

SULFONATED POLYETHERSULFONE AND FUNCTIONALIZED
MULTIWALL CARBON NANOTUBES/POLYVINYLPIRROLIDONE
NANOCOMPOSITE BASED HEMODIALYSIS MEMBRANE

MUHAMMAD IRFAN

A thesis is submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (Bio-process Engineering)

Faculty of Chemical and Energy Engineering,
University Teknologi Malaysia

MARCH 2017

DEDICATION

I would like to dedicate this thesis to my beloved wife,

MASOOMA

Lovely children

Zavier and Zimal,

And especially my **Parents** for their prayers, endless support and encouragement.

ACKNOWLEDGEMENT

In the name of ALLAH, the Almighty, the Most Gracious and the Most Merciful, Alhamdulillah, all praises be to Allah for His countless blessings and granting me the health, strength, and time for the completion of my PhD Thesis.

I would like to take this opportunity to express my appreciation to my supervisor, Prof Dr. Ani Idris for the prestigious guidance, supervision, invaluable advice, practical view, enormous patience, constant motivation and support throughout the development of this research.

Heartfelt thanks also to the Mr. Eric, who has given me valuable discussions and motivation while working together in MTDC-UTM on different projects. My special thanks also go to my colleagues, especially Nur Farah, Hasrul, Ehsan, Rozita, Javid, Nana, Atta, Junaid, Dr. Teo, Dr, Zohre, Kian Hwa Chan and Ting Wong for their help and support throughout my studies. I would like to extend my appreciation for their constant help, much needed motivation, discussion and collaboration in different research work.

My sincere appreciation is also extended to others who have provided assistance directly and indirectly at various occasions. Their views and tips are useful indeed. Last but not least, I would like to thank my lovely and beautiful wife, children, parents, and friends for their vital encouragement and loving care.

ABSTRACT

Chemical modification of polymer and blending of suitable additives are the common methods used to improve the properties of polyethersulfone (PES) based hemodialysis membranes. In this research work, both methods are adopted and novel nanocomposite based additives were synthesized and blended with PES alone; and then with chemically modified PES (sulfonated PES (S-PES)). The whole research work was divided into three phases. In the first phase, the nanocomposites (NCs) were formed by mixing together the acid functionalized multiwall carbon nanotubes (f-MWCNT) and two different grades of polyvinylpyrrolidone (PVP-k90 and PVP-k30) in dimethylformamide and subsequently blended with PES. The f-MWCNT contained some hydrophilic functional groups ($-\text{COOH}$, and $-\text{OH}$) and heredity hydrophobic carbon part, which made it dual nature. On one side, its carbon part created sites for attachment for the hydrophobic polymer (PES) by hydrophobic-hydrophobic interaction and $\pi-\pi$ stacking, whereas on the other side, its hydrophilic acid and hydroxyl groups attracted the hydrophilic sides of PVP by hydrogen bonding, dipole-dipole interaction and dispersion forces. Thus, f-MWCNT acted as the anchoring material between the PVP and PES in the membrane that also greatly reduced the leaching process of the additives and stabilize the membrane composition as shown by elution ratio test. The Fourier transform infrared spectroscopy spectra of fabricated membranes revealed that both types of NCs were physically bonded with PES by hydrogen bonding and the addition of NCs to PES, improved the internal capillary system of membranes as confirmed by field emission scanning electron microscope analysis. The results showed that f-MWCNT/PVP-k90 based membranes exhibited better performance than f-MWCNT/PVP-k30 based membranes in terms of flux rate, rejection rate and biocompatibility. The results from dialysis of uremic solutes unveiled that membrane formed by PVP-k90 based NCs demonstrated superior performance with 56.30%, 55.08% and 27.90% clearance ratio of urea, creatinine and lysozyme solutes, respectively. In the second phase, two best performance membranes of f-MWCNT/PVP-k90 NCs based were selected and then blended with variable ratio of S-PES. The outcome indicated that the blending of S-PES polymer, further enhanced the membrane biocompatibility and reduced the protein adsorption (bovine serum albumin, 55% and lysozyme, 65%), hemolysis process (74.80%) and illustrated longer clotting times than pristine and non-sulfonated membranes. The clearance ratio of uremic solutes was also improved and reached up to 57.3%, 57.1% and 32.4% of urea, creatinine and lysozyme, respectively. Thus, the blending of S-PES and NCs in the PES membrane greatly improved the biocompatibility and removal ability of uremic solutes. In the third and final phase, the hollow fiber (HF) membranes were spun using S-PES and PVPk90/f-MWCNT based NCs and the HF membrane characteristics and dialysis performances were evaluated. The results showed that HF membrane had a good flux rate (29.8l/h.m².bar), low molecular weight cut off (29-34 kDa) than pristine PES membranes. The dialysis tests confirmed that the HF membranes illustrated 72.7%, 75.1% and 35.4% clearance ratio of urea, creatinine and lysozyme solutes, especially. Thus, the blending of S-PES and NCs in the PES membrane highly improved the biocompatibility and removal ability of uremic solutes and it can be used in commercial grade dialyzers.

ABSTRAK

Pengubahsuaian kimia untuk polimer serta pengadunan bahan tambah yang bersesuaian adalah kaedah biasa yang dilakukan untuk menambahbaik ciri-ciri bagi membran hemodialisis yang berasaskan polietersulfon (PES). Dalam kerja penyelidikan ini, kedua-dua kaedah digunakan iaitu nano-komposit (NCs) novel berasaskan bahan tambah telah disintesis kemudian diadunkan dengan PES sahaja; dan kemudiannya PES diubahsuai secara kimia (PES tersulfonat, (S-PES)). Penyelidikan ini telah dibahagikan kepada tiga fasa. Dalam fasa pertama, NCs telah dibentuk dengan mencampurkan bersama-sama tiub nano karbon berbilang dinding yang difungsikan dengan f-MWCNT dan dua jenis polivinilpirolidon yang berbeza gred (PVP-k90 dan PVP-k30) dalam dimetilformamid dan seterusnya diadunkan dengan PES. Asid f-MWCNT mengandungi beberapa kumpulan berfungsi hidrofilik (-COOH dan -OH) dan bahagian karbon hidrofobik, yang menjadikan ia mengandungi dua sifat. Pada satu bahagian, karbon menyediakan tapak untuk lekatan bagi polimer hidrofobik (PES) dengan interaksi hidrofobik-hidrofobik dan penyusunan π - π , manakala di sisi lain, asid hidrofilik dan kumpulan hidroksil ditarik ke bahagian hidrofilik PVP oleh ikatan hidrogen, interaksi dwikutub-dwikutub dan penyebaran daya. Oleh itu, f-MWCNT telah bertindak sebagai bahan teras antara PVP dan PES dalam membran yang juga dengan banyaknya mengurangkan proses larut lesap bahan tambah dan menstabilkan komposisi membran seperti yang ditunjukkan oleh ujian nisbah elusi. Keputusan spektrum spektroskopi inframerah transformasi Fourier untuk membran yang terhasil mendedahkan bahawa kedua-dua jenis NCs ini secara fizikal terikat dengan PES oleh ikatan hidrogen dan penambahan mereka ke PES telah menambah baik sistem kapilari dalaman membran seperti yang disahkan oleh analisis mikroskop elektron pengimbas pancaran medan. Hasil kajian menunjukkan bahawa membran berasaskan f-MWCNT/PVP-k90 menunjukkan prestasi lebih baik daripada membran berasaskan f-MWCNT/PVP-k30 dari segi kadar fluks, kadar penolakan dan keserasian bio. Keputusan dialisis bahan larut uremik menunjukkan bahawa membran dibentuk dengan PVP-k90 berasaskan NCs telah menunjukkan prestasi yang lebih baik dengan 56.30%, 55.08% dan 27.90% masing-masing bagi nisbah kepelepasan bahan larut urea, kreatinin dan lisozim. Dalam fasa kedua, dua membran yang mempunyai prestasi terbaik berasaskan f-MWCNT/PVP-k90 NCs dipilih dan kemudian diadunkan dengan S-PES mengikut nisbah yang berlainan. Hasilnya menunjukkan bahawa pengadunan S-PES polimer telah meningkatkan lagi keserasian bio membran dan telah mengurangkan penjerapan protein (bovin serum albumin, 55% dan lisozim, 65%), proses hemolisis (74.80%) dan menunjukkan pembekuan lebih panjang berbanding asal dan membran tanpa sulfonat. Nisbah pelepasan bahan larut uremik juga bertambah baik dan mencapai sehingga 57.3%, 57.1% dan 32.4% masing-masing untuk urea, kreatinin dan lisozim masing-masing. Oleh itu, pengadunan S-PES dan NCs di dalam membran PES telah menambah baik keserasian bio dan keupayaan penyingkiran bahan larut uremik. Dalam fasa ketiga iaitu fasa terakhir, gentian berongga (HF) membran telah dihasilkan oleh S-PES dan PVPk90/f-MWCNT berasaskan NCs dan ciri-ciri membran HF dan prestasi dialisis telah dinilai. Hasil kajian menunjukkan bahawa membran HF mempunyai kadar fluks yang baik ($29.8 \text{ l / h.m}^2\text{.bar}$), berat molekul dengan had potongan yang rendah (29-34 kDa) berbanding dengan membran PES asli. Ujian dialisis mengesahkan bahawa membran HF menunjukkan 72.7%, 75.1% dan 35.4% masing-masing bagi nisbah pelepasan urea, kreatinin dan lisozim bahan larut. Oleh itu, pengadunan S-PES dan NCs dalam membran PES dapat menambah baik keserasian bio dan keupayaan penyingkiran bahan larut uremik dan ia boleh digunakan dalam dialisis gred komersial.

TABLE OF CONTENT

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENT	vii
	LIST OF TABLES	xiii
	LIST OF FIGURES	xv
	LIST OF ABBREVIATIONS	xxi
	LIST OF SYMBOLS	xxv
	LIST OF APPENDICES	xxvii
1	INTRODUCTION	1
	1.1 Overview	1
	1.2 Problem Statement	6
	1.3 Objective of the Study	9
	1.4 Scope of the Study	10
	1.5 Significance of the Study	11
2	LITRATURE REVIEW	13
	2.1 Renal Failure	13
	2.2 Hemodialysis	15

2.2.1	Solute Transport Mechanism Across the Membrane	16
2.2.2	Dialysis Membrane Transport Model	18
2.2.3	Permeation Rate of HD Membranes	20
2.2.4	Membranes for Blood-Filtering Unit	21
2.2.5	The Blood Dialyzers	21
2.3	Selection of Polymeric Material for HD Treatment	23
2.4	PES: Hemodialysis Polymer	26
2.5	PES-Associated Blood Reaction	27
2.5.1	Thrombogenesis	28
2.5.2	Platelet Adhesion	30
2.5.3	Complement Activation	31
2.5.4	Leukocytes and Endotoxin	33
2.6	Modification Techniques	33
2.6.1	Albumin Immobilization	40
2.6.2	Anticoagulants Immobilization	42
2.6.3	PEG/PEO, Pluronic and Biomimetic Zwitterionic-Cilliry Brushes	45
2.6.4	Polyvinyl Pyrrolidone Amphiphilic Effect	51
2.6.5	Vitamin E-effect	55
2.6.6	Anionic Functional Groups Effects	56
2.6.6.1	Sulfonation	57
2.6.6.2	Carboxylation	59
2.6.7	Physical Blending	60
2.6.8	Hydrophilicity and Hydrophobicity	61

2.7	Nanotechnology in Biomedical Field	63
2.7.1	Structure and Classification of CNTs	64
2.7.2	The CNTs Purification	66
2.7.3	Functionalization of CNTs	67
2.7.3.1	Covalent Functionalization	68
2.7.3.2	Non-Covalent Functionalization	69
2.7.4	Biosafety of CNTs	72
2.8	Conclusion	74
3	METHODOLOGY	77
3.1	Introduction	77
3.2	Materials	77
3.3	MWCNT Functionalization	78
3.4	Synthesis of f-MWCNT/PVP-k90 Based Nanocomposites	79
3.5	Fabrication of PES NCs Hemodialysis Membranes	80
3.6	Synthesis of f-MWCNT/PVP-k30 Based Nanocomposites	82
3.6.1	Membrane Fabrication	82
3.7	Sulfonation	83
3.7.1	Degree of sulfonation (DS) and Ion Exchange Capacity (IEC)	83
3.7.2	Selection of S-PES Based Formulation	84
3.7.3	Preparation of f-MWCNT/PVP-k90 Based Nanocomposites	84

3.7.4	Fabrication of S-PES/nanocomposites Based Membranes	84
3.8	Chemical Characterization of Membrane	85
3.9	Morphology of Membrane	86
3.10	Wettability Properties	86
3.11	Porosity and Pore Size of Membrane	87
3.12	Membrane Performance	87
3.13	Leaching test	89
3.14	Surface Roughness	89
3.15	Biocompatibility	90
3.15.1	Adsorption of Proteins	90
3.15.2	Thrombogenicity (PT, TT, APTT)	90
3.15.3	Hemolysis	92
3.16	Dialysis of Uremic Solutes	92
3.17	Hollow Fiber (HF) Membrane	94
3.17.1	Spinning of HF Membranes	94
3.17.2	Post treatment and Potting	96
3.17.3	Membrane Morphology	96
3.17.4	Flux rate	97
3.17.5	Pore Size and MWCO of HFs	97
3.17.6	The Dialysis Performance of HF	98
3.18	Flow chart	98
4	PES AND NANOCOMPOSITES BASED HEMODIALYSIS MEMBRANE	
4.1	Introduction	100

4.2	Characterization of MWCNT, f-MWCNT and Nanocomposites	100
4.2.1	Characterization of PES-Nanocomposite Membranes	103
4.2.2	Membrane Morphology	104
4.2.3	Contact Angle	107
4.2.4	Porosity and Pore size	110
4.2.5	Solute Rejection Rate and Molecular Weight Cutoff	112
4.2.7	Leaching Ratio	114
4.2.8	Flux Rate	115
4.2.9	Surface Roughness	116
4.2.10	Biocompatibility	118
	4.2.10.1 Protein Adsorption	118
	4.2.10.2 Thrombogenicity	120
4.2.11	Uremic Solutes clearance (Urea, Creatinine, Lysozyme)	122
4.3	Summary	124

5	SULPHONATED-PES AND NANOCOMPOSITE (PVP-K90/F-MWCNT) BASED HEMODIALYSIS MEMBRANE	126
5.1	Introduction	126
5.2	Sulfonation and Acid functionalized MWCNT	126
5.3	FTIR Analysis of NCs and Membranes	127
5.4	Membrane Morphology	131

5.5	Hydrophilicity	133
5.6	Surface Roughness	134
5.7	Solute Rejection and MWCO	137
5.8	Pore size and Porosity and Flux rate	139
5.9	Leaching Ratio	140
5.10	Biocompatibility	141
	5.10.1 Protein adsorption	141
	5.10.2 Thrombogenesis	143
	5.10.3 Hemolysis	145
5.11	Uremic Solutes (Urea, Creatinine, Lysozyme)	146
5.12	Summry	148
6	HOLLOW FIBER HEMODIALYSIS MEMBRANES	149
6.1	Introduction	149
6.2	Hollow Fiber Membrane Morphology	149
6.3	Rejection Rate and MWCO of HF	152
6.4	Flux rate	154
6.5	Dialysis Performances	155
6.6	Summry	157
7	CONCLUSIONS AND FUTURE RECOMMENDATIONS	159
7.1	Conclusions	159
7.2	Recommendation for future work	160
	REFERENCES	162
	Appendix A	195-198

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Uremic Solutes with Potential Toxicity: Uremic Solutes with Potential Toxicity.	14
2.2	Classification of small, middle and large uremic solutes, present in human blood.	17
2.3	The technical specifications of some commercial blood dialyzers.	23
2.4	A list of common polymeric material used for fabrication of hemodialysis membranes .	24
2.5	A comparison of properties between BSA grafted/layered membranes with non-grafted membranes.	36
2.6	Raft and FRSP preparation based additive and their effect on membrane performance.	37
2.7	Monomer, nanoparticles, nano-composite and S-PES containing hemodialysis membrane.	38
2.8	Comparison between SWCNTs and MWCNTs.	65
2.9	The Common methods used for the preparation of CNTs.	66
2.10	The possible impurities associated with non-purified SWCNTs and MWCNTs.	67

