INHIBITION OF MELANOGENESIS THROUGH THE DEACTIVATION OF TYROSINASE ACTIVITY IN MELANOMA CELLS BY *LABISIA PUMILA* FRACTIONS

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Dedicated to my beloved mom, my dad, my wife, Kasmah Manur, and my children, Ahmad Qaid Isyraf and Qaireen Inarah.

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ABSTRACT

Effective and safe tyrosinase inhibitors, especially those from natural sources, are desired for the application in food and whitening cosmetic products. In this research, characterization and in-vitro study of L. pumila methanol fractions have been carried out. L. pumila methanol fractions were prepared using solid phase extraction (SPE) and labelled as 40%, 60% and 100% methanol fractions. From characterization by total phenolic content (TPC) analysis, 40% and 100% methanol fractions were found to give the highest amount of phenolic content. The potential tyrosinase inhibition effects of L. pumila methanol fractions were evaluated using B16F1 melanoma cells. The cytotoxicity of the methanol fractions was evaluated using MTT (3-(4, 5-Dimethylthiazol-2-yl)-2,5) diphenyltetrazolium bromide) assay. The results revealed that concentration up to 0.1% w/v of the *L*. *pumila* methanol fractions did not affect the cell viability. The secreted melanin had gradually decreased after the treatment with 0.025-0.1% (w/v) of L. pumila methanol fractions. L. pumila methanol fractions for both 40% and 100% methanol fractions had displayed an even stronger inhibitory effect on the secretion of melanin. The effect of melanogenesis on B16F1 indicated that both 40% and 100% methanol fractions had shown a decrease in melanin content between 0.025-0.1% (w/v) concentrations which is better than kojic acid. The melanin content for both 40% and 100% methanol fractions were $64.95 \pm 1.87\%$ and $71.91 \pm 1.18\%$, respectively and significantly inhibited the tyrosinase activity in a dose-dependent effect of the L. pumila methanol fractions on the B16F1 melanoma cells. However, the 40% methanol fraction exhibited the highest inhibition up to $73.96 \pm 1.58\%$ of tyrosinase activities. Finally, the study indicates that the L. pumila methanol fractions have good potential as the sources of tyrosinase inhibitor.

ABSTRAK

Perencat tirosinase yang berkesan dan selamat, terutamanya dari sumber semula jadi, sangat dikehendaki bagi penggunaan di dalam produk makanan dan kosmetik pemutihan kulit. Dalam kajian ini, pencirian dan penelitian in-vitro bagi pecahan metanol L. pumila telah dikaji. Pecahan metanol L. pumila telah disediakan dengan menggunakan pengekstrakan fasa pepejal (SPE) dan dilabelkan sebagai 40%, 60% dan 100% pecahan metanol. Keputusan pencirian melalui analisis kandungan jumlah fenol (TPC), 40% dan 100% pecahan metanol telah memberikan jumlah tertinggi kandungan fenolik. Potensi kesan perencatan tirosinase bagi pecahan metanol L. pumila telah dinilai menggunakan sel-sel melanoma B16F1. Kesan sitotoksik pecahan tersebut telah dinilai dengan menggunakan ujian MTT (3-(4, 5 Dimetilthiazol-2-yl)-2,5) dipheniltetrazolium bromida). Keputusan menunjukkan bahawa pada kepekatan sehingga 0.1% w/v bagi pecahan metanol L. pumila tidak menjejaskan kebolehhidupan sel. Rembesan melanin secara beransur-ansur menurun selepas perawatan dengan 0.025-0.1% (w/v) pecahan metanol L. pumila. Pecahan metanol L. pumila untuk kedua-dua iaitu 40% dan 100% menunjukkan kesan perencatan yang lebih kuat pada rembesan melanin. Kesan melanogenesis pada B16F1 menunjukkan bahawa kedua-dua pecahan metanol tersebut mengakibatkan penurunan kandungan melanin antara kepekatan 0.025-0.1% yang lebih baik daripada asid kojic. Kandungan melanin untuk kedua-dua pecahan metanol 40% dan 100% adalah masing-masing $64.95 \pm 1.87\%$ dan $71.91 \pm 1.18\%$ dan secara signifikan menghalang aktiviti tirosinase dengan kebergantungan kesan pecahan metanol L. pumila kepada dos terhadap sel melanoma B16F1. Walau bagaimanapun, pecahan metanol 40% mempamerkan perencatan tertinggi sehingga $73.96 \pm 1.58\%$ bagi aktiviti tirosinase. Secara keseluruhannya, kajian ini menunjukkan bahawa pecahan metanol L. pumila mempunyai potensi yang baik sebagai sumber perencat tirosinase.

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LIST OF ABBREVIATIONS

TPC	-	Total phenolic content
MeOH	-	Methanol
SPE	-	Solid phase Extraction
MTT	-	3-(4,5-Dimethylthiazol-2-yl)-2,5 diphenyl tetrazolium
α-MSH	-	α- Melanocytes-stimulating hormone
MITF	-	Microphtalmia-associated transcription factor
POMC	-	Proopiomelanocortin
MC1R	-	Melanocortin-1 receptor
ATP	-	Adenosine triphosphate
cAMP	-	Cyclic adenosine mono-phosphate
_L -dopa	-	L-3,4-Dihydroxyphenylalanine
TRP	-	Tyrosinase-related protein
DCT	-	Dopachrome tautomerase
DHICA	-	Dihydroxyindole- carboxylic acid
DHI	-	Dihydroxyindole
DMSO	-	Dimetyl sulphoxide
DNA	-	Deoxyribonucleic acid
EDTA	-	Ethylene diamine tetraacetate
UV	-	Ultraviolet
IBMX	-	Isobutyrylxanthine
РКС	-	Protein kinase
ROS	-	Reactive oxygen species
PBS	-	Potassium phosphate buffer saline

CHAPTER 1

INTRODUCTION

1.1 Research background

In recent years, dermatologist and cosmetologist are formulating cosmetics with whitening effect. It has such a big demand especially in the Asian region where people are obsessed with fair and radiant skin. In early 1990s, cosmetic companies had begun to use the term cosmeceuticals to describe skin care products that are claimed to have therapeutic benefits through their bioactive ingredient and exert tangible effects on users. In year 2000, the term cosmeceutic was extended to cover 'the functional cosmetic products' that go above and beyond their intended function by offering an additional therapeutic benefit (Park, 2009).

