EVALUATION OF Justicia gendarussa CRUDE LEAF EXTRACT FOR ENHANCEMENT OF FLAVONOIDS PRODUCTION VIA ADVENTITIOUS ROOT CULTURE AND GENETIC MODIFICATIONS

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Special dedication to

My loving parents Ayob Mat Fatimah Sulieman

My beloved sisters Nurhidayah Ayob Siti Nuradha Ayob

My sweet nieces Wan Nur Hannah Wan Nur Hanis

My future soulmate MJ

For all your love, prayers, support and sacrifice. Thank you so much

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ABSTRACT

Justicia gendarussa extract possesses various bioactivities associated with the availability of flavonoids. Low availability of flavonoids could limit or even hinder the bioactivities effects. Therefore, attempts to enhance the flavonoids production via tissue culture approaches are being studied. This study aimed to optimize flavonoids contents in J. gendarussa using different tissue culture systems (in vitro plant regeneration and adventitious root culture) and genetic transformation methods. The cytotoxicity of plant extracts against various cancer cell lines was also evaluated. Detection and quantification of naringenin and kaempferol were performed using GC-FID. Cytotoxicity tests of crude extract against cancer cell lines (MCF-7, MDA-MB-231, MDA-MB-468, HT-29, HeLa and BxPC-3) were determined by MTT assay. The optimization of elicitors used including yeast extracts (YE), casein hydrolysate (CH) and proline (P) at various concentrations (0, 0.2, 0.4, 0.6, 0.8 or 1.0 mg/L) were examined using nodal explants from in vitro plants. Adventitious roots were inoculated into MS liquid medium supplemented with IBA (2.0-4.0 mg/L). For genetic transformation studies, plasmids pCAMBIA 1305.2, which harbour the PKS gene and plant selectable marker, HPT for hygromycin resistance was used to transform nodal explants of J. gendarussa under the optimized transformation protocol using biolistic and Agrobacterium tumefaciens-mediated transformation. Results showed that mature leaves extract, JG1 had the highest naringenin (444.35 \pm 81.43 mg/kg) and kaempferol (1591.80 \pm 94.91 mg/kg), while the cytotoxicity against BxPC-3 cell was the strongest (IC₅₀~16 μ g/mL). The highest naringenin and kaempferol contents were obtained in leaf crude extracts when treated with 0.6 g/L of CH (1180.30 \pm 50.23 mg/kg) and 0.6 g/L of P (385.01 \pm 13.10 mg/kg), respectively. Adventitious root culture produced high naringenin (97.54 \pm 5.47 mg/kg) and kaempferol (853.82 ± 56.52 mg/kg) when treated with 2.0 mg/L IBA. The optimal parameters for biolistic method were established at 1100 psi helium pressure and 12 cm target distance with 95% of transformation efficiency. Meanwhile, the optimal transformation condition of A. tumefaciens method was bacterial concentration at OD_{600} nm ~ 0.8, 20 minutes of inoculation time, 500 μ M AS and 1 cm explant size with 90% transformation efficiency. Even though A. tumefaciens method produced lower percentage of transient GUS expression than biolistic method, a few transformed explants were successfully produced. The integration of the PKS gene with band size of 1200 bp into the genome of transgenic plants were verified by PCR, sequencing and subsequently confirmed by Southern blot analysis. The content of kaempferol were found to be higher in stem extracts of transgenic plants (450.40 \pm 7.82 mg/kg) than non-transgenic plants (197.13 \pm 2.29 mg/kg). In conclusion, addition of elicitors, establishment of adventitious root culture and A. tumefaciens-mediated transformation could enhance flavonoid contents in J. gendarussa.

ABSTRAK

Ekstrak Justicia gendarussa mempunyai pelbagai bioaktiviti berkaitan dengan ketersediaan flavonoid. Ketersediaan kurang flavonoid boleh menghadkan atau menghalang kesan bioaktiviti. Oleh itu, percubaan untuk meningkatkan kandungan flavonoid melalui teknik kultur tisu sedang dikaji. Kajian ini bertujuan untuk mengotimumkan kandungan flavonoid di dalam J. gendarussa menggunakan sistem kultur tisu yang berbeza (pertumbuhan pokok in vitro dan kultur akar adventitus) dan kaedah transformasi genetik. Kesitotoksikan ekstrak pokok terhadap pelbagai titisan sel kanser juga dinilai. Pengesanan dan pengkuantitian naringenin dan kempferol telah dijalankan menggunakan GC-FID. Ujian kesitotoksikan ekstrak daun mentah J. gendarussa terhadap titisan sel kanser (MCF-7, MDA-MB-231, MDA-MB-468, HT-29, HeLa dan BxPC-3) telah ditentukan oleh asai MTT. Pengoptimuman elisitor iaitu ekstrak yis (YE), kasein hidrolisat (CH) dan prolin (P) pada pelbagai kepekatan (0, 0.2, 0.4, 0.6, 0.8 or 1.0 mg/L) telah diperiksa menggunakan eksplan nodal dari pokok in vitro. Akar adventitus telah diinokulasi di dalam cecair media MS yang ditambah dengan IBA (2.0-4.0 mg/L). Bagi kajian transformasi genetik, plasmid pCAMBIA 1305.2 yang mempunyai gen PKS and gen penanda pemilihan pokok, HPT untuk rintangan higromisin telah digunakan untuk transformasi eksplan nodal J. gendarussa di bawah kaedah transformasi menggunakan biolistik dan transformasi berperantarakan Agrobacterium tumefaciens. Hasil kajian daun matang ekstrak, JG1 menunjukkan naringenin tertinggi (444.35 \pm 81.43 mg/kg) dan kempferol (1591.80 \pm 94.91 mg/kg), manakala kesitotoksikan menentang sel BxPC-3 yang terkuat (IC₅₀~16 µg/mL). Kandungan tertinggi naringenin dan kempferol diperolehi di dalam ekstrak daun apabila dirawat dengan CH 0.6 g/L (1180.30 ± 50.23 mg/kg) dan P pada 0.6 g/L (385.01 ± 13.10 mg/kg). Kultur akar adventitus menghasilkan tinggi naringenin ($97.54 \pm 5.47 \text{ mg/kg}$) dan kempferol (853.82 ± 56.52 mg/kg) apabila dirawat dengan 2 mg/L IBA. Parameter optimum bagi kaedah biolistik adalah tekanan helium 1100 psi dan 12 cm jarak sasaran dengan keberkesanan tranformasi sebanyak 95%. Manakala, parameter optimum bagi kaedah A. tumefaciens adalah apabila dirawat dengan kepekatan bakteria pada OD₆₀₀nm~0.8, 20 minit masa inokulasi, ditambah dengan 500 µM kepekatan AS dan saiz eksplan iaitu 1 cm dengan keberkesanan transformasi sebanyak 90%. Walaupun kaedah A. tumefaciens menghasilkan peratusan gen GUS transien lebih rendah berbanding kaedah biolistik, beberapa eksplan tertransformasi telah berjaya dihasilkan. Kehadiran dan integrasi gen PKS dapat dikenalpasti dengan pengesanan saiz jalur 1200 bp di dalam genom pokok transgenik berdasarkan PCR, penjujukan dan seterusnya disahkan oleh analisis pemblotan Southern. Kandungan kempferol didapati lebih tinggi di dalam ekstrak batang pokok transgenik (450.40 \pm 7.82 mg/kg) berbanding pokok tanpa tertransformasi (197.13 \pm 2.29 mg/kg). Kesimpulannya, penambahan elisitor, penghasilan kultur akar adventitus dan transformasi berperantarakan A. tumefaciens boleh meningkatkan kandungan flavonoid di dalam J. gendarussa.

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LIST OF SYMBOLS

rpm -	-	Rotation per minute
v -	-	Volume
v/v	-	Volume per volume
w/v	-	Weight per volume
µg/mL		Microgram per milliliter

LIST OF ABBREVIATIONS

JG	-	Justicia gendarussa
MTT	-	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium
		bromide
PKS	-	Polyketide synthase
CHS	-	Chalcone synthase
GUS	-	β-glucuronidase
DNA	-	Deoxyribonucleic acid
RNA	-	Ribonucleic acid
PCR	-	Polymerase Chain Reaction
PBS	-	Phosphate buffer saline
FBS	-	Fetal bovine serum
DMSO	-	Dimethyl sulfoxide
LB	-	Luria Bertani
IBA	-	Indole-3-butyric acid
vir	-	virulence
GC-FID	-	Gas Chromatography-Flame Ionization Detector
MS	-	Murashige and Skoog
X-gluc	-	5-bromo-4-chloro-3-indolyl β -D-glucuronic acid
HCl	-	Hydrochloric acid
Na ₂ CO ₃	-	Sodium carbonate
NANO ₂	-	Sodium nitrile
AlCl ₃	-	Aluminium chloride
NaOH	-	Sodium hydroxide
CTAB	-	Cetyltrimethyl ammonium bromide
PVP	-	Polyvinyl pyrrolidone

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CHAPTER 1

INTRODUCTION

1.1 Background of the problem

The updated strategy of World Health Organisation (WHO) from 2014 to 2023 devotes more attention than its predecessor to prioritizing health services and systems, including traditional and complementary medicine products, practices and practitioners (WHO, 2013). Medicinal plants have been used as traditional treatments for numerous human diseases for thousand years. In rural areas of the developing countries, they continue to be used as the primary source of medicine since western pharmaceuticals are often expensive or inaccessible (Ekor, 2014). The natural products derived from medicinal plants have proven to be an abundant source of biologically active compounds, many of which have been the basis for the development of new leading chemicals for pharmaceuticals (Enzo, 2011).

Numerous researchers have shown high interest in plant secondary metabolites particularly flavonoids which have possessed diverse bioactivities and contributed valuable prospects to the pharmaceutical industries. Flavonoids exhibit various biological effects including lowering plasma levels of low-density lipoproteins, inhibiting platelet aggregation, promoting scavenging free radicals and reducing cell proliferation (Woodman and Chan, 2004). Therefore, in this study, flavonoids such as naringenin (flavanone) and kaempferol (flavonol) were given greatattention due to their potential beneficial effects on the human health. Naringenin was reported as strong antioxidant and showed cytotoxicity against human breast cancer cell line, MCF-7 (Cavia-Saiz *et al.*, 2010; Park *et al.*, 2010).

Previous studies reported kaempferol as strong antioxidant, able to prevent arteriosclerosis, inhibits cell proliferation and induces apoptosis in pancreatic cancer cells (Tu *et al.*, 2007; Zhang *et al.*, 2008).

Justicia gendarussa, which is also known by its common name Gendarussa has been investigated as potential medicinal plants in this study. These plants have distributed in many countries such as India, Indonesia, Malaysia and Sri Lanka. The root and leaf extracts of *J. gendarussa* have been used traditionally to treat many ailments such as chronic rheumatism, inflammations, bronchitis, headache, arthritis, vaginal discharges, dyspepsia, eye disease and fever (Janarthanam and Sumanthi, 2010). Leaf and stem extracts of *J. gendarussa* were reported to possess anti-inflammatory, antioxidant, antibacterial, antifungal, antiangiogenis, antiplatelet, antiarthritic, anthelmintic and hepatoprotective activities (Navarro *et al.*, 2001; Paval *et al.*, 2009; Krishna *et al.*, 2010; Saha *et al.*, 2012). Phytochemical studies on leaf extracts of *J. gendarussa* revealed the presence of flavonoids, alkaloids, triterpenoid saponins, amino acids, aromatic amines and sterols (Chakravarty *et al.*, 1982; Ratnasooriya *et al.*, 2007; Bambang Prajogo *et al.*, 2009; Mustafa *et al.*, 2010; Uddin *et al.*, 2011; Kiren *et al.*, 2014).

According to statistics in Malaysia, the incidence of breast and cervical cancers are common among female patients while colon and pancreas cancers are prevalence among men patients (Bachok *et al.*, 2012; Farooqui *et al.*, 2013). Unfortunately, cancer is a public health problem in all over the world affecting all categories of persons (Iweala *et al.*, 2015). Despite the advancement in cancer therapies such as surgery, radiotherapy, hyperthermia, hormone therapy and chemotherapy, these therapies are ineffective in destroying cancer cells and may cause damage to the healthy cells. Examples of the adverse side effects of cancer treatments include mouth sore, tiredness, hair loss, nausea and vomiting (Jones *et al.*, 2004). For many years herbal medicines have been used and are still used in developing countries as the primary source of medical treatment. Thus, research has developed into investigating the potential properties and uses of terrestrial plants extracts for the preparation of potential nanomaterial based drugs for diseases including cancer (Zakaria *et al.*, 2011b). Many plant species are already being used

to treat or prevent development of cancer. Multiple researchers have identified species of plants that have demonstrated anticancer properties with a lot of focus on those that have been used in herbal medicine in developing countries (Greenwell and Rahman, 2015). In this regard, this is the first study to evaluate the cytotoxic activities in *Justicia gendarussa* crude leaf extracts against breast (MCF-7, MDA-MB-231 and MDA-MB-468), colon (HT-29), cervix (HeLa) and pancreas (BxPC-3) cancer cell lines have been investigated. In order to identify the plants with potential bioactive compounds against cancer cell lines, MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay will be performed by using the crude leaf extracts and flavonoids (naringenin and kaempferol) which would give a plausible contribution in cytotoxicity of secondary metabolite towards cancer treatment.

It is well-known that flavonoids possessed remarkable strong anticancer and antioxidant activities (Susanti *et al.*, 2007). The drawbacks of the flavonoids extraction from field-grown plants are low yields and fluctuation in flavonoids concentration due to geographical, seasonal and environmental variations. In addition, the long cultivation period resulting in high-cost commercial drug production (Murthy *et al.*, 2014). Current advances in plant biotechnology offer manipulation of bioactive compounds via tissue culture approaches (Baque *et al.*, 2012). Many strategies for enhancement of plant metabolites such as selection of cell clones, optimization of medium and culture environments, elicitation, nutrient and precursor feeding and biotransformation can be applied. Hence, in this study, alternative methods to enhance the production of flavonoids from *J. gendarussa* were performed through elicitation on *in vitro* plant, adventitious root cultures and genetic transformation methods.

In recent years, many studies on the production of biomass and secondary metabolites through elicitation on *in vitro* plant and adventitious root culture system have been conducted. Elicitors are compounds from various biotic or abiotic sources which may enhance the secondary metabolite in plant cells by triggering the signal in secondary metabolites production (Rao and Ravishankar, 2002). Elicitation can be used as one of the important strategies to increase secondary metabolites production and reduce production cost (Siddiqui *et al.*, 2013). In addition, plant growth

regulators are also one of the most important factors in affecting cell growth, differentiation and metabolites formation (Baque *et al.*, 2012). Previous studies reported that adventitious root tissues are efficient in biomass production because of fast growth rates and stable secondary metabolite productivity (Choi *et al.*, 2000; Kim *et al.*, 2004a; Wu *et al.*, 2006). Hence, in this study, high flavonoids production and biomass can be achieved by optimizing the effects of elicitors and plant growth regulators of *in vitro* plant and adventitious root cultures conditions.

