BIODEGRADATION OF ALKANOLAMINES IN BATCH AND PACKED-BED REACTORS USING FREE LACCASE AND SOL-GEL LACCASE

NUR ATIKAH BINTI MOHIDEM

A thesis submitted in fulfilment of the requirements for the award of the degree of Doctor of Philosophy (Chemical Engineering)

Faculty of Chemical and Energy Engineering Universiti Teknologi Malaysia

FEBRUARY 2018

This thesis is dedicated to Allah swt, prophet Muhammad saw, my beloved husband, parents and grandparents

ACKNOWLEDGEMENT

Alhamdulillah. In the name of Allah S.W.T., the Most Gracious and Merciful. Peace be upon Muhammad S.A.W., the messenger of Allah S.W.T. May Allah grant peace and honour on Him and His family (اللَّهُمُ صَلَّ عَلَى مُحَمَّدٍ وَ عَلَى آلِ مُحَمَّدٍ). Greatest thanks to Almighty Allah S.W.T for His blessing and aid throughout the research. First and foremost, my deepest love and high appreciation to my beloved parents Hj Mohidem and Hjh Safiah, my beloved grandparents, the late Hj Hassan and Hjh Aishah, my beloved husband Suhail, my beloved parents in law Hj Muhammad and Hjh Khodizah, my beloved sister Adibah, my beloved aunties Khuzaimah, Haniza, Haslina and Asiah and my relatives who have given me endless support and love at all times. Thank you so much!

Secondly, I would like to express my deepest gratitude to my supervisor, Associate Professor Dr. Hanapi Mat who has given me guidance, encouragement and confidence to conduct this research under his supervision. Thank you for your willingness to spend time with me in completing this research.

Thirdly, I would like to acknowledge my gratitude to my internal and external examiners, Professor Dr Ida Idayu Muhamad and Professor Dr Robiah Yunus for their guidance throughout my thesis correction. Your comments for thesis improvement were very much appreciated.

Not to forget, thank you to all my colleagues who have supported me and shared their precious ideas and suggestions in solving my research problems. I would like to thank Mada, Nazrah, DrRoket group, Post Graduate Support Group, AMPEN group members, Kak Lin, Kak Syura, and Ilah. I also wish to thank my Head of Department at the Manipal International University, Malaysia, Prof Dr Murthy Velury for giving me the opportunity to use the campus facilities. Thanks to Prof Dr Syed Nur Azman, Dr Yee, Dr Asrul and colleagues at Manipal International University for sharing advice for my PhD thesis writing.

Last but not least, I am grateful to Universiti Teknologi Malaysia (Fundamental Research Grant Scheme) and the Ministry of Higher Education, Malaysia for the financial assistance (MyPhd) used towards completing my studies. Again, thanks to all of you. With all your help and guidance, I managed to complete my Doctor of Philosophy degree successfully and gain optimum benefits.

ABSTRACT

Alkanolamine is commonly used in natural gas processing plant for carbon dioxide removal from natural gas. Alkanolamine may incidentally release and contaminate the surrounding soil and water due to plant operational failure or irresponsible related activities. Thus, the application of laccase for biodegradation of alkanolamine, which has not been reported so far, carried out in batch (shake flask) and continuous (packed-bed) reactors was investigated. Though, biodegradation using laccase may offer many advantages, the free laccase (FL) itself is unstable, cannot be reused and poor of thermal and storage stability. The sol-gel laccase (SGL), i.e. SOLAC04 was therefore synthesized by manipulating triethylamine (TEA) concentration (which was used as a gelating agent), laccase loading (L_L), agitation conditions (with or without sonication), and experimental procedures (onestep or two-step) towards obtaining a higher laccase catalytic activity and stability. The SOLAC04 synthesized using two-step procedure, TEA (0.1 mL), laccase loading, L_L (5 mg/mL) and without sonication had the highest laccase catalytic activity and stability as compared to other synthesized by SGL samples. This result suggested that the entrapment in silica matrix provided an additional framework for the preservation of an active laccase conformation at higher temperature and long storage duration. The biodegradation of alkanolamines: diethanolamine (DEA), ethanolamine and N-methylethanolamine was carried out in batch reactor using both FL and SGL; and optimized using response surface methodology. The results showed that the biodegradation efficiency (μ) and biodegradation rate of DEA using SOLAC04 was higher than other alkanolamines and observed to be higher compared to FL. The µ of DEA in batch reactor using FL and SOLAC04, respectively reached an optimum value at 50 °C and 40 °C. The µ of DEA increased with increasing dosage (D_s) of FL and SOLAC04. It was also revealed that the SOLAC04 are fairly stable and can be used many times. The μ of DEA remained constant at almost the same level after being reused for 5 times. The biodegradation performance and μ of DEA using FL under optimum pH of 5.8, temperature of 45.71°C, D_s of 37.14 mg, and reaction time of 42.66 minutes were 84.8 % and 0.077 mg⁻¹, while for SGL obtained under the optimum pH of 5, temperature of 41°C, D_s of 34.01 mg, and reaction time of 57.59 minutes were 66 % and 1.11 mg⁻¹, respectively. The μ of DEA in packed-bed reactor using SOLAC04 was optimum at pH 6, 250 mL/h and 500 ppm. The μ of DEA increased with increasing D_s of SOLAC04. These experimental results demonstrated the advantages gained from entrapment of laccase in silica matrix and the biodegradation superiority of the SGL over FL for the removal of alkanolamines. Thus, the potential of laccase especially SGL for biodegradation of the alkanolamine was finally demonstrated.

ABSTRAK

Alkanolamina lazimya digunakan dalam loji pemprosesan gas asli untuk penyingkiran karbon dioksida daripada gas asli. Alkanolamina secara tidak sengaja boleh terbebas dan mencemarkan tanah dan air yang berada di persekitaran yang berpunca daripada kegagalan operasi atau berkaitan kecuaian aktiviti. Oleh yang demikian, penggunaan lakase untuk biodegradasi, yang mana belum dilaporkan setakat ini telah dilaksanakan dalam reaktor kelompok (kelalang goncang) dan reaktor berterusan (turus terpadat). Walaupun biodegradasi menggunakan lakase boleh memberikan banyak manfaat, lakase bebas (FL) sendiri tidak stabil, tidak dapat diguna semula, dan kurang kestabilan terma dan penyimpanan. Dengan sebab itu, lakase sol-gel (SGL) seperti SOLAC04 telah disintesis dengan memanipulasi kepekatan trietilamina (TEA) (yang mana digunakan sebagai bahan gelatin), muatan lakase (LL), keadaan pengadukan (bersama atau tanpa sonikasi), dan tatacara eksperimen (satu langkah atau dua langkah) ke arah mendapatkan peningkatan aktiviti bermangkin dan kestabilan. SOLAC04 disintesis menggunakan tatacara dua langkah, TEA (0.1 mL), muatan lakase, LL (5 mg/mL) dan tanpa sonikasi mempunyai lakase aktiviti bermangkin dan kestabilan paling tinggi jika dibandingkan dengan yang disintesis oleh sampel SGL yang lain. Keputusan ini menunjukkan bahawa pemerangkapan dalam matriks silika memberikan rangka tambahan untuk pemeliharaan bentuk lakase aktif pada suhu yang lebih tinggi dan tempoh penyimpanan yang panjang. Biodegradasi alkanolamina: dietanolamina (DEA), etanolamina dan N-metiletanolamina telah dijalankan dalam reaktor kelompok menggunakan kedua-dua FL dan SGL, dan dioptimumkan menggunakan kaedah permukaan tindak balas. Keputusan menunjukkan bahawa kecekapan biodegradasi (µ) dan kadar biodegradasi untuk DEA menggunakan SOLAC04 lebih tinggi berbanding dengan alkanolamina lain, dan juga lebih tinggi berbanding dengan FL. Nilai µ untuk DEA dalam reaktor kelompok menggunakan FL dan SOLAC04 masing-masingnya mencapai optimum pada 50 °C dan 40 °C. Manakala nilai µ untuk DEA meningkat dengan peningkatan muatan (D_s) untuk FL dan SOLAC04. Ianya telah ditunjukkan bahawa SOLAC04 adalah sangat stabil dan boleh digunakan berulang kali, iaitu nilai µ untuk DEA kekal malar pada aras hampir sama selepas µ untuk DEA digunakan sebanyak 5 kali. Perolehan biodegradasi dan nilai menggunakan FL pada pH optimum 5.8, suhu 45.71°C, D_s 37.14 mg, dan masa tindak balas 42.66 minit masing-masing adalah 84.8 % dan 0.077 mg⁻¹, sementara untuk SGL yang telah didapati optimum pada pH 5, suhu pada 41°C, D_s 34.01 mg, dan masa tindak balas 57.59 minit masing-masing adalah 66 % dan 1.11 mg⁻¹. Nilai μ untuk DEA dalam reaktor turus terpadat menggunakan SOLAC04 memiliki tahap optimum pada pH 6, 250 mL/h dan 500 ppm. Nilai µ untuk DEA meningkat dengan peningkatan D_s untuk SOLAC04. Keputusan eksperimen ini telah membuktikan kemanfaatan yang diperolehi daripada pemerangkapan lakase dalam matriks silika dan keunggulan SGL berbanding FL bagi penyingkiran alkanolamina. Oleh yang demikian, akhirnya potensi lakase terutama SGL untuk biodegradasi alkanolamina telah akhirnya dibuktikan.