High awareness among consumers in the past few years has brought about significant development in sales and technology of global cosmetics and toiletries. It may trigger significant growth across markets segments and geographies. The high demands create large opportunities which encourage the emergence of new inventors in this field due to the fast growth of research and development annually (Wang *et al.*, 2006)

One of the primary causes to speed up the market growth in this field is the changing of production unit to be more cost effective as observed to be widely practiced within Asia such as in India and China. Besides, the applications of online selling by retailers can help to speed up the growth of the market. Increasing awareness among consumers towards natural products will force the cosmetics and toiletries manufacturers to go with the flow in order to attract consumers (Transparency Market Research, 2013).

According to Transparency Market Research, development of global cosmetics and toiletries market is affected by economic environment and by the year of 2015 it is estimated to reach USD19.2 billion. However, gaining trust from the consumers is quite challenging but fast development of domestic market can cover up the economic crisis in this field. The market is still at the beginning stage and must be concrete enough for future potential (Transparency Market Research, 2013).

Currently, male products can beat female products and it has experienced quite an important growth in the market. The addition can contribute approximately \$4 billion to the global value size which is 27 billion by 2014 (Carrie, 2010).

High demands of natural products have increased gradually in the European and North American market but the industry is facing high cost challenge. Natural products give high confidence in terms of consumers' health, concerns on environmental, and the carcinogenic nature of the synthesis source. These issues will give major contribution in speeding up the growth of natural products (Transparency Market Research, 2013).

The demand for cosmetics is increasing due to the obsession of having fairer and radiant skin especially in the Asian region. Recently, many whitening products on the shelves have been recalled due to the use of restricted chemical compounds (Eisah, 2013). The main consideration of those called in products is on the safety and effectiveness of the product claims.

In Malaysia it is a trend to have a radiant and fair skin complexion. Most of the products in the local market claim to be effective in lightening the skin. The consumers are also inclined to select natural products over cosmeceutical blending that uses toxic chemicals. Fruits, vegetables and products derived from animals are great sources of antioxidant, rich in substances that can improve human complexion, moisturize skin, attenuate fine lines and wrinkles, and give elasticity to the skin (Ferris, 2007). There is a number of natural ingredients that can be used to remove dark spots and whiten skin (Briganti *et al.*, 2003) and just as many reasons to try them.

There has been a discussion on how to determine the reduction in melanin based on the efficacy of whitening products without human study. Animal tests are also not permitted in most countries. As a result, *in vitro* study of cell culture was identified to be a good alternative to assess the tolerance and efficacy of products to ensure maximum safety (Donald *et al.*, 2006).

Skin pigmentation processes involve the *de novo* synthesis of melanin in melanocytes and transfer of the synthesized melanin packed in melanosome to neighboring keranotycytes, which eventually turns the skin color into a darker tone (Hearing, 2005 and Seiberg *et al.*, 2000). Melanosome contains three types of enzyme, tyrosinase, TRP1, DCT (TRP2) (Kobayashi *et al.*, 2007). Tyrosinase is a rate limiting enzyme involved in melanin synthesis that hydroxylates tyrosinase, a kind of phenylalanine, to L-3, 4-dihydroxyphenylalanine (L-DOPA) and oxidizes L-DOPA to DOPA quinine (Kim *et al.*, 2006). Excessive accumulation of DOPA quinine generated from hydroxylation and oxidation of tyrosinase forms DOPA chrome, which conditionally exhausted systeine, resulted in accumulation of black and brownish pigment called eumelanin. Another type of melanin, pheomelanin is produced through formation of 3 of 5 systeinyl DOPA on condition of existence of

systeine (Kim *et al.*, 2006; Miyamura *et al.*, 2007 and Yamaguchi *et al.*, 2007). These three enzymes determine the types of melanin to eumelanin or pheomelanin (Hearing. 2005). Accordingly, our skin color can be determined by the ratio between the types of melanin, the amount of each type of melanin and the extent of transferring melanosome to keratonocytes (Hamesath *et al.*, 1998).

There are several factors that can modulate the melanin biosynthesis. Melanin biosynthesis can be reduced by avoiding UV exposure, by inhibition of melanocytes metabolism and proliferation (Kim and Uyama, 2005), and by inhibition of tyrosinase or by removal of melanin corneal ablation. Inhibition of tyrosinase oxidation catalyzed by tyrosinase is one of the factors to avoid melanin production (Slominski *et al.*, 2004).

Recently, the demand for natural products that inhibit or prevent skin pigmentation is increasing all over the world. A variety of natural or synthetics substances are currently utilized as ingredients of preparations designed to control hyperpigmentation, but none of these have been proven to be completely satisfactory, either due to limited efficacy or owing to safety concern. For instance, hydroquinone, which has been used widely and until recently was considered the standard depigmenting agent, has now been banned for cosmeticeutical uses in Europe and some Asian countries, and is available only by prescriptions. Kojic acid, also used in skin lightening, is based on blocking the activity of tyrosinase. However, there are reports showing skin sensitization, including dermatitis and cytotoxicity, which have resulted in some countries like Japan, Switzerland and Korea, discontinuity of its use (Serra-Baldrich et. al., 1998). Recently, skin lightening is also a cause of mercury toxicity. The features of mercury toxicity, also known as the 'hatters disease', as immortalized in Alice in Wonderland by Lewis Carroll, consists of psychiatric (disturbance or recent memory, impairment of intellectual function, inattention and depression) and neurological (irritability, memory loss and neuropathies) problems (Dadzie and Petit, 2009).

Thus a study was undertaken to investigate if *L. pumila* possesses any tyrosinase inhibition effects with a view of its possible use as a treatment for hyperpigmentation and its use as a skin whitening agent in cosmetics.

1.2 Problem statement

Over the last few decades, much effort had been exerted to search for tyrosinase inhibitors. A large number of compounds, both from natural products and synthetic compounds were assayed on tyrosinase, but only a limited number of them could fulfill the requirement of efficacy and safety. Common examples of extracts used as whitening is green tea, mulberry, and so on. However, the commercial value of each potential plant is still in a question for its prospects. High efficacy and safety still remain as the main considerations for human use and every active compound identified needs further investigation especially in humans. For those to be applied in food products, it is also important that they do not cause significant negative impact on the sensory properties. Therefore, many plants have been investigated to determine their potential for application as cosmetic agents.

Traditional herbal plants like *L. pumila* are great alternatives since commercial tyrosinase inhibitors such as hydroquinone and kojic acid could bring side effects to human because of their toxicity (Moussy *et al.*, 2004 and Briganti *et al.*, 2003). Previous studies have shown that *L. pumila* contains many phytochemicals that have been identified to exhibit beneficial properties such as antioxidant, antimicrobial and anti-estrogenic diseases (Chua *et al.*, 2012). It has been reported that antioxidant may reduce hyperpigmentation and support skin health (Ma *et al.*, 2001). However, the inhibition activity of tyrosinase by *L. pumila* has not been established.

