LIPID PRODUCTION FROM PALM OIL MILL EFFLUENT BY MICROALGAE

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LIPID PRODUCTION FROM PALM OIL MILL EFFLUENT BY MICROALGAE

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To my beloved family

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ABSTRACT

In tropical countries, the palm oil industry discharges a large amount of wastewater. The wastewater can serve as an economical nutrient source or substrate that can support the cultivation of microalgae. This study aimed to identify the local species of microalgae potentially existing in the industrial wastewater of palm oil mill effluent (POME). POME was selected as the key source of waste due to its higher potential in producing lipids from microalgae as biofuel substrate. A novel green microalgal strain was isolated from POME of Kulai-Johor west Palm Oil Mill in Malaysia and was identified as Chlamydomonas sp. and subsequently named Chlamydomonas sp. UTM 98 with Catalogue No. of KR349061. This strain was cultivated in media with different volume ratios of POME and Basal Bald Medium (BBM). Lipid is generally a group of organic compound that serves as the primary raw material for biofuel. Therefore, this study emphasizes the effectiveness of POME as the main carbon source to maintain the growth of microalgae and simultaneously to increase the lipid content. In addition, glucose (C₆H₁₂O₆) was also used to compare the effectiveness of their cultivations against POME. Furthermore, four selected strains of green microalgae are applied namely Chlorella vulgaris, Chlorella pyrenoidosa, Chlorella sorokiniana, Tetraselmis sp and isolated microalgae from POME. All cultivation of microalgae were initially carried out in 250mL Erlenmeyer flask containing 100 mL medium at ± 30°C with continuous illumination (± 14 µmol⁻¹ m⁻² s⁻¹) and up to 20 days of cultivations. The study demonstrated that Chlamydomona incerta (C. incerta) is the predominant species for specific growth rate (µ), biomass productivity and lipid content in the diluted POME with the value of 0.099/d, 8.0 mg L⁻¹.d, 2.68 mg lipid mg⁻¹ Cell Dry Weight (CDW), respectively. However, C. incerta showed that there was about one and the half times more lipid productivity when the biomass cells utilized glucose as carbon source, compared to POME. The best condition was determined with various carbon-to-total nitrogen (C:TN) ratio and light/dark (L:D) cycles, respectively. As a result, the highest lipid content was achieved when the condition was controlled at C:TN (100:7) and with continuous light (24 hr) which recorded a value of 17 mg lipid mg⁻¹ CDW. These results concluded that C. incerta had the highest growth rates and lipid production in the diluted POME compared to other strains of microalgae. Finally, the study suggested several improvement of the experiment to achieve higher lipid production at steady state condition by manipulating the ratio of carbon-to-total nitrogen and the light intensity on the bio-substrate. The Nile Red method was used to measure the lipid content in the culture. Fatty Acid Methyl Esters (FAMEs) and samples were analyzed via gas chromatography. POME with COD 250mg L⁻¹ concentration showed the greater lipid content with absorbance 3.138a.u. The result showed that Chlamydomonas sp UTM 98 grown in the media of diluted POME exhibited a high potential of microalgae for biomass production and POME nutrients removal.