TABLE OF CONTENTS

CHAPTE	R	TITLE	PAGE
	DEC	LARATION	ii
	DED	ICATION	iii
	ACK	NOWLEDGEMENT	iv
	ABS	ГКАСТ	vi
	ABS	ГКАК	vii
	TAB	LE OF CONTENTS	viii
	LIST	OF TABLES	xii
	LIST	OF FIGURES	xiv
	LIST	OF ABBREVIATIONS	xviii
	LIST	OF SYMBOLS	xix
	LIST	OF APPENDICES	xxi
1	INTF	RODUCTION	1
	1.1	Research Background	1
	1.2	Problem Statement	3
	1.3	Objectives of Research	4
	1.4	Research Scope	5
	1.5	Novelty Statements	6
	1.6	Thesis Outline	7
	1.7	Summary	7
2	LITE	CRATURE REVIEW	8
	2.1	Introduction	8
	2.2	Biodegradation Using Enzymes	8

	2.2.1 Introduction		8
	2.2.2 Catalytic reaction models		10
	2.2.3 Enzymatic degradation by l	accase	12
2.3	Enzyme Immobilisation and Stabili	zation	14
	2.3.1 Enzyme immobilisation		14
	2.3.2 Enzyme immobilisation thro sol-gel silica	ough entrapment in	21
	2.3.3 Characterization of immobi	lised enzyme	26
	2.3.3.1 Physical properties	3	26
	2.3.3.2 Chemical propertie	28	27
	2.3.3.3 Biological property	ies	28
2.4	Biodegradation of alkanolamines		35
	2.4.1 Introduction to alkanolamin	ies	35
	2.4.2 Biodegradation of alkanola	mine	
	using oxidoreductase		35
	2.4.3 Bioreactors for Biodegradat	tion	40
2.5	Optimisation		44
	2.5.1 Introduction to optimisation		44
	2.5.2 Optimisation by Response S	Surface	
	Methodology (RSM)		44
	2.5.3 Box-Behken Design		45
	2.5.4 Statistical design of experim	nent in	
	biodegradation process		46
2.6	Summary		48
			40
	FERIALS AND METHODS		49
3.1	Introduction		49
3.2	Materials		49
3.3	Laccase Entrapment in Sol-ge Characterisation	l Silica and Its	51
	3.3.1 Entrapment procedures		51
	3.3.2 Determination of laccase lea	aching	53
	3.3.3 Laccase catalytic activity ass	ays	55

3

	3.3.4	Characte	risations	of sol-	gel laccases (SGLs)	55
3.4	Biode	gradation	of Alkan	olamin	e		56
	3.4.1	Biodegra	dation in	in bate	ch reactor		56
	3.4.2	Biodegrad	lation in	packed	-bed reactor ((PBR)	58
	3.4.3	Biodegrad	lation op	imisati	on using Res	ponse Surfac	e
		Methodol	ogy (RSN	/I)			60
	3.4.4	Alkanola	mine dete	erminat	tion		64
3.5	Summ	ary					66
RES	ULTS A	ND DISC	CUSSION	1			67
4.1	Introd	uction					67
4.2		cterisation el Laccase	•	ic Act	ivity, and St	ability of	67
	4.2.1	Characte	risation o	of Sol-C	Gel Laccases		68
	4.2.2	Catalytic	activity	of sol–	gel laccase		76
		4.2.2.1	Effect against activity	sol-g	synthesis c el laccase	conditions catalytic	76
		4.2.2.2	FTIR a activity	•	s of laccase	catalytic	80
	4.2.3	Stability	of sol-ge	el lacca	se (SGL)		86
		4.2.3.1	Effect of	of pH			86
		4.2.3.2	Effect of	of temp	erature		88
		4.2.3.3	Effect of	of stora	ge duration		90
4.3	Biode	gradation	of Alkan	olamin	e in Batch Re	actor	94
	4.3.1	Effect of	time cou	rse			94
	4.3.2	Effect of	рН				99
	4.3.3	Effect of	temperat	ure			100
	4.3.4	Effect of	substrate	e (DEA) concentration	on	101
	4.3.5	Effect of	enzyme	loading	gs (Ds)		103
	4.3.6	Reusabil	ity				104
4.4	-	isation dology (R	using SM)	the	Response	Surface	106

4

	4.4.1 Optimisation of DEA biodegradation by free laccase (FL) using RSM	106
	4.4.2 Optimisation of DEA biodegradation by SOLAC04 using RSM	122
4.5	Biodegradation of Alkaolamine in Packed-Bed Reactor (PBR)	139
	4.5.1 Introduction	139
	4.5.2 Effect of reaction time	139
	4.5.3 Effect of pH	141
	4.5.4 Effect of flowrate	143
	4.5.5 Effect of concentration	144
	4.5.6 Effect of enzyme dosage (Ds)	146
4.6	Summary	147
CON	CLUSION AND RECOMMENDATIONS	148
5.1	Introduction	148
5.2	Summary of Reseach Findings	148
5.3	Recommendation for Future Research	150
FERENC	ES	152

REFERENCES
Appendices A-B

5

166-168

xi

LIST OF TABLES

TABLE	NO.
-------	-----

TITLE

PAGE

2.1	Advantages and drawbacks of immobilization techniques	18
2.2	Summary of FTIR spectra of functional group associated with free laccase and SOLAC04	28
2.3	Standard substrates and supports for laccase immobilization	30
2.4	Summary of the procedures of enzyme immobilization and important findings	34
2.5	Biodegradation of alkanolamine using oxidoreductase	39
2.6	Advantages and disadvantages of bioreactors	40
3.1	Source and purpose of chemicals used in this study	50
3.2	Synthesis condition of SGLs	52
3.3	The level of independent variable chosen for the Box- Behnken design	61
3.4	Design matrix in the Box-Behnken model for the biodegradation of DEA using FL	62
3.5	Design matrix in the Box-Behnken model for the biodegradation of alkanolamine using SGL (SOLAC04)	63
4.1	Surface area, pore volume, and pore diameter of sol-gel laccases synthesised without sonication: (a) one-step (SOLAC01-SOLAC03) and two-step (SOLAC04- SOLAC06) procedures.	74
4.2	Surface area, pore volume, and pore diameter of sol-gel laccases synthesis (sonication). (a) One-step (SOLAC07-SOLAC09) and two-step (SOLAC10-SOLAC12).	74
4.3	Biodegradation rates of EA, DEA, and MEA	96
4.4	Experimental conditions in the Box-Behnken design and the corresponding experimental responses	107

4.5	Sequential model sum of squares	108
4.6	Observed and predicted values of the response	109
4.7	Estimated regression coefficient for response	111
4.8	Analysis of variance for DEA biodegradation performance	
4.9	Experimental conditions in the Box-Behnken design and the corresponding experimental responses	124
4.10	Sequential model sum of squares	125
4.11	Observed and predicted values of the response	126
4.12	Estimated regression coefficient for response	128
4.13	Analysis of variance for biodegradation performance of DEA.	129
4.14	Comparison between optimisation of DEA biodegradation performance by FL and SOLAC04 using RSM approach	139

LIST OF FIGURES

FIGURE NO.

TITLE

PAGE

2.1	Reaction coordinate.	10
2.2	Schematic diagram of the lock-and-key model of enzyme catalysis.	10
2.3	A simplified reaction mechanism of laccase oxidation of suitable substrate (Rodríguez-Delgado <i>et al.</i> , 2015).	14
2.4	Principle of enzyme immobilisation techniques (Brady and Jordaan, 2009; Brena <i>et al.</i> , 2013)	17
2.5	Enzyme laccase before and after entrapment (Sassolas <i>et al.</i> , 2013)	21
2.6	Schematic representation of the packed bed bioreactor system for phenol removal using laccase immobilized on alginate beads: (a) untreated sample (phenol model solution); (b) peristaltic pump; (c) column bioreactor packed with immobilized beads; (d) treated sample (Niladevi and Prema, 2008).	43
3.1	Standard curve for laccase determination using biuret assay	54
3.2	Schematic presentation of the PBR system for alkanolamine Biodegradation using SGL: (a) untreated sample (substrate); (b) peristaltic pump; (c) column reactor packed with SGL and silica particles; and (d) feed/substrate sample.	59
3.3	A standard chromatogram of (a) DEA, (b) EA and (c) MEA	65
3.5	Standard calibration curve of alkanolamine: EA; DEA; and MEA	65
4.1	Microscopic structure of sol-gel laccases synthesised without sonication observed by SEM: (a) one-step (SOLAC01-SOLAC03) and two-step (SOLAC04-	
	SOLAC06) procedures.	71