Modification or enhancement of flavonoids production in *Justicia gendarussa* plant through genetic transformation would be a powerful tool in flavonoid biosynthetic pathway. It is important to develop a genetic transformation system by optimizing the transformation efficiency parameters followed by introduction of a polyketide synthase (PKS) gene i.e chalcone synthase (CHS), which is the precursor in the flavonoid biosynthesis pathway (Jamalnasir *et al.*, 2014). There are several methods available for genetic plant transformation such as *Agrobacterium tumefaciens*-mediated transformation, biolistic and electroporation (Yong *et al.*, 2006). In this study, the transformation methods using biolistic and *A. tumefaciens* - mediated transformation had been optimized using β -glucuronidase (GUS) as a reporter gene. Enhancement of the flavonoids production in *J. gendarussa* plant is a highly valuable protocol and thereby this is an ideal platform to improve the genetic transformation system in the medicinal plant.

1.2 Problem Statements

Flavonoids such as kaempferol and naringenin have been reported to have antioxidant, anticancer, antibacterial, antiviral and anti-inflammatory properties (Kumar and Pandey, 2013). These flavonoids also act as plant protective agents against various biotic and abiotic stress and also beneficial in preventing degenerative diseases on human (Kasote *et al.*, 2015). To the best of our knowledge, there is no report on the detection and quantification of naringenin and kaempferol in

young and mature leaves of *Justicia gendarussa* especially from different localities in Johor.

Until now, no study has yet been carried out to determine the cytotoxicity effects of *Justicia gendarussa* leaf extracts against human cancer cell lines. This study was intended to demonstrate the anticancer potential of local *J. gendarussa* as an alternative anticancer agent.

Nowadays, medicinal plant studies have gained considerable attention internationally especially from pharmacology industry because of the extensive research on phytochemical and biological activities. However, the quality of the bioactive compounds derived from field-grown medicinal plants may be affected by environmental factors, physiological and developmental stages of the plant. Field cultivation requires a long growth period and plant management, which is a slow, time-consuming and laborious process (Wang et al., 2015). Therefore, in this study, plant cell culture approach is an alternative for the enhancement of the biomass and secondary metabolites production particularly flavonoids. Studies on the effect of elicitor on *in vitro* plant cultures, plant growth regulators of adventitious root cultures and genetic transformation for the enhancement of flavonoids production were not yet explored. In addition, manipulating the medium compositions supplemented with different concentrations of elicitors and plant growth regulators for in vitro plant and adventitious root cultures with the introduction of polyketide synthase (PKS) gene into J. gendarussa via genetic transformation were attempted in order to enhance the flavonoids production.

1.3 Aim and Objectives of the Study

The aim of the study is to increase the flavonoids (i.e naringenin and kaempferol) contents of *Justicia gendarussa* using tissue culture approaches via adventitious roots cultures and genetic modifications. The study objectives include:

1. To determine flavonoids contents in young and mature leaves of *J*. *gendarussa* plant.

2. To evaluate the cytotoxicity of *J. gendarussa* crude leaf extract against various cancer cell lines.

3. To assess the effects of different concentrations of elicitor and plant growth regulator on flavonoids contents of *in vitro* plant and adventitious roots cultures of *J. gendarussa*.

4. To determine the effects of transformation parameters on the biolistic transformation efficiency of *J. gendarussa* nodal explants

5. To determine the effects of transformation parameters on the *A. tumefaciens*– mediated transformation efficiency of *J. gendarussa* nodal explants

1.4 Scope of the Study

This research was focused on quantification of flavonoids, namely naringenin and kaempferol in young and mature leaf extracts of Justicia gendarussa from different locations in Johor by GC-FID method. The mature leaves that produced high flavonoids content were subjected to cytotoxic MTT assay against various cancer cell lines, MCF-7, MDA-MB-231, MDA-MB-468, BxPC-3, HeLa and HT-29. Plants that produced high flavonoids content and strong cytotoxic activity were selected for further applications of tissue culture approach. In order to enhance the flavonoids content in J. gendarussa plants, in vitro plant culture, adventitious root culture and genetic plant transformation were applied. Firstly, in vitro plants were supplemented with different concentrations of elicitors, while adventitious root cultures were induced and optimized using different concentrations of plant growth regulator in shake flask system. Determination of flavonoids content on in vitro plant and adventitious roots cultures were done using the GC-FID method. Secondly, optimization of biolistic and A. tumefaciens-mediated transformation parameters for J. gendarussa plant by GUS histochemical assay were conducted. The presence of HPT, GUS and PKS genes in transgenic plants were verified by PCR and confirmed using Southern blot analysis. Lastly, the comparison of flavonoids content in transgenic plants and wild-type plants were determined using the GC-FID method.

1.5 Significance of the Study

This study was conducted to determine flavonoids content in Justicia gendarussa plant which contributes to cytotoxic effect against cancer cell lines. High cytotoxic effect of J. gendarussa extracts could be served as a good candidate for the development of new anticancer agents. Besides that, the application of in vitro plant culture, adventitious roots culture and genetic plant transformation could be applied to enhance flavonoids content in J. gendarussa. The establishment of adventitious roots culture system and flavonoids biomass production could enhance the flavonoids content from adventitious roots culture by applying in various strategies such as elicitation, application in a suitable bioreactor and bioprocess technologies. In this study, a fast and reliable method of biolistic and Agrobacterium J. *tumefaciens*-mediated transformation of gendarussa were developed. Establishment of plant transformation system provides the first essential step in the systematic study of the flavonoid biosynthetic pathway. The introduction of desired gene i.e PKS gene into the plant genome would modify flavonoids content in putatively transformed plants. Furthermore, a suitable approach for secondary metabolite production i.e flavonoids in J. gendarussa plants suggested through adventitious root culture and genetic modifications.

REFERENCES

- Abaza, M. S. I., Orabi, K. Y., Al-Quattan, E., and Al-Attiyah, R. J. (2015). Growth Inhibitory and Chemo-sensitization Effects of Naringenin, A Natural Flavanone Purified from *Thymus vulgaris*, on Human Breast and Colorectal Cancer. *Cancer Cell International*. 15(46), 1–19.
- Achakzai, A. K. K., Achakzai, P., Masood, A., Kayani, S. A., and Tareen, R. B. (2009). Response Of Plant Parts And Age On The Distribution Of Secondary Metabolites On Plants Found In Quetta. *Pakistan Journal of Botany*. 41(5), 2129–2135.
- Ahamad, M. S., Siddiqui, S., Jafri, A., Ahmad, S., Afzal, M., and Arshad, M. (2014).
 Induction of Apoptosis and Antiproliferative Activity of Naringenin in Human
 Epidermoid Carcinoma Cell through ROS Generation and Cell Cycle Arrest.
 PLoS ONE. 9(10), 1–8.
- Ahmad, R., Ali, A. M., Israf, D. A., Ismail, N. H., Shaari, K., and Lajis, N. H. (2005). Antioxidant, Radical-Scavenging, Anti-Inflammatory, Cytotoxic and Antibacterial Activities of Methanolic Extracts of Some *Hedyotis* Species. *Life Sciences*. 76(17), 1953–1964.
- Aiswarya, G., Mallika, V., Mur, L. A. J., and Soniya, E. V. (2016). Ectopic Expression and Functional Characterization of Type III Polyketide Synthase Mutants from *Emblica officinalis* Gaertn. *Plant Cell Reports*. 35(10), 2077– 2090.
- Ajungla, L., Patil, P. P., Barmukh, R. B., and Nikam, T. D. (2009). Influence of Biotic and Abiotic Elicitors on Accumulation of Hyoscyamine and Scopolamine in Root Cultures of *Datura metel L. Indian Journal of Biotechnology*. 8(3), 317–322.
- Al-Barazanjy, R. K., Dizaye, K., and Al-Asadye, A. A. (2013). Cytotoxic and Cytogenetic Effects of Salvia officinalis on Different Tumor Cell Lines. *Middle East Journal of International Medicine*. 6(4), 15–25.

- Aldemita, R. R., and Hodges, T. K. (1996). *Agrobacterium tumefaciens* Mediated Transformation of Japonica and Indica Rice Varieties. *Planta*. 199(4), 612–617.
- Ali, M. B., Thanh, N. T., Yu, K., Hahn, E.-J., Paek, K.-Y., and Lee, H. L. (2005). Induction in the Antioxidative Systems and Lipid Peroxidation in Suspension Culture Roots of *Panax ginseng* Induced by Oxygen in Bioreactors. *Plant Science*. 169(5), 833–841.
- Alkhamaiseh, S. I., Taher, M., Ahmad, F., Susanti, D., and Ichwan, S. J. A. (2011). Antioxidant and Cytotoxic Activities of *Calophyllum rubiginosum*. *International Journal of Phytomedicine*. 3(2), 157–163.
- Alkhamaiseh, S. I., Taher, M., Farediah, A., Qaralleh, H., Althunibat, O. Y., Susanti, D., and Ichwan, S. J. A. (2012). The Phytochemical Content and Antimicrobial Activities of Malaysian *Calophyllum canum* (stem bark). *Pakistan Journal of Pharmaceutical Sciences*. 25(3), 555–563.
- Alsemari, A., Alkhodairy, F., Aldakan, A., Al-mohanna, M., Bahoush, E., and Shinwari, Z. (2014). The Selective Cytotoxic Anti-Cancer Properties and Proteomic Analysis of Trigonella Foenum-Graecum. *BMC Complementary and Aternative Medicine*. 14(1), 114–122.
- Arul, D., and Subramanian, P. (2013). Naringenin (Citrus Flavonone) Induces Growth Inhibition, Cell Cycle Arrest and Apoptosis in Human Hepatocellular Carcinoma Cells. *Pathology and Oncology Research*. 19(4), 763–770.
- Asai, K., Moriwaki, S., and Maeda-yamamoto, M. (2005). Kaempferol, a Tea Flavonol, Effect on Interleukin-2 Signal Transduction of Human T Cell Leukemia. *Japan Agricultural Research Quarterly*. 39(3), 175–179.
- Awad, V., Kuvalekar, A., and Harsulkar, A. (2014). Microbial Elicitation in Root Cultures of *Taverniera cuneifolia* (Roth) Arn. for Elevated Glycyrrhizic Acid Production. *Industrial Crops and Products*. 54, 13–16.
- Azad, A. K., Rabbani, G., Amin, L., and Sidik, N. M. (2013). Development of Transgenic Papaya through *Agrobacterium* - Mediated Transformation. *International Journal of Genomics*. 2013, 1–5.
- Azeez, H. A., and Ibrahim, K. M. (2013). Effect of Biotic Elicitors on Secondary Metabolite Production in Cell Suspensions of *Hypericum triquetrifolium* Turra. *Bulletin UASVM Horticulture*. 70(1), 26–33.

- Bachheti, R. K., Pandey, D. P., Joshi, A., and Rana, V. (2011). Chemical Analysis of Aerial Parts of *Justicia gendarussa* Chemical. *International Journal of ChemTech Research*. 3(1), 244–247.
- Bachok, N., Mohd Amin, R., Krishna, G. R., and Aishah, K. (2012). Understanding Barriers to Malaysian Women with Breast Cancer Seeking Help. Asian Pacific Journal of Cancer Prevention. 13(8), 3723–3730.
- Baldi, A., Srivastava, A. K., and Bisaria, V. S. (2009). Fungal Elicitors for Enhanced Production of Secondary Metabolites in Plant Cell Suspension Cultures. In *Symbiotic Fungi* (pp. 373–380). Germany: Springer Berlin Heidelberg.
- Bambang Prajogo, E. W., David, G., Ferreira Queiroz, E., John-Luc, W., Noor Cholies, Z., Aucky, H., and Hostettmann, K. (2009). Isolation of Male Antifertility Compound in N-Butanol Fraction of *Justicia gendarussa* Burm. F. Leaves. *Folia Medica Indonesiana*. 45(1), 28–31.
- Bandyopadhyay, S., Romero, J. R., and Chattopadhyay, N. (2008). Kaempferol and Quercetin Stimulate Granulocyte -Macrophage Colony - Stimulating Factor Secretion in Human Prostate Cancer Cells. *Molecular and Cellular Endocrinology*. 287(1), 57–64.
- Baque, M. A., Hahn, E. J., and Paek, K. Y. (2010a). Growth, Secondary Metabolite Production and Antioxidant Enzyme Response of *Morinda citrifolia* Adventitious Root as Affected by Auxin and Cytokinin. *Plant Biotechnology Reports*. 4(2), 109–116.
- Baque, M. A., Hahn, E., and Paek, K. Y. (2010b). Induction Mechanism of Adventitious Root from Leaf Explants of *Morinda citrifolia* as Affected by Auxin and Light Quality. *In Vitro Cellular and Development Biology-Plant*. 46(1), 71–80.
- Baque, M. A., Lee, E.-J., and Paek, K.-Y. (2010c). Medium Salt Strength Induced Changes in Growth , Physiology and Secondary Metabolite Content in Adventitious Roots of *Morinda citrifolia*: The Role of Antioxidant Enzymes and Phenylalanine Ammonia Lyase. *Plant Cell Reports*. 29(7), 685–694.
- Baque, M. A., Moh, S.-H., Lee, E.-J., Zhong, J.-J., and Paek, K.-Y. (2012). Production of Biomass and Useful Compounds from Adventitious Roots of High-Value Added Medicinal Plants using Bioreactor. *Biotechnology Advances*. 30(6), 1255–1267.

