APPLICATION OF WATER HYACINTH IN WASTEWATER TREATMENT AND CELLULOSIC ETHANOL PRODUCTION

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То

This thesis is dedicated to my family

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ABSTRACT

Water pollution and depletion of fossil fuels are crucial issues which significantly affect sustainability of water resources and usage of non-renewable energies. Phytoremediation technique is an effective method to treat different types of wastewater using plants in an economical way. Lignocellulosic biomass as a renewable resource can be replaced with fossil fuels. In this research, the lignocellulosic aquatic plant, water hyacinth (WH) was propagated in fabricated tanks to evaluate wastewater purification and ethanol production efficiency. Wastewater treatment was performed in two modes (batch and continuous) and the results were compared with the Interim National Water Quality Standards, Malaysia River classification (INWQS) and Water Quality Index (WQI). The matured plants were harvested at maximum growth rate with the highest sugar content. Then, the WH was dried, and four pretreatments, namely ionic liquid, acidic, alkali and microwave-alkali were performed to obtain optimum carbohydrates content. Scanning Electron Microscopy (SEM) and Fourier Transform Infrared (FTIR) were used for compositional and structural analysis on WH before and after pretreatments. Simultaneous Saccharification and Fermentation (SSF) was performed to obtain a high level of sugars which could be converted into ethanol. In the fermentation process, the inoculation of yeast (Saccharomyces cerevisiae) and commercial cellulase enzyme from (Aspergillus niger) were used. Total reducing sugar and ethanol concentration were measured by Dinitrosalicylic Acid (DNS) method and gas chromatography. In the batch system, some parameters such as pH, chemical oxygen demand (COD), phosphate (PO_3^{-4}), nitrate (NO_3^{-}), ammoniacal nitrogen (NH_3) and total organic carbon (TOC) were reduced and the water quality has improved from class III to II. In the continuous system, the removal efficiency 38% to 96% of ten parameters was obtained. The highest ethanol yield was produced by alkali-treated WH with 6.2 ± 0.4 g/L with 0.12 g/g WH/ethanol in 48 hours followed by microwave-alkali, acid and ionic liquid pretreatments.

ABSTRAK

Pencemaran air dan pengurangan bahan api fosil adalah isu-isu penting yang ketara dan memberi kesan kepada kelestarian sumber air dan penggunaan tenaga yang tidak boleh diperbaharui. Teknik Fitopemulihan merupakan kaedah yang berkesan untuk merawat pelbagai jenis tumbuhan air sisa dengan menggunakan cara yang menjimatkan. Lignoselulosa tumbuhan sebagai sumber yang boleh diperbaharui dan boleh diganti dengan bahan api fosil. Dalam kajian ini, akuatik lignoselulosa tumbuhan, iaitu keladi bunting (WH) telah ditanam di dalam tangki yang direka khas untuk menilai pembersihan air sisa dan kecekapan pengeluaran etanol. Rawatan air sisa telah dilaksanakan dalam dua kaedah (kelompok dan berterusan) dan keputusan yang diperolah akan dibandingkan dengan Piawaian Interim Kualiti Air Kebangsaan, klasifikasi River Malaysia (INWQS) dan Indeks Kualiti Air (WQI). Tumbuhtumbuhan yang matang dituai pada kadar pertumbuhan maksimum dengan kandungan gula yang tinggi. Kemudian, WH yang telah kering bersama empat pra-rawatan iaitu cecair ionik, asid, alkali dan ketuhar gelombang mikro-alkali telah dilaksanakan untuk mendapat kandungan karbohidrat yang optimum. Scanning Electron Microscopy (SEM) dan Fourier Transform Infrared (FTIR) telah digunakan untuk analisis komposisi dan struktur ke atas WH sebelum dan selepas pra-rawatan. Saccharification serentak dan Penapaian (SSF) telah dilaksanakan untuk mendapat tahap gula yang tinggi yang boleh ditukar menjadi etanol. Dalam proses penapaian, inokulasi yis (Saccharomyces cerevisiae) dan enzim cellulase komersial dari (Aspergillus niger) telah digunakan. Jumlah pengurangan gula dan kepekatan etanol diukur oleh kaedah Acid Dinitrosalicylic (DNS) dan kromatografi gas. Dalam sistem kumpulan, beberapa parameter seperti pH, keperluan oksigen kimia (COD), fosfat (PO_3^{-4}), nitrat (NO_{3-}), nitrogen ammonia (NH₃) dan jumlah karbon organik (TOC) telah dikurangkan dan kualiti air meningkat daripada kelas III ke II. Dalam sistem yang berterusan, kecekapan penyingkiran 38% kepada 96% daripada sepuluh parameter telah diperolehi. Hasil keputusan menunjukkan bahawa kadar tinggi bagi penghasilan etanol adalah berhubung kait dengan; penggunaan WH dengan 6.2 ± 0.4 g / L dengan 0.12 g / g WH / etanol dalam 48 jam diikuti dengan ketuhar gelombang mikro-alkali, asid dan pra-rawatan cecair ionik.

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LIST OF ABBREVIATIONS

BGL	-	β-glucosidase
BOD	-	Biochemical Oxygen Demand
COD	-	Chemical Oxygen Demand
d	-	Day
DBP	-	Desa Bakti pond
DO	-	Dissolved oxygen
DOE	-	Department of Environment
EC	-	Electricity Conductivity
GC	-	Gas Chromatography
gds	-	gram dry substrate
h	-	Hour
IL	-	Ionic Liquid
INWQS	-	Interim National Water Quality Standards
NB	-	Nutrient Broth
nm	-	Nanometer (10 ⁻⁹ m)
OD	-	Optical Density
PDA	-	Potato Dextrose Agar
RGR	-	Relative growth rate
SHF	-	Separate Hydrolysis and Fermentation
SSF	-	Simultaneous Saccharification and Fermentation
TDS	-	Total Dissolved Solids

TOC	-	Total Organic Carbon
TRS	-	Total Reducing Sugar
TSS	-	Total Suspended Solids
WH	-	Water Hyacinth
WQI	-	Water Quality Index

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CHAPTER 1

INTRODUCTION

1.1 Introduction

Water hyacinth (*Eichhornia crassipes*) has drawn worldwide attention being a harmful weed. It is also considered as a native of South America, invasive and freefloating aquatic macrophyte. Moreover, the rapid growth and quick spreading of Water Hyacinth (WH) cause serious problems for navigation and irrigation (Gopal, 1987). Water hyacinth growth highly depends on the seasonal changes and availability of nutrients in water. For example, the best season for effective cultivation is in summer although it can grow in winter. It competes with almost all the other species growing in their vicinity, which decreases the biodiversity (Craft *et al.*, 2003). It devastates aquatic environment and costs billions of dollars every year in control. As a result, many environmentalists are in favor of the removal of WH from fresh water ecosystem. They discovered that there are several ways to remove this problematic weed from water such as using biological and mechanical methods. However, after removal, it can be used in many useful and eco-friendly processes to keep our environment clean (Gupta *et al.*, 2012).

Water pollution is a significant environmental issue that has been increasing exponentially with urbanization and modernization along with the population growth of the world. As discussed by Ling, (2010), water pollution is a crucial issue in Malaysia which significantly affects the sustainability of water resources and management. Phytoremediation technique is an effective method to treat different types of wastewater using plants in an economical way. Important mechanisms for phytoremediation technique are uptake, accumulation, translocation and metabolism of micro-contaminants (Lv *et al.*, 2016). In this technique, floating aquatic plants such as WH can be used to uptake organic and inorganic matter from wastewater. The abilities of WH such as higher growth rate than other aquatic plants, high absorption efficiency and low operation cost imply that using this plant can be considered as a suitable green plant for the treatment of wastewater (Gupta *et al.*, 2012).

At present, depletion of fossil fuels in near future is predictable by continuous over-utilization of natural energy sources (Jambo *et al.*, 2016). Additionally, lack of energy resources is in an alarming stage due to globalization and increasing demand for energy. In this situation, the researchers tried to find alternative sources of energy that are not related to fossil oil and natural gas (Bradshaw, 2010). On the other hand, the concern on greenhouse effect is an important reason for interest in renewable energy sources. For example, ethanol due to its potential as an alternative automotive fuel has attracted worldwide attention. In addition, concerns towards CO_2 emissions and associated climate change have instigated an accelerated research and production of renewable energy resources (Sanchez and Cardona, 2008). Therefore, these issues have driven many countries to become interested in the use of biofuels as a replacement source of renewable and cheap energy (Tye *et al.*, 2016). Water hyacinth is a type of lignocellulosic material that is potentially available in the tropical region of the world. (Ganguly *et al.*, 2012; Sing and Bishnoi, 2013).

1.2 Background of the study

For a phytoremediation system to work efficiently, an optimal plant growth is the key parameter. The growth rate of WH is found to be dependent on the concentration of dissolved nitrogen (N) and phosphorus (P) present in the wastewater. In addition, the propagation of WH depends on the availability of light, nutrients and the abundance and stability of weevil populations (Williams *et al.*, 2005). The most common aquatic macrophytes among the floating-leaved plants being employed in wastewater treatment are WH and water lettuce (John *et al.*, 2008, Maine *et al.*, 2004; Mishra *et al.*, 2008). There are many studies on the removal of a wide range of pollutants from wastewater by WH (Patel, 2012; Gupta *et al.*, 2012). Significant removal of inorganic nitrogen (nitrate), ammoniacal nitrogen (NH₃-N), total N, phosphorus and total P have been reported by using WH in nutrient rich wastewater (Lu, 2009).

