

Kinetic Modeling of Supercritical Fluid Extraction of Betel Nut

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ABSTRACT

Supercritical fluid extraction is an advanced extraction technique that suitable for heat sensitive and active compound material from plants and herbs. Understanding the effect of extraction parameters on mass transfer coefficient at solid and fluid phase can determine the dominating extraction regime thus performance of the extraction may be enhanced. The aim of this research was to determine the mass transfer coefficient in solid and fluid phase using kinetic modelling approach. Grounded betel nuts were treated with supercritical carbon dioxide with 5% methanol as co-solvent to determine its mass transfer coefficient in solid and fluid phase for the following extraction conditions; pressure, 20 to 30 MPa; temperature, 50 to 70 °C; and flow rate, 2 to 4 mL/min. Simplified Sovová model was coupled with Broken and Intact Cell model to determine the mass transfer coefficients. Results show the mass transfer coefficients of solid phase and liquid phase are in the ranges of 0.00015 to 0.00353 m³/min and 0.3497 to 3.9623 m³/min, respectively. The overall absolute average relative deviation was observed to be 7.39%.

Keywords: Areca catechu; betel nut; supercritical fluid extraction; kinetic model; mass transfer coefficient.

INTRODUCTION

Supercritical fluid extraction (SFE) is an advanced extraction technique with high selectivity. The selectivity of the process depends on the extraction conditions and solvent used as the extractor. Carbon dioxide is widely chosen as the main solvent in SFE due to its low critical condition, inert, inflammable, and inexpensive. While modifier such as methanol can be added to enhance the selectivity and polarity of the fluid [1]. SFE has successfully been conducted on the extraction of interest and bioactive compounds from palm fatty acid [2], cardoon [3], peppermint [4], pitanga fruits [5], peach [6], dandelion leaves [7], abajeru [8], guava seeds [9], coffee beans [10], neem seeds [11], horsetail [12], jatropha seeds [13], garlic flakes [14], tomato [15], Mucuna seeds, [16], vanilla [17], celery seeds [18], paprika [19], hemp seed [20], and coconut flesh [21].

Areca catechu or betel nut is a plant from the arecaceae family which grows widely in India and south-east Asia region. The seeds are traditionally used for cigarette

and the leaves are used as food packaging [22]. One of the compound of interest that can be obtained from betel nut is catechin, is a polyphenols compound which contains anti-oxidant, anti-depressant, hypoglycaemic, wound healing, and anti-HIV properties [23]. Other sources of catechin from local plants are tea leaves and onion skins [24]. The highest catechin content in betel nut was reported as 0.0844 mg catechin/g sample obtained using accelerated water extraction [25]. Apart from having multiple therapeutic properties, the catechin monomers in the form of tannins give the betel a reddish brown colour that is a promising source of natural colorant [25].

Cross over pressure or retrograde solubility plays an important role in determining the suitable operating condition for several sample of extract. It is a phenomenon in SFE process where the dominant effect of solvation power shift to effect of solute vapor pressure. The cross over pressure varies from one sample to the other. To date, no study on determining the cross over pressure for betel nut has been conducted. Thus, determining the cross over pressure was essential in understanding the mass transfer regime for SFE processes.

The SFE experimental results can be interpreted by equation or model developed based on several assumptions to represent the actual extraction process. The models commonly used to describe the SFE data can be categorized as empirical model and kinetic model. The kinetic model is applied to the extraction profile to study the effect of extraction parameter on the mass transfer coefficient of solute in solid phase and fluid phase. The mostly used models are Sovová model [26] and Broken and Intact Cell [27].

To date, there are no study conducted regarding the solubility of betel nuts extract in supercritical fluid and data for mass transfer coefficient in solid phase and fluid phase was not established. This study aims to determine the conditions for the highest mass transfer coefficient in solid phase and fluid phase to preferably demonstrate the best extraction conditions. The effect of extraction parameters was also discussed.

