### STARCH DEGRADING BACTERIA FOR BIOHYDROGEN PRODUCTION

### ATTAHIRU ABUBAKAR

A dissertation submitted in partial fulfilment of the requirements for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

JANUARY 2016

In loving memory of my late grandparents Sheikh Muhammad Bello Gusau and Hajiya Fatima (Inno). May Allah forgive their shortcomings and make Jannatul Firdaus to be their final abode.

#### ACKNOWLEDGEMENT

All thanks are to Allah most Beneficent the most Merciful, for His unquantifiable favour and specifically for keeping me healthy and alive to witness this memorable time of my academic pursuit. May His peace and Blessing be upon the pathfinder, the incomparable General, Prophet Muhammad (S.AW), members of his family, his companions and all those who responded to this call up to the Day of Judgement.

First and foremost, I would like to express my deepest gratitude to my able intellectual supervisor, Dr. Mohd Firdaus Abdul Wahab, I remain forever grateful for his support, guidance, patience and encouragement throughout the period of the research. Thank you for disciplining my professional life. Special thanks and appreciation to my parents, Khalifa Attahiru Bello and Hajiya Fatima, whom their support both financially and morally can never be estimated, and for cultivating in me the habit of early rising. May Allah reward them abundantly AMEEN.

Next, I would like to thank all the staffs in FBME and postgraduate students who had guided me throughout the project. I am grateful for their advice and motivation without which this thesis would not have been the same as presented here.

Last but not least, I would like to extend my deepest and sincere appreciation to my family members. Thank you for their unconditional love and support at all time. Besides, I would like to express my gratitude to all my friends and colleagues for their help and encouragement. May almighty Allah bless and enrich each and every one for his or her contributions.

#### ABSTRACT

Global research is moving forward in developing biological production of hydrogen as a renewable energy source in order to reduce the use of non-renewable energy. Biohydrogen has the potential of replacing the hydrogen production by fossil fuels which is one of the major factors that cause the global warming. The amylolytic activity of several microorganisms capable of utilizing starch in the dark fermentation process has provided a great potential in biohydrogen production by the starch degrading bacteria. In this study, starch utilizing bacteria have been successfully isolated from cassava chips processing wastewater and sludge. Based on the morphological characteristics, the isolate was found to be Gram positive bacteria with spherical cell shape. 16S rRNA analysis identified the strain Lactococcus sp. SDB4 with 92% similarity to Lactococcus lactis. The partial 16S rRNA sequence has been deposited to GenBank with Accession Number KU160544. Furthermore, the isolate was subjected to anaerobic dark fermentation process using synthetic media with tapioca starch as the only source of carbon. The findings in this study indicate that the Lactococcus sp. SDB4 possesses an amylolytic activity and capable of utilizing starch efficiently (75%). This strain has the potential to be used in biohydrogen production using starch containing wastewater as substrate.

### ABSTRAK

Penyelidikan global dalam pembangunan pengeluaran biohydrogen mengurangkan sumber tenaga yang boleh diperbaharui semakin berkembang untuk mengurangkan penggunaan sumber tenaga tidak boleh yang diperbaharui. Biohidrogen berpotensi untuk menggantikan pengeluaran hydrogen daripada bahan api fosil yang merupakan salah satu faktor utama pemanasan global. Pembangunan mikroorganisma yang boleh menggunakan kanji berdasarkan aktiviti amylolytic dalam proses fermentasi gelap menyumbang kepada potensi pengeluaran biohydrogen. Dalam kajian ini, bacteria yang boleh menggunakan kanji telah berjaya diperoleh daripada sisa pemprosesan kerepek ubi kayu. Daripada segi morfologi, sel bacteria tersebut merupakan bakteria Gram positif yang berbentuk sfera. Berdasarkan analisis 16S rRNA, bacteria tersebut merupakan *Lactococcus* sp. SDB4 dan mempunyai 92% persamaan dengan *Lactococcus lactis*. Julukan 16S rRNA bacteria tersebut telah didaftarkan ke dalam GenBank dengan nombor pendaftaran KU160544. Bakteria teersebut diperoleh melalui proses fermentasi gelap yang menggunakan media sintetik yang ditambah kanji ubi kayu sebagai sumber karbon. Kajian ini juga menunjukkan bahawa bakteria Lactococcus sp. SDB4 aktiviti amylolytic dan berupaya untuk menggunakan (75%) mempunyai kanji. Bakteria ini berpotensi untuk diaplikasi dalam proses pengeluaran biohidrogen bersumberkan sisa air mengandungi kanji sebagai substrat.