2.11	The summarized information about different types of covalent functionalized CNTs.	68
2.12	The effect of CNT in biomedical applications.	71
2.13	The bio-safety and toxicity study of CNTs in the biological system.	72
3.1	List of different chemicals that was used in the research work.	78
3.2	Composition of the PES and f-MWCNT/PVP-k90 NC's based hemodialysis membranes.	80
3.3	Composition of the PES and f-MWCNT/PVP-k30 NC's based hemodialysis membranes.	82
3.4	Formulation of different nano-composites (step 1) and their corresponding S-PES and PES based membranes (step 2).	85
3.5	Composition of the PES and S-PES/PES/NC's based HF hemodialysis membranes.	94
3.6	Spinning conditions of hollow fiber membranes.	96
6.1	The dimension of fabricated HF's membranes, measured by FESEM images using the scale mentioned on each figure by Image J software, that included the complete diameter of HF, inner hole diameter and wall thickness.	152

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	The movement of uremic solutes of the blood via a membrane into the dialysate.	17
2.2	Structure of hollow-fiber dialyzer and design factors. of hollow-fiber dialyzer and design factors.	22
2.3	Overview of blood and polymer interaction, which results in the formation of fibrin matrix.	28
2.4	Coagulation cascade model of thrombogenesis.	29
2.5	Reduction of platelet adhesion.	31
2.6	Complement activation values; C3a and C5a (ng/ml) of the various formulated membranes.	32
2.7	BSA grafting scheme with modified PES membranes.	41
2.8	Some important Anticoagulants structures.	43
2.9	Citric acid grafted polyurethane with EG, PEG and BDO.	45
2.10	Diagrammatic representation of a) hydrogel and b) cilia type brush development at PES membrane surface. The hydrogel consists of crosslinked water swellable PEG or PEO chains.	47

2.11	Structure of SMA-g-MPEG, mPEG-PU-mPEG and CA-PU-CA compounds.	48
2.12	The chemical structures of biomimetic and zwitterionic molecules.	50
2.13	Schematic representation of working behavior of PC/MPC containing PES membrane against protein absorption.	50
2.14	Chemical structures of PVP blocks.	55
2.15	Structure of Vitamin E.	56
2.16	Synthetic procedure for the S-PES and CPES polymers.	58
2.17	Diagrammatic representation of polymer brush made of PES hydrophobic polymer and grafted hydrophilic part. The grafted part is stretched away from the base due to absence of hydrophobic-hydrophilic attractions.	62
2.18	Orientation of PVP-PMMA-PVP tri-block in PES polymer after phase separation process. a) In (PVP-b-PMMA-b-PVP) block PMMA components are represented by dark and thick line as compared to PVP; b) Polymer solution containing block additive; c) Membrane surface showing PMMA are more toward PES and PVP tend to move far away than PES polymer.	62
2.19	Conceptual diagrams of (a) SWCNT and (b) MWCNT (He et al., 2013).	65
2.20	Schematic diagrams of different types of functionalizations.	67
2.21	Schematic diagrams of different types of functionalization's.	69

3.1	Acid functionalization of MWCNT produced carboxyl (–COOH) and hydroxyl (–OH) functional groups on the MWCNT surface.	79
3.2	Microwave experimental setup diagram for dope solution preparation .	81
3.3	Pictorial representation of cross flow UF cell.	88
3.4	Schematic diagram of dialysis cell.cell.	93
3.5	The schematic diagram of HF spinning apparatus.	95
3.6	Schematic diagram of HF dialysis system.	98
3.7	The flow chart of the methodology.	98
4.1	FTIR spectra of MWCNT and f-MWCNT.	101
4.2	The XRD analysis of MWCNT and f-MWCNT.	102
4.3	FTIR spectra of the various NC's formulations as listed in step 1 of the Table 3.1.	102
4.4	FTIR spectra of fabricated membranes, a) PES/f-MWCNT/PVP-k90 based membranes (M-05-3k9 to M-2-3k3) and B) PES/f-MWCNT/PVP-k30 based membrane (M-1-3k3).	103
4.5	Schematic representation of PES/f-MWCNT/PVP NC's based membrane.	104
4.6	Cross-sectional and surface FESEM pictures of PES/f-MWCNT/PVP-k90 and PVP-k30 based HD membranes at 500X magnification power.	106
4.7	Cross-sectional FESEM pictures of M-CNT and f-MWCNT/PVP-k30 based NCs based membranes at 500X.	107

4.8	EDX spectra of PES and PES/NCs membranes revealing C, S, O and N as significant elements.	108
4.9	Influence of various combination of f-MWCNT on (a) Contact angles; and (b) water absorbance results on pristine PES, PES-CNT and PES-NC's based membranes (n=3).	109
4.10	Influence of f-MWCNT on (a) pore size diameter; and (b) porosity measurements of different fabricated membranes.	111
4.11	The rejection rates and MWCO graphs; a) PES and PES/f-MWCNT/PVP-k90 based membranes and, b) M-CNT and PES/f-MWCNT/PVP-k30 based membranes, c) MWCO results of all formulated membranes (n=3).	113
4.12	Effect of f-MWCNT on the leaching ratio of all formulated based membrane.	114
4.13	The flux rate of all formulated NC's based membrane.	116
4.14	Surface roughness data and 3D micrograph of PES and PES/NC's based membranes obtained by AFM technique.	117
4.15	Protein adhesion ($\mu\text{g}/\text{cm}^2$) results of PES, PES-CNT and different NCs based membranes ($\text{SD}\pm 0.35$, n=3), (a) BSA adhesions; and (b) Lysozyme adhesions.	119
4.16	The thrombogenic properties of the all fabricated membrane (Table 3.2 and 3.3); (a) prothrombin time (PT), (b) thrombin time (TT), (c) activated partial thrombin time (APTT), and (d) whole blood clotting time (WBCT) results (n=3).	121
4.17	Urea, creatinine and lysozyme dialysis data of PES, PES-CNT and all PES/NC's based membranes.	123
5.1	FTIR spectra of PES and S-PES membranes.	128

5.2	The FTIR spectrums of MWCNT, f-MWCNT, NCs and different PES/ NCs and S-PES membranes.	129
5.3	The schematic representation of the chemical arrangement of S-PES and PES/f-MWCNT/PVP NC's based membrane.	130
5.4	(a) FESEM cross-sectional images of PES and S-PES/PES/NC's based membranes (M-0 to M-1-50); and (b) M-2-0 to M-2-50 membranes.	132
5.5	(a)- The contact angle; and (b)- water absorption results of sulfonated and non-sulfonated membranes.	134
5.6	The quantitative measurements of different statistics of surface roughness parameters, reading via XEI standard software of scanned AFM images that provide the information of; (a) root mean square (Rq), (b) surface roughness (Ra), (c) difference in heigh of peaks (Rpv), and (d) ten points average roughness (Rz).	135
5.7	The 3D-AFM images of fabricated membranes.	136
5.8	The rejection rates of different formulated membranes (n=3).	138
5.9	The MWCO results of PES and sulfonated based membranes (n=3).	138
5.10	(a)- Pore size; and (b)- porosity measurements of the non-sulfonated and sulfonated membranes.	139
5.11	Flux rate estimations of sulfonated and non-sulfonated membranes.	140
5.12	The leaching ratio of all formulated NCs membrane.	141

5.13	Protein adhesion (ug/cm ²) results of different NCs based membranes (SD±0.35, n=3), (a) BSA adhesions; and (b) Lysozyme adhesions.	142
5.14	The thrombogenic properties of the fabricated membrane (Table 3.4); (a) prothrombin time (PT), (b) thrombin time (TT), (c) activated partial thrombin time (APTT), and (d) whole blood clotting time (WBCT) results (n=3).	144
5.15	The hemolysis results of the fabricated membranes (n=3).	145
5.16	The dialysis clearance percentage of urea, creatinine and lysozyme by PES based membranes (n=3).	147
6.1	The side and cross sectional FESEM images of HF membranes at 500 and 300X magnification.	151
6.2	The solute rejection and log normal pore radius graphs of HF membranes.	153
6.3	The MWCO and pore diameter calculations of HF membranes.	154
6.4	The flux rate of fabricated membranes.	155
6.5	The dialysis clearance percentage of urea, creatinine and lysozyme by PES based HF's membranes (n=3).	158

LIST OF ABBREVIATIONS

AA	-	Acrylic acid
AFM	-	Atomic force microscopy
AN	-	Acrylonitrile
APTT	-	Activated partial thrombin time
β 2-m	-	Beta-2- microglobin
BSA	-	Bovine serum albumin
BPA	-	Bisphenol-A
BDO	-	1,4-butanediol
C	-	Carbon
CA	-	Contact angle
CNT	-	Carbon nanotube
-COOH	-	Carboxylic group
ClSO_3H	-	Chlorosulfuric acid
CKD	-	Chronic kidney disease
PEO	-	Polyethylene oxide
CPES	-	Carboxylic polyethersulfone
DNA	-	Deoxyribonucleic acid
DI	-	De-ionized
DMF	-	Dimethyleformamide
DNA	-	Deoxyribonucllic acid

DG	-	Degree of grafting
DS	-	Degree of sulfonation
EDX	-	Energy dispersive X-ray spectroscopy
EG	-	Ethylene glycol
FESEM	-	Field emission scanning electron microscopy
FDA	-	Food and Drug Administration
f-MWCNTs	-	Functionalized multiwall carbon nanotubes
FTIR	-	Fourier transforms infrared spectroscopy
FRSP	-	Free radical solution polymerization
GO	-	Graphene oxide
HD	-	Hemodialysis
HP	-	Hemolytic percentage
IEC	-	Ion exchange capacity
Max	-	Maximum height
mPEG	-	Methoxypoly (ethylene glycol)
M_{dry}	-	Dry membrane
M_{wet}	-	Wet membrane
MF	-	Microfiltration
Mid	-	Average between the minimum and maximum height
Min	-	Minimum height
MPC	-	Methacryloyloxyethyl-phosphoryl-choline
MWCNT	-	Multiwall carbon nanotube
N	-	Nitrogen
NCs	-	Nanocomposites
NF	-	Nanofiltration

Nps	-	Nanoparticles
O	-	Oxygen
OH	-	Hydroxyl
P	-	Poly
PA	-	Polyamides
PAN	-	Polyacrylonitrile
PBS	-	Phosphate-buffered solution
PC	-	Phosphorylcholine
PES	-	Polyethersulfone
PEG	-	Polyethylene glycol
PEO	-	Poly ethyleneoxide
PMMA	-	Polymethyl methacrylate
Pmp	-	Persons per million populations
PP	-	Polypropylene
PT	-	Prothrombin time
PRT	-	Plasma recalcification time
PSf	-	Polysulfone
PU	-	Polyurethanes
PVP	-	Polyvinylpyrrolidone
RAFT	-	Reversible addition fragmentation chain transfer polymerization
RO	-	Reverse osmosis
SO ₃	-	Sulphur trioxide
-SO ₃ H	-	Sulfonic group
SiO ₂	-	Silicon dioxide

SDS	-	Sodium dodesylesulphate
Semi-IPN	-	Semi-interpenetrating network
SMA	-	Poly (styrene-alt-maleic anhydride)
S-PES	-	Sulfonated polyethersulfone
SPEES	-	Sulfonated polyether-ethersulfone
St	-	Styrene
SWCNTs	-	Single-walled carbon nanotubes
TGA	-	Thermogravimetric analysis
TiO ₂	-	Titanium dioxide
TT	-	Thrombin time
UF	-	Ultrafiltration
VP	-	Vinylpyrrolidone
WBCT	-	Whole blood clotting time
XRD	-	X-Ray Diffraction
ZrO ₂	-	Zirconium dioxide

LIST OF SYMBOLS

A	-	Area of the flat sheet membrane (m^2)
C_f	-	Concentrations in feed solution (%)
C_p	-	Concentrations in permeate solution (%)
C_0	-	Concentration, when time = 0 (%)
C_t	-	Concentration, when time = 1-4 hours (%)
d_o	-	Outer diameter of fiber (μm)
D_{pc}	-	Absorbance of the positive control
D_t	-	Absorbance of the negative control
D_w	-	Solute diffusion coefficient
δ	-	Membrane thickness (μm)
ϵ	-	Porosity
$f(q)$	-	Friction coefficients
$-\Delta GSL$	-	Surface free energy
J_p	-	Protein flux ($Lm^{-2}h^{-1}$)
J_w	-	Water flux ($Lm^{-2}h^{-1}$)
L	-	Hydraulic permeability
l	-	Effective length of fiber (m)
M	-	Mole
N	-	Number of the fiber
n	-	Number of membranes

π	-	Pi
P_m	-	Diffusive permeability
R	-	Protein rejection (%)
R_a	-	Roughness average (nm)
R_{FR}	-	Flux recovery ($Lm^{-2}h^{-1}$)
R_{ir}	-	Irreversible resistance
r_p	-	Pore radius (nm)
R_{pv}	-	Peak-to-valley line (nm)
R_q	-	Root-mean-squared roughness (nm)
R_r	-	Reversible resistance (%)
R_t	-	Total resistance (%)
r_m	-	Average pore radius (nm)
r_s	-	Radius of solute (nm)
R_z	-	Ten points average roughness (nm)
S	-	Surface area of the membrane (cm^2)
SF	-	Factor of steric hindrance
σ	-	Staverman reflection coefficient
μ	-	Micro
V	-	Volume (m^3)
δ_w	-	Density of water (g/cm^3)
γ_L^T	-	Total surface tension of water

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	List of publications	197

CHAPTER 1

INTRODUCTION

1.1 Overview

The developments in science and technology have assisted mankind to live longer and consumed a large amount of world's resources. However, the need to sustain life requires a good health condition with better and advance curing methods and therapy. Human life is invaluable and there are various factors that affect the survival and quality of life. Amongst them are diseases that damaged the human kidneys. Kidney, is a bean shaped organ, made up by functional units, called nephrons, which received the blood from the renal arteries and after processing forced the waste solutes and fluids to move into the urinary bladder and return the purified blood to the body circulation. The two key mechanisms are served by the kidney. One is the plasma filtration via UF technique that separate the extracellular fluid in the glomeruli and the second is the removal of extra water through active and passive tubular transport system that also contain dissolved uremic solutes and electrolytes (Findlay et al., 2015).

The origin of kidney disease may be metabolic, vascular, immunologic, degenerative, infectious or genetic. The renal failure might be chronic that last up to months or years or acute stay for some days or weeks. Acute failure is generally related with tubular necrosis, acute glomerulonephritis, ischemia or poisoning with nephrotoxins that involved excessive loads of free hemoglobin, aminoglycosides and heavy metals. The chronic renal failure is typically initiated by hypertension, vascular disease (e.g. diabetes) or chronic glomerulonephritis. If the proper treatment

is not taken for acute renal failure, then it will cause the chronic renal failure. The human kidney can continue to work until 90% of its function has been lost and keep deteriorating and renal failure exceeds 95%. At this stage, survival becomes impossible without the replacement therapy (Findlay et al., 2015).

Supportive therapy like kidney transplant, peritoneal dialysis or hemodialysis (HD) are required to prolong life when the human kidney fails to work either as a result of painful injury or any other disease. Globally, the number of ESRD patients was estimated to be 3,010,000 with \cong 7% growth rate at the end of 2012. Based on a previous report, 89% of dialysis patients opted for HD and only 11% chose peritoneal dialysis (National Institute of Diabetes and Digestive and Kidney Diseases).

Hemodialysis (HD) method is a significant clinical therapy that can eliminate toxic metabolites from the blood of patients whose kidney fail to perform and the patient suffered into end stage renal diseases (ESRD) (Miller et al., 2010). This method has benefited approximately more than one million people per year all over the world and the prevalence rate of ESRD is increasing globally (Saran et al., 2015). The annual renal data report of United State has covered 57 countries and showed that the ESRD incidence rates varied significantly across the globe. United States, Mexico and Taiwan described the maximum occurrence of treated ESRD at 363-458 persons per million population (pmp). Indonesia, Thailand, Portugal, Republic of Korea, Japan and Singapore demonstrated 208–308 pmp, whereas other countries showed less than 96 reported incidence rates of treated ESRD patients. The highest growths in the treatment of ESRD were observed in Mexico (122%), Malaysia (176%), Philippines (185%), Russia (249%), Bangladesh (629%) and Thailand (1210%) in 2012/2013 period (Saran et al., 2015).