EXPERIMENTAL SET UP

Materials

Betel nuts were purchased from local market in Pulau Pinang, Malaysia. The sample was dried and ground using hammer-grinder (POLYMIX® PX-MFC 90D, Switzerland) to a desired average particle size (177.5 μm). A 99.9% purity CO_2 was obtained from KRAS Instrument, Malaysia for SFE process. Methanol, AR grade supplied by QRec was used as the modifier for the study.

Method

Pressurized CO_2 gas was introduced into a 1.5 m coil located inside an oven (Memmert) using constant flow pump (Lab Alliance, Series II Pump) to achieve the desired operating temperature. A 50 mL extraction cell loaded with 10 ± 1 grams of sample with known volume of modifier was fitted between the coil and back pressure regulator (JASCO, Model BP-2080). The experimental set up was as per illustrated in Figure 1. The extraction was conducted for 1 hour of extraction regime with every 10 minutes fractionation. One Factor at a Time (OFAT) design was used for the variation of 3 variables with 3 level values as shown in Table 1.

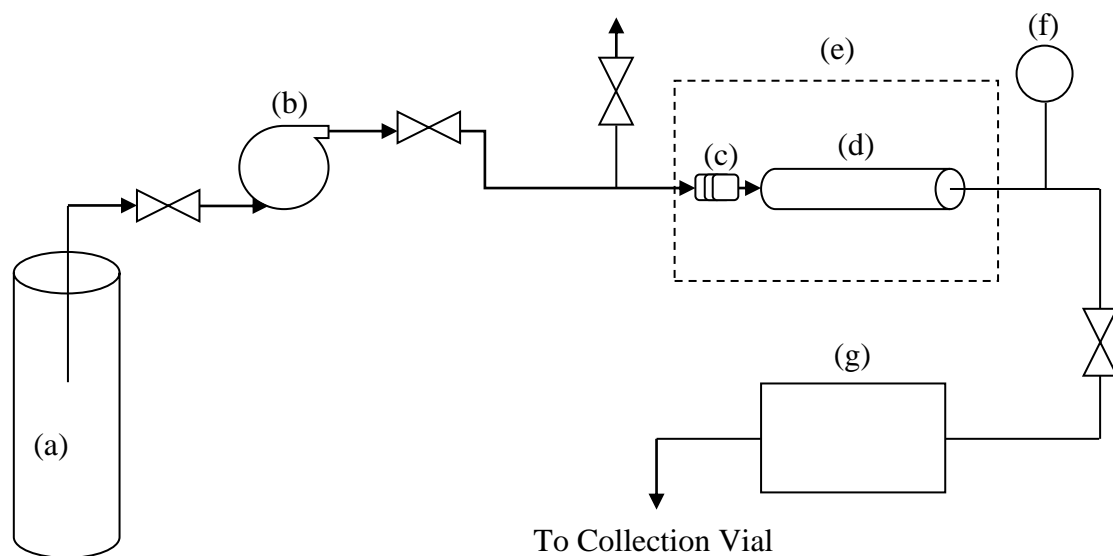


Figure 1: Schematic diagram of SFE apparatus with (a) CO₂ cylinder, (b) CO₂ pump, (c) heated coil, (d) extraction cell, (e) oven, (f) pressure gauge, (g) back pressure regulator [28].

Table 1: Parameters and level values used for the study.

Parameter (unit)	High Value (+1)	Middle Value (0)	Low Value (-1)
Flow Rate (mL/min)	2	3	4
Temperature (°C)	50	60	70
Pressure (MPa)	20	25	30

Kinetic Modelling

Kinetic models were used to fit the extraction profile. The model is generally divided into three equations which represent the three stages of extraction profile. Broken and Intact Cell (BIC) model with 4 adjustable parameters was adapted in this study. Derivation of this model was described elsewhere [29]. The model assumes that the process is isobaric and isothermic, the axial dispersion of solvent in extraction bed is neglected, samples have broken and intact cell, and mass transfer is divided in to solid phase and fluid phase [30]. Extraction regime was divided into three extraction regime namely, equilibrium controlled phase, intermediate phase and diffusion controlled phase. Broken cell is formed due to the grinding effect. High saturation of solute located at the broken cell known as outer layer of sample, while the inner layer of solute located in the intact part of the sample that is not affected or crushed by the grinding process. For kinetic study, the equations for this model are shown in Eq. (1) to Eq. (7).