4.2	Microscopic structure of sol-gel laccases synthesised with sonication observed by SEM: (a) one-step (SOLAC07- SOLAC09) and two-step (SOLAC10-SOLAC12) procedures.	72
4.3	The effect of L_L on the laccase catalytic activity of SOLAC04. Experimental condition: Temperature = 27 °C, substrate = 1mM, pH 5 of 2,6-DMP.	79
4.4	FTIR spectra of the FL and SOLAC04.	82
4.5	FTIR spectra of SOLAC04 having different L _L values.	84
4.6	Effect of pH on the laccase catalytic activity of the free laccase and SOLAC04. Experimental condition: Temperature = 27 °C, substrate = 1mM, pH 5 of 2,6-DMP.; L_L : 5 mg/mL.	87
4.7	Effect of temperature on the laccase catalytic activity of the free laccase and SOLAC04. Experimental condition: Substrate = 1mM, pH 5 of 2,6-DMP.; L_L g: 5 mg/mL.	89
4.8	Effect of storage duration (27°C) on the laccase catalytic activity of the free laccase and SOLAC04. Experimental condition: Substrate = 1mM, pH 5 of 2,6-DMP.; LL: 5 mg/mL; storage time; 34 days.	91
4.9	Fourier transform infrared (FTIR) spectra of the free laccase and SOLAC04 for 34 days of storage duration at 27 °C.	93
4.10	Effect of time course of DEA, EA and MEA on biodegradation efficiency catalysed by FL. [Alkanolamine] = 500 ppm, pH = 5; sample dosage (D_s) = 20 mg; laccase mass (L_m) = 6 mg; reaction temperature (T) = 27 °C; and reaction time (t) = 1 h.	95
4.11	Effect of time course of DEA, EA and MEA on the biodegradation efficiency catalysed by SOLAC04. Experimental conditions: [Alkanolamine] = 500 ppm, pH = 5; sample dosage (D_s) = 20 mg; laccase mass (L_m) = 0.35 mg; reaction temperature (T) = 27 °C; and reaction time (t) = 1 h.	97
4.12	Effect of pH on the biodegradation efficiency catalysed by FL and SOLAC04. Experimental conditions: $[DEA] = 500$ ppm; sample dosage $(D_s) = 20$ mg; laccase mass (L_m) of FL= 6 mg, SGL= 0.35 mg; reaction temperature $(T) = 27$ °C; and reaction time $(t) = 1$ h.	99
4.14	Effect of concentration on biodegradation efficiency catalysed by FL and SOLAC04. Experimental conditions: pH = 5; sample dosage (D _s) = 20 mg; laccase mass (L _m) of	

	FL= 6 mg, SGL= 0.35 mg; reaction temperature (T) = 27 °C and reaction time (t) = 1 h.	102
4.15	Effect of D_s on biodegradation efficiency catalysed by FL and SOLAC04. Experimental conditions: [DEA] = 500 ppm; pH = 5; reaction temperature = 27 °C; and reaction time (t) = 1 h.	104
4.16	Effect of reusability of SOLAC04 on biodegradation efficiency catalysed by SOLAC04. Experimental conditions: [DEA] = 500 ppm; pH = 5; sample dosage (D _s) = 20 mg; laccase mass (L _m) of FL= 6 mg, SGL= 0.35 mg; reaction temperature (T) = 27 °C; and reaction time (t) = 1 h.	105
4.17	Predicted versus actual DEA biodegradation performance of DEA.	113
4.18	3D response surface plot showing the effect of temperature and pH on the biodegradation performance of DEA (%) at D_s of laccase of 40 mg and reaction time of 37.5 minutes.	115
4.19	3D response surface plot showing the effect of D_s and pH on the biodegradation performance of DEA at 55 °C and reaction time of 37.5 minutes.	116
4.20	3D response surface plot showing the effect of reaction time and pH on the biodegradation performance of DEA at 55 °C and 40 mg of $D_{s.}$	118
4.21	3D response surface plot showing the effect of D_s and temperature on the biodegradation performance of DEA at pH 5 and 37.5 of reaction time.	119
4.22	3D response surface plot showing the effect of reaction time and temperature on the biodegradation performance of DEA at pH 5 and 40 mg of $D_{s.}$	120
4.23	3D response surface plot showing the effect of reaction time and Ds on the biodegradation performance of DEA at pH 5 and 55 °C.	121
4.24	Predicted versus actual DEA biodegradation performance.	130
4.25	3D response surface plot showing the effect of temperature and pH on the biodegradation performance of DEA at D_s of laccase of 40 mg and reaction time of 37.5 minutes.	132
4.26	3D response surface plot showing the effect of SOLAC04 D_s and pH on the biodegradation performance of DEA (%) at 55 °C and reaction time of 37.5 minutes.	133

4.27	3D response surface plot showing the effect of reaction time and pH on the biodegradation performance of DEA at 55 °C and 37.5 minutes of reaction time.	134
4.28	3D response surface plot showing the effect of reaction time and pH on the biodegradation performance of DEA at 55 °C and 40 mg of SOLAC04 D _s .	135
4.29	3D response surface plot showing the effect of reaction time and temperature on the biodegradation performance of DEA at pH 5 and D_s of 40 mg.	136
4.30	3D response surface plot showing the effect of reaction time and D_s on the biodegradation performance of DEA at pH 5 and 55 °C.	137
4.31	Effect of DEA, EA and MEA reactions on biodegradation efficiency of DEA degradation in PBR catalysed by SOLAC04. Experimental conditions: [DEA] = 500 ppm; sample dosage $D_s = 20$ mg; pH = 5; reaction temperature (T) = 27 °C; and flowrate (Q) = 100 mL/h.	140
4.32	Effect of pH on biodegradation efficiency of DEA degradation in PBR catalysed by SOLAC04. Experimental conditions: [DEA] = 500 ppm; sample dosage Ds = 20 mg; reaction temperature (T) = 27 °C; flowrate (Q) = 100 mL/h; and reaction time (t) = 1 h.	142
4.33	Effect of flowrate on biodegradation efficiency of DEA degradation in PBR catalysed by SOLAC04. Experimental conditions: [DEA] = 500 ppm; sample dosage $D_s = 20$ mg; pH = 5; reaction temperature (T) = 27 °C; reaction time (t) = 1 h.	143
4.34	Effect of concentration on biodegradation efficiency of DEA degradation in PBR catalysed by SOLAC04. Experimental conditions: Sample dosage $D_s = 20$ mg; pH = 5; reaction temperature (T) = 27 °C; flowrate (Q) = 100 mL/h; and reaction time (t) = 1 h.	145
4.35	Effect of D_s on biodegradation efficiency of DEA degradation in PBR catalysed by SOLAC04. Experimental conditions: [DEA] = 500 ppm; pH = 5; reaction temperature (T) = 27 °C; flowrate (Q) = 100 mL/h; and reaction time (t) = 1 h.	146

LIST OF ABBREVIATIONS

2,6-DMP	-	2,6- dimethoxyphenol
BET	-	Brunauer, Emmett and Teller
BJH	-	Barrett–Joyner–Halenda
CO_2	-	Carbon dioxide
DEA	-	Diethanolamine
DMAMP	-	2-dimethylamino-2-methyl-1-propanol
EA	-	Ethanolamine
FL	-	Free Laccase
FTIR	-	Fourier Transform Infrared Spectrum
H_2S	-	Hydrogen sulfide
HC1	-	Hydrochloric acid
K_2HPO_4	-	di-potassium hydrogen phosphate
KBr	-	Potassium bromine
KH_2PO_4	-	Potassium dihydrogen phosphate
MEA	-	N-methyl ethanolamine
PEG	-	Polyethylene gylcol
PVA	-	Poly (vinyl alcohol)
RSM	-	Response Surface Methodology
SEM	-	Scanning Electron Microscopy
SGL	-	Sol-Gel Laccase
TEA	-	Triethylamine
TEOS	-	Tetraethoxysilane
TMOS	-	Tetramethoxysilane
UV/VIS	-	Ultraviolet/visible light
ANOVA	-	Analysis of Variance

LIST OF SYMBOLS

%	-	Percentage
А	-	Absorbance
Ds	-	Dosage
ka	-	Linear coefficients
k _{aa}	-	Quadratic coefficients
k _{ab}	-	Quadratic coefficients
k _{ac}	-	Quadratic coefficients
k _{ad}	-	Quadratic coefficients
k _b	-	Linear coefficients
k _{bb}	-	Quadratic coefficients
k _{bc}	-	Quadratic coefficients
k _{bd}	-	Quadratic coefficients
kc	-	Linear coefficients
k _{cc}	-	Quadratic coefficients
k _{cd}	-	Quadratic coefficients
k _d	-	Linear coefficients
k _{dd}	-	Quadratic coefficients
k_o	-	Constants
LL	-	Laccase Loadings
L_m	-	Actual Laccase
mg	-	milligram
mL	-	millimetre
°C	-	Degree Celsius
pН	-	Potential of Hydrogen
ppm	-	Part per million
SOLAC	-	Sol-gel laccase

- T Temperature
- v Volume
- *x* Multiple
- *Y* Process response or output
- *Y'* Biodegradation performance of alkanolamine

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
А	Synthesis and Characterisation	166
В	Biodegradation of Alkanolamine in Batch Reactor	168

CHAPTER 1

INTRODUCTION

1.1.1 Research Background

Alkanolamine solutions have been extensively studied during the last 25 years because of their industrial importance in natural gas processing plants, synthetic ammonia plants, fossil-fuel-fired power plants, chemical synthesis, petrochemical, cosmetic formulations, agriculture, and pharmaceutical (Chen *et al.*, 2011; Hansen *et al.*, 2010; Xi *et al.*, 2012; Zurita *et al.*, 2005). Alkanolamines are also used in industrial processing plants for acid gas impurities removal such as carbon dioxide (CO₂), hydrogen sulfide (H₂S), and sulfur dioxide (SO₂) from gas streams (Mundhwa and Henni, 2007).