ABSTRAK

Di negara tropika, industri minyak kelapa sawit melepaskan sejumlah besar air sisa. Air sisa boleh menjadi sumber nutrien ekonomi atau substrat yang boleh menyokong penanaman mikroalga. Kajian ini bertujuan untuk mengenal pasti spesies tempatan mikroalga yang berpontensi dan sedia ada dalam air sisa perindustrian kilang minyak sawit (POME). POME telah dipilih sebagai sumber utama sisa kerana potensinya yang lebih tinggi dalam menghasilkan lipid dari mikroalga yang boleh dijadikan bahan api bio-substrat. Sejenis mikroalga hijau yang baru ditemui telah diasingkan daripada POME Kilang Minyak Sawit barat Kulai-Johor di Malaysia dan telah dikenal pasti sebagai Chlamydomonas sp. dan dinamakan Chlamydomonas sp. UTM 98 dengan No. Catalogue KR349061. Strain ini telah dikultur dalam media POME dan Basal Bald Medium (BBM) dengan nisbah isipadu yang berbeza. Lipid merupakan sebatian organik yang penting sebagai bahan mentah utama banhan api. Oleh itu, kajian ini memberi penekanan kepada keberkesanan POME sebagai sumber karbon utama untuk mengekalkan pertumbuhan mikroalga dan juga untuk meningkatkan kandungan lipid. Di samping itu, glukosa (C₆H₁₂O₆) juga digunakan untuk membandingkan keberkesanan pengkulturan mikroalga berbanding POME. Tambahan pula, empat jenis mikroalga iaitu Chlorella vulgaris, Chlorella pyrenoidosa, Chlorella sorokiniana, Tetraselmis sp dan microalga yang diasing dari POME. Semua pengkulturan mikroalga telah dilakukan di 250 mL kelalang Erlenmeyer yang mengandungi 100 mL media pada ± 30°C suhu dengan pencahayaan yang berterusan (± 14 μmol⁻¹ m⁻² s⁻¹) untuk selama 20 hari. Kajian ini menunjukkan bahawa Chlamydomonas incerta (C. inserta) adalah spesies utama untuk kadar pertumbuhan spesifik (µ), produktiviti biojisin dan kandungan lipid pada kadar pencairan POME adalah masing-masing pada 0.099 / d, 8.0 mg L⁻¹.d, 2.68 mg lipid mg-1 Berat Sel Kering (CDW). Walau bagaimanapun, Chlamydomona incerta menunjukkan bahawa terdapat kira-kira satu setengah kali lebih produktiviti lipid apabila glukos digunakan sebagai sumber karbon, berbanding POME. Keadaan terbaik ditentukan pada pelbagai nisbah karbon / nitrogen (C: TN) dan kitaran cahaya / gelap (L: D). Hasilnya, kandungan lipid yang paling tinggi dicapai apabila keadaan dikawal pada C: TN (100: 7) dan cahaya yang berterusan (24 jam), di mana nilai yang direkodkan sebanyak 17 mg lipid mg-1 CDW. Keputusan ini menyimpulkan bahawa C. incerta mempunyai kadar pertumbuhan yang paling tinggi dan pengeluaran lipid dalam POME yang dicairkan berbanding mikroalga yang lain. Akhir sekali, kajian ini mencadangkan beberapa penambahbaikan eksperimen untuk mencapai pengeluaran lipid yang tinggi pada keadaan mantap dengan memanipulasi nisbah karbon/ nitrogen dan keamatan cahaya ke atas biosubstrat. Kaedah Merah Nile telah digunakan untuk mengukur kandungan lipid dalam media. Fatty Acid Methyl Esters (FAMEs) dan sampel dianalisis melalui kromatografi gas. POME pada kepekatan COD 250mg L⁻¹ menunjukkan kandungan lipid yang lebih besar dengan nilai absorbance 3.138a.u. Hasilnya menunjukkan Chlamydomonas sp. UTM 98 yang dikultur dalam media POME yang dicairkan menunjukkan potensi yang tinggi untuk pengeluaran biojisim dan penyingkiran POME nutrien.

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LIST OF SYMBOLS

 α - alpha

 $C_6H_{12}O_6$ - glucose

CH₄ - methane gas

CI - concentration of lipids at the end of batch run

CO₂ - carbon dioxide

d - day

et.al. - and others

g - gram

g/L.d - gram per liter per day

H₂O - water

H₂S - hydrogen sulphide
μ - specific growth rate

X - biomass

X_m - concentration of biomass at the end of batch run

X₀ - concentration of biomass at the beginning of a batch run

Km - Michaelis-Menten constant, expressed in the same units as X

Ki - dissociation constant for substrate binding in such a way that two

substrates can bind to an enzyme

LN - natural Log m³ - cubic meter

P_{Biomass} - biomass Productivity (mg/L.d)

P_{Lipid} - lipid Productivity (mg/L.d)

t - time, duration (hour,day)

v/v - volume per volume

Vmax - maximum enzyme velocity

LIST OF ABBREVIATIONS

AN - Ammonical nitrogen

ATP - Adenosine TriPhosphate

BBM - Bald's Basal Medium

BOD - Biological Oxygen Demand

C - Carbon

CDW - Cell Dry Weight

CPH - Corn Powder Hydrolysate

COD - Chemical Oxygen Demand

DNA - Deoxyribonucleic acid

EQA - Environmental Quality Acts

FFB - Fresh fruit brunches

LD - Light/dark cycle

MLSS - Mixed Liquor Suspended Solid

MLVSS - Mixed Liquor Volatile Suspended Solid

N - Nitrogen

NADPH - Nicotinamide Adenine Dinucleotide Phosphate-oxidase

nm - nanometer

POME - Palm Oil Mill Effluent
OLR - Organic Loading Rate

OMW - Olive Mill Waste

O&G - Oil & Gas

PAR - Photosynthetically active radiance

RPM - Revolutions per minute (r/min)

