm Sci Nanotech Vol*, 7(4), pp.2650–2665.
- Singh, K. and Arora, S., 2011. Removal of Synthetic Textile Dyes from Wastewaters: A Critical Review on Present Treatment Technologies. *Critical Reviews in Environmental Science and Technology*, 41(9), pp.807–878.
- Singh, O., Kaur, R. and Mahajan, R.K., 2017. Flavonoid-Surfactant Interactions: A Detailed Physicochemical Study. *Spectrochimica Acta - Part A: Molecular and Biomolecular Spectroscopy*, 170, pp.77–88.
- Singh, R., 2012. *Solubilization of Organic Dyes in Surfactant Micelles*. Chalmers University of Technology.
- Suslick, K.S., 1998. Sonochemistry. *Kirk-Othmer Encyclopedia of Chemical Technology*, pp.516–541.
- Suslick, K.S., 1997. Sonoluminescence and Sonochemistry. *1997 IEEE Ultrasonics Symposium Proceedings. An International Symposium (Cat. No.97CH36118)*, 1, pp.523–532.
- Tadros, T.F., 2005. Physical Chemistry of Surfactant Solutions. In: *Applied Surfactants*. Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, FRG, pp. 19–51.
- Tallury, P., Randall, M.K., Thaw, K.L., Preisser, J.S. and Kalachandra, S., 2007. Effects of Solubilizing Surfactants and Loading of Antiviral, Antimicrobial, and Antifungal Drugs on Their Release Rates from Ethylene Vinyl Acetate Copolymer. *Dental Materials: Official Publication of the Academy of Dental Materials*, 23(8), pp.977–82.
- Taurozzi, J.S., Hackley, V.A. and Wiesner, M.R., 2010. Protocol for Preparation of Nanoparticle Dispersions From Powdered Material Using Ultrasonic Disruption. *CEINT, National Institute of Standards and Technology*, pp.1–10.
- Tayade, P. and Modi, A., 2007. A Comparative Solubility Enhancement Profile of Valdecoxib with Different Solubilization Approaches. *Indian Journal of Pharmaceutical Sciences*, 69(2), p.274.
- Tehrani-Bagha, A.R. and Holmberg, K., 2013. Solubilization of Hydrophobic Dyes in Surfactant Solutions. *Materials*, 6(2), pp.580–608.
- Tehrani-Bagha, A.R., Singh, R.G. and Holmberg, K., 2013. Solubilization of Two

- Organic Dyes by Anionic, Cationic and Nonionic Surfactants. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 417, pp.133–139.
- Tinoi, J. and Rakariyatham, N., 2015. Utilization of Pineapple Waste Hydrolysate for Lipid Production by Oleaginous Yeast *Rhodoturula glutinis*. *International Journal of Advanced Research*, 3(3).
- Toraman, O.Y., 2017. Experimental Investigations of Preparation of Calcite Particles by Ultrasonic Treatment. *Physicochemical Problems of Mineral Processing*, 53(2), pp.859–868.
- Torchilin, V.P., 2001. Structure and Design of Polymeric Surfactant-Based Drug Delivery Systems. *Journal of Controlled Release*, 73(2–3), pp.137–172.
- Trivedi, J.S. and Wells, M.L., 2008. Solubilization Using CoSolvent Approach. In: Liu, R., (ed.) *Water-Insoluble Drug Formulation*. CRC Press, pp. 141–168.
- Tuli, H.S., Chaudhary, P., Beniwal, V. and Sharma, A.K., 2015. Microbial Pigments as Natural Color Sources: Current Trends and Future Perspectives. *Journal of Food Science and Technology*, 52(8), pp.4669–4678.
- Tzintzun-Camacho, O., Sánchez-Segura, L., Minchaca-Acosta, A.Z., Rosales-Colunga, L.M., Hernández-Orihuela, A.L. and Martínez-Antonio, A., 2016. Development of a Bacterial Culture Medium from Avocado Seed Waste. *PeerJ Preprints*, 4, p.e2104v1.
- Upadhyay, A., Lama, J.P. and Tawata, S., 2013. Utilization of Pineapple Waste: A Review. *Journal of Food Science and Technology Nepal*, 6(0).
- Venil, C.K., Ainuddin, M., Wahidin, B., Arul, C., Wan, A.& and Ahmad, A., 2017. Production of Bacterial Pigments in Low Cost Medium and Formulation of Biodegradable Ink. *Indian Journal of Experimental Biology*, 55(July), pp.441–447.
- Venil, C.K., Aruldass, C.A., Abd Halim, M.H., Khasim, A.R., Zakaria, Z.A. and Ahmad, W.A., 2015. Spray Drying of Violet Pigment from *Chromobacterium violaceum* UTM 5 and Its Application in Food Model Systems. *International Biodeterioration and Biodegradation*, 102, pp.324–329.
- Venil, C.K. and Lakshmanaperumalsamy, P., 2009. An Insightful Overview on Microbial Pigment, Prodigiosin. *Electronic Journal of Biology*, 5(3), pp.49–61.
- Venil, C.K., Zakaria, Z.A. and Ahmad, W.A., 2013. Bacterial Pigments and Their Applications. *Process Biochemistry*, 48(7), pp.1065–1079.
- Ventura-camargo, B.D.C. and Marin-morales, M.A., 2013. Azo Dyes :

- Characterization and Toxicity – A Review. *Textiles and Light Industrial Science and Technology*, 2(2), 2(2), pp.85–103.
- Vivian, J.T. and Callis, P.R., 2001. Mechanisms of Tryptophan Fluorescence Shifts in Proteins. *Biophysical Journal*, 80(5), pp.2093–2109.
- Wang, H., Jiang, P., Lu, Y., Ruan, Z., Jiang, R., Xing, X.H., Lou, K. and Wei, D., 2009. Optimization of Culture Conditions for Violacein Production by a New Strain of *Duganella* sp. B2. *Biochemical Engineering Journal*, 44(2–3), pp.119–124.
- Wu, L., Zhang, J. and Watanabe, W., 2011. Physical and Chemical Stability of Drug Nanoparticles. *Advanced Drug Delivery Reviews*, 63(6), pp.456–469.
- Wu, W., Ichihara, G., Suzuki, Y., Izuoka, K., Oikawa-Tada, S., Chang, J., Sakai, K., Miyazawa, K., Porter, D., Castranova, V., Kawaguchi, M. and Ichihara, S., 2014. Dispersion Method for Safety Research on Manufactured Nanomaterials. *Industrial Health*, 52(1), pp.54–65.
- Yadav, L.D.S., 2005. Ultraviolet (UV) and Visible Spectroscopy. *Organic Spectroscopy*, pp.7–51.
- Zhang, J., Lv, H., Jiang, K. and Gao, Y., 2011. Enhanced Bioavailability after Oral and Pulmonary Administration of Baicalein Nanocrystal. *International Journal of Pharmaceutics*, 420(1), pp.180–188.