# **TABLE OF CONTENTS**

CHAPTE	R	TITLE	PAGE
	DECI	LARATION	ii
	DEDICATION		
	ACKNOWLEDGEMENT		
	ABSTRACT ABSTRAK		
	TABI	LE OF CONTENTS	vii
	LIST	OF TABLES	Х
	LIST	OF FIGURES	xi
	LIST	OF ABBREVIATIONS AND SYMBOLS	xii
	LIST	OF APPENDICES	XV
1	INTRODUCTION		1
	1.1	Background of the Research	1
	1.2	Production of Hydrogen	2
	1.3	Applications of Hydrogen	3
	1.4	Problem Statement	3
	1.5	Research Objectives	4
	1.6	Significance of the Research	5
	1.7	Scope of the Research	5
2	LITERATURE REVIEW		6
	2.1	Carbohydrate	6
		2.1.1 Starch	7
		2.1.2 Amylose and Amylopectin	9
	2.2	Starch Degrading Enzyme from Starch Utilizing	
		Bacteria	10

2.3	Potential Application of Starch Utilizing Bacteria in	
	Biofuel Production	12
2.4	Biohydrogen	12
2.5	Pathways of Biohydrogen Production	13
2.6	Dark Fermentative Method of Hydrogen Production	15
2.7	Fermentative Microorganisms used in Hydrogen	
	Production	16
2.8	Parameters Affecting Dark Fermentative Hydrogen	
	Production	17
МАТ	TERIALS AND METHOD	19
3.1	Experimental Design	19
3.2	Instruments	21
3.3	Wastewater and Sludge Sample Collection	21
3.4	Preparation of Media	21
3.5	Isolation of Bacteria from Wastewater and Sludge	22
	3.5.1 Bacterial Glycerol Stock	23
3.6	Qualitative Screening for Starch Degrading Ability	
	of Isolates	23
3.7	Identification of Starch Utilizing Bacteria	24
	3.7.1 Morphological Observation	24
	3.7.2 Gram Staining	24
	3.7.3 16S rRNA Bacterial Identification	25
	3.7.3.1 Genomic DNA Extraction	25
	3.7.3.2 Purity Assessment and Quantification	
	of the Extracted DNA	27
	3.7.3.3 Agarose Gel Electrophoresis	27
	3.7.3.4 Polymerase Chain Reaction (PCR)	
	Amplification	28
	3.7.3.5 Purification of PCR Products	30
	3.7.3.6 16S rRNA Sequencing and Analysis	30
	3.7.3.7 Phylogenetic Tree Construction	31
3.8	Preparation of Synthetic Media for Fermentation	
	Process	31

3

	3.9 Determination of Hydrogen Production using Batch			
		Method	32	
4	RES	ULTS AND DISCUSSION	33	
	4.1	Isolation of Bacteria	33	
	4.2	Qualitative Screening for Starch Degrading Ability		
		of Isolates	34	
	4.3	Identification of Selected Starch Degrading Bacteria	35	
		4.3.1 Morphology Observation	36	
		4.3.2 Gram Staining	37	
		4.3.3 16S rRNA Sequence Analysis	38	
		4.3.3.1 Genomic DNA Extraction	39	
		4.3.3.2 PCR Amplification	41	
		4.3.3.3 16S rRNA Sequence BLAST Analysis	42	
		4.3.3.4 Phylogenetic Analysis	44	
	4.4	Growth profile of Lactococcus sp. SDB4	47	
	4.5	Starch Utilization and Hydrogen Production	49	
5	CON	CLUSION AND FUTURE WORK	52	
	5.1	CONCLUSION	52	
	5.2	FUTURE WORK	53	

REFERENCES	54
Appendices A – F	61 – 66

### LIST OF TABLES

TABLE NO	. TITLE	PAGE	
2.1	: Properties of starch granules from different botanical		
	sources.	8	
3.1	: The set of universal primers used for 16S rRNA gene		
	amplification.	28	
3.2	: Components of PCR reaction mixture	29	
3.3	: Thermal cycle profile for PCR	29	
4.1	: Summarized morphological characteristics of isolate		
	SDB4.	36	
4.2	: Selected BLAST search result for isolate SDB4.	44	

