

CHEMICAL CONSTITUENTS AND BIOACTIVITIES OF *Artocarpus scortechinii*

KING. AND *A. hispidus* F.M. JARRETT

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Specially dedicated to my beloved family,

*My husband & my sons,
Mohd Hasrul Ishak*

*Muhammad Hadid Zikry Mohd Hasrul
Muhammad Zayd Zikry Mohd Hasrul*

*My parents and parents in law,
Mohd Arriffin Samsuddin & Zainun Wan Omar
Ishak Sulaiman & Hamidah Johar*

*My siblings and sister in law,
Norzafriza Mohd Arriffin
Mohammad Zafran Mohd Arriffin
Nor Azrita Mohd Arriffin
Nor Haslinda Ishak
Nor Hidayah Ishak*

*For their love, concern, encouragement and continuous prayer for my success in
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ABSTRACT

Two *Artocarpus* species from Malaysia, namely *A. scorchedinii* King and *A. hispidus* F.M. Jarrett were investigated. The aims of this study were to isolate and identify the phytochemicals from different parts of these two species and also to evaluate the biological activities of the crude extracts and isolated compounds using antioxidant, α -glucosidase and anti-tyrosinase inhibition assays. Cold extractions were performed using *n*-hexane, dichloromethane, ethyl acetate and methanol on the stem barks and leaves of *A. scorchedinii* to obtain the respective extracts. Extractions of *A. hispidus* were carried using *n*-hexane, dichloromethane and methanol. Fifteen compounds were isolated from these two *Artocarpus* species. Two new compounds together with thirteen known compounds were identified spectroscopically and by comparison with literature data. New compounds were isolated from the leaves of *A. scorchedinii* and identified as (2*R*,3*R*)-5-hydroxy-6,7-(2,2-dimethylpyrano)-2,3-dihydroflavonol and (2*R*,3*R*)-7-hydroxy-5-methoxy-8-prenyl-2,3-dihydroflavonol together with macakurzin C, flemichapparin A, β -sitosterol, apigenin and luteolin. 4',5-Dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8- γ,γ -dimethylallylflavone, stigmasterol, cudraflavone A, artocarpin, cycloartobiloxanthone, artonin E, oxyresveratrol and engelitin were obtained from the stem barks of *A. scorchedinii*. Purification of the extracts of *A. hispidus* afforded four known compounds and identified spectroscopically as 4',5-dihydroxy-6,7-(2,2-dimethylpyrano)-2'-methoxy-8- γ,γ -dimethylallylflavone, stigmasterol, artocarpin and oxyresveratrol. Extracts from these two *Artocarpus* species were screened for total phenolic contents using Folin Ciocalteau's reagent. Among all extracts tested, methanol extract of the stem barks of *A. hispidus* showed the highest phenolic content with TPC value of 218.93 mg GAE/g dw followed by methanol extract of the stem barks of *A. scorchedinii* with TPC value of 136.84 mg GAE/g dw. The methanol extract of the stem barks of *A. hispidus* also showed the highest scavenging action among all the extracts with an IC₅₀ value of 28.2 μ g/mL towards DPPH radical. Luteolin displayed the best radical scavenging activity compared to other isolated compounds towards DPPH and ABTS radical with IC₅₀ values of 9.51 μ g/mL and 124.4 μ g/mL, respectively. Luteolin also displayed the highest FRAP equivalent value of 4.07 \pm 0.18 mM, which may act as good reducing agent. In α -glucosidase inhibition assay, oxyresveratrol was noticeably having strongest inhibition with an IC₅₀ value of 3.88 μ M. The methanol extract of stem barks of *A. hispidus* displayed the highest percentage inhibition (88.2%) in anti-tyrosinase inhibition activity followed by methanol extract of *A. scorchedinii* (86.7%). Among the isolated compounds, oxyresveratrol exhibited the most potent tyrosinase inhibitory activity (92.3%) higher than the positive control, kojic acid (85.5%).

ABSTRAK

Dua spesies *Artocarpus* Malaysia, iaitu *A. scortechinii* King dan *A. hispidus* F.M. Jarrett telah dikaji. Tujuan kajian ini ialah untuk mengasingkan dan mengenalpasti fitokimia daripada bahagian berbeza dua spesies ini dan juga menilai aktiviti biologi ekstrak mentah dan sebatian terpencil menggunakan pengujian antioksidan, α -glukosidase dan perencatan anti-tirosinase. Pengekstrakan sejuk telah dilakukan menggunakan *n*-heksana, diklorometana, etil asetat, dan metanol ke atas kulit batang dan daun *A. scortechinii* untuk mendapatkan ekstrak masing-masing. Pengekstrakan *A. hispidus* telah dijalankan menggunakan *n*-heksana, diklorometana dan metanol. Lima belas sebatian telah dipencarkan daripada dua spesies *Artocarpus* tersebut. Dua sebatian baharu bersama-sama tiga belas sebatian yang diketahui telah dikenalpasti melalui spektroskopi dan perbandingan dengan data dari literatur. Sebatian baharu telah dipisahkan daripada daun *A. scortechinii* dan dikenalpasti sebagai (*2R,3R*)-5-hidroksi-6,7-(2,2-dimetilpirano)-2,3-dihidroflavonol dan (*2R,3R*)-7-hidroksi-5-metoksi-8-prenil-2,3-dihidroflavonol, bersama-sama dengan macakurzin C, flemichapparin A, β -sitosterol, apigenin dan luteolin. 4',5-Dihidroksi-6,7-(2,2-dimetilpirano)-2'-metoksi-8- γ,γ -dimetilallilflavon, stigmasterol, cudraflavon A, artokarpin, silkoartobiloxanton, artonin E, oksiresveratrol dan engelitin telah diperolehi daripada kulit batang *A. scortechinii*. Penulenan ekstrak *A. hispidus* telah menghasilkan empat sebatian yang diketahui dan dikenalpasti secara spektroskopi sebagai 4',5-dihidroksi-6,7-(2,2-dimetilpirano)-2'-metoksi-8- γ,γ -dimetilallilflavon, stigmasterol, artokarpin dan oksiresveratrol. Ekstrak daripada dua spesies *Artocarpus* tersebut telah disaring untuk jumlah kandungan fenolik menggunakan reagen Folin Ciocalteau. Antara ekstrak yang diuji, ekstrak metanol kulit batang *A. hispidus* menunjukkan kandungan fenolik yang tertinggi dengan nilai TPC 218.93 mg GAE/g dw diikuti oleh ekstrak metanol kulit batang *A. scortechinii* dengan nilai TPC 136.84 mg GAE/g dw. Ekstrak metanol kulit batang *A. hispidus* juga menunjukkan pemerangkapan tertinggi antara semua ekstrak dengan nilai IC₅₀ 28.2 μ g/mL terhadap radikal DPPH. Luteolin memaparkan aktiviti pemerangkapan radikal terbaik berbanding sebatian terpencil yang lain terhadap radikal DPPH dan ABTS dengan nilai IC₅₀ 9.51 μ M dan 124.4 μ M masing-masing. Luteolin juga memaparkan nilai setara FRAP yang tertinggi, iaitu 4.07 \pm 0.18 mM, yang mungkin boleh bertindak sebagai agen penurunan yang baik. Dalam pengujian rencatan α -glukosidase, oksiresveratrol didapati mempunyai perencatan tertinggi dengan IC₅₀ 3.88 μ M. Ekstrak metanol kulit batang *A. hispidus* memaparkan peratus perencatan tertinggi (88.2%), dalam aktiviti rencatan anti-tirosinase diikuti ekstrak metanol *A. scortechinii* (86.7%). Antara sebatian terpencil, oksiresveratrol mempamerkan potensi merencat aktiviti tirosinase tertinggi (92.3%), lebih tinggi daripada kawalan positif, asid kojik (85.5%).