Agricultural products like corn, wheat and sugarcane are defined as the first generation, lignocellulosic biomass as the second generation and algal biomass as the third generation of raw materials for bioethanol production (Baeyens *et al.*, 2015). There are several advantages in the production of ethanol in comparison with gasoline, namely the utilization of an abundant and cheap source of renewable resources, reduction in greenhouse gas emission and toxic substances by carbon neutrality and the significant benefits for rural community and social aspect of sustainability (Lin and Tanaka, 2006). A world production of 50 million m³ in 2007 and in excess of 100 million m³ in 2012 of bio-ethanol was reported as an alternative renewable fuel with the largest potential to replace fossil-derived fuels. Brazil and the United States represent approximately 80% of the world supply of bioethanol (Renewable Fuels Association, 2013). This source of energy also has a potential for a significant reduction of greenhouse gas emissions of fossil fuel origin as well by carbon sequestration (Dias *et al.*, 2013).

Lignocellulosic biomass consists of carbohydrates that can be derived from monosaccharide and disaccharide. On the other side, carbohydrates are comprised of cellulose and hemicellulose that are tightly wrapped by lignin. Water hyacinth, which contains large quantity of cellulose but small content of lignin, can be converted into bioethanol by enzymes. The conversion of biomass into hydrocarbons utilizes a complex but reasonably well understood biochemical route (Shafiei et al., 2015). Large quantities of cellulase and hemicellulose can be produced by genetically engineered microorganisms (Dashtban et al., 2010). Fermentable sugars can be obtained by these enzymes from lignocellulosic compounds like agricultural residues (e.g. corn stover, straw, sugar cane bagasse) and energy crops (e.g. switchgrass) (Mousdale 2008). Genetically modified fermentative and cellulolytic microorganisms are crucial to increase ethanol yield and productivity under stress conditions such as high temperature (Chen, 2009). Due to importance of hydrolytic pre-treatment for cellulose, hemi-cellulose and lignin separation, the conversion of lignocellulosic biomass is an expensive and difficult process. After separation of these materials, the hexose and pentose sugars can be fermented into different alcohols (Anca-Couce, 2016).

Increasing the yield of bioethanol and improvement of the process of simultaneous saccharification and fermentation can be obtained from a study on the decomposition of lignocellulosic biomass (Kotarska et al, 2015). Compositional analysis of lignocellulose before and after pretreatment, is used in almost all second-generation biofuel production studies. According to (Karimi and Taherzadeh, 2016a), an accurate analysis is essential to evaluate the biofuel production. The efficiency of pretreatment methods highly depends on a variety of analytical methods that address their image and composition which can be obtained from Scanning Electron Microscopy (SEM) and Fourier Transform Infrared (FTIR) analysis.

Using appropriate fermentation substrate and suitable process technology can increase the yield of ethanol production by microbial fermentation (Brooks, 2008). In order to increase the bioethanol yield, these pretreatment methods are applied to decrease the recalcitrance of lignocellulosic feedstock. Pretreatment can decrease

crystallinity of cellulose, increase accessibility of polymers by enzymes and reduce lignin content and these functions depend on the mode of the pretreatment methods (Zheng *et al.*, 2014).

1.3 Problem statement

Water is the delicate part of the environment, which is essential for human life and industrial development. Water pollution is not a recent environmental issue and has become a hot topic with urbanization and modernization. Malaysia's major source of surface water contributes some 97% of the total water supply (Gasim et al, 2009). Phytoremediation technique can be applied to treat different types of wastewater that in comparison to other techniques is an environmentally friendly. In literature, WH showed a greater uptake ability in comparison to other aquatic plants like water lettuce (Ismael *et al.*, 2015), water morning glory (Loan *et al.*, 2014) and azolla (Anandha *et al.*, 2015). Water hyacinth has amazing ability to remove different types of pollutants such as heavy metals, organic and inorganic compounds from aquatic environments. It grows quickly even in extreme conditions and no land space is needed for its cultivation (Akinbile *et al.*, 2012; Akinbile *et al.*, 2016).

In many countries, fossil fuel usage is a big problem that is regarded as nonrenewable energy. Malaysia currently relies on fossil fuels for over 90% of its power generation. With such a high demand of energy usage and abundant biomass resources can utilize the biomass residue (Hu, 2016). In contrast to food crops for biofuel production, the utilization of lignocellulosic biomass reduces the carbon footprint and greenhouse gas emission effects (Luque *et al.*, 2008; Govumoni *et al.*, 2013). In this regard, biodiesel and bio-ethanol are the most common biofuels and can replace fossil diesel and gasoline in car engines (Mata *et al.*, 2010). Water hyacinth is an aquatic and abundant plant and a good candidate for production of substitute fuels such as bioethanol due to the high percentage of carbohydrates. Interestingly, WH has a low amount of lignin (10%), high amount of cellulose (25%) and hemicellulose (35%). The economic potential of the biofuel production can be improved by using low-lignin biomass in comparison with lignin-rich biomass (Bhatt and Shilpa, 2014). According to Singh and Bishnoi, (2012), three main operations are needed to convert lignocellulose to bioethanol; the pretreatment is necessary to liberate cellulose and hemicellulose before hydrolysis, the hydrolysis of cellulose and hemicellulose to produce fermentable sugars and fermentation of reducing sugars. In this way, the most common and traditional microorganism used in industrial bioethanol production is the yeast *Saccharomyces cerevisiae* (Dionisi *et al.,* 2015).

1.4 Objectives of the study

The objectives of this research are as follows:

- 1. To determine the maximum rate of organic and nutrient removal from domestic wastewater stabilization pond using local species of WH in batch and continuous system
- 2. To identify the potential parts of WH suitable to produce sugars in large quantities and the correlation to the growth rate of the biomass
- To evaluate the efficiency of different pretreatment methods based on compositional and structural analyses in morphology of raw and treated WH by SEM and FTIR
- 4. To establish high rate production system of ethanol by fermentation by evaluating different pretreatment methods

1.5 Scope of the study

The scope of this research is to study the ability of WH to remove pollutants from domestic wastewater that leads to produce a high amount of bioethanol by simultaneous saccharification and alcohol fermentation. The efficiency of nutrient uptake by matured WH from domestic wastewater in fabricated tanks was compared in batch (no water flow) and continuous system (with water flow). Then, best part of WH in terms of high level of available sugars was selected based on preliminary study. Although, biomass should be harvested at the suitable stage of maturity that could produce highest reducing sugars with a suitable pretreatment.

Various pretreatment methods such as alkali, acidic, Microwave-alkali and Ionic liquid were applied to enhance the recovery ratio of sugars from the biomass. In addition, the effect of different pretreatments on composition of WH was investigated. Then, the conversion ability of glucose fermenting yeast (*Saccharomyces cerevisiae*) for conversion of sugars to ethanol was evaluated. Furthermore, by using cellulase the conversion of sugar to ethanol (yield) would be higher than absence of enzyme. Overall, WH can uptake nutrient from wastewater for its growth that leads to wastewater polishing. It is predicted that the best part in terms of higher sugar and carbohydrates content and lower lignin content would be more feasible for ethanol production.

1.6 Significance of the study

Effluent from oxidation pond with a wide range of pollutants is being released into rivers and has harmful impacts on human health. Studies have shown that WH was effective in removing wide range of nutrients from different types of wastewater. During this study, a free available macrophyte WH was applied as a multi-functional plant to remove contaminants from oxidation pond as well as ethanol production in presence of cellulase. The whole process is like a sequence as: the efficiency of fermentation is based on the amount of the sugars related to nutrient uptake from waste water and it is predicted that more sugar obtained can produce more ethanol. In this study, the conversion of waste to wealth can be considered as the most important issue.

