

DEVELOPMENT AND APPLICATIONS OF ELECTROMEMBRANE  
EXTRACTION (EME) METHODS ACROSS HOLLOW POLYMER  
INCLUSION MEMBRANE (HPIM) FOR ANALYSIS OF  
DRUGS AND HERBICIDES

NOR AKMA BINTI MAMAT

A thesis submitted in fulfilment of the  
requirements for the award of the degree of  
Doctor of Philosophy (Chemistry)

Faculty of Science  
Universiti Teknologi Malaysia

MAY 2017

*Specially dedicated to my beloved parents and families for all support and  
encouragement in completing this study*

## ACKNOWLEDGEMENTS

Firstly, thanks to Allah for giving me strength to finish this study. Special thanks to my supportive supervisor, Dr. See Hong Heng, for his supervision, great information, kindness, help, guidance, invaluable suggestion and for being a constant source of encouragement.

Not forgetting to all my friends in analytical lab for their kind support and advice in completing this research. A million thanks also go to science officers and lab assistants of the Department of Chemistry, Faculty of Science, UTM for their technical assistance throughout the study. I also would like to express my deepest thank to the Ministry of Higher Education (MOHE) for MyPhD scholarship.

Last but not least, I would like to thank my parents and family for constant support, emotional understanding, encouragement that involved indirectly or directly throughout my study.

## ABSTRACT

Electromembrane extraction (EME) has become an effective method in the development of sample preparation technique. In this study, a novel microextraction method based on the EME and employed with hollow polymer inclusion membrane (HPIM) was developed in order to get better stability and reproducibility compared to the conventional EME. HPIM was prepared by dipping the glass capillary tubes into a solution of the desired proportions of cellulose triacetate (CTA), tris(2-ethylhexyl)phosphate (TEHP) and di-(2-ethylhexyl)phosphoric acid (D2EHPA) or Aliquat 336 in dichloromethane. Three basic drugs, namely amphetamine, methamphetamine and 3,4-methylenedioxy-*N*-methylamphetamine (MDMA) were selected as the target analytes to evaluate the extraction efficiency of the new approach. Parameters affecting the extraction efficiency, including the composition of HPIM, pH of sample, extraction voltage and extraction time were investigated in detail. Under the optimized conditions, enrichment factors in the range of 97-103 fold were obtained from 3 mL sample solution with a 10 min extraction time and an applied voltage of 300 V across the HPIM. A comparison was also made between the newly developed approach and the conventional EME as well as standard sample preparation methods (liquid-liquid extraction) used by the Toxicology Unit, Department of Chemistry, Malaysia. The applied voltage in EME is an important parameter for efficient extraction of the analyte, however, when dealing with extremely high voltage, instability occurs due to the formation of bubbles. This limitation has stimulated the development of the exhaustive simultaneous EME across HPIM with the aim of employing a bubbleless electrode for the determination of selected cationic and anionic pesticides present in the environmental water samples. Bubbleless electrode was prepared to solve the bubble formation problem during the extraction process. Cationic herbicides namely paraquat (PQ) and diquat (DQ) as well as anionic herbicides namely (4-chlorophenoxy)acetic acid (4-CPA) and 2-(2, 4-dichlorophenoxy)acetic acid (2,4-D) were selected as the model analytes to evaluate the extraction performance of this new approach. Under the optimized conditions, the enrichment factors in the range of 152–185-fold were obtained from 4 mL of river water sample with a 20 min extraction time and an applied voltage of 3000 V. The proposed method provided good linearity with the correlation coefficients ranging from 0.9982 to 0.9997 over a concentration range of 1–1000 ng/mL. The detection limits of the method for the herbicides were in the range of 0.3–0.4 ng/mL, with the relative standard deviations ranged between 4.8% and 8.5%. A comparison was also made between the newly developed method with that of conventional EME setup using normal electrode.

## ABSTRAK

Kaedah pengekstrakan elektromembran (EME) telah menjadi satu kaedah yang berkesan dalam perkembangan teknik penyediaan sampel. Dalam kajian ini, satu kaedah pengekstrakan mikro baru yang berlandaskan kepada EME dan menggunakan membran terkandung polimer berongga (HPIM) telah dibangunkan untuk memperoleh kestabilan dan kebolehulangan yang lebih baik berbanding dengan EME konvensional. HPIM telah disediakan dengan mencelup tiub kaca kapilari ke dalam larutan selulosa asetat (CTA), tris(2-etilheksil)fosfat (TEHP) dan asid di-(2-etilheksil)fosforik (D2EHPA) atau Aliquat 336 di dalam diklorometana pada perkadaran yang diinginkan. Tiga dadah bebas iaitu amfetamin, metamfetamin dan 3,4-metilenadioksi-*N*-metilamfetamin (MDMA) telah dipilih sebagai analit sasaran untuk menilai kecekapan pengekstrakan bagi pendekatan baharu. Parameter yang mempengaruhi kecekapan pengekstrakan, termasuk komposisi HPIM, pH sampel, voltan pengekstrakan dan masa pengekstrakan telah dikaji dengan mendalam. Di bawah keadaan optimum, faktor pengayaan dalam julat 97-103 kali telah diperolehi daripada 3 mL sampel larutan dengan 10 min masa pengekstrakan dan voltan 300 V dikenakan merentasi HPIM. Satu perbandingan telah dibuat antara pendekatan baharu yang dibangunkan dengan EME konvensional serta kaedah penyediaan sampel piawai (pengekstrakan cecair-cecair) yang digunakan oleh Unit Toksikologi, Jabatan Kimia, Malaysia. Penggunaan voltan dalam EME adalah satu parameter yang penting untuk pengekstrakan analit yang cekap, namun, apabila berurusan dengan voltan yang sangat tinggi, ketidakstabilan berlaku disebabkan oleh pembentukan buih. Pembatasan ini telah merangsang pembangunan EME selari merentasi HPIM yang lengkap dengan tujuan untuk menggunakan elektrod tanpa buih untuk penentuan pestisid kationik dan anionik terpilih yang hadir dalam sampel air persekitaran. Elektrod tanpa buih telah disediakan untuk menyelesaikan masalah pembentukan buih semasa proses pengekstrakan. Herbisid kationik iaitu parakuat (PQ) dan dikuat (DQ) serta herbisid anionik iaitu asid (4-klorofenoksi)asetik (4-CPA) dan asid 2-(2,4-diklorofenoksi)asetik (2,4-D) telah dipilih sebagai model analit untuk menilai prestasi pengekstrakan bagi pendekatan baharu ini. Di bawah keadaan optimum, faktor pengayaan dalam julat 152-185 kali telah diperolehi daripada 4 mL sampel air sungai dengan 20 min masa pengekstrakan dan menggunakan 3000 V. Kaedah yang dicadangkan telah memberikan kelinearan yang baik dengan pekali korelasi antara 0.9982 hingga 0.9997 dalam julat kepekatan 1-1000 ng/mL. Had pengesanan kaedah ini untuk herbisid adalah dalam julat 0.3-0.4 ng/mL, dengan sisihan piawai antara 4.8% dan 8.5%. Satu perbandingan telah dibuat antara kaedah baharu yang telah dibangunkan dengan EME konvensional yang menggunakan elektrod biasa.

## TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	<b>DECLARATION</b>	<b>ii</b>
	<b>DEDICATION</b>	<b>iii</b>
	<b>ACKNOWLEDGEMENT</b>	<b>iv</b>
	<b>ABSTRACT</b>	<b>v</b>
	<b>ABSTRAK</b>	<b>vi</b>
	<b>TABLE OF CONTENTS</b>	<b>vii</b>
	<b>LIST OF TABLE</b>	<b>xiii</b>
	<b>LIST OF FIGURES</b>	<b>xv</b>
	<b>LIST OF ABBREVIATIONS</b>	<b>xx</b>
	<b>LIST OF SYMBOLS</b>	<b>xxii</b>
	<b>LIST OF APPENDICES</b>	<b>xxiii</b>
<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
	1.1 Background	1
	1.2 Problem Statement	4
	1.3 Research Aim and Objectives	5
	1.4 Scopes of the Research	6
	1.5 Significance of Research	7
	1.6 Outline of the Thesis	7
<b>2</b>	<b>LITERATURE REVIEW</b>	<b>9</b>
	2.1 Sample Preparation	9
	2.1.1 Classical Sample Preparation Method	10
	2.1.2 Solid Phase Microextraction (SPME)	10

