

SYNTHESIS AND MECHANICAL PROPERTIES OF CONDUCTIVE
COMPOSITE POLYLACTIC ACID/POLYANILINE SCAFFOLD FOR
POTENTIAL TISSUE ENGINEERING

FARAH NURULJANNAH BINTI DAHLI

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Master of Philosophy

Faculty of Chemical and Energy Engineering
Universiti Teknologi Malaysia

DECEMBER 2016

ACKNOWLEDGEMENTS

In the name of Allah S.W.T the most gracious and the most merciful. Alhamdulillah, with the utmost blessing from Allah and in the remembrance to our prophet Muhammad P.B.U.H His most beloved messenger of all time, the path of my master degree has come to a completion.

I would like to take this opportunity to show my deepest gratitude to Dr. Saiful Izwan Bin Dato' Abd Razak who had stand in front of me to lead my way in achieving my master degree. This thesis has become a reality due to his wholly support in knowledge, guidance and moral support along with the will from Allah S.W.T. The appreciation also goes to my co-supervisor, Assoc. Prof. Dr. Abdul Razak Rahmat and Prof. Dato' Ir. Dr. Mohammed Rafiq bin Dato' Abdul Kadir for their provision on supervising my research.

Not to be forgotten my beloved mother, Zakiah Binti Hashim for placing her highest belief in me to complete this thesis. Same thanks to my family members and friends who are willing to bear with me through thick and torn together in making my dream come true. Finally, tremendous assistance from all of my colleagues in polymeric biomaterials lab would not be forgotten and always be in my thought and prayer.

ABSTRACT

This thesis reports a new composite scaffold material that is conductive and porous made from degradable polylactic acid (PLA) and conducting polyaniline (PANI) which has the potential for use in promoting tissue regeneration. The conductive scaffold was successfully prepared using a simple yet effective method known as freeze extraction method. The doped PANI was synthesised using conventional method of oxidative chemical polymerization. The electrical percolation state was successfully obtained at 3 wt% of PANI inclusion and reached at useable conductivity level for tissue engineering application at 4 wt% PANI, $2.91 \times 10^{-3} \text{ Scm}^{-1}$. 4 wt% inclusion of PANI was justified as the best PLA/PANI composite scaffold because it met the criterion as an electro-responsive material where the conductivity achieved was higher than 10^{-3} Scm^{-1} . It is also much suitable material in the regeneration of skin tissue (fibroblast) because the mean pore size achieved was at $35.82 \mu\text{m}$ and optimum tensile strength at 3.08 MPa. The UV-spectrum of the conductive scaffold displayed transition peaks of PANI indicating the PANI was still in its conducting doped state inside the scaffold. Incubation for 24 weeks for in-vitro degradation revealed that the PANI component delayed the degradation of PLA. Preliminary bioactivity test results indicated that the doping agent able to form chelate at the scaffold surface and this could assist in the formation of in-vitro apatite during the biomimetic immersion.

ABSTRAK

Tesis ini melaporkan bahan komposit perancah terbaharu berkonduktif dan berliang yang diperbuat daripada asid polilaktik (PLA) berdegradasi dan polianilina (PANI) berkonduktif di mana berpotensi untuk menggalakkan pertumbuhan semula tisu. Perancah berkonduktif ini berjaya dihasilkan dengan menggunakan kaedah yang mudah tetapi berkesan yang dikenali sebagai pengekstrakan beku. PANI terdop telah disintesis dengan cara yang konvensional iaitu pemolimeran kimia secara oksidatif. Tahap perkolasi elektrik berjaya diperoleh pada 3% kemasukan PANI dan mencapai tahap konduktiviti yang berguna untuk kejuruteraan tisu pada 4% PANI iaitu $2.91 \times 10^{-3} \text{ Scm}^{-1}$. Kemasukan 4% PANI telah dibuktikan sebagai PLA/PANI perancah komposit yang terbaik kerana ianya memenuhi kriteria sebagai bahan yang elektro-responsif di mana pencapaian konduktiviti adalah lebih tinggi daripada 10^{-3} Scm^{-1} . Ianya juga bahan yang sangat sesuai dalam pertumbuhan semula tisu kulit (fibroblas) kerana purata saiz liang yang dicapai pada $35.82 \mu\text{m}$ dan kekuatan tegangan yang optimum pada 3.08 MPa. Spektra UV perancah berkonduktif ini menunjukkan kewujudan peralihan spektra PANI dan ini menunjukkan bahawa PANI masih berkeadaan terdop di dalam perancah tersebut. Tempoh pengeraman selama 24 minggu untuk degradasi secara in-vitro menunjukkan komponen PANI telah melambatkan degradasi PLA. Keputusan awal ujian bioaktiviti menunjukkan agen dop mampu membentuk sebagai kelat pada permukaan perancah dan ini dapat membantu pembentukan in-vitro apatit ketika rendaman cecair biomimetik.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	ACKNOWLEDGEMENTS	iii
	ABSTRACT	iv
	ABSTRAK	v
	TABLE OF CONTENTS	vi
	LIST OF TABLES	x
	LIST OF FIGURES	xi
	LIST OF ABBREVIATIONS	xiii
	LIST OF SYMBOLS	xiv
		lxi
1	INTRODUCTION	1
	1.1 Overview	1
	1.2 Problem Statement	3
	1.3 Objectives of Study	4
	1.4 Scopes of Study	4
2	LITERATURE REVIEW	6
	2.1 Tissue Engineering	6
	2.2 Scaffolds	7
	2.3 Synthetic Biodegradable Polymers	9
	2.3.1 Polylactic Acid (PLA)	10
	2.4 Scaffolds Preparation	11

