FRACTIONATION OF ROSMARINIC ACID RICH EXTRACT FROM ORTHOSIPHON STAMINEUS FOR ENZYMATIC ANTI-DIABETIC ACTIVITY

NGO YI LEI

A thesis submitted in fulfilment of the requirements for the award of the degree of Master of Philosophy

Faculty of Chemical and Energy Engineering
Universiti Teknologi Malaysia

To my beloved family, supervisor and friends

ACKNOWLEDGEMENTS

For my accomplishment, I would like to express my deepest appreciation and sincerest gratitude to those who supported me throughout my Master degree studies. I would foremost to thank my supervisor, Assoc. Prof. Dr. Chua Lee Suan for her helpful guidance and advice throughout my research project. Her kind motivation, constructive suggestions, and immense knowledge have made my research works completed successfully.

I would also like to extend many thanks to the post-graduate seniors of Institute of Bioproduct Development, Universiti Teknologi Malaysia, for their invaluable guidance and excellent assistance throughout the conduction of bioprocessing works in the laboratory.

Lastly but not least, my special thanks to my family and friends for their constant moral support and encouragement from the beginning until the end of the project. Without their continuous assistances and patience, this study would not have succeeded on time.

ABSTRACT

Orthosiphon stamineus (O. stamineus) is one of the herbal plants containing high amount of rosmarinic acid. However, there is no standard guideline, specifically for rosmarinic acid rich extract from O. stamineus. Therefore, this study was focused on the fractionation of rosmarinic acid from the O. stamineus crude extract using chromatographic methods, namely thin layer chromatography integrated with column chromatography (TLC-CC) and liquid-liquid extraction integrated with solid phase extraction (LLE-SPE). Both fractionation methods were compared, and it was found that TLC-CC performed better than LLE-SPE to recover rosmarinic acid from the O. stamineus crude extract. The purity of the rosmarinic acid rich extract was 100% (fraction 8) which was greater than sub-fraction 1-3 (75%) prepared by LLE-SPE. TLC-CC increased the rosmarinic acid content from 3.6% w/w in crude extract to 100.0% w/w in the fraction 8, while LLE-SPE only increased the content of rosmarinic acid up to 75.0% w/w in the combined sub-fraction 1 to 3. The increase of rosmarinic acid content also simultaneously increased enzymatic anti-diabetic activities compared to the crude extract. The fraction of 100% rosmarinic acid from TLC-CC significantly (P < 0.05) inhibited the activity of α -amylase and α -glucosidase with IC₅₀ values of 2.31 mg/mL and 0.34 mg/mL, respectively. The IC₅₀ values were comparable to those results of standard drug, acarbose which showed 1.03 mg/mL and 1.66 mg/mL for αamylase and α-glucosidase, respectively. Kinetic studies revealed that the 100% rosmarinic acid fraction inhibited α -amylase competitively, but non-competitively for α-glucosidase inhibition. The introduction of rosmarinic acid as an inhibitor slow down the digestion rate of starch by α -amylases, and reduce the performance of α glucosidase by decreasing its V_{max} value to 0.05 mM/min. Understanding the kinetic information of the enzymatic reaction is important to predict in vivo metabolism of rosmarinic acid for drug design. In conclusion, high anti-diabetic property rosmarinic acid rich extract from O. stamineus can be obtained using column chromatography technology.

ABSTRAK

Orthosiphon stamineus (O. stamineus) merupakan salah satu herba yang mengandungi asid rosmarinik yang banyak. Walau bagaimanapun, tiada garis panduan piawai khusus bagi penyediaan asid rosmarinik dari O. stamineus. Oleh itu, kajian ini telah tertumpu kepada pemeringkatan asid rosmarinik daripada ekstrak mentah O. stamineus dengan menggunakan kaedah kromatografi, iaitu kromatografi lapisan nipis bersepadu dengan kromatografi turus (KLN-KT) dan pengekstrakan cecair-cecair bergabung dengan pengekstrakan fasa pepejal (PCC-PFP). Kedua-dua kaedah pemeringkatan ini telah dibandingkan dan didapati bahawa KLN-KT menghasilkan lebih banyak asid rosmarinik berbanding PCC-PFP daripada ekstrak mentah O. stamineus. Ketulenan ekstrak kaya asik rosmarinik adalah 100% (pecahan 8) dan ianya lebih tulen berbanding sub-pecahan 1-3 (75%) yang dihasilkan melalui kaedah PCC-PFP. KLN-KT meningkatkan kandungan asid rosmarinik daripada 3.6% berat dalam ekstrak mentah kepada 100% berat dalam pecahan 8, manakala PCC-PFP hanya meningkatkan kandungan asid rosmarinik kepada 75% berat dalam gabungan sub-pecahan 1 hingga 3. Peningkatan kandungan asid rosmarinik juga meningkatkan aktiviti perencatan enzim kencing manis berbanding dengan ekstrak mentah. Pecahan 100% asid rosmarinik dari KLN-KT merencat aktiviti α-amilase dan αglukosidase secara ketara (P < 0.05) dengan nilai IC₅₀ masing-masing ialah 2.31 dan 0.34 mg/mL. Nilai IC₅₀ ini setanding dengan ubat piawai, acarbose yang menunjukkan nilainya masing-masing pada 1.03 dan 1.66 mg/mL bagi α-amilase dan α-glukosidase. Kajian kinetik menunjukkan bahawa pecahan 100% asik rosmarinik merencat α-amilase secara kompetitif, tetapi tidak bagi α-glukosidase. Penggunaan asid rosmarinik sebagai perencat dapat melambatkan kadar penghadaman kanji oleh α-amilases, dan mengurangkan prestasi α-glukosidase dengan menurunkan nilai V_{max} kepada 0.05 mM/min. Pemahaman tentang kinetik enzim tersebut adalah penting untuk meramalkan metabolisma asid rosmarinik secara in vivo dalam rekaan ubat. Kesimpulannya, pecahan asid rosmarinik yang mempunyai potensi tinggi untuk mengurang gejala kencing manis boleh diperolehi daripada O. stamineus melalui teknologi kromatografi turus.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	V
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xi
	LIST OF FIGURES	xii
	LIST OF APPENDICES	XV
	LIST OF ABBREVIATIONS	xvi
1	INTRODUCTION	1
	1.1 Research Background	1
	1.2 Research Problem	4
	1.3 Research Objectives	5
	1.4 Research Scopes	6
	1.5 Research Significance	7
2	LITERATURE REVIEW	8
	2.1 Background of <i>Orthosiphon stamineus</i>	8
	2.1.1 Phytochemicals in <i>Orthosiphon</i> Stamineus	10

		viii
	2.1.2 Medicinal Uses of <i>Orthosiphon</i> Stamineus	14
2.2	Rosmarinic Acid and Its Pharmacological Activities	15
2.3	Extraction Techniques for Herbal Plant	19
2.4	Fractionation of Crude Extract	22
	2.4.1 Liquid-Liquid Extraction	23
	2.4.2 Solid-Liquid Extraction	24
2.5	Chromatographic Analysis of Herbal Extract and Fractions	32
	2.5.1 Thin Layer Chromatography	32
	2.5.2 High Performance Liquid Chromatography	35
	2.5.3 Liquid Chromatography-Tandem Mass Spectrometer	39
2.6	Pathophysiology of Diabetes Mellitus	42
2.7	Types of Diabetic Mellitus	45
2.8	Anti-diabetic Agents	47
2.9	Techniques of Anti-Diabetic Evaluation	50
	2.9.1 Enzymatic α-amylase Inhibitory Assay	51
	2.9.2 Enzymatic α-glucosidase Inhibitory Assay	5 0
	2.9.3 Kinetics of Enzyme Inhibition	53
2.10	Chapter Summary	54
		60
MAT	TERIALS AND METHODOLOGY	62
3.0	Introduction to Overall Study	62
3.1	Chemicals and Reagents	64
3.2	Reflux Technique for Crude Extract Preparation	64

Column

65

3

Fractionation

Chromatography

Based

on

3.3

		3.3.1	Solvent System Development by Thin Layer Chromatography	65
		3.3.2	Optimization of Solvent System in Column Chromatography	66
		3.3.3	Optimization of Packed Column Height	67
		3.3.4	Stability of Absorbent	68
	3.4		Dimensional Fractionation Based on -Liquid Extraction and Solid Phase tion	68
		3.4.1	Sample Partitioning by Liquid-Liquid Extraction	68
		3.4.2	Second Dimension of Solid Phase Extraction for Rosmarinic Acid Rich Fraction	69
	3.5	Quanti MS/M	fication of Rosmarinic Acid by LC-DAD-S	70
	3.6	Anti-d	iabetic Enzymatic Activity	70
		3.6.1	α-amylase Inhibitory Assay	71
		3.6.2	α-glucosidase Inhibitory Assay	72
		3.6.3	Mode of α-amylase Inhibition	72
		3.6.4	Mode of α -glucosidase Inhibition	73
		3.6.5	Calculation of 50% Inhibitory Concentration	74
	3.7	Design	of Experiments	74
	3.8	Statisti	ical Analysis	74
1	RESU	LTS A	ND DISCUSSION	75
	4.1		Extraction of <i>Orthosiphon Stamineus</i> Extract	75
	4.2	Thin	onation of Rosmarinic Acid Based on Layer Chromatography Integrated with on Chromatography	77
		4.2.1	Development of Solvent System from Thin Layer Chromatography	78