2.1.2.1	Stir Bar Sorptive Extraction (SBSE)	11
2.1.3	Liquid Phase Microextraction (LPME)	12
2.1.3.1	Single Drop Microextraction (SDME)	13
2.1.3.2	Hollow Fiber Liquid Phase Microextraction (HF-LPME)	14
2.2	Electromembrane Extraction (EME)	16
2.2.1	Parameters Influencing Electromigration of Analyte From Sample Solution Across into Acceptor Solution in EME	17
2.2.1.1	Supported Liquid Membrane (SLM)	
2.2.1.2	Effect of pH Sample and Acceptor Solution	18
2.2.1.3	Applied Voltage	18
2.2.1.4	Extraction Time	19
2.2.2	Limitation of EME	20
2.2.3	Alternative of EME	20
2.2.4	Applications of EME	21
2.3	Polymer Inclusion Membrane (PIM)	21
2.3.1	Base Polymer	24
2.3.2	Plasticizer	24
2.3.3	Carrier	25
2.3.4	Mechanism of Transportation across Polymer Inclusion Membrane (PIM)	25
2.3.5	PIM vs SLM	26
2.3.6	Applications of PIM	27
2.3.7	EME across a PIM	28
2.4	Paraquat (PQ) and Diquat (DQ) Herbicides	28
2.5	Amphetamine, Methamphetamine and MDMA	29
2.6	Chlorinated Phenoxyacetic Acid (CPAs) Herbicides	31
2.7	Capillary Electrophoresis	32
2.7.1	Principle of CE	32
2.7.2	Mode of CE	32

	2.7.2.1	Capillary Zone Electrophoresis (CZE)	34
	2.7.2.2	MEKC	34
	2.7.3	Detections of CE	34
	2.7.3.1	CE-C <sup>4</sup> D	35
	2.7.3.2	UV Absorption	35
			35
<b>3</b>		<b>ELECTROMEMBRANE EXTRACTION ACROSS SUPPORTED LIQUID MEMBRANE (SLM) OF DRUGS IN HUMAN PLASMA</b>	<b>36</b>
	3.1	Introduction	36
	3.2	Experimental	38
	3.2.1	Chemicals and Reagents	38
	3.2.2	Sample Preparation	39
	3.2.3	EME-SLM Procedure	39
	3.2.4	Optimization Study	40
	3.2.5	Validation of the Method	41
	3.2.6	CE-C <sup>4</sup> D Condition	41
	3.3	Results and Discussion	42
	3.3.1	Optimization of Supported Liquid Membrane (SLM)	42
	3.3.2	Optimization Sample pH	43
	3.3.3	Optimization of Applied Voltage	45
	3.3.4	Optimization of Extraction Time	46
	3.3.5	Method Validation	47
	3.3.6	Comparison of Proposed Method With Other Existing Method	51
	3.4	Conclusion	52
<b>4</b>		<b>DESIGN OF HOLLOW POLYMER INCLUSION MEMBRANE (HPIM) FOR ELECTROMEMBRANE EXTRACTION (EME)</b>	<b>53</b>
	4.1	Introduction	53



4.2	Experimental	54
4.2.1	Reagents and Chemicals	54
4.2.2	Preparation of Hollow Polymer Inclusion Membrane (HPIM)	55
4.2.3	Thickness Measurement	55
4.2.4	Scanning Electron Microscope (SEM)	55
4.2.5	Contact Angle Measurement	56
4.3	Results and Discussion	56
4.3.1	Preparation of Hollow Polymer Inclusion Membrane (HPIM)	56
4.3.2	Physical Characterizations	58
4.3.2.1	Scanning Electron Micrographs	58
4.3.2.2	Contact Angle	60
4.4	Conclusion	61
<b>5</b>	<b>ELECTROMEMBRANE EXTRACTION OF DRUGS IN HUMAN PLASMA ACROSS A HOLLOW POLYMER INCLUSION MEMBRANE</b>	<b>62</b>
5.1	Introduction	62
5.2	Experimental	64
5.2.1	Chemicals and Reagents	64
5.2.2	Sample Preparation	64
5.2.3	Extraction Procedure	66
5.2.3.1	EME-HPIM	65
5.2.3.2	EME-SLM	65
5.2.3.3	Liquid-Liquid Extraction (LLE)	67
5.2.4	CE-C <sup>4</sup> D Quantification	67
5.2.5	Optimization Studies	68
5.3	Results and Discussion	68
5.3.1	HPIM Composition	68
5.3.1.1	Optimization of Plasticizer	68
5.3.1.2	Optimization of Carrier	69

5.3.2	Extraction Condition	71
5.3.2.1	Sample pH	71
5.3.2.2	Applied Voltage	72
5.3.2.3	Extraction Time	73
5.3.3	Method Validation	74
5.3.4	Spiked Human Plasma	77
5.4	Conclusion	78
<b>6</b>	<b>SIMULTANEOUS ELECTROMEMBRANE EXTRACTION OF CATIONIC AND ANIONIC HERBICIDES ACROSS HOLLOW POLYMER INCLUSION MEMBRANES WITH A BUBBLELESS ELECTRODE</b>	<b>79</b>
6.1	Introduction	79
6.2	Experimental	82
6.2.1	Chemicals and Reagents	82
6.2.2	Sample Preparation	82
6.2.3	Preparation of HPIMs	83
6.2.4	Preparation of Bubbleless Electrode	83
6.2.5	Simultaneous EME-HPIM Procedure	84
6.2.6	CE-C <sup>4</sup> D Quantification	86
6.3	Results and Discussion	87
6.3.1	Preparation of Bubbleless Electrode	87
6.3.1.1	Optimization of Applied Pressure	87
6.3.1.2	Optimization of Combined Capillary for Bubbleless Electrode	
6.3.2	EME-HPIM with Bubbleless Electrode	88
6.3.2.1	HPIM Composition	89
6.3.2.2	Optimization of Sample pH	89
6.3.2.3	Optimization of Applied Voltage	92
6.3.2.4	Optimization of Extraction Time	93
6.3.3	Comparison of EME-HPIM with/without the Bubbleless Electrode	95

		xii
	6.3.4 Analysis of River Water	96
	6.4 Conclusion	99
		101
<b>7</b>	<b>CONCLUSIONS AND FUTURE DIRECTIONS</b>	<b>102</b>
	7.1 Conclusions	102
	7.2 Future Directions	103
	<b>REFERENCES</b>	<b>105</b>
	Appendices A-B	124-125

**LIST OF TABLES**

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	The applications of electromembrane extraction (EME)	23
2.2	Reported PIM studies using base polymer, plasticizer and carrier for certain applications	26
2.3	Applications of PIM	28
2.4	Reported sample preparation method of PQ and DQ	30
2.5	Reported sample preparation method of amphetamine, methamphetamine and MDMA	31
3.1	Molecular structures, dissociation constant and partition coefficient of drugs studied	38
3.2	Quantitative results of EME-SLM for drug abuse in human plasma	49
3.3	Relative recoveries (%) and reproducibilities for the drugs of abuse extracted from spiked plasma samples	50
3.4	Comparison of the proposed method with other published methods for the extraction and determination of amphetamine, methamphetamine and MDMA	51

4.1	Effect of number of dipping cycle to the thickness of membrane	57
4.2	Effect of mass of base polymer on membrane thickness	58
5.1	Linear range, regression data, limit of detection (LODs), limit of quantification (LOQs) and enrichment factors for drugs of abuse in spiked plasma samples	76
5.2	Relative recoveries (%) and reproducibilities for the drugs of abuse extracted from spiked plasma samples	77
6.1	Molecular structures, dissociation constant and partition coefficient of herbicides studied	81
6.2	Effect of the applied pressure on current profile during the synthesis of bubbleless electrode	87
6.3	Effect of the number of capillary used as working electrode on current profile	88
6.4	Linear range, regression data, limits of detection (LODs), limits of quantification (LOQs) and enrichment factors (EFs) for herbicides in spiked river water samples	98
6.5	Relative recoveries (%) and reproducibilities of the spiked cationic and anionic herbicides in river water	99

**LIST OF FIGURES**

<b>FIGURE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Extraction procedure in Stir Bar Sorptive Extraction (SBSE)	11
2.2	Derivative methods under liquid phase microextraction (LPME)	12
2.3	Schematic diagram of SDME	14
2.4	Schematic diagram of HF-LPME	15
2.5	Schematic of EME set up	17
2.6	Schematic diagram of EME across PIM set up	29
2.7	Schematic representation of electrophoresis	33
2.8	Representation of electroosmotic flow (EOF) in capillary	33
3.1	EME-SLM set up	40
3.2	Effect of organic solvent used in EME-SLM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20 $\mu$ L of 0.1 mM KCl; Sample pH: pH 7; Applied voltage: 200 V; Extraction time: 10 min; CE conditions were as described in the text	43

- 3.3 Effect of sample pH on EME-SLM. Extraction condition; sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20  $\mu$ L of 0.1 mM KCl; SLM: 25% TEHP, 25% D2EHPA in NPOE; Applied voltage: 200 V; Extraction time; 10 min. CE conditions were as described in the text. Each data points represent the mean value of five measurements 44
- 3.4 Effect of applied voltage on EME-SLM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20  $\mu$ L of 0.1 mM KCl; SLM: 25% TEHP, 25% D2EHPA in NPOE; Sample pH: no adjustment; Extraction time; 10 min. CE conditions were as described in the text. Each data points represent the mean value of five measurements 45
- 3.5 Effect of extraction time on EME-SLM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20  $\mu$ L of 0.1 mM KCl; SLM: 25% TEHP, 25% D2EHPA in NPOE; Sample pH: no adjustment Extraction voltage; 300 V. CE conditions were as described in the text. Each data points represent the mean value of five measurements 47
- 3.6 Electropherogram of acceptor solution after EME-SLM of spiked human plasma (with an added background of 0.1 mM KCl) containing drugs at 500 ng/ml. Extraction condition: Extraction condition; Acceptor solution: 20  $\mu$ L of 0.1 mM KCl; SLM: 25% TEHP, 25% D2EHPA in NPOE; Sample pH: no adjustment; Extraction voltage; 300 V; Extraction time: 10 min. CE conditions were as described in text 50