S - Substrate

SS - Suspended solid

TCA - Tricarboxylic acid cycle

TS - Total Solid

TSS - Total Suspended Solid

TN - Total Nitrogen

VFA - Volatile fatty acids

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CHAPTER 1

INTRODUCTION

1.1 Introduction

Oil palm is one of the world's most rapidly expanding equatorial crops. In Malaysia, palm plantation currently occupies the largest acre of farmed land and the palm oil industry is growing rapidly. Malaysia is one of the major palm oil producers in the world (Lam *et al.*, 2009; MPOB, 2012). It was estimated that for each tonnes of crude palm oil produced, 5-7.5 tonnes of water are required, and more than 50% of the water will end up as palm oil mill effluent (POME) (Wu *et al.*, 2007 and Singh *et al.*, 2010). While the palm oil industry has been recognized strongly for its contribution towards economic growth and rapid development, it has also contributed to environmental pollution due to the production of large quantities of by products during the process of oil extraction (Singh *et al.*, 2010; Parthasarathy *et al.*, 2016).

Freshwater microalgae are globally ubiquitous and highly deserve, with tens or perhaps hundreds of thousands of species, in a myriad of forms and sizes (Guiry and Guiry, 2014). Current classifications consider most microalgae to be protists with chloroplast, but there also photosynthetic prokaryotes (cyanobacteria) and a subset of land plants (Wehr *et al.*, 2015).

Microalgal culture has received much attention, given its prospects as a source of bioenergy and its potential for wastewater treatment. In this respect, simple and easily cultivated biomass has a number of applications, ranging from its direct use such as biodiesel and various pigments (Fulton, 2004). The demand for these

products is increasing due to their adverse properties, which are economically and environmentally viable options. The complication of cultivation methods and the high cost of growth medium have become a major drawback for the algal industry; nevertheless, the integration along with wastewater treatment has provided a feasible solution due to the fact that exploitation of wastewater as the source of growth medium simultaneously eliminates the requirement for expensive medium and at the same time remediates the wastewater. Recently, much attention has been given to the production of biodiesel from all over the world. The world production of biodiesel was appraised to be around 1.8 billion liters annually (Fulton, 2004; Gnansounou and Raman, 2016). It is also found that biodiesel can be produced using vegetable oils of terraneous oil plants, such as soybean, sunflower and oil palm. However a recent development in the production of biodiesel from microalgae is highly needed to be competitive in the fuel industry.

The current study, investigated the potential, benefits, strategies, and challenges of microalgae to be integrated with wastewater treatment, POME treatment in Malaysia due to the hazardous properties of POME, which may lead to severe environmental pollution. The integration of POME treatment using microalgal culture will potentially reduce the wastewater treatment retention time and eliminate toxic elements, which serve as nutrients for the growth of microalgae. Moreover, microalgae are gaining considerable attention as a feedstock for biodiesel production as they can be grown away from the croplands and hence do not compromise food crop supplies (Liu *et al.*, 2008). The exploration of new microalgae integration methods for the development and formation of valuable products is also being discussed in this study.

1.2 Background of Study

Palm oil mill Effluent (POME) is the wastewater generated by processing oil palm and consists of various suspended materials. POME has a very high biochemical oxygen demand (BOD) and chemical oxygen demand (COD), which is 100 times more than the municipal sewage. The effluent also contains higher

concentration of organic nitrogen, phosphorus, and other nutrient contents (Hadiyanto, 2013). POME is a non-toxic waste, but will pose environmental issue due to large oxygen depleting capability in aquatic system due to organic and nutrient contents. It is also known to be a good source of nutrients (Kamyab *et al.*, 2014a).

The waste products generated during palm oil processing consists of oil palm trunks (OPT), oil palm fronds (OPF), empty fruit bunches (EFB), palm pressed fibers (PPF) and palm kernel shells, less fibrous material such as palm kernel cake and liquid discharge palm oil mill effluent (POME) (Singh *et al.*, 2010). The wastes are in the form of high organic matters concentration, such as cellulosic wastes with a mixture of carbohydrates and oils. The discharge of untreated POME creates adverse impact to the environment (Abdul Aziz, 2007).