According to the U.S. Renal data system (2014) that the medical expenditure of ESRD patients in the USA increased from \$30.4 billion to \$30.9 billion from 2012 to 2013 with 1.6% growth rate. Whereas, the total global expenditures for ESRD patients reached up to \$437 billion. It is also estimated that the growth rate of ESRD patients will increase in the future due to different diseases, poor quality lifestyles

and improper medical treatments (Saran et al., 2015). Since the necessity of HD dialysis will be increasing in the future, thus the dialysis therapy is considered as a multi-million dollar industry (National Institute of Diabetes and Digestive and Kidney Diseases).

In Malaysia in 2014, the total registered patients undergoing dialysis were 34,767 and 6107 new HD cases were registered during the same year. In the last ten years, the rate of dialysis acceptance was doubled and reached 203 pmp. Table 1.1 represents the short summary of the registered and treatment-type of dialysis patients in Malaysia. Moreover, there are 758 dialysis centers in the whole country and 53.2% were funded by government, 12.9% and 31.2% by charity and self supported, respectively (Goh et al., 2016).

Table 1.1: The registered dialysis patients and the type of treatment supplied to ESRD in Malaysia (Goh et al., 2016).

Year	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014
New Dialysis patients	3167	3709	4103	4640	4952	5305	6073	6690	6985	7055
New Transplants	172	151	112	131	141	128	127	107	98	81
Dialysis deaths	1515	1820	1987	2191	2601	3047	3292	3645	4001	4015
Transplant deaths	49	58	47	59	49	48	55	64	56	45
Dialyzing at 31st December	13356	15080	17084	19388	21590	23709	26328	29223	32026	34767
Functioning transplant at 31st December	1716	1771	1788	1808	1852	1881	1907	1891	1870	1844

During the last decades, membrane technology plays a central role in the purification and separation of the biotechnological products. The use of membrane based modules progressively enhances both industrial processes and academic science of engineering (van Reis et al., 2007). The membrane filtration system offers robust performance, easy availability, less processing time and low cost compared to

the other available techniques. These benefits extend the microfiltration (MF) and ultrafiltration (UF) membrane systems into various biomedical applications like hemofiltration, hemodiafiltration, hemodialysis, plasma collection and plasmapheresis (Samtleben et al., 2003; Tullis et al., 2002; Werner et al., 1996; Zhao et al., 2013).

The innovation of asymmetry membrane structures introduced the new era. Nowadays, a number of new polymeric materials are available whose properties can be upgraded by the use of proper additives and new formulation. In 2004, Membrana GmbH launched a performance enhancement technology, in which they used spacer yarns woven pattern in HF bundles and optimized the dialysate flow (Membrana, 2004). Asahi Kasei Medicals Co. Ltd. had launched a PSf dialyzer modified by vitamin-E to reduce the oxidative stress, which reduced the tissue damaging and also shows the good biocompatibility to the body immune system during HD therapy (Sasaki, 2006). Some studies suggested that high flux dialyzer membrane was better due to a lower death rate than the low flux dialyzers (Bloembergen et al., 1999; Woods et al., 2000). The Eknoyan et al. (2002) reported a HD study of 72 patients in 15 clinical centers and checked the mortality and morbidity. He found that neither the high flux rate nor higher dialysis dose enhanced the mortality and morbidity rate among patients. Besides, no much difference was found in the clearance rate of urea solute for the both high and low flux membranes (Eknoyan et al., 2002). The previous study of Bonomini et al. (1995) also provided the same results as Eknoyan et al. (2002) (Bonomini et al., 1995). Moreover, House et al., (2000) found that high flux membranes, increased the cardiovascular risk in the ESRD patients.

The chief constituent of a hemodialysis instrument is a semipermeable membrane, whose chemical composition has great effect on a patient's health either in terms of membrane biocompatibility or uremic waste removal (Daugirdas et al., 2012; Kumar et al., 2014; Nie et al., 2012; Zhao et al., 2013). Some of the most commonly used materials to make HD membranes include PSf, PES, polyamide, ethylene vinyl alcohol copolymers, cellulose triacetate, polymethylmethacrylate and polyacrylonitrile (Matsuda, 2011). Currently, synthetic polymers are used in most of the dialyzer membranes, 93% of which are derived from the parent polyarylsulfone

family, with 22% produced from PES and 71% from PSf (Bowry et al., 2010). Each polymeric membrane has its own advantages or disadvantages and complications may arise, when the membranes are judged only by their polymer names. Due to the varying membrane compositions, membranes with the same polymer names may differ in their adsorption, flux properties and hemocompatibility characteristics. Membranes in the new super high-flux dialyzers are primarily PSf and PES and future trends to use PES as main hemodialyzer materials are increasing because PES has an equivalent property to that of PSf, but it is considered as a bisphenol-A free membrane (Matzke et al., 2011; Yamasaki et al., 2001).

PES has a higher atomic weight ratio of sulfone groups, which makes it more mechanical resistant, heat resistant and additional hydrophilic than that of PSf (Abe et al., 2011). PES is highly amorphous, transparent thermoplastic, comparatively less flammable, chemical resistant, relatively hydrophobic and has less water sorption (0.8% at 50% relative humidity) (Kesting, 1985). In biomedical fields PES membranes are broadly employed for artificial organs and medical devices used for purification of blood like hemofiltration, hemodiafiltration, hemodialysis, plasma collection and plasmapheresis (Samtleben et al., 2003; Tullis et al., 2002; Werner et al., 1996; Zhao et al., 2013).

In order to improve the performance of PES membrane in terms of biocompatibility and uremic solute removal, PVP was frequently used with PES and very little work was reported relating to PVP-K30. PVP is a highly hydrophilic and water-soluble polymer that prevents the protein adsorption and act as a pore forming agent on the membrane surface (Yang et al., 2008). The high affinity of PVP may cause it to swell in aqueous media and then it is eluded during phase inversion and blood dialysis. Thus, it may be possible that HD property of the membrane is altered during dialysis therapy (Sun et al., 2009). In order to maintain the composition of the membrane and retention of PVP as an additive in PES, PVP was used in the form of mono, di and tri-blocks such as poly(styrene-co-acrylic acid)-b-poly(vinyl pyrrolidone)-b-poly(styrene-co-acrylic acid) (Remes et al., 1992), poly(vinyl pyrrolidone)-b-poly(methyl methacrylate)-b-poly(vinyl pyrrolidone) (Ran et al., 2011), poly(vinyl pyrrolidone-acrylonitrile-vinyl pyrrolidone) (Yin et al., 2012),

poly(acrylonitrile-co-acrylic acid) (Fang et al., 2009), PVP nanoparticles (Weifeng-Zhao et al., 2011) and PVP-k90 alone (Barzin et al., 2004).

The use of nanoparticles and nanotubes has revealed outstanding potential in biological systems and membranes. In view of this MWCNTs are verified and reported as a revolutionary choice in biomedical field such as biomolecular transporters and recognition devices (Shi Kam et al., 2004; Wilson et al., 2009), biosensors (Krauss, 2009), cancer therapy and diagnoses (De La Zerda et al., 2008; Liu et al., 2009). It is believed that the high surface to volume ratio of MWCNT can improve the overall performance of many polymer-CNT composite membranes. Moreover, MWCNT can be easily functionalized with different functional groups and provide sites for attachment or bonding to other incoming molecules and open up new applications (Vivekchand et al., 2002). Both covalent and non-covalent alterations of the MWCNT surfaces have been employed to improve the solubility and wetting of MWCNT. The non-covalent approach includes surfactant modifiers (Kang et al., 2003), polymer absorption (Gómez et al., 2003), and polymer wrapping (Star et al., 2001). The benefit of noncovalent connection is that the perfect structure of the MWCNT is maintained and the wrapped or absorbed compound is not damaged, and thus most of the properties remained intact (Chen et al., 2005).

1.2 Problem Statement

Koopman et al., (2008) reported that current HD therapy failed to replace the complicated functions of the kidney in ESRD patients. Numerous research has been carried out in this field, however, patients suffer in chronic kidney disease were still suffering from low sickness situations, normal life quality and low death rate. The ESRD patients also suffered from chronic malnutrition, short life period and high cardiovascular risks (Koopman et al., 2008). Although, the HD technology had been used as a replacement therapy for renal failure for a few decades, it still has a number of unsolved issues.

Blood proteins have a propensity to rapidly adsorb onto the surface of the polymer, when PES-based hemodialysis membranes come into contact with blood (Klinkmann et al., 1987). Consequently the adsorbed protein layer demonstrated unwanted results such as higher platelet adhesion, fast blood coagulation and aggregation (Fang et al., 2009; et al., 2009; Zhu et al., 2007). The biocompatibility of the pristine PES was not satisfying (Klinkmann et al., 1987; Liu et al., 2009; Samtleben et al., 2003; Tullis et al., 2002; Zhao et al., 2013), thus the quest for simple additives and modification methods to improve such property continued and is a challenge for membranologists.

Most of the results obtained from di or tri block of PVP suggested that they can be used in hemodialysis, but their actual dialysis performance in terms of blood compatibility are not well reported in the literature. Moreover, the preparation of the covalently combined di or tri block of PVP require highly specific and controlled conditions. Among the various modification techniques, the blending of PVP, is the simplest method to improve the biocompatibility of PES membranes, but pure PVP cannot be used directly and need slight modification for its permanent residence on the HD membrane (Matsuda et al., 2008).

The MWCNTs are highly hydrophobic and non-polar material and its direct use without surface modification or surfactants may cause agglomeration and poor dispersion in membrane matrix, leading to defective product. The properties of MWCNT can be easily improved or upgraded by the attachment of different functional groups (Bahr et al., 2001; Georgakilas et al., 2002; Pekker et al., 2001; Valcárcel et al., 2008; Ying et al., 2003). Acid treatment is the simplest chemical method to create the polarity and attachment of the hydrophilic functional groups (carboxyl and hydroxyl) with MWCNT surface. This hydrophilic-hydrophobic nature of acid functionalized f-MWCNT provide a dual character, that can attract and bind both hydrophobic (like PES polymer) and hydrophilic types of chemicals (e.g. PVP type hydrophilic compounds) by π - π stacking and dipole-dipole interactions, respectively (Dyke et al., 2004a; Lu et al., 2011; Ma et al., 2010; Qian et al., 2000; Xie et al., 2005).

Most of the blood proteins are negatively charged and thus exhibit reduced adsorption behavior towards anionic character polymers. Sulfonation of PES is the bulk modification method which can increase the percentage of negative charge polarity due to the addition of sulfonated ($-\text{SO}_3\text{H}$) group. The presence of sulfonate functional groups in the polymer reduced the blood and membrane interaction via steric repulsion and showed good anticoagulant activity. Sulfonation is a chemical modification method of the PES that enhanced its biocompatibility and hydrophilicity. Many studies had focused on the sulfonation of polymers that increased the anionic character (Nie et al., 2014; Wang et al., 2009). Gertz et al., (2005) found that S-PES polymer reduced the contact activation of blood, whereas Wang et al., (2009) reported that blending of S-PES reduced the protein adsorption and elevated the blood coagulation time.

There are many dialysis membranes on the market, but the high death ratio of ESRD patients is suggesting that more research need to be done on this issue (Saran et al., 2015). Moreover, a few researchers reported the self-synthesized performance of HD membranes and most of the published work are focused on the performance investigation of commercial dialyzer membranes (Jalal Barzin et al., 2004; Leyboldt et al., 2006; Li et al., 2012; Su et al., 2008). In this research work, keeping in view the above discussion and literature review, an effort has been made to develop a highly biocompatible HD membrane composed of PES as a basic polymer with improved uremic solute permeability. Since the biocompatibility of PES HD membranes does not depend on a particular factor, hence different techniques are utilized to improve this property. Some of the researchers have focussed to chemical modification of PES polymer and others have used different additives (Ran et al., 2011; Wang and Yang, et al., 2009). The use of both methods, polymer modification and blending of additives together; have attracted much attention to improving the PES biocompatibility and uremic solute removal ability.

In this work, effort was made to improve the performance and biocompatibility of PES membranes by the chemically modified sulfonated PES and blending of novel nanocomposites (NCs) additives. The acid functionalized MWCNT and lower and higher molecular weight of PVP grades (PVP-k30 and PVP-

k90) was used for NCs preparation and then they were incorporated into PES and S-PES/PES based membranes. The hydrophilic part of f-MWCNT contributed to the -COOH and -OH groups, whereas sulfonated polymer provided the -SO₃H group in the membrane composition. The S-PES increased the anionic character and hydrophilicity of the HD membrane and favor the reduced protein adhesion. While in the NC's, the PVP tended to enhance the biocompatibility and hydrophilicity and f-MWCNT provided the mechanical strength, reduced the PVP leaching and act as a bridging material between PVP and PES. Moreover, the PVP was also reported as a surfactant for carbon nanotubes that improved its dispersibility and reduced the agglomeration in different solvents. Therefore, PVP in situ with acid treated MWCNT might also improve the dispersion properties of MWCNT in the formulated membranes (Vatanpour et al., 2011).

Thus, the purpose of this research work is to contribute in the development of a new HD membrane with a combination of additives, which might be able to reduce biocompatible issues and exhibits higher rates of uremic solute clearances

1.3 Objective of the Study

The main objective of the study is to develop a high performance, biocompatible dialysis membrane with notable uremic solute sieving properties by blending f-MWCNT into the polymer matrix. In order to achieve this the following objectives, need to be addressed;

1. To formulate and synthesize various flat sheet HD membranes consisting of
i) PES as base polymer and NC's which consists of f-MWCNT/PVP-k90 and f-MWCNT/PVP-k30. ii) Various ratios of PES and S-PES as the base polymer and the NCs which comprise of f-MWCNT/PVP-k90.
2. To characterize the synthesized membranes in terms of chemical composition, biocompatibility and to evaluate their dialysis performance using uremic solute model solution.

3. To fabricate HD hollow fiber membrane using the best formulation and analyse its dialysis performance.

1.4 Scope of the Study

The scope of research is as follows

- i. Acid functionalization of MWCNT and NC's synthesis by blending technique and covalent attachment of sulfonated group to the PES to develop S-PES polymer. The FTIR and XRD were used to characterize the MWCNT, f-MWCNT, NC's and S-PES polymer.
- ii. Preparation of i) various concentrations of NC's and PES alone, ii) various ratios of S-PES and PES polymer with NC's via microwave technique.
- iii. Flat sheet membranes were casted and their chemical properties were analysed by FTIR, whereas FESEM was used for morphological studies.
- iv. The performance of HD membranes was initially evaluated in terms of pure water permeation, the molecular weight cutoff (MWCO) (should be <60kDa), mean pore size, porosity and flux recovery ratio.
- v. The leaching tests were performed to observe the stability of the NC's in the fabricated membranes
- vi. The membrane hydrophilicity were determined by contact angle and water uptake measurements. In addition, AFM technology was utilized for the surface roughness study and 3D micrographs.

- vii. In order to evaluate the biocompatibility of HD membranes, various experiments are performed; protein resistance ability of FSMs, hemolysis (destruction of red blood cells), blood coagulation factors, including Thrombin time (TT), Prothrombin time (PT), Activated partial thrombin time (APTT), fibrin formation, and the plasma re-calcification time.
- viii. The single layer dialysis cell is used to estimate the dialysis properties formulated membranes using a model solution of uremic solutes consisting of urea, creatinine and lysozyme against distilled water as dialysate.
- ix. Finally, hollow fiber membranes were fabricated using the best FSM formulation. Their performances were evaluated in terms of molecular weight cutoff, pure water permeation, pore size, antifouling properties and dialysis ability against uremic solutes.