The main problems associated with alkanolamine gas treatment plants is corrosion that occurs on the cross exchanger rich side, rich-amine piping after cross exchanger, still, and reboiler, where free acid gas and higher temperature are the main driving forces for corrosion (Vaidya and Kenig, 2009). This may result in loss of alkanolamine solution to surrounding soil and groundwater due to the rich-amine piping leakage. Other alkanolamine wastes include from the spillage and spent alkanolamine solution generated during plant shut-down. It was reported that alkanolamine, such as N-methylethanolamine (MEA), diethanolamine (DEA), and triethanolamine, are compounds with potential acute, sub-chronic, and chronic toxicity effects towards aquatic species (Libralato *et al.*, 2010). One of the approaches that can be used to remediate if spillage and leakage of alkanolamine solutions into soil and underground water system or to dispose of the spent alkanolamine solutions is by using biological method (Grace Liu *et al.*, 2011). Enzyme-assisted reaction was investigated in recent years for its advantages in easy operation, high efficiency, economic, versatile, and environmentally sound solution. It can simulate natural processes and result in the complete destruction of hazardous compounds into innocuous products. The use of bioremediation to remove pollutants is typically less expensive than the equivalent physical/chemical methods (Fernández-Fernández *et al.*, 2013).

Laccases enzymes have great biotechnological potential due to their capabilities and potential in various applications, such as in juice manufacturing (Berka *et al.*, 1998), chemical synthesis, and wine stabilization (Fernández-Fernández *et al.*, 2013), dye decolonization (Bayramoğlu *et al.*, 2003; Champagne and Ramsay, 2010), bioleaching (Widsten and Kandelbauer, 2008), enzymatic fuel cells (Cardoso *et al.*, 2013), wastewater treatment (Georgieva *et al.*, 2008), biosensors in nanobiotechnology (Rodríguez Couto and Toca Herrera, 2006), and biopulping have recently attracted considerable research interests.

All laccase applications mentioned above, of especially on an industrial scale, have increased the demand for high amounts of isolated or immobilized laccase production (Birhanli *et al.*, 2013). Meanwhile biodegradation using enzyme has many advantages, the isolated enzymes themselves are unstable, difficult to handle under non-conventional conditions, easily denatured in non-conventional solvents, inhibited by substrates and products, and it can only work well on natural substrates and under physiological conditions. However, it can be improved by enzyme immobilization (Birhanli *et al.*, 2013).

Currently, the stability increase of laccase catalytic activity could be achieved through immobilisation, which has been investigated by researchers ranging from methods of adsorption (Rekuć *et al.*, 2010; Salis *et al.*, 2009) and covalent attachment (Bayramoğlu and Arıca, 2008; Quan and Shin, 2004; Rekuć *et al.*, 2009)

on various supports, cross-linking (Jordaan *et al.*, 2009; Matijošytė *et al.*, 2010; Rasera *et al.*, 2009), and encapsulation in reverse micelles and emulsions (Michizoe *et al.*, 2001; Okazaki *et al.*, 2002), organic polymers such as polyallylamine (Rasera *et al.*, 2009) and inorganic polymers such as sol-gel silicas (Mansor *et al.*, 2016; Mohidem and Mat, 2012b; Nogala *et al.*, 2010).

The common sol-gel materials used in biomolecules encapsulation are silica, aluminum, titanium, zirconium, tin, vanadium, and molybdenum oxides (Debecker *et al.*, 2013; Owens *et al.*, 2016). Among them, the use of silica as precursors, such as tetraethoxysilane (TEOS) and tetramethoxysilane (TMOS), in synthesizing sol-gel could offer numerous advantages, for example improving the mechanical strength and stability. It does not swell in aqueous or organic solvent, thus preventing leaching of encapsulated biomolecules. Silica is not a food source for microorganisms and it is biologically inert. Besides, the organically modified silica, such as TEOS and TMOS, offer tolerable hydrophilic, hydrophobic, and H-bonding capacities, as well as electrochemical activities and display good porosities (Alvarez *et al.*, 2007; Owens *et al.*, 2016; Vera-Avila *et al.*, 2004).

1.2 Problem Statement

The present research investigates the use of potential laccase as an enzyme to degrade alkanolamine solutions such as diethanolamine (DEA), ethanolamine (EA) and N-methylethanolamine (MEA) in batch (shake flask) and continuous (packedbed) reactors. It was reported that alkanolamines, such as diethanolamine (DEA), ethanolamine (EA) and N-methylethanolamine (MEA) are compounds with potential acute, sub-chronic, and chronic toxicity effects towards aquatic species. Generally, alkanolamine is widely used in natural gas processing plant for carbon dioxide removal from natural gas. However, it may incidentally release and contaminate the surrounding soil and water due to the operational plant failure or amine piping leakage. (Libralato *et al.*, 2010). It has been reported that other oxidoreductase enzyme such as ethanolamine oxidase and myeloperoxidase appear to be specific for the oxidative deamination of ethanolamine (Lepaumier *et al.*, 2011).

Although biodegradation by using enzyme has many advantages, the free enzyme themselves are unstable, cannot be reused, poor of thermal and storage stability. However, the catalytic activity, stability, and reusability can be improved by enzyme immobilisation (Brena *et al.*, 2013; Sassolas *et al.*, 2013). One major substantial advantage of immobilization is reusability which drastically cut cost of laccase in treatment plant. Nevertheless, the experimental conditions of the sol-gel technique of immobilisation still require some optimisation to preserve the conformation of the most delicate biomolecule during immobilisation and to recover a high fraction of their catalytic activity (Owens *et al.*, 2016).

Consequently, in the present doctoral research, the sol-gel laccase (SGL) was synthesized by manipulating TEA concentration as a gelating agent, laccase loading (L_L), agitation conditions (with or without ultrasonic), and experimental procedures (one-step or two-step) in order to acquire a higher laccase catalytic activity and stability. TEA is widely used as chelating agents in organic synthesis such as the formation of cobalt ions based catalyst with TEA (Xu *et al.*, 2012) and new magnetic bromochromate hybrid nanomaterial with TEA surface modified iron oxide nanoparticles (Rahimi *et al.*, 2014).

1.3 Objectives of Research

The objectives of current research include:

- a) To synthesize and characterize the sol-gel laccase (SGL);
- b) To evaluate the biodegradation performance and biodegradation efficiency of alkanolamines using free laccase (FL) and SGL in batch reactors;

- c) To optimise the biodegradation performance of alkanolamines using FL and SGL in batch reactors using Response Surface methodology (RSM);
- d) To evaluate the biodegradation performance and biodegradation efficiency of alkanolamines using SGL in PBR.

1.4 Research Scope

The research scope is divided into three main parts:

a) Synthesis and characterisation of the SGL.

The SGL was synthesized by manipulating TEA concentration as a gelating agent, laccase loading (L_L), agitation conditions (with or without ultrasonic), and experimental procedures (one-step or two-step). The characterisation of SGL included particle morphology, Brunauer, Emmett and Teller (BET) surface areas, functional group, effect of pH, effect of temperature and effect of storage durations.

b) To evaluate the biodegradation process (i.e. biodegradation performance and biodegradation efficiency) of alkanolamines using FL and SGL in batch reactors;

In order to evaluate the biodegradation efficiency and biodegradation performance of alkanolamines in a batch reactor, several parameters were explored. These include the effect of reaction time, effect of pH, effect of temperature, effect of substrate concentration, effect of FL and SGL dosage (D_s) and reusability.

c) To optimise the biodegradation performance of alkanolamines using FL and SGL in batch reactors using Response Surface methodology (RSM);

In order to evaluate the biodegradation performance of alkanolamine by FL and SGL and its optimisation by using RSM, the Box–Behnken design was selected. The four parameters such as pH, temperature, reaction time, and FL or SGL dosage (D_s) were chosen based on the produced results.

d) To evaluate the biodegradation performance and biodegradation efficiency of alkanolamines using SGL in PBR.

In order to evaluate the biodegradation performance and biodegradation efficiency of alkanolamines in a PBR, several parameters were investigated. These include the effect of reaction time, effect of pH, effect of substrate concentration, effect of SGL dosage (D_s) and effect of flowrate.

1.5 Novelty Statement

The novelties of the present research are:

- (a) A synthesis method of sol-gel laccase which resulted in high laccase catalytic activity and stability.
- (b) The application of sol-gel laccase for removal of alkanolamines pollutants which is commonly found in the contaminated water from the natural gas processing plant. The alkanolamines are commonly used in removal process of carbon dioxide from natural gas.

1.6 Thesis Outline

This thesis consists of five chapters. Chapter 1 introduces the problem statements and clarified the objectives and scope of research. Chapter 2 provides a review of past research related to enzyme immobilisation and biodegradation of alkanolamines. The materials and methods used in the present research are presented in Chapter 3 while the results and discussion of this research are described in Chapter 4. Conclusion and recommendations of research are presented in Chapter 5.

1.7 Summary

The enzymes immobilisation in the sol-gel matrix have been reported by many researchers to improve their functional characteristics to a large extent. Due to the need for enhancing the catalytic activity and stability of laccase enzyme for wide industrial applications, the synthesis of immobilised laccase in sol-gel silica matrix and its applications was studied. In present research, the SGL was obtained by simple precipitation of TEOS solution by using TEA as a gelating agent in order to degrade alkanolamine in a batch reactor and PBR. Thus the research background, objectives, scope and thesis outline were clearly elaborated in this chapter.