### LIST OF FIGURES

# FIGURE NO.

## TITLE

# PAGE

2.1	: Structure of linear Amylose and Amylopectin	9
3.1	: Overall flow chart of experimental design	20
4.1	: Screening for starch degrading ability of the isolates.	35
4.2	: Isolate SDB4 on enriched starch wastewater agar plate.	37
4.3	: Gram staining of isolate SDB4 under microscope.	38
4.4	: Agarose gel electrophoresis analysis of genomic DNA	
	extracted from isolate SDB4.	40
4.5	: DNA concentration and purity of the extracted DNA	
	from isolate SDB4	41
4.6	: Agarose gel electrophoresis analysis of PCR product	
	for isolate SDB4.	42
4.7	: Phylogenetic tree of starch-degrading bacterium	
	Lactococcus sp. SDB4 and its close relatives based on	
	the 16S rRNA gene sequence.	45
4.8	: Growth profile of <i>Lactococcus</i> sp. SDB4 in ESWM at	
	30°C	47
4.9	: The growth of Lactococcus sp. SDB4 measured by dry	
	cell weight.	48
4.10	:Correlation between microbial growth, starch utilization	
	and hydrogen production of Lactococcus sp. SDB4	50

### LIST OF ABBREVIATIONS AND SYMBOLS

ABI	-	Application Binary Interface
ATP	-	Adenosine Triphosphate
В.	-	Bacillus
BLAST	-	Basic Local Alignment Search Tool
bp	-	Base pairs
CaCl2	-	Calcium chloride
CH4	-	Methane
cm	-	Centimeter
CO2	-	Carbon dioxide
Da	-	Dalton
dNTPs	-	Deoxynucleotide triphosphates
DNA	-	Deoxyribunuclei Acid
E value	-	Expected value
EDTA	-	Ethylenediaminetetraacetic Acid
EtBr	-	Ethidium bromide
et al.	-	And others
ESWA	-	Enriched Starch Wastewater Agar
FeCl3.5H2O	-	Iron (III) chloride
g	-	Gram
g/L	-	Gram per Litre
GC-TCD	-	Gas Chromatography Thermal Conductivity
H2	-	Hydrogen gas
HCl	-	Hydrochloric acid
H <sub>2</sub> O	-	Water
$H_2S$	-	Hydrogen Sulfide
kDa	-	Kilo Dalton

kbp -	Kilo base pair
r	Kilo base pan
KJ g <sup>-1</sup> -	Kilo Jole per gram
K <sub>2</sub> HPO <sub>4</sub> -	Dipotassium hydrogen phosphate
KH <sub>2</sub> PO <sub>4</sub> -	Potassium dihydrogen phosphate
L -	Liter
LAB -	Lactic Acid Bacteria
MEGA -	Molecular Evolutionary Genetics Analysis
MgCl2 -	Magnesium chloride
MgSO4.7H2O -	Magnesium sulphate
μ -	Specific growth rate
μL -	Microliter
μg -	Microgram
μm -	Micrometer
µmax -	Maximum growth rate
mL -	Mililiter
mg/L -	Milligram per liter
mM -	Milimolar
М -	Molar
NaCI -	Sodium chloride
NaOH -	Sodium hydroxide
NCBI -	National Center of Biotechnology Information
NH4NO3 -	Ammonium nitrate
ng/ µL -	Nano gram per microliter
nm -	Nano meter
NO <sub>2</sub> -	Nitrogen
OD -	Optical Density
PCR -	Polymerase Chain Reaction
POME -	Palm Oil Mill Effluent
RNA -	Ribonucleic acid
rpm -	Revolutions per minute
rRNA -	Ribosomal ribonucleic acid
RNase -	Ribonuclease
SDB -	Starch Degrading Bacteria

sp.	-	Species
TAE	-	Tris-Acetate electrophoresis buffer
td	-	Doubling time
UV	-	Ultraviolet
V	-	Voltage
VFA	-	Volatile fatty acids
v/v	-	Volume per volume
w/v	-	Weight per volume
α	-	Alpha
β	-	Beta
°C	-	Degree Celsius
%	-	Percent
$\times$ g	-	Times gravity

# LIST OF APPENDICES

APPENDIX	TITLE	PAGE
А	Components of the Enriched starch wastewater agar.	61
В	Starch Standard Curve.	62
С	16S rRNA sequence of Isolate SDB4.	63
D	Starch concentration in the ESWM at different incubation	64
	time.	
Е	Hydrogen production at different incubation time in batch	65
	fermentation.	
F	Dry cell weight at different incubation time.	66

