

IN VITRO EVALUATION AND WOUND HEALING CREAM FORMULATION
OF *Acalypha Indica Linn* ETHANOLIC EXTRACT

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To my father may Allah bless and have mercy upon his soul. To my beloved mother for her prayers. To my brothers for their support.

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ABSTRACT

Plant-based drugs were used by ancient civilizations for treating various diseases. It is shown scientifically in recent studies that some remedies are effective. *Acalypha indica* Linn, is an annual shrub known in Malaysia as 'Kucing Galak'. It is traditionally used by practitioner for wound-healing. In this study, *Acalypha indica* Linn aerial parts and roots were extracted using 30% ethanol. The extracts were standardized based on protein and polysaccharide content prior to further biochemical and cell tests. Aerial parts extract exhibited higher antioxidant ability compared to roots extract with IC₅₀ of 62 µg/mL and 206 µg/mL, respectively. In cell studies, *In vitro* cytotoxicity was carried out on human skin fibroblast cells (HSF 1189). Both aerial parts and roots extracts showed low cytotoxicities towards (HSF 1189) with 753 µg/mL LD₅₀ for aerial parts extract and undetected on root extract. Further investigation on wound healing showed higher wound closure percentage for both extracts compared to untreated control with 75% at 1 µg/mL for aerial parts, 70% at 0.1 µg/mL for root extract and only 59% closure percentage for untreated control after 48h of the study. Finally, the extracts were formulated into oil-in-water (O/W) cream and then the physicochemical properties such as stability, viscosity and pH were determined.

ABSTRAK

Ubatan berasaskan tumbuhan telah digunakan oleh orang dahulu kala untuk megubati pelbagai penyakit. Kajian saintifik terbaru telah menunjukkan keberkesanan ubatan-ubatan tersebut. *Acalypha indica* Linn ialah tumbuhan renek yang lebih dikenali di Malaysia sebagai 'Kucing Galak'. Tumbuhan ini telah digunakan sebagai ubatan tradisional untuk penyembuhan luka. Dalam kajian ini, bahagian batang dan daun, dan akar pokok *Acalypha indica* Linn telah diekstrak menggunakan 30% etanol. Sebelum menjalankan ujian biokimia dan sel, ekstrak ini telah diselaras berdasarkan kandungan protin dan polisakarida. Ekstrak bahagian batang dan daun mempunyai antioksidan yang lebih tinggi berbanding ekstrak bahagian akar dengan masing-masing menunjukkan nilai IC_{50} 62 $\mu\text{g/mL}$ dan 206 $\mu\text{g/mL}$. Ujian toksik telah dijalankan keatas sel kulit manusia fibroblast (HSF 1189). Ekstrak bahagian batang dan daun menunjukkan nilai toksik terhadap sel HSF 1189 yang rendah dengan nilai LD_{50} 753 $\mu\text{g/mL}$. Ekstrak akar pokok pula tidak menunjukkan sebarang toksik terhadap sel HSF 1189. Ujian lanjut keatas penyembuhan luka menunjukkan bahawa ekstrak bahagian batang dan daun, dan akar pokok mempunyai peratusan penutupan luka yang tinggi berbanding kumpulan yang tidak menerima ekstrak. Masing-masing menunjukkan nilai penutupan luka 75% pada kepekatan 1 $\mu\text{g/mL}$ untuk ekstrak bahagian batang dan daun, 70% pada kepekatan 0.1 $\mu\text{g/mL}$ untuk ekstrak akar pokok dan 59% untuk kumpulan yang tidak menerima sebarang ekstrak. Yang terakhir, ekstrak ini diformulasi kepada krim minyak-dalam-air (O/W) dan diuji dengan ciri-ciri fizikokimia seperti kestabilan, kelekutan dan pH.

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LIST OF ABBREVIATION/ SYMBOLS