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LIST OF ABBREVIATIONS

α	-	Alpha
β	-	Beta
δ	-	Chemical shift
δ_H	-	Chemical shift for proton
δ_C	-	Chemical shift for carbon
AA	-	Ascorbic acid
Abs	-	Absorbance
ABTS	-	2,2'-azinobis (3-ethyl-benzothiazoline-6-sulfonic acid)
APCIMS	-	Atmospheric Pressure Chemical Ionization Mass Spectrometry
ATR	-	Attenuated Total Reflection
BHA	-	Butylated hydroxyanisole
BHT	-	Butylated hydroxytoluene
Br	-	Broad
^{13}C NMR	-	Carbon Nuclear Magnetic Resonance
Calcd.	-	Calculated
CC	-	Column Chromatography
cm^{-1}	-	per centimeter
COSY	-	Correlation Spectroscopy
1D	-	1 Dimension
2D	-	2 Dimension
d	-	Doublet
dd	-	Doublet of Doublets
DEPT	-	Distortionless Enhancement by Polarization Transfer
DMSO	-	Dimethyl sulfoxide
DPPH	-	2,2'-Diphenyl-1-picrylhydrazyl
EIMS	-	Electron Impact Mass Spectrometry

EtOAc	-	Ethyl acetate
FRAP	-	Ferric reducing antioxidant potential
g	-	Gram
GC-MS	-	Gas Chromatography-Mass Spectrometry
¹ H NMR	-	Proton Nuclear Magnetic Resonance
HMBC	-	Heteronuclear Multiple Bond Correlation
HMQC	-	Heteronuclear Multiple Quantum Coherence
HRAPCIMS	-	High Resolution Atmospheric Pressure Chemical Ionization Mass Spectrometry
Hz	-	Hertz
I%	-	Percent Inhibition
IC ₅₀	-	Inhibition Concentration at 50%
int.	-	Intensity
IR	-	Infrared
<i>J</i>	-	Coupling constant
kg	-	Kilogram
lit.	-	Literature
L-DOPA	-	L-tyrosinase-3,4-dihydroxyl-L-phenylalanine
m	-	Multiplet
<i>m/z</i>	-	Mass to charge
M ⁺	-	Molecular ion
MHz	-	Megahertz
mg	-	Milligram
mL	-	Milliliter
mM	-	Milimolar
mp	-	Melting point
MS	-	Mass Spectrometry
µL	-	Microliter
µm	-	Micrometer
µM	-	Micromolar
NMR	-	Nuclear Magnetic Resonance
ppm	-	Parts per million
PTLC	-	Preparative Thin Layer Chromatography
Ref	-	Reference

rel.	-	Relative
R_f	-	Retention factor
s	-	Singlet
sh	-	Shoulder
t	-	Triplet
TLC	-	Thin Layer Chromatography
TPTZ	-	2,4,6-tripyridyl-s-triazine
Trolox	-	6-Hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid
UV	-	Ultraviolet
VLC	-	Vacuum Liquid Chromatography

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CHAPTER 1

INTRODUCTION

1.1 General Introduction

Plants had been used for medicinal purposes by local practitioners since thousands of years ago in whole world and still been used until today. Plants contain thousands valuable treasures that need to be explored. Plants can served as main source of natural product research in discovery of biologically active new drugs [1-3]. Chemical investigations can lead to new findings that can be used in the future by people to heal various illnesses. Nowadays people are exposing to various type of diseases such as HIV/AIDS, Alzheimer, malaria, and cancer which can threat life. Discovery of new medicines and antibiotics are important to treat these serious diseases. High demand on the new generation of antibiotics and medicines had led to more phytochemical study from natural plants. Moreover, about 80% of the population in the world according to The World Health Organization were using plants as traditional medicines [2].

Recently, people prefer using plants origin than chemicals as the source in drug discovery due to safety and less cost. Plants can be serving as important raw materials for production of new medicines because there are many evidences showed that the plant extracts may possess several useful biological activities including antibacterial, antitubercular, antiviral, antifungal, antiplatelet, anti-tyrosinase inhibitory and cytotoxic activities [4]. The discovery of the drug from plants also can depend on the bioassay

guided isolation methods to conduct the isolation of the important and desire drugs [5-6]. This method can lead to isolation of the active compounds.

To date, less than 10% of the world's biodiversity had been explored on the phytochemical and active compounds in plants. Many more discoveries await to be investigated and disinterred for the use of the future [7]. Plants have been used as source of drug discovery for thousands of years. Isolation of the active compounds from plant involves numerous fields and various methods of analysis. It begins with a botanist, ethnobotanist or plant ecologist who identifies the plant of interest. Phytochemist (Natural product chemist) will prepare the extract and conduct the process of isolation. The compounds will be elucidated and appropriate screening assay will be done on the target compounds. Pharmacologist will take further action for the drug discovery [8].

Malaysia is strategically located in the equator with the balance of hot and wet climate that make our country having wide range of plants species. Due to this reason, Malaysia's forests are rich with varieties of plant species that are valuable and suitable for the medicine discovery. Moraceae is one of the plant families that have potential to be explored due to widely used in various traditional medicines [4, 10-12].

1.2 Moraceae Family

Moraceae family is also known as the mulberry family. It comprises of 60 genera and over 1400 species distributed worldwide. Plants of this family are most abundant and ecologically important in tropical rainforest. These plants are usually trees, shrubs or climbers which produce white latex. Moraceae plants are closely related to Ulmaceae and Urticaceae plants except Moraceae plants have latex [9]. Three main genera in Moraceae family are *Ficus*, *Morus* and *Artocarpus*. Several of the genera produce timber and edible.

1.3 Genus *Artocarpus*

Genus *Artocarpus* consists of about 50 species which are widely distributed over tropical rain-forests. *Artocarpus* are known for their edible fruits such as *A. heterophyllus* (jackfruit), *A. altilis* (breadfruit) and *A. integer* (cempedak). This genus had been well known to contain rich sources of prenylated flavonoids, terpenoids, stilbenoids and xanthones [10-13]. More interestingly, they had been reported widely as traditional folk medicine for the treatment of malaria fever, liver cirrhosis, hypertension and diabetes [14-17]. Almost whole parts of this species had its own benefit on human for medicinal purposes. The root can cure asthma and fever, the seed can relieve the diarrhea, the wood act as sedative, the leaves act as antisyphilitic in human and relieve ulcer while the leaf ash can cure wounds. [9]. **Table 1.1** shows several *Artocarpus* species in Malaysia, their common names and the distributions.