4.1	Scanning electron micrographs of HPIM: (a) overall structure; (b) cross-sectional image; (c) outer surface image of HPIM; (d) inner surface image of HPIM	59
4.2	The effect of (a) mass of D2EHPA (b) mass of Aliquat 336 on hydrophilic character of membrane	60
5.1	Electroemembrane extraction across hollow polymer inclusion membrane set up	66
5.2	Effect of amount of TEHP on EME-HPIM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Sceptor solution: 20 $\mu$ L of 0.1 mM KCl; HPIM: 60 mg CTA; Sample pH: pH 7; Applied voltage: 200 V; Extraction time; 10 min. CE conditions were as described in the text. Each data points represent the mean value of five measurements	69
5.3	Effect of amount of D2EHPA on EME-HPIM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20 $\mu$ L of 0.1 mM KCl; HPIM: 60 mg CTA, 15 mg TEHP; Sample pH: pH 7; Applied voltage: 200 V; Extraction time; 10 min. CE conditions were described in text. Each data points represent the mean value of five measurements	70
5.4	Effect of sample pH on EME-HPIM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20 $\mu$ L of 0.1 mM KCl; HPIM: 60 mg CTA, 15 mg TEHP and 40 mg D2EHPA; Applied voltage: 200 V; Extraction time; 10 min. CE conditions were described in text. Each data points represent the mean value of five measurements	71



- 5.5 Effect of applied voltage on EME-HPIM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20  $\mu$ L of 0.1 mM KCl; HPIM: 60 mg CTA, 15 mg TEHP and 40 mg D2EHPA; Sample pH: no adjustment; Extraction time; 10 min. CE conditions were described in text. Each data points represent the mean value of five measurements 72
- 5.6 Effect of extraction time on EME-HPIM. Extraction condition; Sample solution: 500 ng/mL drugs in 3 mL of 0.1 mM KCl; Acceptor solution: 20  $\mu$ L of 0.1 mM KCl; HPIM: 60 mg CTA, 15 mg TEHP and 40 mg D2EHPA; Sample pH: pH 7; Applied voltage: 300 V. CE conditions were described in text. Each data points represent the mean value of five measurements 74
- 5.7 Electropherogram of acceptor solution after EME-HPIM of spiked human plasma containing drugs at 500 ng/ml. Extraction condition: Extraction condition; Acceptor solution: 20  $\mu$ L of 0.1 mM KCl;HPIM: 60 mg of CTA, 15 mg of TEHP and 40 mg of D2EHPA; Sample pH: pH 7; Extraction voltage; 300 V; Extraction time; 10 min. CE conditions were as described in text 78
- 6.1 Simultaneous electromembrane extraction across hollow polymer inclusion membrane set up 85
- 6.2 Effect on the number of capillaries on extraction efficiency. Extraction parameter; sample solution: 200 ng/mL PQ and DQ in 4 mL of 0.1 mM KCl; acceptor solution: 20  $\mu$ L of 0.1 mM KCl; HPIM: 60 mg of CTA, 15 mg of TEHP and 40 mg of D2EHPA; Extraction voltage; 1000 V; Extraction time; 15 min 89

- 6.3 Effect of the carrier amount on the simultaneous EME – HPIM for cationic (a) and anionic (b) herbicides. Sample phase: 200 ng/mL herbicides in 4 mL 100  $\mu$ M KCl. Acceptor phase (both anodic and cathodic): 20  $\mu$ L of 100  $\mu$ M KCl. Sample pH: pH 7. Applied voltage: 1000 V. Extraction time: 15 min. Each data point represents the mean value of five measurements 91
- 6.4 Effect of the applied voltage on simultaneous EME – HPIM for cationic (a) and anionic (b) herbicides Sample phase: 200 ng/mL pesticide in 4 mL 100  $\mu$ M KCl. HPIM; 60 mg of CTA, 15 mg of TEHP, 3.5 mg of Aliquat 336/35 mg of D2EHPA. Acceptor phase (both anodic and cathodic): 20  $\mu$ L of 100  $\mu$ M KCl. Sample pH: no adjustment. Extraction time: 15 min. Each data point represents the mean value of five measurements 94
- 6.5 Effect of the extraction time of simultaneous EME – HPIM for cationic (a) and anionic (b) herbicides Sample phase: 200 ng/mL pesticide in 4 mL 100  $\mu$ M KCl. HPIM; 60 mg of CTA, 15 mg of TEHP, 3.5 mg of Aliquat 336/35 mg of D2EHPA. Acceptor phase (both anodic and cathodic): 20  $\mu$ L of 100  $\mu$ M KCl. Sample pH: no adjustment. Extraction voltage: 3000 V. Each data point represents the mean value of five measurements 95
- 6.6 Electropherogram of the cathodic (a) and anodic (b) acceptor solutions after the simultaneous EME – HPIM of the spiked river water samples (with an added background of 100  $\mu$ M KCl) containing herbicides at 200 mg/mL. Acceptor phase (both anodic and cathodic): 20  $\mu$ L of 100  $\mu$ M KCl. Sample pH: no adjustment. Applied voltage: 3000 V. Extraction time: 20 min. CE conditions were as described in the text 100

**LIST OF ABBREVIATIONS**

Ag(CN) <sub>2</sub>	-	Silver cyanide
AP	-	Amphetamine
Cd	-	Cadmium
CE	-	Capillary electrophoresis
CNT	-	Carbon nanotube
CPA	-	Chlorinated phenoxyacetic acids
Cr	-	Chromium
CTA	-	Cellulose triacetate
CZE	-	Capillary Zone Electrophoresis
C <sup>4</sup> D	-	Capacitively coupled contactless conductivity detector
DLME	-	Dispersive liquid microextraction
D2EHPA	-	Di-(2-ethylhexyl) phosphoric acid
DQ	-	Diquat
EOF	-	Electroosmotic flow
EME	-	Electromembrane Extraction
FESEM	-	Field emission scanning electron microscopy
GC	-	Gas chromatography
HPIM	-	Hollow polymer inclusion membrane
HF-LPME	-	Hollow Fiber- Liquid Phase Microextraction
KCl	-	Potassium chloride
KOH	-	Potassium hydroxide
LC	-	Liquid chromatography
LLE	-	Liquid-Liquid Extraction
LPME	-	Liquid Phase Microextraction
MA	-	Methamphetamine
MDA	-	3,4-methylenedioxy amphetamine
MDMA	-	3,4-methylenedioxy methamphetamine

MEKC	-	Micellar electrokinetic chromatography
NaCl	-	Sodium chloride
NaOH	-	Sodium hydroxide
NPOE	-	2-nitrophenyl octyl ether
PA	-	Polyacrylate
Pb	-	Lead
PDMS	-	Polydimethylsiloxane
PIM	-	Polymer inclusion membrane
PVC	-	Poly(vinyl chloride)
PQ	-	Paraquat
RSD	-	Relative standard deviation
SBSE	-	Stir bar sorptive extraction
SDME	-	Single drop microextraction
SDS	-	Sodium dodecyl sulphate
SLM	-	Supported Liquid Membrane
SPE	-	Solid Phase Extraction
SPME	-	Solid Phase Microextraction
TEHP	-	Tris-(2-ethylhexyl) phosphate
UV	-	Ultra violet
Zn	-	Zinc
2,4- D	-	(4-chlorophenoxy) acetic acid
4-CPA	-	2-(4-dichlorophenoxy) acetic acid

**LIST OF SYMBOLS**

A	-	Ampere
$C_{ih}$	-	Analyte concentration
cm	-	Centimeter
$\text{gmol}^{-1}$	-	Gram per mol
g	-	Gram
h	-	Hour
H	-	Height
$I$	-	Current
I.D	-	Internal diameter
L		Liter
mbar	-	Millibar
mg	-	Milligram
MHz	-	Megahertz
min	-	Minute
mL	-	Milliliter
mM	-	Millimolar
$\text{ng mL}^{-1}$	-	Nanogram per milliliter
nm	-	Nanometer
rpm	-	Revolutions per minute
$\mu\text{A}$	-	Microampere
$\mu\text{L}$	-	Microliter
$\mu\text{g/mL}$	-	Microgram per milliliter
$\mu\text{m}$	-	Micrometer

**LIST APPENDICES**

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
A	List of Publication Related to this Study	124
B	List of Presentation Related to this Study	125

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background

The target analytes (e.g. drugs, pesticides and herbicides) in environmental or biological samples usually occur in complex matrices, which are able to disturb the separation and analysis steps. Therefore, a series of procedures called sample preparation is needed to remove the interference substances, pre-concentrate the target analyte and increase the sensitivity. In addition, the sample preparation method can help to solve the limitations of the sensitivity of the analytical instrument detector.