1.5 Significance of the Study

In this research a novel HD membrane with excellent biocompatibility, good flux rate, better antifouling properties and excellent dialysis performance was synthesized and produced. PVP and the f-MWCNT based di-block nonocomposites were developed by simple blending method and its influence on PES and combine PES/S-PES polymeric membranes were thoroughly investigated with special emphasis on membrane biocompatibility, consistency of chemical composition, dialysis and typical HD performances. The biocompatibility tests include the study of protein adhesion, red blood cell behavior and fibrin formation towards the formulated membranes. Although previous researchers have used different modification techniques to enhance the performance of PES, but none of them has reported a comprehensive study in terms of typical membrane performance, biocompatible studies and dialysis of uremic solutions. Finally, the performance data

REFERENCES

- Abe, T., Kato, K., Fujioka, T., & Akizawa, T. (2011). The blood compatibilities of blood purification membranes and other materials developed in Japan. *International journal of biomaterials*, 2011.
- Acchiardo, S., Kraus Jr, A., & Jennings, B. (1989). Beta 2-microglobulin levels in patients with renal insufficiency. *American journal of kidney diseases: the official journal of the National Kidney Foundation*, 13(1), 70.
- Adeli, M., Mirab, N., Alavidjeh, M. S., Sobhani, Z., & Atyabi, F. (2009). Carbon nanotubes-graft-polyglycerol: biocompatible hybrid materials for nanomedicine. *Polymer*, 50(15), 3528-3536.
- Alshehri, R., Ilyas, A. M., Hasan, A., Arnaout, A., Ahmed, F., & Memic, A. (2016). Carbon Nanotubes in Biomedical Applications: Factors, Mechanisms and Remedies of Toxicity. *Journal of medicinal chemistry*.
- Altinkaya, S. A., & Ozbas, B. (2004). Modeling of asymmetric membrane formation by dry-casting method. *Journal of Membrane Science*, 230(1), 71-89.
- Ambalavanan, S., Rabetoy, G., & Cheung, A. K. (1999). High efficiency and high flux hemodialysis. *Atlas of Diseases of the Kidney*, 5, 1-10.
- Amenta, V., & Aschberger, K. (2015). Carbon nanotubes: potential medical applications and safety concerns. *Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology*, 7(3), 371-386.
- Amiji, M., & Park, K. (1993). Surface modification of polymeric biomaterials with poly (ethylene oxide), albumin, and heparin for reduced thrombogenicity. *Journal of Biomaterials Science, Polymer Edition*, 4(3), 217-234.
- Anderson, J. M. (1993). Mechanisms of inflammation and infection with implanted devices. *Cardiovascular Pathology*, 2(3), 33-41.
- Apartsin, E. K., Buyanova, M. Y., Novopashina, D. S., Ryabchikova, E. I., Filatov, A. V., Zenkova, M. A., & Venyaminova, A. G. (2014). Novel multifunctional hybrids of single-walled carbon nanotubes with nucleic acids: Synthesis and

- interactions with living cells. *ACS applied materials & interfaces*, 6(3), 1454-1461.
- Aucella, F., Gesuete, A., Vigilante, M., & Prencipe, M. (2013). Adsorption dialysis: from physical principles to clinical applications. *Blood purification*, 35(Suppl. 2), 42-47.
- Bahr, J. L., Yang, J., Kosynkin, D. V., Bronikowski, M. J., Smalley, R. E., & Tour, J. M. (2001). Functionalization of carbon nanotubes by electrochemical reduction of aryl diazonium salts: a bucky paper electrode. *Journal of the American Chemical Society*, 123(27), 6536-6542.
- Bai, P., Cao, X., Zhang, Y., Yin, Z., Wei, Q., & Zhao, C. (2010). Modification of a polyethersulfone matrix by grafting functional groups and the research of biomedical performance. *Journal of Biomaterials Science, Polymer Edition*, 21(12), 1559-1572.
- Balan, V., & Verestiuc, L. (2014). Strategies to improve chitosan hemocompatibility: A review. *European Polymer Journal*, 53, 171-188.
- Balasubramanian, K., & Burghard, M. (2005). Chemically functionalized carbon nanotubes. *Small*, 1(2), 180-192.
- Barth, C., Goncalves, M., Pires, A., Roeder, J., & Wolf, B. (2000). Asymmetric polysulfone and polyethersulfone membranes: effects of thermodynamic conditions during formation on their performance. *Journal of Membrane Science*, 169(2), 287-299.
- Barzin, J., Feng, C., Khulbe, K., Matsuura, T., Madaeni, S., & Mirzadeh, H. (2004). Characterization of polyethersulfone hemodialysis membrane by ultrafiltration and atomic force microscopy. *Journal of Membrane Science*, 237(1), 77-85.
- Barzin, J., Madaeni, S., Mirzadeh, H., & Mehrabzadeh, M. (2004). Effect of polyvinylpyrrolidone on morphology and performance of hemodialysis membranes prepared from polyether sulfone. *Journal of applied polymer science*, 92(6), 3804-3813.
- Basmadjian, D., Sefton, M. V., & Baldwin, S. A. (1997). Coagulation on biomaterials in flowing blood: some theoretical considerations. *Biomaterials*, 18(23), 1511-1522.
- Bauer, K., & Rosenberg, R. (1995). Control of coagulation reactions. *Williams Hematology, 5th Ed.(McGraw Hill Inc.)*, 1239-1251.

- Bekyarova, E., Ni, Y., Malarkey, E. B., Montana, V., McWilliams, J. L., Haddon, R. C., & Parpura, V. (2005). Applications of carbon nanotubes in biotechnology and biomedicine. *Journal of biomedical nanotechnology*, *1*(1), 3-17.
- Benavente, J., García, J. M., Riley, R., Lozano, A. E., & de Abajo, J. (2000). Sulfonated poly (ether ether sulfones): characterization and study of dielectrical properties by impedance spectroscopy. *Journal of Membrane Science*, *175*(1), 43-52.
- Besteman, K., Lee, J.-O., Wiertz, F. G., Heering, H. A., & Dekker, C. (2003). Enzyme-coated carbon nanotubes as single-molecule biosensors. *Nano letters*, *3*(6), 727-730.
- Bhushan, B., & Jung, Y. C. (2011). Natural and biomimetic artificial surfaces for superhydrophobicity, self-cleaning, low adhesion, and drag reduction. *Progress in Materials Science*, *56*(1), 1-108.
- Bitter, J. G. (2012). *Transport mechanisms in membrane separation processes*: Springer Science & Business Media.
- Blanco, J., Nguyen, Q., & Schaetzel, P. (2001). Novel hydrophilic membrane materials: sulfonated polyethersulfone Cardio. *Journal of Membrane Science*, *186*(2), 267-279.
- Blockmans, D., Deckmyn, H., & Vermynen, J. (1995). Platelet actuation. *Blood reviews*, *9*(3), 143-156.
- Bloembergen, W. E., Hakim, R. M., Stannard, D. C., Held, P. J., Wolfe, R. A., Agodoa, L. Y., & Port, F. K. (1999). Relationship of dialysis membrane and cause-specific mortality. *American Journal of Kidney Diseases*, *33*(1), 1-10.
- Bonomini, V., Coli, L., Scolari, M., & Stefoni, S. (1995). Structure of dialysis membranes and long-term clinical outcome. *American journal of nephrology*, *15*(6), 455-462.
- Bosi, S., Ballerini, L., & Prato, M. (2013). Carbon nanotubes in tissue engineering *Making and Exploiting Fullerenes, Graphene, and Carbon Nanotubes* (pp. 181-204): Springer.
- Bottini, M., Bruckner, S., Nika, K., Bottini, N., Bellucci, S., Magrini, A., Mustelin, T. (2006). Multi-walled carbon nanotubes induce T lymphocyte apoptosis. *Toxicology letters*, *160*(2), 121-126.

- Bowry, S. K., Gatti, E., & Vienken, J. (2010). Contribution of polysulfone membranes to the success of convective dialysis therapies. *Contributions to nephrology*, 173, 110-118.
- Buang, N. A., Fadil, F., Majid, Z. A., & Shahir, S. (2012). Characteristic of mild acid functionalized multiwalled carbon nanotubes towards high dispersion with low structural defects. *Digest Journal of Nanomaterials and Biostructures*, 7(1), 33-39.
- Byun, I., Kim, I., & Seo, J. (2000). Pervaporation behavior of asymmetric sulfonated polysulfones and sulfonated poly (ether sulfone) membranes. *Journal of applied polymer science*, 76(6), 787-798.
- Caminati, W., Dell'Erba, A., Maccaferri, G., & Favero, P. G. (1998). Free Jet Absorption Millimeter Wave Spectrum of Pyrrolidine: Assignment of a Second, Equatorial, the Most Stable Conformer. *Journal of molecular spectroscopy*, 191(1), 45-48.
- Capila, I., & Linhardt, R. J. (2002). Heparin–protein interactions. *Angewandte Chemie International Edition*, 41(3), 390-412.
- Cassatella, M. A. (1995). The production of cytokines by polymorphonuclear neutrophils. *Immunology today*, 16(1), 21-26.
- Castro Vidaurre, E., Achete, C., Simao, R., & Habert, A. (2001). Surface modification of porous polymeric membranes by RF-plasma treatment. *Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms*, 175, 732-736.
- Chamoulaud, G., & Bélanger, D. (2004). Chemical modification of the surface of a sulfonated membrane by formation of a sulfonamide bond. *Langmuir*, 20(12), 4989-4995.
- Chatterjee, N., Yang, J., Kim, H.-M., Jo, E., Kim, P.-J., Choi, K., & Choi, J. (2014). Potential toxicity of differential functionalized multiwalled carbon nanotubes (MWCNT) in human cell line (BEAS2B) and *Caenorhabditis elegans*. *Journal of Toxicology and Environmental Health, Part A*, 77(22-24), 1399-1408.
- Chen, G.-X., Kim, H.-S., Park, B. H., & Yoon, J.-S. (2005). Controlled functionalization of multiwalled carbon nanotubes with various molecular-weight poly (l-lactic acid). *The Journal of Physical Chemistry B*, 109(47), 22237-22243.

- Chen, Z., Deng, M., Chen, Y., He, G., Wu, M., & Wang, J. (2004). Preparation and performance of cellulose acetate/polyethyleneimine blend microfiltration membranes and their applications. *Journal of Membrane Science*, 235(1), 73-86.
- Cheng, C., Müller, K. H., Koziol, K. K., Skepper, J. N., Midgley, P. A., Welland, M. E., & Porter, A. E. (2009). Toxicity and imaging of multi-walled carbon nanotubes in human macrophage cells. *Biomaterials*, 30(25), 4152-4160.
- Cheng, Y., Li, W., Fan, X., Liu, J., Xu, W., & Yan, C. (2013). Modified multi-walled carbon nanotube/Ag nanoparticle composite catalyst for the oxygen reduction reaction in alkaline solution. *Electrochimica Acta*, 111, 635-641.
- Chiti, F., & Dobson, C. M. (2006). Protein misfolding, functional amyloid, and human disease. *Annu. Rev. Biochem.*, 75, 333-366.
- Choi, J.-H., Jegal, J., & Kim, W.-N. (2006). Fabrication and characterization of multi-walled carbon nanotubes/polymer blend membranes. *Journal of Membrane Science*, 284(1), 406-415.
- Clark, W. R., & Gao, D. (2002). Low-molecular weight proteins in end-stage renal disease: potential toxicity and dialytic removal mechanisms. *Journal of the American Society of Nephrology*, 13(suppl 1), S41-S47.
- Coleman, K. S., Bailey, S. R., Fogden, S., & Green, M. L. (2003). Functionalization of single-walled carbon nanotubes via the Bingel reaction. *Journal of the American Chemical Society*, 125(29), 8722-8723.
- Colman, R. W., Hirsh, J., Marder, V. J., Colman, Hirsh, Marder, George. (2006). Hemostasis and thrombosis: basic principles and clinical practice.
- Cui, D., Tian, F., Ozkan, C. S., Wang, M., & Gao, H. (2005). Effect of single wall carbon nanotubes on human HEK293 cells. *Toxicology letters*, 155(1), 73-85.
- Currie, E., Van der Gucht, J., Borisov, O., & Cohen Stuart, M. (1999). Stuffed brushes: theory and experiment. *Pure and applied chemistry*, 71, 1227-1242.
- Dahe, G. J., Teotia, R. S., Kadam, S. S., & Bellare, J. R. (2011). The biocompatibility and separation performance of antioxidative polysulfone/vitamin E TPGS composite hollow fiber membranes. *Biomaterials*, 32(2), 352-365.
- Dalton, A., Stephan, C., Coleman, J., McCarthy, B., Ajayan, P., Lefrant, S., . . . Byrne, H. (2000). Selective interaction of a semiconjugated organic polymer

- with single-wall nanotubes. *The Journal of Physical Chemistry B*, 104(43), 10012-10016.
- Daugirdas, J. T., Blake, P. G., & Ing, T. S. (2007). *Handbook of dialysis* (Vol. 236): Lippincott Williams & Wilkins.
- Daugirdas, J. T., Blake, P. G., & Ing, T. S. (2012). *Handbook of dialysis*: Lippincott Williams & Wilkins.
- Davoren, M., Herzog, E., Casey, A., Cottineau, B., Chambers, G., Byrne, H. J., & Lyng, F. M. (2007). In vitro toxicity evaluation of single walled carbon nanotubes on human A549 lung cells. *Toxicology in Vitro*, 21(3), 438-448.
- De La Zerda, A., Zavaleta, C., Keren, S., Vaithilingam, S., Bodapati, S., Liu, Z., Oralkan, O. (2008). Carbon nanotubes as photoacoustic molecular imaging agents in living mice. *Nature nanotechnology*, 3(9), 557-562.
- De Paoli Lacerda, S. H., Semberova, J., Holada, K., Simakova, O., Hudson, S. D., & Simak, J. (2011). Carbon nanotubes activate store-operated calcium entry in human blood platelets. *ACS nano*, 5(7), 5808-5813.
- de Vos, W. M., Biesheuvel, P. M., de Keizer, A., Kleijn, J. M., & Cohen Stuart, M. A. (2008). Adsorption of the protein bovine serum albumin in a planar poly (acrylic acid) brush layer as measured by optical reflectometry. *Langmuir*, 24(13), 6575-6584.
- Dember, L. M. (2006). Amyloidosis-associated kidney disease. *Journal of the American Society of Nephrology*, 17(12), 3458-3471.
- Deng, B., Li, J., Hou, Z., Yao, S., Shi, L., Liang, G., & Sheng, K. (2008). Microfiltration membranes prepared from polyethersulfone powder grafted with acrylic acid by simultaneous irradiation and their pH dependence. *Radiation Physics and Chemistry*, 77(7), 898-906.
- Deng, B., Yang, X., Xie, L., Li, J., Hou, Z., Yao, S., Huang, Q. (2009). Microfiltration membranes with pH dependent property prepared from poly (methacrylic acid) grafted polyethersulfone powder. *Journal of Membrane Science*, 330(1), 363-368.
- Donaldson, K., Aitken, R., Tran, L., Stone, V., Duffin, R., Forrest, G., & Alexander, A. (2006). Carbon nanotubes: a review of their properties in relation to pulmonary toxicology and workplace safety. *Toxicological Sciences*, 92(1), 5-22.

- Drews, A. (2010). Membrane fouling in membrane bioreactors—characterisation, contradictions, cause and cures. *Journal of Membrane Science*, 363(1), 1-28.
- Dyck, A., Fritsch, D., & Nunes, S. (2002). Proton-conductive membranes of sulfonated polyphenylsulfone. *Journal of applied polymer science*, 86(11), 2820-2827.
- Dyke, C. A., & Tour, J. M. (2004a). Covalent functionalization of single-walled carbon nanotubes for materials applications. *The Journal of Physical Chemistry A*, 108(51), 11151-11159.
- Dyke, C. A., & Tour, J. M. (2004b). Overcoming the insolubility of carbon nanotubes through high degrees of sidewall functionalization. *Chemistry—A European Journal*, 10(4), 812-817.
- Edwards, H., Brown, D., Dale, J., & Plant, S. (2001). Raman spectroscopic studies of acid dissociation in sulfonated polystyrene resins. *Journal of Molecular Structure*, 595(1), 111-125.
- Ejaz, Shimada, M., Bhagwan, D., & Ahsan, A. (2016). Assessment of elevated creatinine. doi:monograph/935.
- Eknoyan, G., Beck, G. J., Cheung, A. K., Daugirdas, J. T., Greene, T., Kusek, J. W., Depner, T. A. (2002). Effect of dialysis dose and membrane flux in maintenance hemodialysis. *New England Journal of Medicine*, 347(25), 2010-2019.
- Epstein, F. H., Lefkovits, J., Plow, E. F., & Topol, E. J. (1995). Platelet glycoprotein IIb/IIIa receptors in cardiovascular medicine. *New England Journal of Medicine*, 332(23), 1553-1559.
- Fang, B., Cheng, C., Li, L., Cheng, J., Zhao, W., & Zhao, C. (2010). Surface modification of polyethersulfone membrane by grafting bovine serum albumin. *Fibers and Polymers*, 11(7), 960-966.
- Fang, B., Ling, Q., Zhao, W., Ma, Y., Bai, P., Wei, Q., Zhao, C. (2009). Modification of polyethersulfone membrane by grafting bovine serum albumin on the surface of polyethersulfone/poly (acrylonitrile-co-acrylic acid) blended membrane. *Journal of Membrane Science*, 329(1), 46-55.
- Fanizza, C., Casciardi, S., Incoronato, F., Cavallo, D., Ursini, C., Ciervo, A., Lega, D. (2015). Human epithelial cells exposed to functionalized multiwalled carbon nanotubes: interactions and cell surface modifications. *Journal of microscopy*, 259(3), 173-184.