Table 1.1: Several *Artocarpus* species in Malaysia

Species	Common Name	Distributions
<i>A. anisophyllus</i>	Keledang babi	Rare species in lowland forest such as Johor and Negeri Sembilan.
<i>A. altilis</i>	Breadfruit	Frequent in the village in Malaysia.
<i>A. chumpeden</i>	Cempedak	Cultivated throughout Malaysia.
<i>A. communis</i>	Sukun	Frequent in the village in Malaysia.
<i>A. dadah</i>	Tampang bulu	Lowland forest throughout Malaysia.
<i>A. elasticus</i>	Terap nasi	Lowland forest throughout Malaysia.
<i>A. fulvicortex</i>	Tampang gajah	Scattered throughout Malaysia in lowland forest.
<i>A. gomenzianus</i>	Tampang burung	Rare species in lowland forest such as Kedah, Kelantan, Pahang, Negeri Sembilan and Johor.
<i>A. heterophyllus</i>	Nangka	Cultivated throughout Malaysia.
<i>A. hispidus</i>	Temponek	Lowland forest such as Pulau Pinang, Perak, Pahang, Terengganu and Selangor.
<i>A. integer</i>	Cempedak	Cultivated throughout Malaysia.
<i>A. integer</i> var <i>silvestris</i>	Bangkong	Widely distributed in Malaysia from the lowland to mountain forest.
<i>A. kemando</i>	Pudu	Lowland forest including swamps such as Terengganu, Pahang and Selangor.
<i>A. lanceifolius</i>	Keledang	Lowland and hill forest throughout

		Malaysia.
<i>A. lowii</i>	Miku	Rare species in Malaysia and found in lowland forest.
<i>A. scorchedinii</i>	Terap hitam	Rare species in lowland forest in Malaysia.
<i>A. teysmanii</i>	-	Rare species in swampy habitat in Perak and Selangor.

1.3.1 *A. scorchedinii* King.

A. scorchedinii King. (**Figure 1.1**) is locally known as *terap hitam*. It can be found in lowland forest of Malaysia and Sumatra, Indonesia [9]. This species had been identified to be similar to *A. elasticus* (“*terap nasi*”). The difference with *A. elasticus* is on the stem which is darker while the leaves are not too large and wide.

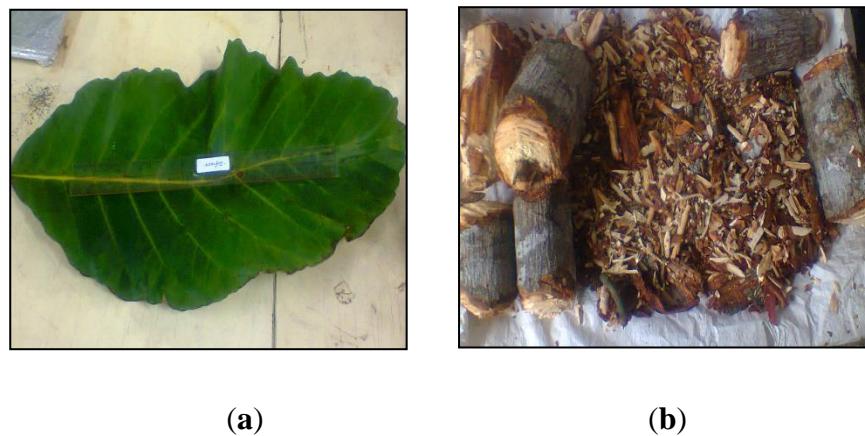


Figure 1.1: (a) Leaves and (b) stem barks of *A. scorchedinii* King.

1.3.2 *A. hispidus* F.M. Jarrett

A. hispidus F.M. Jarrett (**Figure 1.2**) is an evergreen tree commonly known as “temponek”. The tree can reach 20 m tall with the girth of 120 cm. This plant can be found in lowland forest of Pulau Pinang, Perak, Pahang, Terengganu and Selangor. This species had been identified to have similarity with *A. rigidus*. The leaves of *A. hispidus* are densely hairy with stalks about 1-2 cm long [8].

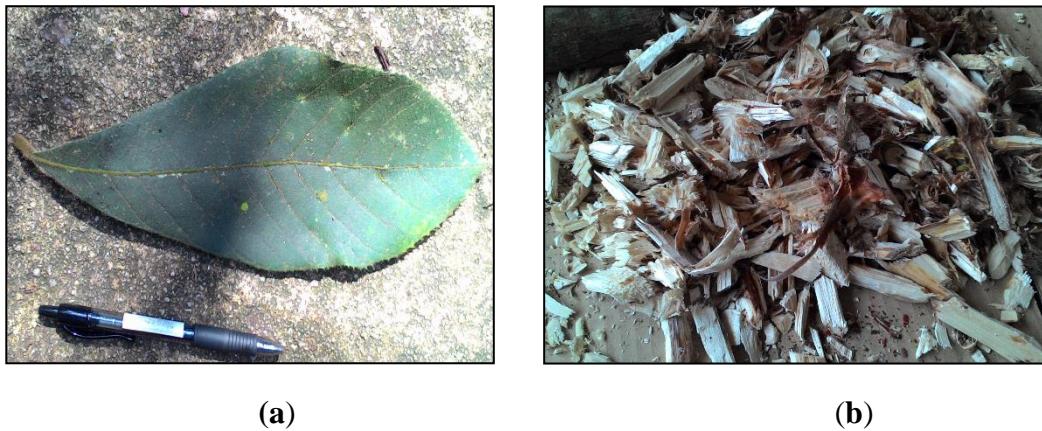


Figure 1.2: (a) Leaves and (b) stem barks of *A. hispidus* F.M. Jarrett

1.4 Problem Statements

Artocarpus species have been proven as rich source of phenolic compounds especially flavonoids based on many previous reports. However, not all species had been scientifically studied for their phytochemicals and bioactivities. Previous studies on *A. scortechinii* were only on the barks which were collected from different location in Malaysia [18-19]. Another study had been carried out by Indonesian researchers on the heartwood [20]. The leaves and stem barks of *A. scortechinii* had not been investigated yet. Thus, the investigation on the presence of secondary metabolites from this species is important since there is still insufficient information regarding the chemical constituents in leaves and stem barks part. The samples were collected from Bukit Fraser, Pahang. Different locality of the sample may result in the isolation of different chemical compounds and bioactivities. To date, no research had been done on *A. hispidus*. A thorough literature search did not find any report on the phytochemical and isolation work of *A. hispidus* thus far. Therefore, extensive studies are needed to provide the data of the chemical compounds present in these two rare species of *Artocarpus*.

1.5 Research Objectives

The objectives of this research are:

- i) To isolate the chemical constituents from the stem barks and leaves of *A. scortechinii* King. and *A. hispidus* F.M. Jarrett.
- ii) To elucidate the structure of pure compounds using spectroscopic methods (FTIR, UV, MS and NMR)
- iii) To evaluate the ability of the extracts and pure compounds on antioxidant, α -glucosidase and anti-tyrosinase inhibition activities.

1.6 Significant of Study

Plants had been well documented for their medical uses and treatments of numerous diseases. In recent years, the awareness of the researchers to explore on the natural based product is increasing. The aim was to give evidence and supported data on the medicinal uses originated from plants. Phytochemical screening and isolation of these two species (*A. scortechinii* and *A. hispidus*) could be used as alternative source of bioactive compounds for further research. The isolation of the pure bioactive compounds could also lead to scientific proof of their medicinal properties and give additional knowledge for pharmaceutical field.