$$\frac{E}{N_{x_0}} = \begin{cases} \psi [1 - \exp(-Z)] & \text{for } \psi < \frac{G}{Z} \\ \psi - \frac{G}{Z} \exp\left[z(h_k - 1)\right] & \text{for } \frac{G}{Z} \leq \psi \leq \psi_k \\ 1 - \frac{1}{Y} \ln\{1 + [\exp(Y) - 1] \exp\left[Y\left(\frac{G}{Z} - \psi\right)\right] (1 - G)\} & \text{for } \psi \geq \psi_k \end{cases} \quad (1)$$

$$\psi = \frac{tQy_f}{N_{x_0}} \quad (2)$$

$$\psi_k = \frac{G}{Z} + \frac{1}{Y} \left[1 + \frac{\{\exp[Y(\psi - \frac{G}{Z})] - 1\}}{G} \right] \quad (3)$$

$$h_k = \frac{1}{Y} \left[1 + \frac{\{\exp[Y(\psi - \frac{G}{Z})] - 1\}}{G} \right] \quad \text{for } \frac{G}{Z} \leq \psi \leq \psi_k \quad (4)$$

$$Y = \frac{Nk_s a x_0}{Q(1-\varepsilon)y_f} \quad (5)$$

$$Z = \frac{Nk_f a \rho_f}{Q(1-\varepsilon)\rho_s} \quad (6)$$

$$\varepsilon = 1 - \frac{\rho_A}{\rho_S} \quad (7)$$

where E (g) is the extraction yield, N (g) is the weight of solute-free sample, ε is the porosity of sample, ρ_A (kg/m³) is apparent density of sample, ρ_S (kg/m³) is real density of sample, ψ and ψ_k are dimensionless times, h_k is the dimensionless coordinate which separates the equilibrium region and diffusion region, Q (g/min) is solvent flow rate, G , Z and Y are adjustable parameters, and k_{fa} and k_{sa} are mass transfer coefficient (m²/m³.min) in fluid phase and solid phase, respectively. Apparent density was determined by the ratio of sample weight to the volume of extraction cell, while real density was measured by the volume of solvent displaced by known weight of sample. G , Z , Y and ψ_k are the adjustable parameters for BIC model. Reducing the number of adjustable parameters will enhance the capability of the model in improving the relative deviation. Thus, the value of G maybe calculated using simplified Sovová model as shown in Eq. (8) until Eq. (12) [31].

$$\frac{E}{N_m} = \begin{cases} q y_s & 0 \leq q < q_c \\ x_u (1 - C_1 \exp(-C_2 q)) & q \geq q_c \end{cases} \quad (8)$$

$$q = M/N_m \quad (9)$$

$$M = \rho V \quad (10)$$

$$N_m = (1 - c_u) N \quad (11)$$

$$x_u = c_u / (1 - c_u) \quad (12)$$

where q is the relative amount of passed solvent, q_c is the relative amount of passed solvent in the fast extraction period, y_s (g extract/g fluid) is the solubility of extract, x_u is the weight fraction of untreated solid, M (g) is the weight of passed solvent, c_u is the weight fraction of total solute content in sample, ρ is density of fluid in g/mL, V (mL) is the volume of passed solvent, N (g) is the weight of solid sample, and C_1 and C_2 are the adjustable parameters for the simplified Sovová model. After the values of C_1 , C_2 and q_c were obtained, Eq. (13) was used to determine the value of G for each extraction condition.