	2.4.1 Freeze Extraction	13
2.5	Composite Scaffolds	14
2.6	Electrically Conductive Polymers	16
	2.6.1 Polyaniline (PANI)	16
	2.6.2 Acid Doping and Bioactivity	19
	2.6.3 PANI in Tissue Engineering	21
2.7	PLA/PANI Composite	22
2.8	Summary of Literature Review	23
3	MATERIALS AND METHODS	24
3.1	Materials And Reagents	24
3.2	Experiment Methods	27
	3.2.1 Synthesis of PANI	27
	3.2.2 Preparation Composite Scaffold by Freeze Extraction	28
3.3	Characteristic and Testing	29
	3.3.1 DC Conductivity Testing	29
	3.3.2 Porosity and Pore Size	30
	3.3.3 Degree of Swelling	30
	3.3.4 Mechanical Testing	31
	3.3.5 UV-vis Spectroscopy	32
	3.3.6 Scanning Electron Microscopy	32
	3.3.7 In-Vitro degradation	33
	3.3.8 In-Vitro Bioactivity	33
3.4	Flow Diagram of the Research Methodology	34
4	RESULTS AND RESULTS	36
4.1	Morphology And Appearance of Synthesized PANI	36
4.2	Electrical Conductivity of Composite Scaffold	37
4.3	Morphology of Composite Scaffold	39
4.4	Porosity, Pore Size And Degree of Swelling	41
4.5	Tensile Properties of Composite Scaffold	42

4.6	UV-vis Spectra of Composite Scaffold	43
4.7	In-vitro degradation of Composite Scaffold	44
4.8	In-vitro Bioactivity of Composite Scaffold	45
5	CONCLUSIONS	48
5.1	Conclusion	48
5.2	Recommendations	49
	REFERENCES	50
	LIST OF PUBLICATIONS	61
	APPENDICES	62-63

LIST OF TABLES

TABLE NO.	TITLE	PAGE
3.1	Chemicals and reagents	26
3.2	Ion concentration of 1.5 SBF	34
4.1	Porosity, pore size and degree of swelling	41
4.2	Tensile properties	42

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Repeating unit of PLA (Gruber et al, 2003)	10
2.2	Different oxidation states in PANI: (a) leucoemeraldine, (b) emeraldine, (c)nigraniline, d) pernigraniline	17
2.3	Reversible transformations from ES to EB of PANI	18
2.4	Reaction mechanism of polyaniline via radical cation polymerization	19
3.1	Chemical structure of aniline	24
3.2	Schematic diagram of the synthesis of PANI	27
3.3	Freeze extraction of scaffold	28
3.4	Flow diagram of the research methodology	35
4.1	(a)SEM image of synthesized PANI at 10,000 magnification (b)PANI suspended in 1,4-dioxane	37
4.2	DC conductivity of PLA scaffold as function of PANI loading	38
4.3	SEM images of neat PLA at (a) 350 (b) 20,000 magnifications	39
4.4	SEM images of PLA/4PANI at (a) 350 (b) 20,000 (c)100,000 and (d) 200,000 magnifications, and (e) 200,000 magnification of PLA/5PANI	40
4.5	UV-vis spectra of (a) neat doped PANI, and (b)PLA/4PANI	43
4.6	SEM images of (a) neat PLA and PLA/4PANI after immersion in PBS for 24 weeks	45

4.7	SEM images of (a) neat PLA and PLA/4PANI after 5 days of soaking in SBF (HA marked in circles)	46
4.8	EDX spectra of PLA/4PANI after 5 days of soaking in SBF	47
4.9	Schematic illustration for the formation of chelation complex of citric acid and calcium ion	47

LIST OF ABBREVIATIONS

1D	-	One dimensional
3D	-	Three dimensional
CA- <i>cit</i>	-	Calcium-citric acid
CHCl ³	-	trichloromethane
DC	-	Direct Current
FC	-	Fast cooling rate
FDA	-	Food and Drug Administration
HA	-	hydroxyl apatite
<i>m</i> -ABA	-	<i>m</i> -Aminobenzoic acid
mmol	-	Milimole
MWCNT	-	Multiwall Carbon Nanotubes
PANI	-	Polyaniline
PANI-CSA	-	PANI-camphor sulfonic acid
PEDOT	-	Poly(3,4-ethylene dioxythiophene)
PLA	-	Poly(lactic Acid)
PLLA	-	Poly-L-Lactic Acid
PGA	-	Polyglycolic Acid
PPY	-	Polypyrrole
PPV	-	Poly(p-phenylene-vinylene)
PYG	-	Polypyrrole/graphene
SBF	-	Simulated body fluid
SC	-	Slow cooling rate
SEM	-	Scanning Electron Microscopy
UV-vis	-	Ultraviolet-Visible Spectroscopy

LIST OF SYMBOLS

α	-	Alpha
A	-	Area
E	-	Electric field intensity
I	-	Current
J	-	Current density
σ	-	Conductivity of the material
M_s	-	Mass of scaffold after immersion in water
M_d	-	Mass of dry scaffold
ρ	-	Resistivity
π	-	Pi bonding orbital
π^*	-	Antibonding Pi orbital
R	-	Resistance
t	-	thickness
V	-	Voltage
V_d	-	Apparent volume
V_p	-	Pore volume
W_d	-	Surplus weight of scaffold after degradation
W_i	-	Initial weight of scaffold before degradation

CHAPTER 1

INTRODUCTION

1.1 Overview

Scaffolds in tissue engineering refer to biodegradable materials which are highly porous that can act as template for tissue regeneration (Yang *et al.*, 2001). Synthetic biodegradable scaffold such as polylactic acid (PLA) has found wide range of pharmaceutical applications in the tissue regeneration of skin (Mohiti-Asli *et al.*, 2015), cartilage (Muhonen *et al.*, 2015), blood vessel (Li *et al.*, 2015) and cardiac valve (Iop and Gerosa 2015). The advantages of PLA are its synthetically controllable degradation rate (Cui *et al.*, 2015), good mechanical properties (Shi *et al.*, 2015) and biocompatibility (Abdal-hay *et al.*, 2015) plus it can be produced from renewable resource (Yang *et al.*, 2015).