This study is significant in regard to:

- Purification of domestic waste water by WH by uptake of organic and inorganic compounds
- Decrease the air pollution and CO₂ fixation by cultivation of this plant in the environment
- Using WH as a raw material which is abundant and free substrate for wide range of application as a multi-functional plant especially in tropical regions like Malaysia
- Ethanol produced from WH as a green fuel to replace petrol and therefore reduction of greenhouse gas emission
- Different methods of pretreatment evaluated to select the most effective method

REFERENCES

- Abdel-Fattah, A. F., Abdel-Naby, M. A. (2012). Pretreatment and enzymic saccharification of water hyacinth cellulose. *Carbohydrate Polymers*. 87(3): 2109-2113.
- Ahn, D. J., Kim, S. K., & Yun, H. S. (2012). Optimization of pretreatment and saccharification for the production of bioethanol from water hyacinth by *Saccharomyces cerevisiae. Bioprocess and Biosystems Engineering.* 35(1-2): 35-41.
- Ajayi, T. O., & Ogunbayio, A. O. (2012). Achieving environmental sustainability in wastewater treatment by phytoremediation with water hyacinth (*Eichhornia crassipes*). Journal of Sustainable Development. 5(7): 80-90.
- Akinbile, C. O., & Yusoff, M. S. (2012). Assessing water hyacinth (*Eichhornia crassopes*) and lettuce (*Pistia stratiotes*) effectiveness in aquaculture wastewater treatment. *International journal of phytoremediation*. 14(3): 201-211.
- Akinbile, C. O., Ogunrinde, T. A., Che bt Man, H., & Aziz, H. A. (2016). Phytoremediation of domestic wastewaters in free water surface constructed wetlands using Azolla pinnata. *International Journal of Phytoremediation* 18(1): 54-61.
- Alam, M. F., Baishnab, A. C., Khan, M. R., & Islam, M. A. (2014). Fermentable Sugar Production and Separation from Water Hyacinth Using Enzymatic Hydrolysis. *Sustainable Energy*. 2(1): 20-24.
- Alizadeh, H., Teymouri, F., Gilbert, T. I., & Dale, B. E. (2005). Pretreatment of switchgrass by ammonia fiber explosion (AFEX). *Applied Biochemistry and Biotechnology*. 124(1-3): 1133-1141.
- Alvi, S. S. M., Ali, M. N., Mohiuddin, M., Khan, M. M., & Khan, M. M. (2014). Eichhornia crassipes - a potential substrate for biofuel production. International Journal of Current Microbiology and Applied Sciences. 3(10): 618-627.

- Alvira, P., Tomás-Pejó, E., Ballesteros, M., & Negro, M. J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: a review. *Bioresource Technology*. 101(13): 4851-4861.
- Amiri, H., & Karimi, K. (2015). Improvement of acetone, butanol, and ethanol production from woody biomass using organosolv pretreatment. *Bioprocess and Biosystems Engineering*. 38(10): 1959-1972.
- Anandha Varun, R., Kalpana, S., (2015). Performance analysis of nutrient removal in pond water using Water Hyacinth and Azolla with papaya stem. *International Research Journal of Engineering Technology*. 2, 444-448.
- Anca-Couce, A. (2016). Reaction mechanisms and multi-scale modelling of lignocellulosic biomass pyrolysis. *Progress in Energy and Combustion Science*. 53: 41-79.
- Ang, S. K., & Abd-Aziz, S. (2015). Potential Uses of Xylanase-Rich Lignocellulolytic Enzymes Cocktail for Oil Palm Trunk (OPT) Degradation and Lignocellulosic Ethanol Production. *Energy & Fuels*. 29(8): 5103-5116.
- Ang, S. K., Shaza, E. M., Adibah, Y., Suraini, A. A., & Madihah, M. S. (2013). Production of cellulases and xylanase by *Aspergillus fumigatus* SK1 using untreated oil palm trunk through solid state fermentation. *Process Biochemistry*. 48(9): 1293-1302.
- APHA. 2005. Standard Methods for the Examination of Water and Waste Water. 21st Edition, American Public Health Association, Washington, DC, 10–15
- Ariunbaatar J, Panico A, Esposito G, Pirozzi F, & Lens PN, (2014). Pretreatment methods to enhance anaerobic digestion of organic solid waste. Applied Energy. 123, 143-156.
- Aswathy, U. S., Sukumaran, R. K., Devi, G. L., Rajasree, K. P., Singhania, R. R., & Pandey, A. (2010). Bio-ethanol from water hyacinth biomass: an evaluation of enzymatic saccharification strategy. *Bioresource Technology*. 101(3): 925-930.
- Awasthi, M., Kaur, J., & Rana, S. (2013). Bioethanol production through water hyacinth, *Eichhornia crassipes* via optimization of the pretreatment conditions. *International Journal of Emerging Technology and Advanced Engineering*. 3(3): 42-46.
- Baeyens, J., Kang, Q., Appels, L., Dewil, R., Lv, Y., & Tan, T. (2015). Challenges and opportunities in improving the production of bio-ethanol *.Progress in Energy* and Combustion Science. 47: 60-88.

- Balat, M. (2011). Production of bioethanol from lignocellulosic materials via the biochemical pathway: a review. *Energy Conversion and Management*. 52(2): 858-875.
- Banerjee, G., Car, S., Scott-Craig, J. S., Borrusch, M. S., & Walton, J. D. (2010). Rapid optimization of enzyme mixtures for deconstruction of diverse pretreatment/biomass feedstock combinations. *Biotechnology for biofuels*. 3(1), 22. DOI: 10.1186/1754-6834-3-22.
- Basha, S. A., Gopal, K. R., & Jebaraj, S. (2009). A review on biodiesel production, combustion, emissions and performance. *Renewable and Sustainable Energy Reviews.* 13(6): 1628-1634.
- Bensah, E. C., & Mensah, M. (2013). Chemical pretreatment methods for the production of cellulosic ethanol: technologies and innovations. *International Journal of Chemical Engineering*. 1-21. http://dx.doi.org/10.1155/2013/719607.
- Bhatt, S. M., & Shilpa. (2014). Lignocellulosic feedstock conversion, inhibitor detoxification and cellulosic hydrolysis–a review. *Biofuels*. 5(6): 633-649.
- Bhattacharya, A., & Kumar, P. (2010). Water hyacinth as a potential biofuel crop. *Electron Journal of Environmental Agricultural Food Chemistry*. 9(1): 112-122.
- Bhattacharya, A., Ganguly, A., Das, S., Chatterjee, P. K., & Dey, A. (2013). Fungal isolates from local environment: isolation, screening and application for the production of ethanol from water hyacinth. *International Journal of Emerging Technology and Advanced Engineering*. 3(3): 58-65.
- Binod, P., Janu, K.U., Sindhu, R., Pandey, A., (2011). In: Hydrolysis of lignocellulosic biomass for bioethanol production. In: Pandey, Ashok, Larroche, Christian, Ricke, Steven C. (Eds.), Biofuels: Alternative Feedstock's and Conversion Processes. *Elsevier Inc., USA*. pp. 229–250
- Biswas, B., Banik, A. K., & Biswas, A. B (2015). Comparative study of various pretreatment techniques for saccharifications of water hyacinth (*eichhornia crassipes*) cellulose. *International Journal of Biotech Trends and Technology*. 12(1): 11-16.
- Biswas, S. R., Choudhury, J. K., Nishat, A., & Rahman, M. M. (2007). Do invasive plants threaten the Sundarbans mangrove forest of Bangladesh? *Forest Ecology and Management*. 245(1): 1-9.
- Bradshaw, M. J. (2010). Global energy dilemmas: a geographical perspective. *The Geographical Journal*. 176(4): 275-290.

Brekke, K. (2005). The Promise of cellulosic ethanol. Ethanol Today. 6, 32-36.

- Brooks, A. A. (2008). Ethanol production potential of local yeast strains isolated from ripe banana peels. *African journal of Biotechnology*. 7(20): 3749-3752.
- Brown, T. R., & Brown, R. C. (2013). A review of cellulosic biofuel commercialscale projects in the United States. *Biofuels, Bioproducts and Biorefining*. 7(3): 235-245.
- Buaban, B., Inoue, H., Yano, S., Tanapongpipat, S., Ruanglek, V., Champreda, V., ...
 & Eurwilaichitr, L. (2010). Bioethanol production from ball milled bagasse using an on-site produced fungal enzyme cocktail and xylose-fermenting *Pichia* stipitis. Journal of Bioscience and Bioengineering. 110(1): 18-25.
- Carvalheiro F, Duarte LC, Girio FM (2008). Hemicellulose biorefineries: a review on biomass pretreatments. *Journal of Scientific and Industrial Research*. 67:849–64.
- Center, T.D., Hill, M.P., Cordo, H., Julien, M.H., (2002). Water hyacinth. In: Van Driesche, R., et al. (Eds.), Biological Control of Invasive Plants in the Eastern United States, 4. USDA Forest Service Publication FHTET. Washington, DC. 41-64.
- Chandel AK, Kapoor RK, Singh,A, & Kuhad RC, (2007). Detoxification of sugarcane bagasse hydrolysate improves ethanol production by *Candida shehatae* NCIM 3501. *Bioresource Technology*. 98(10): 1947-1950.
- Chandra, M., Kalra, A., Sharma, P. K. and Sangwan, R. S. (2009). Cellulase production by six *Trichoderma spp*. fermented on medicinal plant processing. *Journal of Industrial Microbiology & Biotechnology*. 36, 605-609.
- Chang, M. C. (2007). Harnessing energy from plant biomass. *Current opinion in Chemical Biology*. 11(6): 677-684.
- Chen, W. H., Tu, Y. J., & Sheen, H. K. (2011). Disruption of sugarcane bagasse lignocellulosic structure by means of dilute sulfuric acid pretreatment with microwave-assisted heating. *Applied Energy*. 88(8): 2726-2734.
- Chen, X., Chen, X., Wan, X., Weng, B., & Huang, Q. (2010). Water hyacinth (*Eichhornia crassipes*) waste as an adsorbent for phosphorus removal from swine wastewater. *Bioresource Technology*. 101(23): 9025-9030.
- Chen, Y. (2009). Initial Investigation on Xylose Fermentation for Lignocellulosic Bioethanol Production. Doctoral dissertation. Auburn University.