Abc	- Absorbance
ANOVA	- Analysis of Variance
BFF	- Basic Fibroblast Factor
°C	- Degree Celsius
CO ₂	- Carbon Dioxide
DMEM	- Dulbecco's Modified Eagle's Medium
DMSO	- Dimethylsulfoxide
DPPH	- 2,2-diphenyl-1-picrylhydrazyl
ECM	- Extracellular Matrix
EGF	- Epidermal Growth Factor
FBS	- Foetal Bovine Serum
FGF	- Fibroblast Growth Factor
g	- Gram
h	- Hour
H ₂ SO ₄	- Sulfuric Acid
HCl	- Hydrochloric Acid
HSF 1184	- Human Skin Fibroblast
IC ₅₀	- 50% Inhibitory Concentration
LD ₅₀	- 50% Lethal Concentration
mg	- Milligram
mg/L	- Milligram per Liter

min	- Minute
MMPs	- Matrix Metalloproteinases
MTT	- Methyl Tetrazolium
nm	- Nanometer
PBS	- Phosphate Buffered Saline
PDGF	- platelet-derived Growth Factor
ROS	- Reactive Oxygen Species
TGF	- Transforming Growth Factor
UV	- Ultraviolet
pH	- Hydrogen Concentration
Pen Strep	- Penicillin Streptomycin mixtures
RPM	- Revolutions Per Minute
SD	- Standard Deviation
SEM	- Standard Error of the Mean
SPSS	- Statistical Package for the Social Sciences
VEGF	- Vascular Endothelial Growth Factor
μg	- Microgram
μL	- Microliter
WHO	- World Health Organization
%	- Per cent
>	- Greater Than
<	- Less Than

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CHAPTER 1

INTRODUCTION

1.1 Study Background

Prior to this time, the healing process was considered to be a passive process with respect to the physician (Boateng *et al.*, 2008). This theory was summed up by Ambroise Pare, who said "I dressed the wound, God heals it" (Keynes, 1951). The healing process was firstly studied by Winter, and others in 1960s (winter and Scales, 1963), since then much more information has been revealed about cellular and biochemical components of wound healing and factors affecting this process. It is now known that healing is not a passive process, but rather can be accelerated and enhanced by the use of specific wound care/dressing techniques and products (Boateng *et al.*, 2008).

For hundreds of years, plant-based treatments were used by ancient civilizations for curing diseases. Recently most of the research conducted in traditional medicines has shown that some traditional remedies are effective in spite of the fact that there is no clear justification. The greater part of the traditional therapy involves the use of plant extracts on their active principle (WHO, 1993).

Acalypha indica Linn is a herb traditionally used in wound healing management (Upadhyay *et al.*, 2007). New researches conducted on this herb *in vivo* have found evidences that support the wound healing properties of this herb (Reddy *et al.*, 2002; Ganeshkumar *et al.*, 2012). The anti-oxidant properties and the protein

and polysaccharide content of ethanolic extract of this herb have major contribution in wound healing properties. Formulating wound healing cream from the extract of *Acalypha indica* Linn would supplement the wound healing products available in the market with new promising biopharmaceutical product with an active wound healing properties that accelerate the wound healing process.

1.2 Problem Statement

Patients suffering from chronic wounds like diabetes, vascular disease, and pressure ulcer are increasing in numbers. According to American Diabetes Association (2013), 25.8 million Americans suffer from diabetes. Likewise in 2012, two studies conducted by Vascular Disease Foundation and The National Pressure Ulcer Advisory Panel respectively showed that half a million Americans have ulcers on their legs caused by diseased veins, and 2.5 million US residents develop pressure ulcers every year. In Malaysia, according to world health organization 942,000 Malaysian diagnosed with diabetes in 2000 and the number expected to increase to be 2.479.000 in 2030 WHO (2000). A recent study conducted by Mazlina *et al.* (2011) showed that 47.1% of diabetic patient would develop foot ulcers.

Traditional wound management products including cotton wool, natural or synthetic bandages and gauzes do not provide the optimum environment to facilitate the healing process; they only dress the wound and have no biological activity to accelerate wound healing.

In this study, the formulated cream from *Acalypha indica* Linn extracts made use of the wound healing properties of this herb especially the anti-oxidant (Ganeshkumar *et al.*, 2012). The healing process of the ethanolic extract was evaluated *In vitro* on human skin fibroblast cell line (HSF 1189). On the other hand, the moisture provided by the cream itself accelerates the healing process (winter and Scales, 1963). cream will also act as a barrier preventing infection caused by bacteria (Boateng *et al.*, 2008).

1.3 Significance of The Study

Herbs based biopharmaceutical products can be potential agents for wounds healing and are in great demand due to their wide availability, less toxicity, less side effect and cost effective. *Acalypha indica* is a common herb in tropical regions; it is also used traditionally in wound healing management.

Formulating *Acalypha indica* extract in the form of cream will carry the active ingredient of the extract. On the other hand, cream will prevent infection that might happen when applying plants, directly, or as a crude extract, as the crude extract might act as potential source for microorganisms.