As a consequence, many sample preparation methods have been developed for various applications in analytical methods. Most of the developed methods are aimed at searching for more cost effective analyses, higher sample preparation throughput, faster procedures and low consumption of solvents (1). These innovative techniques are derived from traditional sample preparation methods, namely liquid-liquid extraction (LLE) and solid phase extraction (SPE). The major drawbacks of the LLE method are that it is time consuming and not environmentally friendly (2). SPE is relatively easy and it consumes low amounts of organic solvent. Nevertheless, the SPE cartridges are costly and evaporation of the eluent and reconstituents is normally required prior to analysis (3).

To address the problems such as large solvent needs and cost, new research committed towards the development of convenient, efficient, economical and

miniaturized sample preparation methods is required. A miniaturized version of solid-phase extraction (SPE), termed solid-phase microextraction (SPME) (4) and liquid phase microextraction (LPME) (1, 2, 5) were introduced.

Hollow fibre liquid phase microextraction (HF-LPME) is one of the most promising techniques among LPME. In this method, a transport mechanism based on passive diffusion is applied, by adjusting a pH gradient established across the supported liquid membrane (SLM). In this technique, the target analytes are extracted from the sample solution through SLM which is held by capillary forces in the pores of a hollow fibre membrane (support) into the acceptor solution. The SLM is in direct contact with both the aqueous sample and the acceptor solution. Although this method can give good clean-up and also good selectivity by the proper organic solvents (SLM), the extraction time of 20-60 min was considered a major drawback (6).

For this reason, electro-assisted extraction was investigated based on the hypothesis that charged molecules can be transferred faster across SLM by an electric force than passive diffusion. Hence, in 2006, a new extraction principle termed electromembrane extraction (EME) was introduced (7). The equipment for EME was exactly the same as HF-LPME, except for the addition of two electrodes and a D/C power supply. The application of an electrical potential difference as the driving force successfully shortened the extraction time to within the range of 5-20 min (8). In the early development stages of the EME, most of the research focused on the screening and selection of differently composed SLMs. Significant efforts have been made over the course of the past year to expand the EME to a new application area and to improve its performance. The simultaneous electromembrane extraction of cationic and anionic analytes was reported by Safari *et al.* (2013) (9) along with the correlation between the EME and other extraction methods (10). EME has been widely employed in the analysis of drugs (11-14), the extraction of metals (15, 16) and the extraction of ions (17). This technique offered faster extraction time, lower consumption of organic solvent and also high pre-concentrations.



Most EMEs reported the use of water immiscible organic solvents such as 2-nitrophenyl octyl ether (NPOE), 1-octanol, tris (2-ethylhexyl) phosphate (TEHP), and di-hexyl ether (18) immobilized in the pores in the wall of hollow fibre membranes such as SLMs. Unfortunately, SLM was reported to have low mechanical stability, leading to membrane breakdown and leaching of the membrane liquid phase (19, 20). Kim *et al.* (2000) (21) have investigated the stability of polymer inclusion membrane (PIM) and SLM under similar conditions. They reported no flux decline or evidence of material losses within 15 days of continuous transport experiment in PIM. In contrast, leakage of the organic material in SLM after 48 h agitation in aqueous solution was clearly observed.

Consequently, the use of PIM as an alternative for electromembrane extraction has recently been investigated and was reported to be successful for the extraction of inorganic ions and pesticides (19, 22-25). PIM are self-supporting membranes, where a base polymer, plasticizer and carrier are incorporated into homogenous membrane. Therefore, for the first time, in this study, a new variation of the EME approach was created in which a hollow polymer inclusion membrane (HPIM) was developed for the extraction of selected drugs in human plasma. In addition, for the first time, the direct comparison of PIM and SLM for EME was undertaken in this study. The performance of the proposed method was also compared with the standard method (liquid-liquid extraction) used by the Toxicology Unit, Department of Chemistry, Malaysia (26).

The basis of EME is the electromigration of a targeted charged analyte under an electric field. Therefore, electric strength plays a crucial role in EME and depends on the applied voltage. Although it is anticipated that the extraction efficiency will increase as the applied voltage increases, there are some limitations to increasing the voltage; for example, instability of the system due to the formation of bubbles. In addition, the EME also suffered from an increase in the current level when high voltage was applied, especially in the analysis of real samples containing large amounts of ionic components. Therefore, there are several recent publications seeking to overcome the instability problem in the EME, including the application of

a stabiliser circuit in order to prevent the occurrence of an increasing current (27) and pulsed voltage (28). It was shown that the pulsed voltage increased the system stability by decreasing the thickness of the double layer at the interface (29).

For this reason, in this study, a stable EME was achieved by employing bubbleless electrodes to solve the problem of bubble formation during the extraction process. The bubbleless electrode was introduced by Gu *et al.* (2012) (30) and was employed in the electroosmotic pump for the purpose of solution delivery within a microfluidic device. The bubbleless setup facilitated the use of high voltage on the pump without the formation of the bubble in the pump channel.

A new way of thinking about the bubbleless electrode in order for it to be employed in the EME so as to eliminate the bubble formation when high voltage is applied during the extraction process was carried out in this study. A conventional EME approach using platinum electrodes was performed as a comparison.

## 1.2 Problem statement

Various sample preparation methods have been used in the analysis of drugs and pesticides/herbicides. A conventional extraction method like LLE and SPE was used for the analysis of drugs and pesticides. However, these methods are more time-consuming and use large volumes of organic solvents. On the other hand, HF-LPME was introduced to overcome the large consumption of organic solvents. This method offers several advantages, like good enrichment and sample clean-up, thereby reducing potential problems from matrix components (31). However, this method suffered a major drawback of long extraction times

By taking advantage of the electric field to enhance the extraction efficiency, a new extraction, namely EME, was introduced in 2006. In this method, a potential

difference is applied across the SLM which acts as the driving force. This method promises to be fast, simple, selective and rapid. Nevertheless, EME has problems with the mechanical instability of SLM, which can lead to a loss of the SLM under agitation and an electric field (19). Moreover, EME also suffers instability of the system due to the formation of bubbles when high voltages are applied (32). To address these issues, a PIM was introduced as it exhibits excellent stability and versatility compared to SLM.

In this study, a new variation of EME employing a hollow polymer inclusion membrane (HPIM) was developed. In order to get a better understanding, the comparison EME based on SLM was performed. Furthermore, this is the first time that a direct comparison between PIM and SLM was done. The developed methods were applied for the determination of amphetamine, methamphetamine and MDMA abuse in human plasma. In addition, the simultaneous EME of cationic and anionic herbicides was investigated by employing bubbleless electrodes to overcome a bubble formation problem when high voltages are applied during the extraction process. There have been no reports on the use of bubbleless electrodes as an alternative for stable EME.

### **1.3 Research Aim and Objectives**

The aim of this research is to develop simple, miniaturised sample preparation method based on EME for the analysis of drugs and herbicides. The objectives of the study are as follows:

1. To evaluate conventional EME across SLM for the analysis of amphetamine, methamphetamine and MDMA in human plasma
2. To design and characterize new membrane materials, namely HPIM, for the purpose of a highly stable EME approach

3. To develop and evaluate new methods based on EME across the HPIM for the analysis of amphetamine, methamphetamine and MDMA in human plasma
4. To prepare a bubbleless electrode for the simultaneous ultra-high voltage EME of cationic and anionic herbicides across the HPIM

#### **1.4 Scopes of the Research**

In this study, an innovative development of EME with capillary electrophoresis (CE) was developed for the analysis of drugs and herbicides in human plasma and river water samples. The potential to employ HPIM as an alternative to SLM was investigated for the analysis of amphetamine, methamphetamine and MDMA in human plasma in order to achieve greater membrane stability. In this work, the HPIMs were prepared, characterized and applied in EME. The HPIMs were characterized using field emission scanning electron microscopy (FESEM) and contact angle analysis. In order to get a good extraction efficiency, a series of optimizations was done, including optimization of the amount of plasticizer, the amount of carrier, sample pH, extraction voltage and extraction time. Analytical performances of the developed methods were evaluated, validated and applied to the analysis of drug of abuse in human plasma. The study was expanded by introducing a bubbleless electrode to overcome bubble formation during extraction when using a high voltage. The preparation of bubbleless electrodes was characterized in detail. The simultaneous electromembrane extraction of cationic and anionic herbicides employing the bubbleless electrode was developed. Parameters affecting EME were optimized comprehensively and applied to determine cationic and anionic herbicides in river water samples. A conventional EME using normal electrodes was performed as a comparison.

## 1.5 Significance of Research

In this study, EME across HPIM was developed for the first time. HPIMs were prepared as alternatives to SLM. It is expected that HPIM can give better stability compared to SLM due to its physical and chemical features. This proposed method was subsequently employed for the analysis of amphetamine, methamphetamine and MDMA in human plasma. This particular application is important to demonstrate the sample clean-up capability of the new proposed membrane materials against complex matrices present in human biological fluid.

Meanwhile, the introduction of bubbleless electrodes in EME could improve the stability of EME systems when a high voltage was applied. In addition, a simultaneous EME approach was introduced and this unique setup enabled the extraction of positively and negatively charged analytes at the same time. Therefore, it is expected that the extraction process could be achieved in a simple setup without any tedious procedures. In addition, the developed methods are expected to be simple, fast, efficient, sensitive and environmentally friendly, potentially being adopted as established methods for the monitoring of other drugs and organic pollutants such as pesticides.