$$G=1-C_1 \exp \left(\frac{-C_2 q_c}{2} \right) \quad (13)$$

Objective Function

Extraction yield values generated by the models ($y_{\text{calculated}}$) were compared with the experimental data ($y_{\text{experimental}}$). An error minimization method was used to minimize the deviation between the observed and the predicted value. The minimization of Absolute Average Relative Deviation (AARD) value is shown in Eq. (14). The objective was to obtain the smallest possible AARD value rendering a small deviation for the model to be in good agreement with the extraction profile. Thus, the model with the lowest AARD value was chosen as the best model to represent the extraction behavior.

$$\text{AARD} = \frac{1}{N} \sum_{i=1}^N \left| \frac{y_{\text{calculated}} - y_{\text{experimental}}}{y_{\text{experimental}}} \right| \times 100\% \quad (14)$$

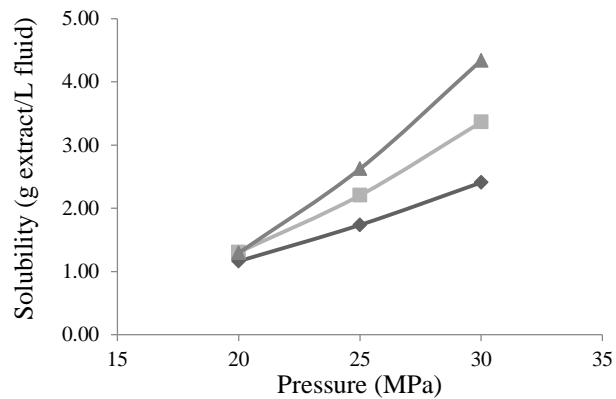
RESULTS AND DISCUSSION

Solubility of Extracts in Supercritical Fluid

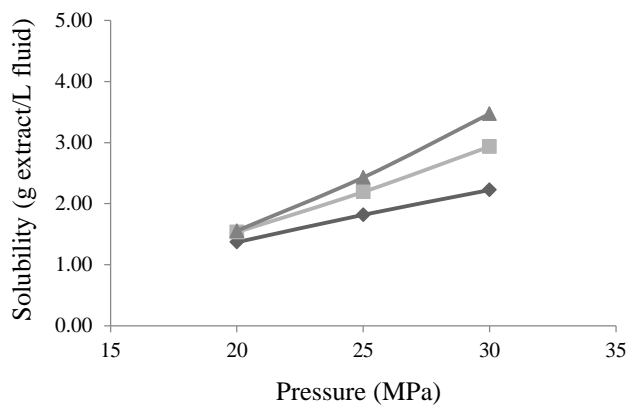
Solubility of extracts in supercritical fluid were obtained from the first part of the extraction curve of extraction yield versus time. Figure 2 shows the solubility of extracts in supercritical fluid at different flow rates. Retrograde solubility was observed at 20 MPa for all flow rates studied. At constant temperature of 50°C, the solubility increases from 1.20 ± 0.30 g extract / L fluid to 2.23 ± 0.17 g extract / L fluid as the pressure increases from 20 to 30 MPa. An average increase of 45% can be observed for all flow rates. However, at 70°C, the increase of 67% may be observed as solubility increases from 1.27 ± 0.30 g extract / L fluid to 3.80 ± 0.50 g extract / L fluid as pressure increases from 20 to 30 MPa.

The increase in temperature increases the solubility of extracts in supercritical fluid. Retrograde solubility or cross over pressure has an inverse dominant effect of pressure to temperature. Below the cross over pressure, the dominant effect is pressure. As the pressure increased, the density of supercritical fluid increases thus, increasing the solvating power of fluid into the sample matrix. The increase in temperature decreases the solvent density, hence lowering the solvating power of fluid. But, above the cross over pressure, the extracts solubility depends on the vapor pressure of the solute. Increasing temperature causes the vapor pressure of solute to increase thus favour the extraction process. This tip off between the major effects of density of solvent to the vapor pressure of solute was referred as retrograde solubility. The cross over pressure is