1.7 Scope of Study

These studies focused on the phytochemicals and bioactivities of two Malaysia's rare *Artocarpus* species i.e. *A. scortechinii* and *A. hispidus*. The parts of the plant investigated are the stem barks and leaves. The dried samples were soak successively using solvents with different polarity to give the extracts. All extracts were fractionated

REFERENCES

1. Kumar, S., Paul, S., Walia, Y. K., Kumar A. and Singhal P. (2015). Therapeutic Potential of Medicinal Plants: A Review, *J. Biol. Chem. Chron.*, 1(1), 46-54.
2. Phukan, A., Borgohain, R., Chutia, P., Saikia, E., Kardong, D. and Chetia, B. (2014). Phytochemical and Antimicrobial Activity of Some Medicinal Plants of North East India. *Asian Journal of Chemistry*, 26(15), 4752-4754.
3. Kartz, L. and Baltz, R. H. (2016). Natural Product Discovery: Past, Present, and Future. *Journal of Industrial Microbiology & Biotechnology*, 43, 155-176.
4. Jagtap, U. B. and Bapat, V. A. (2010). *Artocarpus*: A Review of its Traditional Uses, Phytochemistry and Pharmacology. *Journal of Ethnopharmacology*, 129, 142-166.
5. Lahlou, M. (2013). The Success of Natural Products in Drug Discovery. *Pharmacology & Pharmacy*, 4, 17-31.
6. Jantan, I. (2004). Medicinal Plant Research in Malaysia: Scientific Interests and Advances. *Jurnal Sains Kesihatan Malaysia*, 2(2), 27-46.
7. Dias, D. A., Urban, S. and Roessner. (2012). A Historical Overview of Natural Products in Drug Discovery, *Metabolites*, 2, 303-336.
8. Balunas, M. J. and Kinghorn, A. D. (2005). Drug Discovery from Medicinal plants. *Life Sciences*, 78, 431-441.
9. Kochummen, K.M. (1978). *Tree Flora of Malaya* (Volume 3). Kuala Lumpur, 119-134.
10. Baliga, M. S., Shivashankara, A. R., Haniadka, R., Dsouza, J. and Bhat, H. P. (2011). Phytochemistry, Nutritional and Pharmacological Properties of *Artocarpus heterophyllus* Lam (Jackfruit): A Review. *Food Research International*, 44, 1800-1811.

11. Hakim, A. (2010). Diversity of Secondary Metabolites from Genus *Artocarpus* (Moraceae). *Nusantara Bioscience*, 2(3), 146-156.
12. Sikarwar, M. S., Hui, B. J., Subramaniam, K., Valeisamy, B. D., Yean, L. K. and Balaji K. (2014). A Review on *Artocarpus altilis* (Parkinson) Fosberg (Breadfruit). *Journal of Applied Pharmaceutical Science*, 4, 091-097.
13. Jamil, S, Lathiff, S. M. A., Abdullah, S. A., Jemaon, N. and Sirat, H. M. (2014). Antimicrobial Flavonoids from *Artocarpus anisophyllus* Miq. and *Artocarpus lowii* King. *Jurnal Teknologi*, 71(1), 95-99.
14. Lan, W. C., Tzeng, C. W., Lin, C. C., Yen, F. L and Ko, H. H. (2013). Prenylated Flavonoids from *Artocarpus altilis*: Antioxidant Activities and Inhibitory Effects on Melanin Production. *Phytochemistry*, 89, 78-88.
15. Shahin, N., Kazmi, I and Ali, M. (2012). Glycosidase from the Leaves of *Artocarpus heterophyllus* Lam. *International Journal of Pharmacy and Pharmaceutical Sciences*, 4(3), 449-453.
16. Ren, G., Hu, Z. C., Xiang, H. Y., Peng, J. B., Liu, R. H., Huang, H. L. and Shao, F. (2013). Chemical Constituents from the Fruiting Branches of *Artocarpus nanchuanensis* Endemic to China. *Biochemical Systematics and Ecology*, 51, 98-100.
17. Arung, E. T., Yoshikawa, K., Shimizu, K. and Kondo, R. (2010). Isoprenoid-substituted Flavonoids from Wood of *Artocarpus heterophyllus* on B16 Melanoma Cells: Cytotoxicity and Structural Criteria. *Fitoterapia*, 81, 120-123.
18. Jamil, S., Sirat, H. M., Aimi, N. and Kitajima, M. (2004). Flavones from *Artocarpus scorchedinii* King. *ACGC Chemical Research Communications*, 17, 3-8.
19. Ali, A. H., Hassan, N. M., Shukor, N. I., Embi, N., Latip, J. and Sidek, M. H. M. (2014). Anti-Plasmodial Activity of Engelitin Isolated *Artocarpus scorchedinii*. *Malays. Appl. Biol.*, 43(1), 73-80.
20. Hakim, A. (2009). A Prenylated Flavonoids Heartwood of *Artocarpus scorchedinii* King (Moraceae). *Indo J. Chem.*, 9(1), 146-150.
21. Hakim, E. H., Achmad, S. A., Juliawaty, L. D., Makmur, L., Syah Y. M., Aimi, N., Kitajima, M., Takayama, H. and Ghisalberti, E. L. (2006). Prenylated Flavonoids and Related Compounds of the Indonesian *Artocarpus* (Moraceae). *Journal Natural Medicines*, 60, 161-184.
22. Gang, R., Bing, P. J., Fang, Y. W. and Jun, Y. W. (2014). Advance of Chemical Constituents from *Artocarpus* Plant and Its Bioactivities in Recent Five Years. *Chinese Journal of Experimental Traditional Medical Formulae*. 21.
23. Kumar, S. and Pandey, A. K. (2013). Chemistry and Biological Activities of Flavonoids: An Overview. *The Scientific World Journal*, 1-16.

24. Agrawal, A. D. (2011). Pharmacological Activities of Flavonoids: A Review. *International Journal of Pharmaceutical Sciences and Nanotechnology*, 4(2), 1394-1398.
25. Sandhar, H. K., Kumar, B., Prasher, S., Tiwari, P., Salhan, M. and Sharma, P. (2011). A Review of Phytochemistry and Pharmacology of Flavonoids. *Internationale Pharmaceutica Sciencia*, 1(1), 25-41.
26. Lathiff, S. M., Jemaon, N., Abdullah, S. A. and Jamil, S. (2015). Flavonoids from *Artocarpus anisophyllus* and Their Bioactivities. *Natural Product Communications*, 10(3), 393-396.
27. Amarasinghe, N. R., Jayasinghe, L., Hara, N., and Fujimoto, Y. (2008). Chemical Constituents of the Fruits of *Artocarpus altilis*. *Biochem. Syst. Ecol.*, 36, 323-325.
28. Syah, Y. M., Achmad, S. A., Ghisalberti, E. L., Hakim, E. H., Lukman, M., and Mujahidin, D. (2002). Artoindonesianins Q-T, Four Isoprenylated Flavones from *Artocarpus champeden* Spreng. (Moraceae). *Phytochemistry*, 61, 949-953.
29. Weng, J. R., Chan, S. C., Lu, Y. H., Lin, H. C., Ko, H. H., and Lin, C. N. (2006). Antiplatelet Prenylflavonoids from *Artocarpus communis*. *Phytochemistry*, 67, 824-829.
30. Suhartati, T., Yandri, Suwandi, J. F. and Hadi, S. (2013). Two Flavan Derivatives Isolated from *Artocarpus dadah* Grown in Lampung, Indonesia. *Asian Journal of Chemistry*, 25(2), 1050-1056.
31. Musthapa, I., Juliawaty, L.D., Syah, Y. M., Hakim, E. H., Latip, J. and Ghisalberti, E.L. (2009). An oxepinoflavone from *Artocarpus elasticus* with Cytotoxic Activity Against P-388 Cells. *Archives of Pharmacal Research*, 32(2), 191-194.
32. Soekamto, N. H., Achmad S. A., Ghisalberti, E. L., Hakim, E. H. and Syah, Y. M. (2003). Artoindonesianins X and Y, Two Isoprenylated 2-arylbenzofurans, from *Artocarpus frettessi* (Moraceae). *Phytochemistry*, 64, 831-834.
33. Jamil, S., Taher, M., Sirat. H. M., and Othman N. A. (2012). Flavonoids and Triterpenes from the Leaves of *Artocarpus fulvicortex*. *Natural Product Communication*, 7, 1587-1588.
34. Likhitwitayawuid, K., Chaiwiriya, S., Sritularak, B. and Lipipun, V. (2006). Antiherpetic Flavones from the Heartwood of *Artocarpus gomezianus*. *Chemistry & Biodiversity*, 3, 1138-1143.