1.4 Objectives of The Study

1. To extract and standardize the *Acalypha indica* aerial parts and roots with 30% ethanol, based on protein and polysaccharide content.
2. To evaluate the anti-oxidant properties of *Acalypha indica* extract.
3. To evaluate *In vitro* cytotoxicity activity and wound healing properties of *Acalypha indica* extracts on fibroblast (HSF 1189) cell line.
4. To formulate a wound healing cream comprising *Acalypha indica* extracts into oil-in water (O/W) cream.

1.5 Scope of The Study

This study were conducted to standardize the ethanolic extract based on protein and polysaccharide content, evaluate the anti-oxidant properties of the ethanolic extract of *Acalypha indica*, evaluate the healing process of the extract *In*

vitro on (HSF 1189) Fibroblast cell line, and formulate a wound healing cream from this extract. All results were statistically analysed.

REFERENCES

- Amarowicz, R., Pegg, R. B., Rahimi-Moghaddam, P., Barl, B., & Weil, J. A. (2004). Free-radical Scavenging Capacity and Antioxidant Activity of Selected Plant Species from the Canadian Prairies. *Food Chemistry*, 84(4), 551-562.
- American diabetes dssociation. <http://www.diabetes.org/>
- Amresh, G., Singh, P. N., & Rao, C. V. (2007). Antinociceptive and Antiarthritic Activity Of Cissampelos Pareira Roots. *Journal of Ethnopharmacology*, 111(3), 531-536.
- Aswal, A., Kalra, M., & Rout, A. (2013). Preparation and Evaluation of Polyherbal Cosmetic Cream. *Der Pharmacia Lettre*, 5(1), 83-88.
- Ayyanar, M., (2009). Herbal Medicines for Wound Healing Among Tribal People in Southern India. *Ethnobotanical and Scientific evidences*, 2 (3), 29-42.
- Azlim Almey, A. A., Ahmed Jalal Khan, C., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R., & Kamarul Rahim, K. (2010). Total Phenolic Content and Primary Antioxidant Activity of Methanolic and Ethanolic Extracts of Aromatic Plants Leaves. *International Food Research Journal*, 17(4).
- Barile, F. A., Dierickx, P. J., & Kristen, U. (1994). *In vitro* Cytotoxicity Testing for Prediction of Acute Human Toxicity. *Cell biology and Toxicology*, 10(3), 155-162.
- Beldon, P., (2010). *Basic Science of Wound Healing*. *Surgery (Oxford)*, 28(9), 409-412.
- Biswas, T. K., & Mukherjee, B. (2003). Plant Medicines of Indian Origin For Wound Healing Activity: a Review. *The International Journal of Lower Extremity Wounds*, 2(1), 25-39.
- Boateng, J. S., Matthews, K. H., Stevens, H. N., & Eccleston, G. M. (2008).

Wound Healing Dressings and Drug Delivery Systems: a Review. *Journal of Pharmaceutical Sciences*, 97(8), 2892-2923.

Brighente, I.M.C., Dias, M., Verdi, L.G., & Pizzolatti, M.G., (2007). Antioxidant Activity and Total Phenolic Content of Some Brazilian Species. *Pharmaceutical Biology*, 45 (2), 156–161.

Dhar, M. L., Dhar, M. M., Dhawan, B. N., Mehrotra, B. N., & Ray, C. (1968). Screening of Indian Plants for Biological Activity: Part I. *Indian Journal of Experimental Biology*, 6, 232-247.

Enoch, S. and Leaper, D.J., (2007). *Basic Science of Wound Healing*. Surgery (Oxford).

Evangeline, S., Sundaram, V., Manian, R.P., Kulanthaivelu, K., & Balasundaram, S., (2015). Antioxidant, Antibacterial and Anti-Inflammatory Activity of *Acalypha indica* and *Terminalia chebula*: an *In-Vitro* Analysis. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, 6(3), 388–396.

Falanga, V., (2005). Wound Healing and its Impairment in the Diabetic Foot. *Lancet*, 366(9498), 1736–1743.

Franco, D., Sineiro, J., Rubilar, M., Sánchez, M., Jerez, M., Pinelo, M., & Núñez, M. J. (2008). Polyphenols from Plant Materials: Extraction and Antioxidant Power. *Electron. J. Environ. Agric. Food Chem*, 7, 3210-3216.