		4.2.2	Effect of Solvent Systems on Rosmarinic Acid Recovery from Column Chromatography	82
		4.2.3	Effect of Packed Column Height on Rosmarinic Acid Content	88
		4.2.4	Stability of Adsorbent	90
	4.3	Acid	limensional Fractionation of Rosmarinic Based on Liquid-Liquid Extraction and Phase Extraction	93
		4.3.1	Sample Partitioning by Liquid-Liquid Extraction	93
		4.3.2	Second Dimension of Solid Phase Extraction for Rosmarinic Acid Rich Fraction	95
	4.4	Enzyn	natic Anti-diabetic Activity	98
		4.4.1	<i>In vitro</i> α-amylase Inhibition Study	98
		4.4.2	<i>In vitro</i> α-glucosidase Inhibition Study	103
5	CON	CLUSI	ON	109
	5.1	Concl	usion	109
	5.2	Recon	nmendations	111
REFERENCES	5			112
Appendices A-C				131

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Plant taxonomy of Orthosiphon stamineus	9
2.2	Phytochemicals in Orthosiphon stamineus	12
2.3	Chemical and physical properties of rosmarinic acid	16
2.4	Rosmarinic acid content in different plants	17
3.1	Different solvent ratios of mobile phases	66
4.1	Recovery and concentration of rosmarinic acid from each sub-fraction in solid phase extraction	96
4.2	IC $_{50}$ of samples for α -amylase inhibitory activity	101
4.3	IC ₅₀ of samples for α-glucosidase inhibitory activity	104

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Orthosiphon stamineus in Malaysia	9
2.2	Leaf morphological observation of (a) white and (b) purple varieties of <i>Orthosiphon stamineus</i>	10
2.3	Chemical structure of rosmarinic acid	16
2.4	Hydrogen donation mechanism of rosmarinic acid	18
2.5	Column chromatography involves a mobile phase flowing over a stationary phase	25
2.6	Column chromatography, (a) before fractionation and (b) separation of compound bands after a period of fractionation	26
2.7	Relative polarity of various solvents	29
2.8	A developed TLC plate with spots visualized and $R_{\rm f}$ values determined	34
2.9	Chromatogram of Orthosiphon stamineus leaves extracts	37
2.10	Ultraviolet-visible spectroscopy of rosmarinic acid from (a) <i>Orthosiphon stamineus</i> and (b) <i>Rosmarinus officinalis</i>	38
2.11	Principle of tandem mass spectrometry	40
2.12	ESI-MS/MS spectrum of rosmarinic acid at m/z 359.10	41
2.13	Glycogenolysis pathway	44
2.14	Gluconeogenesis pathway	45
2.15	Use of rosmarinic acid in inhibiting the starch digestion process by α -amylase and α -glucosidase	51

X	1	11	

2.16	Determining the initial velocity of an enzyme reaction from the slope of the graph at the beginning of a reaction		
2.17	Michaelis-Menten kinetics of an enzyme reaction	56	
2.18	Double reciprocal Lineweaver-Burk lines of competitive inhibition	58	
2.19	Double reciprocal Lineweaver-Burk lines of uncompetitive inhibition	59	
2.20	Double reciprocal Lineweaver-Burk lines of non-competitive inhibition	60	
3.1	Flowchart of the overall study	63	
4.1	Chromatogram of <i>Orthosiphon stamineus</i> crude extract at 254 nm	77	
4.2	TLC images of <i>Orthosiphon stamineus</i> crude extract using the mobile phase of ethyl acetate-ethanol (70:30 % v/v) with (a) and without (b) 0.1 % formic acid as additive	78	
4.3	Thin layer chromatogram for standard rosmarinic acid and <i>Orthosiphon stamineus</i> crude extract at different ratios of solvent systems (ethyl acetate and ethanol) with 0.1% v/v formic acid	80	
4.4	$R_{\rm f}$ values of standard rosmarinic acid and rosmarinic acid in crude extract at different concentrations of ethyl acetate	81	
4.5	Eluent volume of different solvent systems required for complete rosmarinic acid elution by packed column fractionator	83	
4.6	Recovery and purity of each fraction at different solvent systems of ethyl acetate and ethanol with 0.1% v/v formic acid	84	
4.7	Comparison of recovery and purity of rosmarinic acid at the first rosmarinic acid containing fraction from all solvent systems	86	
4.8	Chromatograms of the first rosmarinic acid containing fraction at different ethyl acetate concentrations	87	
4.9	Eluent volume of different packed bed heights required for the collection of first and subsequent fractions containing rosmarinic acid	89	
4.10	The effect of packed bed height on recovery and purity of rosmarinic acid content	90	

•	
X1	V

4.11	Comparison of recovery and purity of first rosmarinic acid containing fraction (fraction 8) for each cycle	91
4.12	Liquid-liquid extraction of <i>Orthosiphon stamineus</i> crude extract using different solvent systems to recover rosmarinic acid	94
4.13	Chromatograms of diethyl ether, ethyl acetate and water fractions of <i>Orthosiphon stamineus</i> crude extract after liquid-liquid extraction	95
4.15	Chromatograms of the rosmarinic acid fraction after solid phase extraction with water-ethanol solvent system	97
4.16	Inhibitory activity of crude extract and its rosmarinic acid rich fractions on α -amylase	99
4.17	IC_{50} of samples on α -amylase inhibitory activity	100
4.18	Mode of competitive inhibition for α -amylase by 100% rosmarinic acid fraction from <i>Orthosiphon stamineus</i>	102
4.19	Inhibitory activity of each sample on α -glucosidase inhibition $in\ vitro$	103
4.20	IC $_{50}$ of samples on α -glucosidase inhibitory activity	105
4.21	Mode of non-competitive inhibition for α -glucosidase by 100% rosmarinic acid from <i>Orthosiphon stamineus</i>	106

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Results of thin layer chromatography	131
В	Results of anti-diabetic activity assay	132
C	Calibration curves of reference standard	135

LIST OF ABBREVIATIONS

Arg120 - Arginine 120

CC - Column chromatography

DAD - Diode array detector

DE - Diethyl acetate

DM - Diabetes mellitus

DMH - 1,2-dimethylhydrazine

DNS - 3,5, dinitrosalicylic acid

EA - Ethyl acetate

EC₅₀ - Half maximal effective concentration

ESI - Electrospray ionization

FA - Formic acid

GLUT-4 - Glucose transporter type 4

HDL - High-density lipoprotein

HFD - High-fat diet

HPLC - High-performance liquid chromatography

HSCCC - High-speed counter-current chromatography

IC₅₀ - Half maximal inhibitory concentration

K_m - Michaelis constant

LLE - Liquid-liquid extraction

m/z - Mass-to-charge ratio

MAE - Microwave-assisted extraction

MeOH - Methanol

MS - Mass spectrometer

PE - Petroleum ether

p-NPG - 4-nitrophenyl-α-D-glucopyranoside

R - Recovery

RA - Rosmarinic acid

R_f - Retention factor

Ser353 - Serine 353

SGLT1 - Sodium-glucose transport proteins

SPE - Solid phase extraction

SSL - Spent sulphite liquor

STZ - Streptozocin

TLC - Thin layer chromatography

UAE - Ultrasound assisted extraction

UV - Ultraviolet

V_{max} - Maximum reaction velocity

CHAPTER 1

INTRODUCTION

1.1 Research Background

Orthosiphon stamineus belongs to the family Lamiaceae, and this plant is also known as "Misai Kucing" which is literally meant cat's whisker in Malaysia. It is a perennial herb which can grow well in temperate and tropical areas up to 0.3–1.0 m with 4-angled, poorly ramified and ascending stem (Wiart, 2000). It has been traditionally used as folk medicine for illnesses such as rheumatoid diseases, diabetes, hypertension and renal calculus (Awale *et al.*, 2003). Owing to the promising medicinal values, *O. stamineus* has been formulated into many commercial products in the forms of tea sachets, powdered herb, tablets and capsules (Indubala and Ng, 2000). The extract of *O. stamineus* leaves and stems were found to have more than 20 phenolic compounds including rosmarinic acid, eupatorin, 3'-hydroxy-5, 6, 7, 4'-tetramethoxyflavone and sinensetin (Indubala and Ng, 2000).

Nowadays, entrepreneurs prefer to have herbal extract rich in bioactive compound instead of plant crude extract. The demand of rosmarinic acid rich extract is increasing in herbal products market nowadays (Shekarchi *et al.*, 2012). Therefore, this study was aimed to prepare the rosmarinic acid rich extract from *O. stamineus*.

Plant crude extract is very complex because it contains thousands of phytochemicals with diverse chemical properties. Therefore, a fractionation process is necessary to separate the target compound from the crude extract. Fractionation is a separation process to segregate a complex mixture into smaller fractions with higher quantity of desired compounds according to a gradient of the solvent system (WHO, 2017). The quality of the herbal extract could be enhanced by fractionating them according to their chemical characteristics, usually based on solvent property (WHO, 2017).

In the present study, the fractionation of *O. stamineus* crude extract was conducted by thin layer chromatography integrated with column chromatography (TLC-CC), and liquid-liquid extraction integrated with solid phase extraction (LLE-SPE). Both methods were evaluated and compared in the term of recovery and purity of rosmarinic acid. TLC is used to optimize the conditions of column chromatography by selecting suitable mobile and solid phases (Liu, 2011), while LLE acts as a pretreatment step before SPE to separate the crude extract based on the different distribution and solubility of the compounds between two immiscible liquid solvents (Houghton and Raman, 1998).

CC and SPE are simple fractionation techniques that are widely used for purification and separation of target compounds from the complex mixture which involves the sorption of solutes from a liquid medium into a solid adsorbent. The selection of adsorbent and solvent system are dominant factors affecting the recovery and purity of target compound (Williamson and Masters, 2016). Somehow, the option of impurity retention and target compound collection would always be the method of choice for time saving and effective solvent consumption. Liquid chromatography with diode array detector and tandem mass spectrometer (LC-DAD-MS/MS) was used to identify and quantify the rosmarinic acid.