35. Ren, G., Peng, J., Liu, A., Liang, J., Yuan, W., Wanga, H. and Hea, J. (2015). Structure Elucidation and NMR Assignments of Two New Flavanones from the Roots of *Artocarpus heterophyllus*. *Magnetic Resonance Chemistry*, 53, 872-874.
36. Shimizu, K., Kondo, R., Sakai, K., Buabarn, S. and Dilokkunanant U. (2000). A Geranylated Chalcone with 5 α -reductase Inhibitory Properties from *Artocarpus incisus*. *Phytochemistry*, 54, 737-739.
37. Shah, M. K. K., Sirat, H. M., Jamil, S. and Jalil, J. (2016). Flavonoids from the Bark of *Artocarpus integer* var. *silvestris* and Their Anti-inflammatory Properties. *Natural Product Communications*, 11(9), 1275-1278.
38. Hashim, N. M., Rahmani, M., Shamaun, S. S., Ee, G. C. L., Sukari, M. A., Ali, M. and Go, R. (2011). Dipeptide and Xanthones from *Artocarpus kemando* Miq. *Journal of Medicinal Plants Research*, 5(17), 4224-4230.
39. Syah, Y. M., Achmad, S. A., Aimi, N., Hakim, E. H., Juliawaty, L.D., and Takayama, H. (2006). Two Prenylated Flavones from the Tree Bark of *Artocarpus lanceifolius*, Z. *Naturforsch*, 61b, 1134-1137.
40. Maneechai, S., Eknamkul, W. D., Umehara, K., Noguchi, H. and Likhitwitayawuid, K. (2012). Flavonoid and Stilbenoid Production in Callus Cultures of *Artocarpus lakoocha*. *Phytochemistry*, 81, 42-49.
41. Jamil, S., Sirat, H. M., Jantan, I., Aimi, N. and Kitajima, M. (2008). A New Prenylated Dihydrochalcone from the Leaves of *Artocarpus lowii*. *J. Nat. Med.*, 62, 321-324.
42. Zhang, P. Z., Gu, J. and Zhang, G. L. (2015). Novel Stilbenes from *Artocarpus nanchuanensis*. *Journal of Asian Natural Products Research*, 17(3), 217-223.
43. Jayasinghe, U. L. B., Samarakoon, T. B., Kumarihamy, B. M. M., Hara, N. and Fujimoto, Y. (2008). Four New Prenylated Flavonoids and Xanthones from the Root Bark of *Artocarpus nobilis*. *Fitoterapia*, 79, 37-41.
44. Hashim, N., Rahmani, M., Sukari, M. A., Ali, A. M., Alitheen, N., Go, R. and Ismail, H. B. M. (2010). Two New Xanthones from *Artocarpus obtusus*. *Journal of Asian Natural Products Research*, 12(2), 106-112.
45. Arriffin, N. M., Jamil, S. and Basar, N. (2015). Antioxidant Activities of Extracts from the Leaves and Stem Barks of *Artocarpus scorchedinii* King. *Jurnal Teknologi*, 77(2), 1-5.

46. Ren, G., Xiang, H., Hu, Z., Liu, R., Zhou, Z., Huang, H., Shao, F. and Yang, M. (2013). A New Isoprenylated Flavone from the Root Bark of *Artocarpus styracifolius*. *Biochemical Systematics and Ecology*, 46, 97-100.
47. Jamil, S., Sirat, H. M. Jantan, I., Aimi, N. and Kitajima, M. (2005). Flavonids from *Artocarpus teysmanii* Miq. *Malaysian Journal of Science*, 24, 99-103.
48. Zheng, Z. P., Cheng, K. W., To, J. T. K., Li, H. and Wang, M. (2009). Isolation of Tyrosinase Inhibitors from *Artocarpus heterophyllus* and Use Its Extract as Antibrowning Agent. *Molecular Nutrition & Food Research*, 52, 1530-1538.
49. Syah, Y. M., Achmad, S. A., Ghisalberti, E. L., Hakim, E. H., Lukman, M., and Mujahidin, D. (2004). Two New Cytotoxic Isoprenylated Flavones, Artoindonesianins U and V, from the Heartwood of *Artocarpus champeden*. *Fitoterapia*, 75, 134-140.
50. Jamil. S., Abdullah, S. A., Lathiff, S. M. and Sirat, H. M. (2014). Tyrosinase Inhibitory Activity of Flavonoids from *Artocarpus lowii* King. *Jurnal Teknologi*, 71(1), 55-58.
51. Abdullah, S. A., Jamil. S., Basar, N., Lathiff, S. M. and Arriffin, N. M. (2017). Flavonoids from the Leaves and Heartwood of *Artocarpus lowii* King and Their Bioactivities. *Natural Product Research*, 31(10), 1113-1120.
52. Arriffin, N. M., Jamil. S., Basar, N., Khamis, S., Abdullah, S. A. and Lathiff, S. M. (2017). Phytochemical Studies and Antioxidant Activities of *Artocarpus scortechinii* King. *Record of Natural Product*, 11(3), 299-303.
53. Arung, E. T., Yoshikawa, K., Shimizu, K. and Kondo, R. (2010). Isoprenoid-Substituted Flavonoids from Wood of *Artocarpus heterophyllus* on B16 Melanoma Cells: Cytotoxic and Structural Criteria. *Fitoterapia*, 81, 120-123.
54. Arung, E. T., Shimizu, K., Tanaka, H. and Kondo, R. (2010). 3-Prenyl Luteolin, A New Prenylated Flavone with Melanin Biosynthesis Inhibitory Activity from Wood of *Artocarpus heterophyllus*. *Fitoterapia*, 81, 640-643.
55. Wei, B. L., Weng, J. R., Chiu, P. H., Hung, C. F., Wang, J. P. and Lin, C. N. (2005). Antiinflammatory Flavonoids from *Artocarpus heterophyllus* and *Artocarpus communis*. *J. Agric Food Chem.*, 53(10), 3867-3871.