- Chen, Y. (2011). Development and application of co-culture for ethanol production by co-fermentation of glucose and xylose: a systematic review. *Journal of industrial Microbiology & Biotechnology*. 38(5): 581-597.
- Cheng, J., Wang, X., Huang, R., Liu, M., Yu, C., & Cen, K. (2014). Producing Ethanol from Water Hyacinth through Simultaneous Saccharification and Fermentation with Acclimatized Yeasts. *BioResources*. 9(4): 7666-7680.
- Cheng, Y. S., Chen, K. Y., & Chou, T. H. (2015). Concurrent calcium peroxide pretreatment and wet storage of water hyacinth for fermentable sugar production. *Bioresource Technology*. 176, 267-272.
- Chu, J. J., Ding, Y., & Zhuang, Q. J. (2006). Invasion and control of water hyacinth (*Eichhornia crassipes*) in China. *Journal of Zhejiang University Science B*. 7(8): 623-626.
- Chuang, Y. S., Lay, C. H., Sen, B., Chen, C. C., Gopalakrishnan, K., Wu, J. H., ... & Lin, C. Y. (2011). Biohydrogen and biomethane from water hyacinth (Eichhornia crassipes) fermentation: effects of substrate concentration and incubation temperature. *International Journal of Hydrogen Energy*. 36(21): 14195-14203.
- Claassen, P. A. M., Van Lier, J. B., Contreras, A. L., Van Niel, E. W. J., Sijtsma, L., Stams, A. J. M., ... & Weusthuis, R. A. (1999). Utilisation of biomass for the supply of energy carriers. *Applied Microbiology and Biotechnology*. 52(6): 741-755.
- Craft, C., Megonigal, P., Broome, S., Stevenson, J., Freese, R., Cornell, J., & Sacco, J. (2003). The pace of ecosystem development of constructed Spartina alterniflora marshes. *Ecological Applications*. 13(5): 1417-1432.
- Dadi, A. P., Varanasi, S., & Schall, C. A. (2006). Enhancement of cellulose saccharification kinetics using an ionic liquid pretreatment step. *Biotechnology* and *Bioengineering*. 95(5): 904-910.
- Das, A., Ghosh, P., Paul, T., Ghosh, U., Pati, B. R., & Mondal, K. C. (2016a). Production of bioethanol as useful biofuel through the bioconversion of water hyacinth (*Eichhornia crassipes*). *3 Biotech*. DOI: 10.1007/s13205-016-0385-y.
- Das, S. P., Gupta, A., Das, D., & Goyal, A. (2016b). Enhanced bioethanol production from water hyacinth (*Eichhornia crassipes*) by statistical optimization of fermentation process parameters using Taguchi orthogonal array design. International Biodeterioration & Biodegradation, 109, 174-184.

- Das, S. P., Ravindran, R., Ghosh, A., Deka, D., Das, D., Jawed, M., ... & Goyal, A. (2014a). Efficient pretreatment for bioethanol production from water hyacinth (*Eichhornia crassipes*) involving naturally isolated and recombinant enzymes and its recovery. *Environmental Progress & Sustainable Energy*. 33(4): 1396-1404.
- Das, S., Bhattacharya, A., Ganguly, A., Gu, S., & Chatterjee, P. K. (2014b). Kinetics and genetic algorithm study of acid catalysed hydrolysis of water hyacinth biomass. *Journal of Renewable Sustainable Energy*. 6(6): 063115.
- Das, S., Bhattacharya, A., Haldar, S., Ganguly, A., Gu, S., Ting, Y. P., & Chatterjee,
 P. K. (2015). Optimization of enzymatic saccharification of water hyacinth biomass for bio-ethanol: Comparison between artificial neural network and response surface methodology. *Sustainable Materials and Technologies*. 3, 17-28.
- Dashtban, M., Schraft, H., & Qin, W. (2009). Fungal bioconversion of lignocellulosic residues; opportunities & perspectives. *International Journal of Biological Sciences*. 5(6): 578.
- De Casabianca, M. L., Laugier, T., & Posada, F. (1995). Petroliferous wastewaters treatment with water hyacinths (Raffinerie de Provence, France): Experimental statement. *Waste Management*. 15(8): 651-655.
- De Groote, H., Ajuonu, O., Attignon, S., Djessou, R., & Neuenschwander, P. (2003). Economic impact of biological control of water hyacinth in Southern Benin. *Ecological Economics*. 45(1): 105-117.
- de Souza, A. P., Leite, D. C., Pattathil, S., Hahn, M. G., & Buckeridge, M. S. (2013). Composition and structure of sugarcane cell wall polysaccharides: implications for second-generation bioethanol production. *Bioenergy Research*, 6(2), 564-579.
- De Stefani, G., Tocchetto, D., Salvato, M., and Borin, M. (2011). Performance of a floating treatment wetland for in-stream water amelioration in NE Italy. *Hydrobiologia*. 674, 157-167.
- DellaGreca, M., Previtera, L., & Zarrelli, A. (2009). Structures of new phenylphenalene-related compounds from *Eichhornia crassipes* (water hyacinth). *Tetrahedron*. 65(39): 8206-8208.
- Dhankhar, R., & Dhaka, A. (2014). Bioconversion of Water Hyacinth to Ethanol by Using Cellulase from Trichoderma atroviride AD-130. Advanced Materials Research. 918, 145-148.

- Di Blasi, C., Signorelli, G., Di Russo, C., & Rea, G. (1999). Product distribution from pyrolysis of wood and agricultural residues. *Industrial & Engineering Chemistry Research*. 38(6): 2216-2224.
- Dias, M. O., Junqueira, T. L., Rossell, C. E. V., Maciel Filho, R., & Bonomi, A. (2013). Evaluation of process configurations for second generation integrated with first generation bioethanol production from sugarcane. *Fuel Processing Technology*. 109, 84-89.
- Dionisi, D., Anderson, J. A., Aulenta, F., McCue, A., & Paton, G. (2015). The potential of microbial processes for lignocellulosic biomass conversion to ethanol: a review. *Journal of Chemical Technology and Biotechnology*. 90(3): 366-383.
- Dixit, A., Dixit, S., & Goswami, C. S. (2011). Process and plants for wastewater remediation: a review. *Scientific Reviews & Chemical Communications*. 11, 71-7.
- DOE, (Department of Environment Malaysia), (2007). Malaysia Environmental Quality Report 2006, In: Chapter 3: River Water Quality, Sasyaz Holdings Sdn Bhd, pp. 24.
- Dominguez, H., Núñez, M. N., Chamy, R., & Lema, J. M. (1993). Determination of kinetic parameters of fermentation processes by a continuous unsteady-state method: Application to the alcoholic fermentation of D-xylose by *Pichia stipitis*. *Biotechnology and Bioengineering*. 41(11): 1129-1132.
- Du Preez, J. C., Van Driessel, B., & Prior, B. A. (1989). D-xylose fermentation by *Candida shehatae* and *Pichia stipitis* at low dissolved oxygen levels in fed-batch cultures. *Biotechnology letters*. 11(2): 131-136.
- Eshtiaghi, M. N., Yoswathana, N., Kuldiloke, J., & Ebadi, A. G. (2012). Preliminary study for bioconversion of water hyacinth (*Eichhornia crassipes*) to bioethanol. *African Journal of Biotechnology*. 11(21): 4921-4928.
- Fan, M., Dai, D., Huang, B., 2012. Fourier transform infrared spectroscopy for natural fibers. In: Salih, S.M. (Ed.), Fourier Transform – Materials Analysis. InTech. 46-68.
- Fileto-Pérez, H. A., Rutiaga-Quiñones, J. G., Aguilar-González, C. N., Paéz, J. B., López, J., & Rutiaga-Quiñones, O. M. (2013). Evaluation of *Eichhornia* crassipes as an Alternative Raw Material for Reducing Sugars Production. *BioResources* 8(4): 5340-5348.