### **CHAPTER 1**

#### **INTRODUCTION**

### **1.1 Background of the Research**

The current global economy and energy needs are being covered by fossil fuels. This has led to hyper consumption of these non-renewable resources which in turn has resulted in both the increase in CO<sub>2</sub> concentration in the atmosphere as well as rapid depletion of fossil fuels. The combustion of fossil fuels and its by-product contribute to the increase emission of greenhouse gas (CO<sub>2</sub>, NO<sub>2</sub>, CH<sub>4</sub> and other chemicals) which is responsible for the current global warming and climate change (Show *et al.*, 2012). For these reasons, a novel and safe energy carriers are introduced that substitute fossil fuels. Among the various alternative sources developed, hydrogen is considered the cleanest of all renewable energy source and the most promising fuel with high energy content (142KJ g<sup>-1</sup>) (Rosales-Colunga and Rodríguez, 2014; Thakur *et al.*, 2014)

Hydrogen is considered to be the inspiring fuels for the environment and economy because it produces a non-toxic substance (water) upon combustion and provide all the requirements for a clean and renewable fuel (Ramprakash and Muthukumar, 2014). In addition, hydrogen is safer to handle than domestic natural gas and it can be used directly in internal combustion engines or in fuel cell to generate electricity (Boboescu *et al.*, 2014). Despite the fact that hydrogen has many

social, economic and environmental benefits, but (Kotay and Das, 2008) revealed that the major problem encounter in the utilization of hydrogen gas as a fuel is its non-availability in nature and its production is very expensive.

### 1.2 Production of Hydrogen

Conventionally, hydrogen is produced in different forms of processes which includes electrolysis, photolysis, thermochemical and thermolysis process of water (Audu and Wahab, 2014). (Keskin and Hallenbeck, 2012) revealed that hydrogen can also be produced from fossil fuels by steam reforming or thermal cracking of natural gas, partial oxidation of hydrocarbons and coal gasification or pyrolysis. However, these processes are very expensive, mostly requires an external energy source for the process to function and also not constantly eco-friendly (Kanso *et al.*, 2011).

In the last few years, attention has shifted towards the novel and less energy intensive technologies for the hydrogen production. Biological production of hydrogen arises as an ideal way for hydrogen production due to its flexibility, low energy demand, low cost as its requires organic wastes for the process and also very environmental friendly (Kim *et al.*, 2011).

The biohydrogen production not only provide a clean fuel, but also provide the reduction of environmental pollution from the uncontrolled degradable organic waste (Keskin and Hallenbeck, 2012). As reported by (Rosales-Colunga and Rodríguez, 2014) 40% of hydrogen is produced from natural gases, 30% from heavy oil and naphtha, 18% from coal, 4% from electrolysis and only 1% from biomass.

#### 1.3 Applications of Hydrogen

Hydrogen is a colourless, odourless and non-toxic gas, and also a promising fuel with high energy content which is said to be 2.75 times higher energy yield than hydrocarbon fuels(Ramprakash and Muthukumar, 2014). It has a high calorific value mass  $122 \text{ KJ g}^{-1}$  which is significantly higher than gasoline (Patel *et al.*, 2015).

Hydrogen is widely used in many industries; it is used for the production of ammonia and methanol as well as hydrogenation of fats and oils in the food industry and production of electronic devices (Rosales-Colunga and Rodríguez, 2014). Refineries used it in steel processing, reformulation of gasoline and production of compounds with low molecular weight (Audu and Wahab, 2014; Kapdan and Kargi, 2006).

In the transportation sector, hydrogen also plays a vital role where it can be used as a fuel for vehicles due to its non-polluting nature thereby reducing significantly green house gas emission (Audu and Wahab, 2014; Show *et al.*, 2011). Among the electrical generators, hydrogen serves as an oxygen scavengers and a coolant which can be used in the generation of electricity by converting chemical energy into electrical energy. In addition, hydrogen have a high "low heating value" (LHV) which can be used in fuels cells as well as internal combustion engines (Patel, *et al.*, 2015).

### 1.4 Problem Statement

Over the years fossil fuels have been the main source of energy and global economy. The massive use of these fossil fuels has not only brought environmental threats and health problems, it has also resulted in energy shortage globally and emission of green house gases like carbon dioxide ( $CO_2$ ), nitrogen ( $NO_2$ ) and methane. The emission of these gases has contributed to the current global warming and climate change.