- Batista, D., Fonseca, S., Serrazina, S., Figueiredo, A., and Salome, M. S. (2008). Efficient and Stable Transformation of Hop (*Humulus lupulus* L.) var. Eroica by Particle Bombardment. *Plant Cell Reports*. 27(7), 1185–1196.
- Bestwick, C. S., Milne, L., Pirie, L., and Duthie, S. J. (2005). The Effect of Short-Term Kaempferol Exposure on Reactive Oxygen Levels and Integrity of Human (HL-60) Leukaemic Cells. *Biochimica et Biophysica Acta*. 1740(3), 340–349.
- Bhagya, N., Chandrashekar, K. R., Karun, A., and Bhavyashree, U. (2012). Plantlet Regeneration through Indirect Shoot Organogenesis and Somatic Embryogenesis in *Justicia gendarussa* Burm. f., A Medicinal Plant. *Journal Plant Biochemistry and Biotechnology*. 22(4), 474–482.
- Bhambhani, S., Karwasara, V. S., Dixit, V. K., and Banerjee, S. (2012). Enhanced Production of Vasicine in Adhatoda vasica (L.) Nees. Cell Culture by Elicitation. Acta Physiologiae Plantarum. 34(4), 1571–1578.
- Bhanot, A., Sharma, R., and Noolvi, M. N. (2011). Natural Sources as Potential Anti-Cancer Agents: A Review. *International Journal of Phytomedicine*. 3(1), 9–26.
- Bhat, S. R., and Srinivasan, S. (2002). Molecular and Genetic Analyses of Transgenic Plants : Considerations and Approaches. *Plant Science*. 163(4), 673– 681.
- Brown, T. A. (2001). Southern Blotting and Related DNA Detection Techniques. In Encyclopedia of Life Sciences (pp. 1–6). London: eLS.
- Cailleau, R., Young, R., Olive, M., and Reeves, W. J. (1974). Breast Tumor Cell Lines from Pleural Effusions. *Journal of The National Cancer*. 53(3), 661–674.
- Cailleau, R., Olive, M., and Cruciger, Q. V. J. (1978). Long-Term Human Breast Carcinoma Cell Lines of Metastatic Origin: Preliminary Characterization. *In Vitro*. 14(11), 3–7.
- Campbell, J. K., King, J. L., Harmston, M., Lila, M. A., and Erdman, J. W. (2006). Synergistic Effects of Flavonoids on Cell Proliferation in Hepa-1c1c7 LNCap Cancer Cell Lines. *Journal of Food Science*. 71(4), S358–S363.
- Canini, A., Alesiani, D., Arcangelo, G. D., and Tagliatesta, P. (2007). Gas Chromatography – Mass Spectrometry Analysis of Phenolic Compounds from *Carica papaya* L. Leaf. *Journal of Food Composition and Analysis*. 20(7), 584– 590.

- Cao, J., Han, J., Xiao, H., Qiao, J., and Han, M. (2016). Effect of Tea Polyphenol Compounds on Anticancer Drugs in Terms of Anti-Tumor Activity, Toxicology, and Pharmacokinetics. *Nutrients*. 8(762), 1–10.
- Carlo, G. Di, Mascolo, N., Angelo, A., and Capasso, F. (1999). Minireview Flavonoids : Old and New Aspects of a Class of Natural Therapeutic Drugs. *Life Sciences*. 65(4), 337–353.
- Cavia-Saiz, M., Busto, M. D., Pilar-Izquierdo, M. C., Ortega, N., Perez-Mateos, M., and Muñiz, P. (2010). Antioxidant Properties, Radical Scavenging Activity and Biomolecule Protection Capacity of Flavonoid Naringenin and its Glycoside Naringin: A Comparative Study. *Journal of The Science of Food and Agriculture*. 90(7), 1238–1244.
- Chaichana, N., Dheeranupattana, S., Jatisatienr, A., and Wangkarn, S. (2012). Response of Stemona Alkaloid Production in *Stemona* sp. to Chitosan and Yeast Extract Elicitors. *Current Research Journal of Biological Sciences*. 4(4), 449– 454.
- Chakravarty, A. K., Dastidar, P. P. G., and Pakrashi, S. C. (1982). Simple Aromatic Amines From *Justicia gendarussa*. 13C NMR Spectra of the Bases and Their Analogues. *Tetrahedron*. 38(12), 1797–1802.
- Chen, H., and F, C. (2000). Effect of Yeast Elicitor on the Secondary Metabolism of Ti-Transformed Salvia miltiorrhiza Cell Suspension Cultures. *Plant Cell Reports*. 19(7), 710–717.
- Chen, X., Yang, X., Liu, T., Guan, M., Feng, X., Dong, W., Chu, X., Liu, J., Ci, X., Li, H., Wei, J., Deng, Y., Deng, X., Chi, G., and Sun, Z. (2012). Kaempferol Regulates MAPKs and NF- κ B Signaling Pathways to Attenuate LPS-Induced Acute Lung Injury in Mice. *International Immunopharmacology*. 14(2), 209– 216.
- Chen, H.-J., Lin, C.-M., Lee, C.-Y., Shih, N.-C., Peng, S.-F., Tsuzuki, M., Amagaya, S., Huang, W.-W., and Yang, J.-S. (2013). Kaempferol Suppresses Cell Metastasis Via Inhibition of the ERK-p38-JNK and AP-1 Signaling Pathways in U-2 OS Human Osteosarcoma Cells. *Oncology Reports*. 30(20), 925–932.
- Chen, L., Wang, Q., Chen, H., Sun, G., Liu, H., and Wang, H. (2016). Agrobacterium tumefaciens -Mediated Transformation of Botryosphaeria dothidea. World Journal of Microbiology and Biotechnology. 32(7), 106–111.

- Chinna, M., Devi, P. R., Saravanakumar, A., Femi, V., Hemananthan, E., and Rani, S. S. (2012). Synthesis and Characterization of Gold Nanoparticles by *Justicia* gendarussa Burm F. Leaf Extract. *International Journal of Pharmaceutical Sciences and Research*. 3(2), 623–629.
- Chodisetti, B., Rao, K., Gandi, S., and Giri, A. (2014). Gymnemic Acid Enhancement in the Suspension Cultures of *Gymnema sylvestre* by using the Signaling Molecules-Methyl Jasmonate and Salicylic Acid. In Vitro Cellular and Development Biology-Plant. 51(1), 88–92.
- Choi, S. M., Son, S. H., Yun, S. R., Kwon, O. W., Seon, J. H., and Paek, K. Y. (2000). Pilot-Scale Culture of Adventitious Roots of Ginseng in a Bioreactor System. *Plant Cell*, *T*. 62(3), 187–193.
- Chong, H. Z., Rahmat, A., Yeap, S. K., Akim, A., Alitheen, N. B., Othman, F., and Gwendoline-ee, C. L. (2012). *In Vitro* Cytotoxicity of *Strobilanthes crispus* Ethanol Extract on Hormone Dependent Human Breast Adenocarcinoma MCF-7 Cell. *BMC Complementary and Aternative Medicine*. 12(1), 35–45.
- Cortes, J. R., Perez-g, M., Rivas, M. D., and Zamorano, J. (2007). Kaempferol Inhibits IL-4-Induced STAT6 Activation by Specifically Targeting JAK3 1. *The Journal of Immonology*. 179(6), 3881–3887.
- Coste, A., Vlase, L., Halmagyi, A., Deliu, C., and Coldea, G. (2011). Effects of Plant Growth Regulators and Elicitors on Production of Secondary Metabolites in Shoot Cultures of *Hypericum hirsutum* and *Hypericum maculatum*. *Plant Cell*, *Tissue and Organ Culture*. 106(2), 279–288.
- Cragg, G. M., and Newman, D. J. (2005). Plants as a Source of Anti-Cancer Agents. *Journal of Ethnopharmacology*. 100(1), 72–79.
- Cragg, G. M., Kingston, D. G., and Newman, D. J. (2011). *Anticancer Agents from Natural Products*. United States: CRC press.
- Cui, X.-H., Chakrabarty, D., Lee, E.-J., and Paek, K.-Y. (2010a). Production of Adventitious Roots and Secondary Metabolites by *Hypericum perforatum* L. in a Bioreactor. *Bioresource Technology*. 101(12), 4708–4716.
- Cui, X.-H., Murthy, H. N., Wu, C.-H., and Paek, K.-Y. (2010b). Adventitious Root Suspension Cultures of *Hypericum perforatum*: Effect of Nitrogen Source on Production of Biomass and Secondary Metabolites. *In Vitro Cellular and Development Biology-Plant*. 46(5), 437–444.

- Culea, M., and Gocan, S. (2009). Flavonoids Determination in Herbs by GC and GC/MS. *Journal of Environmental Protection and Ecology*. 10(2), 461–467.
- Dao, T. T. H., Linthorst, H. J. M., and Verpoorte, R. (2011). Chalcone Synthase and Its Functions in Plant Resistance. *Phytochemistry Reviews*. 10(3), 397–412.
- Davies, K. M. (2007). Genetic Modification of Plant Metabolism for Human Health Benefits. *Mutation Research*. 622(2), 122–137.
- De La Riva, G. A., González-Cabrera, J., Vázquez-Padrón, R., and Ayra-Pardo, C. (1998). Agrobacterium tumefaciens: A Natural Tool for Plant Transformation. Electronic Journal of Biotechnology. 1(3), 25–48.
- Denizot, F., and Lang, R. (1986). Rapid Colorimetric Assay for Cell Growth and Survival Modifications to the Tetrazolium Dye Procedure Giving Improved Sensitivity and Reliability. *Journal of Immunilogical Methods*. 89(2), 271–277.
- Diantini, A., Subarnas, A., Lestari, K., Halimah, E., Supriyatna, Y. S., Julaeha, E., Achmad, T. H., Suradji, E. W., Yamazaki, C., Kobayashi, K., Koyama, H., and Abdulah, R. (2012). Kaempferol-3-O-rhamnoside Isolated from the Leaves of *Schima wallichii* Korth. Inhibits MCF-7 Breast Cancer Cell Proliferation through Activation of the Caspase Cascade Pathway. *Oncology Letters*. 3(5), 1069–1072.
- DiCosmo, F., and Misawa, M. (1995). Plant Cell and Tissue Culture: Alternatives for Metabolite Production. *Biotechnology Advances*. 13(3), 425–453.
- Doyle, J. J., and Doyle, J. L. (1990). Isolation of Plant DNA from Fresh Tissue. Focus. 12(1), 13–15.
- Ebrahimi, M. A., and Zarinpanjeh, N. (2015). Bio-Elicitation of β-Carboline Alkaloids in Cell Suspension Culture of *Peganum harmala* L. *Journal of Medicinal Plants*. 14(55), 43–57.
- Ekor, M. (2014). The Growing Use of Herbal Medicines: Issues Relating to Adverse Reactions and Challenges in Monitoring Safety. *Frontiers in Pharmacology*. 4(177), 1–10.
- Elkady, A. I., Hussein, R. A. E. H., and Abu-zinadah, O. A. (2014). Effects of Crude Extracts from Medicinal Herbs *Rhazya stricta* and *Zingiber officinale* on Growth and Proliferation of Human Brain Cancer Cell Line *In Vitro. BioMed Research International*. 2014, 1–16.

- El-Nabarawy, M. A., El-kafafi, S. H., Hamza, M. A., and Omar, M. A. (2015). The Effect of Some Factors on Stimulating the Growth and Production of Active Substances in *Zingiber officinale* Callus Cultures. *Annals of Agricultural Science*. 60(1), 1–9.
- Enzo, A. P. (2011). Traditional Medicinal Plant Extracts and Natural Products with Activity against Oral Bacteria: Potential Application in the Prevention and Treatment of Oral Diseases. *Evidence-Based Complementary and Alternative Medicine*. 2011, 1–15.
- Esmaeili-Mahani, S., Falahi, F., and Yaghoobi, M. M. (2014). Proapoptotic and Antiproliferative Effects of *Thymus caramanicus* on Human Breast Cancer Cell Line (MCF-7) and Its Interaction with Anticancer Drug Vincristine. *Evidence-Based Complementary and Alternative Medicine*. 2014, 1–7.
- Farooqui, M., Hassali, M. A., Knight, A., Shafie, A. A., Farooqui, M. A., Saleem, F., Haq, N., and Aljadhey, H. (2013). A Qualitative Exploration of Malaysian Cancer Patients' Perceptions of Cancer Screening. *BMC Public Health*. 13, 48– 54.
- Fazal, H., Abbasi, B. H., and Ahmad, N. (2014). Optimization of Adventitious Root Culture for Production of Biomass and Secondary Metabolites in *Prunella vulgaris* L. *Applied Biochemistry and Biotechnology*. 174(6), 2086–2095.
- Feild, T. S., Lee, D. W., and Holbrook, N. M. (2001). Why Leaves Turn Red in Autumn. The Role of Anthocyanins in Senescing Leaves of Red-Osier Dogwood. *Plant Physiology*. 127(2), 566–574.
- Fernandes Dos Santos, D. M. M., De Sousa Araújo, S., Roldão Lopes Duque, A. S., and Salema Fevereiro, M. P. (2003). Reverse transcription-PCR assay to verify gene integrity within plasmid constructs for plant transformation. *Plant Cell, Tissue and Organ Culture*. 74(3), 293–296.
- Figueiredo, A. C., Barroso, J. G., Pedro, L. G., and Scheffer, J. J. C. (2008). Factors Affecting Secondary Metabolite Production in Plants : Volatile Components and Essential Oils. *Flavour and Fragrance Journal*. 23(4), 213–226.
- Fior, S., and Gerola, P. D. (2009). Impact of Ubiquitous Inhibitors on the GUS Gene Reporter System: Evidence from the Model Plants Arabidopsis, Tobacco and Rice and Correction Methods for Quantitative Assays of Transgenic and Endogenous GUS. *Plant Methods*. 5, 19–30.

- Firouzi, A., Mohammadi, S. A., Khosrowchahli, M., Movafeghi, A., and Hasanloo, T. (2013). Enhancement of Silymarin Production in Cell Culture of Silybum marianum (L) Gaertn by Elicitation and Precursor Feeding. Journal of Herbs, Spices and Medicinal Plants. 19(3), 262–274.
- Fogh, J., and Trempe, G. (1975). New Human Tumor Cell Lines. In *Human Tumor Cell In Vitro* (pp. 115–159). New York: Springer US.
- Forkmann, G. (1991). Flavonoids as Flower Pigments: The Formation of the Natural Spectrum and its Extension by Genetic Engineering. *Plant Breeding*. 106(1), 1– 26.
- Franklin, G., Oliveira, M., and Dias, A. C. P. (2007). Production of Transgenic *Hypericum perforatum* Plants Via Particle Bombardment-Mediated Transformation of Novel Organogenic Cell Suspension Cultures. *Plant Science*. 172(6), 1193–1203.
- Freshney, R. I. (2005). *Culture of Animal Cells: A Manual of Basic Technique*. United States: John Wiley and Son, Inc.
- Frydoonfar, H. R., Mcgrath, D. R., and Spigelman, A. D. (2003). The Variable Effect on Proliferation of a Colon Cancer Cell Line by the Citrus Fruit Flavonoid Naringenin. *Colorectal Disease*. 5(2), 149–152.
- Füzfai, Z., and Molnár-Perl, I. (2007). Gas chromatographic Mass Spectrometric Fragmentation Study of Flavonoids as Their Trimethylsilyl Derivatives : Analysis of Flavonoids, Sugars, Carboxylic and Amino Acids in Model Systems and in Citrus Fruits. *Journal of Chromatography A*. 1149(1), 88–101.
- Gao, X., Zhu, C., Jia, W., Gao, W., Qiu, M., Zhang, Y., and Xiao, P. (2005). Induction and Characterization of Adventitious Roots Directly from the Explants of *Panax notoginseng*. *Biotechnology Letters*. 27(22), 1771–1775.
- Gao, F., Yong, Y., and Dai, C. (2011). Effects of Endophytic Fungal Elicitor on Two Kinds of Terpenoids Production and Physiological Indexes in *Euphorbia* pekinensis Suspension Cells. Journal of Medicinal Plants Research. 5(18), 4418–4425.
- Geran, R. I., Greenberg, N. H., Macdolnald, M. M., Schumacher, A., and Abbott, B. J. (1972). Protocols for Screening Chemical Agents and Natural Products Against Animal Tumors and Other Biological Systems. *Cancer Chemotherapy Reports*. 3(2), 1–103.