The methods of preparing a porous PLA scaffold are diverse which includes, thermally induced phase separation (Mannella *et al.*, 2015), 3D printing (Rosenzweig *et al.*, 2015), porogen leaching (Choudhury *et al.*, 2015), the highly popular freeze drying (Salerno *et al.*, 2015) and electrospinning (Morelli *et al.*, 2015). Another method to produce polymeric porous scaffold is the simple freeze extraction (Adeli *et al.*, 2011).

Though there are few reports on PLA scaffold prepared by freeze extraction method with the inclusion of other fillers or reinforcements such as carbon nanotubes (Adeli *et al.*, 2011), chitosan and alginate (Yuan *et al.*, 2009), bioactive glass (El-Kady *et al.*, 2010), to date there are no reported studies on the preparation of freeze extracted porous conductive scaffold of PLA with the inclusion of conducting polymers such as of polyaniline (PANI).

PANI is one of the most promising conducting polymers for wide range of applications (Li *et al.*, 2008) mainly due to its ease of synthesis and preparation (Bhadra *et al.*, 2009), excellent electrical properties (Wang *et al.*, 2015) and being biocompatible (Bidez *et al.*, 2006). Inclusion of conductive PANI filler in the PLA scaffold might open up opportunities in many biomedical applications such as tissue engineering. It is only quite recently that the tuneable electroactivity of PANI has been explored in the area of diverse biomedical applications, such as for scaffolds in tissue engineering (Qazi *et al.*, 2014).

Earlier *in vivo* test revealed that various forms of PANI caused minimal inflammation after 50 weeks of implantation beneath the dorsal skin of rats (Wang *et al.*, 1999). It was also shown that PANI can be a good reducing agents and effective scavengers of free radicals when present in biological media (Gizdavic-Nikolaidis *et al.*, 2004). Investigation on adhesion and proliferation of cardiac myocytes on PANI concluded that PANI potential usefulness as an electroactive conductive polymer in cell-culture experiments (Bidez *et al.*, 2006), able to stimulate cell differentiation to cardiomyocytes (Borriello *et al.*, 2011) and biocompatible for both healthy and cancer cells after some modifications (Yslas *et al.*, 2015). However, due to its brittleness and nonprocessability (Saini *et al.*, 2012), it should be incorporated into other polymers that able to be fabricated into a tissue engineered scaffold.

Therefore, the main aim of this study is to prepare and investigate the effects of PANI addition on the properties of PLA scaffold prepared using freeze extraction. This new type of conductive composite scaffold is expected to exhibit new and

enhanced properties including the ease of processing and low cost. Such conductive scaffold may be usable in many applications in tissue engineering and biomedical implants such as controllable electrically responsive cell growth scaffold, controllable drug delivery sites and skin graft for wounds.

1.2 Problem Statements

Most of the research works on PLA composite scaffold are focused on the mechanical and morphology improvement. Nonetheless it was shown that certain material can enhance Schwann cell growths for neural tissue engineering upon applied voltage (Baniasadi *et al.*, 2015). This could decrease the time taken for the cells to fully mature and it could lessen the time for patients to wait for their new regenerate tissue. Thus it seems feasible to induce a certain degree of electrical conductivity to a scaffold material in order to obtain cell responsive properties for tissue engineering. Though being reported, the study on conductive scaffold is still limited to some extent.

Freeze drying is a widely used method to prepare porous scaffold but it is time and energy consuming (Baldino *et al.*, 2015). Plus the resulting freeze dried scaffold usually produced unwanted surface skin which requires additional process thus becomes economically uncompetitive (Sachlos and Czernuszka, 2003). In regards to conductive scaffolds, they have been fabricated using the electrospinning method mainly due to their nanofiber formation which led to high porosity (McKeon 2010, Shokry *et al.*, 2015). Though the electrospinning process seemed feasible, various cumbersome factors should be taken into consideration to obtain its nanofiber form such as applied voltage, solvent mixtures, distance between the tip and the collector, viscosity of the polymer solution, flow rate and even humidity/temperature of the spinning chamber (Subbiah *et al.*, 2005).

Being relatively new in the tissue engineering field, conductive scaffold prepared using freeze extraction has many unexplored features and characteristics. Many aspects that should be studied which includes the electrical conductivity enhancement, morphology, pore size and porosity, electronic transitions, biodegradability and bioactivity.

1.3 Objectives

This study was conducted in order to fulfil the following objectives:

1. To prepare conductive composite scaffold of PLA/PANI via freeze extraction
2. To characterize the electrical, physical and morphological properties of the PLA/PANI scaffold
3. To evaluate the preliminary in-vitro degradation and preliminary bioactivity test

1.4 Scope of Study

In order to satisfy all the outlined objectives, the scopes of this research are undertaken according to the following:

Initially, PANI was synthesized according to conventional method as reported in literatures. The synthesized PANI was characterized for its morphology, color appearance, DC electrical conductivity and UV-vis spectroscopy. Following

that, the as synthesized PANI will be used as fillers in the preparation of conductive scaffold.

Next step was to prepare the scaffold by the inclusions of PANI within the PLA using freeze extraction. Amount of PANI used were (0.5, 1, 2, 3, 4, 5 wt%). The resulting conductive composite scaffolds were evaluated in terms of its DC conductivity, tensile properties, porosity, pore size and degree of swelling. Scaffold of PLA/PANI with a suitable electrical conductivity value and good physical characteristics were identified and further tested using UV-vis spectroscopy and scanning electron microscope.