## 1.6 Outline of the Thesis

This thesis consists of seven chapters. Chapter 1 describes in detail the research background, problem statement, objective, scope and significance of the study. Chapter 2 compiles the literature review on the details of conventional sample preparation method and microextraction of derivative sample preparation methods and focusses on EME and the development of PIM.

Chapter 3 discusses the microextraction of EME across SLM combined with capillary electrophoresis with a  $C^4D$  detector for the analysis of three selected drugs in human plasma. The parameter affecting the extraction efficiency was investigated

## REFERENCES

1. Dadfarnia, S., Haji Shabani, A. M. Recent development in liquid phase microextraction for determination of trace level concentration of metals- A review. *Anal. Chim. Acta.* 2010. 658(2):107-119.
2. Asensio-Ramos, M., Ravelo-Pérez, L. M., González-Curbelo, M. Á., Hernández-Borges, J. Liquid phase microextraction applications in food analysis. *J. Chromatogr. A.* 2011. 1218(42):7415-7437.
3. Augusto, F., Hantao, L. W., Mogollón, N. G. S., Braga, S. C. G. N. New materials and trends in sorbents for solid-phase extraction. *TrAC-Trend in Anal. Chem.* 2013. 43:14-23.
4. Piri-Moghadam, H., Ahmadi, F., Pawliszyn, J. A critical review of solid phase microextraction for analysis of water samples. *TrAC-Trend Anal Chem.* 2016. 85, Part A:133-143.
5. Halvorsen, T. G., Pedersen-Bjergaard, S., Rasmussen, K. E. Liquid-phase microextraction and capillary electrophoresis of citalopram, an antidepressant drug. *J. Chromatogr. A.* 2001. 909(1):87-93.
6. Ghambarian, M., Yamini, Y., Esrafil, A. Developments in hollow fiber based liquid-phase microextraction: principles and applications. *Microchimica Acta.* 2012. 177(3):271-294.
7. Pedersen-Bjergaard, S., Rasmussen, K. E. Electrokinetic migration across artificial liquid membranes: New concept for rapid sample preparation of biological fluids. *J. Chromatogr. A.* 2006. 1109(2):183-190.

8. Gjelstad, A., Jensen, H., Rasmussen, K. E., Pedersen-Bjergaard, S. Kinetic aspects of hollow fiber liquid-phase microextraction and electromembrane extraction. *Anal Chim Acta*. 2012. 742:10-16.
9. Safari, M., Nojavan, S., Davarani, S. S. H., Morteza-Najarian, A. Speciation of chromium in environmental samples by dual electromembrane extraction system followed by high performance liquid chromatography. *Anal Chim Acta*. 2013. 789:58-64.
10. Arjomandi-Behzad, L., Yamini, Y., Rezazadeh, M. Extraction of pyridine derivatives from human urine using electromembrane extraction coupled to dispersive liquid-liquid microextraction followed by gas chromatography determination. *Talanta*. 2014. 126:73-81.
11. Seidi, S., Yamini, Y., Heydari, A., Moradi, M., Esrafil, A., Rezazadeh, M. Determination of thebaine in water samples, biological fluids, poppy capsule, and narcotic drugs, using electromembrane extraction followed by high-performance liquid chromatography analysis. *Anal Chim Acta*. 2011. 701(2):181-188.
12. Eibak, L. E. E., Gjelstad, A., Rasmussen, K. E., Pedersen-Bjergaard, S. Exhaustive electromembrane extraction of some basic drugs from human plasma followed by liquid chromatography-mass spectrometry. *J. Pharmaceut. BioMed*. 2012. 57:33-38.
13. Gjelstad, A., Rasmussen, K. E., Pedersen-Bjergaard, S. Simulation of flux during electro-membrane extraction based on the Nernst-Planck equation. *J. Chromatogr. A*. 2007. 1174(1-2):104-111.
14. Eskandari, M., Yamini, Y., Fotouhi, L., Seidi, S. Microextraction of mebendazole across supported liquid membrane forced by pH gradient and electrical field. *J. Pharmaceut. BioMed*. 2011. 54(5):1173-1179.

15. Kamyabi, M. A., Aghaei, A. Electromembrane extraction and spectrophotometric determination of As (V) in water samples. *Food Chem.* 2016. 212:65-71.
16. Davarani, S. S. H., Moazami, H. R., Keshtkar, A. R., Banitaba, M. H., Nojavan, S. A selective electromembrane extraction of uranium (VI) prior to its fluorometric determination in water. *Anal. Chim. Acta.* 2013. 783:74-79.
17. Tan, T. Y., Basheer, C., Ng, K. P., Lee, H. K. Electro membrane extraction of biological anions with ion chromatographic analysis. *Anal Chim Acta.* 2012. 739:31-36.
18. Yamini, Y., Seidi, S., Reza zadeh, M. Electrical field-induced extraction and separation techniques: Promising trends in Anal Chem– A review. *Anal Chim Acta.* 2014. 814:1-22.
19. See, H. H., Hauser, P. C. Electric field-driven extraction of lipophilic anions across a carrier-mediated polymer inclusion membrane. *Anal Chem.* 2011. 83(19):7507-7513.
20. Nghiem, L. D., Mornane, P., Potter, I. D., Perera, J. M., Cattrall, R. W., Kolev, S. D. Extraction and transport of metal ions and small organic compounds using polymer inclusion membranes (PIMs). *J. Membr. Sci.* 2006. 281(1–2):7-41.
21. Kim, J. S., Kim, S. K., Ko, J. W. Selective transport of cesium ion in polymeric CTA membrane containing calixcrown ethers. *Talanta.* 2000. 52(6):1143-1148.
22. Schmidt-Marzinkowski, J., See, H. H., Hauser, P. C. Electric field driven extraction of inorganic anions across a polymer inclusion membrane. *Electroanal.* 2013. 25(8):1879-1886.
23. See, H. H., Hauser, P. C. Electro-driven extraction of low levels of lipophilic organic anions and cations across plasticized cellulose triacetate membranes: Effect of the membrane composition. *J. Membr. Sci.* 2014. 450:147-152.



24. See, H. H., Stratz, S., Hauser, P. C. Electro-driven extraction across a polymer inclusion membrane in a flow-through cell. *J. Chromatogr. A*. 2013. 1300:79-84.
25. Seip, K. F., Jensen, H., Kieu, T. E., Gjelstad, A., Pedersen-Bjergaard, S. Salt effects in electromembrane extraction. *J. Chromatogr. A*. 2014. 1347:1-7.
26. O'Neil, M. J. The Merck index: An Encyclopedia of Chemicals, Drugs, and Biologicals. 2001.
27. Rezazadeh, M., Yamini, Y., Seidi, S., Arjomandi-Behzad, L. Voltage-step pulsed electromembrane as a novel view of electrical field-induced liquid-phase microextraction. *J. Chromatogr. A*. 2014. 1324:21-28.
28. Arjomandi-Behzad, L., Yamini, Y., Rezazadeh, M. Pulsed electromembrane method for simultaneous extraction of drugs with different properties. *Anal Biochem*. 2013. 438(2):136-143.
29. Rezazadeh, M., Yamini, Y., Seidi, S., Esrafil, A. Pulsed electromembrane extraction: A new concept of electrically enhanced extraction. *J. Chromatogr. A*. 2012. 1262:214-218.
30. Gu, C., Jia, Z., Zhu, Z. Miniaturized electroosmotic pump capable of generating pressures of more than 1200 Bar. *Anal. Chem*. 2012. 84(21):9609-9614.
31. Lee, J., Lee, H. K., Rasmussen, K. E., Pedersen-Bjergaard, S. Environmental and bioanalytical applications of hollow fiber membrane liquid-phase microextraction: A review. *Anal. Chim. Acta*. 2008. 624(2):253-268.
32. Rezazadeh, M., Yamini, Y., Seidi, S. Electromembrane extraction of trace amounts of naltrexone and nalmefene from untreated biological fluids. *J. Chromatogr. A*. 2011. 879(15-16):1143-1148.