- Gey, G., Coffman, W. D., and Kubicek, M. T. (1952). Tissue Culture Studies of the Proliferative Capacity of Cervical Carcinoma and Normal Epithelium. *Cancer Research*. 12(4), 254–265.
- Ghasemzadeh, A., Jaafar, H. Z. E., and Karimi, E. (2012). Involvement of Salicylic Acid on Antioxidant and Anticancer Properties, Anthocyanin Production and Chalcone Synthase Activity in Ginger (*Zingiber officinale* Roscoe) Varieties. *International Journal of Molecular Sciences*. 13(11), 14828–14844.
- Gnasekaran, P., Jeyanthi, J., Antony, J., Uddain, J., and Subramaniam, S. (2014). Agrobacterium - Mediated Transformation of the Recalcitrant Vanda Kasem's Delight Orchid with Higher Efficiency. The Scientific World Journal. 2014, 1– 10.
- Graham, V. A. W. (1988). Delimitation and infra-generic classification of *Justicia* (Acanthaceae). *Kew Bulletin*. 43(4), 551–624.
- Greenwell, M., and Rahman, P. K. S. M. (2015). Medicinal Plants: Their Use in Anticancer Treatment. *International Journal of Pharmaceutical Sciences and Research*. 6(10), 4103–4112.
- Grzegorczyk, I., and Wysokińska, H. (2009). The Effect of Methyl Jasmonate on Production of Antioxidant Compounds in Shoot Cultures of Salvia officinalis L. Herba Polonica. 55(3), 238–243.
- Gupta, P., Sharma, S., and Saxena, S. (2015). Biomass Yield and Steviol Glycoside Production in Callus and Suspension Culture of *Stevia rebaudiana* Treated with Proline and Polyethylene Glycol. *Applied Biochemistry and Biotechnology*. 176(3), 863–874.
- Gutha, L. R., Casassa, L. F., Harbertson, J. F., and Naidu, R. A. (2010). Modulation of Flavonoid Biosynthetic Pathway Genes and Anthocyanins due to Virus Infection in Grapevine (*Vitis vinifera* L.) Leaves. *BMC Plant Biology*. 10(187), 1–18.
- Hamerski, D., Beier, R. C., Kneusel, R. E., Matern, U., and Himmelspach, K. (1990). Accumulation of Coumarins in Elicitor-Treated Cell Suspension Cultures of *Ammi majus*. *Phytochemistry*. 29(4), 1137–1142.
- Han, J., Talorete, T. P. N., Yamada, P., and Isoda, H. (2009). Anti-Proliferative and Apoptotic Effects of Oleuropein and Hydroxytyrosol on Human Breast Cancer MCF-7 Cells. *Cytotechnology*. 59(1), 45–53.

- Hao, X., Shi, M., Cui, L., Xu, C., Zhang, Y., and Kai, G. (2015). Effects of Methyl Jasmonate and Salicylic Acid on Tanshinone Production and Biosynthetic Gene Expression in Transgenic Salvia miltiorrhiza Hairy Roots. Bliotechnology and Applied Biochemistry. 62(1), 24–31.
- Harborne, J. B. (1988). *The Flavonoids: Advance in Research Since 1980*. London: Chapman and Hall.
- Hendra, R., Ahmad, S., Oskoueian, E., Sukari, A., and Shukor, M. Y. (2011). Antioxidant , Anti-Inflammatory and Cytotoxicity of *Phaleria macrocarpa* (Boerl.) Scheff Fruit. *BMC Complementary and Aternative Medicine*. 11(1), 110–120.
- Hichri, I., Barrieu, F., Bogs, J., Kappel, C., Delrot, S., and Lauvergeat, V. (2011).
 Recent Advances in the Transcriptional Regulation of the Flavonoid Biosynthetic Pathway. *Journal of Experimental Botany*. 62(8), 2465–2483.
- Hidayat, T., Abdullah, F. I., Kuppusamy, C., Samad, A. A., and Wagiran, A. (2012).
 Molecular Identification of Malaysian Pineapple Cultivar based on Internal Transcribed Spacer Region. *APCBEE Procedia*. 4(6), 146–151.
- Honda, M., Muramoto, Y., Kuzuguchi, T., Sawano, S., Machida, M., and Koyama,
 H. (2002). Determination of Gene Copy Number and Genotype of Transgenic *Arabidopsis thaliana* by Competitive PCR. *Journal of Experimental Botany*. 53(373), 1515–1520.
- Hu, Z., Wu, Y., Li, W., and Gao, H.-H. (2006). Factors Affecting Agrobacterium tumefaciens -Mediated Genetic Transformation of Lycium barbarum L. In Vitro Cellular and Development Biology-Plant. 42(5), 461–466.
- Huang, W.-W., Chiu, Y.-J., Fan, M.-J., Lu, H.-F., Yeh, H.-F., Li, K.-F., Chen, P.-Y., Chung, J.-G., and Yang, J.-S. (2010). Kaempferol Induced Apoptosis Via Endoplasmic Reticulum Stress and Mitochondria- Dependent Pathway in Human Osteosarcoma U-2 OS Cells. *Molecular Nutrition and Food Research*. 54(11), 1585–1595.
- Huang, Y., Lin, M., Chao, Y., Huang, C., Tsai, Y., and Wu, P. (2014). Anti-Oxidant Activity and Attenuation of Bladder Hyperactivity by the Flavonoid Compound Kaempferol. *International Journal of Urology*. 21(1), 94–98.

- Huet, C., Sahuquillo-Merino, C., Coudrier, E., and Louvard, D. (1987). Absorptive and Mucus-Secreting Subclones Isolated from a Multipotent Intestinal Cell Line (HT-29) Provide New Models for Cell Polarity and Terminal Differentiation. *The Journal of Cell Biology*. 105(1), 345–357.
- Ibrahim, M. J., Wan-Nor Izzah, W. M. Z., Narimah, A. H. H., Nurul Asyikin, Z., Siti-Nur Shafinas, S. A. R., and Froemming, G. A. (2011). Anti-Proliperative and Antioxidant Effects of *Tinospora crispa* (Batawali). *Biomedical Research*. 22(1), 57–62.
- Iida, A., Seki, M., Kamada, M., Yamada, Y., and Morikawa, H. (1990). Gene Delivery into Cultured Plant Cells by DNA-Coated Gold Particles Accelerated by a Pneumatic Particle Gun. *Theoretical and Applied Genetics*. 80(6), 813– 816.
- Ishnava, K. B., Patel, T., and Chauhan, J. B. (2012). Study of Genetic Transformation of Medicinal Plants, Withania somnifera (L.) Dunal by Agrobacterium tumefaciens (MTCC-431). Asian Journal of Experimental Biological Sciences. 3(3), 536–542.
- Islam, T., Bhoo-Pathy, N., Su, T. T., Majid, H. A., Nahar, A. M., Ng, C. G., Dahlui, M., Hussain, S., Cantwell, M., Murray, L., and Taib, N. A. (2015). The Malaysian Breast Cancer Survivorship Cohort (MyBCC): A Study Protocol. *BMJ Open.* 5(10), 1–7.
- Iweala, E. E. J., Liu, F.-F., Cheng, R.-R., Li, Y., Omonhinmin, C. A., and Zhang, Y.-J. (2015). Anti-Cancer and Free Radical Scavenging Activity of Some Nigerian Food Plants *In Vitro*. *International Journal of Cancer Research*. 11(1), 41–51.
- Jalil, M. A. A., Shuid, A. N., and Muhammad, N. (2012). Role of Medicinal Plants and Natural Products on Osteoporotic Fracture Healing. *Evidence-Based Complementary and Alternative Medicine*. 2012, 1–7.
- Jamalnasir, H., Wagiran, A., Shaharuddin, N., and Samad, A. A. (2014). Molecular Cloning and Characterization of a cDNA Encoding a Polyketide Synthase from *Melastoma decemfidum. Biologia*. 69(11), 1482–1491.
- Janarthanam, B., and Sumathi, E. (2010). In Vitro Regeneration of Justicia gendarussa Burm. f. Libyan Agriculture Research Center Journal International. 1(5), 284–287.

- Janna, O. A., Maziah, M., Ahmad Parveez, G. K. A., and Saleh, K. (2006). Factors Affecting Delivery and Transient Expression of β-Glucuronidase Gene in *Dendrobium* Sonia Protocorm- Like-Body. *African Journal of Biotechnology*. 5(2), 88–94.
- Jantan, I., Hikmah, N. H., Septama, A. W., Murad, S., and Mesaik, M. A. (2011). Inhibition of Chemiluminescence and Chemotactic Activity of Phagocytes *In Vitro* by the Extracts of Selected Medicinal Plants. *Journal of Natural Medicines*. 65(2), 400–405.
- Jefferson, R. A. (1987). Assaying Chimeric Genes in Plants : The GUS Gene Fusion System. *Plant Molecular Biology Reporter*. 5(4), 387–405.
- Jefferson, R. A., Kavanagh, T. A., and Bevan, M. W. (1987). GUS Fusions: βglucuronidase as a Sensitive and Versatile Gene Fusion Marker in Higher Plants. *The EMBO Journal*. 6(13), 3901–3907.
- Jiménez-medina, E., Garcia-lora, A., Paco, L., Algarra, I., Collado, A., and Garrido, F. (2006). A New Extract of the Plant *Calendula officinalis* Produces a Dual *In Vitro* Effect: Cytotoxic Anti-Tumor Activity and Lymphocyte Activation. BMC *Cancer.* 14(1), 119–132.
- Jindal, A., Kumar, P., and Singh, G. (2012). In Vitro Antimicrobial Activity of Tribulus terrestris L. International Journal of Pharmacy and Pharmaceutical Sciences. 4(3), 270–272.
- Jones, E. L., Prosnitz, L. R., Dewhirst, M. W., Marcom, P. K., Hardenbergh, P. H., Marks, L. B., Brizel, D. M., and Vujaskovic, Z. (2004). Thermochemoradiotherapy Improves Oxygenation in Locally Advanced Breast Cancer. *Clinical Cancer Research*. 10(13), 4287–4293.
- Jones, L. W., Courneya, K. S., Fairey, A. S., and Mackey, J. R. (2004). Effects of an Oncologist's Recommendation to Exercise on Self-Reported Exercise Behavior in Newly Diagnosed Breast Cancer Survivors : A Single-Blind, Randomized Controlled Trial. *Annals of Behavioral Medicine*. 28(2), 105–113.
- Joshi, C. G., Gopal, M., and Byregowda, S. M. (2011). Cytotoxic Activity of Tragia involucrata Extracts. American-Eurasian Journal of Toxicological Sciences. 3(2), 67–69.
- Joshi, M., Mishra, A., and Jha, B. (2011). Efficient Genetic Transformation of Jatropha curcas L . by Microprojectile Bombardment using Embryo Axes. Industrial Crops and Products. 33(1), 67–77.

- Jothimanivannan, C., Kumar, R. S., and Subramanian, N. (2010). Anti-inflammatory and Analgesic Activities of Ethanol Extract of Aerial Parts of *Justicia gendarussa* Burm. *International Journal of Pharmacology*. 6(3), 278 – 283.
- Jumtee, K., Bamba, T., and Fukusaki, E. (2009). Fast GC-FID Based Metabolic Fingerprinting of Japanese green Tea Leaf for its Quality Ranking Prediction. *Journal of Separation Science*. 32(13), 2296–2304.
- Jumtee, K., Komura, H., Bamba, T., and Fukusaki, E. (2011). Predication of Japanese Green Tea (Sen-cha) Ranking by Volatile Profiling using Gas Chromatography Mass Spectrometry and Multivariate Analysis. *Journal of Bioscience and Bioengineering*. 112(3), 252–255.
- Kalyani, C., Narasu, M. L., and Devi, Y. P. (2015). Effect of Phytochemical Kaempferol on HCT-15 and Lymphocytes. *Indian Journal of Applied Research*. 5(10), 452–454.
- Karami, O. (2008). Factors Affecting Agrobacterium Mediated Transformation of Plants. Transgenic Plant Journal. 2(2), 127–137.
- Karcher, S. J. (2002). Blue Plants: Transgenic Plants with the Gus Reporter Gene. In Association for Biology Laboratory Education (ABLE). Proceedings of the 23rd Workshop, 29–42.
- Karimi, E., Oskoueian, E., Hendra, R., Oskoueian, A., and Jaafar, H. Z. E. (2012). Phenolic Compounds Characterization and Biological Activities of *Citrus aurantium* Bloom. *Molecules*. 17(2), 1203–1218.
- Karuppusamy, S. (2009). A Review on Trends in Production of Secondary Metabolites from Higher Plants by *In Vitro* Tissue, Organ and Cell Cultures. *Journal of Medicinal Plants Research*. 3(13), 1222–1239.
- Kasote, D. M., Katyare, S. S., Hegde, M. V, and Bae, H. (2015). Significance of Antioxidant Potential of Plants and its Relevance to Therapeutic Applications. *International Journal of Biological Sciences*. 11(8), 982–991.
- Kavitha, K., Sridevi sangeetha, K. S., Sujatha, K., and Umamaheswari, S. (2014). Phytochemical and Pharmacological Profile of Justicia gendarussa Burm f. -Review. *Journal of Pharmacy Reseach*. 8(7), 990–997.
- Kibbe, W. A. (2007). OligoCalc: An Online Oligonucleotide Properties Calculator. Nucleic Acids Research. 35(2), 43–46.
- Kikkert, J. R. (1993). The Biolistic ® PDS-1000/He Device. *Plant Cell, Tissue and Organ Culture*. 33(3), 221–226.

