

β -GLUCOSIDASE FROM *Trichoderma harzianum* T12 AS GREEN FUNGICIDE
AGAINST *Macrophomina phaseolina* IN SOYBEAN (*Glycine max* L.)

ELHAM KHALILI

UNIVERSITI TEKNOLOGI MALAYSIA

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ELHAM KHALILI

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*Specially dedicated to:
My family for their endless support and motivation.*

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ABSTRACT

Macrophomina phaseolina (Tassi) Goid remains the prevailing causal agent of charcoal rot disease that can significantly suppress yields of a variety of crops. Its wide host range and survivability under arid conditions as well as the ineffectiveness of fungicides have spurred scientific endeavors in search of alternative avenues to control this phytopathogen. The present study is aimed to provide empirical evidence on the efficacy of β -glucosidase from *Trichoderma harzianum* T12 as a biological control agent against *M. phaseolina*. *In-vitro* pathogenicity tests on 60 isolates of *M. phaseolina* and 30 isolates of *T. harzianum*, collected from different areas of the Mazandaran province in Iran revealed the isolates, M2 of *M. phaseolina* and the T12 of *T. harzianum* were the most virulent and effective in inhibiting growth of *M. phaseolina*, respectively. The present study showed that biochemical and phylogenetic analyses and BIOLOG results confirmed the fungal antagonists and phytopathogen were *T. harzianum* (Rifai) and *M. phaseolina* (Tassi) Goid, respectively. Purified extracellular β -glucosidase of *T. harzianum* inhibited the growth of *M. phaseolina* as seen from the large halo zones, indicating its possible application as a green fungicide against *M. phaseolina*. The β -glucosidase had an optimum pH (7) and temperature 45°C, respectively, remarkably stable up to 240 min with a half-life of $t_{1/2} = 210$ min at 40 °C to 60 °C. Zn^{2+} , Mn^{2+} , and Tween 80 enhanced its activity while was substantially inhibited by Fe^{3+} . Enzyme activity was the highest when wheat bran and $(NH_4)_2SO_4$ were used as carbon and nitrogen sources respectively. The kinetic parameters for β -glucosidase T12, K_m , V_{max} and k_{cat} were estimated as 0.79 mM, 8.45 $mM\ min^{-1}\ mg^{-1}$ protein and 10.69 s^{-1} , respectively, to give a turnover number of 10.69 s^{-1} . Optimization by the Box-Behnken Design (BBD) based on: temperature, carbon sources, inoculum size and pH (7), exhibited the highest β -glucosidase activity (1260 U/mL) at 45°C, pH 7, using a carbon source 10 % (w/v) and inoculum size of 5 % (w/v). The BBD optimization for the application of the β -glucosidase formulation from *T. harzianum* to control infestation of *M. phaseolina* M2 was carried out on soybean plants grown under a greenhouse condition. Under an optimized condition, the lowest plant disease index (PDI) of 4.32% ($R^2 = 0.9676$) was attained using 10 mM Zn^{2+} , Tween 80 at 2 % (w/v) an enzyme concentration at 15 mg/L and an irrigation frequency of 2 times/week. A comparative study showed the developed formulation gave the lowest PDI (4.14 %) ($p < 0.05$) followed by the antagonist *T. harzianum* Rifai (26.13 %) and the commercial fungicide, Carbendazim (32.45 %). The assessments cost revealed that the enzyme formulation only costs at USD34/acre as compared to Carbendazim at USD240/acre. Hence, the findings affirmed that the novel use of crude β -glucosidase from the growth supernatant of *T. harzianum* was efficient in combating charcoal rot disease. Since the enzyme formulation was substantially cheaper and its application combines the practicality of an *in-situ* spraying for rapid control of *M. phaseolina* infestation, the technique proposed here was prospectively feasible to control such disease in crops.

ABSTRAK

Macrophomina phaseolina (Tassi) Goid merupakan penyebab penyakit reput arang yang memberi kesan ketara kepada penghasilan pelbagai tanaman. Kepelbagaian perumah dan keupayaan untuk terus hidup di bawah keadaan tandus, ditambah pula dengan ketidakberkesanan fungisid telah merangsang kajian ini untuk mencari alternatif bagi mengawal fitopatogen ini. Kajian ini bertujuan memberi bukti secara empirikal ke atas keupayaan β -glukosida daripada *Trichoderma harzianum* T12 sebagai agen kawalan biologi terhadap penyakit reput arang. Ujian kepathogenenikan secara *in-vitro* ke atas 60 isolat *M. phaseolina* dan 30 isolat *T. harzianum*, yang dikutip dari kawasan yang berlainan di wilayah Mazandaran di Iran mendedahkan bahawa isolat M2 *M. phaseolina* ialah yang paling patogenik manakala isolat T12 *T. harzianum* paling efektif merencat pertumbuhan *M. phaseolina* M2. Ujian biokimia, analisa filogeni dan keputusan BIOLOG masing-masing mengesahkan bahawa kulat antagonistik dan patogenik tersebut adalah *T. harzianum* (Rifai) dan *M. phaseolina* (Tassi) Goid. β -glukosida ekstrak *T. harzianum* didapati menghalang pertumbuhan *M. phaseolina* seperti yang dilihat dari zon halo yang besar, menunjukkan kemungkinan penggunaannya sebagai fungisida hijau terhadap *M. phaseolina*. β -glukosida tersebut mempunyai pH dan suhu optimum masing-masing pada pH 7 dan 45°C, sangat stabil pada 240 minit dengan separuh hayat $t_{1/2} = 210$ min pada suhu 40 °C to 60 °C. Ion Zn^{2+} , Mn^{2+} and surfaktan Tween 80 didapati dapat meningkatkan aktiviti manakala Fe^{3+} merencat aktiviti dengan ketara. Aktiviti enzim adalah pada tahap tertinggi apabila dedak gandum dan amonium sulfida digunakan sebagai sumber karbon dan nitrogen masing-masing. Parameter kinetik K_m , V_{max} dan k_{cat} β -glukosida T12 yang dijangka masing-masing pada 0.79 mM, 8.45 $mM \min^{-1} mg^{-1}$ protein and 10.69 s^{-1} , unuk memberi jumlah perolehan sebanyak 10.69 s^{-1} . Pengoptimum menggunakan Design Box-Behken (BBD) berdasarkan empat parameter: suhu, sumber karbon, saiz inokulum, pH (7) mendedahkan keadaan optimum. Oleh itu, penghasilan aktiviti β -glukosida (1260 U/mL) adalah pada suhu 45°C, pH 7, dengan sumber karbon 10 % (w/v) dan saiz inokulum 5 % (w/v). Indeks penyakit tumbuhan terendah (PDI) sebesar 4.32% ($R^2 = 0.9676$) dicapai dengan menggunakan Zn^{2+} (10 mM), Tween 80 pada 2% (w/v), 15 mg/L kepekatan enzim dan kekerapan pengairan sekurang-kurangnya 2 kali/minggu. β -glukosida dari *T. harzianum* digunakan untuk mengawal serangan *M. phaseolina* M2 dilakukan pada penggunaan pokok soya yang tumbuh di dalam persekitaran rumah hijau. Pengoptimuman BBD untuk penggunaan formula indeks penyakit tumbuhan yang paling rendah (PDI) ialah pada 4.32%. Kajian komparatif menunjukkan formula β -glucosidase memberikan PDI paling rendah (4.14%) ($p < 0.05$) berbanding antagonis *T. harzianum* Rifai (26.13%) dan racun kulat komersial, Carbendazim (32.45%). Penilaian kos menunjukkan kos formula yang dibangunkan hanya USD34/ekar berbanding Carbendazim USD 240/ekar. Oleh itu, hasil kajian mengesahkan bahawa β -glucosida yang diperolehi dari supernatant pertumbuhan *T. harzianum* adalah cekap dalam memerangi penyakit reput arang. Memandangkan formula β -glukosida yang dibangunkan lebih murah dan penggunaannya yang praktikal dengan hanya semburan secara *in-situ* untuk kawalan segera *M. phaseolina*. Oleh itu, teknik yang dicadangkan di sini adalah berpotensi dan boleh digunakan untuk mengawal penyakit reput arang pada tanaman.

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LIST OF ABBREVIATIONS

°C	-	Centigrade
A_{nm}	-	Absorption Spectroscopy at nm Light Source
ANOVA	-	Analysis of Variance
APS	-	Ammonium Per Sulfate
atm	-	Airborne Thematic Mapper
BLASTn	-	Basic Local Alignment Search Tool – Nucleotide
bp	-	Base Pairs
BSA	-	Bovine Serum Albumin
Ca	-	Calcium
$CaCl_2$	-	Calcium Chloride
CHR	-	Charcoal Rot
cm	-	Centimetre
$CoCl_2$	-	Cobalt Chloride
CRD	-	Completely Randomized Design
Cu	-	Copper
DMRT	-	Duncan's Multiple Range Test
DNA	-	Deoxyribonucleic Acid
dNTPs	-	Deoxynucleotide Triphosphates
E.q	-	Equation
ECe	-	Electrical Conductivity
EDTA	-	Ethylene Diamine Tetraacetic Acid
EtBr	-	Ethidium Bromide
Fe	-	Iron
$FeSO_4$	-	Ferrous Sulfate

FeSO ₄ •7H ₂ O	-	Ferrous Sulfate Heptahydrate
GIP	-	Growth Inhibition Percentage
h	-	Hour
H ₂ O	-	Water
HCl	-	Hydrogen Chloride
ITS	-	Internal Transcribed Space
K	-	Potassium
kDa	-	kilodalton
kg	-	Kilogram
KH ₂ PO ₄	-	Potassium Dihydrogen Phosphate
KNO ₃	-	Potassium Nitrate
Li	-	Lithium
M	-	Molarity (Molar)
MEGA6	-	Molecular Evolutionary Genetics Analysis Software
mg	-	Milligram
Mg	-	Magnesium
MgCl ₂	-	Magnesium Chloride
MgSO ₄ •H ₂ O	-	Magnesium Sulfate
MgSO ₄	-	Magnesium Sulfate
min	-	Minutes
mL	-	Milliliter
mM	-	Millimolar
mm	-	Millimetre
Mn	-	Manganese
Na	-	Sodium
NaCl	-	Sodium Chloride
NaOH	-	Sodium Hydroxide
NaNO ₃	-	Sodium Nitrate
NCBI	-	National Center for Biotechnology Information
(NH ₄) ₂ SO ₄	-	Ammonium Sulfate
NH ₄ Cl	-	Ammonium Chloride

NH_4NO_3	-	Ammonium Nitrate
OVAT	-	One-Variable-at-A-Time
Pb	-	Lead
PCR	-	Polymerase Chain Reaction
PDA	-	Potato Dextrose Agar
PDB	-	Potato Dextrose Broth
PDI	-	Plant Disease Index
pKa	-	Acid Dissociation Constant
pNPG	-	P-nitrophenyl- β -D-glucopyranoside
R^2	-	Coefficient of Determination
R7	-	Yellowing of the Leaves and Yellow Pods at 50% Growing stage
rpm	-	Revolution Per Minute
rRNA	-	Ribosomal Ribonucleic Acid
RSM	-	Response Surface Methodology
s	-	Second
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	-	Manganese Sulfate Dihydrate
ZnSO_4	-	Zinc sulfate
SDS- PAGE	-	Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis
TEMED	-	Tetra Methyl Ethylene Diamine
TNV	-	Total Neutralization Value
U	-	Unite
US	-	United States Dollars
UV	-	Ultraviolet
v/v	-	Volume Percentage per 100mL volume
W	-	Watt
w/v	-	Weight per 100mL Volume Percentage
xg	-	Times Gravity
Zn	-	Zinc
μ	-	Mikro
μg	-	Microgram
μL	-	Microliter

U/ml - Unit Per Millilitre

DNA BASES

A - Adenine

C - Cytosine

G - Guanine

M - Amino; represented by either A or C

N - Any base; A or C or G or T

R - Purine; Represented by Either G or A

T - Thymine

W - Pyrimidine; Represented by Either C or T

AMINO ACIDS

A or Ala - Alanine

C or Cys - Cysteine

D or Asp - Aspartic Acid

E or Glu - Glutamic Acid

F or Phe - Phenylalanine

G or Gly - Glycine

H or His - Histidine

I or Ile - Isoleucine

K or Lys - Lysine

L or Leu - Leucine

N or Asn - Asparagine

P or Pro - Proline

Q or Gln - Glutamine

R or Arg - Arginine

S or Ser - Serine

T or Thr - Threonine

V or Val - Valine

W or Trp - Tryptophan

Y or Tyr - Tyrosine

APPENDICES

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CHAPTER 1

INTRODUCTION

1.1 Background of Study

Among the plant diseases, soil-borne diseases are regarded to be more devastating than air-borne or seed-borne diseases accounting for 10-20% of annual yield losses (Kumar *et al.*, 2016). Such diseases are mainly caused by phytopathogenic fungi that constitutes to approximately 10,000 species of such fungi. Among the phytopathogenic fungi, *Macrophomina phaseolina* (Tassi) Goid is one of the most ubiquitous ones which causes the fatal charcoal rot disease and capable of infecting approximately 700 plant species belonging to over 100 families of monocots and dicots. Such extraordinary adaptability of *M. phaseolina* originates in its significant physiological, morphological and pathogenic diversity that has resulted in its wide distribution across diverse climatic conditions, ranging from arid to tropical regions (Ambrosio *et al.*, 2015).

The fungus is also an exceptionally robust soil-borne pathogen that devastate a myriad of major agricultural crops *viz.* grains, legumes, oil seeds, jute and cotton (Ma *et al.*, 2010; Rayatpanah *et al.*, 2012; Sun *et al.*, 2016) as well as some vegetables and fruits (Ambrosio *et al.*, 2015). The fungus can survive for long periods in soil, as long as 15 years as resistant structures (sclerotia and chlamydospores) without relying on a host (Sanchez *et al.*, 2016). Charcoal rot caused by *M. phaseolina* is also disseminated by wind, soil and contact with infected

plant material (Gaige *et al.*, 2010). Formation of the 'black leg' in infected plants is symptomatic of the charcoal rot disease caused by the formation of sclerotia in the plant crown. Infected plants generally have weak appearances and decolourized leaves with fewer black secondary roots (Santos *et al.*, 2016). The infected plants eventually die from exposure to phytotoxic metabolites released by *M. phaseolina*, such as phaseolinone, as well as other complications, such as vascular blockages that compromise nutrient transport (Santos *et al.*, 2016).