Previous studies reported that rosmarinic acid showed significant antioxidant (Chen *et al.*, 2014), anti-cancer (Venkatachalam *et al.*, 2013), and anti-diabetic activities (Runtuwene *et al.*, 2016). Rosmarinic acid is a phenolic compound

(C₁₈H₁₆O₈) comprised of an ester of caffeic acid and 3,4-dihydroxyphenyllactic acid, which is commonly found in many plant families including Lamiaceae and Boraginaceae (Bandoniene *et al.*, 2005). Lau *et al.* (2014) reported that the *O. stamineus* contains higher amount of rosmarinic acid, approximately 4.1% w/w compared to other plants such as *Origanum vulgare* (0.94% w/w), *Rosmarinus officinalis* (0.2% w/w) and *Satureja macrostema* (0.06% w/w) (Alonso-Carrillo *et al.*, 2017; Jacotet-Navarro *et al.*, 2015; Baranauskaite *et al.*, 2016).

Recently, rosmarinic acid has extensively been studied in the management of diabetic conditions, which indicates that rosmarinic acid could be used to reduce the diabetes-induced disorders and complications (Rao *et al.*, 2014). Diabetes mellitus (DM) is a global health problem and the number of diabetic people is expected to increase to 366 million in year of 2030 and majority of them were type 2 diabetes (Sarah *et al.*, 2004). Hyperglycemia in type 2 diabetic patients is caused by a sudden increase in the blood glucose levels due to starch hydrolysis by enzymes such as α -amylase and α -glucosidases in gastrointestinal tract (Gray, 1975). The pancreatic α -amylase enzymes convert starch to maltose and iso-maltose, which then travel to the small intestine where they are further digested to monosaccharides such as glucose and fructose by α -glucosidase. The glucose and fructose are transported by the intestinal sodium-glucose cotransporter, and thereby increasing blood glucose level (Xiao *et al.*, 2013).

One of the commonly applied therapeutic approaches in the management of blood glucose level in type 2 DM is to control and decrease the postprandial hyperglycemia through inhibition of starch hydrolyzing enzymes such as α -amylase and α -glucosidases (Megh *et al.*, 2008). There are fewer reports available on the activity of rosmarinic acid in inhibiting the α -amylase and α -glucosidases to regulate diabetes mellitus (Azevedo *et al.*, 2011; McCue and Shetty, 2004; Runtuwene *et al.*, 2016). The currently available therapeutic options for diabetes like oral hypoglycemic agents and insulin have limitations of their own, therefore many herbal medicines have been recommended for diabetes treatment (Sharma, 2012). Hence, the inhibitory

activity of crude extract and rosmarinic acid fraction from O. stamineus on α -amylase and α -glucosidase enzymes were also investigated.

1.2 Research Problem

O. stamineus crude extract is very complex in its chemical composition. Hence, the crude extract is needed to fractionate into several fractions to concentrate the rosmarinic acid content. Till to date, there is no standard guideline, specifically for rosmarinic acid rich extract from O. stamineus using thin layer chromatography integrated with column chromatography, and liquid-liquid extraction integrated with solid phase extraction. Furthermore, it is difficult to maintain the quality of herbal extract for product formulation due to the difference in phytochemical profile of the herbal extract from one batch to another batch of processing. Hence, it is important to study the optimal fractionation conditions for the highest recovery and purity, as well as quality control of rosmarinic acid.

Previously, the optimization of the fractionation of rosmarinic acid from *O. stamineus* crude extract using column chromatography was carried out based on single factor (solvent system), which only recovered 4.10 % w/w rosmarinic acid (81.31 ppm) in 80% v/v ethanol fraction with a large consumption of solvent (1000 to 1500 mL) (Chua and Lau, 2016). Another study using solid phase extraction also showed poor separation of rosmarinic acid from *O. stamineus* extract using the solvent system of methanol-acetonitrile as almost all the phenolic compounds were eluted out in fraction 1 (Lau *et al.*, 2015). Therefore, this study was focused on the establishment of an optimal fractionation for the highest yield and recovery of rosmarinic acid.

Researchers have proven that the plants rich in phenolic compounds, especially rosmarinic acid could exhibit high antioxidant (Zheng and Wang, 2001), anti-allergic

(Ito et al., 1998), anti-hyperglycemic (Kumar et al., 2010), anti-inflammatory (Al-Sereiti et al., 1999), and antimicrobial (Nascimento et al., 2009) properties. The O. stamineus crude extract was found to have anti-diabetic activity (Rao et al., 2014). To the knowledge of researchers, there is no previous report on the in vitro inhibitory activity of rosmarinic acid from O. stamineus in α -amylase and α -glucosidases catalysed reaction, especially the kinetic study of the inhibitory mode caused by rosmarinic acid. Therefore, no clear understanding whether the inhibitory activity is contributed by either one or more compounds such as eupatorin and/or sinensetin in the O. stamineus crude extracts (McCue and Shetty, 2004). Possibly, the other compounds in the crude extract also contributed to the enzyme inhibitory activity. Therefore, this study was also determined the inhibitory potential of rosmarinic acid rich extract from O. stamineus on the activities of α -amylase and α -glucosidase concurrently.

1.3 Research Objectives

The followings are the objectives of this study:

- i. To extract rosmarinic acid containing crude extract from *Orthosiphon stamineus* using reflux extraction method.
- ii. To optimize the thin layer chromatography integrated with column chromatography technique in fractionating rosmarinic acid from *Orthosiphon stamineus* crude extract based on the solvent system, packed bed height, stability of adsorbent, and compare this system with liquid-liquid extraction integrated with solid phase extraction techniques.

iii. To determine the kinetics of inhibitory mode caused by rosmarinic acid rich extract in α -amylase and α -glucosidase catalysed anti-diabetic assays using Michaelis-Menten kinetic model.

1.4 Research Scopes

In order to achieve the objectives, the scopes of the study include:

- i. Extraction of rosmarinic acid from O. stamineus using a reflux extraction system with 70% v/v ethanol at 78.4 °C for 1 hour.
- ii. Fractionation of *O. stamineus* crude extract by a normal phase column chromatography using ethyl acetate-ethanol as the eluent system with formic acid as an additive to improve the elution of rosmarinic acid. Investigation of the stability of silica gel adsorbent (0.063-0.2 mm, 70-230 mesh) for rosmarinic acid fractionation by repeating the fractionation process using the same packed column.
- iii. Extraction of rosmarinic acid from *O. stamineus* crude extract using liquid-liquid extraction as pre-treatment step before solid phase extraction. Recovery of rosmarinic acid by solid phase extraction using the solvent system of water-ethanol. Comparison of the performance of both method in term of recovery and purity of rosmarinic acid.
- iv. Investigation of the anti-diabetic capacity of O. stamineus extract and rosmarinic acid rich extract based on the α -amylases and α -glucosidases

inhibitory activities. Determination of the inhibitory mode of rosmarinic acid rich extract in α -amylases and α -glucosidases catalysed reaction through Michaelis–Menten kinetic model to predict the mechanism of inhibition by rosmarinic acid in anti-hyperglycemic treatment.

1.5 Research Significance

In this study, the high yield of rosmarinic acid rich extract could be produced using column chromatography method. This method is considered as cost effective technique due to its minimal solvent consumption and high recovery of rosmarinic acid from the *O. stamineus* crude extract. The optimized fractionation conditions in the column chromatography is also very useful for herbal industry as the optimized conditions can be used to produce high purity of rosmarinic acid rich extract satisfactory. The production of rosmarinic acid from *O. stamineus* is more profitable, since it contains higher amount of rosmarinic acid (Akowuah *et al.*, 2004; Shekarchi *et al.*, 2012). The plant could be cultivated approximately after ten weeks of plantation (Zaharah, 2005).

Rosmarinic acid has been found to have potential health benefits, particularly on its anti-diabetic capacity. The rosmarinic acid rich extract from O. stamineus obtained from CC was found to be the most effective in inhibiting the α -amylases and α -glucosidases. Understanding of the kinetic of inhibitory mode caused by rosmarinic acid rich extract is very important for the early phase of clinical study. The rosmarinic acid rich extract could be the lead source to be developed into a natural anti-diabetic drug replacing the commonly used strong allopathic drugs that possess several harmful side effects. This study brings benefits to the diabetic patients as herbal formulations are lower cost and lesser side effects, so that the low to middle income families can be affordable with the treatment.

REFERENCES

- Abdullah, N. R., Ismail, Z., and Ismail, Z. (2007). Acute Toxicity of *Orthosiphon Stamineus* Benth Standardized Extract in Sprague Dawley Rats. *Phytomedicine*, 16, (2-3), 222-226.
- Afifi, A. F., Kamel, E. M., Khalil, A. A., Foaad, M. A., Fawziand, E. M., and Houseny, M. (2008). Purification and Characterization of A-Amylase from *Penicillium Olsonii* Under the Effect of Some Antioxidant Vitamins. *Global Journal of Biotechnology and Biochemistry*, 3(1), 12–14.
- Ahamed Basheer, M., and Abdul Majid, A. (2010). Medicinal Potentials of *Orthosiphon stamineus* Benth. *Webmed Central CANCER*, 1(12), 1-7.
- Akowuah, G. A., Zhari, I., Norhayati, I., Sadikun, A., and Khamsah, S. M. (2004). Sinensetin, Eupatorin, 3'-Hydroxy-5, 6, 7, 4'-Tetramethoxyflavone and Rosmarinic Acid Contents and Antioxidative Effect of *Orthosiphon stamineus* from Malaysia. *Food Chemistry*, 87(4), 559–566.
- Alexandri, M., Papapostolou, H., Vlysidis, A., Gardeli, C., Komaitis, M., Papanikolaou, S., and Koutinas, A. A. (2016). Extraction of Phenolic Compounds and Succinic Acid Production from Spent Sulphite Liquor. *Journal of Chemical Technology and Biotechnology*, 91(11), 2751-2760.
- Ali, H., Houghton, P. J., and Soumyanath, A. (2006). α-Amylase Inhibitory Activity of Some Malaysian Plants Used to Treat Diabetes; with Particular Reference to *Phyllantus amarus*. *Journal of Ethnopharmacology*, 107(3), 449-455.
- Almatar, M., and Rahmat, Z. (2014). Identifying the Developmental Stages and Optimizing the Sample Preparation for Anatomical Study of *Orthosiphon stamineus*. *Journal of Applied Pharmaceutical Science*, 4(3), 66-74.