1.3 Hypothesis

The central hypothesis of this study is that the methanol fractions of *L*. *pumila* exhibit anti- tyrosinase properties.

1.4 Objective of the study

The aim of this study is to establish the tyrosinase inhibition in melanogenesis by *L. pumila* methanol fractions using B16F1 murine melanoma cells.

1.5 Scopes of the study

This study consists of three main scopes. They are outlined as follows:

- 1. To investigate the effect of *L. pumila* methanol fractions on the cells viability of B16F1 murine melanoma cells.
- 2. To determine the concentration effect of *L. pumila* methanol fractions on the formation of melanin content in B16F1 melanoma cells.
- 3. To investigate the concentration effect of *L. pumila* methanol fractions on the inhibition of tyrosinase activity using enzymatic assay.

REFERENCES

- Abdel-Naser, M.B. (1999). Differential effects on melanocytes growth and melanization of low vs high Ca⁺⁺ keratinocytes conditioned medium. *Brit J Dermatol*, 140:50-55.
- Abdel-Naser, M.B. (2003). Mitogen requirements of normal human melanocytes in a serum and tumor promoter free medium. *European Journal of Dermatology*, 13: 29 – 33.
- Abdel-Naser, M.B., Verma, S.B., and Rahim-Abdallah, M.A. (2005). Common dermatoses in moderately pigmented skin: uncommon presentations. *Clinics in Dermatology*, 23(5): 446-456.
- Aburjai, T., and Natsheh, F.M. (2003). Plants used in cosmetics. *Phytotherapy Research*, 17: 987-1000.
- Ahmad, V.U., Ullah, F., Hussain, J., Farooq, U., Zubair, M., Khan, M.T.H., and Choudhary, M.I. (2004). Tyrosinase inhibitors from *Rhododendron collettianum* and their structure-activity relationship (SAR) studies. *Chem. Pharm. Bull.*, 52: 1458-1461.
- Ali, Z., and Khan, I.A. (2011). Alkyl phenols and saponins from the roots of *Labisia pumila* (Kacip Fatimah). *Phytochemistry*, 77: 2075–2080.
- Ando, H., Kondoh, H., and Ichihashi, M. (2007). Approaches to identify inhibitors of melanin biosynthesis via the quality control of tyrosinase. *J Invest Dermatol*, 127:751–761.
- Aoki, Y., Tanigawa, T., Abe, H., and Fujiwara, Y. (2007). Melanogenesis inhibition by an oolong tea extract in B16 mouse melanoma cells and UV-induced skin pigmentation in brownish guinea pigs. *Bioscience, Biotechnology and Biochemistry*, 71(8):1879–1885.
- Arung, E.T., Kuspradini, H., Kusuma, I.W., Shimizu, K., and Kondo, R. (2012). Validation of Eupatorium triplinerve Vahl leaves, a skin care herb from East Kalimantan, using a melanin biosynthesis assay. *Journal of Acupunctute and Meridian Studies*, 5(2): 87–92.

Azhar-ul-Haq, M.A., Khan, M.T.H., Anwar-ul-Haq, K.S.B., Ahmad, A., and Choudhary, M.I. (2006). Tyrosinase inhibitory lignans from the methanol extract of the roots of *Vitex negundo* Linn and their structure-activity relationship. *Phytomedicine*, 13: 255-260.

- Azidah, A.K., and Nik Hazlina, N.H. (2013). Potential role of *Labisa pumila* in the prevention and treatment of chronic diseases. *Journal of Food Research*, 2(4): 55-60.
- Baek, H.S., Rho, H.S., Yoo, J.W., Ahn, S.M., Lee, J.Y., and Lee, J. (2008). The inhibitory effect of new hydroxamic acid derivatives on melanogenesis. *Buletin of the Korean Chemical Society*, 29: 43-46.
- Bagchi, M,. Mark, M., Casey, W., Jaya, B., Xumei, Y., and Sidney, S. (1999). Acute and chronic stress-induced oxidative gastrointestinal injury in rats and the protective ability of a novel grape seed proanthocyanidin extracts. *Nutr. Res.*, 19: 1189-1119.
- Basf. (2000). Melanomatosis and melanin embedding under the influence of UV light. Retrieved on Mei 13, 2013, from http://www.skin-careforum.basf.com/en/author.
- Beulaja, M., and Manikandan, R. (2012). Detection of natural and induced phenoloxidase activities in human serum. *Human Immunology*, 73(10): 1005– 1010.
- Benech, G. (2002). Novo ativo clareador extraído de cítricos. *Cosmetics* & *Toiletries*, 14: 51-53.
- Bhathena, S.J., and Velasquez, M.T. (2002). Beneficial role of dietary phytoestrogens inobesity and diabetes. *The American Journal of Clinical Nutrition*, 76(6): 1191-1201.
- Biesalski, H.K., Springer, M., and Engelhart, K. (2003). Effects of 3-isobutyl-1methylxanthine and kojic acid on cocultures and skin equivalents composed of HaCat cells and human melanocytes. *Archives of Dermatological Research*, 295(2): 88–91.
- Biotage. (2012). *Introduction to solid phase extraction (SPE)*. Retrieved on June 14, 2013, from http://www.biotage.com
- Boissy, R.E., and Manga, P. (2004). Review on the etiology of contact/ occupational vitiligo. *Pigment Cell Res.*, 17: 208-214.