Various types of agricultural management strategies to control plant diseases caused by *M. phaseolina* using cultural, biological and chemical techniques have been investigated for decades. Cultural practices such as using crop rotation, soil solarization and tolerant cultivars are not sufficiently efficacious on its own and are usually combined with other controls (biological agents, low doses of pesticides) to be more effective. For such technique to work, a higher level of technological support system is needed hence resulting in higher costs of implementation (Patel *et al.*, 2014). So far, the use of fungicides and fumigants to control *M. phaseolina* infestation in crops has been the main choice in the agricultural sector. However, these approaches have been found ineffective and uneconomical. Recent studies have shown that pesticide use per hectare, had generally increased more than proportionally with crop output per hectare. A 1% increase in crop output per hectare was matched with 1.8 % increase in pesticide use per hectare (Patel *et al.*, 2014). This is a particularly worrying trend in agricultural practices.

The indiscriminate use of pesticides has incurred serious concerns over the long term deleterious effects of pesticides on the ecology and human health (Chamorro *et al.*, 2015; Pastrana *et al.*, 2016). Agricultural run-offs containing concentrated amounts of pesticides are highly mobile and can pollute nearby water bodies. These toxic compounds are naturally bioaccumulated through our food chain and ingested by mammals and bird populations (Chua *et al.*, 2014; Edbeib *et al.*, 2016). In one hand, biological techniques using biological control agents and agri-biotechnology i.e. genetically modified crops developed using recombinant DNA technology appears more promising and ecologically friendlier to combat plant diseases i.e. charcoal rot. But both techniques have been subjected to much debate,

due to concerns over new risks on food security as well as the emergence of superweeds and superpest. Even so, the former lacks the consistency in the field studies (Gerbore *et al.*, 2014) which limits its application. Aside from the unknown long-term and other unintended effects on the environment due to the loss of biodiversity, the issue pertaining to food allergies has also been mentioned (Maghari *et al.*, 2011, Arancibia *et al.*, 2013).

In view of the limitations in current management strategies to control charcoal rot disease in crops caused by *M. phaseolina*, the study believes the answer to solving this problem may lie in a simple but effective technique based on a biological control approach using bioactive secretions produced by a well-known antagonistic fungus, *T. harzianum*. For decades, extensive research carried out on the fungus was solely focused on the isolation and *in-situ* cultivation of the fungus to combat *M. phaseolina* infestation in crops. The antagonistic efficacy of the fungus against *M. phaseolina* is attributable to several antagonistic mechanisms such as nutrient competition, antibiotic production and mycoparasitism (Pastrana *et al.*, 2016). However, the study believes the need for *in-situ* propagation of this antagonistic is rather slow and, possibly inadequately effective to confer a comprehensive protection on the crops throughout the different growth stages. But the study notes that the *T. harzianum* secretes an enzyme *via* the mycoparasitism mechanism. It is a well-known fact that the β -glucosidase produced by this fungus explicitly hydrolyzes the β -linkage of the amorphous β -1, 3-glucan filling material in the chitin-based cell wall of *M. phaseolina* (Gajera *et al.*, 2012). This antagonistic mechanism is well-reported for species of *T. harzianum* whereby the β -glucosidase facilitates the penetration of *T. harzianum* into the cytoplasm of the target pathogenic fungi (Kavitha *et al.*, 2012; Andersen *et al.*, 2016). This interesting antagonistic feature of *T. harzianum* to win the biological war against another competing fungus, i.e. *M. phaseolina* seems agronomically useful and scientifically interesting.

1.2 Problem Statement

In view of the limitations and ineffectiveness of current management strategies to control charcoal rot disease caused by the soil borne nature of *M. phaseolina*, and the environmental hazards associated with the use of pesticides, the search for alternative ecologically benign and sustainable strategy, preferably cost-effective, may prove timely and merits agronomical consideration. Herein, a simple but possibly effective bio-based technique of *in-situ* application of the extracellular lytic enzyme i.e. β -glucosidase secreted by a well-known antagonistic fungus, *T. harzianum* is proposed. The technique is justified based on a well-known fact that the *Trichoderma* spp. such as *T. harzianum* has evolved in such a way to specifically secrete a specific β -glucosidase adapted to hydrolyzing the amorphous β -1, 3-glucan cell wall components in *M. phaseolina*. Driven by this fact, the direct spraying of a formulation containing the growth supernatant of the extracellular β -glucosidase would instantaneously destroy the *M. phaseolina*. Hence, further dissemination of the fungal material by wind, soil or by physical contact could be averted and the charcoal disease is halted.

It is hypothesized the extracellular enzyme β -glucosidase isolated from the highly antagonistic *T. harzianum* T12 strain can be effectively used as the bioactive ingredient over the chemically formulated ones for an environmentally benign fungicide formulation. The study strongly believes the enzyme can be formulated as an effective greener fungicide for *in-situ* protection against charcoal rot disease due to *M. phaseolina* infestation for crops at various growth stages.

Although the direct use of the β -glucosidase would probably be cheaper, a comprehensive characterization of its vulnerability against other possible additives in the formulation may prove necessary for this study. This is to ensure its hydrolytic activity is boosted, hence maximizing the efficacy in inhibiting *M. phaseolina* infestation. Furthermore, the specific use of β -glucosidase from *T. harzianum* as the active ingredient in fungicide formulation for agricultural management of *M.*

phaseolina infestation has not been reported. The feasibility of this proposed technique remains to be seen.

1.3 Objectives of Research

This study highlights upon four main research objectives:

1. To collect, isolate and identify the most pathogenic *M. phaseolina* and most effective *T. harzianum* isolates.
2. To purify and characterize the physico- and biochemical properties of the β -glucosidase.
3. To optimize the growth conditions of *T. harzianum* for maximum activity of the produced β -glucosidase.
4. To optimize the application of the β -glucosidase-loaded formulation for maximum inhibition on *M. phaseolina*.

1.4 Aim of Study

The study aimed to prepare a cheaper and greener fungicide using the crude β -glucosidase from *T. harzianum* as the bioactive ingredient over the toxic chemically formulated fungicide for inhibiting the growth of the phytopathogen *M. phaseolina*.

1.5 Scopes of the Study

This study first isolated and screened for fungal strains of the most effective *T. harzianum* and the most pathogenic *M. phaseolina* done under laboratory settings. These *M. phaseolina* strains were isolated from infected plants and *T. harzianum* isolates were isolated from soil samples from different agricultural locations in the Mazandaran province of Iran. The most pathogenic (most prevalent) *M. phaseolina* was identified using pathogenic variability test. It was necessary for the study to isolate the most virulent strain of *M. phaseolina* as the model. If the *T. harzianum* isolate/ β -glucosidase was effective against the most virulent strain of the fungal pathogen, the efficacy of former to inhibit other less virulent ones was highly possible. Similarly, selection of the most effective *T. harzianum* isolate was based on the most prevalent fungus strain found in the soil. This fungus was identified using, dual culture test, hyper parasitism test, and volatile metabolites production. Both fungi were then identified using morphological and biochemical tests.

Prior to formulation, the β -glucosidase in the growth supernatant of the *T. harzianum* was purified using ammonium sulfate precipitation and dialyzed. The enzyme was sequenced to ascertain its classification as a β -glucosidase and subsequently assessed for physicochemical susceptibilities to environmental conditions and possible additives in the fungicide formulation. The molecular weight of the enzyme was evaluated using SDS-PAGE and the assessed physicochemical properties included optimal pH and temperature as well as the effects of metal ions, surfactants, carbon and nitrogen sources, and inoculum size. Since, this was a novel enzyme, the kinetic parameters of the purified β -glucosidase was also evaluated.

The study went on to optimizing the physical and nutritional parameters for batch cultivation of *T. harzianum* to produce the β -glucosidase using the method of Response Surface Methodology (RSM) by the Box-Behnken Design (BBD). A cheap agro-industrial substrate i.e. banana wastes was used as the carbon source for batch cultivation of the *T. harzianum* to produce crude β -glucosidase. The response of this optimization was the highest activity of the harvested β -glucosidase. This was based

on the largest inhibition zone surrounding a paper disc soaked with β -glucosidase, carried out on potato dextrose assay plates.

The study finally optimized the preparation and application of the β -glucosidase loaded fungicide formulation using soybean plants. The plants were grown in soil infected with *M. phaseolina* and the work was evaluated over a period of 8 weeks under a controlled condition in a greenhouse located within the grounds of the Faculty of Bioscience and Medical Engineering, UTM. The efficacy of the crude β -glucosidase to control charcoal rot disease was assessed based on the lowest plant disease index (PDI). Since this study is a pioneering work using β -glucosidase as the fungicide, RSM was again used to evaluate the effects of multiple factors *viz.* metal ions, surfactants, enzyme concentration and irrigation frequency of the formulation on the response *i.e.* PDI. Consequently, a controlled study comparing the optimum protocol of β -glucosidase, a commonly use pesticide *i.e.* carbendazim and the antagonist *T. harzianum* (Rifai) to result in the lowest PDI in growing soy bean plants sown on *M. phaseolina* infected soil was carried out. Cost assessment on the use of carbendazim and β -glucosidase based on per hectare of cultivated soybean was also performed.

1.6 Significance of the Study

The strategy proposed here is potentially more rapid and sustainable to control the spread of charcoal rot disease due to *M. phaseolina* infestation in crops. The approach used in this study is possibly a more practical and cost-effective means to avert charcoal rot disease in other agronomically relevant crops, too.

1.7 Operational Framework of the Research

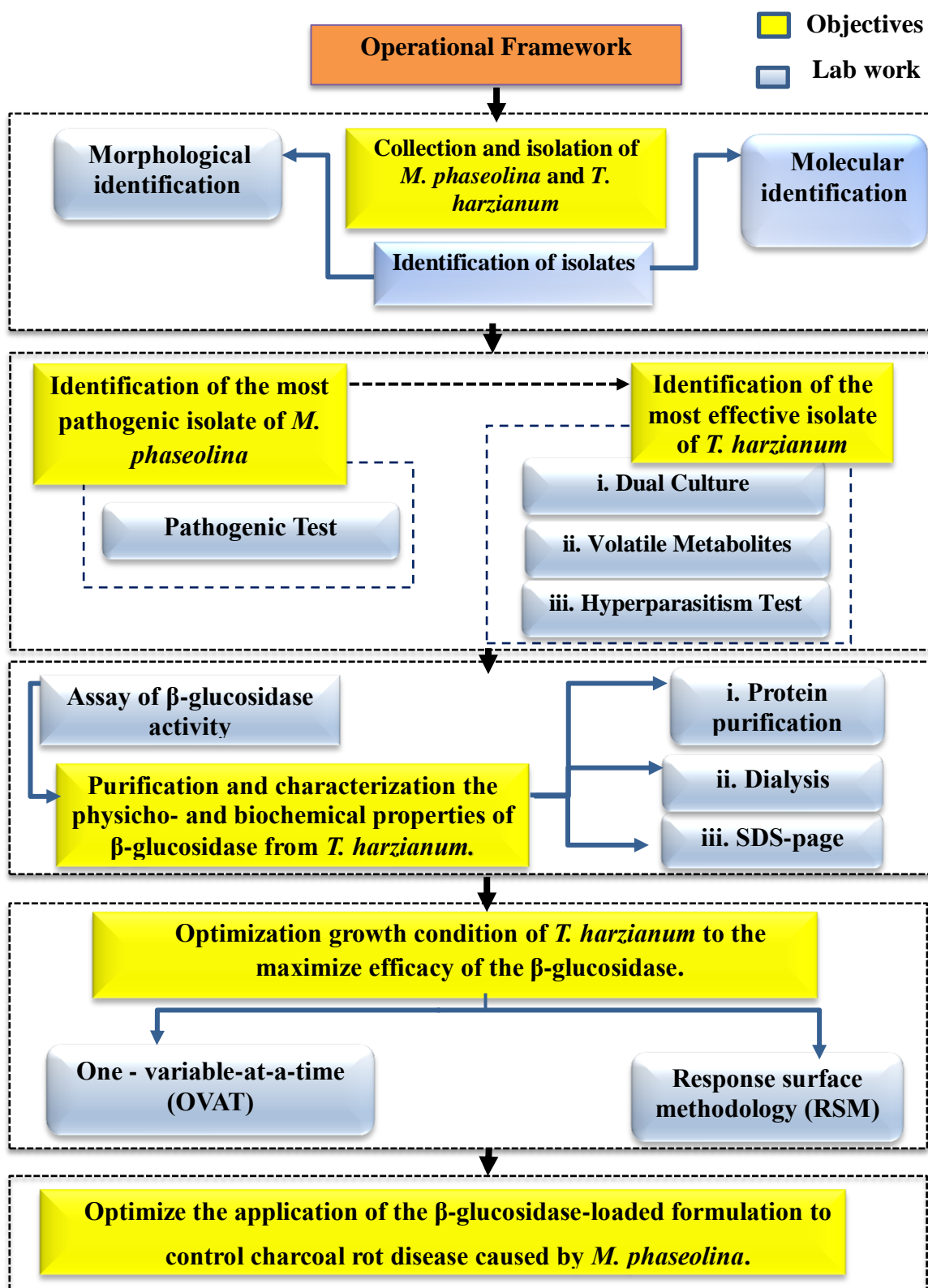


Figure 1.1 Operational framework of the research methodology.