- Alonso-Carrillo, N., Aguilar-Santamaría, M. D. L. Á., Vernon-Carter, E. J., Jiménez-Alvarado, R., Cruz-Sosa, F., and Román-Guerrero, A. (2017). Extraction of phenolic compounds from *Satureja macrostema* using microwave-ultrasound assisted and reflux methods and evaluation of their antioxidant activity and cytotoxicity. *Industrial Crops and Products*, 103, 213-221.
- Al-Sereiti, M. R., Abu-Amer, K. M., and Sen, P. (1999). Pharmacology of Rosemary (*Rosmarinus officinalis* Linn.) and its Therapeutic Potentials. *Indian Journal of Experimental Biology*, 37(2), 124–130.
- Ana, T., and Amadeu, L. (2012). A Prospect for Pyrrolidine Iminosugars as Antidiabetic α-Glucosidase Inhibitors. *Journal of Medicinal Chemistry*, 55(23), 10345-10346.
- Aung, H. T., Nikai, T., Niwa, M., and Takaya, Y. (2010). Rosmarinic acid in Argusia argentea inhibits snake venom-induced hemorrhage. *Journal of Natural Medicines*, 64(4), 482-486.
- Asgar, M. A. (2013). Anti-Diabetic Potential of Phenolic Compounds: A Review. *International Journal of Food Properties*, 16(1), 91-103.
- Awale, S., Tezuka, Y., Banskota, A. H., Shimoji, S., Taira, K., and Kadota, S. (2002). Norstaminane- and Isopimarane-type Diterpenes of *Orthosiphon stamineus* from Okinawa. *Tetrahedron*, 58(27), 5503-5512.
- Awale, S., Tezuka, Y., Banskota, A. H., Adnyana, I. K., and Kadota, S. (2003). Highly Oxygenated Isopimarane-Type Diterpenes from *Orthosiphon stamineus* of Indonesia and Their Nitric Oxide Inhibitory Activity. *Chemical and Pharmaceutical Bulletin*, 51(3), 268-275.
- Azevedo, M. F., Lima, C. F., Fernandes, F. M., Almeida, M. J., Wilson, J. M., and Pereira, W. C. (2011). Rosmarinic Acid, Major Phenolic Constituent of Greek Sage Herbal Tea, Modulates Rat Intestinal SGLT1 Levels with Effects on Blood Glucose. *Molecular Nutrition and Food Research*, 55(1), 15–25.
- Azwanida, N. N. (2015). A Review on the Extraction Methods Use in Medicinal Plants, Principle, Strength and Limitation. *Medicinal and Aromatic Plants*, 4, 196.
- Bandoniene, D., Murkovic, M., and Venskutonis, P. R. (2005). Determination of Rosmarinic Acid in Sage and Borage Leaves by High-Performance Liquid Chromatography with Different Detection Methods. *Journal of Chromatographic Science*, 43(7), 372-376.

- Baranauskaite, J., Jakštas, V., Ivanauskas, L., Kopustinskiene, D. M., Drakšiene, G., Masteikova, R., and Bernatoniene, J. (2016). Optimization of Carvacrol, Rosmarinic, Oleanolic and Ursolic Acid Extraction from Oregano Herbs (*Origanum onites* L., *Origanum vulgare* spp. hirtum and *Origanum vulgare* L.). *Natural Product Research*, 30, 672-674.
- Barrett, M. L., and Udani, K. K. (2011). A Proprietary Alpha-Amylase Inhibitor from White Bean (*Phaseolus vulgaris*): A Review of Clinical Studies on Weight Loss and Glycemic Control. *Nutrition Journal*, 10(24), 1-10.
- Baskan, S., Öztekin, N., and Erim, F. B. (2007). Determination of Carnosic Acid and Rosmarinic Acid in Sage by Capillary Electrophoresis. *Food Chem*istry, 101(4), 1748-1752.
- Bele, A. A., and Khale, A. (2011). An Overview on Thin Layer Chromatography. International Journal of Pharmaceutical Sciences and Research, 2(2), 256-267.
- Berg, J. M., Tymoczko, J. L., and Stryer, L. (2002). *Biochemistry* (5th ed.). New York: W H Freeman.
- Berger, W. (1985). Incidence of Severe Side Effects During Therapy with Sulfonylureas and Biguanides. *Hormone and Metabolic Research*. *Supplement* 1, 15, 111-115.
- Bhalodi, M., Shukla, S., and Saluja, A. K. (2008). *In vitro* Antioxidant Activity of the Flowers of *Ipomoea Aquatic* Forsk. *Pharmacognosy Magazine*, 4, 220-226.
- Bhat, M., Kothiwale, S. K., Tirmale, A. R., Bhargava, S. Y., and Joshi, B. N. (2009)

 Antidiabetic Properties of *Azardiracta indica* and *Bougainvillea*spectabilis: In Vivo Studies in Murine Diabetes Model. Evidence-Based

 Complementary and Alternative Medicine, 2011(2011), 1-9.
- Bhat, M., Zinjarde, S. S., Bhargava, S. Y., Kumar, A. R., and Joshi, B. N. (2011).

 Antidiabetic Indian Plants: A Good Source of Potent Amylase Inhibitors.

 Evidence-Based Complementary and Alternative Medicine, 2011, 1-6.
- Bhatt, R., Mishra, N., and Bansal, P. K. (2013). Phytochemical, Pharmacological and Pharmacokinetics Effects of Rosmarinic Acid. *Journal of Pharmaceutical and Scientific Innovation*, 2(2), 28-34.
- Bischcoff, H. (1994). Pharmacology of Alpha-Glucosidase Inhibition. *European Journal of Clinical Investigation*, 24(3), 3-10.
- Bogusz, M. J. (2000). Foreset Science: Handbook of Analytical Separations.

 Amsterdam: Elsevier Science B. V.

- Boivin, M, Zinsmeister, A. R., Go, V. L., and DiMagno, E. P. (1987). Effect of a Purified Amylase Inhibitor on Carbohydrate Metabolism after a Mixed Meal in Healthy Humans. *Mayo Clinic Proceedings*, 62(4), 249-255.
- Bray, G. A., and Greenway, F. L. (1999). Current and Potential Drugs for Treatment of Obesity. *Endocrine Reviews*, 20, 805-879.
- Buckle, D. R. (2001). *Encyclopedia of Reagents for Organic Synthesis*. Hoboken: John Wiley and Sons, Ltd.
- Buldinia, P. L., Riccib, L., Sharmac, J. L. (2002). Recent Applications of Sample Preparation Techniques in Food Analysis. *Journal of Chromatography A*, 975, 47–70.
- Bulgakov, V. P., Inyushkina, Y. V., and Fedoreyev, S. A. (2012). Rosmarinic Acid and its Derivatives: Biotechnology and Applications. *Critical Reviews in Biotechnology*, 32(3), 203-217.
- Cazes, J., and Scott, R. P. W. (2002). *Chromatography Theory*. New York: Marcel Dekker INC.
- Chan, L. K., and Loo, P. S. (2006). Morphological Similarities and Differences between the Two Varieties of Cat's Whiskers (*Orthosiphon stamineus* Benth.) Grown in Malaysia. *International Journal of Botany*, 2(1), 1-6.
- Che Mansor, C. N. A. N., Latip, J., and Markom, M. (2016). Preparation of *Orthosiphon stamineus* Enriched-Extracts and Evaluation of their Free Radical Scavenging Activity. In 2016 UKM FST Postgraduate Colloquium: Proceedings of the Universiti Kebangsaan Malaysia, Faculty of Science and Technology 2016 Postgraduate Colloquium (Vol. 1784). American Institute of Physics Inc.
- Chen, K. L., Li, H. X., Xu, X. L., and Zhou, G. H. (2014). The Protective Effect of Rosmarinic Acid on Hyperthermia-Induced C2C12 Muscle Cells Damage. *Molecular Biology Report*, 41(8), 5525-5531.
- Chew, K. K., Ng, S. Y., Thoo, Y. Y., Khoo, M. Z., Wan Aida, W. M. and Ho, C. W. (2011). Effect of Ethanol Concentration, Extraction Time and Extraction Temperature on the Recovery of Phenolic Compounds and Antioxidant Capacity of *Centella asiatica* extracts. *International Food Research Journal*, 18, 571-578.