- Kikkert, J. R., Vidal, J. R., and Reisch, B. I. (2005). Stable Transformation of Plant Cells by Particle Bombardment/ Biolistics. In *Methods in Molecular Biology*. *Transgenic Plants: Methods and Protocols* (Vol. 286, pp. 61–78). Totowa, New Jersey: Humana Press Inc.
- Kim, Y.-S., Hahn, E.-J., and Yeung, E. C. (2003). Lateral Root Development and Saponin Accumulation as Affected by IBA or NAA in Adventitious Root Cultures of *Panax ginseng* C. A. Meyer. *In Vitro Cellular and Development Biology-Plant*. 39(2), 245–249.
- Kim, Y.-S., Hahn, E.-J., Murthy, H. N., and Paek, K.-Y. (2004a). Adventitious Root Growth and Ginsenoside Accumulation in *Panax ginseng* Cultures as Affected by Methyl Jasmonate. *Biotechnology Letters*. 26(21), 1619–1622.
- Kim, Y.-S., Hahn, E.-J., Murthy, H. N., and Paek, K.-Y. (2004b). Effect of Polyploidy Induction on Biomass and Ginsenoside Accumulations in Adventitious Roots of Ginseng. *Journal of Plant Biology*. 47(4), 356–360.
- Kim, N. S., Young, W. H., Shin, H., Son, S. H., Wu, S. J., and Kim, Y. S. (2007). Simultaneous Quantification of 14 Ginsenosides in *Panax ginseng* C. A. Meyer (Korean Red Ginseng) by HPLC-ELSD and its Application to Quality Control. *Journal of Pharmaceutical and Biomedical Analysis*. 45(1), 164–170.
- Kirana, C., Record, I. R., Mcintosh, G. H., and Jones, G. P. (2003). Screening for Antitumor Activity of 11 Species of Indonesian Zingiberaceae Using Human MCF-7 and HT-29 Cancer Cells. *Pharmaceutical Biology*. 41(4), 271–276.
- Kiren, Y., Deguchi, J., Hirasawa, Y., Morita, H., and Prajogo, B. (2014). Justidrusamides A-D, New 2-Aminobenzyl Alcohol Derivatives from *Justicia* gendarussa. Journal of Natural Medicines. 68(4), 754–758.
- Kirtikar, K. R., and Basu, B. D. (1993). Indian Medicinal Plants 2nd (Ed) Vol 3. India: Indian Press.
- Kirtikar, K. R., and Basu, B. M. (2005). Indian Medicinal Plants. Vol. 1, Apura Krishna Bose. India: Indian Press.
- Koes, R. E., Quattrocchio, F., and Mol, J. N. M. (1994). The Flavonoid Biosynthesis Pathway in Plants: Function and Evolution. *BioEssays*. 16(2), 123–132.
- Komari, T., Hiei, Y., Saito, Y., Murai, N., and Kumashiro, T. (1996). Vectors Carrying Two Separate T-DNAs for Co-transformation of Higher Plants Mediated by Agrobacterium tumefaciens and Segregation of Transformants Free from Selection Markers. *The Plant Journal*. 10(1), 165–174.

- Kontturi, J., Osama, R., Deng, X., Bashandy, H., Albert, V. A., and Teeri, T. H. (2017). Functional Characterization and Expression of GASCL1 and GASCL2, Two Anther-Specific Chalcone Synthase Like Enzymes from *Gerbera hybrida*. *Phytochemistry*. 134, 38–45.
- Kooter, J. M., Matzke, M. A., and Meyer, P. (1999). Listening to the Silent Genes: Transgene Silencing, Gene Regulation and Pathogen Control. *Trends in Plant Science*. 4(9), 340–347.
- Krishna, K. L., Mruthunjaya, K., and Patel, J. A. (2009). Antioxidant and Hepatoprotective Activity of Leaf Extract of *Juscticia gendarussa* Burm. *International Journal of Biological Chemistry*. 3(3), 99–110.
- Krishna, K. L., Mehta, T. A., and Patel, J. A. (2010). *In-Vitro* Hepatoprotective Activity of *Justicia gendarussa* Stem on Isolated Rat Hepatocytes. *Pharmacologyonline*. 2, 9–13.
- Kubasek, W. L., Shirley, B. W., Mckillop, A., Goodman, H. M., Briggs, W., and Ausubel, F. M. (1992). Regulation of Flavonoid Biosynthetic Genes in Germinating *Arabidopsis* Seedlings. *The Plant Cell*. 4(10), 1229–1236.
- Kumar, K. S., Vijayan, V., Bhaskar, S., Krishnan, K., Shalini, V., and Helen, A. (2012). Anti-Inflammatory Potential of an Ethyl Acetate Fraction Isolated from *Justicia gendarussa* Roots through Inhibition of iNOS and COX-2 Expression Via NF- j B Pathway. *Cellular Immunology*. 272(2), 283–289.
- Kumar, S., and Pandey, A. K. (2013). Chemistry and Biological Activities of Flavonoids: An Overview. *The Scientific World Journal*. 2013, 1–16.
- Kuskoski, E. M., Rios, J. J., Bueno, J. M., Fett, R., Troncoso, A. M., and Asuero, A. G. (2012). Capillary Gas Chromatography-Mass Spectrometry (CGC-MS) Analysis and Antioxidant Activities of Phenolic and Derivatives. *The Open Analytical Chemistry Journal*. 6(1), 1–8.
- Latiff, A. (2005). Valuing the Biodiversity of Medicinal Plant Species in Malaysia. In Sustainable Management and Utilization of Medicinal Plant Resources: Proceedings of the International Conference on Medicinal Plants: Kuala Lumpur (Malaysia). Kuala Lumpur: 3–16.
- Lazzeroni, M., Serrano, D., Dunn, B. K., Heckman-Stoddard, B. M., Lee, O., Khan, S., and Decensi, A. (2012). Oral Low Dose and Topical Tamoxifen for Breast Cancer Prevention: Modern Approaches for an Old Drug. *Breast Cancer Research*. 14(5), 214–225.

- Le Bail, J. C., Varnat, F., Nicolas, J. C., and Habrioux, G. (1998). Estrogenic and Antiproliferative Activities on MCF-7 Human Breast Cancer Cells by Flavonoids. *Cancer Letters*. 130(1), 209–216.
- Lee, N. Y., Jo, C., Sohn, S. H., Kim, J. K., and Byun, M. W. (2006). Effects of Gamma Irradiation on the Biological Activity of Green Tea Byproduct Extracts and a Comparison with Green Tea Leaf Extracts. *Journal of Food Science*. 71(4), C269–C274.
- Lee, Y. S., Yang, T.-J., Park, S.-U., Baek, J. H., Wu, S., and Lim, K.-B. (2011). Induction and Proliferation of Adventitious Roots from *Aloe vera* Leaf Tissues for *In Vitro* Production of Aloe-Emodin. *Plant Omics Journal*. 4(4), 190–194.
- Lee, E.-J., and Paek, K.-Y. (2012). Enhanced Productivity of Biomass and Bioactive Compounds through Bioreactor Cultures of *Eleutherococcus koreanum* Nakai Adventitious Roots Affected by Medium Salt Strength. *Industrial Crops and Products*. 36(1), 460–465.
- Lee, C.-F., Yang, J. A. I. S., Tsai, F.-J., Chiang, N.-N., Lu, C.-C., Huang, Y.-S., Chen, C., and Chen, F.-A. (2016). Kaempferol Induces ATM / p53-Mediated Death Receptor and Mitochondrial Apoptosis in Human Umbilical Vein Endothelial Cells. *International Journal of Oncology*. 48(5), 2007–2014.
- Leung, H. W. C., Lin, C.-J., Hour, M.-J., Yang, W.-H., Wang, M.-Y., and Lee, H.-Z. (2007). Kaempferol Induces Apoptosis in Human Lung Non-Small Carcinoma Cells Accompanied by an Induction of Antioxidant Enzymes. *Food and Chemical Toxicology*. 45(10), 2005–2013.
- Li, X.-G., Chen, S.-B., Lu, Z.-X., and Chang, T.-J. (2002). Impact of Copy Number on Transgene Expression in Tobacco. *Acta Botanica Sinica*. 44(1), 120–123.
- Li, L., and Zhang, C. R. (2006). Production of Puerarin and Isoflavones in Cell Suspension Cultures of *Pueraria lobata* (Willd.): Effects of Medium Supplementation with Casein Hydrolysate and Coconut Milk. *Journal of Environmental Biology*. 27(1), 21–26.
- Li, H. Q., Kang, P. J., Li, M. L., and Li, M. R. (2007). Genetic Transformation of *Torenia fournieri* using the PMI/Mannose Selection System. *Plant Cell, Tissue* and Organ Culture. 90(1), 103–109.
- Li, F., Awale, S., Tezuka, Y., and Kadota, S. (2008). Cytotoxic Constituents from Brazilian Red Propolis and their Structure – Sctivity Relationship. *Bioorganic* and Medicinal Chemistry. 16(10), 5434–5440.

- Li, W., Du, B., Wang, T., Wang, S., and Zhang, J. (2009). Kaempferol Induces Apoptosis in Human HCT116 Colon Cancer Cells Via the Ataxia -Telangiectasia Mutated - p53 Pathway with the Involvement of p53 Upregulated Modulator of Apoptosis. *Chemico-Biological Interactions*. 177(2), 121–127.
- Li, R., Mei, J., and Liu, G. (2011). Kaempferol-Induced Apoptosis of Human Esophageal Squamous Carcinoma Eca-109 Cells and the Mechanism. *Journal of Southern Medical University*. 31(8), 1440–1442.
- Li, J., Wang, J., Li, J., Li, J., Liu, S., and Gao, W. (2015). Salicylic Acid Induces the Change in the Adventitious Root of *Glycyrrhiza uralensis* Fisch.: Bioactive Compounds and Antioxidant Enzymes. *Research on Chemical Intermediates*. 42(2), 1503–1519.
- Li, H., Liang, J., Chen, H., Ding, G., Ma, B., and He, N. (2016). Evolutionary and Functional Analysis of *Mulberry* Type III Polyketide Synthases. *BMC Genomics*. 17(1), 540–558.
- Lievre, K., Hehn, A., Tran, T. L. M., Gravot, A., Thomasset, B., Bourgaud, F., and Gontier, E. (2005). Genetic Transformation of the Medicinal Plant *Ruta* graveolens L. by an Agrobacterium tumefaciens - Mediated Method. Plant Science. 168(4), 883–888.
- Liggins, J., Bluck, L. J. C., Runswick, S., Atkinson, C., Coward, W. A., and Bingham, S. A. (2000). Daidzein and Genistein Content of Fruits and Nuts. *The Journal of Nutrional Biochemistry*. 11(1), 326–331.
- Lijuan, C., Huiming, G., Yi, L., and Hongmei, C. (2015). Chalcone Synthase EaCHS1 from *Eupatorium adenophorum* Functions in Salt Stress Tolerance in Tobacco. *Plant Cell Reports*. 34(5), 885–894.
- Ling, A. P. K., Chin, M. F., and Hussein, S. (2009). Adventitious Root Production of *Centella asiatica* in Response to Plant Growth Regulators and Sucrose Concentrations. *Medicinal and Aromatic Plant Science and Bitechnology*. 3(1), 36–41.
- Liu, H., Zang, C., Fenner, M. H., Possinger, K., and Elstner, E. (2003). PPAR γ Ligands and ATRA Inhibit the Invasion of Human Breast Cancer Cells In Vitro. Breast Cancer Research and Treatment. 79(1), 63–74.
- Liu, E.-H., Qi, L.-W., Cao, J., Li, P., Li, C.-Y., and Peng, Y.-B. (2008). Advances of Modern Chromatographic and Electrophoretic Methods in Separation and Analysis of Flavonoids. *Molecules*. 13(10), 2521–2544.

- Liu, H., Wang, J., Gao, W., Wang, Q., Zhang, L., and Man, S. (2014). Optimization and Quality Assessment of Adventitious Roots Culture in *Panax quinquefolium* L. Acta Physiologiae Plantarum. 36(3), 713–719.
- Liu, Y., An, W., and Gao, A. (2016). Protective Effects of Naringenin in Cardiorenal Syndrome. *Journal of Surgical Research*. 203(2), 416–423.
- Manuhara, Y. S. W., Kristanti, A. N., Utami, E. S. W., and Yachya, A. (2015). Effect of Sucrose and Potassium Nitrate on Biomass and Saponin Content of Talinum *paniculatum* Gaertn. Hairy Root in Balloon-Type Bubble Bioreactor. *Asian Pacific Journal of Tropical Biomedicine*. 5(12), 1027–1032.
- Marfe, G., Tafani, M., Indelicato, M., Sinibaldi-salimei, P., Reali, V., Pucci, B., Fini,
 M., and Russo, M. A. (2009). Kaempferol Induces Apoptosis in Two Different
 Cell Lines Via Akt Inactivation, Bax and SIRT3 Activatin, and Mitochondrial
 Dysfunction. *Journal of Cellular Biochemistry*. 106(4), 643–650.
- Markham, K. R. (1982). *Techniques of Flavonoid Identification*. London: Academic Press.
- Masa, C. V., Díaz, T. S., Gallego, J. C. A., and Lobón, N. C. (2016). Quantitative Variation of Flavonoids and Diterpenes in Leaves and Stems of *Cistus ladanifer* L. at Different Ages Cristina. *Molecules*. 21(275), 1–14.
- Matchett, M. D., Mackinnon, S. L., Sweeney, M. I., Gottschall-Pass, K. T., and Hurta, R. A. R. (2005). Blueberry Flavonoids Inhibit Matrix Metalloproteinase Activity in DU145 Human Prostate Cancer Cells. *Biochemistry and Cell Biology*. 643(5), 637–643.
- Mazid, M., Khan, T. A., and Mohammad, F. (2011). Role of Nitric Oxide in Regulation of H₂O₂ Mediating Tolerance of Plants to Abiotic Stress: A Synergistic Signalling Approach. *Journal of Stress Physiology and Biochemistry*. 7(2), 34–74.
- Meyer, P., Heidmann, I., Forkmann, G., and Saedler, H. (1987). A New Petunia Flower Colour generated by Transformation of a Mutant with a Maize Gene. *Nature*. 330(6149), 677–678.
- Minuti, L., and Pellegrino, R. (2008). Determination of Phenolic Compounds in Wines by Novel Matrix Solid-Phase Dispersion Extraction and Gas Chromatography/Mass Spectrometry. *Journal of Chromatography A*. 1185(1), 23–30.