REFERENCES

- Abdel-Fatah, O. M., Hassan, M. M., Elshafei, A. M., Haroun, B. M., Atta, H. M. and Othman, A. M. (2012). Physiological studies on carboxymethyl cellulase formation by *Aspergillus terreus* DSM 826. *Brazilian Journal of Microbiology*. 43(1), 01-11.
- Abdella, A., El-Sayed Mazeed, T., Yang, S. T. and F El-Baz, A. (2014). Production of β -glucosidase by *Aspergillus niger* on wheat bran and glycerol in submerged culture: factorial experimental design and process optimization. *Curent Biotechnology*. 3(2), 197-206.
- Abdulgader Edbeib, M. F., Wahab, R. A. and Huyop, F. (2016). Characterization of an α -haloalkanoic acid-degrading *Pseudomonas aeruginosa* MX1 isolated from contaminated seawater. *Bioremediation Journal*. 20(2), 89-97.
- Abdullah, M. T., Ali, N. Y. and Suleman, P. (2008). Biological control of *Sclerotinia sclerotiorum* (Lib.) de Bary with *Trichoderma harzianum* and *Bacillus amyloliquefaciens*. *Crop Protection*. 27(10), 1354-1359.
- Ab-Majid, A. H., Zahran, Z., Rahim, A. H. A., Ismail, N. A., Rahman, W. A., Zubairi, K. S. M. and Satho, T. (2015). Morphological and molecular characterization of fungus isolated from tropical bed bugs in Northern Peninsular Malaysia, *Cimex hemipterus* (Hemiptera: Cimicidae). *Asian Pacific Journal of Tropical Biomedicine*. 5(9), 707-713.
- Aboshosha, S. S., Attaalla, S. I., El-Korany, A. E. and El-Argawy, E. (2007). Characterization of *Macrophomina phaseolina* isolates affecting sunflower growth in El-Behera governorate, Egypt. *International Journal of Agriculture and Biology*. 9(6), 807-815.
- Adav, S. S., Ravindran, A., Chao, L. T., Tan, L., Singh, S. and Sze, S. K. (2011). Proteomic analysis of pH and strains dependent protein secretion of *Trichoderma reesei*. *Journal of Proteome Research*. 10(10), 4579-4596.

- Adekunle, A. T., Ikotun, T., Florini, D. A. and Cardwell, K. F. (2006). Field evaluation of selected formulations of *Trichoderma* species as seed treatment to control damping-off of cowpea caused by *Macrophomina phaseolina*. *African Journal of Biotechnology*. 5(5), 419-424.
- Afouda, L. C., Schulz, D., Wolf, G. and Wydra, K. (2012). Biological control of *Macrophomina phaseolina* on cowpea (*Vigna unguiculata*) under dry conditions by bacterial antagonists. *International Journal of Biological and Chemical Sciences*. 6(6), 5068-5077.
- Agarwal, V. K. and Sinclair, J. B. (1996). *Principles of seed pathology*. CRC Press. 253-543.
- Ahamed, A. and Vermette, P. (2009). Effect of culture medium composition on *Trichoderma reesei*'s morphology and cellulase production. *Bioresource Technology*. 100(23), 5979-5987.
- Ahmad, S. S. and Dalby, P. A. (2011). Thermodynamic parameters for salt-induced reversible protein precipitation from automated microscale experiments. *Biotechnology and Bioengineering*. 108 (2), 322-332.
- Ait-Lahsen, H., Soler, A., Rey, M., de la Cruz, J., Monte, E. and Llobell, A. (2001). An antifungal exo- α -1, 3-glucanase (AGN13. 1) from the biocontrol fungus *Trichoderma harzianum*. *Applied Environmental Microbiology*. 67(12), 5833-5839.
- Akhtar, S. Shoaib, A., Akhtar, N. and Mehmood, R. (2016). Separate and Combined Effects of *Macrophomina phaseolina* and Copper on Growth, Physiology and Antioxidative Enzymes in *Vigna Mungo* L. *Journal of Animal and Plant Sciences*. 26(5). 1339-1345.
- Ali, M. B., Irshad, M., Anwar, Z., Zafar, M. and Imran, M. (2016). Screening and Statistical Optimization of Physiochemical Parameters for the Production of Xylanases from Agro-Industrial Wastes. *Advances in Enzyme Research*. 4(1), 20-23.
- Al-Jazairi, M., Abou-Ghorra, S., Bakri, Y. and Mustafa, M. (2015). Optimization of β -galactosidase production by response surface methodology using locally isolated *Kluyveromyces marxianus*. *International Food Research Journal*. 22(4), 1361-1367.

- Al-Rajhi, A. M. (2013). Impact of biofertilizer *Trichoderma harzianum* Rifai and the biomarker changes in *Eruca sativa* L. plant grown in metal-polluted soils. *World Applied Sciences Journal*. 22(2), 171-180.
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W. and Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology*. 215(3), 403-410.
- Alvarado-Carrillo, M., Diaz-Franco, A., Delgado-Aguirre, E. and Montes-Garcia, N. (2010). Impact of corn agronomic management on aflatoxin (*Aspergillus flavus*) contamination and charcoal stalk rot (*Macrophomina phaseolina*) incidence. *Tropical and Subtropical Agroecosystems*. 12 (3), 575-582.
- Alvandia, D. G. and Hirooka, Y. (2011). Identification of *Clonostachys* and *Trichoderma* spp. from banana fruit surfaces by cultural, morphological and molecular methods. *Mycology*. 2(2), 109-115.
- Ambrosio, M. M., Dantas, A. C., Martinez-Perez, E., Medeiros, A. C., Nunes, G. H. and Pico, M. B. (2015). Screening a variable germplasm collection of *Cucumis melo* L. for seedling resistance to *Macrophomina phaseolina*. *Euphytica*. 206(2), 287-300.
- Andersen, B., Poulsen, R. and Hansen, G. H. (2016). Cellulolytic and xylanolytic activities of common indoor fungi. *International Biodeterioration and Biodegradation*. 107, 111-116.
- Anderson, P. K., Cunningham, A. A., Patel, N. G., Morales, F. J., Epstein, P. R. and Daszak, P. (2004). Emerging infectious diseases of plants: pathogen pollution, climate change and agrotechnology drivers. *Trends in Ecology and Evolution*. 19(10), 535-544.
- Anees, M., Tronsmo, A., Edel-Hermann, V., Hjeljord, L. G., Heraud, C. and Steinberg, C. (2010). Characterization of field isolates of *Trichoderma* antagonistic against *Rhizoctonia solani*. *Fungal biology*. 114(9), 691-701.
- Arancibia, R. A., Main, J. L. and Clark, C. A. (2013). Sweetpotato tip rot incidence is increased by preharvest applications of ethephon and reduced by curing. *Hort Technology*. 23(3), 288-293.
- Askari, H., Shahbazi, S., Naseripour, T., Moosavi-Nasab, M. and Bakhtiyari, M. (2014). The impact of extracellular enzymes of *Trichoderma viride* and *Trichoderma harzianum* on succinoglycan produced from *Agrobacterium*

- radiobacter*. *International Journal of Agriculture and Crop Sciences*. 7(8), 488-498.
- Aviles, M., Castillo, S., Bascon, J., Zea-Bonilla, T., Martin-Sanchez, P. M. and Perez-Jimenez, R.M. (2008). First report of *Macrophomina phaseolina* causing crown and root rot of strawberry in Spain. *Plant Pathology*. 57(2), 382.
- Babalola, O. O. and Glick, B. R. (2012). The use of microbial inoculants in African agriculture: current practice and future prospects. *Journal of Food, Agriculture and Environment*. 10(3-4), 540-549.
- Bai, H., Wang, H., Sun, J., Irfan, M., Han, M., Huang, Y. and Yang, Q. (2013). Production, purification and characterization of novel beta glucosidase from newly isolated *Penicillium simplicissimum* H-11 in submerged fermentation. *EXCLI Journal*. 12, 528-540.
- Baino, O. M., Salazar, S. M., Ramallo, A. C. and Kirsch Baum, D. S. (2016). First report of *Macrophomina phaseolina* causing strawberry crown and root rot in North Western Argentina. *Journal of Berry Research Sep*. 6(3), 345-354.
- Bas, D. and Boyaci, I. H. (2007). Modeling and optimization I: Usability of response surface methodology. *Journal of food engineering*. 78(3), 836-845.
- Bashour, I. I. and Sayegh, A. H. (2007). Methods of analysis for soils of arid and semi-arid regions. Food and Agriculture Organization (FAO) of the United Nations, Rome. 4,64-65
- Batra, J., Beri, D. and Mishra, S. (2014). Response surface methodology based optimization of β -glucosidase production from *Pichia pastoris*. *Applied Biochemistry and Biotechnology*. 172(1), 380-393.
- Beas-Fernandez, R., De Santiago-De Santiago, A., Hernandez-Delgado, S. and Mayek-Perez, N. (2006). Characterization of Mexican and non-Mexican isolates of *Macrophomina phaseolina* based on morphological characteristics, pathogenicity on bean seeds and endoglucanase genes. *Journal of Plant Pathology*. 88(1), 53-60.
- Bennett, A. J., Bending, G. D., Chandler, D., Hilton, S. and Mills, P. (2012). Meeting the demand for crop production: the challenge of yield decline in crops grown in short rotations. *Biological Reviews*. 87(1), 52-71.

- Berbee, M. L. and Taylor, J. W. (1992). Two ascomycete classes based on fruiting-body characters and ribosomal DNA sequence. *Molecular Biology and Evolution*. 9(2), 278-284.
- Betiku, E., Okunsolawo, S. S., Ajala, S. O. and Odedele, O. S. (2015). Performance evaluation of artificial neural network coupled with generic algorithm and response surface methodology in modeling and optimization of biodiesel production process parameters from shea tree (*Vitellaria paradoxa*) nut butter. *Renewable Energy*. 76, 408-417.
- Bhatti, H. N., Batool, S. and Afzal, N. (2013). Production and Characterization of a Novel β -glucosidase from *Fusarium solani*. *International Journal of Agriculture and Biology*. 1, 1-15.
- Bhiri, F., Chaabouni, S. E., Limam, F., Ghrir, R. and Marzouki, N. (2008). Purification and biochemical characterization of extracellular β -glucosidases from the hypercellulolytic Pol6 mutant of *Penicillium occitanis*. *Applied Biochemistry and Biotechnology*. 149(2), 169-182.
- Bisby, G. R. (1939). *Trichoderma viride* Pers. ex Fries, and notes on Hypocrea. *Transactions of the British Mycological Society*. 23(2), 149-168.
- Bissett, J. (1991). A revision of the genus *Trichoderma*. II. Infrageneric classification. *Canadian Journal of Botany*. 69(11), 2357-2372.
- Biswanger, H. (2014). Enzyme Assays. *Perspective in Science*. 1, 41-55.
- Blanco-Canqui, H. (2010). Energy crops and their implications on soil and environment. *Agronomy Journal*. 102(2), 403-419.
- Box, G. E. and Behnken, D. W. (1960). Some new three level designs for the study of quantitative variables. *Technometrics*. 2(4), 455-475.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*. 72(1-2), 248-254.
- Bressano, M., Giachero, M.L., Luna, C.M. and Ducasse, D.A. (2010). An in vitro method for examining infection of soybean roots by *Macrophomina phaseolina*. *Physiological and Molecular Plant Pathology*. 74, 201-204.
- Cabanillas, H. E. and Jones, W. A. (2009). Effects of temperature and culture media on vegetative growth of an entomopathogenic fungus *Isaria* sp.

- (Hypocreales: Clavicipitaceae) naturally affecting the whitefly, *Bemisia tabaci* in Texas. *Mycopathologia*. 167(5), 263.
- Cai, F., Chen, W., Wei, Z., Pang, G., Li, R., Ran, W. and Shen, Q. (2015). Colonization of *Trichoderma harzianum* strain SQR-T037 on tomato roots and its relationship to plant growth, nutrient availability and soil microflora. *Plant and Soil*. 388(1-2), 337-350.
- Cantarel, B.L., Coutinho, P.M., Rancurel, C., Bernard, T., Lombard, V. and Henrissat B. (2009). The Carbohydrate-Active Enzymes database (CAZy): an expert resource for glycogenomics. *Nucleic Acids Res*. 37, 233-238.
- Cardina, J., Herms, C. P. and Doohan, D. J. (2002). Crop rotation and tillage system effects on weed seedbanks. *Weed Science*. 50(4), 448-460.
- Cardoza, R. E., Malmierca, M. G., Hermosa, M. R., Alexander, N. J., McCormick, S. P., Proctor, R. H. and Gutiérrez, S. (2011). Identification of loci and functional characterization of trichothecene biosynthesis genes in filamentous fungi of the genus *Trichoderma*. *Applied and Environmental Microbiology*. 77(14), 4867-4877.
- Chamorro, M., Miranda, L., Dominguez, P., Medina, J. J., Soria, C., Romero, F. and De los Santos, B. (2015). Evaluation of biosolarization for the control of charcoal rot disease (*Macrophomina phaseolina*) in strawberry. *Crop Protection*. 67, 279-286.
- Chang, K.F., Hwang, S.F., Wang, H.P., Turnbull, G. and Howard, R. (2006). Etiology and biological control of sclerotinia blight of coneflower using *Trichoderma* species. *Plant Pathology Journal*. 5, 15-19.
- Chauhan, B. S., Singh, V. P., Kumar, A. and Johnson, D. E. (2011). Relations of rice seeding rates to crop and weed growth in aerobic rice. *Field Crops Research*. 121(1), 105-115.
- Chauhan, P. S., Bharadwaj, A., Puri, N. and Gupta, N. (2014). Optimization of medium composition for alkali-thermostable mannanase production by *Bacillus nealsonii* PN-11 in submerged fermentation. *International Journal of Current Microbiology and Applied Sciences*. 3(10), 1033-1045.
- Chaverri, P. and Samuels, G. J. (2004). *Hypocrea/Trichoderma* (Ascomycota, Hypocreales, Hypocreaceae): species with green ascospores. *Studies in Mycology*. 48, 1-36.