- Chin, J. H., Abas, H. H., and Sabariah, I. (2008). Toxicity study of *Orthosiphon stamineus* Benth (Misai Kucing) on Sprague Dawley rats. *Tropical Biomedicine*, 25(1), 9-16.
- Chirinos, R., Rogez, H., Campos, D., Pedreschi, R. and Larondelle, Y. (2007). Optimization of Extraction Conditions of Antioxidant Phenolic Compounds from Mashua (*Tropaeolum tuberosum* Ruíz and Pavón) Tubers. *Separation and Purification Technology*, 55(2), 217-225.
- Choudhury, A., Maeda, K., Murayama, R., and DiMagno, E. P. (1996). Character of a Wheat Amylase Inhibitor Preparation and Effects on Fasting Human Pancreaticobiliary Secretions and Hormones. *Gastroenterology*, 111(5), 1313-1320.
- Christy, A. A. (2012). Effect of Heat on the Adsorption Properties of Silica Gel. *International Journal of Engineering and Technology*, 4(4), 484-488.
- Chua, L. S., and Lau, C. H. (2016). Reflux Extraction and Column Chromatography for Rosmarinic Acid-Rich Fraction from *Orthosiphon stamineus*. *The Natural Products Journal*, 6, 1-7.
- Chuichulcherma, S., Prommakortb, S., Srinophakunb, P., and Thanapimmetha, A. (2013). Optimization of Capsaicin Purification from Capsicum frutescens Linn. with Column Chromatography using Taguchi Design. Industrial Crops and Products, 44, 473-479.
- Collier, C. A., Bruce, C. R., Smith, A. C., Lopaschuk, G., and Dyck, D. J. (2006). Metformin Counters the Insulin-Induced Suppression of Fatty Acid Oxidation and Stimulation of Triacylglycerol Storage in Rodent Skeletal Muscle. *American Journal of Physiology. Endocrinology and Metabolism*, 291(1), 182–189.
- Conn, E. E., and Stumph, P. K. (1987). *Outline of Biochemistry* (5th ed.). New York: John Wiley and Sons.
- Cooper, E. L. (2004). Drug discovery, CAM and Natural Products. *Evidence-Based Complementary and Alternative Medicine*, 1(3), 215-217.
- Culberson, C. F. (1974). Conditions for the Use of Merck Silica Gel 60 F254 Plates in the Standardized Thin-Layer Chromatographic Technique for Lichen Products. *Chromatography*, 97, 107-108.
- Dell'Aglio, D. M., Perino, L. J., Kazzi, Z., Abramson, J., Schwartz, M. D., and Morgan, B. W. (2009). Acute Metformin Overdose: Examining Serum pH, Lactate

- Level, and Metformin Concentrations in Survivors versus None Survivors: A Systematic Review of the Literature. *Annals of Emergency Medicine*, 54(6), 818-823.
- Dent, M., Uzelac, V. D., Penić, M., Brnčić, M., Bosiljkov, T., and Levaj, B. (2013).
 The Effect of Extraction Solvents, Temperature and Time on the Composition and Mass Fraction of Polyphenols in Dalmatian Wild Sage (Salvia officinalis L.) Extracts. Food Technology and Biotechnology, 51(1), 84–91.
- de Villiers, A., Kalili, K. M., Malan, M., and Roodman, J. (2010). Improving HPLC Separation of Polyphenols. *LCGC Europe*, 23(10), 466–478.
- Dhanani, T., Shah, S., Gajbhiye, N. A., and Kumar, S. (2013). Effect of Extraction Methods on Yield, Phytochemical Constituents and Antioxidant Activity of Withania somnifera. Arabian Journal of Chemistry, 6, 1-7.
- El Nahhal, I. M., Zourab, S. M., Kodeh, F. S., and Qudaih, A. I. (2012). Thin Film Optical BTB pH Sensors using Sol–Gel Method in Presence of Surfactants. *International Nano Letters*, 2(16), 1-9.
- Ellis, B. E., and Towers, G. H. N. (1970). Biogenesis of Rosmarinic Acid in Mentha. *Biochemical Journal*, 118(2), 291-297.
- Englert, J., and Harnischfeger, G. (1992). Diuretic Action of Aqueous Orthosiphun Extract in Rats. *Planta medica*, 58(3), 237-238.
- Ermer, J. (1998). The Use of Hyphenated LC-MS Technique for Characterisation of Impurity Profiles during Drug Development. *Journal of Pharmaceutical and Biomedical Analysis*, 18(4), 707–714.
- Eurich, D. T., McAlister, F. A., and Blackburn, D. F. (2007). Benefits and Harms of Anti-Diabetic Agents in Patients with Diabetics and Heart Failure. *Systematic Review*, 335(7618), 497–499.
- Ferreira, L. G., Celotto, A. C., Capellini, V. K., Albuquerque, A. A. S., de Nadai, T., de Carvalho, M. T. M., and Evora, P. R. B. (2013). Does Rosmarinic Acid Underestimate as an Experimental Cardiovascular Drug? *Acta Cirurgica Brasileira*, 28, 83-87.
- Fetterolf, D. M. (2009). Column Chromatography. *Journal of Validation Technology*, 15(3), 43-48.
- Firenzuoli, F., and Gori, L. (2007). Herbal Medicine Today: Clinical and Research Issues. *Evidence-Based Complementary and Alternative Medicine*, 4(1), 37-40.

- GBIF backbone taxonomy. (2016). Retrieved on October 31, 2016, from http://www.gbif.org/species/3891871
- Glowniak, K., Zgórka, G., and Kozyra, M. (1996). Solid-Phase Extraction and Reversed-Phase High-Performance Liquid Chromatography of Free Phenolic Acids in Some Echinacea Species. *Journal of Chromatography A*, 730(1996), 25-29.
- Gray, G. M. (1975). Carbohydrate Digestion and Absorption. Role of the Small Intestine. *The New England Journal of Medicine*, 292(23), 1225-1230.
- Guthrie, R.A., and Guthrie, D.W. (2004). Pathophysiology of Diabetes Mellitus. *Critical Care Nursing Quarterly*, 27(2), 113-125.
- Haffner, S. M., Kahn, S. E., Zinman, B., Holman, R. R., Viberti, G. F., Herman, W.
 H., Lachin, J. M., Kravitz, B. G., and Heise, M. A. (2007). Greater Reductions in C-Reactive Protein with Rosiglitazone than with Glyburide or Metformin Despite Greater Weight Gain. *Diabetologia*, 50(1), 502.
- Hamann, O. (1988). The Joint IUCN-WWF Plants Conservation Program and its Interest in Medicinal Plants. In Olayiwola, A., Vernon, H, and High, S. (Eds.). Conservation of medicinal plant (pp. 3-11). New York: Cambridge University Press.
- Handa, S. S., Khanuja, S. P. S., Longo, G., and Rakesh, D. D. (2008). Extraction Technologies for Medicinal and Aromatic Plants. Trieste: United Nations Industrial Development Organization and the International Centre for Science and High Technology.
- Hegyi, G., Kardos, J., Kovács, M., Csizmadia, A. M., Nyitray, L., Pál, G., Radnai, L.,Reményi, A., and Venekei, I. (2013). *Introduction to Practical Biochemistry*.Budapest, Hungary: Eötvös Loránd University.
- Hernández-Hernández, E., Ponce-Alquicira, E., Jaramillo-Flores, M. E., and Guerrero Legarreta, I. (2009). Antioxidant Effect Rosemary (*Rosmarinus officinalis* L.) and Oregano (*Origanum vulgare* L.) Extracts on TBARS and Colour of Model Raw Pork Batters. *Meat Science*, 81, 410-417.
- Herrero, M., Plaza, M., Cifuentes, A., and Ibáñez, E. (2010). Green Processes for the Extraction of Bioactives from Rosemary: Chemical and Functional Characterization via Ultra-Performance Liquid Chromatography-Tandem Mass Spectrometry and *In-vitro* Assays. *Journal of Chromatography A*, 1217(16), 2512-2520.

- Himani, B., Seemab, B., Bholec, N., Mayanka, Y., Vinodd, S., and Mamta, S. (2013).
 Misai Kuching: A Glimpse of Maestro. *International Journal of Pharmaceutical Sciences Review and Research*, 22(2), 55-59.
- Hodgson, E. (2010). A Textbook of Modern Toxicology. Hoboken, NJ: John Wiley & Sons
- Hossain, M. A., and Ismail, Z. (2005). New Lupene-Type Triterpene from the Leaves of *Orthosiphon stamineus*. *Indian Journal of Chemistry*, 44(B), 436-437.
- Hossain, M. A., Ismail, Z., Rahman, A., and Kang, S. C. (2008). Chemical Composition and Anti-Fungal Properties of the Essential Oils and Crude Extracts of *Orthosiphon stamineus* Benth. *Industrial Crops and Products*, 27(3), 328-334.
- Hossain, M. A., and Ismail, Z. (2009). High Performance Thin Layer Chromatographic Determination of Caffeic Acid and Rosmarinic Acid from the Leaves of *Orthosiphon stamineus*. *Indonesian Journal of Chemistry*, 9 (1), 137 141.
- Hossain, M., Rai, D., Brunton, N., Martin-Diana, A. B., and Barry-Ryan, C. (2010).Characterization of Phenolics Composition in Lamiaceae Spices by LC-ESI-MS/MS. *Journal of Agricultural and Food Chemistry*, 58(19), 10576-10581.
- Houghton, P. J., and Raman, A. (1998). *Laboratory Handbook for the Fractionation of Natural Extracts*. London: Chapman and Hall.
- Hsuan, K. (1986). *Order dan Famili Tumbuhan Berbiji di Tanah Melayu*. Kuala Lumpur: Dewan Bahasa dan Pustaka.
- Hyun, T. K., Eom, S. H., and Kim, J. S. (2014). Molecular Docking Studies for Discovery of Plant-Derived α-glucosidase Inhibitors. *Plant Omics Journal*, 7(3),166-170.
- Ignat, I., Volf, I., and Popa, V. I. (2011). A Critical Review of Methods for Characterisation of Polyphenolic Compounds in Fruits and Vegetables. *Food Chemistry*, 126(4), 1821–1835.
- Indubala J, and Ng, L. T. (2000). *The Green Pharmacy of Malaysia*. Kuala Lumpur: Vinpress Sdn. Bhd.
- Ito, H., Miyazaki, T., Ono, M., and Sakurai, H. (1998). Antiallergic Activities of Rabdosiin and its Related Compounds: Chemical and Biochemical Evaluations. *Bioorganic & Medicinal Chemistry*, 6(7), 1051–1056.
- Jacotet-Navarro, M., Rombaut, N., Fabiano-Tixier, A. S., Danguien, M., Bily, A., and Chemat, F. (2015). Ultrasound Versus Microwave as Green Processes for