33. Li, N., Song, Y. P., Tang, H., Wang, Y. Recent developments in sample preparation and data pre-treatment in metabonomics research. *Archives of Biochemistry and Biophysics*. 2016. 589:4-9.
34. Psillakis, E., Kalogerakis, N. Developments in liquid-phase microextraction. *TrAC-Trend in Anal. Chem.* 2003. 22(9):565-574.
35. Xiong, J., Chen, J., He, M., Hu, B. Simultaneous quantification of amphetamines, caffeine and ketamine in urine by hollow fiber liquid phase microextraction combined with gas chromatography-flame ionization detector. *Talanta*. 2010. 82(3):969-975.
36. Arthur, C. L., Pawliszyn, J. Solid phase microextraction with thermal desorption using fused silica optical fibers. *Anal. Chem.* 1990. 62(19):2145-2148.
37. Spietelun, A., Marcinkowski, L., de la Guardia, M., Namieśnik, J. Green aspects, developments and perspectives of liquid phase microextraction techniques. *Talanta*. 2014. 119:34-45.
38. Terzopoulou, Z., Papageorgiou, M., Kyzas, G. Z., Bikiaris, D. N., Lambropoulou, D. A. Preparation of molecularly imprinted solid-phase microextraction fiber for the selective removal and extraction of the antiviral drug abacavir in environmental and biological matrices. *Anal. Chim. Acta*. 2016. 913:63-75.
39. Zhang, S., Yang, Q., Yang, X. A zeolitic imidazolate framework based nanoporous carbon as a novel fiber coating for solid-phase microextraction of pyrethroid pesticides. *Talanta*. 2017. 166:46-53.
40. Baltussen, E., Sandra, P., David, F., Cramers, C. Stir bar sorptive extraction (SBSE), a novel extraction technique for aqueous samples: Theory and principles. *J. Microcolumn Sep.* 1999. 11(10):737-747.
41. Mao, X., He, M., Chen, B., Hu, B. Membrane protected C18 coated stir bar sorptive extraction combined with high performance liquid chromatography-

- ultraviolet detection for the determination of non-steroidal anti-inflammatory drugs in water samples. *J. Chromatogr. A*. 2016. 1472:27-34.
42. Liu, H., Dasgupta, P. K. Anal chemin a drop. solvent extraction in a microdrop. *Anal. Chem.* 1996. 68(11):1817-1821.
  43. Jeannot, M. A., Cantwell, F. F. Solvent microextraction into a single drop. *Anal Chem.* 1996. 68(13):2236-2240.
  44. Psillakis, E., Kalogerakis, N. Developments in single-drop microextraction. *TrAC- Trend in Anal. Chem.* 2002. 21(1):54-64.
  45. Xu, L., Basheer, C., Lee, H. K. Developments in single-drop microextraction. *J. Chromatogr. A*. 2007. 1152(1-2):184-192.
  46. Wang, X., Li, Y., Cai, F. Fully automatic single-drop microextraction with one-step extraction and derivatization and its application for rapid analysis of hydroxylated polycyclic aromatic hydrocarbons in seawaters. *Talanta*. 2017. 164:727-734.
  47. Hadjmohammadi, M., Karimiyan, H., Sharifi, V. Hollow fibre-based liquid phase microextraction combined with high-performance liquid chromatography for the analysis of flavonoids in *Echinophora platyloba* DC. and *Mentha piperita*. *Food Chem.* 2013. 141(2):731-735.
  48. Hadjmohammadi, M. R., Soltani, M., Sharifi, V. Use of hollow fiber liquid phase microextraction and HPLC for extraction and determination of apigenin in human urine after consumption of *Satureja sahendica* Bornm. *J. Chromatogr. A*. 2012. 900:85-88.
  49. Pedersen-Bjergaard, S., Rasmussen, K. E. Liquid-phase microextraction with porous hollow fibers, a miniaturized and highly flexible format for liquid-liquid extraction. *J. Chromatogr. A*. 2008. 1184(1-2):132-142.
  50. Richoll, S. M., Colón, I. Determination of triphenylphosphine oxide in active pharmaceutical ingredients by hollow-fiber liquid-phase microextraction

- followed by reversed-phase liquid chromatography. *J. Chromatogr. A.* 2006. 1127(1–2):147-153.
51. Gjelstad, A., Andersen, T. M., Rasmussen, K. E., Pedersen-Bjergaard, S. Microextraction across supported liquid membranes forced by pH gradients and electrical fields. *J. Chromatogr. A.* 2007. 1157(1–2):38-45.
  52. Esrafil, A., Yamini, Y., Ghambarian, M., Moradi, M., Seidi, S. A novel approach to automation of dynamic hollow fiber liquid-phase microextraction. *J. Sep. Sci.* 2011. 34(8):957-964.
  53. Huang, S., Hu, D., Wang, Y., Zhu, F., Jiang, R., Ouyang, G. Automated hollow-fiber liquid-phase microextraction coupled with liquid chromatography/tandem mass spectrometry for the analysis of aflatoxin M1 in milk. *J. Chromatogr. A.* 2015. 1416:137-140.
  54. Gjelstad, A., Rasmussen, K. E., Pedersen-Bjergaard, S. Electrokinetic migration across artificial liquid membranes: Tuning the membrane chemistry to different types of drug substances. *J. Chromatogr. A.* 2006. 1124(1–2):29-34.
  55. Middelthon-Bruer, T. M., Gjelstad, A., Rasmussen, K. E., Pedersen-Bjergaard, S. Parameters affecting electro membrane extraction of basic drugs. *J. Sep. Sci.* 2008. 31(4):753-759.
  56. Fashi, A., Yaftian, M.R., Zamani, A. Determination of melamine in dairy products using electromembrane–LPME followed by HPLC. *Food Chem.* 2015. 188:92-98.
  57. Zahedi, P., Davarani, S. S. H., Moazami, H. R., Nojavan, S. Surfactant assisted pulsed two-phase electromembrane extraction followed by GC analysis for quantification of basic drugs in biological samples. *J. Pharmaceut. BioMed.* 2016. 117:485-491.

58. Payán, M. D. R., Li, B., Petersen, N. J., Jensen, H., Hansen, S. H., Pedersen-Bjergaard, S. Nano-electromembrane extraction. *Anal Chim Acta*. 2013. 785:60-66.
59. Alhooshani, K., Basheer, C., Kaur, J. Electromembrane extraction and HPLC analysis of haloacetic acids and aromatic acetic acids in wastewater. *Talanta*. 2011. 86:109-113.
60. Balchen, M., Halvorsen, T. G., Reubsaet, L., Pedersen-Bjergaard, S. Rapid isolation of angiotensin peptides from plasma by electromembrane extraction. *J. Chromatogr. A*. 2009. 1216(41):6900-6905.
61. Hasheminasab, K. S., Fakhari, A. R. Development and application of carbon nanotubes assisted electromembrane extraction (CNTs/EME) for the determination of buprenorphine as a model of basic drugs from urine samples. *Anal. Chim. Acta*. 2013. 767:75-80.
62. Šlampoová, A., Kubáň, P., Boček, P. Electromembrane extraction using stabilized constant d.c. electric current—A simple tool for improvement of extraction performance. *J. Chromatogr. A*. 2012. 1234:32-37.
63. Seidi, S., Yamini, Y., Rezazadeh, M. Combination of electromembrane extraction with dispersive liquid–liquid microextraction followed by gas chromatographic analysis as a fast and sensitive technique for determination of tricyclic antidepressants. *J. Chromatogr. A*. 2013. 913–914:138-146.
64. Kjelsen, I. J. Ø., Gjelstad, A., Rasmussen, K. E., Pedersen-Bjergaard, S. Low-voltage electromembrane extraction of basic drugs from biological samples. *J. Chromatogr. A*. 2008. 1180(1–2):1-9.
65. Balchen, M., Reubsaet, L., Pedersen-Bjergaard, S. Electromembrane extraction of peptides. *J. Chromatogr. A*. 2008. 1194(2):143-149.
66. Seidi, S., Yamini, Y., Rezazadeh, M. Electrically enhanced microextraction for highly selective transport of three  $\beta$ -blocker drugs. *J. Pharmaceut BioMed*. 2011. 56(5):859-866.

67. Payán, M. R., López, M. Á. B., Torres, R. F., Navarro, M. V., Mochón, M. C. Electromembrane extraction (EME) and HPLC determination of non-steroidal anti-inflammatory drugs (NSAIDs) in wastewater samples. *Talanta*. 2011. 85(1):394-399.
68. Seidi, S., Yamini, Y., Rezazadeh, M., Esrafil, A. Low-voltage electrically-enhanced microextraction as a novel technique for simultaneous extraction of acidic and basic drugs from biological fluids. *J. Chromatogr. A*. 2012. 1243:6-13.
69. Balchen, M., Lund, H., Reubsæet, L., Pedersen-Bjergaard, S. Fast, selective, and sensitive analysis of low-abundance peptides in human plasma by electromembrane extraction. *Anal Chim Acta*. 2012. 716:16-23.
70. Davarani, S. S. H., Najarian, A. M., Nojavan, S., Tabatabaei, M. A. Electromembrane extraction combined with gas chromatography for quantification of tricyclic antidepressants in human body fluids. *Anal Chim Acta*. 2012. 725:51-56.
71. Walkowiak, W., Bartsch, R. A., Kozłowski, C., Gega, J., Charewicz, W. A., Amiri-Eliasi, B. Separation and removal of metal ionic species by polymer inclusion membranes. *J. Radioanal Nuclear Chem*. 2005. 246(3):643-650.
72. Raut, D. R., Mohapatra, P. K., Ansari, S. A., Sarkar, A., Manchanda, V. K. Selective transport of radio-cesium by supported liquid membranes containing calix[4]crown-6 ligands as the mobile carrier. *Desalination*. 2008. 232(1-3):262-271.
73. Tabani, H., Fakhari, A. R., Shahsavani, A. Combination of graphene oxide-based solid phase extraction and electro membrane extraction for the preconcentration of chlorophenoxy acid herbicides in environmental samples. *J. Chromatogr. A*. 2013. 1300:227-235.
74. Kebiche-Senhadji, O., Mansouri, L., Tingry, S., Seta, P., Benamor, M. Facilitated Cd(II) transport across CTA polymer inclusion membrane using