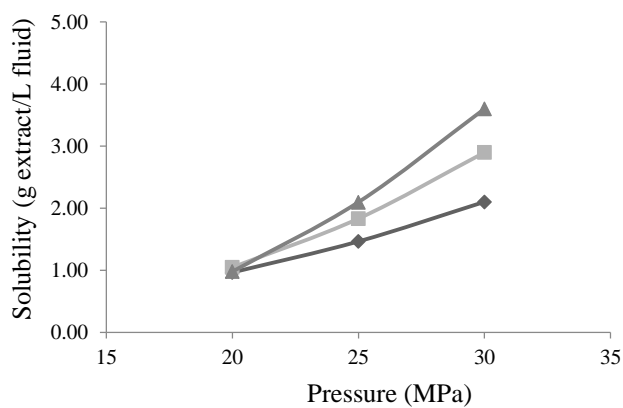
not similar for all samples. As reported by previous researchers, the cross over pressure values of cinnamic acid, phenoxyacetic acid, 4-methoxyphenylacetic acid and clotrimazole are 16.2 to 16.8 MPa, 13.4 to 14 MPa, 16 to 16.7 MPa and 20 MPa, respectively [31, 32].



(a)



(b)



(c)

Figure 2. Solubility of extract in supercritical fluid at constant flow rate of (a) 2 mL/min (b) 3 mL/min (c) 4 mL/min (♦ for 50 °C, ■ for 60 °C and ▲ for 70 °C)

Kinetic Models

The extraction curve was divided into three phases as shown in Figure 3. Phase I is the equilibrium region where the saturated solute present at the outer layer of sample was extracted (0 – 20 min). It can be observed that 70% from the total extractable content is extracted in the first 20 minutes for all cases. While the other 30% extracted within the next 40 minutes. This shows that most of the extracts is located in the outer layer of the sample matrix. Equilibrium controlled region dictates the extraction regime during the first 20 minutes of extraction. Table 2 shows the k_{fa} value which represents the equilibrium-controlled region, is higher compared to the k_{sa} value which represent the diffusion-controlled region. At constant temperature and pressure of 70°C and 30 MPa, the k_{fa} value is 1.20, 2.82 and 3.93 m^3/min for flow rate of 2, 3 and 4 mL/min, respectively. Comparatively to the k_{sa} , the value is 0.00094, 0.00086 and 0.0011 m^3/min as the flow rate increases from 2 to 4 mL/min. This data supports the plotted data by showing the equilibrium-controlled region dominates the extraction regime substantially.

The extracts are mostly located at the outer layer of the surface matrix making it easily accessible for the fluid to extract it. Several reports on other mass transfer coefficient in solid and fluid phase have been reported such as 7.85×10^{-6} m/s and 6.56×10^{-9} m/s for *Pistacia khinjuk* [33], 14.1×10^{-3} s⁻¹ and 1.26×10^{-4} s⁻¹ for sunflower seed [34], 6.298×10^{-3} min⁻¹ and 1.541×10^{-2} min⁻¹ for crambe seed [35], 5.73×10^{-3} s⁻¹ and 2.23×10^{-4} s⁻¹ for hops [36] and 0.333 hr⁻¹ and 13.2×10^{-2} h⁻¹ for *Quercus cerrus* cork [37], respectively. For this study, as the increases of pressure, temperature and flow rate increases, the k_{fa} value also increases. This is because the condition permits more interaction between solute and solvents by altering the density of solvents, increasing the solvation power of solvent, increasing the vapour pressure of solute and removing the film resistance for better mass transfer.