Ganeshkumar, M., Ponrasu, T., Krithika, R., Iyappan, K., Gayathri, V.S., & Suguna, L., (2012). Topical Application of *Acalypha Indica* Accelerates Rat Cutaneous Wound Healing by Up-regulating The Expression of Type I and III Collagen. *Journal of ethnopharmacology*, 142(1), 14–22.

Ghasemi, A. and Zahediasl, S., (2012). Normality Tests for Statistical Analysis: a Guide for Non-Statisticians. *International Journal of Endocrinology and Metabolism*, 10(2), 486–489.

Green, R. J. (2004). Antioxidant Activity of Peanut Plant Tissues. Master's

Thesis. NC State University.

- Guo, S. and Dipietro, L. a, (2010). Factors Affecting Wound Healing. *Journal of Dental Research*, 89(3), 219–229.
- Gurib-Fakim, A., Sewraj, M., Gueho, J., & Dulloo, E., (1993). Medicaethnobotany of Some Weeds of Mauritius and Rodrigues. *Journal of Ethnopharmacology*, 39(3), 175-185.
- Gurtner, G. C., Werner, S., Barrandon, Y., & Longaker, M. T. (2008). Wound Repair and Regeneration. *Nature*, 453(7193), 314-321.
- Hiremath, S. P., Rudresh, K., Badami, S., Patil, S. B., & Patil, S. R. (1999). Post-Coital Antifertility Activity of *Acalypha indica* L. *Journal of Ethnopharmacology*, 67(3), 253-258.
- Houghton, P.J., and Raman, A. (1998). *Laboratory Handbook for the Fractionation of Natural Extracts 1st Edition*. New York, NY: Chapman
- Hussain, A.Z. & Ignatius, A., (2010). GC-MS Analysis and Antimicrobial Activity of *Acalypha indica* Linn. *Asian Journal Of Chemistry*, 22(5), 3591–3595.
- Hussain, K., Ismail, Z., Sadikun, A., Ibrahim, P., & Malik, A. (2008). Analysis of Proteins, Polysaccharides, Glycosaponins Contents of *Piper Sarmentosum* Roxb. and Anti-TB Evaluation for Bioenhancing/Interaction Effects of Leaf Extracts with Isoniazid (INH). *Natural Product Radiance*, 7(5), 402-408
- Hutt, M. J., & Houghton, P. J. (1998). A Survey from the Literature of Plants Used to Treat Scorpion Stings. *Journal of Ethnopharmacology*, 60(2), 97-110.
- Keynes, G. (1951). *The apology and treatise of ambrose pare*. Falcon Educational Books: London.
- Koleckar, V., Kubikova, K., Rehakova, Z., Kuca, K., Jun, D., Jahodar, L., & Opletal, L. (2008). Condensed and Hydrolysable Tannins as Antioxidants

- Influencing the Health. *Mini Reviews in Medicinal Chemistry*, 8(5), 436-447.
- Kumar, B., Vijayakumar, M., Govindarajan, R., & Pushpangadan, P. (2007). Ethnopharmacological Approaches to Wound healing—exploring Medicinal Plants of India. *Journal of Ethnopharmacology*, 114(2), 103-113.
- Kumar, K. K., Sasikanth, K., Sabareesh, M., & Dorababu, N. (2011). Formulation and Evaluation of Diacerein Cream. *Asian J Pharm Clin Res*, 4(2), 93-98.
- Kupcsik, L. (2011). Estimation of Cell Number Based on Metabolic Activity: the MTT Reduction Assay in Mammalian Cell Viability (pp. 13-19). Humana Press.
- Lambers, H., Piessens, S., Bloem, A., Pronk, H., & Finkel, P. (2006). Natural Skin Surface Ph is on Average Below 5, Which is Beneficial for its Resident flora. *International Journal of Cosmetic Science*, 28(5), 359-370.
- Liang, C.-C., Park, A.Y., & Guan, J.-L., (2007). *In vitro* Scratch Assay: a Convenient and Inexpensive Method for Analysis of Cell Migration *in vitro*. *Nature Protocols*, 2(2), 329–333.
- Mahishi, P., Srinivasa, B. H., & Shivanna, M. B. (2005). Medicinal Plant Wealth of Local Communities in Some Villages in Shimoga District of Karnataka, India. *Journal of Ethnopharmacology*, 98(3), 307-312.
- Manjunatha, B. K., Vidya, S. M., Krishna, V., Mankani, K. L., Singh, S. D., & Manohara, Y. N. (2007). Comparative Evaluation of Wound Healing Potency of *Vitex Trifolia* L. and *Vitex Altissima* L. *Phytotherapy Research*, 21(5), 457-461.
- Mathivanan, N., Surendiran, G., Srinivasan, K., & Malarvizhi, K. (2006). *Morinda Pubescens* JE Smith (*Morinda Tinctoria* Roxb.) Fruit Extract Accelerates Wound Healing in Rats. *Journal of Medicinal Food*, 9(4), 591-593.