- Chaverri, P., Branco-Rocha, F., Jaklitsch, W., Gazis, R., Degenkolb, T. and Samuels, G. J. (2015). Systematics of the *Trichoderma harzianum* species complex and the re-identification of commercial biocontrol strains. *Mycologia*. 107(3), 558-590.
- Chen, S. F., Morgan, D., Beede, R. H. and Michailides, T. J. (2015). First report of *Lasiodiplodia theobromae* associated with stem canker of almond in California. *Plant Disease*. 99(12), 1678-1688.
- Chowdhury, S., Basu, A. and Kundu, S. (2014). Green synthesis of protein capped silver nanoparticles from phytopathogenic fungus *Macrophomina phaseolina* (Tassi) Goid with antimicrobial properties against multidrug-resistant bacteria. *Nanoscale Research Letters*. 9(1), 1-11.
- Chua, E. M., Shimeta, J., Nugegoda, D., Morrison, P. D. and Clarke, B. O. (2014). Assimilation of polybrominated diphenyl ethers from microplastics by the marine amphipod, *Allorchestes compressa*. *Environmental Science and Technology*. 48(14), 8127-8134.
- Contreras-Cornejo, H. A., Macias-Rodriguez, L., Cortes-Penagos, C. and Lopez-Bucio, J. (2009). *Trichoderma virens*, a plant beneficial fungus, enhances biomass production and promotes lateral root growth through an auxin-dependent mechanism in *Arabidopsis*. *Plant Physiology*. 149(3), 1579-1592.
- Cornish-Bowden, A. (2014). Current IUBMB recommendations on enzyme nomenclature and kinetics. *Perspectives in Science*. 1(1), 74-87.
- Corpet, F. (1988). Multiple sequence alignment with hierarchical clustering. *Nucleic Acids Research*. 16(22), 10881-10890.
- Costa, S. S., Matos, K. S., Tessmann, D. J., Seixas, C. D. and Pfenning, L. H. (2016). *Fusarium paranaense* sp. nov., a member of the *Fusarium solani* species complex causes root rot on soybean in Brazil. *Fungal Biology*. 120(1), 51-60.
- Crous, P. W., Slippers, B., Wingfield, M. J., Rheeder, J., Marasas, W. F., Philips, A. J. and Groenewald, J. Z. (2006). Phylogenetic lineages in the Botryosphaeriaceae. *Studies in Mycology*. 55, 235-253.
- Crucello, A., Sforça, D. A., Horta, M. A. C., dos Santos, C. A., Viana, A. J. C., Beloti, L. L. and de Souza, A. P. (2015). Analysis of genomic regions of

- Trichoderma harzianum* IOC-3844 related to biomass degradation. *PLoS One*. 10(4), 122.
- Csodes, I., Cseh, A., Taller, J. and Poczai, P. (2012). Genetic diversity and effect of temperature and pH on the growth of *Macrophomina phaseolina* isolates from sunflower fields in Hungary. *Molecular Biology Reports*. 39(3), 3259-3269.
- Danielson, R. M. and Davey, C. B. (1973). Carbon and nitrogen nutrition of *Trichoderma*. *Soil Biology and Biochemistry*. 5(5), 505-515.
- Da-Silva Delabona, P., Pirota, R. D. P. B., Codima, C. A., Tremacoldi, C. R., Rodrigues, A. and Farinas, C. S. (2013). Effect of initial moisture content on two Amazon rainforest *Aspergillus* strains cultivated on agro-industrial residues: Biomass-degrading enzymes production and characterization. *Industrial Crops and Product*. 42, 236-242.
- Daynes, C. N., Zhang, N., Saleeba, J. A. and McGee P.A. (2012). Soil aggregates formed in vitro by saprotrophic *Trichocomaceae* have transient water-stability. *Soil Biol Biochem*. 48,151-161.
- Deka, D., Bhargavi, P., Sharma, A., Goyal, D., Jawed, M. and Goyal, A. (2011). Enhancement of cellulase activity from a new strain of *Bacillus subtilis* by medium optimization and analysis with various cellulosic substrates. *Enzyme Research*. 2011(2011),1-8.
- Dexter, A. R. (1988). Advances in characterization of soil structure. *Soil and Tillage Research*. 11(3-4), 199-238.
- Dhawane, S.H., Kumar. T. and Halder. G. (2015). Central composite design approach towards optimization of flamboyant pods derived steam activated carbon for its use as heterogeneous catalyst in transesterification of *Hevea brasiliensis* oil. *Energy Conversion and Management*. 100, 277-287.
- Dhingra, O. D. and Sinclair, J. B. (1978). Biology and pathology of *Macrophomina phaseolina*. Universidade Federal de Vicosa. Minas Gerais. 14, 166-279.
- Domingues, F.C., Queiroz, J.A., Cabral, J.M. and Fonseca, L.P. (2000). The influence of culture conditions on mycelial structure and cellulase production by *Trichoderma reesei* Rut C-30. *Enzyme Microb Technol*. 26,394-401.

- Druzhinina, I. S., Kopchinskiy, A. G., Komon, M., Bissett, J., Szakacs, G. and Kubicek, C. P. (2005). An oligonucleotide barcode for species identification in *Trichoderma* and *Hypocrea*. *Fungal Genetics and Biology*. 42(10), 813-828.
- Dube, M. P. (2000). Disorders of glucose metabolism in patients infected with human immunodeficiency virus. *Clinical Infectious Diseases*. 31(6), 1467-1475.
- Dubey, R. C., Kumar, H. and Pandey, R. R. (2009). Fungi toxic effect of neem extracts on growth and sclerotial survival of *Macrophomina phaseolina* in vitro. *Journal of American Science*. 5, 17-24.
- El-Fadaly, H. M., El-Kadi, S. M., Hamad, M. N. and Habib, A. A. (2015). Isolation and Identification of Egyptian Ras Cheese (Romy) Contaminating Fungi during Ripening Period. *Journal of Microbiology Research*. 5(1), 1-10.
- El-Fiki, A. I. I., Mohamed, F. G., El-Deeb, A. A. and Khalifa, M. M. A. (2004). Some applicable methods for controlling sesame charcoal rot disease (*Macrophomina phaseolina*) under greenhouse conditions. *Egyptian Journal of Phytopathology*. 32(1-2), 87-101.
- El-Hawary, F. I. and Mostafa, Y. S. (2001). Factors affecting cellulase production by *Trichoderma koningii*. *Acta Alimentaria*. 30(1), 3-13.
- Ellis, M. L., Broders, K. D., Paul, P. A. and Dorrance, A. E. (2011). Infection of soybean seed by *Fusarium graminearum* and effect of seed treatments on disease under controlled conditions. *Plant Disease*. 95(4), 401-407.
- Erskine, W., Muehlbauer, F. J. and Short, R. W. (1990). Stages of development in lentil. *Experimental Agriculture*. 26(03), 297-302.
- Etebarian, H. R. (2006). Evaluation of *Trichoderma* isolates for biological control of charcoal stem rot in melon caused by *Macrophomina phaseolina*. *Journal of Agricultural Science and Technology*. 8, 243-250.
- Falconer, R. E., Otten, W. and White, N. A. (2015). Chapter One-Toward Modeling the Resistance and Resilience of “Below-ground” Fungal Communities: A Mechanistic and Trait-Based Approach. *Advances in Applied Microbiology*. 93, 1-44.
- Fernandez, F. G. and Hoeft, R. G. (2009). Managing soil pH and crop nutrients. *Illinois Agronomy Handbook*. 8, 91-112.

- Fernando, W. D., Ramarathnam, R., Krishnamoorthy, A. S. and Savchuk, S. C. (2005). Identification and use of potential bacterial organic antifungal volatiles in biocontrol. *Soil Biology and Biochemistry*. 37(5), 955-964.
- Ferreira, S. C., Bruns, R. E., Ferreira, H. S., Matos, G. D., David, J. M., Brandao, G. C. and Dos Santos, W. N. L. (2007). Box-Behnken design: an alternative for the optimization of analytical methods. *Analytica Chimica Acta*. 597(2), 179-186.
- Fiers, M., Edel-Hermann, V., Chatot, C., Le Hingrat, Y., Alabouvette, C. and Steinberg, C. (2012). Potato soil-borne diseases. A review. *Agronomy for Sustainable Development*. 32(1), 93-132.
- Froeliger, E. H., Carpenter, B. E. and Froeliger, E. (1996). NUT1, a major nitrogen regulatory gene in *Magnaporthe grisea*, is dispensable for pathogenicity. *Molecular and General Genetics MGG*. 251(6), 647-656.
- Gaetan, S.A., Fernande, L. and Madia, M. (2006). Occurrence of charcoal rot caused by *Macrophomina phaseolina* on canola in argentina. *Plant Disease*. 90, 524-524.
- Gaige, A. R., Ayella, A. and Shuai, B. (2010). Methyl jasmonate and ethylene induce partial resistance in *Medicago truncatula* against the charcoal rot pathogen *Macrophomina phaseolina*. *Physiological and Molecular Plant Pathology*. 74(5), 412-418.
- Gajera, H. P., Bambarolia, R. P., Patel, S.V., Khatrani, T. J. and Goalkiya, B. A. (2012). Antagonism of *Trichoderma* spp. against *Macrophomina phaseolina*: Evaluation of coiling and cell wall degrading enzymatic activities. *Plant Pathology and Microbiology*. 3(7), 1-7.
- Gams, W. and Bissett, J. (1998). Morphology and identification of *Trichoderma*. *Trichoderma and Gliocladium*. 1, 3-34.
- Garcia, N. F. L., Santos, F. R. D. S., Gonçalves, F. A., Paz, M. F. D., Fonseca, G. G. and Leite, R. S. R. (2015). Production of β -glucosidase on solid-state fermentation by *Lichtheimia ramosa* in agroindustrial residues: Characterization and catalytic properties of the enzymatic extract. *Electronic Journal of Biotechnology*. 18(4), 314-319.
- Gardes, M., White, T. J., Fortin, J. A., Bruns, T. D. and Taylor, J. W. (1991). Identification of indigenous and introduced symbiotic fungi in

- ectomycorrhizae by amplification of nuclear and mitochondrial ribosomal DNA. *Canadian Journal of Botany*. 69(1), 180-190.
- Gautam, S. P., Bundela, P. S., Pandey, A. K., Awasthi, M. K. and Sarsaiya, S. (2010). Optimization of the medium for the production of cellulase by the *Trichoderma viride* using submerged fermentation. *International Journal of Environmental Sciences*. 1(4), 656.
- Gemell, L. G., Hartley, E. J. and Herridge, D. F. (2005). Point-of-sale evaluation of preinoculated and custom-inoculated pasture legume seed. *Animal Production Science*. 45(3), 161-169.
- Gent, D. H., Mahaffee, W. F., McRoberts, N. and Pfender, W. F. (2013). The use and role of predictive systems in disease management. *Annual Review of Phytopathology*. 51, 267-289.
- George, E., Tamerler, C., Martinez, A., Martinez, M. J. and Keshavarz, T. (1999). Influence of growth medium composition on the lipolytic enzyme activity of *Ophiostoma piliferum* (Cartapip™). *Journal of Chemical Technology and Biotechnology*. 74(2), 137-140.
- Gerbore, J., Benhamou, N., Vallance, J., Le Floch, G., Grizard, D., Regnault-Roger, C. and Rey, P. (2014). Biological control of plant pathogens: advantages and limitations seen through the case study of *Pythium oligandrum*. *Environmental Science and Pollution Research*. 21(7), 4847-4860.
- Ghangaokar, N. M. and Kshirsagar, A. D. (2013). Study of seed borne fungi of different legumes. *Trends in Life Sciences*. 2(1), 32-35.
- Ghazanfar, M. U., Hussain, M., Hamid, M. I. and Ansari, S. U. (2016). Utilization Of Biological Control Agents For The Management Of Postharvest Pathogens Of Tomato. *Pakistan Journal of Botany*. 48(5), 2093-2100.
- Ghosh, T., Mukherji, N. and Basak, M. (1964). On the occurrence of a new species of *Orbilia* Fr. *Jute Bulletin*. 27, 134-141.
- Goujon, M., McWilliam, H., Li, W., Valentin, F., Squizzato, S., Paern, J. and Lopez, R. (2010). A new bioinformatics analysis tools framework at EMBL-EBI. *Nucleic Acids Research*. 38(2), 695-699.
- Goyari, S., Devi, S. S., Kalita, M. C. and Talukdar, N. C. (2014). Population, diversity and characteristics of cellulolytic microorganisms from the Indo-Burma Biodiversity hotspot. *Springer Plus*. 3(1), 1-9.

- Graber, E. R., Frenkel, O., Jaiswal, A. K. and Elad, Y. (2014). How may biochar influence severity of diseases caused by soilborne pathogens? *Carbon Management*. 5(2), 169-183.
- Gray, K. A., Zhao, L. and Emptage, M. (2006). Bioethanol. *Current Opinion in Chemical Biology*. 10(2), 141-146.
- Groenewald, J. Z. and Crous, P. W. (2014). Genetic diversity in *Macrophomina phaseolina*, the causal agent of charcoal rot. *Phytopathologia Mediterranea*. 53(2), 250-268.
- Gupta, G. K., Sharma, S. K. and Ramteke, R. (2012). Biology, epidemiology and management of the pathogenic fungus *Macrophomina phaseolina* (Tassi) Goid with special reference to charcoal rot of soybean (*Glycine max* (L.) Merrill). *Journal of Phytopathology*. 160(4), 167-180.
- Gupta, P., Chakraborty, D. and Mittal, R. K. (2015). Anti-fungal activity of medicinal plants leaf extracts on growth of *Macrophomina phaseolina*. *Agricultural Science Digest-A Research Journal*. 35(3), 211-214.
- Haaland, P. D. (1989). Experimental design in biotechnology (Vol. 105). *CRC Press*. 105, 55-74
- Haddadin, M. S., Haddadin, J., Arabiyat, O. I. and Hattar, B. (2009). Biological conversion of olive pomace into compost by using *Trichoderma harzianum* and *Phanerochaete chrysosporium*. *Bioresource Technology*. 100(20), 4773-4782.
- Hallsworth, J.E. and Magan, N. (1996) Culture age, temperature, and pH affect the polyol and trehalose contents of fungal propagules. *Appl Environ Microbiol*. 62, 2435-2442.
- Harighi, M. J., Zamani, M. R. and Motallebi, M. (2007). Evaluation of antifungal activity of purified chitinase 42 from *Trichoderma atroviride* PTCC5220. *Biotechnology*. 6(1), 28-33.
- Harman, G. E., Howell, C. R., Viterbo, A., Chet, I. and Lorito, M. (2004). *Trichoderma* species-opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*. 2(1), 43-56.
- Hartman, G. L., Bowen, C. R., Haudenschild, J. S., Fox, C. M., Cary, T. R. and Diers, B. W. (2015). Evaluation of disease and pest damage on soybean cultivars released from 1923 through 2008 under field conditions in Central Illinois. *Agronomy Journal*. 107(6), 2373-2380.