- Feng, B., Weng, J., Yang, B., Qu, S., & Zhang, X. (2003). Characterization of surface oxide films on titanium and adhesion of osteoblast. *Biomaterials*, 24(25), 4663-4670.
- Fenoglio, I., Aldieri, E., Gazzano, E., Cesano, F., Colonna, M., Scarano, D., Lison, D. (2011). Thickness of multiwalled carbon nanotubes affects their lung toxicity. *Chemical research in toxicology*, 25(1), 74-82.
- Ferrari, M. (2005). Cancer nanotechnology: opportunities and challenges. *Nature Reviews Cancer*, 5(3), 161-171.
- Feynman, R. P. (1961). There's plenty of room at the bottom. *Miniaturization"*(*HD Gilbert, ed.*) Reinhold, New York.
- Findlay, M., & Isles, C. (2015). Structure and Function of the Kidney *Clinical Companion in Nephrology* (pp. 3-9): Springer.
- Finelli, L., Miller, J. T., Tokars, J. I., Alter, M. J., & Arduino, M. J. (2005). *National surveillance of dialysis-associated diseases in the United States, 2002*. Paper presented at the Seminars in dialysis.
- Fowkes, F. M. (1990). Quantitative characterization of the acid-base properties of solvents, polymers, and inorganic surfaces. *Journal of Adhesion Science and Technology*, 4(1), 669-691.
- Fry, A. K., Schilke, K. F., McGuire, J., & Bird, K. E. (2010). Synthesis and anticoagulant activity of heparin immobilized "end-on" to polystyrene microspheres coated with end-group activated polyethylene oxide. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 94(1), 187-195.
- Fujimori, A., Naito, H., & Miyazaki, T. (1998). Adsorption of complement, cytokines, and proteins by different dialysis membrane materials: evaluation by confocal laser scanning fluorescence microscopy. *Artificial organs*, 22(12), 1014-1017.
- Galli, F., Rovidati, S., Chiarantini, L., Campus, G., Canestrari, F., & Buoncristiani, U. (1998). Bioreactivity and biocompatibility of a vitamin E-modified multi-layer hemodialysis filter. *Kidney international*, 54(2), 580-589.
- Georgakilas, V., Kordatos, K., Prato, M., Guldi, D. M., Holzinger, M., & Hirsch, A. (2002). Organic functionalization of carbon nanotubes. *Journal of the American Chemical Society*, 124(5), 760-761.

- Gertz, M. A., Comenzo, R., Falk, R. H., Fermand, J. P., Hazenberg, B. P., Hawkins, P. N., Sanchorawala, V. (2005). Definition of organ involvement and treatment response in immunoglobulin light chain amyloidosis (AL): a consensus opinion from the 10th International Symposium on Amyloid and Amyloidosis. *American journal of hematology*, 79(4), 319-328.
- Gholami, M., Nasser, S., Feng, C., Matsuura, T., & Khulbe, K. (2003). The effect of heat-treatment on the ultrafiltration performance of polyethersulfone (PES) hollow-fiber membranes. *Desalination*, 155(3), 293-301.
- Giri, N., Natarajan, R., Gunasekaran, S., & Shreemathi, S. (2011). ¹³C NMR and FTIR spectroscopic study of blend behavior of PVP and nano silver particles'. *Archives of Applied Science Research*, 3(5), 624-630.
- Godo, M. N., & Sefton, M. V. (1999). Characterization of transient platelet contacts on a polyvinyl alcohol hydrogel by video microscopy. *Biomaterials*, 20(12), 1117-1126.
- Goh, B., Ong, L., & Lim, Y. (2016). Malaysian Society Nephrology. *22th Report of Malaysian Dialysis & Transplant Registry 2014*. Retrieved from <http://www.msn.org.my/fwbPagePublic.jsp?fwbPageId=pMdtr2014>
- Gómez, F. J., Chen, R. J., Wang, D., Waymouth, R. M., & Dai, H. (2003). Ring opening metathesis polymerization on non-covalently functionalized single-walled carbon nanotubes. *Chem. Commun.*(2), 190-191.
- Gong, K., & Ci, L. (2015). Process for purification of carbon nanotubes, *U.S. Patent Application No. 14/608,123*.
- Gorbet, M., & Sefton, M. (2003). Material-induced tissue factor expression but not CD11b upregulation depends on the presence of platelets. *Journal of Biomedical Materials Research Part A*, 67(3), 792-800.
- Gorbet, M. B., & Sefton, M. V. (2004). Biomaterial-associated thrombosis: roles of coagulation factors, complement, platelets and leukocytes. *Biomaterials*, 25(26), 5681-5703.
- Guan, R., Zou, H., Lu, D., Gong, C., & Liu, Y. (2005). Polyethersulfone sulfonated by chlorosulfonic acid and its membrane characteristics. *European Polymer Journal*, 41(7), 1554-1560.
- Guignard, J.-P., & Drukker, A. (1999). Why do newborn infants have a high plasma creatinine? *Pediatrics*, 103(4), e49-e49.

- Guiver, M., Croteau, S., Hazlett, J., & Kutowy, O. (1990). Synthesis and characterization of carboxylated polysulfones. *British Polymer Journal*, 23(1-2), 29-39.
- Guldi, D. M., Taieb, H., Rahman, G. A., Tagmatarchis, N., & Prato, M. (2005). Novel Photoactive Single-Walled Carbon Nanotube–Porphyrin Polymer Wraps: Efficient and Long-Lived Intracomplex Charge Separation. *Advanced Materials*, 17(7), 871-875.
- Gwinner, W., & Gröne, H. J. (2000). Role of reactive oxygen species in glomerulonephritis. *Nephrology Dialysis Transplantation*, 15(8), 1127-1132.
- Hakim, R. M., Held, P. J., Stannard, D. C., Wolfe, R. A., Port, F. K., Daugirdas, J. T., & Agodoa, L. (1996). Effect of the dialysis membrane on mortality of chronic hemodialysis patients. *Kidney international*, 50(2), 566-570.
- Hamciuc, C., Bruma, M., & Klapper, M. (2001). Sulfonated poly (ether-ketone) s containing hexafluoroisopropylidene groups. *Journal of Macromolecular Science, Part A*, 38(7), 659-671.
- Hamilton, R. F., Wu, Z., Mitra, S., Shaw, P. K., & Holian, A. (2013). Effect of MWCNT size, carboxylation, and purification on in vitro and in vivo toxicity, inflammation and lung pathology. *Particle and fibre toxicology*, 10(1), 1.
- Han, Y.-J., Wang, K.-H., Lai, J.-Y., & Liu, Y.-L. (2014). Hydrophilic chitosan-modified polybenzimidazole membranes for pervaporation dehydration of isopropanol aqueous solutions. *Journal of Membrane Science*, 463, 17-23.
- Hanson, S. R. (1993). Device thrombosis and thromboembolism. *Cardiovascular Pathology*, 2(3), 157-165.
- He, H., Pham-Huy, L. A., Dramou, P., Xiao, D., Zuo, P., & Pham-Huy, C. (2013). Carbon nanotubes: applications in pharmacy and medicine. *BioMed research international*, 2013.
- Higuchi, A., Shirano, K., Harashima, M., Yoon, B. O., Hara, M., Hattori, M., & Imamura, K. (2002). Chemically modified polysulfone hollow fibers with vinylpyrrolidone having improved blood compatibility. *Biomaterials*, 23(13), 2659-2666.
- Himmelfarb, J. (2005). Relevance of oxidative pathways in the pathophysiology of chronic kidney disease. *Cardiology clinics*, 23(3), 319-330.

- Hirlekar, R., Yamagar, M., Garse, H., Vij, M., & Kadam, V. (2009). Carbon nanotubes and its applications: a review. *Asian Journal of Pharmaceutical and Clinical Research*, 2(4), 17-27.
- Hoffman, M. (2003). Remodeling the blood coagulation cascade. *Journal of thrombosis and thrombolysis*, 16(1-2), 17-20.
- Hoffman, M., Meng, Z. H., Roberts, H. R., & Monroe, D. M. (2005). Rethinking the coagulation cascade. *日本血栓止血学会誌*, 16(1), 70-81.
- Holzinger, M., Vostrowsky, O., Hirsch, A., Hennrich, F., Kappes, M., Weiss, R., & Jellen, F. (2001). Sidewall functionalization of carbon nanotubes. *Angewandte Chemie International Edition*, 40(21), 4002-4005.
- House, A. A., Wells, G. A., Donnelly, J. G., Nadler, S. P., & Hébert, P. C. (2000). Randomized trial of high-flux vs low-flux haemodialysis: effects on homocysteine and lipids. *Nephrology Dialysis Transplantation*, 15(7), 1029-1034.
- Hu, H., Ni, Y., Montana, V., Haddon, R. C., & Parpura, V. (2004). Chemically functionalized carbon nanotubes as substrates for neuronal growth. *Nano letters*, 4(3), 507-511.
- Huang, J., Xue, J., Xiang, K., Zhang, X., Cheng, C., Sun, S., & Zhao, C. (2011). Surface modification of polyethersulfone membranes by blending triblock copolymers of methoxyl poly (ethylene glycol)–polyurethane–methoxyl poly (ethylene glycol). *Colloids and Surfaces B: Biointerfaces*, 88(1), 315-324.
- Huang, R., Shao, P., Burns, C., & Feng, X. (2001). Sulfonation of poly (ether ether ketone)(PEEK): kinetic study and characterization. *Journal of applied polymer science*, 82(11), 2651-2660.
- Huang, Y., Wong, C., Zheng, J., Bouwman, H., Barra, R., Wahlström, B., Wong, M. (2012). Bisphenol A (BPA) in China: a review of sources, environmental levels, and potential human health impacts. *Environment international*, 42, 91-99.
- Humes, H., Fissell, W., & Tiranathanagul, K. (2006). The future of hemodialysis membranes. *Kidney international*, 69(7), 1115-1119.
- Idris, A., & Ahmed, I. (2008). Viscosity behavior of microwave-heated and conventionally heated poly (ether sulfone)/dimethylformamide/lithium

- bromide polymer solutions. *Journal of applied polymer science*, 108(1), 302-307.
- Idris, A., Ahmed, I., & Misran, M. (2009). Novel high performance hollow fiber ultrafiltration membranes spun from LiBr doped solutions. *Desalination*, 249(2), 541-548.
- Idris, A., Hew, K. Y., & Chan, M. K. (2009). Preparation of cellulose acetate dialysis membrane using d-glucose monohydrate as additive. *Jurnal Teknologi (Kejuruteraan)*(51F), 67-76.
- Idris, A., Lee, K. Y., Noordin, M., & Chan, M. K. (2008). Response surface methodology approach to study the influence of PEG and water in cellulose acetate dialysis membranes. *Jurnal Teknologi F*(49F), 39-49.
- Idris, A., Mat Zain, N., & Noordin, M. (2007). Synthesis, characterization and performance of asymmetric polyethersulfone (PES) ultrafiltration membranes with polyethylene glycol of different molecular weights as additives. *Desalination*, 207(1), 324-339.
- Idris, A., & Yet, L. K. (2006). The effect of different molecular weight PEG additives on cellulose acetate asymmetric dialysis membrane performance. *Journal of Membrane Science*, 280(1), 920-927.
- Iijima, S. (1991). Helical microtubules of graphitic carbon. *nature*, 354(6348), 56-58.
- Iojoiu, C., Maréchal, M., Chabert, F., & Sanchez, J. Y. (2005). Mastering sulfonation of aromatic polysulfones: crucial for membranes for fuel cell application. *Fuel Cells*, 5(3), 344-354.
- Irfan, M., & Idris, A. (2015). Overview of PES biocompatible/hemodialysis membranes: PES–blood interactions and modification techniques. *Materials Science and Engineering: C*, 56, 574-592.
- Irfan, M., Idris, A., Yusof, N. M., Khairuddin, N. F. M., & Akhmal, H. (2014). Surface modification and performance enhancement of nano-hybrid f-MWCNT/PVP90/PES hemodialysis membranes. *Journal of Membrane Science*, 467, 73-84.
- Irvine, D. J., Mayes, A. M., & Griffith, L. G. (2001). Nanoscale clustering of RGD peptides at surfaces using comb polymers. 1. Synthesis and characterization of comb thin films. *Biomacromolecules*, 2(1), 85-94.

- Ishihara, K., Hasegawa, T., Watanabe, J., & Iwasaki, Y. (2002). Protein Adsorption-Resistant Hollow Fibers for Blood Purification. *Artificial organs*, 26(12), 1014-1019.
- Ishihara, K., Nomura, H., Mihara, T., Kurita, K., Iwasaki, Y., & Nakabayashi, N. (1998). Why do phospholipid polymers reduce protein adsorption? *Journal of biomedical materials research*, 39(2), 323-330.
- Ismail, A. F., Mustaffar, M., Illias, R. M., & Abdullah, M. S. (2006). Effect of dope extrusion rate on morphology and performance of hollow fibers membrane for ultrafiltration. *Separation and purification technology*, 49(1), 10-19
- Israëls, R., Leermakers, F., & Fler, G. (1995). Adsorption of charged block copolymers: effect on colloidal stability. *Macromolecules*, 28(5), 1626-1634.
- Iwasaki, Y., Yamato, H., Nii-Kono, T., Fujieda, A., Uchida, M., Hosokawa, A., Fukagawa, M. (2006). Uremic toxin and bone metabolism. *Journal of bone and mineral metabolism*, 24(2), 172-175.
- Jain, K. (2012). Advances in use of functionalized carbon nanotubes for drug design and discovery. *Expert opinion on drug discovery*, 7(11), 1029-1037.
- Jia, Z., & Tian, C. (2009). Quantitative determination of polyethylene glycol with modified Dragendorff reagent method. *Desalination*, 247(1), 423-429.
- Johnell, M., Larsson, R., & Siegbahn, A. (2005). The influence of different heparin surface concentrations and antithrombin-binding capacity on inflammation and coagulation. *Biomaterials*, 26(14), 1731-1739.
- Johnson, B. D., Kip, K. E., Marroquin, O. C., Ridker, P. M., Kelsey, S. F., Shaw, L. J., .Sopko, G. (2004). Serum Amyloid A as a Predictor of Coronary Artery Disease and Cardiovascular Outcome in Women The National Heart, Lung, and Blood Institute-Sponsored Women's Ischemia Syndrome Evaluation (WISE). *Circulation*, 109(6), 726-732.
- Kaeselev, B., Pieracci, J., & Belfort, G. (2001). Photoinduced grafting of ultrafiltration membranes: comparison of poly (ether sulfone) and poly (sulfone). *Journal of Membrane Science*, 194(2), 245-261.
- Kaiser, V., & Stropnik, C. (2000). Membranes from polysulfone/N, N-Dimethylacetamide/water system; structure and water Flux. *Acta Chimica Slovenica*, 47(2), 205-214.