56. Ee, G. C. L., Teo, S. H., Rahmani, M., Lim, C. K., Lim, Y. M and Go, R. (2011). Artomandin, A New Xanthone from *Artocarpus kemando* (Moraceae). *Natural Product Research*, 25(10), 995-1003.
57. Di, X., Wang, S., Wang, B., Liu, Y., Yuan, H., Lou, H. and Wang, X. (2013). New Phenolic Compounds from the Twigs of *Artocarpus heterophyllus*. *Drug Discoveries & Therapeutics*, 7(1), 24-28.
58. Ramli, F., Rahmani, M., Kassim, N. K., Hashim, N. M., Sukari, M. A., Akim, A. M. and Go, R. (2013). New Diprenylated Dihydrochalcone from Leaves of *Artocarpus elasticus*. *Phytochemistry Letters*, 6, 582-585.
59. Daus, M., Chaithada, P., Phongpaichit, S., Watanapokasin, R., Carroll, A. R. and Mahabusarakam, W. (2017). New Prenylated Dihydrochalcone from the Leaves of *Artocarpus elasticus*. *Phytochemistry Letters*, 19, 226-230.
60. Jin, J. J., Lin, C. C., Lu, T. M., Li, J. H., Chen, I. S., Kuo, Y. H. and Ko, H. H. (2015). Chemical Constituents Derived from *Artocarpus xanthocarpus* as Inhibitors of Melanin Biosynthesis. *Phytochemistry*, 117, 424-435.
61. Shah, M. K. K., Sirat, H. M. and Jamil, S. (2016). Cholinesterase Inhibitors from Heartwood of *Artocarpus fulvicortex* F. M. Jarret (Moraceae). *Jurnal Teknologi*, 78(6), 185-189.
62. Shen, H. and Hou, A. J. (2008). Prenylated 2-arybenzofurans from *Artocarpus petelotii*. *Natural Product Research*, 22(16), 1451-1456.
63. Likhithwitayawuid, K., Sornsu, A., Sritularak, B. and Ploypradith, P. (2006). Chemical Transformations of Oxyreveratrol into a Potent Tyrosinase Inhibitor and Strong Cytotoxic Agent. *Bioorganic & Medicine Letter*, 16, 5650-5653.
64. Boonlaksiri, C., Oonanant, W., Kongsaeree, P., Kittakoop, P., Tantcharoen, M. and Thebtaranonth, Y. (2000). An Antimalarial Stilbene from *Artocarpus integer*. *Phytochemistry*, 54, 415-417.
65. Tsai, P. W., Cruz, K. A. D. C., Shen, C. C. and Chiou, C. T. (2013). Chemical Constituents of *Artocarpus camansi*. *Pharmacognosy Journal*, 5, 80-82.
66. Ragasa, C. Y., Caro, J. L. and Shen, C. C. (2014). Triterpenes and Sterol from *Artocarpus ovatus*. *Journal of Applied Pharmaceutical Science*, 4(10), 007-011.
67. Ko, F. N., Cheng, Z. J., Lin, C. N. and Teng, C. M. (1998). Scavenger and Antioxidant Properties of Prenylflavones Isolated from *Artocarpus Heterophyllus*. *Free Radical Biology & Medicine*, 25(2), 160-168.
68. Hashim, N. M., Rahmani, M., Ee, G. C. L., Sukari, M. A., Yahayu, M., Amin, M. A. M., Ali, A. M., and Go, R. (2012). Antioxidant, Antimicrobial and Tyrosinase Inhibitory Activities of Xanthones Isolated from *Artocarpus obtusus* F. M. Jarret, *Molecules*, 17, 6071-6082.

69. Kamboj, A. and Salujai, A. K. (2011). Isolation of Stigmasterol and β -Sitosterol from Petroleum Ether Extract of Aerial Parts of *Ageratum conyzoides* (Asteraceae). *International Journal of Pharmacy and Pharmaceutical Sciences*, 3(1), 94-96.
70. Omar, M. N., Muzammil, A., Khan, N. T., Zahid, M. and Kamal, T. (2013). Isolation and Identification of Bioactive Compounds from Twigs of *Artocarpus altilis*. *Merit Research Journal*, 1(8), 147-155.
71. Musthapa, I., Hakim, E. H., Syah, Y. M. and Juliawaty, L. D. (2016). Cytotoxic Activities of Prenylated Flavonoids from *Artocarpus heterophyllus*. *ARPN Journal of Engineering and Applied Sciences*, 11(16), 9754-9758.
72. Lin, K. W., Liu, C. H., Tu, H. Y., Ko, H. H. and Wei, B. L. (2009). Antioxidant Prenylflavonoids from *Artocarpus communis* and *Artocarpus elasticus*. *Food Chemistry*, 115, 558-562.
73. Povichit, N., Phrutivorapongkul, A., Suttajit, M. and Leelapornpisid, P. (2010). Antiglycation and Antioxidant Activities of Oxyresveratrol Extracted from the Heartwood of *Artocarpus lakoocha* Roxb. *Maejo International Journal of Science and Technology*, 4(03), 454-461.
74. Markham K. R. (1982) *Techniques of flavonoids Identification*. London: Academic Press Inc, 36-49.
75. Shi, H., Liua, M., Wang, R., Gaoa, B., Zhang, Z., Niua, Y., and Yu, L. (2015). Separating Four Diastereomeric Pairs of Dihydroflavonol Glycosides from *Engelhardia roxburghiana* Using High Performance Counter-current Chromatography. *Journal of Chromatography A*, 1383, 79-87.
76. Thanh, V. T. T., Mai, H. D. T., Pham, V. C., Litaudon, M., Dumontet, V., Gueritte, F., Nguyen, V. H. and Chau, V. M. (2012). Acetylcholinesterase Inhibitors from the Leaves of *Macaranga kurzii*. *Journal of Natural Product*, 75, 2012-2015.
77. Moghaddam, F. M., Farimani, M. M., Salahvarzi, S. and Amin, G. (2007). Chemical Constituents of Dichloromethane Extract of Cultivated *Satureja khuzistanica*. *Advance Access Publication*, 4(1), 95-98.
78. Ragasa, C. Y., Ng, V. A., Park, J. H., Kim, D. W., Cornelio, K. and Shen, C. C. (2014). Chemical Constituents of *Artocarpus altilis* and *Artocarpus odoratissimus*. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 5(4), 1081-1087.
79. Awantu, A. F., Lenta, B. N., Donfack, E. V., Wansi, J. D., Neumann, B., Stammler, H., Noungoue, D. T., Tsamo, E. and Sewald, N. (2011). Flavonoids and Other Constituents of *Hymenostegia afzelii* (Caesalpiniaceae). *Phytochemistry Letters*, 4, 315–319.
80. Tajuddeen, N., Sallaua, M. S., Musab, A. M., Yahayab, S. M., Habilaa, J. D. and Ismail, A. M. (2016). A Novel Antimicrobial Flavonoid from the Stem Bark of