- Extraction of Rosmarinic, Carnosic and Ursolic Acids from Rosemary. *Ultrasonics Sonochemistry*, 27, 102-109.
- Jayanthy, G., and Subramanian, S. (2015). RA Abrogates Hepatic Gluconeogenesis and Insulin Resistance by Enhancing IRS-1 and AMPK Signalling in Experimental Type 2 Diabetes. *RSC Adv.*, 5, 44053–44067.
- Jiang, Z., Lu, Z., and Bing, Y. (2004). Strategies and Methods for Purifying Organic Compounds and Combinatorial Libraries. In Bing, Y. (Ed.). Analysis and Purification Methods in Combinatorial Chemistry (pp. 267-268). Hoboken: John Wiley & Sons, Inc.
- Jork, H., Funk, W., Fishcer, W., and Wimmer, H. (1990). *Thin-Layer Chromatography Reagents and Detection Methods-Physical and Chemical Detection Methods: Fundamentals, Reagents I.* New York: VCH Publishers.
- Jung, M., Park, M., Chul, H. L., Kang, Y., Seok-Kang, E., Ki-Kim, S. (2006).
 Antidiabetic Agents from Medicinal Plants. *Current Medicinal Chemistry*, 13, 1-16.
- Justesen, U. (2000). Negative Atmospheric Pressure Chemical Ionization Low-Energy Collision Activation Mass Spectrometry for The Characterization of Flavonoids in Extracts of Fresh Herbs. *Journal of Chromatography A*, 902(2), 369-379.
- Kaufmann, B., and Christen, P. (2002). Recent Extraction Techniques for Natural Products: Microwave-Assisted Extraction and Pressurized Solvent Extraction. *Phytochemical Analysis*, 13, 105-113.
- Khairana, H., Juriyati, J., Jamia, A. J., Ibrahim, J., and Azean, A. G. (2000).
 Consumption Plant Medicines to Treat Disease Diabetes in Malay Traditional Medicine. *Proceedings of the Seminar on Medicinal and Aromatic Plants: Towards Bridging Science and Herbal Industry*. 12-13 September. Kuala Lumpur, Malaysia: Forest Research Institute, 154-157.
- Kim, Y. M., Jeong, Y. K., Wang, M. H., Lee, W. Y., and Rhee, H. I. (2005). Inhibitory Effects of Pine Bark Extract on Alpha Glucosidase Activity and Postprandial Hyperglycemia. *Nutrition*, *21*, 756-761.
- Koay, Y. C., and Amir, F. A. (2012). Survey of the Chemical Constituents and Biological Activities of *Orthosiphon stamineus*. Science International, 24(2), 133-138.

- Kubínová, R., Pořízková, R., Navrátilová, A., Farsa, O., Hanáková, Z., Bačinská, A., Čížek, A., and Valentová, M. (2013). Antimicrobial and Enzyme Inhibitory Activities of the Constituents of *Plectranthus madagascariensis* (Pers.) Benth. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 29(5), 749-752.
- Kumanan, R., Manimaran, S., Saleemulla, K., Dhanabal, S. P., and Nanjan, M. J. (2010). Screening of Bark of *Cinnamomum tamala* (Lauraceae) by using α-amylase Inhibition Assay for Anti-diabetic Activity. *International Journal of Pharmaceutical and Biomedical Research*, 1(2), 69-72.
- Kumar, V., Khanna, A. K., Khan, M. M., Singh, R., Singh, S., Chander, R., and Singh,
 R. K. (2009). Hypoglycemic, Lipid Lowering and Antioxidant Activities in
 Root Extract of Anthocephalus Indicus in Alloxan Induced Diabetic Rats.
 Indian Journal of Clinical Biochemistry, 24(1), 65-69.
- Kumar, P. M., Sasmal, D., and Mazumder, P. M. (2010). The Antihyperglycemic Effect of Aerial Parts of *Salvia Splendens* (Scarlet Sage) in Streptozotocin-Induced Diabetic-Rat. *Pharmacognosy Research*, 2(3), 190–194.
- Kumar, S., and Pandey, A. K. (2013). Chemistry and Biological Activities of Flavonoids: An Overview. *The Scientific World Journal*, 2013(2013), 1-16.
- Kupiec, T. (2004). Quality-Control Analytical Methods: High-Performance Liquid Chromatography. *International Journal of Pharmaceutical Compounding*, 8(3), 223-227.
- Kwon, Y. I, Apostolidis, E., and Shetty, K. (2008). Inhibitory Potential of Wine and Tea against A-Amylase And A-Glucosidase for Management of Hyperglycemia Linked to Type 2 Diabetes. *Journal of Food Biochemistry*, 32(2008), 15–31.
- Lau, C. H., Chua, L. S., Lee, C. W., and Aziz, R. (2015). Fractionation of Rosmarinic Acid from Extract of *Orthosiphon stamineus* by Solid Phase Extraction. *Journal of Engineering Science and Technology*, 1(1), 104-112.
- Lau, C. H., Chua, L. S., Lee, C. T, and Aziza, R. (2014). Optimization and Kinetic Modeling of Rosmarinic Acid Extraction from *Orthosiphon stamineus*. *Current Bioactive Compounds*, 10, 271-285.
- Li, A. F., Sun, A. L., and Liu, R.M. (2005). Preparative Isolation and Purification of Costunolide and Dehydrocostuslactone from *Aucklandia Lappa Decne* by High-Speed Counter Current Chromatography. *Journal of Chromatography A*, 1076(2005), 193–197.

- Liu, H. W. (2011). Extraction and Isolation of Compounds from Herbal Medicines. In Liu, W. J. H. (Ed). *Traditional Herbal Medicine Research Methods* (pp.107-123). Hoboken: John Wiley and Sons, Inc.
- Lovegrove, A., Edwards, C. H., De Noni, I., Patel, H., El, S. N., Grassby, T., Zielke,
 C., Ulmius, M., Nilsson, L., Butterworth, P. J., Ellis, P. R., and Shewry, P. R.
 (2017). Role of Polysaccharides in Food, Digestion, and Health. *Critical Reviews in Food Science and Nutrition*, 57(2), 237–253.
- Lyckander, I., and Maltreud, K. (1996). Liphophilic Flavonoids from *Orthosiphon* spicatus Prevent Oxidative Inactivation of 15-Lypooxygenase. *Prostaglandins* Leukotrienes and Essential Fatty Acids, 54(4), 239-246.
- Malbaša, R. V., Lončar, E. S., and Kolarov, L. A. (2004). TLC Analysis of Some Phenolic Compounds in Kombucha Beverage. *Acta periodica technologica*, 35, 199-205.
- Manohar, V., Talpur, N. A., Echard, B. W., Lieberman, S, and Preuss, H. G. (2002). Effects of a Water-Soluble Extract of Maitake Mushroom on Circulating Glucose/Insulin Concentrations in KK Mice. *Diabetes, Obesity and Metabolism*, 4(1), 43–48.
- Masharani, U., and German, M. S. (2011). Pancreatic Hormones and Diabetes Mellitus. In Shoback, D. G., and Gardner, D. (eds.), *Greenspan's Basic & Clinical Endocrinology 9e*, New York, NY: McGraw-Hill, pp.605-615.
- Matkowski, A. (2008). Antioxidant Activity of Extracts and Different Solvent Fractions of *Glechoma hederacea* L. and *Orthosiphon stamineus* (Benth.) Kudo. *Advances in Clinical and Experimental Medicine*, 17(6), 615-624.
- Matsubara, T., Bohgaki, T., Watarai, M., Suzuki, H., Ohashi, K., and Shibuya, H. (1999). Antihypertensive Actions of Methylripariochromene A from *Orthosiphon aristatus*, An Indonesian Traditional Medicinal Plant. *Biological and Pharmaceutical Bulletin*, 22(10), 1083-1088.
- McCue, P. P., and Shetty, K. (2004). Inhibitory effects of rosmarinic acid extracts on porcine pancreatic amylase in vitro. *Asia Pacific Journal of Clinical Nutrition*, 13(1), 101-106.
- McDougall, G. J, and Stewart, D. (2005). The Inhibitory Effects of Berry Polyphenols on Digestive Enzymes. *BioFactors*, 23(4), 189-195.