- Kamal, A. H., Tefferi, A., & Pruthi, R. K. (2007). *How to interpret and pursue an abnormal prothrombin time, activated partial thromboplastin time, and bleeding time in adults*. Paper presented at the Mayo Clinic Proceedings.
- Kang, Y., & Taton, T. A. (2003). Micelle-encapsulated carbon nanotubes: a route to nanotube composites. *Journal of the American Chemical Society*, *125*(19), 5650-5651.
- Kee, C. M., & Idris, A. (2010). Permeability performance of different molecular weight cellulose acetate hemodialysis membrane. *Separation and Purification Technology*, *75*(2), 102-113.
- Keen, M., Lancaster, L., & Binkley, L. (1995). Section IX-Hemodialysis. *ANNA core curriculum for nephrology nursing*, 207-258.
- Kelly, S. T., & Zydney, A. L. (1997). Protein fouling during microfiltration: comparative behavior of different model proteins. *Biotechnology and bioengineering*, *55*(1), 91-100.
- Kenausis, G. L., Vörös, J., Elbert, D. L., Huang, N., Hofer, R., Ruiz-Taylor, L., . . . Spencer, N. D. (2000). Poly (L-lysine)-g-poly (ethylene glycol) layers on metal oxide surfaces: attachment mechanism and effects of polymer architecture on resistance to protein adsorption. *The Journal of Physical Chemistry B*, *104*(14), 3298-3309.
- Kerr, P. G., & Huang, L. (2010). Review: membranes for haemodialysis. *Nephrology*, *15*(4), 381-385.
- Kesting, R. E. (1985). *Synthetic polymeric membranes: a structural perspective*: Wiley NY etc.
- Khabashesku, V., Barrera, E., McIntosh, D., & Para-Pena, L. (2006). Carbon nanotube reinforced thermoplastic polymer composites achieved through benzoyl peroxide initiated interfacial bonding to polymer matrices, *U.S. Patent Application No. 11/411,730*.
- Khabashesku, V. N., Peng, H., Billups, W. E., Ying, Y., & Margrave, J. L. (2006). Method for functionalizing carbon nanotubes utilizing peroxides, *U.S. Patent No. 7,125,533*.
- Kim, I., Choi, J., & Tak, T. (1999). Sulfonated polyethersulfone by heterogeneous method and its membrane performances. *Journal of applied polymer science*, *74*(8), 2046-2055.

- Kim, J., & Kim, C. (2005). Ultrafiltration membranes prepared from blends of polyethersulfone and poly (1-vinylpyrrolidone-co-styrene) copolymers. *Journal of Membrane Science*, 262(1), 60-68.
- Kim, K., Lee, K., Cho, K., & Park, C. (2002). Surface modification of polysulfone ultrafiltration membrane by oxygen plasma treatment. *Journal of Membrane Science*, 199(1), 135-145.
- Klinkmann, H., Falkenhagen, D., & Courtney, J. (1987). Clinical relevance of biocompatibility—the material cannot be divorced from the device *Uremia Therapy* (pp. 125-140): Springer.
- Kong, N., Shimpi, M. R., Park, J. H., Ramström, O., & Yan, M. (2015). Carbohydrate conjugation through microwave-assisted functionalization of single-walled carbon nanotubes using perfluorophenyl azides. *Carbohydrate research*, 405, 33-38.
- Koopman, B., Kim, S., Partch, R., & El-Shall, H. (2008). Synthesis and Engineering of Polymeric Latex Particles for Hemodialysis Part I?A Review *Particulate Systems in Nano- and Biotechnologies* (pp. 53-83): CRC Press.
- Kopple, J. D. (2001). National kidney foundation K/DOQI clinical practice guidelines for nutrition in chronic renal failure. *American Journal of Kidney Diseases*, 37(1), S66-S70.
- Krauss, T. D. (2009). Biosensors: Nanotubes light up cells. *Nature nanotechnology*, 4(2), 85-86.
- Kruger, N. J. (2009). The Bradford method for protein quantitation *The protein protocols handbook* (pp. 17-24): Springer.
- Kuijpers, T., Hakkert, B., Van Mourik, J., & Roos, D. (1990). Distinct adhesive properties of granulocytes and monocytes to endothelial cells under static and stirred conditions. *The Journal of Immunology*, 145(8), 2588-2594.
- Kumar, M., & Ulbricht, M. (2014). Novel ultrafiltration membranes with adjustable charge density based on sulfonated poly (arylene ether sulfone) block copolymers and their tunable protein separation performance. *Polymer*, 55(1), 354-365.
- Kumar, R., Dhanawat, M., Kumar, S., N Singh, B., K Pandit, J., & R Sinha, V. (2014). Carbon nanotubes: a potential concept for drug delivery applications. *Recent patents on drug delivery & formulation*, 8(1), 12-26.

- Lee, K. M., Li, L., & Dai, L. (2005). Asymmetric end-functionalization of multi-walled carbon nanotubes. *Journal of the American Chemical Society*, 127(12), 4122-4123.
- Lemke, H. (1993). Methods for the detection of endotoxins present during extracorporeal circulation. *Nephrology, dialysis, transplantation: official publication of the European Dialysis and Transplant Association-European Renal Association*, 9, 90-95.
- Leng, Y., Chen, J., Yang, P., Sun, H., Wan, G., & Huang, N. (2003). Mechanical properties and platelet adhesion behavior of diamond-like carbon films synthesized by pulsed vacuum arc plasma deposition. *Surface science*, 531(2), 177-184.
- Lesaffer, G., De Smet, R., Lameire, N., Dhondt, A., Duym, P., & Vanholder, R. (2000). Intradialytic removal of protein-bound uraemic toxins: role of solute characteristics and of dialyser membrane. *Nephrology Dialysis Transplantation*, 15(1), 50-57.
- Leung, L. L. (2006). Perioperative evaluation of bleeding diathesis. *ASH Education Program Book*, 2006(1), 457-461.
- Leyboldt, J. (2000). Solute fluxes in different treatment modalities. *Nephrology Dialysis Transplantation*, 15(suppl 1), 3-9.
- Leyboldt, J. K., & Cheung, A. K. (1996). Characterization of molecular transport in artificial kidneys. *Artificial organs*, 20(6), 381-389.
- Leyboldt, J. K., & Cheung, A. K. (2006). *Unresolved issues in dialysis: Revisiting the Hemodialysis Dose*. Paper presented at the Seminars in dialysis.
- Leyboldt, J. K., Cheung, A. K., Carroll, C. E., Stannard, D. C., Pereira, B. J., Agodoa, L. Y., & Port, F. K. (1999). Effect of dialysis membranes and middle molecule removal on chronic hemodialysis patient survival. *American Journal of Kidney Diseases*, 33(2), 349-355.
- Li, D., Chen, H., Glenn McClung, W., & Brash, J. L. (2009). Lysine-PEG-modified polyurethane as a fibrinolytic surface: Effect of PEG chain length on protein interactions, platelet interactions and clot lysis. *Acta Biomaterialia*, 5(6), 1864-1871.
- Li, L., Cheng, C., Xiang, T., Tang, M., Zhao, W., Sun, S., & Zhao, C. (2012). Modification of polyethersulfone hemodialysis membrane by blending citric

- acid grafted polyurethane and its anticoagulant activity. *Journal of Membrane Science*, 405, 261-274.
- Li, L., Li, G., Jiang, J., Liu, X., Luo, L., & Nan, K. (2012). Electrospun fibrous scaffold of hydroxyapatite/poly (ϵ -caprolactone) for bone regeneration. *Journal of Materials Science: Materials in Medicine*, 23(2), 547-554.
- Li, L., Yan, G., & Wu, J. (2009). Modification of polysulfone membranes via surface-initiated atom transfer radical polymerization and their antifouling properties. *Journal of applied polymer science*, 111(4), 1942-1946.
- Li, L., Yin, Z., Li, F., Xiang, T., Chen, Y., & Zhao, C. (2010). Preparation and characterization of poly (acrylonitrile-acrylic acid-N-vinyl pyrrolidinone) terpolymer blended polyethersulfone membranes. *Journal of Membrane Science*, 349(1), 56-64.
- Li, Y., Guan, H.-M., Chung, T.-S., & Kulprathipanja, S. (2006). Effects of novel silane modification of zeolite surface on polymer chain rigidification and partial pore blockage in polyethersulfone (PES)–zeolite A mixed matrix membranes. *Journal of Membrane Science*, 275(1), 17-28.
- Lim, W. (2009). Antiphospholipid antibody syndrome. *ASH Education Program Book*, 2009(1), 233-239.
- Liu, A., Watanabe, T., Honma, I., Wang, J., & Zhou, H. (2006). Effect of solution pH and ionic strength on the stability of poly (acrylic acid)-encapsulated multiwalled carbon nanotubes aqueous dispersion and its application for NADH sensor. *Biosensors and Bioelectronics*, 22(5), 694-699.
- Liu, B., Li, X., Li, B., Xu, B., & Zhao, Y. (2009). Carbon nanotube based artificial water channel protein: membrane perturbation and water transportation. *Nano letters*, 9(4), 1386-1394.
- Liu, K., Tian, Y., & Jiang, L. (2013). Bio-inspired superoleophobic and smart materials: design, fabrication, and application. *Progress in Materials Science*, 58(4), 503-564.
- Liu, L., Wang, T., Li, J., Guo, Z.-X., Dai, L., Zhang, D., & Zhu, D. (2003). Self-assembly of gold nanoparticles to carbon nanotubes using a thiol-terminated pyrene as interlinker. *Chemical physics letters*, 367(5), 747-752.
- Liu, Z.-B., Deng, X.-P., & Zhao, C.-S. (2006). BSA hybrid synthesized polymer. *Chinese Chemical Letters*, 17(11), 1519-1522.

- Liu, Z., Deng, X., Wang, M., Chen, J., Zhang, A., Gu, Z., & Zhao, C. (2009). BSA-modified polyethersulfone membrane: preparation, characterization and biocompatibility. *Journal of Biomaterials Science, Polymer Edition*, 20(3), 377-397.
- Liu, Z., Sun, X., Nakayama-Ratchford, N., & Dai, H. (2007). Supramolecular chemistry on water-soluble carbon nanotubes for drug loading and delivery. *ACS nano*, 1(1), 50-56.
- Lu, D., Zou, H., Guan, R., Dai, H., & Lu, L. (2005). Sulfonation of polyethersulfone by chlorosulfonic acid. *Polymer Bulletin*, 54(1-2), 21-28.
- Lu, Q., Moore, J. M., Huang, G., Mount, A. S., Rao, A. M., Larcom, L. L., & Ke, P. C. (2004). RNA polymer translocation with single-walled carbon nanotubes. *Nano letters*, 4(12), 2473-2477.
- Lu, W., & Chou, T.-W. (2011). Analysis of the entanglements in carbon nanotube fibers using a self-folded nanotube model. *Journal of the Mechanics and Physics of Solids*, 59(3), 511-524.
- Lufrano, F., Baglio, V., Staiti, P., Arico, A. S., & Antonucci, V. (2006). Development and characterization of sulfonated polysulfone membranes for direct methanol fuel cells. *Desalination*, 199(1), 283-285.
- Ma, P.-C., Siddiqui, N. A., Marom, G., & Kim, J.-K. (2010). Dispersion and functionalization of carbon nanotubes for polymer-based nanocomposites: a review. *Composites Part A: Applied Science and Manufacturing*, 41(10), 1345-1367.
- Ma, X., Su, Y., Sun, Q., Wang, Y., & Jiang, Z. (2007). Preparation of protein-adsorption-resistant polyethersulfone ultrafiltration membranes through surface segregation of amphiphilic comb copolymer. *Journal of Membrane Science*, 292(1), 116-124.
- Macey, R. L., & Farmer, R. E. (1970). Inhibition of water and solute permeability in human red cells. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 211(1), 104-106.
- Maher, J. F. (1989). *Replacement of renal function by dialysis: a text book of dialysis*: Springer.
- Mann, K. G. (2003). Thrombin formation. *CHEST Journal*, 124(3_suppl), 4S-10S.
- Mansourpanah, Y., Madaeni, S., Rahimpour, A., & Farhadian, A. (2009). The effect of non-contact heating (microwave irradiation) and contact heating (annealing

- process) on properties and performance of polyethersulfone nanofiltration membranes. *Applied Surface Science*, 255(20), 8395-8402.
- Mansourpanah, Y., Madaeni, S. S., Adeli, M., Rahimpour, A., & Farhadian, A. (2009). Surface modification and preparation of nanofiltration membrane from polyethersulfone/polyimide blend—Use of a new material (polyethyleneglycol-triazine). *Journal of applied polymer science*, 112(5), 2888-2895.
- Marshall, A., Munro, P., & Trägårdh, G. (1993). The effect of protein fouling in microfiltration and ultrafiltration on permeate flux, protein retention and selectivity: a literature review. *Desalination*, 91(1), 65-108.
- Matsuda, K. S. M. (2011). Solute Removal Efficiency and Biocompatibility of the High-Performance Membrane—From Engineering Points of View. *High-Performance Membrane Dialyzers*, 173, 11-22.
- Matsuda, M., Sato, M., Sakata, H., Ogawa, T., Yamamoto, K., Yakushiji, T., Sakai, K. (2008). Effects of fluid flow on elution of hydrophilic modifier from dialysis membrane surfaces. *J Artif Organs*, 11(3), 148-155.
- Matzke, G. R., Aronoff, G. R., Atkinson, A. J., Bennett, W. M., Decker, B. S., Eckardt, K.-U., Keller, F. (2011). Drug dosing consideration in patients with acute and chronic kidney disease—a clinical update from Kidney Disease: Improving Global Outcomes (KDIGO). *Kidney international*, 80(11), 1122-1137.
- Mavroidis, C. (2006). *Artificial Kidney Tissue Engineering and Artificial Organs* (pp. 67-61-67-24): CRC Press.
- Mehra, N. K., & Jain, N. K. (2016). Multifunctional hybrid-carbon nanotubes: new horizon in drug delivery and targeting. *Journal of drug targeting*, 24(4), 294-308.
- Membrana, G. (2004). Membrane letter news for nephrologist-The world of dialysis membrane (1/04).
- Mickelson, E., Huffman, C., Rinzler, A., Smalley, R., Hauge, R., & Margrave, J. (1998). Fluorination of single-wall carbon nanotubes. *Chemical physics letters*, 296(1), 188-194.
- Mielcarek, J., & Skupin, P. (2010). [Functionalization of carbon nanotubes for multimodal drug delivery]. *Przegląd lekarski*, 68(3), 167-170.

- Miller, J. E., Kovesdy, C. P., Nissenson, A. R., Mehrotra, R., Streja, E., Van Wyck, D., . . . Kalantar-Zadeh, K. (2010). Association of hemodialysis treatment time and dose with mortality and the role of race and sex. *American Journal of Kidney Diseases*, 55(1), 100-112.
- Misoph, M., Schwender, S., & Babin-Ebell, J. (1998). Response of the cellular immune system to cardiopulmonary bypass is independent of the applied pump type and of the use of heparin-coated surfaces. *The Thoracic and cardiovascular surgeon*, 46(04), 222-227.
- Mitchell, C. A., Bahr, J. L., Arepalli, S., Tour, J. M., & Krishnamoorti, R. (2002). Dispersion of functionalized carbon nanotubes in polystyrene. *Macromolecules*, 35(23), 8825-8830.
- Mittal, V. (2011). Carbon nanotubes surface modifications: an overview. *Surface Modification of Nanotube Fillers*, 1-23.
- Möckel, D., Staude, E., & Guiver, M. D. (1999). Static protein adsorption, ultrafiltration behavior and cleanability of hydrophilized polysulfone membranes. *Journal of Membrane Science*, 158(1), 63-75.
- Mody, N., Tekade, R. K., Mehra, N. K., Chopdey, P., & Jain, N. K. (2014). Dendrimer, liposomes, carbon nanotubes and PLGA nanoparticles: one platform assessment of drug delivery potential. *Aaps Pharmscitech*, 15(2), 388-399.
- Moghaddam, M. J., Taylor, S., Gao, M., Huang, S., Dai, L., & McCall, M. J. (2004). Highly efficient binding of DNA on the sidewalls and tips of carbon nanotubes using photochemistry. *Nano letters*, 4(1), 89-93.
- Moore Jr, J. E., & Maitland, D. J. (2013). *Biomedical technology and devices*: CRC press.
- Morti, S., Shao, J., & Zydney, A. L. (2003). Importance of asymmetric structure in determining mass transport characteristics of hollow fiber hemodialyzers. *Journal of Membrane Science*, 224(1), 39-49.
- Morena, M., Delbosc, S., Dupuy, A. M., Canaud, B., & Cristol, J. P. (2005). Overproduction of reactive oxygen species in end-stage renal disease patients: A potential component of hemodialysis-associated inflammation. *Hemodialysis international*, 9(1), 37-46.