- Briganti, S., Camera, E., and Picardo, M. (2003). Chemical and instrumental approaches to treat hyperpigmentation. *Journal of Pigment Cell Research*, 16: 101 110.
- Burkill, I.H. (1966). A dictionary of the economic product of the Malay Peninsula. Vol.II (I-Z) Government of Malaysia and Singapore by the Ministry of Agricultural and Cooperative, Kuala Lumpur.
- Busca, R., and Ballotti, R. (2000). Cyclic AMP a key messenger in the regulation of skin pigmentation. *Pigment Cell Res.*, 13: 60–69.
- Cabanes, J., Chazarra, S., and Garcia-Carmona, F. (1994). Kojic acid, a cosmetic skin whitening agent, is a slow-binding inhibitor of catecholase activity of tyrosinase. *J. Pharm. Pharmacol*, *46*: 982-985.
- Carrie Lennard. (2010, December 6). Masculine Dynamism—Men's Care Growing Fast. *GCI Magazine*. Retrieved on June 18, 2013, from http://www.gcimagazine.com
- Cerenius, L., and Söderhäll, K. (2004). The prophenoloxidase-activating system in invertebrates. *Immunological reviews*, 198: 116–126.
- Chan, Y.Y., Sim, K.H., and Cheah, S.H. (2011). Inhibitory effects of Sargassum polycystum on tyrosinase activity and melanin formation in B16F10 murine melanoma cell. J. of Ethnopharmacology, 137:1183-1188.
- Chen, J.S., Wei, C.I., and Marshall, M.R. (1991). Inhibition-mechanism of kojic acid on polyphenol oxidase. *J. Agric. Food Chem.*, 39: 1897–1901.
- Chen, L., Jiang, W., and Hou, A. J. (2006). Novel 2-arylbenzofuran derivatives from *Artocarpus petelotii. Helv. Chim. Acta*, 89: 1000-1007.
- Chen, L.G., Chang, W.L., Lee, C.J., Lee, L.T., Shih, C.M., and Wang, C.C. (2009). Melanogenesis inhibition by gallotannins from Chinese galls in B16 mouse melanoma cells. *Biol. Pharm. Bull.*, 32:1447–1452.
- Choi, H.K., Kim, D.H., Kim, J.W., Ngadiran, S., Sarmidi, M.R., and Park, C.S. (2010). *Labisia pumila* extract protects skin cells from photoaging caused by UVB irradiation. *J. Biosci. Bioeng.*, 109: 291-296.
- Chua, L.S., Abdul Latiff, N., Lee, S.Y., Lee, C.T., Sarmidi, M.R., and Abdul Aziz, R. (2011). Flavonoids and phenolic acids from *Labisia pumila* (Kacip Fatimah). *Food Chem.*, 127: 1186-1192.
- Chua, L.S., Sze Yean, Lee., Norhanisah Abdullah, and Mohamad Roji Sarmidi. (2012). Review on *Labisia pumila* (Kacip Fatimah): Bioactive

phytochemicals and skin collagen synthesis promoting herb. *Fitoterapia*, 83: 1322-1335.

- Chung, S.Y., Seo, Y.K., Park, J.M., Seo, M.J., Park, J.K., Kim, J.W., and Park, C.S. (2009). Fermented rice bran downregulates MITF expression and leads to inhibition of α-MSH-induced melanogenesis in B16F1 melanoma. *Biosci.*, *Biotechnol.*, and Biochem., 73(8): 1704-1710.
- Cook, N.C., and Samman, S. (1996). Flavonoids-chemistry, metabolism, cardioprotective effects, and dietary sources. *J. Nutr. Biochem.*, 7: 66-76.
- Costin, G.E., and Raabe, H. (2013). Optimizied in vitro pigmentation screening assay using a reconstructed three dimensional human skin model. *Rom. J. Biochem.*, 50 (1): 15-27.
- Crozier, A., Jaganath, I.B., and Clifford, M.N. (2009). Dietary phenolics: chemistry, bioavailability and effects on health. *Nat. Prod. Rep.*, 26:1001-1043.
- Dadzie, O.E., and Petit, A. (2009). Skin bleaching: Highlithing the misuse of cutaneous depigmenting agents. *Journal of the European Academy of Dermatology and Venereology*, 23(7): 741-750.
- Dai, J., and Mumper, R.J. (2010). Plant phenolics: Extraction, Analysis and their antioxidant and anticancer properties. *Molecules*, 15: 7313-7352.
- Dermamedics. (2013). *Hyperpigmentation: An overview of human pigmentation*. Retrieved on May 12, 2013, from http://www.dermamedics.com/ hyperpigmentation_id60.html
- Donald, M.B., Colin, T., Nathalie, M., and Egbert, A. (2006). *Ethical eye-Animal welfare*. Strasbourg Cedex, France: Council of Europe Publishing.
- Donsing, P., Limpeanchob, N., and Viyoch, J. (2008). Evaluation of the effect of Thai breadfruit's heartwood extract on melanogenesis-inhibitory and antioxidant activities. *J Cosmet Sci.*, 59: 41-58.
- Dodonne, D., Poupard, P., Coutiere, P., Woillez, M., Richard, T., Merillon, J.M., and Vitrac, X. (2011). Phenolic composition and antioxidant properties of poplar bud (*Populus nigra*) extract: Individual antioxidant contribution of phenolics and transcriptional effect on skin aging. J Agric Food Chem., 59: 4527-4536.
- Dooley, T.P. (1997). Topical skin depigmentation agents: current products and discovery of novel inhibitors of melanogenesis. *Journal of Dermatological Treatment*, 8(4): 275-283.

- Duval, C., Schmidt, R., Regnier, M., Facy, V., Asselineau, D., and Bernard, F. (2003). The use of reconstructed human skin to evaluate UV – induced modifications and sunscreen efficacy. *Experimental Dermatology*, 12: 64-70.
- Ehrhardt Proksch, Johanna, M., Brandner, and Jens-Michael Jensen (2008). The Skin: an indispensable barrier. *Experimental Dermatology*, 17: 1063–1072.
- Eisah, A.R. (2013). Kenyataan Akhbar: Pengguna di nasihatkan untuk mengelakkan dari mengguna produk kosmetik yang dikesan mengandungi racun berjadual. Retrieved on April 12, 2013, from http://portal.bpfk.gov.my
- Elias, P.M., Menon, G., Wetzel, B.K., and Williams, J.J. (2009). Evidence that stress to the epidermal barrier influenced the development of pigmentation in human. *Pigment cell Melanoma Res.*, 22(4): 420-434.
- Englaro, W., Bertolotto, C., Busca, R., Brunet, A., Pages, G., Ortonne, J.P., and Balloti, R. (1998). Inhibition of the mitogen activated protein kinase pathway triggers B16 melanoma cell differentiation. *J. Biol. Chem.*, 273:1966-1970.
- Fazliana, M., Gu, H.F., Ostenson, C.G., Yusoff, M.M., and Wan Nazaimoon, W.M. (2012). Labisia pumila extract down-regulates hydroxysteroid (11-beta) dehydrogenase 1 expression and corticosterone levels in ovariectomized rats. Journal of Natural Medicine, 66: 257–264.
- Ferris, P. (2007). Idebenone, green tea, and Coffebery extract: new and innovative antioxidant. *Dermatol Ther.*, 20(5): 322-329.
- Frank, L., Meyskens, J.R., Patrick, F., and John, P. Freuhauf (2001). Redox regulation in human melanocytes and melanoma, *Pigmen Cell Res.*, 14: 148-154.
- Freedberg, I.M., and Fitzpatrick, T.B., (1999). *Fitzpatrick's dermatology in general medicine*. Health Professions Division, New York: McGraw-Hill.
- Freshney, R.I. (2000). *Freshney's culture of animal cell: A manual of basic technique* (2nd ed). Hoboken, N.J.: Wiley publication.
- Freshney, R.I. (2005). Freshney's culture of animal cell: A manual of basic technique (5th ed). Hoboken, N.J.: Wiley publication.
- Garcia-Borron, J.C., Sanchez-Laorden, B.L., and Jimenez-Cervantez, C. (2005). Melanocortin-1 receptor structure and functional regulation. *Pigment Cell Research*, 18: 393-410.
- Greco, G., Panzella, L., Verotta, L., d'Ischia, M., and Napolitano, A. (2011). Uncovering the structure of human red hair pheomelanin:

benzothiazolylthiazinodihydroisoquinolines as key building blocks. *J. Nat. Prod.* 74:675–682.