- Hartman, G. L., Pawlowski, M. L., Herman, T. K. and Eastburn, D. (2016). Organically grown soybean production in the USA: Constraints and management of pathogens and insect pests. *Agronomy*. 6(1), 1-18.
- Hartman, G. L., West, E. D. and Herman, T. K. (2011). Crops that feed the World 2. Soybean-worldwide production, use, and constraints caused by pathogens and pests. *Food Security*. 3(1), 5-17.
- Hassan, N., Elsharkawy, M. M., Shimizu, M. and Hyakumachi, M. (2014). Control of root rot and wilt disease of roselle under field conditions. *Mycobiology*. 42(4), 376-384.
- Helali, I., Ferchichi, S., Maaouia, A., Aouni, M. and Harizi, H. (2016). Modulation of macrophage functionality induced in vitro by chlorpyrifos and carbendazim pesticides. *Journal of Immunotoxicology*. 13(5), 745-750.
- Hermosa, R., Rubio, M. B., Cardoza, R. E., Nicolas, C., Monte, E. and Gutierrez, S. (2013). The contribution of *Trichoderma* to balancing the costs of plant growth and defense. *International Microbiology*. 16(2), 69-80.
- Hermoso, J., Pignol, D., Kerfelec, B., Crenon, I., Chapus, C. and Fontecilla-Camps, J. C. (1996). Lipase activation by nonionic detergents the crystal structure of the porcine lipase-colipase-tetraethylene glycol monoethyl ether complex. *Journal of Biological Chemistry*. 271(30), 18007-18016.
- Heydari, A. and Pessarakli, M. (2010). A review on biological control of fungal plant pathogens using microbial antagonists. *Journal of Biological Sciences*. 10(4), 273-290.
- Hmad, I. B., Abdeljalil, S., Saibi, W., Amouri, B. and Gargouri, A. (2014). Medium initial pH and carbon source stimulate differential alkaline cellulase time course production in *Stachybotrys microspora*. *Applied Biochemistry and Biotechnology*. 172(5), 2640-2649.
- Holliday, P. and Punithalingam, E. (1970). *Macrophomina phaseolina*. Descriptions of Fungi and Bacteria. *IMI Descriptions of Fungi and Bacteria*. 275,1-28.
- Hossain, M. T., Hossain, S. M. M., Bakr, M. K., Rahman, A. M. and Uddin, S. N. (2010). Survey on major diseases of vegetable and fruit crops in Chittagong region. *Bangladesh Journal of Agricultural Research*. 35(3), 423-429.

- Hymowitz, T. and Bernard, R. L. (1991). Origin of the soybean and germplasm introduction and development in North America. Use of Plant Introductions in Cultivar Development Part. 1, 147-164.
- Ijaz, S., Sadaqat, H. A. and Khan, M. N. (2013). A review of the impact of charcoal rot (*Macrophomina phaseolina*) on sunflower. *The Journal of Agricultural Science*. 151(02), 222-227.
- Iqbal, H. M. N., Asgher, M. and Bhatti, H. N. (2011). Optimization of physical and nutritional factors for synthesis of lignin degrading enzymes by a novel strain of *Trametes versicolor*. *Bio Resources*. 6(2), 1273-1287.
- Iqbal, S., Nguyen, T. H., Nguyen, T. T., Maischberger, T. and Haltrich, D. (2010). β -Galactosidase from *Lactobacillus plantarum* WCFS1: biochemical characterization and formation of prebiotic galacto-oligosaccharides. *Carbohydrate Research*. 345(10), 1408-1416.
- Iqbal, U. and Mukhtar, T. (2014). Morphological and pathogenic variability among *Macrophomina phaseolina* isolates associated with mungbean (*Vigna radiata* L.) Wilczek from Pakistan. *The Scientific World Journal*. 14,1-9.
- Iqbal, U., Mukhtar, T. and Iqbal, S. M. (2014). *In vitro* and *in vivo* evaluation of antifungal activities of some antagonistic plants against charcoal rot causing fungus, *Macrophomina phaseolina*. *Pakistan Journal of Agricultural Sciences*. 51(3), 691-696.
- Iriarte, F., Roskopf, E., Hilf, M., McCollum, G., Albano, J. and Adkins, S. (2007). First report of *Macrophomina phaseolina* causing leaf and stem blight of tropical soda apple in Florida. *Plant Health Progress*. 10, 1-3.
- Irshad, M., Anwar, Z., Ramzan, M., Mahmood, Z. and Nawaz, H. (2013). Characterization of purified β -glucosidase produced from *Trichoderma viride* through bio-processing of orange peel waste. *Advances in Bioscience and Biotechnology*. 4(10), 941-944.
- Isah, A. A., Mahat, N. A., Jamalis, J., Attan, N., Zakaria, I. I., Huyop, F. and Wahab, R. A. (2016). Synthesis of geranyl propionate in a solvent-free medium using *Rhizomucor miehei* lipase covalently immobilized on chitosan-graphene oxide beads. *Preparative Biochemistry and Biotechnology*. 47(2)1-12.
- Islam, M. S., Haque, M. S., Islam, M. M., Emdad, E. M., Halim, A., Hossen, Q. M. M. and Alam, M. M. (2012). Tools to kill: genome of one of the most

- destructive plant pathogenic fungi *Macrophomina phaseolina*. *Bmc Genomics*. 13 (1), 493.
- Jabbour, D., Klippel, B. and Antranikian, G. (2012). A novel thermostable and glucose-tolerant β -glucosidase from *Fervidobacterium islandicum*. *Applied Microbiology and Biotechnology*. 93, 1947-1956.
- Jabeen, K., Javaid, A., Ahmad, E. and Athar, M. (2011). Antifungal compounds from *Melia azedarach* leaves for management of *Ascochyta rabiei*, the cause of chickpea blight. *Natural Product Research*. 25(3), 264-276.
- Jaspers, M. V., Seyb, A. M., Trought, M. C. T. and Balasubramaniam, R. (2013). Overwintering grapevine debris as an important source of *Botrytis cinerea* inoculum. *Plant Pathology*. 62(1), 130-138.
- Jeya, M., Joo, A. R., Lee, K. M., Tiwari, M. K., Lee, K. M., Kim, S. H. and Lee, J. K. (2010). Characterization of β -glucosidase from a strain of *Penicillium purpurogenum* KJS506. *Applied Microbiology and Biotechnology*. 86(5), 1473-1484.
- Jimenez, D. R. C. (2011). Influence of soils, nutrition, and water relations upon charcoal rot disease processes in Kansas (Doctoral dissertation, Kansas State University).1-26.
- Jimenez-Diaz, R. M., Blanco-Lopez, M. A. and Sackston, W. E. (1983). Incidence and distribution of charcoal rot of sunflower caused by *Macrophomina phaseolina* in Spain. *Plant Disease*. 67(9), 1033-1036.
- Job, J., Sukumaran, R.K. and Jayachandran, K. (2010). Production of a highly glucose tolerant β -glucosidase by *Paecilomyces variotii* MG3: optimization of fermentation conditions using Plackett-Burman and Box-Behnken experimental designs. *World Journal Microbiol Biotechnol*. 26(8),1385-1391.
- Jordaan, E. and Van der Waals, J. (2016). With drought comes charcoal rot: agronomy. *Oilseeds Focus*. 2(3), 6-7.
- Karboune, S., Geraert, P. A. and Kermasha, S. (2008). Characterization of selected cellulolytic activities of multi-enzymatic complex system from *Penicillium funiculosum*. *Journal of Agriculture and Food Chemistry*. 56(3), 903-909.
- Karnchanatat, A., Petsom, A., Sangvanich, P., Piaphukiew, J., Whalley, A. J., Reynolds, C. D. and Sihanonth, P. (2007). Purification and biochemical characterization of an extracellular β -glucosidase from the wood-decaying

- fungus *Daldinia eschscholzii* (Ehrenb.: Fr.) Rehm. *FEMS Microbiology Letters*. 270(1), 162-170.
- Kaur, J., Chadha, B. S., Kumar, B. A., Kaur, G. and Saini, H. S. (2007). Purification and characterization of β -glucosidase from *Melanocarpus* sp. MTCC 3922. *Electronic Journal of Biotechnology*. 10(2), 260-270.
- Kaur, S., Dhillon, G. S., Brar, S. K., Vallad, G. E., Chand, R. and Chauhan, V. B. (2012). Emerging phytopathogen *Macrophomina phaseolina*: biology, economic importance and current diagnostic trends. *Critical Reviews in Microbiology*. 38(2), 136-151.
- Kavitha, T., Gopalan, A. I., Lee, K. P. and Park, S. Y. (2012). Glucose sensing, photocatalytic and antibacterial properties of grapheme-ZnO nanoparticle hybrids. *Carbon*. 50(8), 2994-3000.
- Keswani, C., Mishra, S., Sarma, B. K., Singh, S. P. and Singh, H. B. (2014). Unraveling the efficient applications of secondary metabolites of various *Trichoderma* spp. *Applied Microbiology and Biotechnology*. 98(2), 533-544.
- Khaledi, N., Taheri, P. and Tarighi, S. (2015). Antifungal activity of various essential oils against *Rhizoctonia solani* and *Macrophomina phaseolina* as major bean pathogens. *Journal of Applied Microbiology*. 118 (3), 704-717.
- Khan, N., Jadhav, S. and Rathod, V. K. (2016). Enzymatic synthesis of n-butyl palmitate in a solvent-free system: RSM optimization and kinetic studies. *Biocatalysis and Biotransformation*. 34(3), 99-109.
- Khangura, R. and Aberra, M. (2009). First report of charcoal rot on canola caused by *Macrophomina phaseolina* in Western Australia. *Plant Disease*. 93, 666-667.
- Klein, D. and Eveleigh, D. E. (2002). Ecology of *Trichoderma*. In: Kubicek, C. P. and Harman, G. E. (eds.). *Trichoderma and Gliocladium: Basic Biology, Taxonomy and Genetics*. Taylor and Francis Ltd. 57-69.
- Kocher, G., Kalra, K. and Banta, G. (2008). Optimization of cellulase production by submerged fermentation of rice straw by *Trichoderma harzianum* Rut-C 8230. *International Journal of Medical Microbiology*. 5(2)1-7.
- Kubicek, C. P., Mach, R. L., Peterbauer, C. K. and Lorito, M. (2001). *Trichoderma*: from genes to biocontrol. *Journal of Plant Pathology*. 1, 11-23.

- Kumar, A., Kumar, S., Kumar, S. and Kumar, B. (2016). Diseases of Chickpea Crop and Their Management. In Center Crop Diseases and Their Management: Integrated Approaches *Apple Academic Press*. 27, 39-55.
- Kumar, S. (2016). Diseases of Soybean and Their Management. In Center Crop Diseases and Their Management: Integrated Approaches . *Apple Academic Press*. 27, 95-125
- Kusumawati, D. E., Hadiastono, T. and Aini, L. Q. (2012). Leaf Extract of *Mirabilis jalapa* L. Induced Defense of Tomato Plant (*Lycopersicon esculentum* Mill.) Against Cucumber Mosaic Virus (CMV) Infection. *Journal of Tropical Plant Protection*. 1(1), 46-51.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*. 227, 680-685.
- Landa, B.B., Navas-Cortes, J.A. and Jimenez-Diaz, R.M. (2012). Biological control of *Macrophomina phaseolina* on cowpea (*Vigna unguiculata*). *International Journal of Biological and Chemical Sciences*. 94, 946-960.
- Landmeyer, J. E. (2012). Plant and Groundwater Interactions Under Pristine Conditions. In Introduction to Phytoremediation of Contaminated Groundwater. *Springer Netherlands*. 115-127.
- Larralde-Corona, C. P., Santiago-Mena, M. R., Sifuentes-Rincon, A. M., Rodriguez-Luna, I. C., Rodriguez-Perez, M. A., Shirai, K. and Narvaez-Zapata, J. A. (2008). Biocontrol potential and polyphasic characterization of novel native *Trichoderma* strains against *Macrophomina phaseolina* isolated from sorghum and common bean. *Applied Microbiology and biotechnology*. 80(1), 167-177.
- Latge, J. P. (2010). Tasting the fungal cell wall. *Cellular Microbiology*. 12(7), 863-872.
- Lee, H., Lee, Y. M., Heo, Y. M., Hong, J. H., Jang, S., Ahn, B. J. and Kim, J. J. (2017). Optimization of Fungal Enzyme Production by *Trichoderma harzianum* KUC1716 through Surfactant-Induced Morphological Changes. *Mycobiology*. 45(1), 48-51.
- Li, S., Hartman, G. L. and Boykin, D. L. (2010). Aggressiveness of *Phomopsis longicolla* and other *Phomopsis* spp. on soybean. *Plant Disease*. 94(8), 1035-1040.