- Mosqueda-Jimenez, D. B., Narbaitz, R. M., & Matsuura, T. (2006). Effects of preparation conditions on the surface modification and performance of polyethersulfone ultrafiltration membranes. *Journal of applied polymer science*, 99(6), 2978-2988.
- Murthy, R., Shell, C. E., & Grunlan, M. A. (2009). The influence of poly (ethylene oxide) grafting via siloxane tethers on protein adsorption. *Biomaterials*, 30(13), 2433-2439.
- Nabe, A., Staude, E., & Belfort, G. (1997). Surface modification of polysulfone ultrafiltration membranes and fouling by BSA solutions. *Journal of Membrane Science*, 133(1), 57-72.
- Nakatan, T., & Takemoto, Y. (2003). The Effect of Vitamin E-Bonded Dialyzer Membrane on Red Blood Cell Survival in Hemodialyzed Patients. *Artificial organs*, 27(3), 214-217.
- National Institute of Diabetes and Digestive and Kidney Diseases, N. I. o. H. (March 23, 2012). hemodialysis dose and adequacy Retrieved from http://kidney.niddk.nih.gov/kudiseases/pubs/hemodialysisdose/hemodialysis_dose_508.pdf
- Ngadiman, N. H. A., Idris, A., Irfan, M., Kurniawan, D., Yusof, N. M., & Nasiri, R. (2015). γ -Fe₂O₃ nanoparticles filled polyvinyl alcohol as potential biomaterial for tissue engineering scaffold. *Journal of the mechanical behavior of biomedical materials*, 49, 90-104.
- Nie, S., Tang, M., Yin, Z., Wang, L., Sun, S., & Zhao, C. (2014). Biologically inspired membrane design with a heparin-like interface: prolonged blood coagulation, inhibited complement activation, and bio-artificial liver related cell proliferation. *Biomaterials Science*, 2(1), 98-109.
- Nie, S., Xue, J., Lu, Y., Liu, Y., Wang, D., Sun, S., Zhao, C. (2012). Improved blood compatibility of polyethersulfone membrane with a hydrophilic and anionic surface. *Colloids and Surfaces B: Biointerfaces*, 100, 116-125.
- Niyogi, S., Hamon, M., Hu, H., Zhao, B., Bhowmik, P., Sen, R., Haddon, R. (2002). Chemistry of single-walled carbon nanotubes. *Accounts of Chemical Research*, 35(12), 1105-1113.
- Nugent, L., & Jain, R. (1984). Plasma pharmacokinetics and interstitial diffusion of macromolecules in a capillary bed. *American Journal of Physiology-Heart and Circulatory Physiology*, 246(1), H129-H137.

- Okpalugo, T., Ogwu, A., Maguire, P., & McLaughlin, J. (2004). Platelet adhesion on silicon modified hydrogenated amorphous carbon films. *Biomaterials*, 25(2), 239-245.
- Pantarotto, D., Singh, R., McCarthy, D., Erhardt, M., Briand, J. P., Prato, M., Bianco, A. (2004). Functionalized carbon nanotubes for plasmid DNA gene delivery. *Angewandte Chemie*, 116(39), 5354-5358.
- Park, J. Y., Acar, M. H., Akthakul, A., Kuhlman, W., & Mayes, A. M. (2006). Polysulfone-*graft*-poly (ethylene glycol) graft copolymers for surface modification of polysulfone membranes. *Biomaterials*, 27(6), 856-865.
- Pekker, S., Salvetat, J.-P., Jakab, E., Bonard, J.-M., & Forro, L. (2001). Hydrogenation of carbon nanotubes and graphite in liquid ammonia. *The Journal of Physical Chemistry B*, 105(33), 7938-7943.
- Peng, N., Chung, T.-S., & Wang, K. Y. (2008). Macrovoid evolution and critical factors to form macrovoid-free hollow fiber membranes. *Journal of Membrane Science*, 318(1), 363-372.
- Pinnau, I., & Freeman, B. (2000). *Formation and modification of polymeric membranes: overview*. Paper presented at the ACS Symposium series.
- Qian, D., Dickey, E. C., Andrews, R., & Rantell, T. (2000). Load transfer and deformation mechanisms in carbon nanotube-polystyrene composites. *Applied physics letters*, 76(20), 2868-2870.
- Ran, F., Nie, S., Lu, Y., Cheng, C., Wang, D., Sun, S., & Zhao, C. (2012). Comparison of surface segregation and anticoagulant property in block copolymer blended evaporation and phase inversion membranes. *Surface and Interface Analysis*, 44(7), 819-824.
- Ran, F., Nie, S., Zhao, W., Li, J., Su, B., Sun, S., & Zhao, C. (2011). Biocompatibility of modified polyethersulfone membranes by blending an amphiphilic triblock co-polymer of poly (vinyl pyrrolidone)-*b*-poly (methyl methacrylate)-*b*-poly (vinyl pyrrolidone). *Acta Biomaterialia*, 7(9), 3370-3381.
- Rana, D., & Matsuura, T. (2010). Surface modifications for antifouling membranes. *Chemical reviews*, 110(4), 2448-2471.

- Raravikar, N. R., Schadler, L. S., Vijayaraghavan, A., Zhao, Y., Wei, B., & Ajayan, P. M. (2005). Synthesis and characterization of thickness-aligned carbon nanotube-polymer composite films. *Chemistry of materials*, 17(5), 974-983.
- Rebuck, N., & Finn, A. (1994). Polymorphonuclear granulocyte expression of CD11a/CD18, CD11b/CD18 and L-selectin in normal individuals. *FEMS immunology and medical microbiology*, 8(3), 189-195.
- Remes, A., & Williams, D. (1992). Immune response in biocompatibility. *Biomaterials*, 13(11), 731-743.
- Rixman, M., & Ortiz, C. (2002). *Exploring the molecular origins of bio (in) compatibility: Adhesion between proteins and poly (ethylene oxide)*. Paper presented at the abstract of the papers of the american chemical society.
- Rizvi, Z. A., Puri, N., & Saxena, R. K. (2015). Lipid antigen presentation through CD1d pathway in mouse lung epithelial cells, macrophages and dendritic cells and its suppression by poly-dispersed single-walled carbon nanotubes. *Toxicology in Vitro*, 29(6), 1275-1282.
- Rodríguez-Yáñez, Y., Bahena-Urbe, D., Chávez-Munguía, B., López-Marure, R., González-Monroy, S., Cisneros, B., & Albores, A. (2015). Commercial single-walled carbon nanotubes effects in fibrinolysis of human umbilical vein endothelial cells. *Toxicology in Vitro*, 29(5), 1201-1214.
- Rogers Nieman, G. M., & Dinu, C. Z. (2014). Therapeutic applications of carbon nanotubes: opportunities and challenges. *Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology*, 6(4), 327-337.
- Rovira-Bru, M., Giralt, F., & Cohen, Y. (2001). Protein adsorption onto zirconia modified with terminally grafted polyvinylpyrrolidone. *Journal of colloid and interface science*, 235(1), 70-79.
- Sajid, M. I., Jamshaid, U., Jamshaid, T., Zafar, N., Fessi, H., & Elaissari, A. (2016). Carbon nanotubes from synthesis to in vivo biomedical applications. *International journal of pharmaceutics*, 501(1), 278-299.
- Sakai, K. (1994). Determination of pore size and pore size distribution: 2. Dialysis membranes. *Journal of Membrane Science*, 96(1-2), 91-130.
- Samtleben, W., Dengler, C., Reinhardt, B., Nothdurft, A., & Lemke, H.-D. (2003). Comparison of the new polyethersulfone high-flux membrane DIAPES® HF800 with conventional high-flux membranes during on-line haemodiafiltration. *Nephrology Dialysis Transplantation*, 18(11), 2382-2386.

- Saran, R., Li, Y., Robinson, B., Ayanian, J., Balkrishnan, R., Bragg-Gresham, J., He, K. (2015). US Renal Data System 2014 Annual Data Report. *American Journal of Kidney Diseases*, 66(1), Svii.
- Sasaki, M. (2006). Development of vitamin E-modified polysulfone membrane dialyzers. *Journal of Artificial Organs*, 9(1), 50-60.
- Saxena, N., Prabhavathy, C., De, S., & DasGupta, S. (2009). Flux enhancement by argon-oxygen plasma treatment of polyethersulfone membranes. *Separation and Purification Technology*, 70(2), 160-165.
- Schmaier, A. (1997). Contact activation: a revision. *Thrombosis and haemostasis*, 78(1), 101-107.
- Schulze, A., Marquardt, B., Kaczmarek, S., Schubert, R., Prager, A., & Buchmeiser, M. R. (2010). Electron Beam-Based Functionalization of Poly (ethersulfone) Membranes. *Macromolecular rapid communications*, 31(5), 467-472.
- Sedighi, O., Abediankenari, S., & Omranifar, B. (2015). Association between plasma Beta-2 microglobulin level and cardiac performance in patients with chronic kidney disease. *Nephro-urology monthly*, 7(1). DOI: 10.5812/numonthly.23563
- Sepahvand, R., Adeli, M., Astinchap, B., & Kabiri, R. (2008). New nanocomposites containing metal nanoparticles, carbon nanotube and polymer. *Journal of Nanoparticle Research*, 10(8), 1309-1318.
- Shah, T., Goodwin, J., & Ritchie, S. (2005). Development and characterization of a microfiltration membrane catalyst containing sulfonated polystyrene grafts. *Journal of Membrane Science*, 251(1), 81-89.
- Shen, Y. B., Zhang, Y. T., Qiu, J. H., Zhang, Y. W., & Zhang, H. Q. (2011). Hydrophilic modification of PES hollow fiber membrane via surface-initiated atom transfer radical polymerization. *Advanced Materials Research*, 150, 565-570.
- Sheppard, J., McClung, W., & Feuerstein, I. (1994). Adherent platelet morphology on adsorbed fibrinogen: effects of protein incubation time and albumin addition. *Journal of biomedical materials research*, 28(10), 1175-1186.
- Shi Kam, N. W., Jessop, T. C., Wender, P. A., & Dai, H. (2004). Nanotube molecular transporters: internalization of carbon nanotube-protein conjugates into mammalian cells. *Journal of the American Chemical Society*, 126(22), 6850-6851.

- Shi, Q., Su, Y., Ning, X., Chen, W., Peng, J., & Jiang, Z. (2010). Graft polymerization of methacrylic acid onto polyethersulfone for potential pH-responsive membrane materials. *Journal of Membrane Science*, 347(1), 62-68.
- Singh, B., Baburao, C., Pispati, V., Pathipati, H., Muthy, N., Prassana, S., & Rathode, B. G. (2012). Carbon nanotubes. A novel drug delivery system. *International Journal of Research in Pharmacy and Chemistry*, 2(2), 523-532.
- Sinha, A., & Bagga, A. (2015). Maintenance dialysis in developing countries. *Pediatric Nephrology*, 30(2), 211-219.
- Smith, J. A. (1994). Neutrophils, host defense, and inflammation: a double-edged sword. *Journal of Leukocyte Biology*, 56(6), 672-686.
- Sokolov, A., Hellerud, B., Tønnessen, T., Johannessen, E., & Mollnes, T. (2013). Activation of coagulation and platelets by candidate membranes of implantable devices in a whole blood model without soluble anticoagulant. *Journal of Biomedical Materials Research Part A*, 101(2), 575-581.
- Sperling, C., Houska, M., Brynda, E., Streller, U., & Werner, C. (2006). In vitro hemocompatibility of albumin–heparin multilayer coatings on polyethersulfone prepared by the layer-by-layer technique. *Journal of Biomedical Materials Research Part A*, 76(4), 681-689.
- Star, A., & Stoddart, J. F. (2002). Dispersion and solubilization of single-walled carbon nanotubes with a hyperbranched polymer. *Macromolecules*, 35(19), 7516-7520.
- Star, A., Stoddart, J. F., Steuerman, D., Diehl, M., Boukai, A., Wong, E. W., Heath, J. R. (2001). Preparation and properties of polymer-wrapped single-walled carbon nanotubes. *Angewandte Chemie International Edition*, 40(9), 1721-1725.
- Stevens, P. E., & Levin, A. (2013). Evaluation and management of chronic kidney disease: synopsis of the kidney disease: improving global outcomes 2012 clinical practice guideline. *Annals of internal medicine*, 158(11), 825-830.
- Steffens, G., Nothdurft, L., Buse, G., Thissen, H., Höcker, H., & Klee, D. (2002). High density binding of proteins and peptides to poly (D, L-lactide) grafted with polyacrylic acid. *Biomaterials*, 23(16), 3523-3531.

- Su, Bai-hai., Fu, Ping., Li, Q., Tao, Y., Li, Z., Zao, H.-s., & Zhao, C.-s. (2008). Evaluation of polyethersulfone highflux hemodialysis membrane in vitro and in vivo. *Journal of Materials Science: Materials in Medicine*, 19(2), 745-751.
- Sun, M., Su, Y., Mu, C., & Jiang, Z. (2009). Improved antifouling property of PES ultrafiltration membranes using additive of silica– PVP nanocomposite. *Industrial & Engineering Chemistry Research*, 49(2), 790-796.
- Sun, S., Yue, Y., Huang, X., & Meng, D. (2003). Protein adsorption on blood-contact membranes. *Journal of Membrane Science*, 222(1), 3-18.
- Tagmatarchis, N., & Prato, M. (2004). Functionalization of carbon nanotubes via 1, 3-dipolar cycloadditions. *Journal of materials chemistry*, 14(4), 437-439.
- Tang, M., Xue, J., Yan, K., Xiang, T., Sun, S., & Zhao, C. (2012a). Heparin-like surface modification of polyethersulfone membrane and its biocompatibility. *Journal of Colloid and Interface Science*, 386(1), 428-440.
- Tang, M., Xue, J., Yan, K., Xiang, T., Sun, S., & Zhao, C. (2012b). Heparin-like surface modification of polyethersulfone membrane and its biocompatibility. *Journal of colloid and interface science*, 386(1), 428-440.
- Taniguchi, M., & Belfort, G. (2004). Low protein fouling synthetic membranes by UV-assisted surface grafting modification: varying monomer type. *Journal of Membrane Science*, 231(1), 147-157.
- Taniguchi, M., Kilduff, J. E., & Belfort, G. (2003). Low fouling synthetic membranes by UV-assisted graft polymerization: monomer selection to mitigate fouling by natural organic matter. *Journal of Membrane Science*, 222(1), 59-70.
- Tokars, J. I., Alter, M. J., Favetro, M. S., Moyer, L. A., Miller, E., & Bland, L. A. (1994). National surveillance of dialysis associated diseases in the United States, 1992. *ASAIO journal*, 40(4), 1020-1031.
- Tripathi, B. P., Chakrabarty, T., & Shahi, V. K. (2010). Highly charged and stable cross-linked 4, 4'-bis (4-aminophenoxy) biphenyl-3, 3'-disulfonic acid (BAPBDS)-sulfonated poly (ether sulfone) polymer electrolyte membranes impervious to methanol. *Journal of Materials Chemistry*, 20(37), 8036-8044.
- Tripathi, B. P., Dubey, N. C., & Stamm, M. (2014). Polyethylene glycol cross-linked sulfonated polyethersulfone based filtration membranes with improved antifouling tendency. *Journal of Membrane Science*, 453, 263-274.