To overcome the global warming and energy shortage crisis, there is a need to search for alternative renewable energy sources. Hydrogen is considered as one of the promising fuel in the future for its renewable energy sources, low cost and inexhaustible. Microbial hydrogen production through fermentation using waste materials is an attractive option as the waste materials are readily available and the fermentation process is technically feasible. Several studies have been done on the biohydrogen production using different substrate but the relatively low yield of hydrogen, cost of production and unstable hydrogen production are the major challenges of biohydrogen production (Show, *et al.*, 2012). The capability of starch utilizing bacteria to hydrolyze starch in wastewater and sludge and produce hydrogen via anaerobic fermentation provides an advantage for cost competitive and sustainable hydrogen production. Hence, this underscores the need to identify and characterize the bacterial strains that can utilize starch efficiently with a hydrogen production potential.

#### **1.5** Research Objectives

This study was designed to address the following objectives:

- (1) To isolate starch degrading bacteria from cassava chips industrial waste
- (2) To screen for starch utilizing ability qualitatively
- (3) To identify phylogenetic characteristics of the selected isolate based on molecular techniques.

### **1.6** Significance of the Research

The increasing public concerns on the environmental problems and energy crisis has urged the development of an alternative clean fuel to substitute the conventional fossil fuels, in which biohydrogen is among the promising ones. Therefore, starch utilizing bacteria isolated from cassava chips processing wastewater and sludge is identified and characterized using 16S rRNA sequencing. The preliminary study conducted here will provide an insight for possibility of hydrogen gas production via dark fermentation by using starch-based waste materials as substrate. From the isolation and identification of the bacteria isolated, further studies on the hydrogen-producing ability of the bacteria can be carried out, allowing the hydrogen gas to be commercially available as clean fuel in the future.

### **1.7** Scope of the Research

This study focused on the isolation of starch degrading bacteria from cassava chips processing industrial waste sample (wastewater and sludge) in enriched starch wastewater media. The isolates obtained were screened for starch utilization and 16S rRNA sequence analysis. The isolates will then be evaluated for hydrogen production using gas chromatography (GC) equipped with thermal conductivity detector (TCD).

### REFERENCES

- Abdeshahian, P., Al-Shorgani, N. K. N., Salih, N. K., Shukor, H., Kadier, A., Hamid, A. A. and Kalil, M. S. (2014). The production of biohydrogen by a novel strain Clostridium sp. YM1 in dark fermentation process. *International Journal of Hydrogen Energy*. 39(24), 12524-12531.
- Adessi, A. and De Philippis, R. (2012). Hydrogen production: photofermentation *Microbial Technologies in Advanced Biofuels Production* (pp. 53-75).
- Aiyer, P. V. (2005). Amylases and their applications. African Journal of Biotechnology. 4(13).
- Alvarado-Cuevas, Z. D., López-Hidalgo, A. M., Ordoñez, L. G., Oceguera-Contreras, E., Ornelas-Salas, J. T. and De León-Rodríguez, A. (2014).
  Biohydrogen production using psychrophilic bacteria isolated from Antarctica. *International Journal of Hydrogen Energy*.
- Audu, J. O. and Wahab, M. F. A. (2014). Biohydrogen production by bacteria isolated from manures of three different bovines using synthetic starch wastewater as substrate. *Malaysian Journal of Microbiology*. 10(1), 38-47.
- Axelsson, L. (2004). Lactic acid bacteria: classification and physiology. *Food Science And Technology-New York-Marcel Dekker-*. 139, 1-66.
- Balat, M. (2008). Potential importance of hydrogen as a future solution to environmental and transportation problems. *International Journal of Hydrogen Energy*. 33(15), 4013-4029.
- Bao, M., Su, H. and Tan, T. (2012). Biohydrogen Production by Dark Fermentation of Starch Using Mixed Bacterial Cultures of Bacillus sp and Brevumdimonas sp. *Energy & Fuels*. 26(9), 5872-5878.