- Gilchrest, B.A., Park, H.Y., Eller, M.S., and Yaar, M. (1996). Mechanisms of ultraviolet light-induced pigmentation. *Photochemistry and Photobiology*, 63(1): 1–10.
- Grimes, P.E. (1999). The safety and efficacy of salicylic acid chemical peels in darker racial-ethnic groups. *Dermatol. Surg.*, 25: 18-22.
- Gupta, S. (2001). Plant-based Skin Whitening Cosmetics, Happi, 90.
- Gursoy, N., Sarikurkcu, C., Cengiz, M., and Solak, M.H. (2009). Antioxidantactivities, metal contents, total phenolics and flavonoids of seven Morchella species. *Food Chem Toxicol*, 47: 2381-2388.
- Halaban, R., Pomerantz, S.H., Marshall, S., Lambert, D.T., and Lerner, A.B. (1983).
 Regulation of tyrosinase in human melanocytes grown in culture. *Journal of Cell Biology*, 97, 480 488.
- Hamesath, T.J., Price, E.R., Takemoto, C., Badalian, T., and Fisher, D.E. (1998). MAP kinase links the transcriptions factors Microphthalmia to c-Kit signaling in melanocytes. *Nature*, 391(6664): 298-301.
- Hamid, M.A., Sarmidi, M.R., and Park, C.S. (2011). Mangosteen leaf extract increases melanogenesis in B16F1 melanoma cells by stimulating tyrosinase activity *in vitro* and by up-regulating tyrosinase gene expression. *Int. J. Mol. Med.*, 29: 209-217.
- Harborne, J.B, and Williams, C. (2000). Advances in flavonoid research since 1992. *Phytochem* 55: 481–504
- Hearing, V.J., and Jimenez, M. (1987). Mammalian tyrosinase: the critical regulatory control point in melanocyte pigmentation. *International Journal of Biochemical*, 19: 1141 – 1147.
- Hearing, V.J., and Tsukamoto, K. (1991). Enzymatic control of pigmentation in mammals. *FASEB J.*, 14: 2902-2909.
- Hearing, V.J. (2005). Biogenesis of pigment granules: a sensitive way to regulate melanocytes function. *J Dermatol Sci.*, 37(1): 3-14.
- Hermanns, J.F., Petit, L., Martalo, O., Pierard-Franchiomont, C., Cauwenbergh, G., and Pierard, G.E. (2000). Unraveling the patterns subclinical pheomelaninenriched facial hyperpigmentation: effect of dipegmentating agent. *Journal of Dermatology*, 201: 118 – 122.

- Hoffhines, A.D., Damoc, U., Bridges, K.G., Leary, J.A., and Moore, K.L. (2006). Detection and purification of tyrosinase-sulphated proteins using a novel antisulfotyrosinase monoclonal antibody. *Journal of Biological Chemistry*, 281:37877-37887.
- Hirobe, T. (2005). Role of keratinocyte-derived factors involved in regulating the proliferation and differentiation of mammalianepidermal melanocytes. *Journal of Pigment Cell Research*, 18: 2 – 12.
- Holbrook. (1997). KA: Development of human skin. Retinoids, 13: 47–53.
- Hu, D.N. (2008). Methodology for evaluation of melanin content and production of pigment cells *in vitro*. *Photochemistry and Photobiology*, 84(3): 645–649.
- Jablonski, N.G., and Chaplin, G. (2012). Human skin pigmentation and disease susceptibility. *Phil. Trans. R. Soc. B.*, 367: 785-792.
- Jamal, J.A., Houghton, P.J., Miligan, S.R., and Jantan, I. (2003). The oestrogenic and cytotxic effects of the extracts of *Labisia pumila* var. *alata* and *Labisia pumila* var. *pumila* in vitro. *Sains kesihatan*, 1: 53-60.
- Karimi, E., Jaafar, H.Z.E., and Ahmad, S. (2011). Phytochemical analysis and antimicrobial activities of methanolic extracts of leaf, stem and root from different varieties of *Labisia pumila Benth. Molecules*, 16: 4438-4450.
- Karimi, E., and Jaafar H.Z.E. (2011). HPLC and GC-MS determination of bioactive compounds in microwave obtained extracts of three varieties of *Labisia pumila Benth. Molecules*, 16: 6791-6805.
- Karimi, E., Jaafar, H.Z.E., and Ahmad, S. (2013). Antifungal, anti-inflammatory and cytotocity activities of three varieties of *Labisia pumila* benth: from microwave obtained extracts. *BMC Complement Altern Med.*, 13:20.
- Karioti, A., Protopappa, A., Megoulas, N., and Skaltsa, H. (2007). Identification of tyrosinase inhibitors from *Marrubium velutinum* and *Marrubium cylleneum*. *Bioorg. Med. Chem.*, 15: 2708-2714.
- Kasree, B., Handjani, F., and Aslani, F.S. (2003). Enhancement of the depigmenting effect of hydroquinone and 4-hydroxyanisole by all-trans-retinoic acid (tretinoin): the impairment of glutathione-dependent cytoprotection. *Dermatology*, 206: 289-291.
- Khan, K.M., Maharvi, G.M., Abbaskhan, A., Hayat, S., Khan, M.T.H., Makhmoor, T., Choudhary, M.I., Shaheen, F., and Rahman, A. (2003). Three tyrosinase

inhibitors and antioxidant compounds from *Salsola foetida*. *Helv. Chim. Acta,* 86: 457-462.