- Mizukami, Y., Kohata, K., Yamaguchi, Y., Hayashi, N., Sawai, Y., Chuda, Y., Ono, H., Yada, H., and Yoshida, M. (2006). Analysis of Acrylamide in Green Tea by Gas Chromatography – Mass Spectrometry. *Journal of Agricultural and Food Chemistry*. 54(19), 7370–7377.
- Moeso, S., and Agus, P. (1985). Report of The Trip to Jayapura Sentani (Irian Jaya). Faculty of Biology Gadjah Mada University, Yogyakarta.
- Mohamed, S. M., Ali, A. M., Rahmani, M., Dhaliwal, J. S., and Yusoff, K. (2000). Apoptotic and Neurotic Cell Death Manifestations in Leukemic Cells Treated with Methylgerambulin a Sulphone from *Glycosmis calcicola*. *The Journal of Biochemistry Molecular Biology and Biophysics*. 4(4), 253–261.
- Molnár-Perl, I., and Füzfai, Z. (2005). Chromatographic, Capillary Electrophoretic and Capillary Electrochromatographic Techniques in the Analysis of Flavonoids. *Journal of Chromatography A*. 1073(1-2), 201–227.
- Mondal, T. K., Bhattacharya, A., Ahuja, P. S., and Chand, P. K. (2001). Transgenic Tea [*Camellia sinensis* (L.) O. Kuntze cv. Kangra Jat] Plants obtained by *Agrobacterium*-Mediated Transformation of Somatic Embryos. *Plant Cell Reports*. 20(8), 712–720.
- Moniruzzaman, M., Alamgir, M., Chowdhury, Z., Rahman, M. A., Sulaiman, S. A., and Gan, S. H. (2014a). Determination of Mineral , Trace Element , and Pesticide Levels in Honey Samples Originating from Different Regions of Malaysia Compared to Manuka Honey. *BioMed Research International*. 2014, 1–10.
- Moniruzzaman, M., An, C. Y., Rao, P. V., Hawlader, M. N. I., Azlan, S. A. B. M., Sulaiman, S. A., and Gan, S. H. (2014b). Identification of Phenolic Acids and Flavonoids in Monofloral Honey from Bangladesh by High Performance Liquid Chromatography: Determination of Antioxidant Capacity. *BioMed Research International*. 2014, 1–11.
- Moon, J.-K., and Shibamoto, A. (2009). Antioxidant Assays for Plant and Food Components. *Journal of Agricultural and Food Chemistry*. 57(5), 1655–1666.
- Mosmann, T. (1983). Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity Assays. *Journal of Immunological Methods*. 65(1-2), 55–63.

- Mousavi, M., Mousavi, A., Habashi, A. A., and Arzani, K. (2009). Optimization of Physical and Biological Parameters for Transient Expression of uidA Gene in Embryogenic Callus of Date Palm (*Phoenix dactylifera* L.) Via Particle Bombardment. *African Journal of Biotechnology*. 8(16), 3721–3730.
- Mruthunjaya, K., and Hukkeri, V. I. (2007). Antioxidant and Free Radical Scavenging Potential of *Justicia gendarussa* Burm. Leaves *In Vitro. Natural Product Sciences*. 13(3), 199–206.
- Muñoz-Espada, A. C., and Watkins, B. A. (2006). Cyanidin Attenuates PGE 2 Production and Cyclooxygenase-2 Expression in LNCaP Human Prostate Cancer Cells. *Journal of Nutrition Biochemistry*. 17(9), 589–596.
- Murashige, T., and Skoog, F. (1962). A Revised Medium for Rapid Growth and Bio Assays with Tobacco Tissue Cultures. *Physiologia Plantarum*. 15(3), 473–497.
- Murthy, H. N., Lee, E.-J., and Paek, K.-Y. (2014). Production of Secondary Metabolites from Cell and Organ Cultures: Strategies and Approaches for Biomass Improvement and Metabolite Accumulation. *Plant Cell, Tissue and Organ Culture*. 118(1), 1–16.
- Mustafa, R. A., Abdul Hamid, A., Mohamed, S., and Bakar, F. A. (2010). Total Phenolic Compounds, Flavonoids, and Radical Scavenging Activity of 21 Selected Tropical Plants. *Journal of Food Science*. 75(1), C28–C35.
- Naik, P. M., and Al-Khayri, J. M. (2016). Abiotic and Biotic Elicitors Role in Secondary Metabolites Production through *In Vitro* Culture of Medicinal Plants. In *Abiotic and Biotic Stress in Plants - Recent Advances and Future Perspectives* (pp. 247–277). Croatia: InTech.
- Nakatsuma, A., Fukami, T., Suzuki, T., Furuishi, T., Tomono, K., and Hidaka, S. (2010). Effects of Kaempferol on the Mechanisms of Drug Resistance in the Human Glioblastoma Cell Line T98G. *Pharmazie*. 65(5), 379–383.
- Namdeo, A. G. (2007). Plant Cell Elicitation for Production of Secondary Metabolites : A Review. *Pharmacognosy Reviews*. 1(1), 69–79.
- Navarro, E., Alonso, S. J., Trujillo, J., Jorge, E., and Perèz, C. (2001). General Behavior, Toxicity, and Cytotoxic Activity of Elenoside, a Lignan from *Justicia hyssopifolia*. *Journal of Natural Products*. 64(1), 134–135.

- Nguyen, T. T. T., Tran, E., Ong, C. K., Lee, S. K., Do, P. T., Huynh, T. T., Nguyen, T. H., Lee, J. J., Tan, Y., Ong, C. S., and Huynh, H. (2003). Kaempferol-Induced Growth Inhibition and Apoptosis in A549 Lung Cancer Cells is Mediated by Activation of MEK-MAPK. *Journal of Cellular Physiology*. 197(1), 110–121.
- Niestroy, J., Barbara, A., Herbst, K., Rode, S., Liempt, M. Van, and Roos, P. H. (2011). Single and Concerted Effects of Benzo[a]pyrene and Flavonoids on the AhR and Nrf2-Pathway in the Human Colon Carcinoma Cell Line Caco-2. *Toxicology in Vitro*. 25(3), 671–683.
- Ningsih, I. Y., Purwanti, D. I., Wongso, S., Prajogo, B. E. W., and Indrayanto, G. (2015). Metabolite Profiling of *Justicia gendarussa* Burm. f. Leaves using UPLC-UHR-QTOF-MS. *Scientia Pharmaceutica*. 83(3), 489–500.
- Nisha, K. K., Seettha, K., Rajmohan, K., and Purushothama, M. G. (2003). Agrobacterium tumefaciens - Mediated Transformation of Brahmi [Bacopa monniera (L.) Wettst.], a Popular Medicinal Herb of India. Current Science. 85(1), 85–89.
- Ochiai, A., Miyata, S., Iwase, M., Shimizu, M., Inoue, J., and Sato, R. (2016). Kaempferol Stimulates Gene Expression of Low-Density Lipoprotein Receptor through Activation of Sp1 in Cultured Hepatocytes. *Scientific Reports*. 6, 1–10.
- Ogo, Y., Ozawa, K., Ishimaru, T., Murayama, T., and Takaiwa, F. (2013). Transgenic Rice Seed Synthesizing Diverse Flavonoids at High Levels: A New Platform for Flavonoid Production with Associated Health Benefits. *Plant Biotechnology Journal*. 11(6), 734–746.
- Osborn, J. (2000). A Review of Radioactive and Non-Radioactive-Based Techniques used in Life Science Applications-Part I: Blotting Techniques. *Life Science News (Amersham Biosciences)*. 6, 1–4.
- Pallant, J. (2007). SPSS Survival Manual. United States: McGraw-Hill.
- Park, J.-H., Lee, J.-W., Paik, H.-D., Cho, S. G., Nah, S.-Y., Park, Y.-S., and Han, Y. S. (2010). Cytotoxic Effects of 7- O -Butyl Naringenin on Human Breast Cancer. *Food Science and Biotechnology*. 19(3), 717–724.
- Park, K.-I., Park, H., Kim, M.-K., Hong, G.-E., Nagappan, A., Lee, H., Won, C.-K., Shin, S.-C., and Kim, G.-S. (2014). Flavonoids Identified from Korean *Citrus aurantium* L. Inhibit Non-Small Cell Lung Cancer Growth In Vivo and In Vitro. Journal of Functional Foods. 7(1), 287–297.

- Parveez, G. K. A., Chowdhury, M. K. U., and Saleh, N. M. (1997). Physical Parameters Affecting Transient GUS Gene Expression in Oil Palm (*Elaeis* guineensis Jacq.) using the Biolistic Device. Industrial Crops and Products. 6(1), 41–50.
- Patel, S. S., and Zaveri, M. N. (2012). Cytotoxic Activity to find Bioactive Compound from Justicia gendarussa using Brine Shrimp Lethality Assay. Asian Journal of Traditional Medicines. 7(3), 102–108.
- Patil, J. G., Ahire, M. L., Nitnaware, K. M., Panda, S., Bhatt, V. P., Kishor, P. B. K., and Nikam, T. D. (2013). *In Vitro* Propagation and Production of Cardiotonic Glycosides in Shoot Cultures of *Digitalis purpurea* L. by Elicitation and Precursor Feeding. *Applied Microbiology and Biotechnology*. 97(6), 2379– 2393.
- Paval, J., Kaitheri, S. K., Potu, K. B., Govindan, S., Kumar, R. S., Narayanan, S. N., and Moorkoth, S. (2009). Anti-Arthritic Potential of the Plant *Justicia* gendarussa Burm F. Clinics. 64(4), 357–362.
- Peer, W. A., Brown, D. E., Tague, B. W., Muday, G. K., Taiz, L., and Murphy, A. S. (2001). Flavonoid Accumulation Patterns of Transparent Testa Mutants of Arabidopsis. *Plant Physiology*. 126(2), 536–548.
- Periyanayagam, K., Umamaheswari, B., Suseela, L., Padmini, M., and Ismail, M. (2009). Evaluation of Antiangiogenic Effect of the Leaves of *Justicia* gendarussa (Burm. f) (Acanthaceae) by Chrio Allontoic Membrane Method. *American Journal of Infectious Diseases*. 5(3), 180–182.
- Philip, K., Malek, S. N. A., Sani, W., Shin, S. K., Kumar, S., Lai, H. S., Serm, L. G., and Rahman, S. N. S. A. (2009). Antimicrobial Activity of Some Medicinal Plants from Malaysia. *American Journal of Applied Sciences*. 6(8), 1613–1617.
- Phromnoi, K., Yodkeeree, S., Anuchapreeda, S., and Limtrakul, P. (2009). Inhibition of MMP-3 Activity and Invasion of the MDA- MB-231 Human Invasive Breast Carcinoma Cell Line by Bioflavonoids. *Acta Pharmacologica Sinica*. 30(8), 1169–1176.
- Prakash, G., and Srivastava, A. K. (2008). Statistical Elicitor Optimization Studies for the Enhancement of Azadirachtin Production in Bioreactor Azadirachta indica Cell Cultivation. Biochemical Engineering Journal. 40(2), 218–226.

- Qin, J. B., Wang, Y., and Zhu, C. Q. (2014). Biolistic Transformation of Wheat using the HMW-GS 1Dx5 Gene without Selectable Markers. *Genetics and Molecular Research*. 13(2), 4361–4371.
- Rahman, Z. A., Seman, Z. A., Roowi, S., Basirun, N., and Subramaniam, S. (2010).
 Production of Transgenic Indica Rice (*Oryza sativa* L.) Cv. MR 81 Via Particle
 Bombardment System. *Emirates Journal of Food and Agriculture*. 22(5), 353–366.
- Rahman, Z. A., Seman, Z. A., Basirun, N., Julkifle, A. L., Zainal, Z., and Subramaniam, S. (2011). Preliminary Investigations of Agrobacterium -Mediated Transformation in Indica Rice MR219 Embryogenic Callus using GusA Gene. African Journal of Biotechnology. 10(40), 7805–7813.
- Rajendran, P., Rengarajan, T., Nandakumar, N., Palaniswami, R., Nishigaki, Y., and Nishigaki, I. (2014). Kaempferol, a Potential Cytostatic and Cure for Inflammatory Disorders. *European Journal of Medicinal Chemistry*. 86, 103– 112.
- Ramirez-Estrada, K., Vidal-Limon, H., Hidalgo, D., Moyano, E., Golenioswki, M., Cusidó, R. M., and Palazon, J. (2016). Elicitation , an Effective Strategy for the Biotechnological Production of Bioactive High-Added Value Compounds in Plant Cell Factories. *Molecules*. 21(2), 182–206.
- Rao, S. R., and Ravishankar, G. A. (2002). Plant Cell Cultures: Chemical Factories of Secondary Metabolites. *Biotechnology Advances*. 20(2), 101–153.
- Rashid, H., Khan, M. H., Chaudhry, Z., Bano, R., and Raja, N. I. (2012). An Improved Agrobacterium - Mediated Transformation System in Wheat. *Pakistan Journal of Botany*. 44(1), 297–300.
- Ratnasooriya, W. D., Deraniyagala, S. A., and Dehigaspitiya, D. C. (2007). Antinociceptive Activity and Toxicological Study of Aqueous Leaf Extract of *Justicia gendarussa* Burm. F. in Rats. *Pharmacognosy Magazine*. 3(11), 145– 155.
- Ravanfar, S. A., Aziz, M. A., Saud, H. M., and Abdullah, J. O. (2015). Optimization of *In Vitro* Regeneration and *Agrobacterium tumefaciens* Mediated Transformation with Heat-Resistant cDNA in *Brassica oleracea* subsp. italica cv. Green Marvel. *Current Genetics*. 61(4), 653–663.

- Reddy, Y. S., Anitha, G., Nagulu, M., Reddy, M. R., Prasad, P. H., Jagath, M. J., Kumar, V.R., and Reddy, G. P. C. S. (2013). *In Vitro* Antibacterial Activity of Leaf Extracts of *Justicia gendarussa* Wild. *Der Pharmacia Lettre*. 5(5), 101– 103.
- Ren, H. J., Hao, H. J., Shi, Y. J., Meng, X. M., and Han, Y. Q. (2010). Apoptosis-Inducing Effect of Quercetin and Kaempferol on Human HL-60 Cells and its Mechanism. *Journal of Experimental Hematology/Chinese Association of Pathophysiology*. 18(3), 629–633.
- Ren, B., Qin, W., Wu, F., Wang, S., Pan, C., Wang, L., Zeng, B., Ma, S., and Liang, J. (2016). Apigenin and Naringenin Regulate Glucose and Lipid Metabolism, and Ameliorate Vascular Dysfunction in Type 2 Diabetic Rats. *European Journal of Pharmacology*. 773, 13–23.
- Riasat, R., Riasat, Z., Abbasi, B. H., Liu, C., and Khan, M. A. (2015). Silybum marianum: Adventitious Roots Induction along with Free Radical Scavenging Activity. Journal of Plant Biology Research. 4(1), 12–21.
- Rijke, E. De, Out, P., Niessen, W. M. A., Ariese, F., Gooijer, C., and Brinkman, U.
 A. T. (2006). Analytical Separation and Detection Methods for Flavonoids. *Journal of Chromatography A*. 1112(1-2), 31–63.
- Rivera, L. A., Gómez-lim, M., Fernández, F., and Loske, A. M. (2012). Physical Methods for Genetic Plant Transformation. *Physics of Life Reviews*. 9(3), 308– 345.
- Ruh, M., Zacharewski, T., Connor, K., Howell, J., Chen, I., and Safe, S. (1995). Naringenin: A Weakly Estrogenic Exhibits Antiestrogenic Bioflavonoid Activity that Exhibits Antiestrogenic Activity. *Bichemical Pharmacology*. 50(9), 1485–1493.
- Ruiz-May, E., Galaz-Ávalos, R. M., and Loyola-Vargas, V. M. (2009). Differential Secretion and Accumulation of Terpene Indole Alkaloids in Hairy Roots of *Catharanthus roseus* Treated with Methyl Jasmonate. *Molecular Biotechnology*. 41(3), 278–285.
- Russell, J. A., Roy, M. K., and Sanford, J. C. (1992). Physical Trauma and Tungsten Toxicity Reduce the Efficiency of Biolistic Transformation. *Plant Physiology*. 98(3), 1050–1056.