- Li, X. H., Zhang, P., Wang, M. X., Zhou, F., Malik, F. A., Yang, H. J. and Miao, Y. G. (2011). Expression of *Trichoderma aviride* endoglucanase III in the larvae of silkworm, *Bombyx mori* L. and characteristic analysis of the recombinant protein. *Molecular Biology Reports*. 38(6), 3897-3902.
- Lieberman, R. L., Wustman, B. A., Huertas, P., Powe Jr, A. C., Pine, C. W., Khanna, R. and Petsko, G. A. (2007). Structure of acid [beta]-glucosidase with pharmacological chaperone provides insight into Gaucher disease. *Nature Chemical Biology*. 3(2), 101-107.
- Lieckfeldt, E., Kuhls, K. and Muthumeenakshi, S. (2002). Molecular taxonomy of *Trichoderma* and *Gliocladium* and their teleomorphs. *Trichoderma and Gliocladium*. 1, 35-56.
- Liu, K. H., Yeh, Y. L. and Shen, W. C. (2011). Fast preparation of fungal DNA for PCR screening. *Journal of Microbiological Methods*. 85(2), 170-172.
- Liu, W., Yin, P., Liu, X. and Qu, R. (2014). Design of an effective bifunctional catalyst organotriphosphonic acid-functionalized ferric alginate (ATMP-FA) and optimization by Box–Behnken model for biodiesel esterification synthesis of oleic acid over ATMP-FA. *Bioresource Technology*. 173, 266-271.
- Lopes, F. A. C., Steindorff, A. S., Geraldine, A. M., Brandao, R. S., Monteiro, V. N., Júnior, M. L. and Silva, R. N. (2012). Biochemical and metabolic profiles of *Trichoderma* strains isolated from common bean crops in the Brazilian Cerrado, and potential antagonism against *Sclerotinia sclerotiorum*. *Fungal Biology*. 116(7), 815-824.
- Lorito, M., Woo, S. L., Harman, G. E. and Monte, E. (2010). Translational research on *Trichoderma*: from momics to the field. *Annual Review of Phytopathology*. 48, 395-417.
- Lopez, M.R., Ros, M. and Pascual, J.A., (2011). Mycoparasitism-related genes expression of *Trichoderma harzianum* isolates to evaluate their efficacy as biocontrol agent. *Biological Control* .56, 59-66.
- Lynd, L. R., Weimer, P. J., Van Zyl, W. H. and Pretorius, I. S. (2002). Microbial cellulose utilization: fundamentals and biotechnology. *Microbiology and Molecular Biology Reviews*. 66(3), 506-577.

- Ma, J., Hill, C. B. and Hartman, G. L. (2010). Production of *Macrophomina phaseolina* conidia by multiple soybean isolates in culture. *Plant Disease*. 94(9), 1088-1092.
- Ma, S. J., Leng, B., Xu, X. Q., Zhu, X. Z., Shi, Y., Tao, Y. M. and Chen, Q. X. (2011). Purification and characterization of b-1, 4-glucosidase from *Aspergillus glaucus*. *African Journal of Biotechnology*. 10(84), 19607-19614.
- Maghari, B. M. and Ardekani, A. M. (2011). Genetically modified foods and social concerns. *Avicenna Journal of Medical Biotechnology*. 3(3), 109-117.
- Mahajan, P. M., Desai, K. M. and Lele, S. S. (2012). Production of cell membrane-bound α - and β -glucosidase by *Lactobacillus acidophilus*. *Food and Bioprocess Technology*. 5(2), 706-718.
- Mahapatra, S., Vickram, A. S., Sridharan, T. B., Parameswari, R. and Pathy, M. R. (2016). Screening, production, optimization and characterization of β -glucosidase using microbes from shellfish waste. *3 Biotech*. 6(213),2-10.
- Maki, M., Leung, K. T. and Qin, W. (2009). The prospects of cellulase-producing bacteria for the bioconversion of lignocellulosic biomass. *International Journal of Biological Sciences*. 5(5), 500-516.
- Malik, G. and Dawar, S. (2003). Biological control of root infecting fungi with *Trichoderma harzianum*. *Pakistan Journal of Agricultural Sciences*. 35, 971-975.
- Manan, F. M. A., Rahman, I. N. A., Marzuki, N. H. C., Mahat, N. A., Huyop, F. and Wahab, R. A. (2016). Statistical modelling of eugenol benzoate synthesis using *Rhizomucor miehei* lipase reinforced nanobioconjugates. *Process Biochemistry*. 51(2), 249-262.
- Manici, L. M., Bregaglio, S., Fumagalli, D. and Donatelli, M. (2014). Modelling soil borne fungal pathogens of arable crops under climate change. *International Journal of Biometeorology*. 58(10), 2071-2083.
- Manici, L. M., Caputo, F. and Cerato, C. (1992). Pathogenic and biological variability of *Macrophomina phaseolina* (Tassi) Goid. isolates in different areas of sunflower cultivation in Italy. *Sunflower Conf*. 1, 779-784.
- Marco, J. L. D. and Felix, C. R. (2007). Purification and characterization of a beta-Glucanase produced by *Trichoderma harzianum* showing biocontrol potential. *Brazilian Archives of Biology and Technology*. 50(1), 21-29.

- Martins, I., Garcia, H., Varela, A., Nunez, O., Planchon, S., Galceran, M. T. and Pereira, C. S. (2014). Investigating *Aspergillus nidulans* secretome during colonisation of cork cell walls. *Journal of Proteomics*. 98, 175-188.
- Marzuki, N. H. C., Mahat, N. A., Huyop, F., Aboul-Enein, H. Y. and Wahab, R. A. (2015). Sustainable production of the emulsifier methyl oleate by *Candida rugosa* lipase nanoconjugates. *Food Bioproducts and Processing*. 96, 211-220.
- Mateos, S. E., Cervantes, C. A. M., Zenteno, E., Slomianny, M. C., Alpuche, J., Hernandez-Cruz, P. and Mayoral, L. P. C. (2015). Purification and partial characterization of β -glucosidase in chayote (*Sechium edule*). *Molecules*. 20 (10), 19372-19392.
- Mayek-Perez, N., Garcia-Espinosa, R., Lopez-CastaNeda, C., Acosta-Gallegos, J. A. and Simpson, J. (2002). Water relations, histopathology and growth of common bean (*Phaseolus vulgaris* L.) during pathogenesis of *Macrophomina phaseolina* under drought stress. *Physiological and Molecular Plant Pathology*. 60(4), 185-195.
- Mbarga, J.B., Hoopen, G.M.T., kuate, J., Adiobo, A., Ngonkeu, M.E.L., Ambang, Z., Akoa, A., Tondje, P.R. and Begoude, B.A.D., (2012). *Trichoderma asperellum*: a potential biocontrol agent for *Pythium myriotylum*, causal agent of cocoyam (*Xanthosoma sagittifolium*) root rot disease in Cameroon. *Crop Protection*. 36, 18-22.
- McCain, A. H. and Smith, R. S. (1972). Quantitative assay of *Macrophomina phaseoli* from soil. *Phytopathology*. (62), 1062-1980.
- Mc-Fadden, A. G. and Sutton, J. C. (1975). Relationship of population of *Trichoderma* spp. in soil to disease in maize. *Canadian Journal of Plant Science*. 55(2), 579-586.
- Medic, J., Atkinson, C. and Hurburgh, C. R. (2014). Current knowledge in soybean composition. *Journal of the American Oil Chemists Society*. 91(3), 363-384.
- Meena, A. K. and Meena, A. K. (2016). Characterization and antagonistic effect of isolated *Trichoderma* sp. against pathogens under clusterbean (*Cyamopsis tetragonoloba* L.). *Indian Journal Of Agricultural Research*. 50 (3) 249-253.

- Mengistu, A., Reddy, K. N., Zablotowicz, R. M. and Wrather, A. J. (2009). Propagule densities of *Macrophomina phaseolina* in soybean tissue and soil as affected by tillage, cover crop, and herbicide. *Plant Health Progress. Online Publication.* 30(10), 1-12.
- Mergel, O., Gelissen, A. P., Wünnemann, P., Boker, A., Simon, U. and Plamper, F. A. (2014). Selective packaging of ferricyanide within thermoresponsive microgels. *The Journal of Physical Chemistry.* 118(45), 26199-26211.
- Mihail, J.D. (1992). *Macrophomina*. In Methods for research on soilborne phytopathogenic fungi. *American Phytopathological Society.* 134-136.
- Mikiashvili, N., Elisashvili, V., Wasser, S. and Nevo, E. (2005). Carbon and nitrogen sources influence the ligninolytic enzyme activity of *Trametes versicolor*. *Biotechnology Letters.* 27(13), 955-959.
- Mirkhani, F. and Alaei, H. (2015). Species diversity of indigenous *Trichoderma* from alkaline pistachio soils in Iran. *Mycologia Iranica.* 2(1), 22-37.
- Mohamad, N. R., Buang, N. A., Mahat, N. A., Lok, Y. Y., Huyop, F., Aboul-Enein, H. Y. and Wahab, R. A. (2015). A facile enzymatic synthesis of geranyl propionate by physically adsorbed *Candida rugosa* lipase onto multi-walled carbon nanotubes. *Enzyme and Microbial Technology.* 72, 49-55.
- Mokhtar, H. and Dehimat, L. (2015). *In vitro* and *In vivo* Efficiency of *Trichoderma harzianum* against Phoma and Glocladium Soft Rot Occurred on Tomato Fruits (*Lycopersicon esculentum*). *International Journal of Current Microbiology and Applied.* 4(8), 141-147.
- Monis, P. T., Caccio, S. M. and Thompson, R. A. (2009). Variation in *Giardia*: towards a taxonomic revision of the genus. *Trends in Parasitology.* 25(2), 93-100.
- Monte, E. and Llobell, A. (2003). *Trichoderma* in organic agriculture. In *Proceeding V World Avocado Congress.* 19, 725-733.
- Montealegre, J., Valderrama, L., Herrera, R., Besoain, X. and Perez, L. M. (2009). Biocontrol capacity of wild and mutant *Trichoderma harzianum* (Rifai) strains on *Rhizoctonia solani* 618: effect of temperature and soil type during storage. *Electronic Journal of Biotechnology.* 12(4), 2-3.
- Morris, N. L., Miller, P. C. H., Orson, J. H. and Froud-Williams, R. J. (2010). The adoption of non-inversion tillage systems in the United Kingdom and the

- agronomic impact on soil, crops and the environment-A review. *Soil and Tillage Research*. 108(1), 1-15.
- Mueller, D. S., Pierson, W. L. and Wiggs, S. N. (2013). Evaluation of Foliar Fungicides and Insecticides on Soybeans in Central Iowa. *Iowa State University Research and Demonstration Farms*. 30, 13-16.
- Mukherjee, D. and Haque, Z. Z. (2015). Antioxidant activity and persistence of cottonseed protein and oil from two cultivars as determined by their ability to scavenge peroxy and alkoxy radicals. *Journal of Agricultural and Life Sciences*. 2, 6-10.
- Mukherjee, P. K., Horwitz, B. A., Herrera-Estrella, A., Schmoll, M. and Kenerley, C. M. (2013). *Trichoderma* research in the genome era. *Annual Review of Phytopathology*. 51, 105-129.
- Mwamburi, Lizzy A., Mark D. Laing. and Ray M. Miller. (2015). Effect of surfactants and temperature on germination and vegetative growth of *Beauveria bassiana*. *Brazilian Journal of Microbiology*. 46 (1) 67-74.
- Nakazawa, H., Kawai, T., Ida, N., Shida, Y., Kobayashi, Y., Okada, H. and Ogasawara, W. (2012). Construction of a recombinant *Trichoderma reesei* strain expressing *Aspergillus aculeatus* β -glucosidase 1 for efficient biomass conversion. *Biotechnology and Bioengineering*. 109(1), 92-99.
- Naranjo, S. E., Ellsworth, P. C. and Frisvold, G. B. (2015). Economic value of biological control in integrated pest management of managed plant systems. *Annual Review of Entomology*. 60, 621-645.
- Narayan, M.S., Thimmaraju, R. and Bhagyalakshmi, N. (2005) Interplay of growth regulators during solid-state and liquid-state batch cultivation of anthocyanin producing cell line of *Daucus carota*. *Process Biochem*. 40,351-358.
- Naseri, B. (2014). Charcoal rot of bean in diverse cropping systems and soil environments. *Journal of Plant Diseases and Protection*. 121(1), 20-25.
- Ndiaye, M., Sarr, M. P., Cisse, N. and Ndoeye, I. (2015). Is the recently described *Macrophomina pseudophaseolina* pathogenically different from *Macrophomina phaseolina*? *African Journal of Microbiology Research*. 9(45), 2232-2238.
- Newman, J. P. and Molinar, R. H. (2014). Safe and Sustainable Pest Management. *California Master Gardener Handbook*. 3382, 219.

- Nicot, P. C., Blum, B., Kohl, J. and Ruocco, M. (2011). Conclusions and perspectives for future research-and-development projects on biological control of plant pests and diseases. *Classical and Augmentative Biological Control Against Diseases and Pests: Critical Status Analysis and Review of Factors*. 1-11.
- Niranjane, A. P., Madhou, P. and Stevenson, T. W. (2007). The effect of carbohydrate carbon sources on the production of cellulase by *Phlebia gigantea*. *Enzyme and Microbial Technology*. 40(6), 1464-1468.
- Nitta, M., Furukawa, T., Shida, Y., Mori, K., Kuhara, S., Morikawa, Y. and Ogasawara, W. (2012). A new Zn (II) 2 Cys 6-type transcription factor BglR regulates β -glucosidase expression in *Trichoderma reesei*. *Fungal Genetics and Biology*. 49(5), 388-397.
- Noyrod, P., Chailapakul, O., Wonsawat, W. and Chuanuwatanakul, S. (2014). The simultaneous determination of isoproturon and carbendazim pesticides by single drop analysis using a graphene-based electrochemical sensor. *Journal of Electroanalytical Chemistry*. 719, 54-59.
- Olajuyigbe, F. M., Nlekerem, C. M. and Ogunyewo, O. A. (2016). Production and characterization of highly thermostable β -glucosidase during the biodegradation of methyl cellulose by *Fusarium oxysporum*. *Biochemistry Research International*. 2016, 1-8.
- Oliveira-Garcia, E. and Deising, H. B. (2013). Infection Structure-Specific Expression of β -1, 3-Glucan Synthase Is Essential for Pathogenicity of *Colletotrichum graminicola* and Evasion of β -Glucan-Triggered Immunity in Maize. *The Plant Cell*. 25(6), 2356-2378.
- Opassiri, R., Pomthong, B., Akiyama, T., Nakphaichit, M., Onkoksoong, T., Cairns, M. K. and Cairns, J. R. K. (2007). A stress-induced rice (*Oryza sativa* L.) β -glucosidase represents a new subfamily of glycosyl hydrolase family 5 containing a fascin-like domain. *Biochemical Journal*. 408(2), 241-249.
- Pal, K. K. and Gardener, B. M. (2006). Biological control of plant pathogens. *The Plant Health Instructor*. 2(2006), 1117-1142.
- Pan, S. Y., Gao, S. H., Lin, R. C., Zhou, S. F., Dong, H. G., Tang, M. K. and Ko, K. M. (2015). New perspectives on dietary-derived treatments and food safety-antinomy in a new era. *Critical Reviews in Food Science and Nutrition*. 55(13), 1836-1859.