REFERENCES

- Alvarez, G. S., Desimone, M. F., and Diaz, L. E. (2007). Immobilization of Bacteria in Silica Matrices Using Citric Acid in the Sol–Gel Process. *Applied Microbiology and Biotechnology*, 73(5): 1059-1064.
- Avnir, D., Coradin, T., Lev, O., and Livage, J. (2006). Recent Bio-Applications of Sol-Gel Materials. *Journal of Materials Chemistry*, 16(11): 1013-1030.
- Bai, X., Gu, H., Chen, W., Shi, H., Yang, B., Huang, X., and Zhang, Q. (2014). Immobilized Laccase on Activated Poly(Vinyl Alcohol) Microspheres for Enzyme Thermistor Application. *Applied Biochemistry and Biotechnology*, 173(5): 1097-1107.
- Balusu, R., Paduru, R. R., Kuravi, S. K., Seenayya, G., and Reddy, G. (2005). Optimization of Critical Medium Components Using Response Surface Methodology for Ethanol Production from Cellulosic Biomass by Clostridium Thermocellum Ss19. *Process Biochemistry*, 40(9): 3025-3030.
- Basak, B., Bhunia, B., Dutta, S., and Dey, A. (2013). Enhanced Biodegradation of 4-Chlorophenol by Candida Tropicalis Phb5 Via Optimization of Physicochemical Parameters Using Taguchi Orthogonal Array Approach. *International Biodeterioration & Biodegradation*, 78: 17-23.
- Bayramoğlu, G., Altınok, H., Bulut, A., Denizli, A., and Arıca, M. Y. (2003). Preparation and Application of Spacer-Arm-Attached Poly(Hydroxyethyl Methacrylate-Co-Glycidyl Methacrylate) Films for Urease Immobilisation. *Reactive and Functional Polymers*, 56(2): 111-121.
- Bayramoğlu, G., and Arıca, M. Y. (2008). Enzymatic Removal of Phenol and P-Chlorophenol in Enzyme Reactor: Horseradish Peroxidase Immobilized on Magnetic Beads. *Journal of Hazardous Materials*, 156(1–3): 148-155.
- Bayramoglu, G., Yilmaz, M., and Yakup Arica, M. (2010). Preparation and Characterization of Epoxy-Functionalized Magnetic Chitosan Beads: Laccase

Immobilized for Degradation of Reactive Dyes. *Bioprocess and Biosystems Engineering*, 33(4): 439-448.

- Benzina, O., Frikha, F., Zouari-Mechichi, H., Woodward, S., Belbahri, L., Mnif, E., and Mechichi, T. (2012). Enhanced Decolourization of the Azo Dye Sirius Rose Bb by Laccase–Hbt System. *3 Biotech*, 2(2): 149-157.
- Berg, J. M., Tymoczko, J. L., and Stryer, L. (2002). Biochemistry.
- Berka, R. M., Brown, S. H., Xu, F., Schneider, P., Ll, K. M. O., and Aaslyng, D. A. (1998). Purified Myceliophthora Laccases and Nucleic Acids Encoding Same: Google Patents.
- Beyer, R. E. (1983). A Rapid Biuret Assay for Protein of Whole Fatty Tissues. Analytical Biochemistry, 129(2): 483-485.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., and Escaleira, L. A. (2008). Response Surface Methodology (Rsm) as a Tool for Optimization in Analytical Chemistry. *Talanta*, 76(5): 965-977.
- Birhanli, E., Erdogan, S., Yesilada, O., and Onal, Y. (2013). Laccase Production by Newly Isolated White Rot Fungus Funalia Trogii: Effect of Immobilization Matrix on Laccase Production. *Biochemical Engineering Journal*, 71: 134-139.
- Bradford, M. M. (1976). A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical Biochemistry*, 72(1): 248-254.
- Brady, D., and Jordaan, J. (2009). Advances in Enzyme Immobilisation. Biotechnology Letters, 31(11): 1639.
- Brakstad, O. G., Booth, A., Eide-Haugmo, I., Skjæran, J. A., Sørheim, K. R., Bonaunet, K., Vang, S.-H., and Da Silva, E. F. (2012). Seawater Biodegradation of Alkanolamines Used for Co2-Capture from Natural Gas. *International Journal of Greenhouse Gas Control*, 10: 271-277.
- Brandi, P., D'annibale, A., Galli, C., Gentili, P., and Pontes, A. S. N. (2006). In Search for Practical Advantages from the Immobilisation of an Enzyme: The Case of Laccase. *Journal of Molecular Catalysis B: Enzymatic*, 41(1–2): 61-69.
- Brena, B., González-Pombo, P., and Batista-Viera, F. (2013). Immobilization of Enzymes: A Literature Survey. In J. M. Guisan (Ed.), *Immobilization of Enzymes and Cells: Third Edition* (pp. 15-31). Totowa, NJ: Humana Press.

- Brennan, J.D., Benjamin, D., DiBattista, E., and Gulcev, M.D. (2003). Using Sugar and Amino Acid Additives to Stabilize Enzymes within Sol-Gel Derived Silica. *Chem. Mater.*, 15, 737-745.
- Campás, M., and Marty, J.-L. (2006). Encapsulation of Enzymes Using Polymers and Sol-Gel Techniques. In J. M. Guisan (Ed.), *Immobilization of Enzymes* and Cells (pp. 77-85). Totowa, NJ: Humana Press.
- Cardoso, F. P., Aquino Neto, S., Fenga, P. G., Ciancaglini, P., and De Andrade, A.
 R. (2013). Electrochemical Characterization of Methanol/O2 Biofuel Cell: Use of Laccase Biocathode Immobilized with Polypyrrole Film and Pamam Dendrimers. *Electrochimica Acta*, 90: 90-94.
- Cavazzuti, M. (2013). Design of Experiments Optimization Methods: From Theory to Design Scientific and Technological Aspects in Mechanics (pp. 13-42).
 Berlin, Heidelberg: Springer Berlin Heidelberg.
- Champagne, P. P., and Ramsay, J. A. (2010). Dye Decolorization and Detoxification by Laccase Immobilized on Porous Glass Beads. *Bioresource Technology*, 101(7): 2230-2235.
- Chen, J.P. and Lin, W.S. (2003). Sol-gel Powders and Supported Sol-Gel Polymers for Immobilization of Lipase in Ester Synthesis. *Enzyme Microb. Technol.*, 32, 801-811.
- Chen, L., Zhu, S. Y., Wang, H. M., and Wang, Y. M. (2011). One-Step Synthesis of Hierarchical Aluminosilicate Aggregates Using Bifunctional Alkanolamine as Single Template. *Solid State Sciences*, 13(11): 2024-2029.
- Chi, S., and Rochelle, G. T. (2002). Oxidative Degradation of Monoethanolamine. *Industrial & Engineering Chemistry Research*, 41(17): 4178-4186.
- Chutipongtanate, S., Watcharatanyatip, K., Homvises, T., Jaturongkakul, K., and Thongboonkerd, V. (2012). Systematic Comparisons of Various Spectrophotometric and Colorimetric Methods to Measure Concentrations of Protein, Peptide and Amino Acid: Detectable Limits, Linear Dynamic Ranges, Interferences, Practicality and Unit Costs. *Talanta*, 98: 123-129.
- Claus, H. (2004). Laccases: Structure, Reactions, Distribution. *Micron*, 35(1–2): 93-96.
- Crestini, C., Perazzini, R., and Saladino, R. (2010). Oxidative Functionalisation of Lignin by Layer-by-Layer Immobilised Laccases and Laccase Microcapsules. *Applied Catalysis A: General*, 372(2): 115-123.

- Cristóvão, R. O., Amaral, P. F. F., Tavares, A. P. M., Coelho, M. A. Z., Cammarota, M. C., Loureiro, J. M., Boaventura, R. A. R., Macedo, E. A., and Pessoa, F. L. P. (2010). Optimization of Laccase Catalyzed Degradation of Reactive Textile Dyes in Supercritical Carbon Dioxide Medium by Response Surface Methodology. *Reaction Kinetics, Mechanisms and Catalysis*, 99(2): 311-323.
- Daâssi, D., Zouari-Mechichi, H., Frikha, F., Martinez, M. J., Nasri, M., and Mechichi, T. (2013). Decolorization of the Azo Dye Acid Orange 51 by Laccase Produced in Solid Culture of a Newly Isolated Trametes Trogii Strain. 3 Biotech, 3(2): 115-125.
- David, A. E., Yang, A. J., and Wang, N. S. (2011). Enzyme Stabilization and Immobilization by Sol-Gel Entrapment. In S. D. Minteer (Ed.), *Enzyme Stabilization and Immobilization: Methods and Protocols* (pp. 49-66). Totowa, NJ: Humana Press.
- David, A., Thibaud, C., Ovadia, L. and Jacques, L. (2006). Recent Bio-Applications of Sol–Gel Materials. *J Mater. Chem.*, 16, 1013–1030.
- Davis, G. B., Laslett, D., Patterson, B. M., and Johnston, C. D. (2013). Integrating Spatial and Temporal Oxygen Data to Improve the Quantification of in Situ Petroleum Biodegradation Rates. *Journal of Environmental Management*, 117: 42-49.
- De Stefano, L., Rea, I., De Tommasi, E., Rendina, I., Rotiroti, L., Giocondo, M., Longobardi, S., Armenante, A., and Giardina, P. (2009). Bioactive Modification of Silicon Surface Using Self-Assembled Hydrophobins from Pleurotus Ostreatus. *The European Physical Journal E*, 30(2): 181.
- Debecker, D. P., Hulea, V., and Mutin, P. H. (2013). Mesoporous Mixed Oxide Catalysts Via Non-Hydrolytic Sol–Gel: A Review. *Applied Catalysis A: General*, 451: 192-206.
- Eldridge, H. C., Milliken, A., Farmer, C., Hampton, A. S., Wendland, N., Coward, L., Gregory, D. J., and Johnson, C. M. (2017). Efficient Remediation of 17α-Ethinylestradiol by Lentinula Edodes (Shiitake) Laccase. *Biocatalysis and Agricultural Biotechnology*, 10: 64-68.
- Fernández-Fernández, M., Sanromán, M. Á., and Moldes, D. (2013). Recent Developments and Applications of Immobilized Laccase. *Biotechnology Advances*, 31(8): 1808-1825.