Consequently the conductive composite scaffold was tested for in-vitro degradation; evaluating the weight loss and the resulting morphology. Bioactivity test of the conductive scaffold was done by immersion in simulated body fluid solution (SBF), followed by the evaluation of hydroxyl apatite growth on the sample.

REFERENCES

- Abdal-hay, A., Hussein, K. H., Casettari, L., Khalil, K. A., and Hamdy, A. S. (2016). Fabrication of novel high performance ductile poly (lactic acid) nanofiber scaffold coated with poly (vinyl alcohol) for tissue engineering applications. *Materials Science and Engineering: C*. 60, 143–150.
- Adeli, H., Zein, S. H. S., Tan, H. S., Akil, H. M., and Ahmad, A. L. (2011). Synthesis, characterization and biodegradation of novel poly (L-lactide)/multiwalled carbon nanotube porous scaffolds for tissue engineering applications. *Current Nanoscience*. 7(3), 323–332.
- Azmi, S., Razak, S. I. A., Kadir, M. R. A., Iqbal, N., Hassan, R., Nayan, N. H. M., Wahab, A. H. A., and Shaharuddin, S. (2016). Reinforcement of Poly (vinyl alcohol) Hydrogel with Halloysite Nanotubes as Potential Biomedical Materials. *Soft Materials*. In Press. DOI: 10.1080/1539445X.2016.1242500
- Badami, A. S., Kreke, M. R., Thompson, M. S., Riffle, J. S., and Goldstein, A. S. (2006). Effect of fiber diameter on spreading, proliferation, and differentiation of osteoblastic cells on electrospun poly (lactic acid) substrates. *Biomaterials*. 27(4), 596-606.
- Baldino, L., Concilio, S., Cardea, S., De Marco, I., and Reverchon, E. (2015). Complete glutaraldehyde elimination during chitosan hydrogel drying by SC-CO₂ processing. *The Journal of Supercritical Fluids*. 103, 70–76.
- Baniasadi, H., Ramazani S. A. A., and Mashayekhan, S. (2015). Fabrication and characterization of conductive chitosan/gelatin-based scaffolds for nerve tissue engineering. *International Journal of Biological Macromolecules*. 74, 360-366.
- Baniasadi, H., Ramazani S. A. A., Mashayekhan, S., Farani, M. R., Ghaderinezhad, F., and Dabaghi, M. (2015). Design, fabrication, and characterization of novel porous conductive scaffolds for nerve tissue engineering. *International Journal of Polymeric Materials and Polymeric Biomaterials*. 64(18), 969-977.

- Bean, A. C., and Tuan, R. S. (2015). Fiber diameter and seeding density influence chondrogenic differentiation of mesenchymal stem cells seeded on electrospun poly (ϵ -caprolactone) scaffolds. *Biomedical Materials*. 10(1), 015018.
- Bhadra, S., Khastgir, D., Singha, N. K., and Lee, J. H. (2009). Progress in preparation, processing and applications of polyaniline. *Progress in Polymer Science*. 34(8), 783–810.
- Bidez, P. R., Li, S., MacDiarmid, A. G., Venancio, E. C., Wei, Y., and Lelkes, P. I. (2006). Polyaniline, an electroactive polymer, supports adhesion and proliferation of cardiac myoblasts. *Journal of Biomaterials Science, Polymer Edition*. 17(1-2), 199-212.
- Borriello, A., Guarino, V., Schiavo, L., Alvarez-Perez, M. A., and Ambrosio, L. (2011). Optimizing PANi doped electroactive substrates as patches for the regeneration of cardiac muscle. *Journal of Materials Science:Materials in Medicine*. 22(4), 1053–1062.
- Budyanto, L., Goh, Y. Q., and Ooi, C. P. (2009). Fabrication of porous poly(L-lactide)(PLLA) scaffolds for tissue engineering using liquid-liquid phase separation and freeze extraction. *Journal of Material Science: Material Medicine*. 20, 105-111.
- Choudhury, M., Mohanty, S., and Nayak, S. (2015). Effect of different solvents in solvent casting of porous scaffolds – in biomedical and tissue engineering applications. *Journal of Tissue Science & Engineering*. 5, 1–9.
- Chutipakdeevong, J., Ruktanonchai, U., and Supaphol, P. (2015). Hybrid biomimetic electrospun fibrous mats derived from poly (ϵ -caprolactone) and silk fibroin protein for wound dressing application. *Journal of Applied Polymer Science*, 132(11).
- Coleman, J. N., Khan, U., Blau, W. J., and Gun'ko, Y. K. (2006). Small but strong: a review of the mechanical properties of carbon nanotube–polymer composites. *Carbon*. 44(9), 1624-1652.
- Cui, M., Liu, L., Guo, N., Su, R., and Ma, F. (2015). Preparation, cell compatibility and degradability of collagen-modified poly (lactic acid). *Molecules*. 20(1), 595–607.
- Cui, W., Zhou, Y., and Chang, J. (2016). Electrospun nanofibrous materials for tissue engineering and drug delivery. *Science and Technology of Advanced Materials*. 11(1), 014108.