- Mazlina, M., Shamsul, A. S., & Jeffery, F. S. (2011). Health-Related Quality of Life in Patients with Diabetic Foot Problems in Malaysia. *Med J Malaysia*, 66(3), 234-8.
- Montesano, R. & Orci, L., (1988). Transforming Growth Factor Beta Stimulates Collagen-Matrix Contraction by Fibroblasts: Implications for Wound Healing. *Proceedings of the National Academy of Sciences of the United States of America*, 85(13), 4894–4897.
- Nahrstedt, a., Hungeling, M., & Petereit, F., (2006). Flavonoids from *Acalypha indica*. *Fitoterapia*, 77(6), 484–486.
- Nayak, B. S., Udupa, A. L., & Udupa, S. L. (1999). Effect of *Ixora Coccinea* Flowers on Dead Space Wound Healing in Rats. *Fitoterapia*, 70(3), 233-236.
- Nirmal, N., Praba, G.O., & Velmurugan, D., (2008). Modeling Studies on Phospholipase A2-Inhibitor Complexes. *Indian Journal of Biochemistry & Biophysics*, 45(4), 256–62.
- Nigam, A., & Ayyagari, A. (2008). *Lab manual in biochemistry, immunology, biotechnology Tata*. McGraw Hill.
- Owoyele, V. B., Adediji, J. O., & Soladoye, A. O. (2005). Anti-Inflammatory Activity of Aqueous Leaf Extract of *Chromolaena Odorata*. *Inflammopharmacology*, 13(5-6), 479-484.
- Panthong, A., Kanjanapothi, D., Taesotikul, T., & Taylor, W. C. (1991). Ethnobotanical Review of Medicinal Plants from Thai Traditional Books, Part II: Plants with Antidiarrheal, Laxative and Carminative Properties. *Journal of Ethnopharmacology*, 31(2), 121-156.
- Phan, T. T., Hughes, M. A., & Cherry, G. W. (1998). Enhanced Proliferation of Fibroblasts and Endothelial Cells Treated with an Extract of the Leaves of *Chromolaena Odorata* (Eupolin), an Herbal Remedy for Treating Wounds. *Plastic and Reconstructive Surgery*, 101(3), 756-765.

- Pietta, P. G. (2000). Flavonoids as Antioxidants. *Journal of Natural Products*, 63(7), 1035-1042.
- Principe, P. P. (1996). *Monetizing the Pharmacological Benefits of Plants. Medicinal Resources of the Tropical Forest: Biodiversity and its Importance to Human Health*. Columbia University Press, New York, 191-219.
- Prior, R. L., Wu, X., & Schaich, K. (2005). Standardized Methods for the Determination of Antioxidant Capacity and Phenolics in Foods and Dietary Supplements. *Journal of Agricultural and Food Chemistry*, 53(10), 4290-4302.
- Rahman, M.A., Bachar, S.C., & Rahmatullah, M., (2010). Analgesic and Antiinflammatory Activity of Methanolic Extract of *Acalypha indica* Linn. *Pakistan Journal of Pharmaceutical Sciences*, 23(3), 256–258.
- Reddy, J.S., Rao, P.R., & Reddy, M.S., (2002). Wound Healing Effects of *Heliotropium Indicum*, *Plumbago Zeylanicum* and *Acalypha Indica* in Rats. *Journal of Ethnopharmacology*, 79(2), 249–251.
- Raj, J., & Singh, K. P. (2000). *Acalypha indica*. *Central Council for Research in Homeopathy*, 22(1), 1-6.
- Rasik, A. M., & Shukla, A. (2000). Antioxidant Status in Delayed Healing Type of Wounds. *International journal of Experimental Pathology*, 81(4), 257-263.
- Samy, R. P., Thwin, M. M., Gopalakrishnakone, P., & Ignacimuthu, S. (2008). Ethnobotanical Survey of Folk Plants for the Treatment of Snakebites in Southern Part of Tamilnadu, India. *Journal of Ethnopharmacology*, 115(2), 302-312.
- San Miguel, S. M., Opperman, L.A., Allen, E. P., Zielinski, J., & Svoboda, K. K. (2010). Antioxidant Compounds Increased Wound Healing Migration Via Rac-GTP Activation in Nicotine-Treated Human Gingival and PDL