- Megh, R. B., Nilubon, J. A., Gao, H., and Jun, K. (2008). α-Glucosidase and α-amylase Inhibitory Activities of Nepalese Medicinal Herb Pakhanbhed (*Bergenia ciliata*, Haw.). *Food Chemistry*, 106(1), 247-252.
- Mohamed, E. A. H., Mohamed, A. J., Asmawi, M. Z., Sadikun, A., Ebrika, O. S., and Yam M. F. (2011). Antihyperglycemic Effect of *Orthosiphon stamineus* Benth Leaves Extract and its Bioassay Guided Fractions. *Molecules*, 16, 3787-3801.
- Mohamed, E. A. H., Siddiqui, M. J. A., Ang, L. F., Sadikun, A., Chan, S. H., Tan, S. C., Asmawi, M. Z., and Yam, M. F. (2012). Potent α-glucosidase and α-amylase Inhibitory Activities of Standardized 50% Ethanolic Extracts and Sinensetin from *Orthosiphon stamineus* Benth as Anti-diabetic Mechanism.
 BMC Complementary and Alternative Medicine, 12, 1-7.
- Mohammed, S. A., Yaqub, A. G., Sanda1, K. A., Nicholas, A. O., Arastus, W., Muhammad, M., and Abdullahi, S. (2013). Review on Diabetes, Synthetic Drugs and Glycemic Effects of Medicinal Plants. *Journal of Medicinal Plants Research*, 7(36), 2628-2637.
- Mogale, A. M., Lebelo, L. S., Thovhogi, N., de Freitas, A. N., and Shai, L. J. α-amylase and α-glucosidase Inhibitory Effects of *Sclerocarya Birrea* [(A. Rich.) Hochst.] Subspecies Caffra (Sond) Kokwaro (Anacardiaceae) Stem-Bark Extracts. (2011). *African Journal of Biotechnology*, 10(66), 15033-15039.
- Muhit, M. A., Tareq, S. M., Apu, A. S., Basak, D., and Islam, M. S. (2010). Isolation and Identification of Compounds from the Leaf Extract of *Dillenia indica* Linn. *Bangladesh Pharmaceutical Journal*, 13(1), 49-53.
- Nascimento, E. M., Rodrigues, F. F., Campos, A. R., and Costa, J. G. (2009). Phytochemical Prospection, Toxicity and Antimicrobial Activity of *Mentha Arvensis* (Labiatae) from Northeast of Brazil. *Journal of Young Pharmacists*, 1(3), 210–212.
- Neergheen, V. S., Soobrattee, M. A., Bahorun, T., Aruoma, O. I. (2006). Characterization of the Phenolic Constituents in Mauritian Endemic Plants as Determinants of their Antioxidant Activities *in vitro*. *Journal of Plant Physiology*, 163(8), 787-799.
- Nguyen, M. T. T., Awale, S., Tezuka, Y., Chien-Hsiung, C., and Kadota, S. (2004). Staminane- and Isopimarane-Type Diterpenes from *Orthosiphon Stamineus* of Taiwan and their Nitric Oxide Inhibitory Activity. *Journal of Natural Products*, 67(4), 654-658.

- Ode, O. J., Asuzu, I. U., and Ajayi, I. E. (2011). Bioassay-Guided Fractionation of the Crude Methanol Extract of *Cassia Singueana* Leaves. *Journal of Advanced Scientific Research*, 2(4), 81-86.
- Ohashi, K., Bohgaki, T., Matsubara, T., and Shibuya, H. (2000). Indonesian Medicinal Plants XXIII. Chemical Structured of Two New Migrated Pimarane-Type Diterpenes, Neoorthosiphols A and B, and Suppressive Effects on Rat Thoracic Aorta of Chemical Constituents Isolated from the Leaves of *Orthosiphon aristatus* (Lamiaceae). *Chemical and Pharmaceutical Bulletin*, 48(3), 433-435.
- Ogunwande, I. A., Matsui, T., Fujise, T., and Matsumoto, K. (2007). α-glucosidase Inhibitory Profile of Nigerian Medicinal Plants in Immobilized Assay System. *Food Science and Technology Research*, 13(2), 169-172.
- Olah, N. K., Radu, L., MogoÅŸan, C., Hanganu, D., and Gocan, S. (2003). Phytochemical and Pharmacological Studies on *Orthosiphon stamineus* Benth. (Lamiaceae) Hydroalcoholic Extracts. *Journal of Pharmaceutical and Biomedical Analysis*, 33(1), 117-123.
- Ondrejovič, M., Benkovičová, H., Šilhár, S. (2009). Optimization of Rosmarinic Acid Extraction from Lemon Balm (*Melissa Officinalis*). *Nova Biotechnologica*, 9(2), 175-182.
- Pare, J. J. R., Belanger, J. M. R., and Stafford, S. S. (1994). Microwave-Assisted Process (MapTM): A New Tool for the Analytical Laboratory. *Trends in Analytical Chemistry*, 13(4), 176–184.
- Parris, N. A. (1984). Instrumental Liquid Chromatography: A Practical Manual on High-Performance Liquid Chromatographic Methods. *Journal of Chromatography Library*, 27, 135-174.
- Pitt, J. J. (2009). Principles and applications of Liquid Chromatography-Mass Spectrometry in Clinical Biochemistry. *Clinical Biochemistry Review*, 30(1), 19-34.
- Prabhu, K., Karar, P. K., Hemalatha, S., and Ponnudurai, P. (2011). A Preliminary Chromatographic Detection of Phenolic Compounds from Ethanolic Stem Extracts of *Viburnum* Linn. species by TLC and PC. *Der Pharmacia Sinica*, 2(3), 74-80.
- Ramirez-Coronel, M. A., Marnet, N., Kolli, V. S., Roussos, S., Guyot, S., Augur, C. (2004). Characterization and Estimation of Proanthocyanidins and Other Phenolics in Coffee Pulp (*Coffea arabica*) by Thiolysis-High-Performance

- Liquid Chromatography. *Journal of Agricultural and Food Chemistry*, 52(5), 1344-1349.
- Rao, M. K., Bethala, K., Sisinthy, S.P., and Rajeswari, K. S. (2014). Antidiabetic Activity of *Orthosiphon stamineus* Benth Roots in Streptozotocin Induced Type 2 Diabetic Rats. *Asian Journal of Pharmaceutical and Clinical Research*, 7(1), 149-153.
- Reichardt, C., and Welton, T. (2011). Solvents and Solvent Effects in Organic Chemistry (4th ed.). Weinheim: Wiley-VCH.
- Rendell, M. (2000). Editorial: Dietary Treatment of Diabetes Mellitus. *The New England Journal of Medicine*, 342, 1440-1441.
- Ridgway, K., Lalljie, S. P. D., and Smith, R. M. (2008). Microextraction Methods in Food Analysis. In Otles, S. (Ed.). *Handbook of Food Analysis Instruments* (pp.7-20). Boca Raton: Taylor and Francis.
- Robbins, R. J. (2003). Phenolic Acids in Foods: An Overview of Analytical Methodology. *Journal of Agricultural and Food Chemistry*, 51(10), 2866–2887.
- Runtuwene, J., Cheng, K. C., Asakawa, A., Amitani, H., Amitani, M., Morinaga, A., and Inui, A. (2016). Rosmarinic Acid Ameliorates Hyperglycemia and Insulin Sensitivity in Diabetic Rats, Potentially by Modulating the Expression of PEPCK and GLUT4. *Drug Design, Development and Therapy*, 10, 2193–2202.
- Saidan, N. H., Hamil, M. S. R., Memon, A. H., Abdelbari, M. M, Hamdan, M. R., Mohd, K. S., Majid, A. M. S. A., and Ismail, Z. (2015a). Selected Metabolites Profiling of *Orthosiphon Stamineus* Benth Leaves Extracts Combined with Chemometrics Analysis and Correlation with Biological Activities. *BMC Complementary and Alternative Medicine*, 15(350), 1-12.
- Saidan, N. H., Aisha, A. F. A., Hamil, M. S. R., Majid, A. M. S. A., and Ismail, Z. (2015b). A Novel Reverse Phase High-Performance Liquid Chromatography Method for Standardization of *Orthosiphon stamineus* Leaf Extracts. Pharmacognosy *Research*, 7(1), 23–31.
- Sarah, W., Gojka, R., Anders, G., Richard, S., and Hilary, K. (2004). Global Prevalence of Diabetes. *Diabetes Care*. 27(5), 1047-1053.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M., and Latha, L. Y. (2011). Extraction, Isolation and Characterization of Bioactive Compounds from

- Plants' Extracts. *African Journal of Traditional, Complementary and Alternative Medicines*, 8(1), 1-10.
- Scarpati, M. L., and Oriente, G. (1958). Isolamento E Costituzione Dell'acido Rosmarinico (*Dal Rosmarinus* off.). *Rice Science*, 28, 2329–2333.
- Shahidi, F., and Wanasundara, P. K. (1992). Phenolic Antioxidants. *Critical Reviews* in Food Science and Nutrition, 32, 67-103.
- Shai, L. J., Masoko, P., Mokgotho, M. P., Magano, S. R., Mogale, A. M., Boaduo, N., and Eloff, J. N. (2010). Yeast Alpha Glucosidase Inhibitory and Antioxidant Activities of Six Medicinal Plants Collected in Phalaborwa, South Africa. South African Journal of Botany, 76(3), 465-470.
- Sharma, R. R. (2012). Enzyme Inhibition and Bioapplications. In Sharma, R. R. (Ed). *Enzyme inhibition: mechanisms and scope* (pp.4-35). Rijeka: InTech.
- Shekarchi, M., Hajimehdipoor, H., Saeidnia, S., Gohari, A.R., and Hamedani, M.P. (2012). Comparative Study of Rosmarinic Acid Content in Some Plants of Labiatae Family. *Pharmacognosy Magazine*, 8(29), 37-41.
- Shen, D., Pan, M. H., Wu, Q. L., Park, C. H., Juliani, H. R., Ho, C. T., and Simon, J. E. (2010). LC-MS Method for the Simultaneous Quantitation of the Anti-Inflammatory Constituents in Oregano (*Origanum Species*). *Journal of Agricultural and Food Chemistry*, 58(12), 7119-7125.
- Shobana, S., Sreerama, Y. N., and Malleshi, N. G. (2009). Composition and Enzyme Inhibitory Properties of Finger Millet (*Eleusine coracana* L.) Seed Coat Phenolics: Mode of Inhibition of α-Glucosidase and Pancreatic Amylase. *Food Chemistry*, 115, 1268-1273.
- Siddiqui, M. R., AlOthman, Z. A., and Rahman, N. (2013). Analytical Techniques in Pharmaceutical Analysis: A Review. *Arabian Journal of Chemistry*, 1, 1-13.
- Silva, E. M., Rogez, H. and Larondelle, Y. 2007. Optimization of Extraction of Phenolics from Inga Edulis Leaves using Response Surface Methodology. Separation and Purification Technology, 55(3), 381-387.
- Simpson, N. J. K. (2000). *Solid Phase Extraction Principles, Techniques, and Applications*. New York: Marcel Dekker.
- Snyder, L. R., and Kirkland, J. J. (1979). Introduction to Modern Liquid Chromatography (2nd ed.). New York: John Wiley and Sons Inc.
- Soxhlet, F. (1879). Die Gewichtsanalytische Bestimmung des Milchfettes. *Dingler's Polytechnisches Journal*, 232, 461–465.