Nowadays biological process is the most common practice for the treatment of POME based on anaerobic and aerobic ponding system (Singh *et al.*, 2010). While the emerging technologies for the treatment of POME, the notion of nurturing POME and its derivatives as valuable resources should not be dismissed. Furthermore, it is necessary to properly address the POME treatment so as not to contribute to human health hazards and environmental pollution. When compared to the conventional wastewater treatment process which introduces activated sludge and biological floc to degrade organic carbonaceous matter to CO₂, microalgae can assimilate organic pollutants into cellular constituents such as lipid and carbohydrate, thus achieving pollutant reduction in a more environmental friendly way (de Andrade *et al.*, 2016). In fact, microalgae have become the focus of attention for both wastewater treatment and biomass production as early as 1950s (Oswald and Gotaas, 1957).

Microalgae can assist the treatment and purification of wastewater such as municipal, animal and industrial runoff, while benefiting from using the nutrients. For several years, it has been studied that microalgae have the potential to remove organic and inorganic matters present in the polluted water. It is also concluded that this method is an economic method for removing inorganic and organic materials

from the wastewater, resulting in better quality water discharge and obtaining valuable algal biomass which could be useful for different purposes such as the production of biofuel, fertilizer, animal feedstock, biogas etc., (Becker, 2007; Gonçalves *et al.*, 2016).

Several studies have shown that microalgae are able to remove nitrogen and phosphorus from the wastewater (Aslan and Kapdan, 2006; Gonçalves et al., 2016; Lu et al., 2016). Microalgae grow rapidly; able to divide once every 3-4 h, but mostly would divide every 1-2 days under favorable growth conditions (Griffiths and Harrison, 2009; Huang et al, 2010). Moreover, parameters such as temperature, irradiance and, most markedly, nutrient availability have been shown to affect both lipids composition and content in many microalgae (Rodolfi et al., 2009; Karpagam et al., 2015; Suganya et al., 2016). The average lipid content of algal cells varies between 1% and 70%, but can reach 90% of dry weight under certain conditions (Xin et al., 2010; Lordan and Stanton 2011). The total content of lipids in microalgae may vary from about 1-85% of the dry weight, with values higher than 40% being typically achieved under nutrient limitation. The interest in microalgae for biodiesel production is due to the presence of high amount of lipid content in some species, and also due to the fact that lipid synthesis, especially of non-polar TAGs (triacylglycerols), which are considered to be the best substrate for producing biodiesel, can be modulated by varying the growth conditions (Monari et al., 2016).

1.3 Problem statement

In 2011, Malaysia was the second largest palm oil producer in the world, with a total of 16.6 million tonnes, an amount lesser than 1% from the total world's supply behind Indonesia. Since the palm oil industry is huge, with 67% of agricultural land covered with oil palm tree, biomass from oil palm contributes the most. Currently, 85.5% of the biomass residues are coming from the palm oil industry. Palm oil has a very good potential in producing alternative energy due to its calorific content (Ahmad *et al.*, 2011 and MPOB, 2012). More than 85% of palm oil mills in Malaysia have adopted ponding system for treating POME (Ma *et al.*,

1993; Chin *et al.*,2013) while, the rest have opted for open digesting tank (Yacob *et al.*, 2005). These methods are regarded as conventional POME treatment method, whereby longer retention time and large treatment areas are required (Poh and Chong 2009). The effluent that comes out from palm oil mill is hazardous to the ecosystem. The discharge can lead to land and aquatic pollution if it is left untreated (Salihu, 2012).

Based on the statistic value of total crude palm oil production in May 2001, the production of 985,063 tonnes of crude palm oil means a total of 1,477,595 m³ of water is being used, and 738,797 m³ released as POME, in a month. Without proper treatment, this wastewater will pollute the nearby watercourses. The current treatment technology of POME typically consists of biological aerobic and anaerobic digestion. Biologically treated effluent is disposed of via land application system, thus providing essential nutrients for growing plants. This method may be a good choice for the disposal of treated effluent. However, considering the rate of daily wastewater production, for example, approximately 26 m³/d for an average palm oil mill with an operating capacity of 35 t/d FFB, it is doubtful that the surrounding plantations receiving it could efficiently absorb all the treated effluent (Wah *et al.*, 2002).