- Densakulprasert, N., Wannatong, L., Chotpattananont, D., Hiamtup, P., Sirivat, A., and Schwank, J. (2005). Electrical conductivity of polyaniline/zeolite composites and synergetic interaction with CO. *Materials Science and Engineering: B*. 117(3), 276-282.
- Dhandayuthapani, B., Yoshida, Y., Maekawa, T., and Kumar, D. S. (2011). Polymeric scaffolds in tissue engineering application: a review. *International Journal of Polymer Science*. 2011.
- Ding, H. J., Shen, J. Y., Wan, M. X., and Chen, Z. J. (2008). Formation mechanism of polyaniline nanotubes by a simplified template-free method. *Macromolecular Chemistry and Physics*. 209(8), 864-871.
- El-Kady, A. M., Saad, E. A., El-Hady, B. M. A., and Farag, M. M. (2010). Synthesis of silicate glass/poly (L-lactide) composite scaffolds by freeze-extraction technique: characterization and in vitro bioactivity evaluation. *Ceramics International*. 36(3), 995-1009.
- Gao, J., Sansinena, J. M., and Wang, H. L. (2003). Chemical vapor driven polyaniline sensor/actuators. *Synthetic Metals*. 135, 809-810.
- Garlotta, D. (2001). A literature review of poly(lactic acid). *Journal of Polymers and the Environment*. 9(2), 63-84.
- Ghanbari, K., Mousavi, M. F., Shamsipur, M., and Karami, H. (2007). Synthesis of polyaniline/graphite composite as a cathode of Zn-polyaniline rechargeable battery. *Journal of Power Sources*. 170(2), 513-519.
- Gizdavic-Nikolaidis, M., Travas-Sejdic, J., Bowmaker, G. A., Cooney, R. P., Thompson, C., and Kilmartin, P. A. (2004). The antioxidant activity of conducting polymers in biomedical applications. *Current Applied Physics*. 4(2), 347-350.
- Green, R. A., Baek, S., Poole-Warren, L. A., and Martens, P. J. (2016). Conducting polymer-hydrogels for medical electrode applications. *Science and Technology of Advanced Materials*. 11(1), 014107.
- Gruber, P., and O'Brien, M. (2003). Polylactides "NatureWorks™ PLA". *Biopolymers*. 4, 235-252.
- Guo, B., Glavas, L., and Albertsson, A. C. (2013). Biodegradable and electrically conducting polymers for biomedical applications. *Progress in Polymer Science*. 38(9), 1263-1286.

- Guo, Y., and Zhou, Y. (2007). Polyaniline nanofibers fabricated by electrochemical polymerization: a mechanistic study. *European Polymer Journal*. 43(6), 2292-2297.
- Heeger, A. J. (2010). Semiconducting polymers: the third generation. *Chemical Society Reviews*. 39(7), 2354-2371.
- Ho, M., Kuo, P., Hsieh, H., Hsien, T., Hou, L., Lai, J., and Wang, D. (2004). Preparation of porous scaffolds by using freeze-extraction and freeze-gelation methods. *Journal of Biomaterials*. 25, 129-138.
- Huang, J. (2006). Synthesis and applications of conducting polymer polyaniline nanofibers. *Pure and Applied Chemistry*. 78(1), 15-27.
- Huang, J., and Kaner, R. B. (2004). A general chemical route to polyaniline nanofibers. *Journal of the American Chemical Society*. 126(3), 851-855.
- Humpolicek, P., Kasparkova, V., Saha, P., and Stejskal, J. (2012). Biocompatibility of polyaniline. *Journal of Synthetic Materials*. 162, 722-727.
- Iop, L., and Gerosa, G. (2015). Guided tissue regeneration in heart valve replacement: from preclinical research to first-in-human trials. *BioMed Research International*, 2015. ID:432901.
- Jang, K. S., Lee, H., and Moon, B. (2004). Synthesis and characterization of water soluble polypyrrole doped with functional dopants. *Synthetic Metals*. 143(3), 289-294.
- Jiang, Y., Wang, Z. H., and Cromack, K. R. (2002). Effect of sulfonic acid group on polyaniline backbone. *J Am Chem Soc*. 113(7), 2665-2671.
- Joziassse, C. A. P., Grijpma, D. W., Bergsma, J. E., Cordewener, F. W., Bos, R. R. M., and Pennings, A. J. (1998). The influence of morphology on the (hydrolytic degradation of as-polymerized and hot-drawn poly (L-lactide)). *Colloid and Polymer Science*. 276(11), 968-975.
- Kai, D., Prabhakaran, M. P., Jin, G., and Ramakrishna, S. (2013). Biocompatibility evaluation of electrically conductive nanofibrous scaffolds for cardiac tissue engineering. *Journal of Materials Chemistry B*. 1(17), 2305-2314.
- Kim, B. J., Oh, S. G., Han, M. G., and Im, S. S. (2000). Preparation of polyaniline nanoparticles in micellar solutions as polymerization medium. *Langmuir*. 16(14), 5841-5845.
- Kokubo, T., and Takadama, H. (2006). How Useful is SBF in predicting in vivo bone bioactivity?. *Biomaterials*. 27(15), 2907-2915.

- Kokubo, T., Ito, S., Shigematsu, M., Sanka, S., and Yamamuro, T. (1987). Fatigue and life-time of bioactive glass-ceramic AW containing apatite and wollastonite. *Journal of Materials Science*. 22(11), 4067-4070.
- Kulkarni, M. V., Viswanath, A. K., Marimuthu, R., and Seth, T. (2004). Synthesis and characterization of polyaniline doped with organic acids. *Journal of Polymer Science Part A: Polymer Chemistry*. 42(8), 2043-2049.
- Laska, J., Zak, K., and Proń, A. (1997). Conducting Blends of Polyaniline with Conventional Polymers. *Synthetic Metals*. 84(1), 117-118.
- Lasprilla A. J. R, Martinez, G. A. R., and Lunelli, B. H. (2012). Poly-lactic acid synthesis for application in biomedical devices – A review. *Journal of Biotechnology Advances*. 30, 321-328.
- Leong, W. S., Wu, S. C., Ng, K., and Tan, L. P. (2016). Electrospun 3D multi-scale fibrous scaffold for enhanced human dermal fibroblast infiltration. *International Journal of Bioprinting*. 2(1).
- Li, C., Bai, H., and Shi, G. Q. (2009). Conducting polymer nanomaterials: electrosynthesis and applications. *Chemical Society Reviews*. 38(8), 2397-2409.
- Li, D., Huang, J., and Kaner, R. B. (2008). Polyaniline nanofibers: a unique polymer nanostructure for versatile applications. *Accounts of Chemical Research*. 42(1), 135-145.
- Li, Z., Zhao, X., Ye, L., Coates, P., Caton-Rose, F., and Martyn, M. (2015). Fibrillation of chain branched poly (lactic acid) with improved blood compatibility and bionic structure. *Chemical Engineering Journal*. 279, 767–776.
- Lu, X. F., Zhang, W. J., Wang, C., Wen, T. C., and Wei, Y. (2010). One-dimensional conducting polymer nanocomposites: synthesis properties and applications. *Progress in Polymer Science*. 36(5), 671-712.
- Luo, Y., Engelmayer, G., Auguste, D. T., Ferreira, L. D. S., Karp, J. M., Saigal, R., and Langer, R. (2007). Three-dimensional scaffolds. Lanza, Langer and Vacanti (Eds.) *Principles of Tissue Engineering, 3rd Edition* (pp. 3-6). London: Academic Press.
- MacDiarmid, A. G. (2001). “Synthetic metals”: a novel role for organic polymers. *Current Applied Physics*. 1(4), 269-279.