- Stalikas, C. D. (2007). Review: Extraction, Separation, and Detection Methods for Phenolic Acids and Flavonoids. *Journal of Separation Science*, 30(18), 3268–3295.
- Still, W. C., Kahn, M., and Mitra, A. (1978). Rapid Chromatography Technique for Preparative Separations with Moderate Resolution. *Journal of Organic Chemistry*, 43(14), 2923-2925.
- Strelow, J., Dewe, W., Iversen, P. W., Brooks, H. B., Radding, J. A., McGee, J, and Weidner, J. (2012). Mechanism of Action Assays for Enzymes. In Sittampalam, G. S., Coussens, N. P., and Brimacombe, K. (Eds.). Assay Guidance Manual. (pp. 1-27). Bethesda: Eli Lilly & Company and the National Center for Advancing Translational Sciences.
- Subramanian, R., Asmawi, A. Z., and Sadikun, A. (2008). *In Vitro* Alpha-Glucosidase and Alpha-Amylase Enzyme Inhibitory Effects of *Andrographis paniculata* Extract and Andrographolide. *The Journal of Polish Biochemical Society*, 55, 391-398.
- Sumaryono, W., Proksch, P., Wray, V., Witte, L., and Hartmann, T. (1991).

 Qualitative and Quantitative Analysis of the Phenolic Constituents from *Orthosiphon aristatus. Planta Medica*, 57(2), 176–180.
- Sundarammal, S., Thirugnanasampandan, and Selvi, M. T. (2012). Chemical Composition Analysis and Antioxidant Activity Evaluation of Essential Oil from *Orthosiphon thymiflorus* (Roth.) Sleesen. *Asian Pacific Journal of Tropical Biomedicine*, 2(1), 112-115.
- Takeda, Y., Matsumoto, T., Terao, H., Shingu, T., Futatsuishi, Y., Nohara, T., and Kajimoto, T. (1993). Orthosiphol D and E, Minor Diterpenes from Orthosiphon stamineus. Phytochemistry, 33(2), 411-415.
- Tao, L., Wang, S., Zhao, Y., Sheng, X., Wang, A., Zheng, S., and Lu, Y. (2014).
 Phenolcarboxylic Acids from Medicinal Herbs Exert Anticancer Effects
 through Disruption of COX-2 Activity. *Phytomedicine*, 21(11), 1473-1482.
- Tarling, C. A., Woods, K., Zhang, R., Brastianos, H. C., Brayer, G. D., Andersen, R. J., and Withers, S. G. (2008). The Search for Novel Human Pancreatic Alpha-Amylase Inhibitors: High-Throughput Screening of Terrestrial and Marine Natural Product Extracts. *Chembiochem*, 9(3), 433-438.

- Tekel, J., and Hatrík, Š. (1996). Pesticide Residue Analyses in Plant Material by Chromatographic Methods: Clean-Up Procedures and Selective Detectors. *Journal of Chromatography A*, 754, 397–410.
- Tezuka, Y., Stampoulis, P., Banskota, A. H., Awale, S., Tran, K. Q., Saiki, I., and Kadota, S. (2000). Constituents of the Vietnamese Medicinal Plant *Orthosiphon stamineus*. *Chemical and Pharmaceutical Bulletin*, 48, 1711–1719.
- Thurman, E. M., and Mills, M. S. (1998). *Solid-phase Extraction Principles and Practice*. New York: John Wiley.
- Truong, D. M, Bich Thu, N. T., and Nghi, L. T. (2010). Accumulation and Variation of Rosmarinic Acid Content in *Orthosiphon stamineus Benth*. based on Phenological Stages. *Journal of Biology*, 32(3), 65-71.
- Trusheva, B., Trunkova, D., and Bankova, V. (2007). Different Extraction Methods of Biologically Active Components from Propolis: A Preliminary Study. *Chemistry Central Journal*, 1, 13.
- Tsuda, Y. (2004). *Isolation of Natural Products*. Tokyo: Japan Analytical Industry Limited Company.
- Venkatachalam, K., Gunasekaran, S., Jesudoss, V. A., and Namasivayam, N. (2013).
 The Effect of Rosmarinic Acid on 1,2-Dimethylhydrazine Induced Colon Carcinogenesis. *Experimental and Toxicologic Pathology*, 65, 409-418.
- Vogeser, M., and Parhofer, K. G. (2007). Liquid Chromatography Tandem-Mass Spectrometry (LC-Ms/Ms)-Technique and Applications in Endocrinology. *Experimental and Clinical Endocrinology Diabetes*, 115(9), 559 570.
- Wang, H. F., Provan, G. J., and Helliwell, K. (2004). Determination of Rosmarinic Acid and Caffeic Acid in Aromatic Herbs by HPLC. *Food Chemistry*, 87, 307–311.
- Wei, L., Mouming, Z., Bao, Y., Guanglin, S., and Guohua, R. (2008). Identification of Bioactive Compounds in *Phyllenthus emblica* L. Fruit and their Free Radical Scavenging Activities. *Food Chemistry*, 114(2009), 499–504.
- WHO. (2017). WHO Guidelines on Good Herbal Processing Practices (GHPP) for Herbal Medicines. World Health Organization: Geneva.
- Wiart, C. (2000). *Medicinal Plant of Southeast Asia*. Selangor: Pelanduk Publication Sdn. Bhd.

- Wilkin, T. (2001). The Accelerator Hypothesis: Weight Gain as the Missing Link between Type I and Type Ii Diabetes. *Diabetologia*, 44(7), 914-922.
- Williamson, K. L., and Masters, K. M. (2016). *Macroscale and Microscale Organic Experiments*. Boston: Cengage Learning.
- Wright, E. M. I. (1998). Glucose Galactose Malabsorption. *American Journal of Physiology*, 275, 879-882.
- World Health Organization (2016). *Global Report on Diabetes*. Retrieved on October 31, 2016 from http://www.who.int/diabetes/action_online/basics/en/.
- Worthley, L. I. G. (2003). *The Australian Short Course on Intensive Care Medicine* 2004 Handbook. Underdale: Gillingham Printers.
- Xiao, J., Kai, G., Yamamoto, K., and Chen, X. (2013). Advance in Dietary Polyphenols as α-Glucosidases Inhibitors: A Review on Structure-Activity Relationship Aspect. *Critical Review in Food Science and Nutrition*, 53, 818–836.
- Xu, R. S., Ye, Y., and Zhao, W. M. (2011). *Introduction to Naturals Products Chemistry*. Boca Raton: Taylor and Francis Group.
- Yam, M. F., Asmawi, M. Z., and Basir, R. (2008). An Investigation of the Anti-Inflammatory and Analgesic Effects of *Orthosiphon stamineus* Leaf Extract. *Journal of Medicinal Food*, 11(2), 362-368.
- Yuan, H., Shan, L., Sun, Q., and Han, W. (2011). Total Synthesis of Rosmarinic Acid. *Acta Chimica Sinica*, 69(8), 945-948.
- Yuliana, N., Khatib, A., Regina, A. M., Ijzerman, A. P., Zakaria, F. R., Choi, Y. H., and Verpoorte, R. (2009). Adenosine A1 Receptor Binding Activity of Methoxy Flavonoids from *Orthosiphon stamineus*, *Planta Medica*, 75(2), 132-136.
- Zaharah, A. (2005). Misai Kucing (*Orthosiphon stamineus*). In Musa, Y., Ghawas, M.M., and P. Mansor (Eds.). *Penanaman Tumbuhan Ubatan and Beraroma* (pp. 14-20). Serdang: MARDI.
- Zahradníková, L., Schmidt, Š., Sékelyová, Z., and Sekretár, S. (2008). Fractionation and Identification of Some Phenolics Extracted from Evening Primrose Seed Meal. *Czech Journal of Food Science*, 26, 58–64.
- Zhang, Y., Seeram, N. P., Lee, R., Feng, L., Heber, D. (2008). Isolation and Identification of Strawberry Phenolics with Antioxidant and Human Cancer

- Cell Antiproliferative Properties. *Journal of Agricultural and Food Chemistry*, 56(3), 670-675.
- Zheng, W., and Wang, S. Y. (2001). Antioxidant Activity and Phenolic Compounds in Selected Herbs. *Journal of Agricultural and Food Chemistry*, 49(11), 5165–5170.
- Żwir-Ferenc, A., and Biziuk, M. (2006). Solid Phase Extraction Technique-Trends, Opportunities and Applications. *Polish Journal of Environmental Studies*, 15(5), 677-690.