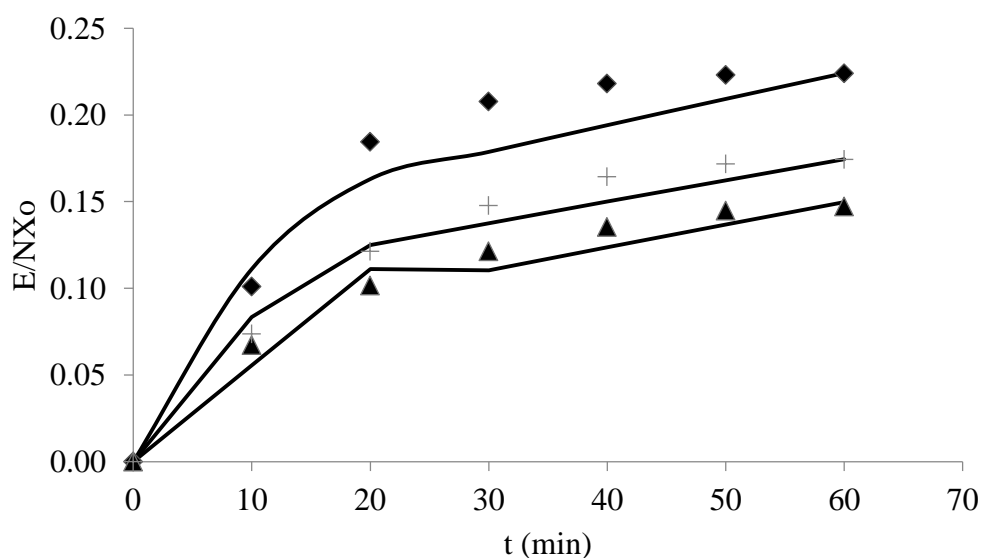


Figure 3. Extraction profile of betel nuts using supercritical fluid at constant temperature of 70°C and 30 MPa (with (◆) 2mL/min (+) 3mL/min (▲) 4mL/min (—) BIC model).

Table 2: Mass transfer coefficient in fluid phase and solid phase.

Flow Rate (mL/min)	Temperature (°C)	Pressure (MPa)	k_{sa} (m ³ /min)	k_{fa} (m ³ /min)	AARD (%)
2	50	20	0.00149	0.3497	4.48
		25	0.00243	0.4773	9.52
		30	0.00063	0.5325	6.37
	60	20	0.00015	0.6796	5.14
		25	0.00071	0.7677	6.42
		30	0.00144	0.8517	18.25
	70	20	0.00191	0.8735	4.79
		25	0.00085	0.9698	5.00
		30	0.00094	1.2016	8.60
3	50	20	0.00207	1.2667	11.87
		25	0.00353	1.9571	6.76
		30	0.00069	2.4555	6.33
	60	20	0.00156	1.6409	13.68
		25	0.00124	2.0053	5.72
		30	0.00128	2.7116	5.77
	70	20	0.00123	1.7452	7.29
		25	0.00104	2.1591	7.74
		30	0.00086	2.8185	6.18
4	50	20	0.00276	1.9989	7.84
		25	0.00081	2.2817	4.23
		30	0.00082	3.2728	2.87
	60	20	0.00141	2.1924	5.92
		25	0.00104	2.6658	3.20
		30	0.00258	3.6185	12.53
	70	20	0.00252	2.3319	7.33
		25	0.00107	2.7996	6.81
		30	0.00111	3.9623	8.82
Overall AARD					7.39

The extraction rate was controlled by the solubility of solute in the supercritical fluid until it reaches G/Z (dimensionless time). As the solute from the outer layer depletes, the intermediate phase begins. Separation of phases is given by a transition separator h_k , which divided the equilibrium-controlled region with the diffusion-controlled region. The extraction phase was from 20 – 30 min as shown in Figure 3. Phase II given by the transition region indicates the flow of solvent through the inner layer of sample matrix until it reaches ψ_k (dimensionless time). The extraction then moves Phase III, i.e., the diffusion-controlled region. Here, the extraction is highly dependent on the diffusivity of solvent in solid matrix to extract solute located in the intact cell. In normal cases, the value of k_{fa} will be greater than k_{sa} because the mass transfer in fluid phase (equilibrium-controlled region) is faster than that in solid phase (diffusion-controlled region). Extraction was rapid in the fluid phase due to abundant amount of solute present in the outer layer of sample. Also, the diffusion path from

solvent to solute was shorter compared to the penetration of solute to the inner layer of sample in solid phase.