- Mannella, G. A., Conoscenti, G., Pavia, F. C., La Carrubba, V., and Brucato, V. (2015). Preparation of polymeric foams with a pore size gradient via thermally induced phase separation (TIPS). *Materials Letters*. 160, 31–33.
- Martínez-Pérez, C. A., Olivás-Armendariz, I., Castro-Carmona, J. S., and García-Casillas, P. E. (2011). Scaffolds for tissue engineering via thermally induced phase separation. *Advances in Regenerative Medicine: InTech*. 275-294.
- Mattioli-Belmonte, M., Giavaresi, G., Biagini, G., Virgili, L., Giacomini, M., Fini, M., Giantomassi, D. Natali, P. Torricelli, and Giardino, R. (2003). Tailoring biomaterial compatibility: in vivo tissue response versus in vitro cell behavior. *The International Journal of Artificial Organs*. 26(12), 1077-1085.
- McKeon, K. D., Lewis, A., and Freeman, J. W. (2010). Electrospun poly (D, L-lactide) and polyaniline scaffold characterization. *Journal of Applied Polymer Science*. 115(3), 566–1572.
- Meszynska, A., Pollet, E., Odelius, K., Hakkarainen, M., and Avérous, L. (2015). Effect of Oligo-Hydroxyalkanoates on Poly (3-Hydroxybutyrate-co-4-Hydroxybutyrate)-Based Systems. *Macromolecular Materials and Engineering*. 300(6), 661-666.
- Meyer, J. L., and Thomas Jr, W. C. (1982). Trace metal-citric acid complexes as inhibitors of calcification and crystal growth. I. Effects of Fe (III), Cr (III) and Al (III) complexes on calcium phosphate crystal growth. *The Journal of Urology*. 128(6), 1372-1375.
- Mohiti-Asli, M., Saha, S., Murphy, S. V., Gracz, H., Pourdeyhimi, B., Atala, A., and Lobo, E.G. (2015). Ibuprofen loaded PLA nanofibrous scaffolds increase proliferation of human skin cells in vitro and promote healing of full thickness incision wounds in vivo. *Journal of Biomedical Materials Research Part B: Applied Biomaterials*. 2015(00B), 000–000.
- Morelli, S., Salerno, S., Holopainen, J., Ritala, M., and De Bartolo, L. (2015). Osteogenic and osteoclastogenic differentiation of co-cultured cells in polylactic acid–nanohydroxyapatite fiber scaffolds. *Journal of Biotechnology*. 204, 53–62.
- Muhonen, V., Salenius, E., Haaparanta, A. M., Järvinen, E., Paatela, T., Meller, A., and Kiviranta, I. (2015). Articular cartilage repair with recombinant human type II collagen/polylactide scaffold in a preliminary porcine study. *Journal of Orthopaedic Research*. 34, 745–753

- Najim, T. S., and Salim, A. J. (2014). Polyaniline nanofibers and nanocomposites: preparation, characterization and application for Cr(IV) and phosphate ions removal from aqueous solution. *Arabian Journal of Chemistry*.
- Nakanishi, K., Okuma, M., and Katayama, S. (1993). U.S. Patent No. 5,259,985. Washington, DC: U.S. Patent and Trademark Office.
- Nam, Y. S., and Park, T. G. (1999). Porous biodegradable polymeric scaffolds prepared by thermally induced phase separation. *Journal of Biomedical Materials Research*. 47(1), 8-17.
- O'Brien, F. J. (2011). Biomaterials & scaffolds for tissue engineering. *Materials Today*. 14(3), 88-95.
- O'Brien, F. J., Harley, B. A., Yannas, I. V., and Gibson, L. J. (2005). The effect of pore size on cell adhesion in collagen-GAG scaffolds. *Biomaterials*. 26(4), 433-441.
- Odedra, D., Chiu, L., Reis, L., Rask, F., Chiang, K., and Radisic, M. (2011). Cardiac tissue engineering. Burdick, J. A. and Mauck, R.L. (Eds.). *Biomaterials for Tissue Engineering Applications*. (pp. 421-456). London: Springer-Verlag.
- Pandey, S. S., Annapoorni, S., and Malhotra, B. D. (1993). Synthesis and characterization of poly (aniline-co-o-anisidine). A processable conducting copolymer. *Macromolecules*. 26(12), 3190-3193.
- Pham, Q. P., Sharma, U., and Mikos, A. G. (2006). Electrospinning of polymeric nanofibers for tissue engineering applications: a review. *Tissue Engineering*. 12(5), 1197-1211.
- Picciani, P. H., Medeiros, E. S., Pan, Z., Orts, W. J., Mattoso, L. H., and Soares B. G. (2009). Development of conducting polyaniline/poly(lactic acid) nanofibers by electrospinning. *Journal of Applied Polymer Science*. 112(2), 744-753.
- Pollet, E., and Avérous, L. (2012). Biodegradable polymers. Pollet, E. and Avérous, L. (Eds.) *Environmental Silicate Nano-biocomposites*. (pp. 13-39). London: Springer.
- Qazi, T. H., Rai, R., and Boccaccini, A. R. (2014). Tissue engineering of electrically responsive tissues using polyaniline based polymers: a review. *Biomaterials*. 35(33), 9068-9086.
- Rahman, N. A., Gizdavic-Nikolaidis, M., Ray, S., Easteal, A. J., and Travas-Sejdic, J. (2010). Functional electrospun nanofibers of poly(lactic acid) blends with