The waste water treatment technologies are expensive, dependent on skilled personnel and hard to carry out (Darajeh *et al.*, 2014). Furthermore, the common conventional treatment is unable to meet the regulations set by the Department of Environment (DOE) with the level of BOD at 100 mg/L. According to Ahmad *et al.*, 2003, large quantities of water are used during the extraction of crude palm oil from the fresh fruit bunch, and about 50% of the water results in POME. POME is a thick brownish liquid that contains high amount of total solids (40,500 mg/L), oil and grease (4000 mg/L), COD (50,000 mg/L) and BOD (25,000 mg/L). The disposal of this highly polluting effluent is becoming a major problem if it is not being treated properly besides a stringent standard limit imposed by the Malaysian Department of Environment for the discharge of effluent. A POME treatment system based on membrane technology shows high potential for eliminating the environmental

problem, and in addition, this alternative treatment system offers water recycling (Ahmad *et al.*, 2003).

POME contains high content of degradable organic matter, which could become one of the promising sources for renewable energy in Malaysia (Ahmad *et al.*, 2011; Chin *et al.*, 2013). The discharge of improperly treated POME creates adverse impact to the environment. However, the substances in POME are able to support the growth of microalgae. Microalgae naturally exist in many palm oil mill processes, phenomenon known as "algae bloom", hence declining the water quality. Because POME consists of large amount of organic compounds and inorganics which is hazardous to environmental health, therefore microalgae have been suggested as a potential candidate to remove these pollutants and able to breakdown the organic compounds present in it (Munoz and Guieysse, 2006; Kamyab *et al.*, 2015a).

On the other hand, culturing microalgae in wastewater offers an inexpensive alternative to the conventional forms of wastewater treatment (Hoh *et al.*, 2016; Ge *et al.*, 2016). At the same time microalgae can utilize the nitrogen and phosphorus compound in wastewater to generate microalgae biomass for different types of lipid production, which can serve as a substrate for biofuel production (Huang *et al.*, 2010; Kamyab *et al.*, 2014a).

In addition, still there is a need to investigate an efficient microalgae candidate to apply in wastewater treatment method for remediation and simultaneously produce lipid. Utilizing microalgae into the treatment system cause to several advantageous comprise enhancing treatment method, microalgae growth, decreasing nutrients, reducing cost and time saving.

1.4 Objectives of study

The main objectives of this study are as follows:

- 1. To isolate, identify and screen the algal species that can grow in different concentrations of POME.
- 2. To investigate the growth and the physio-chemical parameters in achieving higher lipid from the selected micro algal species.
- 3. To determine the lipid content from the selected microalgae based on different conditions (carbon to total nitrogen ratio, photo periods, and organic loading rate).
- 4. To quantify and characterize the fatty acid content of the selected microalgae to serve as substrate for the production of biodiesel.

1.5 Scope of study

This study aimed to isolate and identify the potential microalgae isolated from the palm oil mill located in Johor, Malaysia (geographical location, latitude N 3° 57' 20.01 and longitude E 101° 11' 55.69) which has the existing POME that are capable of producing high lipid using microalgae. The scope of this study is to assess the main component of POME to be used as organic carbon for microalgae. The research is mainly focused in developing a lab scale prototype to produce high lipid content from the microalgae. The applicable parameters such as optical density (OD), Light intensity, Chlorophyll content, Mixed Liquor Suspended Solid (MLSS), Mixed Liquor Volatile Suspended Solid (MLVSS), Cell Dry Weight (CDW), Scanning Electronic Microscope (SEM) and FTIR, were also investigated in this study to enhance the lipid production. The study is more focused on lipid production of POME using different species of microalgae at laboratory scale.

1.6 Significance of the study

The use of wastewater for the growth of microalgal cultures is considered beneficial for minimizing the use of freshwater, reducing the cost of nutrient addition, removing nitrogen and phosphorus from wastewater and producing microalgal biomass as bio resources for biofuel or value added by products. There are three main sources of wastewater (municipal (domestic), agricultural and industrial wastewater) which contains a variety of ingredients. Some components in the wastewater, such as nitrogen and phosphorus, are useful ingredients for microalgal cultures (Chiu *et al.*, 2015).

There are several important aspects to be considered during the current study. POME is the major source of water pollutant in Malaysia (Kamarudin *et al.*, 2015). For example, in a conventional palm oil mill, 600-700 kg of POME is generated for every 1000 kg of processed FFB (fresh fruit bunches) (Aini *et al.*, 1999; MPOB, 2014). By utilizing the ingredients present in POME, this study will play a major role to solve the pollution problem resulting from the POME as it will pollute the environment if it is improperly discharged into the environment.