- Tsuruoka, S., Kawaguchi, A., Nishiki, K., Hayasaka, T., Fukushima, C., Sugimoto, K., Fujimura, A. (2002). Vitamin E-bonded hemodialyzer improves neutrophil function and oxidative stress in patients with end-stage renal failure. *American Journal of Kidney Diseases*, 39(1), 127-133.
- Tu, X., & Zheng, M. (2008). A DNA-based approach to the carbon nanotube sorting problem. *Nano Research*, 1(3), 185-194.
- Tullis, R. H., Duffin, R. P., Zech, M., & Ambrus, J. L. (2002). Affinity Hemodialysis for Antiviral Therapy. I. Removal of HIV-1 from Cell Culture Supernatants, Plasma, and Blood. *Therapeutic Apheresis*, 6(3), 213-220.
- Tur, E., Onal-Ulusoy, B., Akdogan, E., & Mutlu, M. (2012). Surface modification of polyethersulfone membrane to improve its hydrophobic characteristics for waste frying oil filtration: Radio frequency plasma treatment. *Journal of applied polymer science*, 123(6), 3402-3411.
- Ueda, T., Oshida, H., Kurita, K., Ishihara, K., & Nakabayashi, N. (1992). Preparation of 2-methacryloyloxyethyl phosphorylcholine copolymers with alkyl methacrylates and their blood compatibility. *Polymer Journal-Tokoyo-*, 24, 1259-1259.
- Ulbricht, M., & Riedel, M. (1998). Ultrafiltration membrane surfaces with grafted polymer 'tentacles': preparation, characterization and application for covalent protein binding. *Biomaterials*, 19(14), 1229-1237.
- Umeyama, T., Tezuka, N., Fujita, M., Matano, Y., Takeda, N., Murakoshi, K., . . . Imahori, H. (2007). Retention of intrinsic electronic properties of soluble single-walled carbon nanotubes after a significant degree of sidewall functionalization by the Bingel reaction. *The Journal of Physical Chemistry C*, 111(27), 9734-9741.
- Unveren, E., Erdogan, T., Celebi, S. S., & Inan, T. Y. (2010). Role of post-sulfonation of poly (ether ether sulfone) in proton conductivity and chemical stability of its proton exchange membranes for fuel cell. *International Journal of Hydrogen Energy*, 35(8), 3736-3744.
- Ursini, C. L., Maiello, R., Ciervo, A., Fresegna, A. M., Buresti, G., Superti, F., Cavallo, D. (2015). Evaluation of uptake, cytotoxicity and inflammatory effects in respiratory cells exposed to pristine and-OH and-COOH functionalized multi-wall carbon nanotubes. *Journal of Applied Toxicology*.

- Valcárcel, M., Cárdenas, S., Simonet, B., Moliner-Martinez, Y., & Lucena, R. (2008). Carbon nanostructures as sorbent materials in analytical processes. *TrAC Trends in Analytical Chemistry*, 27(1), 34-43.
- Van der Heiden, A., Willems, G., Lindhout, T., Pijpers, A., & Koole, L. (1998). Adsorption of proteins onto poly (ether urethane) with a phosphorylcholine moiety and influence of preadsorbed phospholipid. *Journal of biomedical materials research*, 40(2), 195-203.
- van Reis, R., & Zydney, A. (2007). Bioprocess membrane technology. *Journal of Membrane Science*, 297(1), 16-50.
- Vandenberg, L. N., Hauser, R., Marcus, M., Olea, N., & Welshons, W. V. (2007). Human exposure to bisphenol A (BPA). *Reproductive toxicology*, 24(2), 139-177.
- Vandentorren, S., Zeman, F., Morin, L., Sarter, H., Bidondo, M.-L., Oleko, A., & Leridon, H. (2011). Bisphenol-A and phthalates contamination of urine samples by catheters in the Elfe pilot study: implications for large-scale biomonitoring studies. *Environmental research*, 111(6), 761-764.
- Vanholder, R., De Smet, R., Glorieux, G., Argilés, A., Baurmeister, U., Brunet, P., . . . Deppisch, R. (2003). Review on uremic toxins: classification, concentration, and interindividual variability. *Kidney international*, 63(5), 1934-1943.
- Vanholder, R. C., De Smet, R. V., & Ringoir, S. (1992). Assessment of urea and other uremic markers for quantification of dialysis efficacy. *Clinical chemistry*, 38(8), 1429-1436.
- Vardharajula, S., Ali, S. Z., Tiwari, P. M., Eroğlu, E., Vig, K., Dennis, V. A., & Singh, S. R. (2012). Functionalized carbon nanotubes: biomedical applications. *International journal of nanomedicine*, 7, 5361.
- Vatanpour, V., Madaeni, S. S., Moradian, R., Zinadini, S., & Astinchap, B. (2011). Fabrication and characterization of novel antifouling nanofiltration membrane prepared from oxidized multiwalled carbon nanotube/polyethersulfone nanocomposite. *Journal of Membrane Science*, 375(1), 284-294.
- Verniory, A., Du Bois, R., Decoodt, P., Gasee, J., & Lambert, P. (1973). Measurement of the Permeability of Biological Membranes Application to the glomerular wall. *The Journal of general physiology*, 62(4), 489-507.

- Vivekchand, S., Sudheendra, L., Sandeep, M., Govindaraj, A., & Rao, C. (2002). A Study of Polyaniline–Carbon Nanotube Composites. *Journal of nanoscience and nanotechnology*, 2(6), 631-635.
- Von Albertini, B., & Bosch, J. (1991). Short hemodialysis. *American journal of nephrology*, 11(3), 169-173.
- Wachtfogel, Y., Bischoff, R., Bauer, R., Hack, C., Nuijens, J., Kucich, U., . . . Colman, R. (1994). Alpha 1-antitrypsin Pittsburgh (Met358--> Arg) inhibits the contact pathway of intrinsic coagulation and alters the release of human neutrophil elastase during simulated extracorporeal circulation. *Thrombosis and haemostasis*, 72(6), 843.
- Wang, D.-a., Williams, C. G., Li, Q., Sharma, B., & Elisseeff, J. H. (2003). Synthesis and characterization of a novel degradable phosphate-containing hydrogel. *Biomaterials*, 24(22), 3969-3980.
- Wang, D., Zou, W., Li, L., Wei, Q., Sun, S., & Zhao, C. (2011). Preparation and characterization of functional carboxylic polyethersulfone membrane. *Journal of Membrane Science*, 374(1), 93-101.
- Wang, H., Yang, L., Zhao, X., Yu, T., & Du, Q. (2009). Improvement of hydrophilicity and blood compatibility on polyethersulfone membrane by blending sulfonated polyethersulfone. *Chinese Journal of Chemical Engineering*, 17(2), 324-329.
- Wang, H., Yu, T., Zhao, C., & Du, Q. (2009). Improvement of hydrophilicity and blood compatibility on polyethersulfone membrane by adding polyvinylpyrrolidone. *Fibers and polymers*, 10(1), 1-5.
- Wang, P., Tan, K., Kang, E., & Neoh, K. (2002). Plasma-induced immobilization of poly (ethylene glycol) onto poly (vinylidene fluoride) microporous membrane. *Journal of Membrane Science*, 195(1), 103-114.
- Wang, Y.-Q., Su, Y.-L., Ma, X.-L., Sun, Q., & Jiang, Z.-Y. (2006). Pluronic polymers and polyethersulfone blend membranes with improved fouling-resistant ability and ultrafiltration performance. *Journal of Membrane Science*, 283(1), 440-447.
- Wang, Y.-q., Wang, T., Su, Y.-l., Peng, F.-b., Wu, H., & Jiang, Z.-y. (2006). Protein-adsorption-resistance and permeation property of polyethersulfone and soybean phosphatidylcholine blend ultrafiltration membranes. *Journal of Membrane Science*, 270(1), 108-114.

- Wang, Y.-X., Robertson, J. L., Spillman Jr, W. B., & Claus, R. O. (2004). Effects of the chemical structure and the surface properties of polymeric biomaterials on their biocompatibility. *Pharmaceutical research*, 21(8), 1362-1373.
- Ward, R. A., & McLeish, K. R. (2003). Oxidant stress in hemodialysis patients: what are the determining factors? *Artificial organs*, 27(3), 230-236.
- Wavhal, D. S., & Fisher, E. R. (2002). Hydrophilic modification of polyethersulfone membranes by low temperature plasma-induced graft polymerization. *Journal of Membrane Science*, 209(1), 255-269.
- Wavhal, D. S., & Fisher, E. R. (2003). Membrane surface modification by plasma-induced polymerization of acrylamide for improved surface properties and reduced protein fouling. *Langmuir*, 19(1), 79-85.
- Wei, H., Han, L., Ren, J., & Jia, L. (2013). Anticoagulant surface coating using composite polysaccharides with embedded heparin-releasing mesoporous silica. *ACS applied materials & interfaces*, 5(23), 12571-12578.
- Werner, C., Jacobasch, H.-J., & Reichelt, G. (1996). Surface characterization of hemodialysis membranes based on streaming potential measurements. *Journal of Biomaterials Science, Polymer Edition*, 7(1), 61-76.
- Wick, P., Manser, P., Limbach, L. K., Dettlaff-Weglikowska, U., Krumeich, F., Roth, S., Bruinink, A. (2007). The degree and kind of agglomeration affect carbon nanotube cytotoxicity. *Toxicology letters*, 168(2), 121-131.
- Williams, J. A., Hyland, R., Jones, B. C., Smith, D. A., Hurst, S., Goosen, T. C., Ball, S. E. (2004). Drug-drug interactions for UDP-glucuronosyltransferase substrates: a pharmacokinetic explanation for typically observed low exposure (AUC_i/AUC) ratios. *Drug Metabolism and Disposition*, 32(11), 1201-1208.
- Wilson, N. R., & Macpherson, J. V. (2009). Carbon nanotube tips for atomic force microscopy. *Nature nanotechnology*, 4(8), 483-491.
- Witham, M. J., & Johnson, J. S. (2001). Non-cracking hydrophilic polyethersulfone membranes. *U.S. Patent No. 6,193,077*. 27.
- Wolff, S. H., & Zydny, A. L. (2004). Effect of bleach on the transport characteristics of polysulfone hemodialyzers. *Journal of Membrane Science*, 243(1), 389-399.

- Woods, H., & Nandakumar, M. (2000). Improved outcome for haemodialysis patients treated with high-flux membranes. *Nephrology Dialysis Transplantation*, 15(suppl 1), 36-42.
- Xie, X.-L., Mai, Y.-W., & Zhou, X.-P. (2005). Dispersion and alignment of carbon nanotubes in polymer matrix: a review. *Materials Science and Engineering: R: Reports*, 49(4), 89-112.
- Xu, Z.-K., Nie, F.-Q., Qu, C., Wan, L.-S., Wu, J., & Yao, K. (2005). Tethering poly (ethylene glycol) s to improve the surface biocompatibility of poly (acrylonitrile-co-maleic acid) asymmetric membranes. *Biomaterials*, 26(6), 589-598.
- Y. Ho, J., Matsuura, T., & P. Santerre, J. (2000). The effect of fluorinated surface modifying macromolecules on the surface morphology of polyethersulfone membranes. *Journal of Biomaterials Science, Polymer Edition*, 11(10), 1085-1104.
- Yamasaki, H., Nagake, Y., & Makino, H. (2001). Determination of bisphenol a in effluents of hemodialyzers. *Nephron*, 88(4), 376-378.
- Yamashita, A. C., & Tomisawa, N. (2009). Importance of membrane materials for blood purification devices in critical care. *Transfusion and Apheresis Science*, 40(1), 23-31.
- Yang, Q., Chung, T.-S., Chen, S. B., & Weber, M. (2008). Pioneering explorations of rooting causes for morphology and performance differences in hollow fiber kidney dialysis membranes spun from linear and hyperbranched polyethersulfone. *Journal of Membrane Science*, 313(1), 190-198.
- Yang, Q., Chung, T.-S., & Santoso, Y. (2007). Tailoring pore size and pore size distribution of kidney dialysis hollow fiber membranes via dual-bath coagulation approach. *Journal of Membrane Science*, 290(1), 153-163.
- Yang, Q., Chung, T.-S., & Weber, M. (2009). Microscopic behavior of polyvinylpyrrolidone hydrophilizing agents on phase inversion polyethersulfone hollow fiber membranes for hemofiltration. *Journal of Membrane Science*, 326(2), 322-331.
- Yehia, H. N., Draper, R. K., Mikoryak, C., Walker, E. K., Bajaj, P., Musselman, I. H., . . . Pantano, P. (2007). Single-walled carbon nanotube interactions with HeLa cells. *Journal of Nanobiotechnology*, 5(1), 1.

- Yin, G., Janson, J.-C., & Liu, Z. (2000). Characterization of protein adsorption on membrane surface by enzyme linked immunoassay. *Journal of Membrane Science*, 178(1), 99-105.
- Yin, Z., Su, B., Nie, S., Wang, D., Sun, S., & Zhao, C. (2012). Poly (vinylpyrrolidone-co-acrylonitrile-co-vinylpyrrolidone) modified polyethersulfone hollow fiber membranes with improved blood compatibility. *Fibers and Polymers*, 13(3), 269-276.
- Ying, Y., Saini, R. K., Liang, F., Sadana, A. K., & Billups, W. (2003). Functionalization of carbon nanotubes by free radicals. *Organic letters*, 5(9), 1471-1473.
- Zhang, H., Li, H. X., & Cheng, H. M. (2006). Water-soluble multiwalled carbon nanotubes functionalized with sulfonated polyaniline. *The Journal of Physical Chemistry B*, 110(18), 9095-9099.
- Zhang, Q., Zhang, S., Dai, L., & Chen, X. (2010). Novel zwitterionic poly (arylene ether sulfone) s as antifouling membrane material. *Journal of Membrane Science*, 349(1), 217-224.
- Zhang, W., Zhang, Z., & Zhang, Y. (2011). The application of carbon nanotubes in target drug delivery systems for cancer therapies. *Nanoscale research letters*, 6(1), 1.
- Zhang, Z., Chao, T., Chen, S., & Jiang, S. (2006). Superlow fouling sulfobetaine and carboxybetaine polymers on glass slides. *Langmuir*, 22(24), 10072-10077.
- Zhao, C., Liu, X., Nomizu, M., & Nishi, N. (2003). Blood compatible aspects of DNA-modified polysulfone membrane—protein adsorption and platelet adhesion. *Biomaterials*, 24(21), 3747-3755.
- Zhao, C., Liu, X., Rikimaru, S., Nomizu, M., & Nishi, N. (2003). Surface characterization of polysulfone membranes modified by DNA immobilization. *Journal of Membrane Science*, 214(2), 179-189.
- Zhao, C., Xue, J., Ran, F., & Sun, S. (2013). Modification of polyethersulfone membranes—a review of methods. *Progress in Materials Science*, 58(1), 76-150.
- Zhao, C. S., Xue, J. M., Ran, F., & Sun, S. D. (2013). Modification of polyethersulfone membranes - A review of methods. *Progress in Materials Science*, 58(1), 76-150.

- Zhao, W., Huang, J., Fang, B., Nie, S., Yi, N., Su, B., Zhao, C. (2011). Modification of polyethersulfone membrane by blending semi-interpenetrating network polymeric nanoparticles. *Journal of Membrane Science*, 369(1), 258-266.
- Zhao, W., Mou, Q., Zhang, X., Shi, J., Sun, S., & Zhao, C. (2013). Preparation and characterization of sulfonated polyethersulfone membranes by a facile approach. *European Polymer Journal*, 49(3), 738-751.
- Zhao, W., Su, Y., Li, C., Shi, Q., Ning, X., & Jiang, Z. (2008). Fabrication of antifouling polyethersulfone ultrafiltration membranes using Pluronic F127 as both surface modifier and pore-forming agent. *Journal of Membrane Science*, 318(1), 405-412.
- Zhou, H., Cheng, C., Qin, H., Ma, L., He, C., Nie, S., Zhao, C. (2014). Self-assembled 3D biocompatible and bioactive layer at the macro-interface via graphene-based supermolecules. *Polymer Chemistry*, 5(11), 3563-3575.
- Zhu, B., Eurell, T., Gunawan, R., & Leckband, D. (2001). Chain-length dependence of the protein and cell resistance of oligo (ethylene glycol)-terminated self-assembled monolayers on gold. *Journal of biomedical materials research*, 56(3), 406-416.
- Zhu, L.-P., Yi, Z., Liu, F., Wei, X.-Z., Zhu, B.-K., & Xu, Y.-Y. (2008). Amphiphilic graft copolymers based on ultrahigh molecular weight poly (styrene-*alt*-maleic anhydride) with poly (ethylene glycol) side chains for surface modification of polyethersulfone membranes. *European Polymer Journal*, 44(6), 1907-1914.
- Zhu, L.-P., Zhang, X.-X., Xu, L., Du, C.-H., Zhu, B.-K., & Xu, Y.-Y. (2007). Improved protein-adsorption resistance of polyethersulfone membranes via surface segregation of ultrahigh molecular weight poly (styrene-*alt*-maleic anhydride). *Colloids and Surfaces B: Biointerfaces*, 57(2), 189-197.
- Zhu, L., Wang, J., Zhu, B., & Xu, Y. (2008). Molecular design and synthesis of amphiphilic copolymers, and the performances of their blend membranes. *Acta Polymerica Sinica*, 4, 309.