- Commiphora pedunculata* (Kotschy & Peyr.) Engl. *Natural Product Research*, 30(10), 1109–1115.
81. Argaiza, R. B., Diaz, M. E. P., Waterman, P. G. and Rodriguez, L. M. P. (2005). Additional Flavonoids from *Lonchocarpus yucatanensis* and *L. xuul*. *J. Braz. Chem. Soc.*, 16(5), 1078-1081.
82. Lee, Y. R. and Xia, L. (2007). An Efficient and Concise Synthesis of Biologically Interesting Natural Flemichapparin A, Flemingin A, Flemingin D, and Their Non-natural Analogues, *Bull. Korean Chem. Soc.*, 28(9), 1579-1584.
83. Bhattacharyya, J., Majetich, G., Spearing, P. and Almeida, R. N. Dioclenol, (1997). A Minor flavanone from the Root-Bark of *Dioclea grandiflora*. *Phytochemistry*, 46(2), 385-387.
84. Capistrano, I. R., Wouters, A., Foubert, K., Balde, A. M., Sandra Apers, S., Lardon, F., Pieters, L. and Exarchou, V. (2015). Phytochemical Characterisation of a Cytotoxic Stem Bark Extract of *Steganotaenia araliacea* and Identification of a Protoflavanone by LC–SPE–NMR. *Phytochemistry Letters*, 12, 119–124.
85. Karim, A., Amani, H., Hafez, M. A. and Kalid, M. (2016). Isolation, Characterization and Biological Activity of a Dihydroflavonol from *Tamarix nilotica* (Tamaricaceae) Leaves. *International Journal of Advanced Research*, 4(6), 2048-2056.
86. Noro, T., Oda, Y., Miyase, T., Ueno, A. and Fukushima, S. (1983). Inhibitors of Xanthine Oxidase from the Flowers and Buds of *Daphne genkwa*. *Chem. Pharm. Bull.*, 31(11), 3984-3987.
87. Alwahsh, M. A. A., Khairuddean, M. and Chong, W. K. (2015). Chemical Constituents and Antioxidant Activity of *Teucrium barbeyanum* Aschers, *Record of Natural Products*, 9(1), 159-163.
88. Lin, L. C., Pai, Y. F. and Tsai, T. H. (2015). Isolation of Luteolin and Luteolin-7-O-glucoside from *Dendranthema morifolium* Ramat Tzvel and Their Pharmacokinetics in Rats. *Journal of Agricultural and Food Chemistry*, 63(35), 7700–7706.
89. Bhoyar, M. S., Mishra, G. P., Naik, P. K. and Srivastava, R. B. (2011). Estimation of Antioxidant Activity and Total Phenolics Among Natural

- Populations of Caper (*Capparis spinosa*) Leaves Collected from Cold Arid Desert of Trans-Himalayas. *Australian Journal of Crop Sciences*, 5(7), 912-919.
90. Shafiq, M., Mehmood, S., Yasmeen, A., Khan, S. J., Khan, N. H. and Ali, S. (2017). Evaluation of Phytochemical, Nutritional and Antioxidant Activity of Indigenously Grown Jackfruit (*Artocarpus heterophyllus* Lam). *J. Sci. Res.*, 9(1), 135-143.
91. Lee, S. Y., Mediani, A., Nur, A. A. H., Azliana, A. B. S. and Abas, F. (2014). Antioxidant and α -Glucosidase Inhibitory Activities of the Leaf and Stem of Selected Traditional Medicinal Plants. *International Food Research Journal*, 21(1), 165-172.
92. Moukette, B. M., Pieme, C. A., Biapa, P. C. N., Njimou, J. R., Moor, V. J. A., Stoller, M., Bravi, M. and Ngogang, J. Y. (2014). Phenolic Content of *Hypodaphnis Zenkeri* and Its Antioxidant Effects against Fenton Reactions' Mediated Oxidative Injuries on Liver Homogenate. *Antioxidants*, 3, 866-889.
93. Saha, R. K., Jamiruddin, M., Acharya, S. and Roy, P. (2013). Phytochemical, Antioxidant, Antimicrobial and Receptor Binding Activities of the Methanolic Extract from the Testa of *Artocarpus heterophyllus* Lam. *Archives*, 2, 128-140.
94. Tirzitis, G. and Bartosz, G. (2010). Determination of Antiradical and Antioxidant Activity: Basic Principles and New Insights. *Acta Biochimica Polonica*, 57, 139-142.
95. Pour, B. M., Jothy, S. L., Latha, L. Y., Chen, Y. and Sasidharan, S. (2012). Antioxidant Activity of Methanol Extracts of Different Parts of *Lantana camara*. *Asian Pacific Journal of Tropical Biomedicine*, 2(12), 960-965.
96. Pisoschi, A. M. and Negulescu, G. P. (2011). Methods for Total Antioxidant Activity Determination: A Review. *Biochemistry & Analytical Biochemistry*, 1-10.
97. Faujan, N. H., Rahim, Z. A., Rehan, M. M. and Ahmad, F. (2015). Comparative Analysis Phenolic Content and Antioxidative Activities of Eight Malaysian Traditional Vegetables. *Malaysian Journal of Analytical Sciences*, 19(3), 611-624.
98. Butsat, S. and Siriamornpun, S. (2016). Effect of Solvent Types and Extraction Times on Phenolic and Flavonoid Contents and Antioxidant Activity in Leaf Extracts of *Amomum chinense* C. *International Food Research Journal*, 23(1), 180-187.
99. Babbar, N., Oberoi, H. S., Sandhu, S. K. and Bhargav, V. K. (2014). Influence of Different Solvents in Extraction of Phenolic Compounds from Vegetable Residues and Their Evaluation as Natural Sources of Antioxidants. *J. Food Sci. Technol.*, 51(10), 2568-2575.

100. Alberti, A., Zielinski, A. A. F., Zardo, D. M., Demiate, I. M., Nogueira, A. and Mafra, L. I. (2014). Optimisation of the Extraction of Phenolic Compounds from Apples Using Response Surface Methodology. *Food Chemistry*, 149, 151-158.
101. Okawa, M., Kinjo, J., Nohara, T. and Ono, M. (2001). DPPH (1,1-Diphenyl-2-Picrylhydrazyl) Radical Scavenging Activity of Flavonoids Obtained from Some Medicinal Plants. *Biol. Pharm. Bull.*, 24(10), 1202-1205.
102. Jothy, S. L., Zuraini, Z. and Sasidharan, S. (2011). Phytochemicals Screening, DPPH Free Radical Scavenging and Xanthine Oxidase Inhibitory Activities of *Cassia fistula* Seeds Extract. *Journal of Medicinal Plants Research*, 5(10), 1941-1947.
103. Liang, N. and Kitts, D. D. (2014). Antioxidant Property of Coffee Components: Assessment of Methods that Define Mechanisms of Action. *Molecules*, 19, 19180-19208.
104. Blois, M. S. (1958). Antioxidant Determinations by the Use of a Stable Free Radical. *Nature*, 181, 1199-1200.
105. Shalaby, E. A. and Shanab, S. M. M. (2013). Comparison of DPPH and ABTS Assays for Determining Antioxidant Potential of Water and Methanol Extracts of *Spirulina platensis*. *Indian Journal of Geo-Marine Sciences*, 42(5), 556-564.
106. Rebaya, A., Belghith, S. I., Baghdikian, B., Leddet, V. M., Mabrouki, F., Olivier, E., Cherif, J. K. and Ayadi, M. T. (2014). Total Phenolic, Total Flavonoid, Tannin Content, and Antioxidant Capacity of *Halimium halimifolium* (Cistaceae). *Journal of Applied Pharmaceutical Science*, 5(01), 052-057.
107. Biskup, I., Golonka, I., Gamian, A. and Sroka, Z. (2013). Antioxidant Activity of Selected Phenols Estimated by ABTS and FRAP methods, *Postepy Hig Med Dosw.*, 67, 958-963.
108. Pawlak, K., Bylka, W., Jazurek, B., Matlawska, I., Sikorska, M., Manikowski, H. and Byla, G. B. (2010). Antioxidant Activity of Flavonoids of Different Polarity, Assayed by Modified ABTS Cation Radical Decolorization and Technique. *Acta biologica Cracoviensia Series Botanica*, 52(1), 97-104.