Furthermore, culturing microalgae in wastewater offers an inexpensive alternative in comparison to conventional forms of biological wastewater treatments in reducing the main contents such as nitrogen and phosphorus compounds present in the wastewater. Therefore, this study using POME has the potential to offer carbon source for the microalgal growth in promoting high lipid content (Kamyab *et al.*, 20154b; Chiu *et al.*, 2015). Microalgae are able to produce the highest potential outputs of oil. Furthermore, the characteristics of microalgae include all year long growth and a short life cycle that makes it the fastest growing plant on Earth; microalgae growth is 100 times faster than trees. Additionally, microalgae cultivation is inexpensive with its necessities only being readily available raw materials such as sunlight, water, carbon dioxide and nutrients. Besides that, the information and knowledge gained through this research will provide a strong foundation for further research in biochemistry, genetic engineering, and technology enhancement of oil producing microalgae (Clarens *et al.*, 2010).

REFRENCES

- Abdul Aziz, H. (2007). Reactive Extraction Of Sugars From Oil Palm Empty Fruit Bunch Hydrolysate Using Naphthalene-2-Boronic Acid (Doctoral dissertation, Universiti Sains Malaysia).
- Abdullah, A. Z., Salamatinia, B., Mootabadi, H., & Bhatia, S. (2009). Current status and policies on biodiesel industry in Malaysia as the world's leading producer of palm oil. *Energy Policy*, 37(12), 5440-5448.
- Ahluwalia, S. S., & Goyal, D. (2007). Microbial and plant derived biomass for removal of heavy metals from wastewater. *Bioresource technology*, 98(12), 2243-2257.
- Ahmad, A. D., Salihon, J., & Tao, D. G. (2015, July). Evaluation of CO₂ Sequestration by Microalgae Culture in Palm Oil Mill Effluent (POME) Medium. In *Advanced Materials Research* (Vol. 1113, pp. 311-316).
- Ahmad, A. L., Ismail, S., & Bhatia, S. (2003). Water recycling from palm oil mill effluent (POME) using membrane technology. *Desalination*, 157(1), 87-95.
- Ahmad, A. L., Sumathi, S., & Hameed, B. H. (2006). Coagulation of residue oil and suspended solid in palm oil mill effluent by chitosan, alum and PAC. *Chemical Engineering Journal*, 118(1), 99-105.
- Ahmad, S., Ab Kadir, M. Z. A., & Shafie, S. (2011). Current perspective of the renewable energy development in Malaysia. *Renewable and Sustainable Energy Reviews*, 15(2), 897-904.
- Ajayan, K. V., Selvaraju, M., Unnikannan, P., and Sruthi, P. (2015). Phycoremediation of Tannery Wastewater Using Microalgae *Scenedesmus* Species. *International Journal of Phytoremediation*, 17,907–916.
- Alabi AO, Bibeau E, Tampier M, Council BCI (2009). Microalgae technologies & processes for biofuels-bioenergy production in British Columbia: Current technology, suitability & barriers to implementation. *The British Columbia Innovation Council, University of Manitoba*, Manitoba, Canada

- Alonso, D.L., Belarbi, E.H., Rodríguez-Ruiz, J., Segura, C.I., Giménez, A. (1998). Acyl lipids of three microalgae. *Phytochemistry*, 47, 1473–1481.
- Alvarez, D. A., Petty, J. D., Huckins, J. N., & Manahan, S. E. (2000). Development of an integrative sampler for polar organic chemicals in water. In *ACS Division of Environmental Chemistry, Preprints.* 40(3), 71-74.
- Anton, A., & Duthie, H. C. (1981). Use of cluster analysis in the systematics of the algal genus *Cryptomonas*. *Canadian Journal of Botany*, 59(6), 992-1002.
- APHA (2005). Standard methods for the examination of water and wastewater. (21st ed.). Washington, D.C: American Public Health Association (APHA).
- Arif, S., Tengku Mohd Ariff, T.A., (2001). The Case Study on the Malaysian Palm Oil, Paper prepared for the UNCTAD/ESCAP Regional Workshop on Commodity Export Diversification and Poverty Reduction in South and South-East Asia, Bangkok 3-5 April.
- Ashokkumar, V., & Rengasamy, R. (2012). Mass culture of *Botryococcus braunii* Kutz. under open raceway pond for biofuel production. *Bioresource technology*, 104, 394-399.
- Aslan, S., Kapdan, I.K. (2006). Batch kinetics of nitrogen and phosphorus removal from synthetic wastewater by algae. *Ecological Engineering*, 28, 64-70.
- Balduyck, L., Veryser, C., Goiris, K., Bruneel, C., Muylaert, K., & Foubert, I. (2015). Optimization of a Nile Red method for rapid lipid determination in autotrophic, marine microalgae is species dependent. *Journal of microbiological methods*, 118, 152-158.
- Banerjee, A., Sharma, R., Chisti, Y., & Banerjee, U. C. (2002). *Botryococcus braunii*: a renewable source of hydrocarbons and other chemicals. *Critical reviews in biotechnology*, 22(3), 245-279.
- Barclay, W. R., Meager, K. M., & Abril, J. R. (1994). Heterotrophic production of long chain omega-3 fatty acids utilizing algae and algae-like microorganisms. *Journal of Applied Phycology*, 6(2), 123-129.
- Barsanti, L., Gualtieri, P. (2006). Algae: Anatomy, Biochemistry, and Biotechnology, First Edition: *CRC Press Taylor & Francis Group*, New York, pp. 301.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109(4), 289-295.