- Ganguly A, Das S, Bhattacharya A, Dey A, Chatterjee PK (2013b). Enzymatic hydrolysis of water hyacinth biomass for the production of ethanol: Optimization of driving parameters. *Indian Journal of Experimental Biology*. (51): 556-566.
- Ganguly, A., Chatterjee, P. K., & Dey, A. (2012). Studies on ethanol production from water hyacinth—A review. *Renewable and Sustainable Energy Reviews*. 16(1): 966-972.
- Ganguly, A., Halder, S., Laha, A., Saha, N., Chatterjee, P. K., & Dey, A. (2013a). Effect of Alkali Pretreatment on Water Hyacinth Biomass for Production of Ethanol. Advanced Chemical Engineering Research. 2(2):40-44.
- Gao, J., Chen, L., Yan, Z., & Wang, L. (2013a). Effect of ionic liquid pretreatment on the composition, structure and biogas production of water hyacinth (*Eichhornia crassipes*). *Bioresource Technology*. 132, 361-364.
- Gao, J., Chen, L., Yuan, K., Huang, H., Yan, Z., (2013b). Ionic liquid pretreatment to enhance the anaerobic digestion of lignocellulosic biomass. *Bioresource Technology*. 150, 352-358.
- Gao, J., Chen, L., Zhang, J., & Yan, Z. (2014). Improved enzymatic hydrolysis of lignocellulosic biomass through pretreatment with plasma electrolysis. *Bioresource Technology*. 171, 469-471.
- Gao, J., Dang, H., Liu, L., & Jiang, L. (2015). Remediation effect of contaminated water by water hyacinth (*Eichhornia crassipes* (Mart.) Solms). *Desalination and Water Treatment*. 55(2): 381-388.
- Gasim, M. B., Ismail Sahid, E. T., Pereira, J. J., Mokhtar, M., & Abdullah, M. P. (2009). Integrated water resource management and pollution sources in Cameron Highlands, Pahang, Malaysia. *American-Eurasian Journal Agricultural Environmental Science*, 5, 725-732.
- Ghaly, A. E., Kamal, M., & Mahmoud, N. S. (2005). Phytoremediation of aquaculture wastewater for water recycling and production of fish feed. *Environment International*. 31(1): 1-13.
- Ghose, T. (1987). Measurement of cellulase activities. *Pure and Applied Chemistry*. 59, 257-268.
- Gopal, B. (1987). Aquatic Plant Studies 1. Water Hyacinth .Elsevier Oxford. 471 pp.
- Govumoni, S. P., Koti, S., Kothagouni, S. Y., Venkateshwar, S., & Linga, V. R. (2013). Evaluation of pretreatment methods for enzymatic saccharification of wheat straw for bioethanol production. *Carbohydrate polymers*. 91(2): 646-650.

- Gunnarsson, C. C., & Petersen, C. M. (2007). Water hyacinths as a resource in agriculture and energy production: A literature review. Waste Management. 27(1): 117-129.
- Guo, B. H., Tang, H. C., Song, Z. W., & Xi, J. X. (2003). Theory of wastewater treatment by constructed wetlands and removal of nitrogen and phosphorus. *Pollution Control Technology*. 16(4): 119-121.
- Gupta, A., & Verma, J. P. (2015). Sustainable bio-ethanol production from agroresidues: a review. *Renewable and Sustainable Energy Reviews*, 41, 550-567.
- Gupta, P., Roy, S., & Mahindrakar, A. B. (2012). Treatment of water using water hyacinth, water lettuce and vetiver grass-A review. *Resources and Environment*. 2(5): 202-215.
- Guragain, Y. N., De Coninck, J., Husson, F., Durand, A., & Rakshit, S. K. (2011). Comparison of some new pretreatment methods for second generation bioethanol production from wheat straw and water hyacinth. *Bioresource Technology*. 102(6): 4416-4424.
- Hamelinck, C. N., Van Hooijdonk, G., & Faaij, A. P. (2005). Ethanol from lignocellulosic biomass: techno-economic performance in short-, middle-and long-term. *Biomass and Bioenergy*. 28(4): 384-410.
- Harun, M. Y., Radiah, A. D., Abidin, Z. Z., & Yunus, R. (2011). Effect of physical pretreatment on dilute acid hydrolysis of water hyacinth (*Eichhornia crassipes*). *Bioresource Technology*. 102(8): 5193-5199.
- Hassan, O., Ling, T. P., Maskat, M. Y., Illias, R. M., Badri, K., Jahim, J. and Mahadi, N. M. (2013). Optimization of pretreatments for the hydrolysis of oil palm empty fruit bunch fiber (EFBF) using enzyme mixtures. *Biomass and Bioenergy*. 56, 137-146
- Heard, T. A., & Winterton, S. L. (2000). Interactions between nutrient status and weevil herbivory in the biological control of water hyacinth. *Journal of Applied Ecology*. 37(1): 117-127.
- Hendriks, A. T. W. M., Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*. 100(1): 10–18.
- Ho, L. W. (2016). Wind energy in Malaysia: Past, present and future. *Renewable and Sustainable Energy Reviews*. 53, 279-295.
- Hoffmann, W. A., & Poorter, H. (2002). Avoiding bias in calculations of relative growth rate. *Annals of Botany*. 90(1): 37-42.

- Hossain, R., Chowdhury, M. K., Yeasmin, S., & Hoq, M. M. (2010). Production of ethanol using yeast isolates on water hyacinth and Azolla. *Bangladesh Journal of Microbiology*. 27(2): 56-60.
- Hu, G., Heitmann, J. A., & Rojas, O. J. (2008). Feedstock pretreatment strategies for producing ethanol from wood, bark, and forest residues. *BioResources*. 3(1): 270-294
- Hu, Z., & Wen, Z. (2008). Enhancing enzymatic digestibility of switchgrass by microwave-assisted alkali pretreatment. *Biochemical Engineering Journal*. 38(3): 369-378.
- Huang, Y. H., Jin, Y. L., Fang, Y., Li, Y. H., Zhang, G. H., Xiao, Y., et al. (2013). Simultaneous saccharification and fermentation (SSF) of non-starch polysaccharides and starch from fresh tuber of Canna edulis ker at a high solid content for ethanol production. *Biomass Bioenergy*. 52, 8–14.
- Idi, A., & Mohamad, S. E. (2011). Bioethanol from second generation feedstock (lignocellulose biomass). *Interdisciplinary Journal of Contemporary Research in Business*. 3(8): 919-935.
- Idrees, M., Adnan, A., Sheikh, S., & Qureshi, F. A. (2013). Optimization of dilute acid pretreatment of water hyacinth biomass for enzymatic hydrolysis and ethanol production. *EXCLI Journal*. 12, 30-40.
- Imman, S., Arnthong, J., Burapatana, V., Champreda, V., & Laosiripojana, N. (2015). Fractionation of rice straw by a single-step solvothermal process: Effects of solvents, acid promoters, and microwave treatment. *Renewable Energy*, 83, 663-673.
- Isarankura-Na-Ayudhya, C., Kongpanpee, T., Prabkate, P., Prachayasittikul, V., & Tantimongcolwat, T. (2007). Appropriate Technology for the Bioconversion of Water Hyacinth (*Eichhornia crassipes*) to Liquid Ethanol, *EXCLI Journal*. 6, 167-176.
- Ismail, Z., Othman, S. Z., Law, K. H., Sulaiman, A. H., & Hashim, R. (2015). Comparative Performance of Water Hyacinth (*Eichhornia crassipes*) and Water Lettuce (*Pista stratiotes*) in Preventing Nutrients Build-up in Municipal Wastewater. *CLEAN–Soil. Air. Water.* 43, 521-531.
- Jafari, N. (2010). Ecological and socio-economic utilization of water hyacinth (*Eichhornia crassipes* Mart Solms). *Journal of Applied Sciences and Environmental Management.* 14(2): 43-49.

- Jambo, S. A., Abdulla, R., Azhar, S. H. M., Marbawi, H., Gansau, J. A., & Ravindra, P. (2016). A review on third generation bioethanol feedstock. *Renewable and Sustainable Energy Reviews*, 65, 756-769.
- Jamuna, S., Noorjahan, C.M. (2009). Treatment of sewage waste water using water hyacinth - Eichhornia sp and its reuse for fish culture. *Toxicology International*. 16, 103-106.
- Jimenez, M. M., & Balandra, M. A. G. (2007). Integrated control of Eichhornia crassipes by using insects and plant pathogens in Mexico. *Crop Protection*. 26(8): 1234-1238.
- John, R., Ahmad, P., Gadgil, K., & Sharma, S. (2008). Effect of cadmium and lead on growth, biochemical parameters and uptake in Lemna polyrrhiza L. *Plant Soil* and Environment. 54(6): 262-270.
- Kang, S. W., Park, Y. S., Lee, J. S., Hong, S. I., & Kim, S. W. (2004). Production of cellulases and hemicellulases by *Aspergillus niger* KK2 from lignocellulosic biomass. *Bioresource Technology*. 91(2): 153-156.
- Karimi, K., & Taherzadeh, M. J. (2016a). A critical review of analytical methods in pretreatment of lignocelluloses: Composition, imaging, and crystallinity. *Bioresource Technology*. 200, 1008-1018.
- Karimi, K., & Taherzadeh, M. J. (2016b). A critical review on analysis in pretreatment of lignocelluloses: Degree of polymerization, adsorption/desorption, and accessibility. *Bioresource Technology*. 203, 348-356.
- Kateregga, E., & Sterner, T. (2007). Indicators for an invasive species: Water hyacinths in Lake Victoria. *Ecological Indicators*. 7(2): 362-370.
- Keshwani, D. R., & Cheng, J. J. (2009). Switchgrass for bioethanol and other valueadded applications: a review. *Bioresource Technology*. 100(4), 1515-1523.
- Khanna, S., Santos, M. J., Ustin, S. L., & Haverkamp, P. J. (2011). An integrated approach to a biophysiologically based classification of floating aquatic macrophytes. *International Journal of Remote Sensing*. 32(4): 1067-1094.
- Klumpp, A., Bauer, K., Franz-Gerstein, C., & de Menezes, M. (2002). Variation of nutrient and metal concentrations in aquatic macrophytes along the Rio Cachoeira in Bahia (Brazil). *Environment International*. 28(3): 165-171.
- Ko, J. K., Bak, J. S., Jung, M. W., Lee, H. J., Choi, I. G., Kim, T. H., & Kim, K. H. (2009). Ethanol production from rice straw using optimized aqueous-ammonia

soaking pretreatment and simultaneous saccharification and fermentation processes. *Bioresource Technology*. 100(19): 4374-4380.