109. Philips, A., Philip, S., Arul, V., Padmakeerthiga, B., Renju, V., Santha, S. and Sethupathy, S. (2010). Free Radical Scavenging Activity of Leaf Extracts of *Indigofera Aspalathoides* - An in vitro Analysis, *J. Pharm. Sci. & Res.*, 2(6), 322-328.
110. Lou, S. N. and Ho, C. T. (2017). Phenolic Compounds and Biological Activities of Small-size Citrus: Kumquat and Calamondin. *Journal of Food and Drug Analysis*, 25, 162-175
111. Prior, R. L., Wu, X. and Schaich, K. (2005). Standardized Methods for the Determination of Antioxidant Capacity and Phenolics in Foods and Dietary Supplements. *J. Agric. Food Chem.*, 53, 4290–4302.
112. Assanga, S. B. I., Lujan, L. M. L., Espinoza, C. L. L., Salido, A. A. G., Angulo, D. F., Pino, J. L. R. and Haines, D. D. (2015). Solvent Effects on Phytochemical Constituent Profiles and Antioxidant Activities, using Four Different Extraction Formulations for Analysis of *Bucida buceras* L. and *Phoradendron californicum*. *BMC Res Notes*, 8, 1-14.
113. Proenca, C., Freitas, M., Ribeiro, D., Oliveira, E. F. T., Sousa, J. L. C., Tome, S. M., Ramos, M. J., Silva, A. M. S., Fernandes, P. A. and Fernandes, E. (2017). α -Glucosidase Inhibition by Flavonoids: An In Vitro and In Silico Structure-Activity Relationship Study. *Journal of Enzyme Inhibition and Medicinal Chemistry*, 32(1), 1216-1228.
114. Asgharia, B., Salehib, P., Sonbolic, A. and Ebrahimib, S. N. (2015). Flavonoids from *Salvia chloroleuca* with α -Amylsae and α -Glucosidase Inhibitory Effect. *Iranian Journal of Pharmaceutical Research*, 14(2), 609-615.
115. Gu, C., Zhang, H., Putri, C. Y. and Ng, K. (2015). Evaluation of α -Amylase and α -Glucosidase Inhibitory Activity of Flavonoids. *International Journal of Food and Nutritional Science*, 2(6), 1-6.
116. Elya, B., Malik, A., SeptiMahanani, P. I. and Loranza, B. (2012). Antidiabetic Activity Test by Inhibition of α -Glucosidase and Phytochemical Screening from the Most Active Fraction of Buni (*Antidesma bunius* L.) Stem Barks and Leaves. *Int. J. PharmTech. Res.*, 4(4), 1667-1671.

117. Nguyen, H. X., Nguyen, N. T., Nguyen, M. H. K., Le, T. H., Do, T. N. V., Hung, T. M. and Nguyen, M. T. T. (2016). Tyrosinase Inhibitory Activity of Flavonoids from *Artocarpus heterophyllous*. *Chemistry Central Journal* 10(2), 1-6.
118. Likhitwitayawuid, K. (2008). Stilbenes with Tyrosinase Inhibitory Activity. *Current Science*, 94(1), 44-52.
119. Lai, J. S., Lin, C. C. and Chiang, T. M. (2014). Tyrosinase Inhibitory Activity and Thermostability of the Flavonoid Complex from *Sophora japonica L* (Fabaceae). *Tropical Journal of Pharmaceutical Research*, 13(2), 243-247.
120. Kim, S. J., Son, K. H., Chang, H. W., Kang, S. S. and Kim, H. P. (2003). Tyrosinase Inhibitory Prenylated Flavonoids from *Sophora flavescens*. *Biol. Pharm. Bull.*, 26(9) 1348-1350.
121. Kassim, N. K., Rahmani, M., Ismail, A., Sukari, M. A., Ee, G. C. L., Nasir, N. M. and Awang, K. (2013). Antioxidant Activity-Guided Separation of Coumarins and Lignan from *Melicope glabra* (Rutaceae). *Food Chem*, 139, 87-92.
122. Fu, R., Zhang, Y., Guo, Y., Liu, F. and Chen, F. (2014). Determination of Phenolic Contents and Antioxidant Activities of Extracts of *Jatropha curcas L.* Seed Shell, A By-Product, A New Source of Natural Antioxidant. *Industrial Crops and Products*, 58, 265-270.
123. Zou, Y., Chang, S. K. C., Gu, Y. and Qian, S. Y. (2011). Antioxidant Activity and Phenolic Composition of Lentils (*Lens culinaris* var. Morton) Extracts and its Fractions. *Journal of Agriculture Food Chemistry*, 59(6), 2268-2276.
124. Channarong, S., Jutiviboonsuk, A. and Korsanan, S. (2012). Total Reducing Antioxidant Capacity of Thai Herbal Aromatic Powder (Ya Hom) Measured by FRAP Assay. *Thai Pharmaceutical and Health Science Journal*, 7(3), 111-114.
125. Shahwar, D., Raza, M. A., Bukhari, S. and Bukhari, G. (2012). Ferric Reducing Antioxidant Power of Essential Oils Extracted from *Eucalyptus* and *Curcuma* Species. *Asian Pacific Journal of Tropical Biomedicine*, 1633-1636.
126. Lee, S. Y., Median, A., Nur Ashikin, A. H., Azliana, A. B. S., Abas, F. (2014). Antioxidant and α -Glucosidase Inhibitory Activities of the Leaf and Stem of Selected Traditional Medicinal Plants. *Int. Food Res. J.*, 21, 165-172.

127. Chai, T. T., Chiam, M. J., Lau C. H., Ismail N. I. M., Ong, H. C., Manan, F. A, Wong, F. C. (2015). Alpha-glucosidase Inhibitory and Antioxidant Activity of Solvent Extracts and Fractions of *Typha domingensis* (Typhaceae) Fruit. *Tropical Journal of Pharmaceutical Research*, 14, 1983-1990.
128. Kim, S., Jo, S., Kwon, Y. and Hwang, J. (2011). Effects of Onion (*Allium cepa* L.) Extract Administration on Intestinal Alpha-glucosidases Activities and Spikes in Postprandial Blood Glucose Levels in SD Rats Model. *International Journal of Molecular Sciences*, 12, 3757-3769.
129. Likhitwitayawuid, K. and Sritulak, B. (2001). A new Dimeric Stilbene with Tyrosinase Inhibitory Activity from *Artocarpus gomezianus*. *J. Nat. Prod.*, 64, 1457-1459.
130. Kamkaen, N., Mulsri, N. and Treesak, C. (2007). Screening of Some Tropical Vegetables for Anti-tyrosinase Activity. *Thai Pharm. Health Sci. J.*, 2 (1), 15-19.