- anion (Aliquat 336) and cation (D2EHPA) metal carriers. *J. Membr. Sci.* 2008. 310(1–2):438-445.
75. Annane, K., Sahmoune, A., Montels, P., Tingry, S. Polymer inclusion membrane extraction of cadmium(II) with Aliquat 336 in micro-channel cell. *Chem Eng Research and Design.* 2015. 94:605-610.
76. O'Bryan, Y., Cattrall, R. W., Truong, Y. B., Kyratzis, I. L., Kolev, S. D. The use of poly(vinylidene fluoride-co-hexafluoropropylene) for the preparation of polymer inclusion membranes. Application to the extraction of thiocyanate. *J. Membr. Sci.* 2016. 510:481-488.
77. Kebiche-Senhadji, O., Tingry, S., Seta, P., Benamor, M. Selective extraction of Cr(VI) over metallic species by polymer inclusion membrane (PIM) using anion (Aliquat 336) as carrier. *Desalination.* 2010. 258(1–3):59-65.
78. Yilmaz, A., Arslan, G., Tor, A., Akin, I. Selectively facilitated transport of Zn(II) through a novel polymer inclusion membrane containing Cyanex 272 as a carrier reagent. *Desalination.* 2011. 277(1–3):301-307.
79. Mahmoudi, D. H., Merzouk, D. N. K., Spahis, D. N. SWEE'11 Cationic dye (MB) removal using polymer inclusion membrane (PIMs). *Proc Eng.* 2012. 33:38-46.
80. Zawierucha, I., Kozłowski, C., Malina, G. Removal of toxic metal ions from landfill leachate by complementary sorption and transport across polymer inclusion membranes. *Waste Management.* 2013. 33(10):2129-2136.
81. Konczyk, J., Kozłowski, C., Walkowiak, W. Removal of chromium(III) from acidic aqueous solution by polymer inclusion membranes with D2EHPA and Aliquat 336. *Desalination.* 2010. 263(1–3):211-216.
82. Arous, O., Amara, M., Trari, M., Bouguelia, A., Kerdjoudj, H. Cadmium (II) and lead (II) transport in a polymer inclusion membrane using tributyl phosphate as mobile carrier and CuFeO<sub>2</sub> as a polarized photo electrode. *J. Hazard Mater.* 2010. 180(1–3):493-498.

83. St John, A. M., Catrall, R. W., Kolev, S. D. Transport and separation of uranium (VI) by a polymer inclusion membrane based on di-(2-ethylhexyl) phosphoric acid. *J. Membr. Sci.* 2012. 409–410:242-250.
84. Iben Nasser, I., Ibn El Haj Amor, F., Donato, L. Removal and recovery of from synthetic electroplating baths by polymer inclusion membrane containing Aliquat 336 as a carrier. *J. Chem Eng.* 2016. 295:207-217.
85. Schow, A. J., Peterson, R. T., Lamb, J. D. Polymer inclusion membranes containing macrocyclic carriers for use in cation separations. *J. Membr. Sci.* 1996. 111(2):291-295.
86. Ulewicz, M., Szczygelska-Tao, J., Biernat, J. F. Selectivity of Pb (II) transport across polymer inclusion membranes doped with imidazole azothiacrown ethers. *J. Membr. Sci.* 2009. 344(1–2):32-38.
87. Pantůčková, P., Kubáň, P., Boček, P. In-line coupling of microextractions across polymer inclusion membranes to capillary zone electrophoresis for rapid determination of formate in blood samples. *Anal Chim Acta.* 2015. 887:111-117.
88. Jones, G. M., Vale, J. A. Mechanisms of toxicity, clinical features, and management of diquat poisoning: A Review. *J.Toxic: Clinical Toxic.* 2000. 38(2):123-128.
89. Lock, E. A., Wilks, M. F. Chapter 83 - Paraquat A2 - Krieger, Robert. Hayes' Handbook of pesticide toxicology (Third Edition). Academic Press, New York, . 2010. pp. 1771-1827.
90. Classification of pesticides by hazard and guidelines to classification. 2009.
91. EEC Drinking Water Guidelines 80/779/EEC. 1980.
92. Ruan, X. L., Qiu, J. J., Wu, C., Huang, T., Meng, R. B., Lai, Y. Q. Magnetic single-walled carbon nanotubes–dispersive solid-phase extraction method



- combined with liquid chromatography–tandem mass spectrometry for the determination of paraquat in urine. *J. Chromatogr. A*. 2014. 965:85-90.
93. Grey, L., Nguyen, B., Yang, P. Liquid chromatography–electrospray ionization isotope dilution mass spectrometry analysis of paraquat and diquat using conventional and multilayer solid-phase extraction cartridges. *J. Chromatogr. A*. 2002. 958(1–2):25-33.
94. Zeng, J., Chen, J., Li, M. Determination of amphetamines in biological samples using electro enhanced solid-phase microextraction-gas chromatography. *J. Chromatogr. A*. 2015. 1000:169-175.
95. Chiang, J. S., Huang, S. D. Simultaneous derivatization and extraction of amphetamine and methylenedioxyamphetamine in urine with headspace liquid-phase microextraction followed by gas chromatography–mass spectrometry. *J. Chromatogr. A*. 2008. 1185(1):19-22.
96. Ahmadi-Jouibari, T., Fattahi, N., Shamsipur, M. Rapid extraction and determination of amphetamines in human urine samples using dispersive liquid-liquid microextraction and solidification of floating organic drop followed by high performance liquid chromatography. *J. Pharmaceut BioMed*. 2014. 94:145-151.
97. Seidi, S., Yamini, Y., Baheri, T., Feizbakhsh, R. Electrokinetic extraction on artificial liquid membranes of amphetamine-type stimulants from urine samples followed by high performance liquid chromatography analysis. *J. Chromatogr. A*. 2011. 1218(26):3958-3965.
98. Pantaleão, L. D. N., Paranhos, B. A. P. B., Yonamine, M. Hollow-fiber liquid-phase microextraction of amphetamine-type stimulants in human hair samples. *J. Chromatogr. A*. 2012. 1254:1-7.
99. Kohler, I., Schappler, J., Sierro, T., Rudaz, S. Dispersive liquid–liquid microextraction combined with capillary electrophoresis and time-of-flight mass spectrometry for urine analysis. *J. Pharmaceut. BioMed*. 2013. 73:82-89.

100. Ridgway, K., Lalljie, S. P. D., Smith, R.M. Sample preparation techniques for the determination of trace residues and contaminants in foods. *J. Chromatogr. A*. 2007. 1153(1–2):36-53.
101. National Primary Drinking Water REGULATIONS. Enviromental Protection Agency (USEPA), Washington, DC, Office of water, U.S. 2009.
102. He, Y., Kang, Y. J. Single drop liquid–liquid–liquid microextraction of methamphetamine and amphetamine in urine. *J. Chromatogr. A*. 2006. 1133(1–2):35-40.
103. Grønhaug Halvorsen, T., Pedersen-Bjergaard, S., Rasmussen, K. E. Reduction of extraction times in liquid-phase microextraction. *J. Chromatogr B*. 2001. 760(2):219-226.
104. Jamt, R. E. G., Gjelstad, A., Eibak, L. E. E. Electromembrane extraction of stimulating drugs from undiluted whole blood. *J. Chromatogr. A*. 2012. 1232:27-36.
105. Huang, Z., Zhang, S. Confirmation of amphetamine, methamphetamine, MDA and MDMA in urine samples using disk solid-phase extraction and gas chromatography–mass spectrometry after immunoassay screening. *J. Chromatogr. A*. 2003. 792(2):241-247.
106. Basheer, C., Lee, H. K. Hollow fiber membrane-protected solid-phase microextraction of triazine herbicides in bovine milk and sewage sludge samples. *J. Chromatogr. A*. 2004. 1047(2):189-194.
107. Sarafraz-Yazdi, A., Amiri, A., Rounaghi, G., Eshtiagh-Hosseini, H. Determination of non-steroidal anti-inflammatory drugs in water samples by solid-phase microextraction based sol–gel technique using poly(ethylene glycol) grafted multi-walled carbon nanotubes coated fiber. *Anal Chim Acta*. 2012. 720:134-141.

108. Gjelstad, A., Pedersen-Bjergaard, S. Electromembrane extraction: a new technique for accelerating bioanalytical sample preparation. *Bioanal.* 2011. 3(7):787-797.
109. Seip, K. F., Gjelstad, A., Pedersen-Bjergaard, S. Electromembrane extraction from aqueous samples containing polar organic solvents. *J. Chromatogr. A.* 2013. 1308:37-44.
110. Mahanty, B. N., Mohapatra, P. K., Raut, D. R. Polymer inclusion membrane containing a diglycolamide-functionalized calix[4]arene for actinide ion uptake and transport. *J. Membr. Sci.* 2016. 516:194-201.
111. Mukhtar, N. H., See, H. H. Carbonaceous nanomaterials immobilised mixed matrix membrane microextraction for the determination of polycyclic aromatic hydrocarbons in sewage pond water samples. *Anal Chim Acta.* 2016. 931:57-63.
112. Park, M. J., Nisola, G. M., Vivas, E. L. Mixed matrix nanofiber as a flow-through membrane adsorber for continuous  $\text{Li}^+$  recovery from seawater. *J. Membr. Sci.* 2016. 510:141-154.
113. Kamaruzaman, S., Hauser, P. C., Sanagi, M.M., Ibrahim, W. A. W., Endud, S., See, H. H. A simple microextraction and preconcentration approach based on a mixed matrix membrane. *Anal Chim Acta.* 2013. 783:24-30.
114. Kemperman, A. J. B., Bargeman, D., Van Den Boomgaard, T., Strathmann, H. Stability of supported liquid membranes: State of the art. *Sep Sci. and Tech.* 1996. 31(20):2733-2762.
115. Danesi, P. R., Reichley-Yinger, L., Rickert, P. G. Lifetime of supported liquid membranes: the influence of interfacial properties, chemical composition and water transport on the long-term stability of the membranes. *J. Membr. Sci.* 1987. 31(2):117-145.