- polyaniline or poly(aniline-co-benzoic acid). *Journal of Synthetic Metals*. 160, 2015-2022.
- Rakhmatia, Y. D., Ayukawa, Y., Atsuta, I., Furuhashi, A., and Koyano, K. (2015). Fibroblast attachment onto novel titanium mesh membranes for guided bone regeneration. *Odontology*. 103(2), 218-226.
- Razak, S. I. A., Abdul Rahman, W. A. W., Hashim, S., and Yahya, M. Y. (2013). Enhanced interfacial interaction and electronic properties of novel conducting kenaf/polyaniline biofibers. *Polymer-Plastics Technology and Engineering*. 52(1), 51-57.
- Razak, S. I. A., Ahmad, A. L., Zein, S. H. S., and Boccaccini, A. R. (2009). MnO₂-filled multiwalled carbon nanotube/polyaniline nanocomposites with enhanced interfacial interaction and electronic properties. *Scripta Materialia*. 61(6), 592-595.
- Razak, S. I. A., Dahli, F. N., Wahab, I. F., Abdul Kadir, M. R., Muhamad, I. I., Yusof, A. H. M., and Adeli, H. (2016). A Conductive polylactic acid/polyaniline porous scaffold via freeze extraction for potential biomedical applications. *Soft Materials*. 14(2), 78-86.
- Razak, S. I. A., Rahman, W. A. W. A., and Yahya, M. Y. (2012). Electrically conductive nanocomposites of epoxy/polyaniline nanowires doped with formic acid: effect of loading on the conduction and mechanical properties. *NANO*. 7, 1250039.
- Razak, S. I. A., Sharif, N. F. A., and Muhamad, I. I. (2014). Polyaniline coated halloysite nanotubes: effect of para-hydroxybenzene sulfonic acid doping. *Composite Interfaces*. 21(8), 715-722.
- Razak, S. I. A., Wahab, I. F., Kadir, M. R. A., Khudzari, A. Z. M., Yusof, A. H. M., Dahli, F. N., Nayan, H. N. M., and Anand, T. J. S. (2016). Biomimetic growth of hydroxyapatite on kenaf fibers. *BioResources*. 11(1), 1971-1981.
- Rengier, F., Mehndiratta, A., von Tengg-Kobligk, H., Zechmann, C. M., Unterhinninghofen, R., Kauczor, H. U., and Giesel, F. L. (2010). 3D printing based on imaging data: review of medical applications. *International Journal of Computer Assisted Radiology and Surgery*. 5(4), 335-341.
- Rhee, S. H., and Tanaka, J. (1999). Effect of citric acid on the nucleation of hydroxyapatite in a simulated body fluid. *Biomaterials*. 20(22), 2155-2160.

- Rosenzweig, D. H., Carelli, E., Steffen, T., Jarzem, P., and Haglund, L. (2015). 3D-printed ABS and PLA scaffolds for cartilage and nucleus pulposus tissue regeneration. *International Journal of Molecular Sciences*. 16(7), 15118-15135.
- Sachlos, E., and Czernuszka, J. T. (2003). Making tissue engineering scaffolds work. Review: the application of solid freeform fabrication technology to the production of tissue engineering scaffolds. *European Cells & Materials*. 5(29), 39–40.
- Saini, P., Choudhary, V., Vijayan, N., and Kotnala, R. K. (2012). Improved electromagnetic interference shielding response of poly (aniline)-coated fabrics containing dielectric and magnetic nanoparticles. *The Journal of Physical Chemistry C*. 116(24), 13403–13412.
- Salerno, A., Fernández-Gutiérrez, M., del Barrio, J. S. R., and Domingo, C. (2015). Bio-safe fabrication of PLA scaffolds for bone tissue engineering by combining phase separation, porogen leaching and scCO₂ drying. *The Journal of Supercritical Fluids*. 97, 238–246.
- Serrano, W., Melendez, A., Ramos, I., and Pinto, N. J. (2014). Electropsun composite poly(lactic acid)/polyaniline nanofibers from low concentrations in CHCl₃: Making a biocompatible polyester electro-active. *Journals of Polymer*. 55, 5727-5733.
- Sharifian, I. (2011). Conductive and biodegradable polyaniline/starch blends and their composites with polystyrene. *Iran Polymers Journals*. 20(4), 319-328.
- Shi, H., Gan, Q., Liu, X., Ma, Y., Hu, J., Yuan, Y., and Liu, C. (2015). Poly (glycerol sebacate)-modified polylactic acid scaffolds with improved hydrophilicity, mechanical strength and bioactivity for bone tissue regeneration. *RSC Advances*. 5(97), 79703–79714.
- Shokry, H., Vanamo, U., Wiltschka, O., Niinimäki, J., Lerche, M., Levon, K., and Sahlgren, C. (2015). Mesoporous silica particle-PLA–PANI hybrid scaffolds for cell-directed intracellular drug delivery and tissue vascularization. *Nanoscale*. 7(34), 14434–14443.
- Stejskal, J., and Gilbert, R. G. (2002). Polyaniline. Preparation of a conducting polymer (IUPAC technical report). *Pure Applied Chemistry*. 74, 857-867