- Kim, Y.J., and Uyama, H. (2005). Tyrosinase inhibitors from natural and synthetic sources: structure, inhibition mechanism and perspective for the future. *Cellular and Molecular Life Sciences*, 62: 1707 – 1723.
- Kim, D.S., Park, S.H., Kwon, S.B., Park, E.S., Huh, C.H., Youn, S.W., and Park, K.C. (2006). Sphingosylphosphorylcholine-induced ERK activation inhibits melanin synthesis in human melanocytes. *Pigment Cell Res.*, 19(2): 146-153.
- Kim, E.O., Choi, S.W., and Lee, S.K. (2007). Antioxidant and antimelanogenic activities of polyamine conjugates from corn bran and related hydroxycinnamic acids. *Journal of Agricultural and Food Chemistry*, 55(10): 3920–3925.
- Kim, Y.J., Kang, K.S., and Yokozawa, T. (2008). The anti –meanogenic effect of pycnogenol by its anti-oxidative action. *Food and chemical toxicology*, 46: 2466-2471.
- Kobayashi, T., and Hearing, V.J. (2007). Direct interaction of tyrosinase with Tyrpl to form heterodimeric complexes in vitro. *J Cell Sci.*, 120(24): 4261-4268.
- Krishna, P.L., Bhat, J.W., and Kosmeder II, J.M.P. (2001). Biological effects of resveratrol. Antioxid. Redox Signal, 3: 1041-1064.
- Kubo, I., Kinst-Hori, I., Chaudhuri, S.K., Kubo, Y., Sánchez, Y., and Ogura, T. (2000). Flavonols from *Heterotheca inuloides*: tyrosinase inhibitory activity and structural criteria. *Bioorg. Med. Chem.*, 8:1749–1755.
- Kubo, I., Masuoka, N., Ha, T.J., and Tsujimoto, K., (2006). Antioxidant activity of anacardic acids. *Food Chemistry*, 99: 555–562.
- Kumaran, A. and Karunakaran, R.J. (2007). In vitro antioxidant activities of methanol extracts of five Phyllanthus species from India. *LWT-Food Science* and Technology, 40: 344–352.
- Lam, P. K., (1999). Evaluation of Human Skin Substitute for Burn wound Coverage based on Cultured Epidermal Autograft. Doctor Philosophy, The Chinese University of Hong Kong.
- Larsen, W.J. (1993). Human embryology. New York: Churchill Livingstone.
- Lassalle, M.W., Igarashi, S., Sasaki, M., Wakamatsu, K., Ito, S., and Horikoshi, T. (2003) Effects of melanogenesis-inducing nitric oxide and histamine on the

production of eumelanin and pheomelanin in cultured human melanocytes. *Pigment Cell Res.*, 16: 81–84.

- Lee, J.H., Park, H., and Chung, H. (2009). Syndecan-2 regulates the migratory potential of melanoma cells. *Journal of Biological Chemistry*, 284(40): 27167–27175.
- Lee, K.K, and Choi, J.D. (1999). The effecta of *Areca catechu L* extract on antiinflammation and anti-melanogenesis. *Int. J. Cosmet. Sci.*, 21(4): 275-284.
- Lee, M.H., Lin, Y.P., Hsu, F.L., Zhan, G.R., and Yen, K.Y. (2006). Bioactive constituents of *Spatholobus suberectus* in regulating tyrosinase-related proteins and mRNA in HEMn cells. *Phytochemistry*, 67: 1260-1270.
- Lee, K.T., Kim, B.J., Kim, H.J., Heo, M.Y., Kim, H.P. (1997). Biological screening of 100 plant extracts for cosmetic use (I): Inhibitory activities of tyrosinase and DOPA auto-oxidation. *Int J Cosmet Sci.*, 19: 291–298.
- Lin, V.C., Ding, H.Y., Tsai, P.C., Wu, J.Y., Lu, Y.H., Chang, T.S.(2011). In Vitro and in Vivo melanogenesis inhibition by biochanin a from Trifolium pratense. Bioscience, Biotechnology and Biochemistry, 75(5): 914–918.
- Liu, Y., Hong, L., Kempf, V.R., Wakamatsu, K., Ito, S., and Simon, J.D. (2004). Ionexchange and adsorption of Fe(III) by Sepia melanin. *Pigment Cell Research*, 17(3): 262–269.
- Li, S.B., Xue, Y., Lv, X.Y., Nie, H.L., Zhu, L.M., Zhang, H.T., Qiu, T., and Zhou, L.M. (2009). *In vitro* effect of Ozagrel on Mushroom tyrosinase. *Protein J.*, 28(3-4): 182-188.
- Ma, W., Wlaschek, M., Tantcheva-Poor, I., Schneider, L.A., and Naderi, L. (2001). Chronological aging and photoageing of the fibroblasts and the dermal connective tissue. *Clin. Exp. Dermatol.*, 26: 592-599.
- Madani, W., Kermasha, S., and Bisakowki, B. (1999). Inhibition of tyrosinase activity by polyphenol esterase using selected phenolic substances. *Phytochemistry*, 52: 1001-1008.
- Marieb, E.N. (1997). *Essentials of human anatomy and physiology (5th Edi.)*. Menlo Park, California: Addison Wesley Longman.
- Mariani Abdul Hamid. (2012). Study on Malaysian tropical plantmodulate melanogenesis in B16F1 melanoma cells. Doctor Philosophy, Dongguk University, Seoul.

- Matsuda, H., Higashino, M., Chen, W., Tosa, H., Iinuma, M., and Kubo, M. (1995).
 Studies of cuticle drugs from natural sources. III. Inhibitory effect of *Myrica rubra* on melanin biosynthesis. *Biol. Pharm. Bull.*, 18: 1148–1150.
- Miyamura, Y., Coelho, S.G., Wolber, R., Miller, S.A., Wakamatsu, K., Zmudzka, B.Z., Ito, S., Smuda, C., Passeron, T., Choi, W., Batzer, J., Yamaguchi, Y., Beer, J.Z., and Hearing, V.J. (2007). Regulation of human skin pigmentation and responses to ultraviolet radiation. *Pigment Cell Research*, 20(1): 2-13.
- Mohd Fuad, W.E., Sulaiman, S.A., Islam, M.N., Abdul Wahab, M.S., and Jamalullail, S.M.S. (2005). Evaluation of the teratogenicity of aqueous extract of *Labisia pumila* var. *alata* in rats. *Malaysia J Med Sci*, 12: 13-21.
- Mohd Mukrish bin Mohd Hanafi @ Hanafiah (2012). The photoprotective and collagen stimulatory effects of Labisia pumila extract on UVB irradiated human skin fibroblast (HSF1184) cells. Master, Universiti Teknologi Malaysia, Skudai.
- Mousy., Alain., Kinet., and Jean Pierre. (2004). *European Patent No. EP1427422*. Retrieved on August 5, 2007 from http://www.freepatentonline.com
- Murphy, G. F. (1995). *Dermatopathology, A practical guide to common disorders*. *1st edition*. Philadelphia, Pennyslavania: W.B. Sanders Company,
- Nerys Ohad., Ramadan Musa., Soliman Khatib., Snait Tamir., and Jacob Vaya (2004). Chalcones as potent tyrosinase inhibitors: the effect of hydroxyl positions and numbers. *Journal of Phytochemistry*, 65: 1389 1395.
- Ni, G., Zhang, Q.J., Zheng, Z.F., Chen, R.Y., and Yu, D.Q. (2009). 2-Arylbenzofuran derivatives from *Morus cathayana*. J. Nat. Prod., 72: 966-968.
- Ngoc, T.M., Lee, I., Ha, D.T., Kim, H.J., Min, B.S., and Bae, K.H. (2009). Tyrosinase-inhibitory constituents from the twigs of *Cinnamonum cassia*. J. *Nat. Prod.*, 72: 1205-1208.
- Nicolaus, R.A., Piattelli, M., and Fattorusso, E. (1964) The structure of melanins and melanogenesis-IV: On some natural melanins. *Tetrahedron*, 20: 1163–1172.
- Norhaiza, M., Maziah, M., and Hakiman, M. (2009). Antioxidative properties of leaf extracts of a popular Malaysian herbs, *Labisia pumila*. *J Med Plant Res.*, 3: 217-223.