- Pardo, A.G. (1996). Effect of surfactants on cellulase production by *Nectria catalinensis*. *Curr Microbiol.* 33(4), 275-278.
- Park, A. R., Park, J. H., Ahn, H. J., Jang, J. Y., Yu, B. J., Um, B. H. and Yoon, J. J. (2015). Enhancement of β -Glucosidase activity from a brown rot fungus *Fomitopsis pinicola* KCTC 6208 by medium optimization. *Mycobiology.* 43 (1), 57-62.
- Pastrana, A. M., Basallote-Ureba, M. J., Aguado, A., Akdi, K. and Capote, N. (2016). Biological control of strawberry soil-borne pathogens *Macrophomina phaseolina* and *Fusarium solani*, using *Trichoderma asperellum* and *Bacillus* spp. *Phytopathologia Mediterranea.* 55(1), 109.
- Patel, N., Desai, P., Patel, N., Jha, A. and Gautam, H. K. (2014). Agronanotechnology for plant fungal disease management: a review. *International Journal Current Microbiology and Applied Sciences.* 3(2014), 71-84.
- Pearson, C. A. S., Schwenk, F. W., Crowe, F. J. and Kelley, K. (1984). Colonization of soybean roots by *Macrophomina phaseolina*. *Plant Disease.* 68(12), 1086-1088.
- Pereira, L. S., Oweis, T. and Zairi, A. (2002). Irrigation management under water scarcity. *Agricultural Water Management.* 57(3), 175-206.
- Perez-Brandan, C., Arzeno, J. L., Huidobro, J., Grümberg, B., Conforto, C., Hilton, S. and Vargas-Gil, S. (2012). Long-term effect of tillage systems on soil microbiological, chemical and physical parameters and the incidence of charcoal rot by *Macrophomina phaseolina* (Tassi) Goid in soybean. *Crop Protection.* 40(2012), 73-82.
- Pratt, R. G. (2006). A direct observation technique for evaluating sclerotium germination by *Macrophomina phaseolina* and effects of biocontrol materials on survival of sclerotia in soil. *Mycopathologia.* 162(2), 121-131.
- Puglisi, M. P., Sneed, J. M., Sharp, K. H., Ritson-Williams, R. and Paul, V. J. (2014). Marine chemical ecology in benthic environments. *Natural Product Reports.* 31(11), 1510-1553.
- Qadir, M., Wichelns, D., Raschid-Sally, L., McCornick, P. G., Drechsel, P., Bahri, A. and Minhas, P. S. (2010). The challenges of wastewater irrigation in developing countries. *Agricultural Water Management.* 97(4), 561-568.

- Qing, Q., Yang, B. and Wyman, C.E. (2010). Impact of surfactants on pretreatment of corn stover. *Bioresour Technol.* 101(15),5941–5951.
- Raaijmakers, J. M., Paulitz, T. C., Steinberg, C., Alabouvette, C. and Moënne-Loccoz, Y. (2009). The rhizosphere: a playground and battlefield for soilborne pathogens and beneficial microorganisms. *Plant and Soil.* 321(1-2), 341-361.
- Radwan, O., Rouhana, L. V., Hartman, G. L. and Korban, S. S. (2014). Genetic mechanisms of host-pathogen interactions for charcoal rot in soybean. *Plant Molecular Biology Reporter.* 32(3), 617-629.
- Rafi, H., Dawar, S. and Zaki, M. J. (2015). Seed priming with extracts of *Acacia nilotica* (L.) willd. ex Delile and *Sapindus mukorossi* (L.) plant parts in the control of root rot fungi and growth of plants. *Pakistan Journal of Botany.* 47(3), 1129-1135.
- Raghunath, M. and Viswanathan, C. L. (2014). Benzimidazole-2-carbamic acid as a privileged scaffold for antifungal, anthelmintic and antitumor activity: A Review. *International Journal of Pharmacy and Pharmaceutical Sciences.* 6(5), 17-25.
- Ramamurthy, R. K., Jedlicka, J., Graef, G. L. and Waters, B. M. (2014). Identification of new QTLs for seed mineral, cysteine, and methionine concentrations in soybean [*Glycine max* (L.) Merr.]. *Molecular Breeding.* 34(2), 431-445.
- Ramani, G., Meera, B., Vanitha, C., Rao, M. and Gunasekaran, P. (2012). Production, purification, and characterization of a β -glucosidase of *Penicillium funiculosum* NCL1. *Applied Biochemistry and Biotechnology.* 167(5), 959-972.
- Ramos, A. M., Gally, M., Szapiro, G., Itzcovich, T., Carabajal, M. and Levin, L. (2016). In vitro growth and cell wall degrading enzyme production by Argentinean isolates of *Macrophomina phaseolina*, the causative agent of charcoal rot in corn. *Revista Argentina De Microbiologia.* 48(4), 267-273.
- Rasheed, T., Mathew, A., Mathew, M., Sukumaran, R. K. and Elyas, K. K. (2016). Optimization of process parameters for β -glucosidase production by *Byssoschlamys fulva* in slurry state fermentation. *Biotechnological Research.* 1(1), 101-107.

- Rayatpanah, S., Alavi, V. and Arab, G. (2007). Reaction of some Soybean Advanced Lines to Charcoal Rot Disease. *Seed and Plant Improvement Journal*. 23(2), 181-189.
- Rayatpanah, S., Nanagulyan, S. G. and Alavi, S. V. (2009). Chlorate sensitivity, temperature responses and pathogenicity of some Iranian isolates of *Macrophomina phaseolina* from oilseed plants. *Biological Journal of Armenia*. 61 (3), 23-26.
- Rayatpanah, S., Nanagulyan, S. G., Alavi, S. V., Razavi, M. and Ghanbari-Malidarreh, A. (2012). Pathogenic and genetic diversity among Iranian isolates of *Macrophomina phaseolina*. *Chilean Journal of Agricultural Research*. 72 (1), 40-44.
- Razak, M. N. A., Ibrahim, M. F., Yee, P. L., Hassan, M. A. and Abd-Aziz, S. (2012). Utilization of oil palm decanter cake for cellulase and polyoses production. *Biotechnology and Bioprocess Engineering*. 17(3), 547-555.
- Reddy, P. P. (2016). Crop Rotation. In Sustainable Intensification of Crop Production . *Springer Singapore*. 95-107.
- Reichert, I. and Hellinger, E. (1947). On the occurrence, morphology and parasitism of *Sclerotium bataticola*. *Palestine Journal Botany*. 6, 107-147 .
- Reicosky, D. C. and Saxton, K. E. (2007). The benefits of no-tillage. *No-tillage Seeding in Conservation Agriculture*. 11-20.
- Reis, E. M., Boaretto, C. and Danelli, A. L. D. (2014). *Macrophomina phaseolina*: density and longevity of microsclerotia in soybean root tissues and free on the soil, and competitive saprophytic ability. *Summa Phytopathologica*. 40 (2), 128-133.
- Reusche, M., Truskina, J., Thole, K., Nagel, L., Rindfleisch, S., Tran, V. T. and Lipka, V. (2014). Infections with the vascular pathogens *Verticillium longisporum* and *Verticillium dahliae* induce distinct disease symptoms and differentially affect drought stress tolerance of *Arabidopsis thaliana*. *Environmental and Experimental Botany*. 31(108), 23-37.
- Reyes- Franco, M. C., Hernandez- Delgado, S., Beas- Fernandez, R., Medina- Fernandez, M., Simpson, J. and Mayek- Perez, N. (2006). Pathogenic and genetic variability within *Macrophomina phaseolina* from Mexico and other countries. *Journal of Phytopathology*. 154(7- 8), 447-453.

- Rittmann, B. E. and McCarty, P. L. (2012). Environmental biotechnology: principles and applications. *Trends in Biotechnology*. 24(6), 261-266.
- Rocha, V. A. L., Maeda, R. N., Santa Anna, L. M. M. and Pereira, N. (2013). Sugarcane bagasse as feedstock for cellulase production by *Trichoderma harzianum* in optimized culture medium. *Electronic Journal of Biotechnology*. 16(5), 1-13.
- Roy, A., Chakraborty, S., Kundu, S. P., Adhikari, B. and Majumder, S. B. (2012). Adsorption of anionic-azo dye from aqueous solution by lignocellulose-biomass jute fiber: equilibrium, kinetics, and thermodynamics study. *Industrial and Engineering Chemistry Research*. 51(37), 12095-12106.
- Saharan, V., Mehrotra, A., Khatik, R., Rawal, P., Sharma, S. S. and Pal, A. (2013). Synthesis of chitosan based nanoparticles and their *in vitro* evaluation against phytopathogenic fungi. *International Journal of Biological Macromolecules*. 62, 677-683.
- Saibi, W. and Gargouri, A. (2011). Purification and biochemical characterization of an atypical β -glucosidase from *Stachybotrys microspora*. *Journal of Molecular Catalysis B: Enzymatic*. 72(3), 107-115.
- Sakayaroj, J., Preedanon, S., Supaphon, O., Jones, E. G. and Phongpaichit, S. (2010). Phylogenetic diversity of endophyte assemblages associated with the tropical seagrass *Enhalus acoroides* in Thailand. *Fungal Diversity*. 42 (1), 27-45.
- Sakkas, V. A., Islam, M. A., Stalikas, C. and Albanis, T. A. (2010). Photocatalytic degradation using design of experiments: a review and example of the Congo red degradation. *Journal of Hazardous Materials*. 175(1), 33-44.
- Saleh, A. A., Ahmed, H. U., Todd, T. C., Travers, S. E., Zeller, K. A., Leslie, J. F. and Garrett, K. A. (2010). Relatedness of *Macrophomina phaseolina* isolates from tallgrass prairie, maize, soybean and sorghum. *Molecular Ecology*. 19(1), 79-91.
- Sanchez, S., Gambardella, M., Henríquez, J. L. and Diaz, I. (2016). First report of crown rot of strawberry caused by *Macrophomina phaseolina* in Chile. *Journal of Berry Research*. 6(3), 345-354.

- Sangeetha, J. and Thangadurai, D. (2013). Staining techniques and biochemical methods for the identification of fungi. *In Laboratory Protocols in Fungal Biology*. 237-257.
- Santos, C. A., Zanphorlin, L. M., Crucello, A., Tonoli, C. C., Ruller, R., Horta, M. A. and Souza, A. P. (2016). Crystal structure and biochemical characterization of the recombinant ThBgl, a GH1 β -glucosidase overexpressed in *Trichoderma harzianum* under biomass degradation conditions. *Biotechnology for Biofuels*. 9(71), 1-11.
- Sarkar, T. S., Biswas, P., Ghosh, S. K. and Ghosh, S. (2014). Nitric oxide production by necrotrophic pathogen *Macrophomina phaseolina* and the host plant in charcoal rot disease of jute: Complexity of the interplay between necrotroph–host plant interactions. *Plos One*. 9 (9), 107-348.
- Schroder, C., Elleuche, S., Blank, S. and Antranikian, G. (2014). Characterization of a heat-active archaeal β -glucosidase from a hydrothermal spring metagenome. *Enzyme and Microbial Technology*. 57, 48-54.
- Schuster, A. and Schmoll, M. (2010). Biology and biotechnology of *Trichoderma*. *Applied Microbiology and Biotechnology*. 87(3), 787-799.
- Senthilkumar, M. (2005). Studies on the Biocontrol Mechanisms of Soybean Bacterial Endophytes against Charcoal Rot Fungus-*Rhizoctonia Bataticola* (Doctoral dissertation, Indian Agricultural Research Institute, Division Of Microbiology Indian Agricultural Research Institute: New Delhi). 1-54.
- Seshadri, S., Akiyama, T., Opassiri, R., Kuaprasert, B. and Cairns, J. K. (2009). Structural and enzymatic characterization of Os3BGlu6, a rice β -glucosidase hydrolyzing hydrophobic glycosides and (1 \rightarrow 3)-and (1 \rightarrow 2)-linked disaccharides. *Plant Physiology*. 151(1), 47-58
- Sexton, Z. F., Hughes, T. J. and Wise, K. A. (2016). Analyzing Isolate Variability of *Macrophomina phaseolina* from a regional perspective. *Crop Protection*. 31(81), 9-13.
- Shafawati, S. N. and Siddiquee, S. (2013). Composting of oil palm fibres and *Trichoderma spp.* as the biological control agent: A review. *International Biodeterioration and Biodegradation*. 85, 243-253.
- Shah, S., Nasreen, S. and Sheikh, P.A. (2012). Cultural and morphological characterization of *Trichoderma spp.* associated with green mold disease

- of *Pleurotus* spp. in Kashmir. *Research Journal of Microbiology*. 7(2), 139-144.
- Shahidi, F. and Ambigaipalan, P. (2015). Phenolics and polyphenolics in foods, beverages and spices: Antioxidant activity and health effects-A review. *Journal of Functional Foods*. 18, 820-897.
- Shahid, A. A., Rao, Q. A., Bakhsh, A. and Husnain, T. (2012). Entomopathogenic fungi as biological controllers: new insights into their virulence and pathogenicity. *Archives of Biological Sciences*. 64(1), 21-42.
- Shahzadi, T., Anwar, Z., Iqbal, Z., Anjum, A., Aqil, T., Afzal, B. A. and Irshad, M. (2014). Induced production of exoglucanase, and β -glucosidase from fungal co-culture of *T. viride* and *G. lucidum*. *Advances in Bioscience and Biotechnology*. 5, 426-433.
- Sharma, M., Ghosh, R. and Pande, S. (2015). Dry root rot (*Rhizoctonia bataticola* (Taub.) Butler): an emerging disease of chickpea—where do we stand?. *Archives of Phytopathology and Plant Protection*. 48(13-16), 797-812.
- Sharma, R. R., Singh, D. and Singh, R. (2009). Biological control of postharvest diseases of fruits and vegetables by microbial antagonists: A Review *Biological Control*. 50(3), 205-221.
- Sharma, S. K. and Srivastava, S. K. (2014). Fungi Associated with Soybean [*Glycine max* (L.) Merrill] Diseases. *Fungi from Different Substrates*. 27(3), 3-31.
- Sharma, S. K., Srivastava, S. K. and Saxena, M. (2016). Integrated management of fungal diseases of soybean [*Glycine max* (L.) Merrill] occurring in India. In *Fungi: Applications and Management Strategies*. CRC Press. 21, 432-472.
- Shata, H. M., El-Deen, A. M. N., Nawwar, G. A. and Farid M.A. (2014). β -Glucosidase production by mixed culture using crude hemicellulose from rice straw black liquor and peat moss as an inert support. *Egypt Pharmaceut Journal*. 1(13), 121-129.
- Short, G. E., Wyllie, T. D. and Bristow, P. R. (1980). Survival of *Macrophomina phaseolina* in soil and in residue of soybean. *Survival*. 1(7), 13-17.
- Sinclair, J. B. and Backman, P. A. (1989). Compendium of soybean Diseases. 3edn St Paul. Minnesota: *American Phytopathological Society*. 106, 101-106.