- Saha, M. R., Debnath, P. C., Rahman, M. A., and Islam, M. A. U. (2012). Evaluation of *In Vitro* Anthelmintic Activities of Leaf and Stem Extracts of *Justicia* gendarussa. Bangladesh Journal Pharmacological. 7(1), 50–53.
- Sahoo, L., Sugla, T., and Jaiwal, P. K. (2003). In Vitro Regeneration and Genetic Transformation of Cowpea, Mungbean, Urdbean and Azuki Bean. In Applied Genetics of Leguminosae Biotechnology (pp. 89–120). New York: Springer Netherlands.
- Sak, K. (2014). Cytotoxicity of Dietary Flavonoids on Different Human Cancer Types. *Pharmacognosy Reviews*. 8(16), 122–146.
- Sampaio, B. L., Edrada-ebel, R., and Costa, F. B. D. (2016). Effect of the Environment on the Secondary Metabolic Profile of *Tithonia diversifolia* : A Model for Environmental Metabolomics of Plants. *Scientific Reports*. 6(10), 1– 11.
- Sanford, J. C., Klein, T. M., Wolf, E. D., and Allen, N. (1987). Delivery of Substances into Cells and Tissues using a Particle Bombardment Process. *Particulate Science and Technology : An International Journal*. 5(1), 27–37.
- Sanford, J. C. (1988). The Biolistic Process. *Trends in Biotechnology*. 6(12), 299–302.
- Sanford, J. C. (1990). Biolistic Plant Transformation. *Physiologia Plantarum*. 79(1), 206–209.
- Sanford, J. C., Smith, F. D., and Russell, J. A. (1993). Optimizing the Biolistic Process for Different Biological Applications. *Methods in Enzymology*. 217, 483–509.
- Sarju, N., Samad, A. A., Ghani, M. A., and Ahmad, F. (2012). Detection and Quantification of Naringenin and Kaempferol in *Melastoma decemfidum* Extracts by GC-FID and GC-MS. *Acta Chromatographica*. 24(2), 221–228.
- Saslowsky, D., and Winkel-Shirley, B. (2001). Localization of Flavonoid Enzymes in *Arabidopsis* Roots. *The Plant Journal*. 27(1), 37–48.
- Satyavathi, V. V, Prasad, V., Lakshmi, B. G., and Sita, G. L. (2002). High Efficiency Transformation Protocol for Three Indian Cotton Varieties Via Agrobacterium tumefaciens. Plant Science. 162(2), 215–223.
- Sautter, C. (1993). Development of A Microtargeting Device for Particle Bombardment of Plant Meristems. *Plant Cell, Tissue and Organ Culture*. 33(3), 251–257.

- Schijlen, E. G. W. M., Ric De Vos, C. H., van Tunen, A. J., and Bovy, A. G. (2004). Modification of Flavonoid Biosynthesis in Crop Plants. *Phytochemistry*. 65(19), 2631–2648.
- Sharma, V., Joseph, C., Ghosh, S., Agarwal, A., Mishra, M. K., and Sen, E. (2007). Kaempferol Induces Apoptosis in Glioblastoma Cells through Oxidative Stress. *Molecular Cancer Therapeutics*. 6(9), 2544–2553.
- Sharma, K. K., Saikia, R., Kotoky, J., Kalita, J. C., and Devi, R. (2011). Antifungal Activity of Solanum melongena L, Lawsonia inermis L. and Justicia gendarussa B. against Dermatophytes. International Journal of PharmTech Research. 3(3), 1635–1640.
- Sharma, P., Yadav, S., Srivastava, A., and Shrivastava, N. (2013). Methyl Jasmonate Mediates Upregulation of Bacoside A Production in Shoot Cultures of *Bacopa monnieri*. *Biotechnology Letters*. 35(7), 1121–1125.
- Sharma, S. N., Jha, Z., Sinha, R. K., and Geda, A. K. (2015). Jasmonate-Induced Biosynthesis of Andrographolide in Andrographis paniculata. Physiologia Plantarum. 153(2), 221–229.
- Sharon, I., Birkland, A., Chang, K., El-Yaniv, R., and Yona, G. (2005). Correcting BLAST e-Values for Low-Complexity Segments. *Journal of Computational Bology*. 12(7), 980–1003.
- Shih, C. H., Chu, H., Tang, L. K., Sakamoto, W., Maekawa, M., Chu, I. K., ... Lo, C. (2008). Functional Characterization of Key Structural Genes in Rice Flavonoid Biosynthesis. *Planta*. 228(6), 1043–1054.
- Shirley, B. W. (1996). Flavonoid Biosynthesis: "New"Function for an "Old" Pathway. *Trends in Plant Science*. 1(11), 377–382.
- Shrawat, A. K., Becker, D., and Lörz, H. (2007). Agrobacterium tumefaciens -Mediated Genetic Transformation of Barley (Hordeum vulgare L.). Plant Science. 172(2), 281–290.
- Siddiqui, Z. H., Mujid, A., Mahmooduzzafar, Aslam, J., Hakeem, K. R., and Parween, T. (2013). In Vitro Production of Secondary Metabolites Using Elicitor in *Catharanthus roseus*: A Case Study. In *Crop Improvement* (pp. 401– 419). United States: Springer US.
- Sijam, K., and Lim, T. K. (1989). A Rust Disease on *Gendarussa vulgaris* Nees. Caused by Puccinia thwaitesii Berk. *Pertanika*. 12(1), 7–10.

- Silja, P. K., Gisha, G. P., and Satheeshkumar, K. (2014). Enhanced Plumbagin Accumulation in Embryogenic Cell Suspension Cultures of *Plumbago rosea* L. following Elicitation. *Plant Cell, Tissue and Organ Culture*. 119(3), 469–477.
- Silva, T. E. R., Cidade, L. C., Alvim, F. C., Cascardo, J. C. M., and Costa, M. G. C. (2009). Studies on Genetic Transformation of *Theobroma cacao* L.: Evaluation of Different Polyamines and Antibiotics on Somatic Embryogenesis and the Efficiency of uidA Gene Transfer by *Agrobacterium tumefaciens*. *Plant Cell, Tissue and Organ Culture*. 99(3), 287–298.
- Sivakumar, B. G., Yu, K. W., and Paek, K. Y. (2005). Production of Biomass and Ginsenosides from Adventitious Roots of *Panax ginseng* in Bioreactor Cultures. *Engineering in Life Sciences*. 5(4), 333–342.
- Sivanandhan, G., Arun, M., Mayavan, S., Rajesh, M., Jeyaraj, M., Dev, G. K., Manickavasagam, M., Selvaraj, N., and Ganapathi, A. (2012). Optimization of Elicitation Conditions with Methyl Jasmonate and Salicylic Acid to Improve the Productivity of Withanolides in the Adventitious Root Culture of *Withania somnifera* (L.) Dunal. *Applied Biochemistry and Biotechnology*. 168(3), 681– 696.
- Sivanandhan, G., Dev, G. K., Jeyaraj, M., Rajesh, M., Arjunan, A., Muthuselvam, M., Manickavasagam, M., Selvaraj, N., and Ganapathi, A. (2013). Increased Production of Withanolide A, Withanone, and Withaferin A in Hairy Root Cultures of *Withania somnifera* (L.) Dunal Elicited with Methyl Jasmonate and Salicylic Acid. *Plant Cell, Tissue and Organ Culture*. 114(1), 121–129.
- Sivasakthi, A., and Vijayalakshmi, M. (2014). Antibacterial Activities Of Phytochemical Extracts from the Leaves of Justicia gendarussa Burm . F. International Journal of Pharma and Bio Sciences. 5(2), 433–438.
- Slater, T. F., Sawyer, B., and Sträuli, U. (1963). Studies on Succinate-Tetrazolium Reductase Systems. III. Points of Coupling of Four Different Tetrazolium Salts. *Biochimica Et Biophysica Acta*. 77, 383–393.
- Slater, A., Scott, N. W., and Fowler, M. R. (2008). Plant Biotechnology: The Genetic Manipulation of Plants. London: Oxford University Press.
- Slavov, S., Valkov, V., Batchvarova, R., Atanassova, S., Alexandrova, M., and Atanassov, A. (2005). Chlorsulfuron Resistant Transgenic Tobacco as a Tool for Broomrape Control. *Transgenic Research*. 14(3), 273–278.

- Smith, E. F., and Townsend, C. O. (1907). A Plant-Tumor of Bacterial Origin. American Association for the Advancement of Science. 25(643), 671–673.
- So, F. V, Guthrie, N., Chambers, A. F., and Carroll, K. K. (1997). Inhibition of Proliferation of Estrogen Receptor-Positive MCF-7 Human Breast Cancer Cells by Flavonoids in the Presence and Absence of Excess Estrogen. *Cancer Letters*. 112(2), 127–133.
- Sonal, P., and Maitreyi, Z. (2011a). Pharmacognostic Study of the Root of *Justicia* gendarussa Burm. Asian Journal of Traditinal Medicines. 6(2), 1–12.
- Sonal, P., Nayana, K., Bakula, S., and Mamta, S. (2011b). Botanical Identification and Physicochemical Investigation of Leaf of Nili-Nirgundi (*Justicia* gendarussa). International Journal of Pharmaceutical Sciences Review and Research. 10(1), 116–121.
- Song, H. M., Park, G. H., Eo, H. J., Lee, J. W., Kim, M. K., Lee, J. R., Koo, J. S., and Jeong, J. B. (2015). Anti-Proliferative Effect of Naringenin through p38-Dependent Downregulation of Cyclin D1 in Human Colorectal Cancer Cells. *Biomolecules and Therapeutics*. 23(4), 339–344.
- Soule, H. D., Vazquez, J., Long, A., Albert, S., and Brennan, M. (1973). A Human Cell Line from a Pleural Effusion Derived from a Breast Carcinoma. *Journal of The National Cancer Institute*. 51(5), 1409–1416.
- Stachel, S. E., Messens, E., Montagu, M. Van, and Zambryski, P. (1985). Identification of the Signal Molecules Produced by Wounded Plant Cells that Activate T-DNA Transfer in Agrobacterium Tumefaciens. Nature. 318(6047), 624–629.
- Subarnas, A., Diantini, A., Abdulah, R., Zuhrotun, A., Yamazaki, C., Nakazawa, M., and Koyama, H. (2012). Antiproliferative Activity of Primates-Consumed Plants against MCF-7 Human Breast Cancer Cell Lines. *E3 Journal of Medical Research*. 1(4), 38–43.
- Subramaniam, S., Mahmood, M., Meon, S., and Rathinam, X. (2010a). Genetic Engineering for Tolerance to Fusarium Wilt Race 1 in *Musa sapientum* cv. Rastali (AAB) using Biolistic Gun Transformation System. *Tree and Forestry Science and Biotechnology*. 4(2), 65–75.

- Subramaniam, S., and Rathinam, X. (2010b). Emerging Factors that Influence Efficiency of T-DNA Gene Transfer into *Phalaenopsis violacea* Orchid via *Agrobacterium tumefaciens* – mediated Transformation System. *International Journal of Biology*. 2(2), 64–73.
- Subramanian, N Jothimanivannan, C., and Moorthy, K. (2012). Antimicrobial Activity and Preliminary Phytochemical Screening of *Justicia gendarussa* (Burm. F.) against Human Pathogens. *Asian Journal of Pharmaceutical and Clinical Research*. 5(3), 229–233.
- Subramanian, N., Jothimanivannan, C., Senthil Kumar, R., and Kameshwaran, S. (2013). Evaluation of Anti-anxiety Activity of *Jucticia gendarussa* Burm. *Pharmacologia*. 4(5), 404–407.
- Sudhakar, A. (2009). History of Cancer, Ancient and Modern Treatment Methods. Journal of Cancer Science and Therapy. 1(2), 1–4.
- Sui, N., Li, M., Zhao, S., Li, F., Liang, H., and Meng, Q. (2007). Overexpression of Glycerol-3-Phosphate Acyltransferase Gene Improves Chilling Tolerance in Tomato. *Planta*. 226(5), 1097–1108.
- Sultana, S., Ho, C., Namasivayam, P., and Napis, S. (2014). Genotypic Differences in Response to Hygromycin Effect on Untransformed Calli Death and Rice Germination. *Bangladesh Rice Journal*. 18(1-2), 38–43.
- Sun, W., Meng, X., Liang, L., Jiang, W., Huang, Y., He, J., Hu,Y., Almqvist, J., Gao, X., and Wang, L. (2015). Molecular and Biochemical Analysis of Chalcone Synthase from *Freesia hybrid* in Flavonoid Biosynthetic Pathway. *PLoS ONE*. 10(3), 1–18.
- Suratman, F., Huyop, F., Wagiran, A., Rahmat, Z., Ghazali, H., and Parveez, G. K. A. (2010a). Biolistic Transformation of *Citrullus vulgaris* Schrad (Watermelon). *Biotechnology*. 9(2), 119–130.
- Suratman, F., Huyop, F., Wagiran, A., Rahmat, Z., Ghazali, H., and Parveez, G. K. A. (2010b). Cotyledon with Hypocotyl Segment as an Explant for the Production of Transgenic *Citrullus vulgaris* Schrad (Watermelon) Mediated by *Agrobacterium Tumefaciens*. *Biotechnology*. 9(2), 106–118.
- Susanti, D., Sirat, H. M., Ahmad, F., Ali, R. M., Aimi, N., and Kitajima, M. (2007). Antioxidant and Cytotoxic Flavonoids from the Flowers of *Melastoma malabathricum* L. *Food Chemistry*. 103(3), 710–716.