- Frauenkron, M., Melder, J.-P., Ruider, G., Rossbacher, R., and Höke, H. (2000). Ethanolamines and Propanolamines Ullmann's Encyclopedia of Industrial Chemistry: Wiley-VCH Verlag GmbH & Co. KGaA.
- Gad, S. C. (2014). Ethanolamine A2 Wexler, Philip Encyclopedia of Toxicology (Third Edition) (pp. 492-495). Oxford: Academic Press.
- Georgieva, S., Godjevargova, T., Portaccio, M., Lepore, M., and Mita, D. G. (2008).
 Advantages in Using Non-Isothermal Bioreactors in Bioremediation of Water
 Polluted by Phenol by Means of Immobilized Laccase from Rhus Vernicifera.
 Journal of Molecular Catalysis B: Enzymatic, 55(3–4): 177-184.
- Gill, I. (2001). Bio-doped Nanocomposite Polymers: Sol-Gel Bioencapsulates. *Chem. Mater.*, 13, 3404-3421.
- Gouedard, C., Picq, D., Launay, F., and Carrette, P. L. (2012). Amine Degradation in Co2 Capture. I. A Review. *International Journal of Greenhouse Gas Control*, 10: 244-270.
- Grace Liu, P.-W., Chang, T. C., Whang, L.-M., Kao, C.-H., Pan, P.-T., and Cheng, S.-S. (2011). Bioremediation of Petroleum Hydrocarbon Contaminated Soil: Effects of Strategies and Microbial Community Shift. *International Biodeterioration & Biodegradation*, 65(8): 1119-1127.
- Guisán, J. M., Penzol, G., Armisen, P., Bastida A, Blanco R. M., Fernandez-Lafuente, R., and García Junceda, E. (1997) Immobilization of enzymes acting on macromolecular substrates.. In: *Immobilization of Enzymes and Cells*, (Bickerstaff, G. F., ed.), Humana Press, Totowa, NJ, 261–275.
- Hansen, B. H., Altin, D., Booth, A., Vang, S.-H., Frenzel, M., Sørheim, K. R., Brakstad, O. G., and Størseth, T. R. (2010). Molecular Effects of Diethanolamine Exposure on Calanus Finmarchicus (Crustacea: Copepoda). Aquatic Toxicology, 99(2): 212-222.
- Hayashi, M., Uchida, R., Unemoto, T., and Miyaki, K. (1964). Enzymic Oxidation of Ethanolamine by Beef Serum. *Chemical and Pharmaceutical Bulletin*, 12(2): 223-227
- Homaei, A. A., Sariri, R., Vianello, F., and Stevanato, R. (2013). Enzyme Immobilization: An Update. *Journal of Chemical Biology*, 6(4): 185-205.
- Huajun Qiu, C. X., Xirong Huang, Yi Ding, Yinbo Qu and Peiji Gao. (1997). Immobilization of Laccase on Nanoporous Gold: Comparative Studies on the

Immobilization Strategies and the Particle Size Effects. *The Journal of physical chemistry*, 113: 2521-2525.

- Hwang, S.Y., Kim, H.K., Choo, J., Seong, G.H., Hien, T.B.D. and Lee, E.K. (2012).
 Effects of Operating Parameters on the Efficiency of Liposomal Encapsulation of Enzymes. *Colloids Surf., B: Biointerfaces, 94, 296-303*.Homaei, A. A., Sariri, R., Vianello, F., and Stevanato, R. (2013). Enzyme Immobilization: An Update. *Journal of Chemical Biology*, 6(4): 185-205.
- Itabaiana Jr, I., Miranda, L. S. d. M., and de Souza R. O. M. A. (2013). Towards a Continuous Flow Environment for Lipase-Catalyzed Reactions. J. Mol. Catal. B: Enzym., 85–86, 1-9.
- Jakobsen, H. A. (2008). Packed Bed Reactors Chemical Reactor Modeling: Multiphase Reactive Flows (pp. 953-984). Berlin, Heidelberg: Springer Berlin Heidelberg.
- Jin, W. and Brennan, J.D. (2002). Properties and Applications of Proteins Encapsulated within Sol-Gel Derived Materials. *Anal. Chim. Acta.*, 461, 1-36.
- Jordaan, J., Mathye, S., Simpson, C., and Brady, D. (2009). Improved Chemical and Physical Stability of Laccase after Spherezyme Immobilisation. *Enzyme and Microbial Technology*, 45(6–7): 432-435.
- Kandimalla, V. B., Tripathi, V. S. and Huangxian, J. (2006) Immobilization of Biomolecules in Sol-Gels: Biological and Analytical Applications. *Crit. Rev. Anal. Chem.*, 36, 73-106.
- Kato, K., Kawachi, Y., and Nakamura, H. (2014). Silica–Enzyme–Ionic Liquid Composites for Improved Enzymatic Activity. *Journal of Asian Ceramic Societies*, 2(1): 33-40.
- Keeling-Tucker, T., Rakic, M., Spong, C. and Brennan, J.D. (2000). Controlling the Material Properties and Biological Activity of Lipase within Sol-Gel derived Bioglasses via Organosilane and Polymer Doping. *Chem. Mater.*, 12, 3695-3704.
- Khan, J. A., and Vulfson, E. N. (2001). Microencapsulation of Enzymes and Cells for Nonaqueous Biotransformations. In E. N. Vulfson, P. J. Halling andH. L. Holland (Eds.), *Enzymes in Nonaqueous Solvents: Methods and Protocols* (pp. 31-40). Totowa, NJ: Humana Press.
- Khani, Z., Jolivalt, C., Cretin, M., Tingry, S., and Innocent, C. (2006). Alginate/Carbon Composite Beads for Laccase and Glucose Oxidase

Encapsulation: Application in Biofuel Cell Technology. *Biotechnology Letters*, 28(22): 1779-1786.

- Kim, J., Grate, J.W. and Wang, P. (2006). Nanostructure for Enzyme Stabilization. *Chem. Eng. Sci.*, 61, 1017-1026.
- Kumar, V., Jahan, F., Raghuwanshi, S., Mahajan, R. V., and Saxena, R. K. (2013).
 Immobilization of Rhizopus Oryzae Lipase on Magnetic Fe3o4-Chitosan
 Beads and Its Potential in Phenolic Acids Ester Synthesis. *Biotechnology and Bioprocess Engineering*, 18(4): 787-795.
- Lepaumier, H., Grimstvedt, A., Vernstad, K., Zahlsen, K., and Svendsen, H. F. (2011). Degradation of Mmea at Absorber and Stripper Conditions. *Chemical Engineering Science*, 66(15): 3491-3498.
- Li, B., Chen, Y., Chen, X., Liu, D., Niu, H., Xiong, J., Wu, J., Xie, J., Bai, J., and Ying, H. (2012). A Novel Immobilization Method for Nuclease P1 on Macroporous Absorbent Resin with Glutaraldehyde Cross-Linking and Determination of Its Properties. *Process Biochemistry*, 47(4): 665-670.
- Li, J., Cai, J., Zhong, L., and Du, Y. (2012). Immobilization of a Protease on Modified Chitosan Beads for the Depolymerization of Chitosan. *Carbohydrate Polymers*, 87(4): 2697-2705.
- Li, Q.-Y., Wang, P.-Y., Zhou, Y.-L., Nie, Z.-R., and Wei, Q. (2016). A Magnetic Mesoporous Sio₂/Fe₃O₄ Hollow Microsphere with a Novel Network-Like Composite Shell: Synthesis and Application on Laccase Immobilization. *Journal of Sol-Gel Science and Technology*, 78(3): 523-530.
- Libralato, G., Volpi Ghirardini, A., and Avezzù, F. (2010). Seawater Ecotoxicity of Monoethanolamine, Diethanolamine and Triethanolamine. *Journal of Hazardous Materials*, 176(1–3): 535-539.
- Livage, J., Coradin, T. and Rouz, C. (2001). Encapsulation of Biomolecules in Silica Gels. J. Phys.: Condens. Matter., 13, R763-R691.
- Lu, C.-S., Chen, C.-C., Mai, F.-D., and Li, H.-K. (2009). Identification of the Degradation Pathways of Alkanolamines with Tio2 Photocatalysis. *Journal of Hazardous Materials*, 165(1–3): 306-316.
- Lundberg, M., and Borowski, T. (2013). Oxoferryl Species in Mononuclear Non-Heme Iron Enzymes: Biosynthesis, Properties and Reactivity from a Theoretical Perspective. *Coordination Chemistry Reviews*, 257(1): 277-289.