- Kola K. 1988. Aspects of the ecology of water hyacinth *Eichhornia crassipes* (Martius) Solms. In: Farri TA, editors. The Lagos Lagoon System. Lagos (Nigeria): Proceedings of the International Workshop on Water Hyacinth-Menace and Resource, *Nigerian Federal Ministry of Science and Technology*. p. 80–84.
- Kootstra, A. M. J., Beeftink, H. H., Scott, E. L., & Sanders, J. P. (2009). Comparison of dilute mineral and organic acid pretreatment for enzymatic hydrolysis of wheat straw. *Biochemical Engineering Journal*. 46(2): 126-131.
- Kotarska, K., Świerczyńska, A., & Dziemianowicz, W. (2015). Study on the decomposition of lignocellulosic biomass and subjecting it to alcoholic fermentation: study on the decomposition of lignocellulosic biomass. *Renewable Energy*. 75: 389-394.
- Koutika, L.S., Rainey, H.J. (2015). A Review of the Invasive, Biological and Beneficial Characteristics of Aquatic Species *Eichhornia Crassipes* and *Salvinia Molesta. Applied Ecology and Environmental Research.* 13, 263-275.
- Kumar, A., Singh, L. K., & Ghosh, S. (2009). Bioconversion of lignocellulosic fraction of water-hyacinth (*Eichhornia crassipes*) hemicellulose acid hydrolysate to ethanol by Pichia stipitis. *Bioresource Technology*. 100(13): 3293-3297.
- Kumar, D., & Murthy, G. S. (2011). Impact of pretreatment and downstream processing technologies on economics and energy in cellulosic ethanol production. *Biotechnology for biofuels*. 4(1), 27. DOI: 10.1186/1754-6834-4-27.
- Kumar, P., Barrett, D. M., Delwiche, M. J., & Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial & Engineering Chemistry Research*. 48(8): 3713-3729.
- Kumari, M., & Tripathi, B. D. (2014). Effect of aeration and mixed culture of *Eichhornia crassipes* and *Salvinia natans* on removal of wastewater pollutants. *Ecological Engineering*. 62, 48-53.
- Kumari, N., Bhattacharya, A., Dey, A., Ganguly, A., & Chatterjee, P. K. (2014b). Bioethanol production from water hyacinth biomass using isolated fungal strain from local environment. *Biolife*. 2(2): 516-522.
- Kunatsa, T., Madiye, L., Chikuku, T., Shonhiwa, C., & Musademba, D. (2013). Feasibility study of biogas production from water hyacinth, A case of Laka

Chivero-Harare, Zimbabwe. International Journal of Engineering and Technology. 3(2): 119-28.

- Li, X., Kim, T. H., & Nghiem, N. P. (2010). Bioethanol production from corn stover using aqueous ammonia pretreatment and two-phase simultaneous saccharification and fermentation (TPSSF). *Bioresource Technology*. 101(15): 5910-5916.
- Liao, S., Chang, W. (2004). Heavy metal phytoremediation by water hyacinth at constructed wetlands in Taiwan. *Photogramm Engineering Remote Sensing*. 54, 177-185.
- Lin, R., Cheng, J., Song, W., Ding, L., Xie, B., Zhou, J., & Cen, K. (2015). Characterization of water hyacinth with microwave-heated alkali pretreatment for enhanced enzymatic digestibility and hydrogen/methane fermentation. Bioresource Technology. 182, 1-7.

Lin, Y., & Tanaka, S. (2006). Ethanol fermentation from biomass resources: current state and prospects. *Applied Microbiology and Biotechnology*. 69(6): 627-642.

- Ling, J. K. B. (2010). Water quality study and its relationship with high tide and low tide at Kuantan River. Doctoral dissertation. Universiti Malaysia Pahang.
- Lissy, P.N.M., Madhu, G. (2011). Removal of Heavy Metals from Waste Water Using Water Hyacinth, ACEEE. *International Journal on Transportation and Urban Development*. 1, 48-52.
- Loan, N. T., Phuong, N. M., & Anh, N. T. N. (2014). The role of aquatic plants and microorganisms in domestic wastewater treatment. *Environmental Engineering* and Management Journal. 13(8); 2031-2038.
- Lu, Q., (2009). Evaluation of aquatic plants for phytoremediation of eutrophic storm waters. PhD Thesis. University of Florida, Florida
- Luque, R., Herrero-Davila, L., Campelo, J. M., Clark, J. H., Hidalgo, J. M., Luna, D., ... & Romero, A. A. (2008). Biofuels: a technological perspective. *Energy & Environmental Science*. 1(5): 542-564.
- Lv, T., Zhang, Y., Casas, M. E., Carvalho, P. N., Arias, C. A., Bester, K., & Brix, H. (2016). Phytoremediation of imazalil and tebuconazole by four emergent wetland plant species in hydroponic medium. *Chemosphere*, 148, 459-466.
- M. Mandels, M., & Weber, J. (1969). The production of cellulases. Advances in Chemistry Series. 95, 394–414.

- Ma, F., Yang, N., Xu, C., Yu, H., Wu, J., & Zhang, X. (2010). Combination of biological pretreatment with mild acid pretreatment for enzymatic hydrolysis and ethanol production from water hyacinth. *Bioresource Technology*. 101(24): 9600-9604.
- Maine, M.A., Sune, N.L. and Lagger, S.C., (2004). Chromium bioaccumulation: Comparison of the capacity of two floating aquatic macrophytes. *Water Research*. 38, 1494-1501.
- Malik, A. (2007). Environmental challenge vis a vis opportunity: the case of water hyacinth. *Environment International*. 33(1): 122-138.
- Mandels, M., & Weber, J. (1969). The production of cellulases.
- Manivannan A, Narendhirakannan RT (2015). Bioethanol Production from Aquatic Weed Water Hyacinth (*Eichhornia crassipes*) by Yeast Fermentation. *Waste & Biomass Valorization*. 6(2): 209-216.
- Masami, G. O., Usui, I., & Urano, N. (2008). Ethanol production from the water hyacinth *Eichhornia crassipes* by yeast isolated from various hydrospheres. *African Journal of Microbiology Research*. 2(5): 110-113.
- Mata, T. M., Martins, A. A., & Caetano, N. S. (2010). Microalgae for biodiesel production and other applications: a review. *Renewable and Sustainable Energy Reviews*. 14(1): 217-232.
- Mbati, G., & Neuenschwander, P. (2005). Biological control of three floating water weeds, *Eichhornia crassipes, Pistia stratiotes*, and *Salvinia molesta* in the Republic of Congo. *BioControl.* 50(4): 635-645.
- Menon, V., & Rao, M. (2012). Trends in bioconversion of lignocellulose: biofuels, platform chemicals & biorefinery concept. *Progress in Energy and Combustion Science*. 38(4), 522-550.
- Milano, J., Ong, H. C., Masjuki, H. H., Chong, W. T., Lam, M. K., Loh, P. K., & Vellayan, V. (2016). Microalgae biofuels as an alternative to fossil fuel for power generation. *Renewable and Sustainable Energy Reviews*, 58, 180-197.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*. 31(3): 426-428.
- Mishima, D., Kuniki, M., Sei, K., Soda, S., Ike, M., & Fujita, M. (2008). Ethanol production from candidate energy crops: water hyacinth (*Eichhornia crassipes*) and water lettuce (*Pistia stratiotes L.*). *Bioresource Technology*. 99(7): 2495-2500.