- Sutthanut, K., Sripanidkulchai, B., Yenjai, C., and Jay, M. (2007). Simultaneous Identification and Quantitation of 11 Flavonoid Constituents in *Kaempferia parviflora* by Gas Chromatography. *Journal of Chromatography A*. 1143(1-2), 227–233.
- Taha, A. M., Wagiran, A., Ghazali, H., Huyop, F., and Parveez, G. K. A. (2009).
 Optimization and Transformation of Garden Balsam, *Impatiens balsamina*, Mediated by Microprojectile Bombardment. *Biotechnology*. 8(1), 1–12.
- Tai, D., Tian, J., Zhang, J., Song, T., and Yao, Y. (2014). A *Malus* Crabapple Chalcone Synthase Gene, McCHS, Regulates Red Petal Color and Flavonoid Biosynthesis. *PLoS ONE*. 9(10), 1–13.
- Tan, M. H., Nowak, N. J., Loor, R., Ochi, H., Sandberg, A. A., Lopez, C., Pickren, J. Berjian, R., and Douglass, H., and W., Chu, T. M. (1986). Characterization of a New Primary Human Pancreatic Tumor Line. *Cancer Investigation*. 4(1), 15– 23.
- Tate, C. R., Rhodes, L. V, Segar, H. C., Driver, J. L., Pounder, F. N., Burow, M. E., and Collins-burow, B. M. (2012). Targeting Triple-Negative Breast Cancer Cells with the Histone Deacetylase Inhibitor Panobinostat. *Breast Cancer Research*. 14(3), 79–94.
- Taurino, M., Ingrosso, I., D'amico, L., Domenico, S. De, Nicoletti, I., Corradini, D., Santino, A., and Giovinazzo, G. (2015). Jasmonates Elicit Different Sets of Stilbenes in *Vitis vinifera* cv. Negramaro Cell Cultures. *SpringerPlus*. 4, 49–60.
- Tee, C. S., and Maziah, M. (2005). Optimization of Biolistic Bombardment Parameters for *Dendrobium* Sonia 17 Calluses using GFP and GUS as the Reporter System. *Plant Cell, Tissue and Organ Culture*. 80(1), 77–89.
- Tel-Zur, N., Abbo, S., Myslabodski, D., and Mizrahi, Y. (1999). Modified CTAB Procedure for DNA Isolation from Epiphytic Cacti of the Genera Hylocereus and Selenicereus (Cactaceae). *Plant Molecular Biology Reporter*. 17(3), 249– 254.
- Thilip, C., Raju, C. S., Varutharaju, K., Aslam, A., and Shajahan, A. (2015). Improved Agrobacterium Rhizogenes - Mediated Hairy Root Culture System of Withania somnifera (L.) Dunal using Sonication and Heat Treatment. 3 Biotech. 5(6), 949–956.

- Thomas, T. D., and Yoichiro, H. (2010). In Vitro Propagation for the Conservation of a Rare Medicinal Plant Justicia gendarussa Burm. f. by Nodal Explants and Shoot Regeneration from Callus. Acta Physiologiae Plantarum. 32(5), 943–950.
- Tian, L., Wan, S.-B., Pan, Q.-H., Zheng, Y.-J., and Huang, W.-D. (2008). Plant Science Short communication A Novel Plastid Localization of Chalcone Synthase in Developing Grape Berry. *Plant Science*. 175(3), 431–436.
- Tim Cushnie, T. P., and Lamb, A. J. (2005). Antimicrobial Activity of Flavonoids. *International Journal of Antimicrobial Agents*. 26(5), 343–356.
- Torre, L. A., Bray, F., Siegel, R. L., Ferlay, J., Lortet-Tieulent, J., and Jemal, A. (2015). Global Cancer Statistics , 2012. CA: A Cancer Journal for Clinicians. 65(2), 87–108.
- Tu, Y.-C., Lian, T.-W., Yen, J.-H., Chen, Z.-T., and Wu, M.-J. (2007). Antiatherogenic Effects of Kaempferol and Rhamnocitrin. *Journal of Agricultural and Food Chemistry*. 55(24), 9969–9976.
- Tu, L.-Y., Bai, H.-H., Cai, J.-Y., and Deng, S.-P. (2016). The Mechanism of Kaempferol Induced Apoptosis and Inhibited Proliferation in Human Cervical Cancer SiHa Cell : From Macro to Nano. *Scanning*. 38(6), 644–653.
- Twentyman, P. R., and Luscombe, M. (1987). A Study of Some Variables in a Tetrazolium Dye (MTT) Based Assay for Cell Growth and Chemosensitivity. *British Journal of Cancer*. 56(3), 279–285.
- Tzfira, T., and Citovsky, V. (2006). Agrobacterium mediated Genetic Transformation of Plants: Biology and Biotechnology. Current Opinion in Biotechnology. 17(2), 147–154.
- Ucar, M. B., and Ucar, G. (2011). Characterization of Methanol Extracts from Quercus hartwissiana Wood and Bark. Chemistry of Natural Compounds. 47(5), 697–703.
- Uddin, M. R., Sinha, S., Hossain, M. A., Kaisar, M. A., Hossain, M. K., and Rashid,
 M. A. (2011). Chemical and Biological Investigations of *Justicia gendarussa* (Burm . f). *Dhaka University Journal of Pharmaceutical Sciences*. 10(1), 53–57.
- Van der Krieken, W. M., Breteler, H., Visser, M. H. M., and Mavridou, D. (1993). The Role of the Conversion of IBA into IAA on Root Regeneration in Apple: Introduction of a Test System. *Plant Cell Reports*. 12(4), 203–206.

- Van der Krol, A. R., Mur, L. A., Beld, M., Moi, J. N. M., and Stuitje, A. R. (1990). Flavonoid Genes in Petunia: Addition of a Limited Number of Gene Copies May Lead to a Suppression of Gene Expression. *The Plant Cell*. 2(4), 291–299.
- Vannozzi, A., Dry, I. B., Fasoli, M., Zenoni, S., and Lucchin, M. (2012). Genome-Wide Analysis of the Grapevine Stilbene Synthase Multigenic Family: Genomic Organization and Expression Profiles upon Biotic and Abiotic Stresses. *BMC Plant Biology*. 12(1), 130–152.
- Vázquez-Flota, F., Hernández-Domínguez, E., De Lourdes Miranda-Ham, M., and Monforte-González, M. (2009). A Differential Response to Chemical Elicitors in *Catharanthus roseus In Vitro* Cultures. *Biotechnology Letters*. 31(4), 591– 595.
- Wagiran, A., Ismail, I., Zain, C. R. C. M., and Abdullah, R. (2010). Agrobacterium tumefaciens - Mediated Transformation of the Isopentenyltransferase Gene in Japonica Rice Suspension Cell Culture. Australian Journal of Crop Science. 4(6), 421–429.
- Wang, C., and Zuo, Y. (2011). Ultrasound-Assisted Hydrolysis and Gas Chromatography – Mass Spectrometric Determination of Phenolic Compounds in Cranberry Products. *Food Chemistry*. 128(2), 562–568.
- Wang, J., Qian, J., Yao, L., and Lu, Y. (2015). Enhanced Production of Flavonoids by Methyl Jasmonate Elicitation in Cell Suspension Culture of *Hypericum perforatum. Bioresources and Bioprocessing*. 2, 5–13.
- Wang, X., Zhang, Z., Dong, X., Feng, Y., and Liu, X. (2017). Identification and Functional Characterization of Three Type III Polyketide Synthases from Aquilaria sinensis Calli. Biochemical and Biophysical Research Communications. 17, 1–12.
- Winkel-Shirley, B. (2002). Biosynthesis of Flavonoids and Effects of Stress. *Plant Biology*. 5(3), 218–223.
- Woodman, O. L., and Chan, E. C. H. (2004). Vascular and Anti-Oxidant Actions of Flavonols and Flavones. *Clinical and Experimental Pharmacology and Physiology*. 31(11), 786–790.
- Wu, C.-H., Dewir, Y. H., Hahn, E.-J., and Paek, K.-Y. (2006). Optimization of Culturing Conditions for the Production of Biomass and Phenolics from Adventitious Roots of *Echinacea angustifolia*. *Journal of Plant Biology*. 49(3), 193–199.

- Xu, Y. C., Leung, S. W. S., Yeung, D. K. Y., Hu, L. H., Chen, G. H., Che, C. M., and Man, R. Y. K. (2007). Structure – Activity Relationships of Flavonoids for Vascular Relaxation in Porcine Coronary Artery. *Phytochemistry*. 68(8), 1179– 1188.
- Xu, A., Zhan, J., and Huang, W.-D. (2015). Effects of Ultraviolet C , Methyl Jasmonate and Salicylic Acid, Alone or In Combination, on Stilbene Biosynthesis in Cell Suspension Cultures of *Vitis vinifera* L. cv. Cabernet Sauvignon. *Plant Cell, Tissue and Organ Culture*. 122(1), 197–211.
- Yadav, S. K., Katikala, S., Yellisetty, V., Kannepalle, A., Maddi, J. L. N. V., Mandapaka, M., Shanker, A. K., and Bandi, V., and Bharadwaja, K. P. (2012).
 Optimization of *Agrobacterium* Mediated Genetic Transformation of Cotyledonary Node Explants of *Vigna radiata*. *SpringerPlus*. 1, 59–66.
- Yáñez, J., Vicente, V., Alcaraz, M., Castillo, J., Benavente-Garcia, O., Canteras, M., and Teruel, J. A. L. (2004). Cytotoxicity and Antiproliferative Activities of Several Phenolic Compounds Against Three Melanocytes Cell Lines: Relationship Between Structure and Activity. *Nutrition and Cancer*. 49(2), 191–199.
- Yao, Q., Cong, L., Chang, J. L., Li, K. X., Yang, G. X., and He, G. Y. (2006). Low Copy Number Gene Transfer and Stable Expression in a Commercial Wheat Cultivar Via Particle Bombardment. *Journal of Experimen*. 57(14), 3737–3746.
- Yin, K. B. (2011). The Mesenchymal-Like Phenotype of the MDA-MB-231 Cell Line The Mesenchymal-Like Phenotype of the MDA-MB-231 Cell Line. In Breast Cancer-Focusing Tumor Microenvironment, Stem Cells and Metastasis (pp. 385–402). Croatia:InTech.
- Yong, W. T. L., Abdullah, J. O., and Mahmood, M. (2006). Optimization of *Agrobacterium* -Mediated Transformation Parameters for Melastomataceae spp. using Green Fluorescent Protein (GFP) as a Reporter. *Scientia Horticulturae*. 109(1), 78–85.
- Yong, W. T. L., Abdullah, J. O., and Mahmood, M. (2009). Agrobacterium -Mediated Transformation of Melastoma malabathricum and Tibouchina semidecandra with Sense and Antisense Dihydroflavonol-4-Reductase (DFR) Genes. Plant Cell, Tissue and Organ Culture. 96(1), 59–67.

- Yoshida, T., Konishi, M., Horinaka, M., Yasuda, T., Goda, A. E., Taniguchi, H., Yano, K., Wakada, M., and Sakai, T. (2008). Kaempferol Sensitizes Colon Cancer Cells to TRAI-Induced Apoptosis. *Biochemical and Biophysical Research Communications*. 375(1), 129–133.
- Yu, K.-W., Gao, W., Hahn, E.-J., and Paek, K.-Y. (2002). Jasmonic Acid Improves Ginsenoside Accumulation in Adventitious Root Culture of *Panax ginseng* C.
 A. Meyer. *Biochemical Engineering Journal*. 11(2-3), 211–215.
- Yu, D., Xu, F., Zeng, J., and Zhan, J. (2012). Type III Polyketide Synthases in Natural Product Biosynthesis. *IUBMB Life*. 64(4), 285–295.
- Zakaria, Z. A., Mohamed, A. M., Jamil, N. S. M., Rofiee, M. S., Hussain, M. K., Sulaiman, M. R., Teh, L. K., and Salleh, M. Z. (2011a). *In Vitro* Antiproliferative and Antioxidant Activities of the Extracts of *Muntingia calabura* Leaves. *The American Journal of Chinese Medicine*. 39(1), 183–200.
- Zakaria, Z. A., Mohamed, A. M., Jamil, N. S. M., Rofiee, M. S., Somchit, M. N., Zuraini, A., Arifah, A. K., and Sulaiman, M. R. (2011b). *In Vitro* Cytotoxic and Antioxidant Properties of the Aqueous, Chloroform and Methanol Extracts of *Dicranopteris linearis* Leaves. *African Journal of Biotechnology*. 10(2), 273– 282.
- Zakaria, Z. A., Rofiee, M. S., Mohamed, A. M., Teh, L. K., and Salleh, M. Z. (2011c). *In Vitro* Antiproliferative and Antioxidant Activities and Total Phenolic Contents of the Extracts of *Melastoma malabathricum* Leaves. *Journal* of Acupuncture and Meridian Studies. 4(4), 248–256.
- Zakaria, I., Ahmat, N., Jaafar, F. M., and Widyawaruyanti, A. (2012). Fitoterapia Flavonoids with Antiplasmodial and Cytotoxic Activities of *Macaranga triloba*. *Fitoterapia*. 83(5), 968–972.
- Zaker, A., Sykora, C., Gössnitzer, F., Abrishamchi, P., Asili, J., Mousavi, S. H., and Wawrosch, C. (2015). Effects of Some Elicitors on Tanshinone Production in Adventitious Root Cultures of *Perovskia abrotanoides* Karel. *Industrial Crops* and Products. 67, 97–102.
- Zand, R. S., Jenkins, D. J., and Diamandis, E. P. (2000). Steroid Hormone Activity of Flavonoids and Related Compounds. *Breast Cancer Research and Treatment*. 62(1), 35–49.

- Zeef, L. A. H., Christou, P., and Leech, M. J. (2000). Transformation of the Tropane Alkaloid-Producing Medicinal Plant *Hyoscyamus muticus* by Particle Bombardment. *Transgenic Research*. 9(3), 163–168.
- Zhang, Y., Chen, A. Y., Li, M., Chen, C., and Yao, Q. (2008). Ginkgo biloba Extract Kaempferol Inhibits Cell Proliferation and Induces Apoptosis in Pancreatic Cancer Cells. *Journal of Surgical Research*. 148(1), 17–23.
- Zhao, Z. Y., Gu, W., Cai, T., Tagliani, L., Hondred, D., Bond, D., Schroeder, S., Rudert, M., and Pierce, D. (2001). High throughput Genetic Transformation Mediated by Agrobacterium tumefaciens in Maize. Molecular Breeding. 8(4), 323–333.
- Zhao, J., Davis, L. C., and Verpoorte, R. (2005). Elicitor Signal Transduction Leading to Production of Plant Secondary Metabolites. *Biotechnology Advances*. 23(4), 283–333.
- Zheng, X., and Xing, F. (2009). Ethnobotanical Study on Medicinal Plants Around Mt. Yinggeling, Hainan Island, China. *Journal of Ethnopharmacology*. 124(2), 197–210.
- Ziemienowicz, A. (2014). Agrobacterium-Mediated Plant Transformation: Factors, Applications and Recent Advances. *Biocatalysis and Agricultural Biotechnology*. 3(4), 95–102.
- Zupan, J., Muth, T. R., Draper, O., and Zambryski, P. (2000). The Transfer of DNA from Agrobacterium tumefaciens into Plants: A Feast of Fundamental Insights. *The Plant Journal*. 23(1), 11–28.
- Zuraida, A. R., Rahiniza, K., Nurul Hafiza, M. R., Suri, R., Zamri, Z., and Sreeramanan, S. (2010). Factors Affecting Delivery and Transient Expression of GusA Gene in Malaysian Indica Rice MR 219 Callus Via Biolistic Gun System. *African Journal of Biotechnology*. 9(51), 8810–8818.