- Nurhanani, R., Rasyidah, R., Sarni, M.J., and Azlina A.A. (2008). Radical scavenging and reducing properties of extracts of cashew shoots (*Anacardium occindentale*). *Journal of Food Chemistry*, 111: 38 44.
- Ohguchi, K., Tanaka, T., Ito, T., Iinuma, M., Matsumoto, K., Akao, Y., and Nozawa, Y. (2003). Inhibitory effects of resveratrol derivatives from dipterocarpaceae plants on tyrosinase activity. *Biosci. Biotechnol. Biochem.*, 67: 1587-1589.
- Ohguchi, K., Banno, Y., Akao, Y., and Nozawa, Y. (2004). Involvement of phospholipase D1 in melanogenesis of mouse B16 melanoma cells. *Journal* of Biological Chemistry, 279(5): 3408–3412.
- Oh, M.J., Abdul Hamid, M., Ngadiran, S., Seo, Y.K., Sarmidi, M.R., and Park, C.S. (2011). Ficus deltoidea (Mas Cotek) extract exerted anti-melanogenic activity by preventing tyrosinase activity in vitro and suppressing tyrosinase gene expression in B16F1 melanoma cells. *Arch Dermatol Res.*, 303: 161-170.
- Okombi Sabrina., Delphine Rival., Sebastian Bonnet., Anne-Marie Marotte., eric Perrier., and Ahcene Boumendjel. (2006). Analogues of N-hydroxy cinnamoyl phenalkylamides as inhibitors of human melanocyte-tyrosinase. *Journal of Bioorganis and Medicinal Chemistry Letters*, 16: 2252 – 2255.
- O'neil, D. (2013). *Skin color adaptation*. Retrieved on Januari 26, 2013, from http://shttp://anthro.palomar.edu/adapt/adapt_4.htm
- Ortonne, J.P., and Bissett, D.L. (2008). Latest insights into skin hyperpigmentation. Journal of Investigative Dermatology Symposium Proceedings, 13(1):10–14.
- Pandey, G., and Madhuri, S. (2009). Some medicinal plants as natural anticancer agents. *Pharmacogn Rev.*, 3: 259-263.
- Park, C.S. (2009). New generation of anti-ageing cosmeceutical. 2nd International conference on biotechnology for the wellness industry. 23rd – 26th July 2009. Kuala Lumpur, 23.
- Park, Y.D., So-Yeon Kim., You-Jeong Lyou., Dong-Youn Lee., and Jun-Mo Yang. (2006). TXM13 human melanoma cells: a novel source for the inhibition kinetics of human tyrosinase and for screening whitening agents. *Journal of Biochemical Cell Biology*, 84: 112 – 116.
- Park, H.Y., Wu, C., Yonemoto, L., Murphy-Smith, M., Wu, H., Starchur, C.M., and Gilchrest, B.A. (2006) MITF mediates cAMP-induced protein kinase C-beta expression in human melanocytes. *J, Biochem.*, 395: 571-578.

- Perluigi, M., De Marco., Foppoli, F., Coccia, C., Balrzino, R., Marcante, C., and Cini, M.L. (2003) Tyrosinase protects human melanocytes from ROSgenerating compounds. *Journal of Biochemical and Biophysical Research Communication*, 305: 205 – 256.
- Phetdee, K., Rattnamanee, K., Teaktong, T., and Voyoch, J. (2012). Tamarind seed coat extract reduce melanin production via tyrosinase in melanocytes. *Biological sciences*, 12(4): 239-245.
- Piao, X.L., Baek, S.H., Park, M.K., and Park, J.H. (2004). Tyrosinase-inhibitory furanocoumarin from *Angelica dahurica*. *Biol. Pharm. Bull.*, 27: 1144-1146.
- Prasad, N.K., Yang, B., Dong, X., Jiang, G., Zhang, H., and Xie, H. (2009). Flavonoid contents and antioxidant activities from Cinnamomum species. *Innovat. Food Science and Emerging Technologies*, 10: 627–632.
- Prota, G., Novellino, L., and Napolitano, A. (1999). 5,6-Dihydroxyindoles in the fenton reaction: a model study of the role of melanin precursors in oxidative stress and hyperpigmentary processes. *Chemical Research in Toxicology*, 12(10): 985–992.
- Roa, B. (2003). Bioactive phytochemicals in Indian foods and their potential health promotion and disease prevention. *Asia pacific Journal of Clinical Nutrition*, 12: 9-22.
- Runnie, I., Salleh, M.N., Mohamed, S., Head, R.J., and Abeywardena, M.Y. (2004). Vasorelaxation induced by common edible tropical plant extracts in isolated rat aorta and mesenteric vascular bed. *J. Ethnopharmacol*, 92: 311-316.
- Rusciano, D., Lorenzoni, P., and Burger, M.M. (1999). Regulation of c-met expression in B16 murine melanoma cells by melanocyte stimulating hormone. *Journal of Cell Science*, 112(5): 623–630.
- Sanchez-Ferrer, A., Rodriguea-Lopez, J.N., Garcia-Canovas, F., and Garcia-Carmona, F. (1995). Tyrosinase: a comprehensive review of its mechanism. *Biochim Biophys Acta*, 1247: 1-11.
- Sato, K., and Toriyama, M. (2009). Depigmenting effect of Catechins. *Molecules*, 14(11): 4425-4432.
- Schallreuter, K.U., and Wood, J.W. (1990). A possible mechanism of action for azelaic acid in the human epidermis. *Arch. Dermatol. Res.*, 282: 168-171.
- Serra-Baldrich, E., Tribo, M.J., and Camarasa, J.G. (1998). Allergic contact dermatitis from kojic acid. *Contact Dermatitis*, *39*: 86-87.