- Barnwal, B. and Sharma, M. (2005). Prospects of biodiesel production from vegetable oils in India. *Renewable and Sustainable Energy Reviews*. 9(4), 363-378.
- Berry, V. and Gascuel, O. (2000). Inferring evolutionary trees with strong combinatorial evidence. *Theoretical computer science*. 240(2), 271-298.
- Beveridge, T. J. (1999). Structures of gram-negative cell walls and their derived membrane vesicles. *Journal of bacteriology*. 181(16), 4725-4733.
- Bhanwar, S. and Ganguli, A. (2014). α-amylase and β-galactosidase production on Potato starch waste by Lactococcus lactis subsp lactis isolated from pickled yam. *Journal of Scientific & Industrial Research*. 73(5).
- Biliaderis, C. (1998). Structures and phase transitions of starch polymers. *ChemInform*. 29(47).
- Boboescu, I. Z., Ilie, M., Gherman, V. D., Mirel, I., Pap, B., Negrea, A., Kondorosi, É., Bíró, T. and Maróti, G. (2014). Revealing the factors influencing a fermentative biohydrogen production process using industrial wastewater as fermentation substrate. *Biotechnology for Biofuels*. 7(1), 139.
- Boesenberg-Smith, K. A., Pessarakli, M. M. and Wolk, D. M. (2012). Assessment of DNA yield and purity: an overlooked detail of PCR troubleshooting. *Clinical Microbiology Newsletter*. 34(1), 1-6.
- Bohak, I., Back, W., Richter, L., Ehrmann, M., Ludwig, W. and Schleifer, K. H. (1998). Lactobacillus amylolyticus sp. nov., isolated from beer malt and beer wort. *Systematic and Applied Microbiology*. 21(3), 360-364.
- Buléon, A., Colonna, P., Planchot, V. and Ball, S. (1998). Starch granules: structure and biosynthesis. *International Journal of Biological Macromolecules*. 23(2), 85-112.
- Cardoso, V., Romão, B. B., Silva, F. T., Santos, J. G., Batista, F. R. and Ferreira, J.S. (2014). Hydrogen Production by Dark Fermentation.
- Chong, M.-L., Rahim, R. A., Aziz, S. A., Shirai, Y. and Hassan, M. A. (2009). Optimization of biohydrogen production by Clostridium butyricum EB6 from palm oil mill effluent using response surface methodology. *International Journal of Hydrogen Energy*. 34(17), 7475-7482.
- Claus, D. (1992). A standardized Gram staining procedure. World journal of Microbiology and Biotechnology. 8(4), 451-452.

- Collins, P. M. and Ferrier, R. J. (1996). *Monosaccharides: their chemistry and their roles in natural products*. John Wiley & Sons.
- Davidson, B., Llanos, R., Cancilla, M., Redman, N. and Hillier, A. (1995). Current research on the genetics of lactic acid production in lactic acid bacteria. *International Dairy Journal*. 5(8), 763-784.
- De Gioannis, G., Muntoni, A., Polettini, A. and Pomi, R. (2013). A review of dark fermentative hydrogen production from biodegradable municipal waste fractions. *Waste management*. 33(6), 1345-1361.
- Dutta, T., Das, A. K. and Das, D. (2009). Purification and characterization of [Fe]hydrogenase from high yielding hydrogen-producing strain, Enterobacter cloacae IIT-BT08 (MTCC 5373). *International Journal of Hydrogen Energy*. 34(17), 7530-7537.
- French, D. (1973). Chemical and physical properties of starch. *Journal of Animal Science*. 37(4), 1048-1061.
- Ghimire, A., Frunzo, L., Pontoni, L., d'Antonio, G., Lens, P. N., Esposito, G. and Pirozzi, F. (2015). Dark fermentation of complex waste biomass for biohydrogen production by pretreated thermophilic anaerobic digestate. *Journal of Environmental Management*. 152, 43-48.
- Giraud, E., Brauman, A., Keleke, S., Lelong, B. and Raimbault, M. (1991). Isolation and physiological study of an amylolytic strain of Lactobacillus plantarum. *Applied Microbiology and Biotechnology*. 36(3), 379-383.
- Hay, J. X. W., Wu, T. Y. and Juan, J. C. (2013). Biohydrogen production through photo fermentation or dark fermentation using waste as a substrate: overview, economics, and future prospects of hydrogen usage. *Biofuels, Bioproducts* and Biorefining. 7(3), 334-352.
- Hongoh, Y., Yuzawa, H., Ohkuma, M. and Kudo, T. (2003). Evaluation of primers and PCR conditions for the analysis of 16S rRNA genes from a natural environment. *FEMS Microbiology Letters*. 221(2), 299-304.
- Idi, A., Nor, M. H. M., Wahab, M. F. A. and Ibrahim, Z. (2014). Photosynthetic bacteria: an eco-friendly and cheap tool for bioremediation. *Reviews in Environmental Science and Bio/Technology*. 1-15.
- Kanso, S., Dasri, K., Tingthong, S. and Watanapokasin, R. Y. (2011). Diversity of cultivable hydrogen-producing bacteria isolated from agricultural soils, waste

water sludge and cow dung. *International Journal of Hydrogen Energy*. 36(14), 8735-8742.