Table 2 shows the result of kinetic modelling using simplified Sovová model and BIC model with overall AARD value of 7.39%. The kinetic modelling was conducted to obtain the mass transfer coefficient values in solid and fluid phase. It can be seen in Table 2 that the values of mass transfer coefficient in solid phase are significantly lower than the mass transfer coefficient on fluid phase. This indicates that the extraction regime is dominated by the equilibrium-controlled region instead of the diffusion-controlled region. This shows that the solubility of solute in supercritical fluid is essential in the extraction process instead of the diffusivity of solute in the solid matrix. Thus, the discussions are focused on the fluid phase instead of the solid phase.

At constant temperature of 70°C and pressure if 30 MPa, as flow rate increase from 2 to 4 mL/min, the mass transfer coefficient in fluid phase increases from 1.2016 m³/min to 3.9623 m³/min. The increase of flow rate shows a growth of 69.9%. Increasing the solvent flow rate increases the saturation of solvent in the extraction cell. The increase on volume increases the interaction of solute and solvent hence enhancing the mass transfer from the solid matrices to the fluid bulk. Increasing the solvent flow rate also reduces the film resistance between the wall of sample thus hence increasing the mass transfer rate.

As the increase of operating pressure at constant temperature and solvent flow rate, the mass transfer fluid phase increases. Increment of 20 to 40% can be observe Table 2. At 70°C and 4 mL/min, the mass transfer coefficient in fluid phase increases from 2.3319 m³/min to 3.98623 m³/min as pressure was increase from 20 to 30 MPa. Increasing the operating pressure increases the density of solvent. This causes the solvent to be packed together and increases the solvating power of solvent. The condensed fluid extracts the solute from sample efficiently, thus increasing the mass transfer coefficient as the pressure increased at constant temperature and flow rate. The results obtained was similar with a trend observe in extraction of rosemary using supercritical carbon dioxide [38]. The highest yield was observed at approximately 2.1 wt% of extraction yield operated at 50°C and 400 bar. This condition is the highest conditions compared to only 1.9 wt% yield was obtained when extraction condition reduces to 200 bar. Another recent study also reporting on the same trend whereby the extraction of *Portulaca oleracea* at constant temperature of 333 K increases it yield from 49.75 % to 85.0% when pressure was increased from 8 MPa to 24 MPa [39].

Temperature significantly affects the extraction rate. At constant pressure and flow rate of 30 MPa and 4 mL/min, respectively, the extraction yield and mass transfer increases with increasing temperature from 50°C to 70°C. The increment observed was from 3.2728 m³/min to 3.9623 m³/min. The increases of temperature cause the vapour pressure of solute to increase thus, the extraction became rapid. Increase in temperature also increases the diffusivity of solute into the sample matrix. The same result was observed on the extraction of *Quercus infectoria* galls where, as the extraction temperature increases from 50°C to 70°C at constant pressure of 30 MPa, the k_{fa} value increases from $6.72 \times 10^{-6} \text{ ms}^{-1}$ to $8.20 \times 10^{-6} \text{ ms}^{-1}$ [40]. A similar behaviour also may be observed on the extraction of *Pithecellobium jiringan* jack seed. As the pressure was kept constant at 41.38 MPa, the yield doubles from 3% to 6% when temperature was increased from 40°C to 80°C [41].

CONCLUSION

In a nut shell, the incorporation of simplified Sovová model with BIC model was successful in determining the mass transfer coefficient in solid and fluid phase. Furthermore, the AARD obtained in this study also shows that the data is acceptable to be use within the operating condition studied. The effect of the operating pressure, temperature and solvent flow rate on the mass transfer coefficient on solid and fluid phases was also discussed. The cross over pressure was found to be at 20 MPa. As for the future, it is suggested that the data is taken to be used for up-scaling as the solubility and mass transfer data is valuable for that purposes.

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