- Bastert, J., Korting, H.C., Traenkle, P., Schmalreck, A. F. (1999). Identification of dermatophytes BY Fourier Transform Infrared Spectroscopy, Mycoses, ISSN 09333.42:525-528.
- Becker, E. W. (2007). Micro-algae as a source of protein. *Biotechnology* advances, 25(2), 207-210.
- Bellinger, E. G. (1992). A key to common algae: Freshwater, estuarine and some coastal species. Institution of Water and Environmental Management.
- Bello, M. M., Nourouzi, M. M., Abdullah, L. C., Choong, T. S., Koay, Y. S., & Keshani, S. (2013). POME is treated for removal of color from biologically treated POME in fixed bed column: applying wavelet neural network (WNN). *Journal of hazardous materials*, 262, 106-113.
- Bellorin, A. M., Oliveira, M. C., & Oliveira, E. C. (2002). Phylogeny and systematics of the marine algal family Gracilariaceae (*Gracilariales, Rhodophyta*) based on small subunit RDNA and its sequences of Atlantic and Pacific species. *Journal of Phycology*, 38(3), 551-563.
- Ben-Amotz, A., Tornabene, T. G., & Thomas, W. H. (1985). Chemical profile of selected species of microalgae with emphasis on lipids. *Journal of Phycology*, 21(1), 72-81.
- Bi, Z., & He, B. B. (2013). Characterization of microalgae for the purpose of biofuel production. *Transactions of the ASABE*, 56(4), 1529-1539.
- Bigogno, C., Khozin-Goldberg, I., Cohen, Z. (2002). Accumulation of arachidonic acid-rich triacylglycerols in the microalga Parietochloris incisa (trebuxiophyceae, chlorophyta). *Phytochemistry*, 60, 135–143.
- Bird, D. F., & Kalff, J. (1987). Algal phagotrophy: regulating factors and importance relative to photosynthesis in Dinobryon (Chrysophyceae). *Limnology and Oceanography*, 32(2), 277-284.
- Blackburn, S. I., Dunstan, G. A., Frampton, D. M. F., et al. (2009). Australian strain selection and enhancement for biodiesel from algae. *Phycologia*, 48(4): 8–9.
- Blanco, C., Petkova, E., Ibáñez, A., & Sáiz-Ruiz, J. (2002). A pilot placebocontrolled study of fluvoxamine for pathological gambling. *Annals of Clinical Psychiatry*, 14(1), 9-15.
- Bligh, E.G., and Dyer, W.J (1959). A rapid method of total lipid extraction and purification. *Can J. Biochem physiol*, 37,911-923.