- Singhal, L., Bagga, S., Kumar, R. and Chauhan R. (2003). Down-regulation of humoral immunity in chickens due to carbendazim. *Toxicol In Vitro*. 17,687-692.
- Singhania, R. R., Patel, A. K., Sukumaran, R. K., Larroche, C. and Pandey, A. (2013). Role and significance of beta-glucosidases in the hydrolysis of cellulose for bioethanol production. *Bioresource Technology*. 31(127), 500-507.
- Sjaarda, C. P., Abubaker, K. S. and Castle, A. J. (2015). Induction of lcc2 expression and activity by *Agaricus bisporus* provides defence against *Trichoderma aggressivum* toxic extracts. *Microbial Biotechnology*. 8(6), 918-929.
- Smith, A., Beltran, C. A., Kusunoki, M., Cotes, A. M., Motohashi, K., Kondo, T. and Deguchi, M. (2013). Diversity of soil-dwelling *Trichoderma* in Colombia and their potential as biocontrol agents against the phytopathogenic fungus *Sclerotinia sclerotiorum* (Lib.) de Bary. *Journal of General Plant Pathology*.79(1), 74-85.
- Souza, F. H. M., Nascimento, C. V., Rosa, J. C., Masui, D. C., Leone, F. A., Jorge, J. A. and Furriel, R. P. M. (2010). Purification and biochemical characterization of a mycelial glucose-and xylose-stimulated β -glucosidase from the thermophilic fungus *Humicola insolens*. *Process Biochemistry*. 45(2), 272-278.
- Spagnoletti, F. N., Balestrasse, K., Lavado, R. S. and Giacometti, R. (2016). *Arbuscular mycorrhiza* detoxifying response against arsenic and pathogenic fungus in soybean. *Ecotoxicology and Environmental Safety*.133(2016), 47-56.
- Stanger, T. F. and Lauer, J. G. (2008). Corn grain yield response to crop rotation and nitrogen over 35 years. *Agronomy Journal*. 100(3), 643-650.
- Steffens, M., Kolbl, A., Totsche, K. U. and Kogel-Knabner, I. (2008). Grazing effects on soil chemical and physical properties in a semiarid steppe of Inner Mongolia (PR China). *Geoderma*. 143(1), 63-72.
- Stockley, R. A. Vestbo, J., Hurd, S. S., Agusti, A. G., Jones, P. W., Vogelmeier, C. and Anzueto, A. (2013). Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: GOLD executive summary. *American Journal of Respiratory and Critical Care Medicine*. 187(4), 347-365.

- Su, G., Suh, S. O., Schneider, R. W. and Russin, J. S. (2001). Host specialization in the charcoal rot fungus, *Macrophomina phaseolina*. *Phytopathology*. 91(2), 120-126.
- Sun, S., Wang, X., Zhu, Z., Wang, B. and Wang, M. (2016). Occurrence of Charcoal Rot Caused by *Macrophomina phaseolina*, an Emerging Disease of Adzuki Bean in China. *Journal of Phytopathology*. 164 (3), 212-216.
- Susanto, A., Sudharto, P. S. and Purba, R. Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*. 159(1), 153-157.
- Tamura, K., Stecher, G., Peterson, D., Filipinski, A. and Kumar, S. (2013). MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*. 30(12), 2725-2729.
- Tan, S. H. (2013). Morphological characterization and sequence analysis of 5.8 s-its region of *Trichoderma* species (Doctoral dissertation, UTAR). 17-45.
- Tarjuelo, J. M., Rodriguez-Diaz, J. A., Abadia, R., Camacho, E., Rocamora, C. and Moreno, M. A. (2015). Efficient water and energy use in irrigation modernization: Lessons from Spanish case studies. *Agricultural Water Management*. 162, 67-77.
- Terra, V. S., Homer, K. A., Rao, S. G., Andrew, P. W. and Yesilkaya, H. (2010). Characterization of novel β -galactosidase activity that contributes to glycoprotein degradation and virulence in *Streptococcus pneumoniae*. *Infection and Immunity*. 78(1). 348-357.
- Tiwari, P., Misra, B. N. and Sangwan, N. S. (2013). β -Glucosidases from the fungus *Trichoderma*: an efficient cellulase machinery in biotechnological applications. *BioMed Research International*. 20, 1-10.
- Tonin, R. F. B., Avozani, A., Danelli, A. L. D., Reis, E. M., Zoldan, S. M. and Garces-Fiallos, F. R. (2013). *In vitro* mycelial sensitivity of *Macrophomina phaseolina* to fungicides. *Pesquisa Agropecuária Tropical*. 43(4), 460-466.
- Triplett, G. B. and Dick, W. A. (2008). No-tillage crop production: a revolution in agriculture. *Agronomy Journal*. 100 (3), 153-165.
- Troian, R. F., Steindorff, A. S., Ramada, M. H. S., Arruda, W. and Ulhoa, C. J. (2014). Mycoparasitism studies of *Trichoderma harzianum* against

- Sclerotinia sclerotiorum*: evaluation of antagonism and expression of cell wall-degrading enzymes genes. *Biotechnology Letters*. 36(10), 2095-2101.
- Tronsmo, A. and Dennis, C. (1978) Effect of temperature on antagonistic properties of Trichoderma species. *Transactions of the British Mycological Society*. 71(3), 469-474.
- Valiente, C., Diaz, K., Gacitua, S., Martinez, M. and Sanfuentes, E. (2008). Control of charcoal root rot in *Pinus radiata* nurseries with antagonistic bacteria. *World Journal of Microbiology and Biotechnology*. 24(4), 557-568.
- Van Der Plank, J. E. (2012). *Disease resistance in plants*. Elsevier. 23-76.
- Vasebi, A., Bathaee, S. M. T. and Partovibakhsh, M. (2008). Predicting state of charge of lead-acid batteries for hybrid electric vehicles by extended Kalman filter. *Energy Conversion and Management*. 49(1), 75-82.
- Vasebi, Y., Safaie, N. and Alizadeh, A. (2013). Biological control of soybean charcoal root rot disease using bacterial and fungal antagonists *In Vitro* and greenhouse condition. *Journal of Crop Protection*. 2(2), 139-150.
- Vikash, K., Irshita, V., Deepak, K. and Sanjay, G. (2013). Cellulase and β -glucosidase production by *Trichoderma viride* and *Aspergillus wentii* in sub-merged fermentation utilizing pretreated lignocellulosic biomass. *Journal of Microbiology and Biotechnology Research*. 3(5), 63-78.
- Viktorov, A.V. and Yurkiv, V. A. (2013). Effects of carbendazim on Kupffer cell functioning. *Bulletin of Experimental Biology and Medicine*. 154,438-440.
- Vinale, F., Sivasithamparam, K., Ghisalberti, E. L., Marra, R., Woo, S. L. and Lorito, M. (2008). *Trichoderma*-plant-pathogen interactions. *Soil Biology and Biochemistry*. 40(1), 1-10. 568.
- Vishnavat, K. (2013). Smut fungi: potential pathogens and biocontrol agents. *Centre of Advanced Faculty Training in Plant Pathology*. 11,131-135.
- Viterbo, A., Ramot, O., Chernin, L. and Chet, I. (2002). Significance of lytic enzymes from *Trichoderma* spp. in the biocontrol of fungal plant pathogens. *Antonie Van Leeuwenhoek*. 81(1-4), 549-556.
- Vu, V. H., Pham, T. A. and Kim, K. (2011). Improvement of fungal cellulase production by mutation and optimization of solid state-fermentation. *Mycobiology*. 39(1), 20-25.
- Wahab, R. A., Basri, M., Rahman, R. N. Z. R. A., Salleh, A. B., Rahman, M. B. A., Chaibakhsh, N. and Leow, T. C. (2012). Enzymatic production of a

- solvent-free menthyl butyrate via response surface methodology catalyzed by a novel thermostable lipase from *Geobacillus zalihae*. *Biotechnology and Biotechnological Equipment*. 28(6), 1065-1072.
- Wang, Y., Xu, Y. and Li, J. (2012). A Novel Extracellular β - Glucosidase from *Trichosporon asahii*: Yield Prediction, evaluation and application for aroma enhancement of cabernet sauvignon. *Journal of Journal of Food Science*. 77(8), 505-515.
- White, T. J., Bruns, T., Lee, S. J. W. T. and Taylor, J. W. (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR Protocols: a Guide to Methods and Applications*. 18(1), 315-322.
- Wragg, P., Randall, L. and Whatmore, A. M. (2014). Comparison of Biolog GEN III MicroStation semi-automated bacterial identification system with matrix-assisted laser desorption ionization-time of flight mass spectrometry and 16S ribosomal RNA gene sequencing for the identification of bacteria of veterinary interest. *Journal of Microbiological Methods*. 105(2014), 16-21.
- Wrather, J. A. and Koenning, S. R. (2006). Estimates of disease effects on soybean yields in the United States 2003 to 2005. *Journal of Nematology*. 38(2), 173-180.
- Xiao, J., Zhang, H., Niu, L., Wang, X. and Lu, X. (2011). Evaluation of detoxification methods on toxic and antinutritional composition and nutritional quality of proteins in *Jatropha curcas* meal. *Journal of Agricultural and Food Chemistry*. 59(8), 4040-4044.
- Yang, C., Hamel, C., Vujanovic, V. and Gan, Y. (2011). Fungicide: modes of action and possible impact on nontarget microorganisms. *ISRN Ecology*. 10, 13-43.
- Yin, W., Zhou, L., Ma, Y., Tian, G., Zhao, J., Yan, L. and Zhao, Y. (2015). Phytotoxicity, translocation, and biotransformation of NaYF₄ upconversion nanoparticles in a soybean plant. *Small*. 11(36), 4774-4784.
- Yun, S. I., Jeong, C. S., Chung, D. and Choi, H. S. (2001). Purification and some properties of a β -glucosidase from *Trichoderma harzianum* type C-4. *Biosci Biotechnol Biochem*. 65(9): 2028-2032.
- Zahoor, S., Javed, M. M., Aftab, S. and Latif, F. (2011). Metabolic engineering and thermodynamic characterization of an extracellular β -glucosidase

- produced by *Aspergillus niger*. *African Journal of Biotechnology*. 10(41), 8107-8116.
- Zain, M. E. (2011). Impact of mycotoxins on humans and animals. *Journal of Saudi Chemical Society*. 15(2), 129-144.
- Zegada-Lizarazu, W. and Monti, A. (2011). Energy crops in rotation. *Biomass and Bioenergy*. 35(1), 12-25.
- Zhang, H., Yao, M., Morrison, R. A. and Chong, S. (2003). Commonly used surfactant, Tween 80, improves absorption of P-glycoprotein substrate, digoxin, in rats. *Archives of Pharmacal Research*. 26(9), 768-772.
- Zhang, J., Zhao, S., Yin, P., Yan, L., Han, J., Shi, L. and Ma, C. (2014). α -Glucosidase inhibitory activity of polyphenols from the burs of *Castanea mollissima* Blume. *Molecules*. 19(6), 8373-8386.
- Zhang, Z., Liu, J. L., Lan, J. Y., Duan, C. J., Ma, Q. S. and Feng, J. X. (2014). Predominance of *Trichoderma* and *Penicillium* in cellulolytic aerobic filamentous fungi from subtropical and tropical forests in China, and their use in finding highly efficient β -glucosidase. *Biotechnology for Biofuels*. 7(107), 1-14.
- Zhou, Q., Xu, J., Kou, Y., Lv, X., Zhang, X., Zhao, G. and Liu, W. (2012). Differential involvement of β -glucosidases from *Hypocrea jecorina* in rapid induction of cellulase genes by cellulose and cellobiose. *Eukaryotic Cell*. 11(11), 1371-1381.
- Zhu, H., Fu, Y., Jiang, R., Yao, J., Xiao, L. and Zeng, G. (2014). Optimization of copper (II) adsorption onto novel magnetic calcium alginate/maghemite hydrogel beads using response surface methodology. *Industrial and Engineering Chemistry Research*. 53(10), 4059-4066.
- Zimbardi, A. L., Sehn, C., Meleiro, L. P., Souza, F. H., Masui, D. C., Nozawa, M. S. and Furriel, R. P. (2013). Optimization of β -glucosidase, β -xylosidase and xylanase production by *Colletotrichum graminicola* under solid-state fermentation and application in raw sugarcane trash saccharification. *International Journal of Molecular Sciences*. 14(2), 2875-2902.