- Kapdan, I. K. and Kargi, F. (2006). Bio-hydrogen production from waste materials. *Enzyme and Microbial Technology*. 38(5), 569-582.
- Keskin, T. and Hallenbeck, P. C. (2012). Enhancement of biohydrogen production by two-stage systems: dark and photofermentation *Biomass Conversion* (pp. 313-340)Springer.
- Kim, J. Y., Jo, B. H. and Cha, H. J. (2011). Production of biohydrogen by heterologous expression of oxygen-tolerant Hydrogenovibrio marinus [NiFe]hydrogenase in Escherichia coli. *Journal of Biotechnology*. 155(3), 312-319.
- Kitahata, S., Tsuyama, N. and Okada, S. (1974). Purification and some properties of cyclodextrin glycosyltransferase from a strain of Bacillus species. *Agricultural and Biological Chemistry*. 38(2), 387-393.
- Kolbert, C. P. and Persing, D. H. (1999). Ribosomal DNA sequencing as a tool for identification of bacterial pathogens. *Current opinion in Microbiology*. 2(3), 299-305.
- Konsoula, Z. and Liakopoulou-Kyriakides, M. (2007). Co-production of α-amylase and β-galactosidase by Bacillus subtilis in complex organic substrates. *Bioresource Technology*. 98(1), 150-157.
- Kotay, S. M. and Das, D. (2007). Microbial hydrogen production with Bacillus coagulans IIT-BT S1 isolated from anaerobic sewage sludge. *Bioresource Technology*. 98(6), 1183-1190.
- Kotay, S. M. and Das, D. (2008). Biohydrogen as a renewable energy resource prospects and potentials. *International Journal of Hydrogen Energy*. 33(1), 258-263.
- Levin, D. B., Pitt, L. and Love, M. (2004). Biohydrogen production: prospects and limitations to practical application. *International journal of hydrogen energy*. 29(2), 173-185.
- Lin, Y.-H., Juan, M.-L. and Hsien, H.-J. (2011). Effects of temperature and initial pH on biohydrogen production from food-processing wastewater using anaerobic mixed cultures. *Biodegradation*. 22(3), 551-563.
- Liu, D., Angelidaki, I., Zeng, R. J. and Min, B. (2008). *Bio-hydrogen production by dark fermentation from organic wastes and residues*. Technical University of

DenmarkDanmarks Tekniske Universitet, Department of Systems BiologyInstitut for Systembiologi.

- Liu, G. and Shen, J. (2004). Effects of culture and medium conditions on hydrogen production from starch using anaerobic bacteria. *Journal of Bioscience and Bioengineering*. 98(4), 251-256.
- Ma, Y., Huang, A., Zhu, D., Pan, G. and Wang, G. (2015). Biohydrogen production via the interaction of nitrogenase and anaerobic mixed-acid fermentation in marine bacteria. *International Journal of Hydrogen Energy*. 40(1), 176-183.
- McKinney, R. E. (2004). Environmental Pollution Control Microbiology: a fifty-year perspective. CRC Press.
- Mishra, S. and Behera, N. (2008). Amylase activity of a starch degrading bacteria isolated from soil receiving kitchen wastes. *African Journal of Biotechnology*. 7(18).
- Morlon-Guyot, J., Guyot, J., Pot, B., De Haut, I. J. and Raimbault, M. (1998). Lactobacillus manihotivorans sp. nov., a new starch-hydrolysing lactic acid bacterium isolated during cassava sour starch fermentation. *International Journal of Systematic Bacteriology*. 48(4), 1101-1109.
- Murray, R. K., Granner, D., Mayes, P. and Rodwell, V. (1990). Harper's Biochemistry. Englewood Cliffs, New Jersey.
- Newman, M. M., Feminella, J. W. and Liles, M. R. (2010). Purification of genomic DNA extracted from environmental sources for use in a polymerase chain reaction. *Cold Spring Harbor Protocols*. 2010(2), pdb. prot5383.
- Nitipan, S., Mamimin, C., Intrasungkha, N., Birkeland, N. K. and Sompong, O. (2014). Microbial community analysis of thermophilic mixed culture sludge for biohydrogen production from palm oil mill effluent. *International Journal* of Hydrogen Energy. 39(33), 19285-19293.
- Nwankwo, D., Anadu, E. and Usoro, R. (1989). Cassava-fermenting organisms. MIRCEN Journal of Applied Microbiology and Biotechnology. 5(2), 169-179.
- Olsen, G. J., Lane, D. J., Giovannoni, S. J., Pace, N. R. and Stahl, D. A. (1986). Microbial ecology and evolution: a ribosomal RNA approach. Annual Reviews in Microbiology. 40(1), 337-365.
- Patel, A. K., Debroy, A., Sharma, S., Saini, R., Mathur, A., Gupta, R. and Tuli, D. K. (2015). Biohydrogen production from a novel alkalophilic isolate Clostridium sp. IODB-O3. *Bioresource Technology*. 175, 291-297.