- Mishra, V. K., Upadhyay, A. R., Pandey, S. K., & Tripathi, B. D. (2008). Concentrations of heavy metals and aquatic macrophytes of Govind Ballabh Pant Sagar an anthropogenic lake affected by coal mining effluent. *Environmental monitoring and assessment*. 141(1-3): 49-58.
- Mood, S. H., Golfeshan, A. H., Tabatabaei, M., Jouzani, G. S., Najafi, G. H., Gholami, M., & Ardjmand, M. (2013). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renewable and Sustainable Energy Reviews*. 27, 77-93.
- Moretti, M. M. S., Bocchini-Martins, D. A., Nunes, C. d. C. C., Villena, M. A., Perrone, O. M., Silva, R. D., Boscolo, M., Gomes, E. (2014). Pretreatment of sugarcane bagasse with microwaves irradiation and its effects on the structure and on enzymatic hydrolysis. *Applied Energy*. 122, 189–195.
- Mousdale, D. M. (2008). *Biofuels: biotechnology, chemistry, and sustainable development*. CRC press. Taylor & Francis Group, New York.
- Moyo, P., Chapungu, L., & Mudzengi, B. (2013). Effectiveness of water Hyacinth (*Eichhornia crassipes*) in remediating polluted water: The case of Shagashe river in Masvingo, Zimbabwe. *Advances in Applied Science Research.* 4 (4): 55-62.
- Mukhopadhyay, S. B., & Chatterjee, N. C. (2010). Bioconversion of water hyacinth hydrolysate into ethanol. *BioResources*. 5(2): 1301-1310.
- Munavalli, G., & Saler, P. (2009). Treatment of dairy wastewater by water hyacinth. *Water Science & Technology*. 5(4): 713–722.
- Naik, S. N., Goud, V. V., Rout, P. K., & Dalai, A. K. (2010). Production of first and second generation biofuels: a comprehensive review. *Renewable and Sustainable Energy Reviews*. 14(2), 578-597.
- Nasir, N. F., Daud, W. R. W., Kamarudin, S. K., & Yaakob, Z. (2013). Process system engineering in biodiesel production: A review. *Renewable and Sustainable Energy Reviews*. 22, 631-639.
- Nguyen, M. T., Choi, S. P., Lee, J., Lee, J. H., & Sim, S. J. (2009). Hydrothermal acid pretreatment of *Chlamydomonas reinhardtii* biomass for ethanol production. Journal of Microbiology and Biotechnology. 19(2): 161-166.
- Nigam, J. N. (2002). Bioconversion of water-hyacinth (Eichhornia crassipes) hemicellulose acid hydrolysate to motor fuel ethanol by xylose–fermenting yeast. *Journal of Biotechnology*. 97(2): 107-116.

Nigam, P. S., & Singh, A. (2011). Production of liquid biofuels from renewable resources. *Progress in Energy and Combustion Science* 37(1): 52-68.

OECD-FAO. (2012). Agricultural outlook, chapter 3 biofuels. 87-117

- Olivares, E., Colonnello, G. (2000). Salinity gradient in the Manamo River, a dammed distributary of the Orinoco Delta, and its influence on the presence of *Eichhornia crassipes* and *Paspalum repens. Interciencia.* 25(5): 242-248.
- Oudhia, P. (1999). Medicinal weeds in rice fields of Chhattisgarh, India. *International Rice Research Notes*. 24, 40-46.
- Patel, S. (2012). Threats, management and envisaged utilizations of aquatic weed Eichhornia crassipes: an overview. Reviews in Environmental Science and Bio/Technology. 11(3): 249-259.
- Phitsuwan, P., Sakka, K., & Ratanakhanokchai, K. (2013). Improvement of lignocellulosic biomass in planta: a review of feedstocks, biomass recalcitrance, and strategic manipulation of ideal plants designed for ethanol production and processability. *Biomass and Bioenergy*. 58, 390-405.
- Poletto, M., Pistor, V., & Zattera, A. J. (2013). Structural characteristics and thermal properties of native cellulose. Cellulose-fundamental aspects. Ed. Van de Ven, T. and Gdbout, L. InTech, 45-68.
- Priya, E.S., Selvan, P.S. (2014). Water hyacinth (*Eichhornia crassipes*)-An efficient and economic adsorbent for textile effluent treatment–A review. *Arabian. Journal. Chemistry.* <u>http://dx.doi.org/10.1016/j.arabjc.2014.03.002</u>
- Puls, J., Poutanen, K., Korner, H.U., Viikari, L. (1985). Biotechnical utilization of wood carbohydrates after steaming pretreatment. *Applied Microbiology and Biotechnology*. 22, 416–423.
- Reales-Alfaro, J. G., Trujillo-Daza, L. T., Arzuaga-Lindado, G., Castaño-Peláez, H.
 I., & Polo-Córdoba, Á. D. (2013). Acid hydrolysis of water hyacinth to obtain fermentable sugars. *CT&F-Ciencia, Tecnología y Futuro*. 5(2): 101-111.
- Reddy K.R., Agami M., Angelo E.M.D. Tucker, J.C. (1991). Influence of potassium supply on growth and nutrient storage by water hyacinth, *Bioresource Technology*. (37):79-84.
- Reddy R.R. and Tucker J.C. (1983). Productivity and Nutrient Uptake of Water Hyacinth, *Eichhornia crassipes*, I. Effect of Nitrogen Source. *Economic Botany*. 37 (2): 237-247.

- Renewable Fuels Association. (2013). *World fuel ethanol production*, Retrieved from: http://ethanolrfa.org/pages/World-Fuel-Ethanol-Production. [04.06.14]
- Rezania S, Ponraj M, Din MFM, Songip AR, Md Sairan F, Chelliapan S, (2015a). The diverse applications of water hyacinth with main focus on sustainable energy and production for new era: An overview. *Renewable and Sustainable Energy Reviews.* 41, 943-954.
- Rezania, S., Din, M. F. M., Ponraj, M., Sairan, F. M., & binti Kamaruddin, S. F. (2013). Nutrient uptake and wastewater purification with Water Hyacinth and its effect on plant growth in batch system. *Journal of Environmental Treatment Techniques*. 1(2): 81-85.
- Rezania, S., Din, M. F. M., Taib, S. M., Dahalan, F. A., Songip, A. R., Singh, L., & Kamyab, H. (2016b). The Efficient Role of Aquatic Plant (Water Hyacinth) in Treating Domestic Wastewater in Continuous System. *International journal of phytoremediation*. 18(7): 679-685.
- Rezania, S., Ponraj, M., Fadhil Md Din, M., Chelliapan, S., & Md Sairan, F. (2016a). Effectiveness of *Eichhornia crassipes* in nutrient removal from domestic wastewater based on its optimal growth rate. *Desalination and Water Treatment*. 57(1): 360-365.
- Rezania, S., Ponraj, M., Talaiekhozani, A., Mohamad, S. E., Din, M.F.M., Taib, S.M, Sabbagh F, Sairan, F. M. (2015b). Perspectives of phytoremediation using water hyacinth for removal of heavy metals, organic and inorganic pollutants in wastewater. *Journal of Environmental Management*. 163, 125-133.
- Rodrigues, A. J., Odero, M. O., Hayombe, P. O., Akuno, W., Kerich, D., & Maobe, I. (2014). Converting water hyacinth to briquettes: a beach community based approach. *International Journal of Science: Basic Applied Research*. 15(1): 358-78.
- Roongtanakiat, N., Tangruangkiat, S., & Meesat, R. (2007). Utilization of vetiver grass (Vetiveria zizanioides) for removal of heavy metals from industrial wastewaters. *Science Asia.* 33, 397-403.
- Ruan, T., Zeng, R., Yin, X. Y., Zhang, S. X., & Yang, Z. H. (2016). Water Hyacinth (*Eichhornia crassipes*) Biomass as a Biofuel Feedstock by Enzymatic Hydrolysis. *BioResources*. 11(1): 2372-2380.

- Sanchez, O. J., & Cardona, C. A. (2008). Trends in biotechnological production of fuel ethanol from different feedstocks. *Bioresource Technology*. 99(13): 5270-5295.
- Sarkar, N., & Aikat, K. (2012). Cellulase and xylanase production from rice straw by a locally isolated fungus *Aspergillus fumigatus* NITDGPKA3 under solid state fermentation-Statistical optimization by response surface methodology. *Journal of Technology Innovations in Renewable Energy*. 1(1): 54-62.
- Sarkar, N., Ghosh, S. K., Bannerjee, S., & Aikat, K. (2012). Bioethanol production from agricultural wastes: An overview. *Renewable Energy*. 37(1): 19-27.
- Satyanagalakshmi K, Sindhu R, Binod P, Kanakambaran U, Janu RK, Sukumaran A (2011). Bioethanol production from acid pretreated water hyacinth by separate hydrolysis and fermentation. Journal of Scientific and Industrial Research. 70, 156-161.
- Schell, D. J., Dowe, N., Chapeaux, A., Nelson, R. S., & Jennings, E. W. (2016). Accounting for all Sugars Produced during Integrated Production of Ethanol from Lignocellulosic Biomass. *Bioresource Technology*. 205, 153-158.
- Shabana, Y. M., & Mohamed, Z. A. (2005). Integrated control of water hyacinth with a mycoherbicide and a phenylpropanoid pathway inhibitor. *Biocontrol science and Technology*. 15(7): 659-669.
- Shafiei, M., Kumar, R., & Karimi, K. (2015). Pretreatment of lignocellulosic biomass. In Lignocellulose-Based Bioproducts. Springer International Publishing. (pp. 85-154).
- Shah, R. A., Kumawat, D. M., Singh, N., & Wani, K. A. (2010). Water hyacinth (*Eichhornia crassipes*) as a remediation tool for dye-effluent pollution. *International Journal of Science and Nature*. 1(2): 172-178.
- Shanab, S. M., Shalaby, E. A., Lightfoot, D. A., & El-Shemy, H. A. (2010). Allelopathic effects of water hyacinth (*Eichhornia crassipes*). PLoS One. 5(10): e13200. doi:10.1371/journal.pone.0013200.
- Singh, A., & Bishnoi, N. R. (2012). Optimization of ethanol production from microwave alkali pretreated rice straw using statistical experimental designs by *Saccharomyces cerevisiae. Industrial Crops and Products.* 37(1): 334-341.
- Singh, A., & Bishnoi, N. R. (2013). Comparative study of various pretreatment techniques for ethanol production from water hyacinth. *Industrial Crops and Products.* 44, 283-289.