- Fibroblasts. *Journal of Periodontology*, 81(11), 1675-1690.
- Sanseera, D., Niwatananun, W., Liawruangrath, B., Liawruangrath, S., Baramee, A., Trisuwan, K., & Pyne, S.G., (2012). Antioxidant and Anticancer Activities from Aerial Parts of *Acalypha indica* Linn. *Chiang Mai University Journal of Natural Sciences*, 11(2), 157–168.
- Sarker, S. D., Latif, Z., & Gray, A. I. (2005). *Natural Product Isolation*. (pp. 1-25). Humana Press.
- Sawhney, S. K., & Singh, R. (2000). *Introductory Practical Biochemistry*. Alpha Science Int'l Ltd..
- Saxena, M., Saxena, J., Nema, R., Singh, D., & Gupta, A. (2013). Phytochemistry of Medicinal Plants. *Journal of Pharmacognosy and Phytochemistry*.
- Shirwaikar, A., Rajendran, K., Bodla, R., & Kumar, C. D. (2004). Neutralization Potential of Viper Russelli Russellii (Russell's Viper) Venom by Ethanol Leaf Extract of *Acalypha indica*. *Journal of Ethnopharmacology*, 94(2), 267-273.
- Siddiqui, M. B., & Husain, W. (1990). Traditional Antidotes of Snake Poison in Northern India. *Fitoterapia*, 61(1), 41-44.
- Sinha, T., & Bandyopadhaya, A. (2012). Ethno-Pharmacological Importance and Valuable Phytochemicals of *Acalypha indica* (L.) a Review. *International Journal Res Pharm Sci*, 3, 360-368.
- Srinivasan, K., Muruganandan, S., Lal, J., Chandra, S., Tandan, S. K., & Prakash, V. R. (2001). Evaluation of Anti-Inflammatory Activity of *Pongamia Pinnata* Leaves in Rats. *Journal of Ethnopharmacology*, 78(2), 151-157.
- Sultana, B., Anwar, F., & Przybylski, R. (2007). Antioxidant Activity of Phenolic Components Present in Barks of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica*, and *Eugenia jambolana* Lam. Trees. *Food*

Chemistry, 104(3), 1106-1114

Susanti, D., Sirat, H. M., Ahmad, F., Ali, R. M., Aimi, N., & Kitajima, M. (2007). Antioxidant and Cytotoxic Flavonoids from the Flowers of *Melastoma Malabathricum* L. *Food Chemistry*, 103(3), 710-716.

Takle, V.V., Savad, R.V., Kandalkar, A.M., Akarte, A.M., and Patel, A.M., (2011). Pharmacognostic and Phytochemical Investigations of Aerial Parts Of *Acalypha Indica* Linn. *Pharmacognosy Journal*, 3(21), 33–35.

Taufiq-Yap, Y. H., Peh, T. H., Ee, G. C. L., & Rahmani, M. (2000). Chemical Investigation of *Acalypha indica* (Euphorbiaceae). *Oriental Journal of Chemistry*, 16(2), 249-251.

Tromp, J. (1983). Nutrient Reserves in Roots of Fruit Trees, In Particular Carbohydrates and Nitrogen in Tree root systems and their mycorrhizas. In D. Atkinson. *Developments in Plant and Soil Sciences*, (pp. 401-413). Springer Netherlands.

Upadhyay, B., Roy, S., & Kumar, A. (2007). Traditional Uses of Medicinal Plants Among the Rural Communities of Churu District in the Thar Desert, India. *Journal of Ethnopharmacology*, 113(3), 387-399.

Winter, G. D., & Scales, J. T. (1963). Effect of Air Drying and Dressings on the Surface of a Wound. *Nature*, 197, 91 – 92.

WHO <http://www.who.int/>

WHO, IUCN, WWF (1993) Guidelines on the Conservation of Medicinal Plants. P-1 Osler W. Chemotherapy. In: Thomas CG. *Medical microbiology*. 5th edition (1983). Baillier Tindall, London, pp, 192.