- Petrov, K., Urshev, Z. and Petrova, P. (2008). L (+)-Lactic acid production from starch by a novel amylolytic Lactococcus lactis subsp. lactis B84. Food Microbiology. 25(4), 550-557.
- Phankhamla, P., Sawaengkaew, J., Buasri, P. and Mahakhan, P. (2014). Biohydrogen production by a novel thermotolerant photosynthetic bacterium Rhodopseudomonas pentothenatexigens strain KKU-SN1/1. *International Journal of Hydrogen Energy*. 39(28), 15424-15432.
- Pigman, W. (2012). The carbohydrates: chemistry and biochemistry. Elsevier.
- Ramprakash, B. and Muthukumar, K. (2014). Comparative study on the production of biohydrogen from rice mill wastewater. *International Journal of Hydrogen Energy*. 39(27), 14613-14621.
- Reddy, G., Altaf, M., Naveena, B., Venkateshwar, M. and Kumar, E. V. (2008). Amylolytic bacterial lactic acid fermentation—a review. *Biotechnology Advances*. 26(1), 22-34.
- Redwood, M. D., Mikheenko, I. P., Sargent, F. and Macaskie, L. E. (2008). Dissecting the roles of Escherichia coli hydrogenases in biohydrogen production. *FEMS Microbiology Letters*. 278(1), 48-55.
- Rosales-Colunga, L. M. and Rodríguez, A. D. L. (2014). Escherichia coli and its application to biohydrogen production. *Reviews in Environmental Science* and Bio/Technology. 14(1), 123-135.
- Samaržija, D., Antunac, N. and Havranek, J. L. (2001). Taxonomy, physiology and growth of Lactococcus lactis: a review. *Mljekarstvo*. 51(1), 35-48.
- Scheffers, D.-J. and Pinho, M. G. (2005). Bacterial cell wall synthesis: new insights from localization studies. *Microbiology and Molecular Biology Reviews*. 69(4), 585-607.
- Show, K.-Y., Lee, D.-J. and Chang, J.-S. (2011). Bioreactor and process design for biohydrogen production. *Bioresource Technology*. 102(18), 8524-8533.
- Show, K., Lee, D., Tay, J., Lin, C. and Chang, J. (2012). Biohydrogen production: current perspectives and the way forward. *International Journal of Hydrogen Energy*. 37(20), 15616-15631.
- Souza, P. M. d. (2010). Application of microbial α-amylase in industry-A review. *Brazilian Journal of Microbiology*. 41(4), 850-861.

- Stamatelatou, K., Antonopoulou, G. and Michailides, P. (2014). 15-Biomethane and biohydrogen production via anaerobic digestion/fermentation. Advances in Biorefineries, edited by K. Waldron, Woodhead Publishing. 476-524.
- Stiles, M. E. and Holzapfel, W. H. (1997). Lactic acid bacteria of foods and their current taxonomy. *International Journal of Food Microbiology*. 36(1), 1-29.
- Thakur, V., Jadhav, S. and Tiwari, K. (2014). Optimization of different parameters for biohydrogen production by Klebsiella oxytoca ATCC 13182. *Trends in Applied Sciences Research*. 9(5), 229-237.
- Vashist, H., Sharma, D. and Gupta, A. (2013). A review on commonly used biochemical test for bacteria. *Innovare Journal of Life Sciences*. 1(1).
- Vidyalakshmi, R., Paranthaman, R. and Indhumathi, J. (2009). Amylase production on submerged fermentation by Bacillus spp. World Journal of Chemistry. 4(1), 89-91.
- Wang, A.-J., Cao, G.-L. and Liu, W.-Z. (2012). Biohydrogen Production from Anaerobic Fermentation *Biotechnology in China III: Biofuels and Bioenergy* (pp. 143-163).
- Whistler, R. L. and BeMiller, J. N. (1997). *Carbohydrate Chemistry for Food Scientists*. Eagan press.
- Widdel, F. (2007). Theory and measurement of bacterial growth. *Di dalam Grundpraktikum Mikrobiologie*. 4, 1-11.
- Zanganeh, K. E. and Shafeen, A. (2007). A novel process integration, optimization and design approach for large-scale implementation of oxy-fired coal power plants with CO 2 capture. *International Journal of Greenhouse Gas Control*. 1(1), 47-54.