- Maleki, A., Rahimi, R., Maleki, S., and Hamidi, N. (2014). Synthesis and Characterization of Magnetic Bromochromate Hybrid Nanomaterials with Triphenylphosphine Surface-Modified Iron Oxide Nanoparticles and Their Catalytic Application in Multicomponent Reactions. *RSC Advances*, 4(56): 29765-29771.
- Mansor, A. F., Mohidem, N. A., Wan Mohd Zawawi, W. N. I., Othman, N. S., Endud, S., and Mat, H. (2016). The Optimization of Synthesis Conditions for Laccase Entrapment in Mesoporous Silica Microparticles by Response Surface Methodology. *Microporous and Mesoporous Materials*, 220: 308-314.
- Mateo, C., Palomo, J. M., Fernandez-Lorente, G., Guisan, J. M., and Fernandez-Lafuente, R. (2007). Improvement of Enzyme Activity, Stability and Selectivity Via Immobilization Techniques. *Enzyme and Microbial Technology*, 40(6): 1451-1463.
- Matijošytė, I., Arends, I. W. C. E., De Vries, S., and Sheldon, R. A. (2010).
 Preparation and Use of Cross-Linked Enzyme Aggregates (Cleas) of Laccases. *Journal of Molecular Catalysis B: Enzymatic*, 62(2): 142-148.
- Matsune, H., Jogasaki, H., Date, M., Takenaka, S., and Kishida, M. (2006). One-Pot Synthesis and Characterization of Laccase-Entrapped Magnetic Nanobeads. *Chemistry Letters*, 35(12): 1356-1357.
- Matsuura, S.-I., El-Safty, S. A., Chiba, M., Tomon, E., Tsunoda, T. and Hanaoka, T A. (2012). Enzyme Encapsulation using Highly Ordered Mesoporous
 Monoliths. *Mater. Lett.*, 89, 184-187.
- Michizoe, J., Goto, M., and Furusaki, S. (2001). Catalytic Activity of Laccase Hosted in Reversed Micelles. *Journal of Bioscience and Bioengineering*, 92(1): 67-71.
- Moehlenbrock, M. J., and Minteer, S. D. (2011). Introduction to the Field of Enzyme Immobilization and Stabilization. In S. D. Minteer (Ed.), *Enzyme Stabilization and Immobilization: Methods and Protocols* (pp. 1-7). Totowa, NJ: Humana Press.
- Mohajershojaei, K., Mahmoodi, N. M., and Khosravi, A. (2015). Immobilization of Laccase Enzyme onto Titania Nanoparticle and Decolorization of Dyes from Single and Binary Systems. *Biotechnology and Bioprocess Engineering*, 20(1): 109-116.

- Mohamed, M. F., and Hollfelder, F. (2013). Efficient, Crosswise Catalytic Promiscuity among Enzymes That Catalyze Phosphoryl Transfer. *Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics*, 1834(1): 417-424.
- Mohidem, and Mat. (2012a). Catalytic Activity and Stability of Laccase Entrapped in Sol–Gel Silica with Additives. *Journal of Sol-Gel Science and Technology*, 61(1): 96-103.
- Mohidem, and Mat. (2012b). The Catalytic Activity Enhancement and Biodegradation Potential of Free Laccase and Novel Sol–Gel Laccase in Non-Conventional Solvents. *Bioresource Technology*, 114: 472-477.
- Montgomery, D.C. and Runger, G.C. (2002). Applied Statistics and Probability for Engineers, third ed. John Wiley and Sons (Asia) Pvt. Ltd., Singapore.
- Mrklas, O., Chu, A., Lunn, S., and Bentley, L. R. (2004). Biodegradation of Monoethanolamine, Ethylene Glycol and Triethylene Glycol in Laboratory Bioreactors. *Water, Air, and Soil Pollution*, 159(1): 249-263.
- Mundhwa, M., and Henni, A. (2007). Molar Heat Capacity of Various Aqueous Alkanolamine Solutions from 303.15 K to 353.15 K. Journal of Chemical & Engineering Data, 52(2): 491-498.
- Najafpour, G. D. (2015). Chapter 2 Enzyme Technology. *Biochemical Engineering and Biotechnology*(Second Edition): 19-49.
- Narrod, S. A., and Jakoby, W. B. (1964). Metabolisme of Ethanolamine. An Ethanolamine Oxidase. . *The Journal of Biological Chemistry*, 239(7): 2189-2193.
- Nelson, D. L., and Cox, M. M. (2008). Principle of Biochemistry (Vol. Fifth).
- Niladevi, K. N., and Prema, P. (2008). Immobilization of Laccase from Streptomyces Psammoticus and Its Application in Phenol Removal Using Packed Bed Reactor. World Journal of Microbiology and Biotechnology, 24(7): 1215-1222.
- Nogala, W., Szot, K., Burchardt, M., Jönsson-Niedziolka, M., Rogalski, J., Wittstock, G., and Opallo, M. (2010). Scanning Electrochemical Microscopy Activity Mapping of Electrodes Modified with Laccase Encapsulated in Sol– Gel Processed Matrix. *Bioelectrochemistry*, 79(1): 101-107.
- Noor, E., Flamholz, A., Liebermeister, W., Bar-Even, A., and Milo, R. (2013). A Note on the Kinetics of Enzyme Action: A Decomposition That Highlights Thermodynamic Effects. *FEBS Letters*, 587(17): 2772-2777.

- Noureddini, H., and Gao, X. (2007). Characterization of Sol-Gel Immobilized Lipases. *Journal of Sol-Gel Science and Technology*, 41(1): 31-41.
- Nthumbi, R. M., and Ngila, J. C. (2016). Electrospun and Functionalized Pvdf/Pan Nanocatalyst-Loaded Composite for Dechlorination and Photodegradation of Pesticides in Contaminated Water. *Environmental Science and Pollution Research*, 23(20): 20214-20231.
- Okazaki, S.-y., Goto, M., Wariishi, H., Tanaka, H., and Furusaki, S. (2000). Characterization and Catalytic Property of Surfactant-Laccase Complex in Organic Media. *Biotechnology Progress*, 16(4): 583-588.
- Okazaki, S.-y., Michizoe, J., Goto, M., Furusaki, S., Wariishi, H., and Tanaka, H. (2002). Oxidation of Bisphenol a Catalyzed by Laccase Hosted in Reversed Micelles in Organic Media. *Enzyme and Microbial Technology*, 31(3): 227-232.
- Ota, H., Tamezane, H., Sasano, Y., Hokazono, E., Yasuda, Y., S-I, S., Imamura, S., Tamura, T., and Osawa, S. (2008). Enzymatic Characterization of an Amine
- Owens, G. J., Singh, R. K., Foroutan, F., Alqaysi, M., Han, C.-M., Mahapatra, C., Kim, H.-W., and Knowles, J. C. (2016). Sol–Gel Based Materials for Biomedical Applications. *Progress in Materials Science*, 77: 1-79.
 Oxidase from Arthrobacter Sp. Used to MeasurePhosphatidylethanolamine. *Bioscience, Biotechnology, and Biochemistry*, 72(10).
- Pabby, A. K. and Sastre, A. M. (2013). State-Of-The-Art Review on Hollow Fibre Contactor Technology and Membrane-Based Extraction Processes. J Membran. Sci., 430, 263-303.
- Panchal, S., and Verma, R. J. (2016). Effect of Diethanolamine on Testicular Steroidogenesis and Its Amelioration by Curcumin. Asian Pacific Journal of Reproduction, 5(2): 128-131.
- Park, J. H., Xue, H., Jung, J. S., and Ryu, K. (2012). Immobilization of Laccase on Carbon Nanomaterials. *Korean Journal of Chemical Engineering*, 29(10): 1409-1412.
- Pelley, J. W., Garner, C. W., and Little, G. H. (1978). A Simple Rapid Biuret Method for the Estimation of Protein in Samples Containing Thiols. *Analytical Biochemistry*, 86(1): 341-343.
- Perry, C. C. and Keeling-Tucker, T. (2000). Aspects of the Bioinorganic Chemistry of Silicon in Conjunction with Biometals. *J. Biol. Inorg. Chem.*, 5, 537-550.