- Sheila, M. (2008). Biometerials for tissue engineering of skin. *Materialstoday*, 11(5): 26-35.
- Sieberg, M., Paine, C., Sharlow, E., Andrade-Gordon, Costanzo, P., Eisniger, M., and Saphiro, M. (2000). Inhibition of melanosome transfer results in skin whitening. *Journal of Investigative Dermatology*, 115: 162 – 167.
- Singh, G.D., Ganjoo, M., Yousouf, M.S., Koul, A., Sharma, R., Singh, S., Sangwan, P.L., Kaol, S., Ahamad, D.B., and Johri, R.K. (2009). Sub-acute toxicity evaluation of an aqueous extract of *Labisia pumila*, A Malaysian herb. *Food Chem Toxical*, 47: 2661-2665.
- Slominski, A., Tobin, D.J., Shibahara, S., and Wortsman, J. (2004). Melanin pigmentation in mammalian skin and its hormonal regulation. *Physiol. Rev.*, 84: 1154-1228.
- Son, J.K., Park, J.S., Kim, J.A., Kim, Y., Chung, S.R., and Lee, H.S. (2003). Prenylated flavonoids from the roots of *Sophora flavescens* with tyrosinase inhibitory activity. *Planta Med.*, 69: 559-561.
- Stone, B.C. (1988) J. Malayan Nat., 42: 43-51.
- Sugumaran, M. (2002). Comparative biochemistry of eumelanogenesis and the protective roles of phenoloxidase and melanin in insects. *Pigment Cell Research*, 15: 2-9.
- Sunarno, B. (2005). Revision of the genus *Labisia* (Myrsinaceae). *Blumea*, 50:579-597.
- Supelco, (1998). *Guide to solid phase extraction*. [Brochure]. Bellefonte, PA: Sigma-Aldrich.
- Te-Sheng Chang,. (2009). An update review of tyrosinase inhibitors. *International Journal of molecular sciences*, 10: 2440-2475.
- Toriyama, M., Sato, K., Takahashi, H., and Iraha, R. (2008). Down-regulation of tyrosinase expression by acetylsalicylic acid in murine B16 melanoma. *Biol Pharm Bull*, 31(1): 33–37.
- Transperancy Market Research (2013). Global Cosmetic & Toiletries Market Analysis and Forecast, 2011 – 2017. Retrieved on July 14, 2013, from http://transparencyresearch.wordpress.com.
- Veitch, N.C., and Grayer, R.J. (2008). Flavonoids and their glycosides, including anthocyanins. *Nat. Prod. Rep.*, 25: 555-611.

- Victor, F.C., Gelber, J., and Roa, B. (2004). Melasma: a review. *Journal of Cutaneous Medicine and Surgery*, 8: 97-102.
- Wan Ezumi, M.F., Siti Amrah, S., Suhaime, A.W.M., and Mohsin, S.S.J. (2007). Evaluation of the female reproductive toxicity of aqueous extracts of *Labisia pumila* var. *alata* in rats. *Indian J Pharmacol*, 39: 30-32.
- Wang, L., and Weller, C.L. (2006). Recent advances in extraction of nutraceuticals from plants. *Trend in Food Science & Technology*, 17: 300-312.
- Wang, K.H., Lin, R.D., Hsu, F.L., Huang, Y.H., Chang, H.C., Huang, C.Y., and Lee, M.H. (2006). Cosmetic applications of selected traditional Chinese herbal medicines. J. Ethnopharmacol, 106: 353–359.
- West MD. (1994). The cellular and molecular biology of skin aging. *Arch Dermatol*, 130: 87-95.
- Xie, L.P., Chen, Q.X., Huang, H., Wang, H.Z., and Zhang, R.Q. (2003). Inhibitory effects of some flavonoids on the activity of mushroom tyrosinase. *Biochemistry*, 68: 487–491.
- Yamaguchi, Y., Brenner, M., and Hearing, V.J. (2007). The regulation of skin pigmentation. *Journal of Biological Chemistry*, 282(38): 27557-27561.
- Yanez, J., Vicente, V., Alcaraz, M., Castillo, J., Garcia, O.B., Canteras, M., and Taruel, J.A.L. (2004). Cytotoxicity and proliferation activities of several phenolic compound against three melanocytes cell lines: Relationship between structure and activity. *Nutrition and canser*, 49(2): 191-199.
- Yokata, T., Nishio, H., Kubota, Y., and Mizoguchi, M. (1998). The inhibitory effect of glabridin from licorice extracts on melanogenesis and inflammation. *Pigment Cell Research*, 11: 355-361.
- Yoo, K.H., Choi, Y.K., and Rho, Y.K. (2010). Effects of vitamin C vs. multivitamin on melanogenesis: comparative study in vitro and in vivo. *International Journal of Dermatology*, 49(2): 218–226.
- Yoon, T.J., Lei, T.C., Yamaguchi, Y., Batzer, J., Wolber, R., and Hearing, V.J. (2003). Reconstituted 3-dimensional human skin of various ethnic origins as an in vitro model for studies of pigmentation. *Anal Biochem*, 318: 260-269.
- Yoshinori, M., Coelho, S.G., Wolber, R., Miller, S.A., Kazumasa, W., Zmudzka, B.Z., Shosuke, I., Smuda, C., Passeron, T., Choi, W., Batzer, J., Yuji, Y., Beer, J.Z., and Hearing, V.J. (2007). Regulation of human skin pigmentation and responses to ultraviolet radiation. *Pigment Cell Research*, 20(1): 2-13.

- Zheng, Z.P., Cheng, K.W., James To, T.K., Li, H., and Wang, M. (2008). Isolation of tyrosinase inhibitors from *Artocarpus heterophyllus* and use of its extract as antibrowning agent .*Mol. Nutr. Food Res.*, 52:1530 – 1538.
- Zheng, Z.P., Zhu, Q., Fan, C.L., Tan, H.Y., and Wang, M. (2011). Phenolic tyrosinase inhibitors from the stems of *Cudrania cochinchinensis*. *Food Funct.*, 2: 259-264.