- Stejskal, J., Sapurina, I., and Trchoya, M. (2010). Polyaniline nanostructures and the role of aniline oligomers in their formation. *Progress Polymer Science*. 35, 1420–1481.
- Subbiah, T., Bhat, G. S., Tock, R. W., Parameswaran, S., and Ramkumar, S. S. (2005). Electrospinning of nanofibers. *Journal of Applied Polymer Science*. 96(2), 557–569.
- Vacanti, J., and Vacanti, C. A. (2007). The history and scope of tissue engineering. Lanza, Langer and Vacanti (Ed.) *Principles of Tissue Engineering, 3rd Edition* (pp. 3-6). London: Academic Press.
- Van Lieshout, M. I., Vaz, C. M., Rutten, M. C. M., Peters, G. W. M., and Baaijens, F. P. T. (2006). Electrospinning versus knitting: two scaffolds for tissue engineering of the aortic valve. *Journal of Biomaterials Science, Polymer Edition*. 17(1-2), 77-89.
- Wan, Y., Huang, W., Wang, Z., and Zhu, X.X. (2004). Preparation and characterization of high loading porous crosslinked poly (vinyl alcohol) resins. *Polymer*. 45(1), 71–77.
- Wang, N., Li, H., Chen, T., Wang, J., and Shen, Q. (2014). Formation and comparison of poly(L-lactide)-guided polyaniline morphology via normal and phase alternated interfacial polymerization. *Journal of Materials Letters*. 137, 203-205.
- Wang, W., Li, C., Shi, X., Liu, H., and Sun, L. (2016). Synthesis and characterization of polyaniline coating modification micro copper powder. *In Advanced Graphic Communications, Packaging Technology and Materials* (pp. 1031-1037). Springer Singapore.
- Wang, Y., Ji, H., Shi, H., Zhang, T., and Xia, T. (2015). Fabrication and characterization of stearic acid/polyaniline composite with electrical conductivity as phase change materials for thermal energy storage. *Energy Conversion and Management*. 98, 322-330.
- Weiller, B. H., Virji, S., Baker, C., Huang, J., Li, D., and Kaner, R. B. (2016). Polyaniline nanofibers and composite materials for chemical detection. *Une*. 13, 15.
- Whang, K., Thomas, C. H., Healy, K. E., and Nuber, G. (1995). A novel method to fabricate bioabsorbable scaffolds. *Polymers*. 36(4), 837-842.

- Williams, D. F. (Ed.) (1987). Definitions in biomaterials. *Proceedings of a consensus conference of the European Society for Biomaterials*. 3-5 March 1986. Chester, England. Volume 4.
- Woodruff, M. A., and Hutmacher, D. W. (2010). The return of a forgotten polymer— polycaprolactone in the 21st century. *Progress in Polymer Science*. 35(10), 1217-1256.
- Yang, J., Shi, G., Bei, J., Wang, S., Cao, Y., Shang, Q., and Wang, W. (2002). Fabrication and surface modification of macroporous poly (l-lactic acid) and poly (l-lactic-co-glycolic acid) (70/30) cell scaffolds for human skin fibroblast cell culture. *Journal of Biomedical Materials Research*. 62(3), 438-446.
- Yang, J., Webb, A. R., and Ameer, G. A. (2004). Novel citric acid-based biodegradable elastomers for tissue engineering. *Advanced Materials*. 16(6), 511-516.
- Yang, S., Leong, K. F., Du, Z., and Chua, C. K. (2001). The design of scaffolds for use in tissue engineering. Part I. Traditional factors. *Tissue Engineering*. 7(6), 679–689.
- Yang, S., Madbouly, S. A., Schrader, J. A., Srinivasan, G., Grewell, D., McCabe, K. G., and Graves, W. R. (2015). Characterization and biodegradation behavior of biobased poly (lactic acid) and soy protein blends for sustainable horticultural applications. *Green Chemistry*. 17(1), 380–393.
- Yslas, E.I., Cavallo, P., Acevedo, D. F., Barbero, C. A., and Rivarola, V. A. (2015). Cysteine modified polyaniline films improve biocompatibility for two cell lines. *Materials Science and Engineering: C*. 51, 51–56.
- Yuan, N. Y., Lin, Y. A., Ho, M. H., Wang, D. M., Lai, J. Y., and Hsieh, H. J. (2009). Effects of the cooling mode on the structure and strength of porous scaffolds made of chitosan, alginate, and carboxymethyl cellulose by the freeze-gelation method. *Carbohydrate Polymers*. 78(2), 349-356.
- Yuan, X., Mak, A. F., and Yao, K. (2002). Comparative observation of accelerated degradation of poly (L-lactic acid) fibres in phosphate buffered saline and a dilute alkaline solution. *Polymer degradation and stability*. 75(1), 45-53.
- Zhang, H. Z., Liu, X., Yang, M., and Zhu, L. (2015). Silk fibroin/sodium alginate composite nano-fibrous scaffold prepared through thermally induced phase-separation (TIPS) method for biomedical applications. *Materials Science and Engineering C*. 55, 8-13.

- Zhang, R., and Ma, P. X. (2004). Biomimetic polymer/apatite composite scaffolds for mineralized tissue engineering. *Macromolecular Bioscience*. 4(2), 100-111.
- Zhang, X., Qi, H., Wang, S., Feng, L., Ji, Y., Tao, L., and Wei, Y. (2012). Cellular responses of aniline oligomers: a preliminary study. *Toxicology Research*. 1(3), 201-205.