- Singh, O. V., Labana, S., Pandey, G., Budhiraja, R., & Jain, R. K. (2003). Phytoremediation: an overview of metallic ion decontamination from soil. *Applied Microbiology and Biotechnology*. 61(5-6): 405-412.
- Singh, P., Suman, A., Tiwari, P., Arya, N., Gaur, A., & Shrivastava, A. K. (2008). Biological pretreatment of sugarcane trash for its conversion to fermentable sugars. *World Journal of Microbiology and Biotechnology*. 24(5): 667-673.
- Sooknah, R. D., & Wilkie, A. C. (2004). Nutrient removal by floating aquatic macrophytes cultured in anaerobically digested flushed dairy manure wastewater. *Ecological Engineering*. 22(1): 27-42.
- Sornvoraweat, B., & Kongkiattikajorn, J. (2010). Separated hydrolysis and fermentation of water hyacinth leaves for ethanol production. *KKU Research Journal*. 15(9): 794-802.
- Su, H., Cheng, J., Zhou, J., Song, W., & Cen, K. (2010). Hydrogen production from water hyacinth through dark-and photo-fermentation. *International Journal of Hydrogen Energy*. 35(17): 8929-8937.
- Sukumaran, R. K., Singhania, R. R., Mathew, G. M., & Pandey, A. (2009). Cellulase production using biomass feed stock and its application in lignocellulose saccharification for bio-ethanol production. *Renewable Energy*. 34(2): 421-424.
- Sun, S., Sun, S., Cao, X., & Sun, R. (2016). The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials. *Bioresource Technology*. 199, 49-58.
- Sun, Y. C., Wen, J. L., Xu, F., & Sun, R. C. (2011). Structural and thermal characterization of hemicelluloses isolated by organic solvents and alkaline solutions from *Tamarix austromongolica*. *Bioresource Technology*. 102(10): 5947-5951.
- Sun, Y., & Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource Technology*. 83(1): 1-11.
- Sundari, M. T., & Ramesh, A. (2012). Isolation and characterization of cellulose nanofibers from the aquatic weed water hyacinth—*Eichhornia crassipes*. *Carbohydrate Polymers*. 87(2): 1701-1705.
- Taherzadeh, M. J., & Karimi, K. (2007). Acid-based hydrolysis processes for ethanol from lignocellulosic materials: a review. *BioResources*. 2(3): 472-499.

- Taherzadeh, M. J., & Karimi, K. (2008). Pretreatment of lignocellulosic wastes to improve ethanol and biogas production: a review. International Journal of Molecular Sciences. 9(9): 1621-1651.
- Takagi, T., Uchida, M., Matsushima, R., Ishida, M., & Urano, N. (2012). Efficient bioethanol production from water hyacinth *Eichhornia crassipes* by both preparation of the saccharified solution and selection of fermenting yeasts. *Fisheries science*. 78(4): 905-910.
- Talebnia, F., Karakashev, D., & Angelidaki, I. (2010). Production of bioethanol from wheat straw: an overview on pretreatment, hydrolysis and fermentation. *Bioresource Technology*. 101(13): 4744-4753.
- Tan, H. T., Lee, K. T., & Mohamed, A. R. (2011). Pretreatment of lignocellulosic palm biomass using a solvent-ionic liquid [BMIM] Cl for glucose recovery: An optimisation study using response surface methodology. *Carbohydrate Polymers*. 83(4): 1862-1868.
- Tellez, T. R., López, E. M., Granado, G. L., Pérez, E. A., López, R. M., & Guzmán, J. M. S. (2008). The water hyacinth, Eichhornia crassipes: an invasive plant in the Guadiana River Basin (Spain). *Aquatic Invasions*. 3(1): 42-53.
- Tiwari, S., Dixit, S., & Verma, N. (2007). An effective means of biofiltration of heavy metal contaminated water bodies using aquatic weed *Eichhornia* crassipes. Environmental Monitoring and Assessment. 129(1-3): 253-256.
- Tye, Y. Y., Lee, K. T., Abdullah, W. N. W., & Leh, C. P. (2016). The world availability of non-wood lignocellulosic biomass for the production of cellulosic ethanol and potential pretreatments for the enhancement of enzymatic saccharification. *Renewable and Sustainable Energy Reviews*, 60, 155-172.
- Uday, U. S. P., Choudhury, P., Bandyopadhyay, T. K., & Bhunia, B. (2016). Classification, mode of action and production strategy of xylanase and its application for biofuel production from water hyacinth. *International Journal of Biological Macromolecules*. 82, 1041-1054.
- Valentine, J., Clifton-Brown, J., Hastings, A., Robson, P., Allison, G., & Smith, P. (2012). Food vs. fuel: the use of land for lignocellulosic 'next generation'energy crops that minimize competition with primary food production. *GCB Bioenergy*. 4(1): 1-19.

- Valipour, A., Raman, V.K., & Ahn, Y.H. (2015). Effectiveness of Domestic Wastewater Treatment Using a Bio-Hedge Water Hyacinth Wetland System. *Water*. 7, 329-347.
- Vani, S., Sukumaran, R. K., & Savithri, S. (2015). Prediction of sugar yields during hydrolysis of lignocellulosic biomass using artificial neural network modeling. *Bioresource Technology*. 188, 128-135.
- Verma, R., Singh, S. P., & Ganesha Raj, K. (2003). Assessment of changes in waterhyacinth coverage of water bodies in northern part of Bangalore city using temporal remote sensing data. *Current science*. 84(6): 795-804
- Vidya, S., and Girish, L. (2014). Water Hyacinth as a green manure for Organic farming, *International Journal of Research in Applied*, *Natural and Social Sciences*. 2(6); 65-72.

Vymazal, J. (2007). Removal of nutrients in various types of constructed wetlands. *Science of the Total Environment*. 380(1-3): 48-65.

- Weerachanchai, P., Leong, S. S. J., Chang, M. W., Ching, C. B., & Lee, J. M. (2012). Improvement of biomass properties by pretreatment with ionic liquids for bioconversion process. *Bioresource Technology*. 111, 453-459.
- Williams, A. E., Duthie, H. C., & Hecky, R. E. (2005). Water hyacinth in Lake Victoria: Why did it vanish so quickly and will it return? *Aquatic Botany*. 81(4): 300-314.
- Wilson, J. R., Holst, N., & Rees, M. (2005). Determinants and patterns of population growth in water hyacinth. *Aquatic Botany*. 81(1): 51-67.
- Wilson, J. R., Rees, M., Holst, N., Thomas, M. B., & Hill, G. (2000). Water hyacinth population dynamics. Biological and Integrated Control of Water Hyacinth, *Eichhornia crassipes. ACIAR Proceedings*. 102, 96-104.
- Xia, A., Cheng, J., Song, W., Yu, C., Zhou, J., & Cen, K. (2013). Enhancing enzymatic saccharification of water hyacinth through microwave heating with dilute acid pretreatment for biomass energy utilization. *Energy*. 61, 158-166.
- Xu, F., Chen, L., Wang, A., & Yan, Z. (2016). Influence of surfactant-free ionic liquid microemulsions pretreatment on the composition, structure and enzymatic hydrolysis of water hyacinth. *Bioresource Technology*. 208, 19-23.
- Yan, J., Wei, Z., Wang, Q., He, M., Li, S., & Irbis, C. (2015). Bioethanol production from sodium hydroxide/hydrogen peroxide-pretreated water hyacinth via simultaneous saccharification and fermentation with a newly isolated

thermotolerant Kluyveromyces marxianu strain. *Bioresource Technology*. 193, 103-109.

- Yang, B., & Wyman, C. E. (2008). Pretreatment: the key to unlocking low-cost cellulosic ethanol. Biofuels. *Bioproducts and Biorefining*. 2(1): 26-40.
- Zhang, Q., Weng, C., Huang, H., Achal, V., & Wang, D. (2015). Optimization of bioethanol production using whole plant of Water Hyacinth as substrate in Simultaneous Saccharification and Fermentation process. *Frontiers in Microbiology*. 6: 1411. doi: 10.3389/fmicb.2015.01411
- Zhao, H., Jones, C. L., Baker, G. A., Xia, S., Olubajo, O., & Person, V. N. (2009). Regenerating cellulose from ionic liquids for an accelerated enzymatic hydrolysis. *Journal of Biotechnology*. 139(1): 47-54.
- Zheng, Y., Pan, Z., & Zhang, R. (2009). Overview of biomass pretreatment for cellulosic ethanol production. *International Journal of Agricultural and Biological Engineering*. 2(3): 51-68.
- Zheng, Y., Zhao, J., Xu, F., & Li, Y. (2014). Pretreatment of lignocellulosic biomass for enhanced biogas production. *Progress in Energy and Combustion Science*. 42, 35-53.
- Zhu, L., O'Dwyer, J. P., Chang, V. S., Granda, C. B., & Holtzapple, M. T. (2008). Structural features affecting biomass enzymatic digestibility. *Bioresource Technology*. 99(9): 3817-3828.