- Piontek, K., Antorini, M., and Choinowski, T. (2002). Crystal Structure of a Laccase from the FungusTrametes Versicolor at 1.90-Å Resolution Containing a Full Complement of Coppers. *The Journal of Biological Chemistry*, 277: 37663.
- Qiu, L., and Huang, Z. (2010). The Treatment of Chlorophenols with Laccase Immobilized on Sol–Gel-Derived Silica. World Journal of Microbiology and Biotechnology, 26(5): 775-781.
- Quan, D., and Shin, W. (2004). Modification of Electrode Surface for Covalent Immobilization of Laccase. *Materials Science and Engineering: C*, 24(1–2): 113-115.
- Rahimi, R., Maleki, A., and Maleki, S. (2014). Synthesis and Characterization of a New Magnetic Bromochromate Hybrid Nanomaterial with Triethylamine Surface Modified Iron Oxide Nanoparticles. *Chinese Chemical Letters*, 25(6): 919-922.
- Raseda, N., Park, J., and Ryu, K. (2016). Laccase-Catalyzed Polymerization of M-Phenylenediamine in Aqueous Buffers. *Korean Journal of Chemical Engineering*, 33(10): 3011-3015.
- Rasera, K., Ferla, J., Dillon, A. J. P., Riveiros, R., and Zeni, M. (2009). Immobilization of Laccase from Pleurotus Sajor-Caju in Polyamide Membranes. *Desalination*, 245(1–3): 657-661.
- Ravindra, P., and Jegannathan, K. R. (2015). Production of Biodiesel Using Lipase Encapsulated in K-Carrageenan. *SpringerBriefs in Bioengineering*: 23-63.
- Reetz, M. T. (2006). Practical Protocols for Lipase Immobilization Via Sol-Gel Techniques. In J. M. Guisan (Ed.), *Immobilization of Enzymes and Cells* (pp. 65-76). Totowa, NJ: Humana Press.
- Rekuć, A., Bryjak, J., Szymańska, K., and Jarzębski, A. B. (2009). Laccase Immobilization on Mesostructured Cellular Foams Affords Preparations with Ultra High Activity. *Process Biochemistry*, 44(2): 191-198.
- Rekuć, A., Bryjak, J., Szymańska, K., and Jarzębski, A. B. (2010). Very Stable Silica-Gel-Bound Laccase Biocatalysts for the Selective Oxidation in Continuous Systems. *Bioresource Technology*, 101(7): 2076-2083.
- Rodríguez Couto, S., and Toca Herrera, J. L. (2006). Industrial and Biotechnological Applications of Laccases: A Review. *Biotechnology Advances*, 24(5): 500-513.

- Rodríguez-Delgado, M. M., Alemán-Nava, G. S., Rodríguez-Delgado, J. M., Dieck-Assad, G., Martínez-Chapa, S. O., Barceló, D., and Parra, R. (2015). Laccase-Based Biosensors for Detection of Phenolic Compounds. *TrAC Trends in Analytical Chemistry*, 74: 21-45.
- Salis, A., Pisano, M., Monduzzi, M., Solinas, V., and Sanjust, E. (2009). Laccase from Pleurotus Sajor-Caju on Functionalised Sba-15 Mesoporous Silica: Immobilisation and Use for the Oxidation of Phenolic Compounds. *Journal* of Molecular Catalysis B: Enzymatic, 58(1–4): 175-180.
- Sánchez-Moreno, I., Oroz-Guinea, I., Iturrate, L., and García-Junceda, E. (2012).
 7.20 Multi-Enzyme Reactions *Comprehensive Chirality* (pp. 430-453).
 Amsterdam: Elsevier.
- Sassolas, A., Hayat, A., and Marty, J.-L. (2013). Enzyme Immobilization by Entrapment within a Gel Network. In J. M. Guisan (Ed.), *Immobilization of Enzymes and Cells: Third Edition* (pp. 229-239). Totowa, NJ: Humana Press.
- Sauvant, P., Cansell, M., Sassi, A.H. and Atgié, C. (2012).Vitamin A Enrichment: Caution with Encapsulation Strategies used for Food Applications. *Food Res. Int.*, 46, 469-479.
- Seoud, M. A. and Maachi, R. (2003). Biodegradation of Naphthalene by Free and Alginate Entrapped *Pseudomonas sp.* Z. Naturforsch. C 58 (9-10), 726-731, 2003.
- Shuler, and Kargi. (2005). Bioprocess Engineering. (Second).
- Soleimani, M., Khani, A., and Najafzadeh, K. (2012). A-Amylase Immobilization on the Silica Nanoparticles for Cleaning Performance Towards Starch Soils in Laundry Detergents. *Journal of Molecular Catalysis B: Enzymatic*, 74(1–2): 1-5.
- Stelmaszyńska, T., and Zgliczyński, J. M. (1974). Myeloperoxidase of Human Neutrophilic Granulocytes as Chlorinating Enzyme. 45: 305-312.
- Tang, K., Baskaran, V., and Nemati, M. (2009). Bacteria of the Sulphur Cycle: An Overview of Microbiology, Biokinetics and Their Role in Petroleum and Mining Industries. *Biochemical Engineering Journal*, 44(1): 73-94.
- Torres, E., and Ayala, M. (2010). Biocatalysis Based on Heme Peroxidases. Springer-Verlag Berlin Heidelberg.
- Trevan, M. D. (1988). Enzyme Immobilization by Entrapment. In J. M. Walker (Ed.), New Protein Techniques (pp. 491-494). Totowa, NJ: Humana Press.

- Twala, B. V., Sewell, B. T., and Jordaan, J. (2012). Immobilisation and Characterisation of Biocatalytic Co-Factor Recycling Enzymes, Glucose Dehydrogenase and Nadh Oxidase, on Aldehyde Functional Resyn[™] Polymer Microspheres. *Enzyme and Microbial Technology*, 50(6–7): 331-336.
- Upadhyay, P., Shrivastava, R., and Agrawal, P. K. (2016). Bioprospecting and Biotechnological Applications of Fungal Laccase. *3 Biotech*, 6(1): 15.
- Vaidya, P. D., and Kenig, E. Y. (2009). Kinetics of Carbonyl Sulfide Reaction with Alkanolamines: A Review. *Chemical Engineering Journal*, 148(2–3): 207-211.
- Vera-Avila, L. E., Morales-Zamudio, E., and Garcia-Camacho, M. P. (2004). Activity and Reusability of Sol-Gel Encapsulated A-Amylase and Catalase. Performance in Flow-through Systems. *Journal of Sol-Gel Science and Technology*, 30(3): 197-204.
- Wang, Y., Chen, X., Liu, J., He, F., and Wang, R. (2013). Immobilization of Laccase by Cu2+ Chelate Affinity Interaction on Surface-Modified Magnetic Silica Particles and Its Use for the Removal of 2,4-Dichlorophenol. *Environmental Science and Pollution Research*, 20(9): 6222-6231.
- Werther, J. (2000). Fluidized-Bed Reactors Ullmann's Encyclopedia of Industrial Chemistry: Wiley-VCH Verlag GmbH & Co. KGaA.
- Wesenberg, D., Kyriakides, I., and Agathos, S. N. (2003). White-Rot Fungi and Their Enzymes for the Treatment of Industrial Dye Effluents. *Biotechnology Advances*, 22(1–2): 161-187.
- Widsten, P., and Kandelbauer, A. (2008). Laccase Applications in the Forest Products Industry: A Review. *Enzyme and Microbial Technology*, 42(4): 293-307.
- Williams, G. R., and Callely, A. G. (1982). The Biodegradation of Diethanolamine and Triethanolamine by a Yellow Gram-Negative Rod. *Microbiology*, 128(6): 1203-1209.
- Wu, X.-c., Zhang, Y., Wu, C.-y., and Wu, H.-x. (2012). Preparation and Characterization of Magnetic Fe3o4/Crgo Nanocomposites for Enzyme Immobilization. *Transactions of Nonferrous Metals Society of China*, 22, Supplement 1: s162-s168.

- Xi, H., Wang, Z., Chen, Y., Li, W., Sun, L., and Fang, L. (2012). The Relationship between Hydrogen-Bonded Ion-Pair Stability and Transdermal Penetration of Lornoxicam with Organic Amines. *European Journal of Pharmaceutical Sciences*, 47(2): 325-330.
- Xu, A., Li, X., Xiong, Z., Wang, Q., Cai, Y., and Zeng, Q. (2012). High Catalytic Activity of Cobalt (Ii)-Triethylamine Complex Towards Orange Ii Degradation with H2o2 as an Oxidant under Ambient Conditions. *Catalysis Communications*, 26(Supplement C): 44-47.
- Xue, R. and Woodley, J.M. (2012). Review. Process Technology for Multi-Enzymatic Reaction Systems. *Bioresour. Technol.*, 115, 183–195.
- Yi, Y., Neufeld, R. and Kermasha, S. (2007). Controlling Sol-Gel Properties Enhancing Entrapped Membrane Protein Activity through Doping Additives. *Sol-Gel Sci. Technol.*, 43, 161-170.
- Yu, X.-L., and He, Y. (2017). Application of Box-Behnken Designs in Parameters Optimization of Differential Pulse Anodic Stripping Voltammetry for Lead(Ii) Determination in Two Electrolytes. *Scientific Reports*, 7(1): 2789.
- Zhou, H., Li, W., Shou, Q., Gao, H., Xu, P., Deng, F., Liu, H. (2012). Immobilization of Penicillin G Acylase on Magnetic Nanoparticles Modified by Ionic Liquids. *Chin. J. Chem. Eng.*, 20, 146-151.
- Zurita, Repetto, G., Jos, A., Del Peso, A., Salguero, M., López-ArtíGuez, M., Olano,
 D., and Cameán, A. (2005). Ecotoxicological Evaluation of Diethanolamine
 Using a Battery of Microbiotests. *Toxicology in Vitro*, 19(7): 879-886.