116. Baker, R. W., Cussler, E. L., Eykamp, W., Koros, W. J. R. R. L., Strathmann, H. Membrane Separation Systems - Recent Developments and Future Directions. William Andrew Publishing/Noyes.
117. Forch, R., Schonherr, H., Tobias, A., Jenkins, A. Surface design; Applications in Bioscience and Nanotechnology. p.47 ed. Wiley-VCH: Weinheim. 2009.
118. He, Y., Huang, K. L. Liquid-phase microextraction in a single drop of organic solvent by using a conventional microsyringe. *Anal. Chem.* 1997. 69(22):4634-4640.
119. Rezaee, M., Assadi, Y., Milani Hosseini, M.-R., Aghaee, E., Ahmadi, F., Berijani, S. Determination of organic compounds in water using dispersive liquid-liquid microextraction. *J. Chromatogr. A.* 2006. 1116(1-2):1-9.
120. Pedersen-Bjergaard, S., Kramula, E. R. Liquid-liquid-liquid microextraction for sample preparation of biological fluids prior to capillary electrophoresis. *Anal. Chem.* 1999. 71:2650-2656.
121. Basheer, C., Rao, S. Valliyaveetil, M., Lee, H. K. Development and application of porous membrane-protected carbon nanotube micro-solid-phase extraction combined with gas chromatography/mass Spectrometry. *Anal. Chem.* 2006. 78:2853-2858.
122. Bruheim, L., Pawliszyn, J. Thin-film microextraction. *Anal. Chem.* 2003. 75:2976-2983.
123. Morales-Cid, G., Simonet, B. M., Valcárcel, M. Sample treatments improved by electric fields. *Trends Anal. Chem.* 2010. 29:158-165.
124. Pedersen-Bjergaard, S., Rasmussen, K. E. Electrical potential can drive liquid-liquid extraction for sample preparation in chromatography. *Trends Anal. Chem.* 2008. 27:934-941.

125. Kubán, P., Bocek, P. Electric field-enhanced transport across phase boundaries and membranes and its potential use in sample pretreatment for bioanalysis. *Electrophoresis*. 2010. 31:768-785.
126. Collins, C., Moradi, A. D. A review of recent advances in electrochemically modulated extraction methods. *Anal. Bioanal. Chem.* 2009. 393:835-845.
127. Gjelstad, A., Pedersen-Bjergaard, S. Electromembrane extraction – three-phase electrophoresis for future preparative applications. *Electrophoresis*. 2014. 35:2421-2428.
128. Gjelstad, A., Pedersen.-Bjergaard, S. Recent developments in electromembrane extraction. *Anal. Methods*. 2013. 5:4549-4557.
129. Balchen, M., Jensen, H., Reubsaet, L., Pedersen-Bjergaard, S. Potential-driven peptide extractions across supported liquid membranes: Investigation of principal operational parameters. *J. Sep. Sci.* 2010. 33(11):1665-1672.
130. See, H. H., Hauser, P. C. Automated electric-field-driven membrane extraction system coupled to liquid chromatography-mass spectrometry. *Anal Chem.* 2014. 86(17):8665-8670.
131. Jeannot, M. A., Folin, F., C. Solvent microextraction into a single drop. *Anal. Chem.* 1996. 17(68):2236-2240.
132. See, H. H., Hausert, P. C. Electric field-driven extraction of lipophilic anions across a carrier-mediated polymer inclusion membrane. *Anal Chem.* 2011. 83(19):7507-7513.
133. Gardner, J. S., Walker, J. O., Lamb, J. D. Permeability and durability effects of cellulose polymer variation in polymer inclusion membranes. *J. Membr. Sci.* 2004. 229(1–2):87-93.
134. Campos, C. D. M., de Campos, R. P. S., da Silva, J. A. F., Jesus, D. P., Orlando, R. M. Sample preparation assisted by electric fields: fundamentals, advances, applications, and trends. *Quim. Nova.* 2015. 38(8):1093-1106.

135. Oedit, A., Ramautar, R., Hankemeier, T., Lindenburg, P. W. Electroextraction and electromembrane extraction: Advances in hyphenation to analytical techniques. *Electrophoresis*. 2016. 37(9):1170-1186.
136. Wuethrich, A., Haddad, P. R., Quirino, J. P. The electric field - An emerging driver in sample preparation. *Trac-Trends in Anal. Chem.* 2016. 80:604-611.
137. Rezazadeh, M., Yamini, Y., Seidi, S. Electrically stimulated liquid-based extraction techniques in bioanalysis. *Bioanal.* 2016. 8(8):815-828.
138. Yamini, Y., Seidi, S., Rezazadeh, M. Electrical field-induced extraction and separation techniques: Promising trends in Anal Chem– A review. *Anal. Chim. Acta*. 2014. 814:1-22.
139. Morales-Cid, G., Cardenas, S., Simonet, B. M., Valcarcel, M. Sample treatments improved by electric fields. *Trac-Trends in Anal. Chem.* 2010. 29(2):158-165.
140. Kuban, P., Slampova, A., Bocek, P. Electric field-enhanced transport across phase boundaries and membranes and its potential use in sample pretreatment for bioanalysis. *Electrophoresis*. 2010. 31(5):768-785.
141. See, H. H., Hauser, P. C., Sanagi, M. M., Ibrahim, W. A. W. Dynamic supported liquid membrane tip extraction of glyphosate and aminomethylphosphonic acid followed by capillary electrophoresis with contactless conductivity detection. *J. Chromatogr. A*. 2010. 1217(37):5832-5838.
142. Huang, C. X., Jensen, H., Seip, K. F., Gjelstad, A., Pedersen-Bjergaard, S. Mass transfer in electromembrane extraction-The link between theory and experiments. *J. Sep. Sci.* 2016. 39(1):188-197.
143. Huang, C., Seip, K. F., Gjelstad, A., Pedersen-Bjergaard, S. Electromembrane extraction for pharmaceutical and biomedical analysis - Quo vadis. *J. Pharmaceut. Biomed. Anal.* 2015. 113:97-107.

144. Seip, K. F., Gjelstad, A., Pedersen-Bjergaard, S. The potential of electromembrane extraction for bioanalytical applications. *Bioanal.* 2015. 7(4):463-480.
145. Kemperman, A. J. B., Bargeman, D., Van Den Boomgaard, T., Strathmann, H. Stability of supported liquid membranes: State of the art. *Sep. Sci. Tech.* 1996. 31(20):2733-2762.
146. See, H. H., Hauser, P. C. Electro-driven extraction of low levels of lipophilic organic anions and cations across plasticized cellulose triacetate membranes: Effect of the membrane composition. *J. Membr. Sci.* 2014. 450:147-152.
147. See, H. H., Stratz, S., Hauser, P. C. Electro-driven extraction across a polymer inclusion membrane in a flow-through cell. *J. Chromatogr. A.* 2013. 1300:79-84.
148. Mamat, N. A., See, H. H. Development and evaluation of electromembrane extraction across a hollow polymer inclusion membrane. *J. Chromatogr. A.* 2015. 1406:34-39.
149. Wang, W., Gu, C., Lynch, K., B. High-pressure open-channel on-chip electroosmotic pump for nanoflow high performance liquid chromatography. *Anal. Chem.* 2014. 86(4):1958-1964.
150. Gu, C., Jia, Z., Zhu, Z., et al.; Miniaturized electroosmotic pump capable of generating pressures of more than 1200 mbar. *Anal. Chem.* 2012. 84(21):9609-9614.
151. Buglione, L., See, H. H., Hauser, P. C. Study on the effects of electrolytes and solvents in the determination of quaternary ammonium ions by nonaqueous capillary electrophoresis with contactless conductivity detection. *Electrophoresis.* 2013. 34(2):317-323.
152. Nghiem, L. D., Mornane, P., Potter, I. D., Perera, J. M., Cattrall, R. W., Kolev, S. D. Extraction and transport of metal ions and small organic

compounds using polymer inclusion membranes (PIMs). *J. Membr. Sci.* 2006. 281(1-2):7-41.

153. De Gaetano, Y., Hubert, J., Mohamadou, A. Removal of pesticides from wastewater by ion pair Centrifugal Partition Extraction using betaine-derived ionic liquids as extractants. *Chem. Eng. J.* 2016. 285:596-604.