- Bouterfas, R., Belkoura, M., Dauta, A. (2002). Light and temperature effects on the growth rate of three freshwater (2pt) algae isolated from a eutrophic lake. *Hydrobiologia*, 489(1-3): 207-217.
- Bradley, Mike. 2007. Biodiesel (FAME) Analysis by FT-IR. Ph.D., Thermo Fisher Scientific, Madison, WI, US, Application Note: 51258.
- Brandenburg, K., Seydel, U. (1996). Fourier Transform Infrared Spectroscopy of cell surface polysaccharides. In Manstsch, H.H., and Chapman, D. (eds.), *Infrared Spectroscopy of biomolicules*, Wiley: chichester, pp. 203-278.
- Braun, B. Z. (1974). Light absorption, emission and photosynthesis. *Botanical monographs*.
- Brennan, L., & Owende, P. (2010). Biofuels from microalgae—a review of technologies for production, processing, and extractions of biofuels and coproducts. *Renewable and sustainable energy reviews*, 14(2), 557-577.
- Brown, L. M. (1996). Uptake of carbon dioxide from flue gas by microalgae. *Energy Conversion and Management*, 37(6), 1363-1367.
- Brune, D. E., Lundquist, T. J., Benemann, J. R. (2009). Microalgal biomass for greenhouse gas reductions: Potential for replacement of fossil fuels and animal feeds. *Journal of Environmental Engineering-Asce*, 135(11): 1136–1144.
- Bulgariu, D., & Bulgariu, L. (2012). Equilibrium and kinetics studies of heavy metal ions biosorption on green algae waste biomass. *Bioresource technology*, 103(1), 489-493.
- Burlew, J. S. (1953). Algal culture. From Laboratory to Pilot Plant, Carnegie Inst. Washington Publ, 600(1).
- Burtner, C. R., Murakami, C. J., Kennedy, B. K., & Kaeberlein, M. (2009). A molecular mechanism of chronological aging in yeast. *Cell cycle*, 8(8), 1256-1270.
- Cai, T., Park, S. Y., & Li, Y. (2013). Nutrient recovery from wastewater streams by microalgae: status and prospects. *Renewable and Sustainable Energy Reviews*, 19, 360-369.
- Cakmak, T., Angun, P., Demiray, Y. E., Ozkan, A. D., Elibol, Z., & Tekinay, T. (2012). Differential effects of nitrogen and sulfur deprivation on growth and biodiesel feedstock production of Chlamydomonas reinhardtii. *Biotechnology and bioengineering*, 109(8), 1947-1957.

- Carioca J.O.B., Hiluy J.J., Leal M., et al. (2009). The hard choice for alternative biofuels to diesel in Brazil. *Biotechnol Adv*, 27(6): 1043–1050.
- Carvalho, P. A., Silva, O. S., Baptista, M. Jo., Malcata, F. X. (2011). Light Requirements in Microalgal Photobioreactors: An Overview of Biophotonic Aspects. *Appl Microbiol Biotechnol.*, 89, 1275–1288.
- Cavonius, L., Fink, H., Kiskis, J., Albers, E., Undeland, I., & Enejder, A. (2015). Imaging of Lipids in Microalgae with Coherent Anti-Stokes Raman Scattering Microscopy. *Plant physiology*, 167(3), 603-616.
- Çelekli, A., & Yavuzatmaca, M. (2009). Predictive modeling of biomass production by Spirulina platensis as function of nitrate and NaCl concentrations. *Bioresource technology*, 100(5), 1847-1851.
- Chen, C. Y., Yeh, K. L., Aisyah, R., Lee, D. J., & Chang, J. S. (2011). Cultivation, photobioreactor design and harvesting of microalgae for biodiesel production: a critical review. *Bioresource technology*, 102(1), 71-81.
- Chen, J. P., & Yang, L. (2005). Chemical modification of Sargassum sp. for prevention of organic leaching and enhancement of uptake during metal biosorption. *Industrial & engineering chemistry research*, 44(26), 9931-9942.
- Chen, W., Zhang, C., Song, L., Sommerfeld, M. and Hu Q. (2009). A High Throughput Nile red Method for Quantitative Measurement of Neutral Lipids in Microalgae. *Microbiological Methods*.77:41-47.
- Cheng, J., Huang, R., Yu, T., Li, T., Zhou, J., & Cen, K. (2014). Biodiesel production from lipids in wet microalgae with microwave irradiation and biocrude production from algal residue through hydrothermal liquefaction. *Bioresource technology*, 151, 415-418.
- Chin, M. J., Poh, P. E., Tey, B. T., Chan, E. S., & Chin, K. L. (2013). Biogas from palm oil mill effluent (POME): Opportunities and challenges from Malaysia's perspective. *Renewable and Sustainable Energy Reviews*, 26, 717-726.
- Chisti, Y. (2007). Biodiesel from microalgae. *Biotechnology Advances*, 25(3), 294-306.
- Chisti, Y., & Moo-Young, M. (1986). Disruption of microbial cells for intracellular products. *Enzyme and Microbial Technology*, 8(4), 194-204.

- Chiu, S. Y., Kao, C. Y., Chen, T. Y., Chang, Y. B., Kuo, C. M., & Lin, C. S. (2015). Cultivation of microalgal *Chlorella* for biomass and lipid production using wastewater as nutrient resource. *Bioresource technology*, 184, 179-189.
- Chiu, S. Y., Kao, C. Y., Tsai, M. T., Ong, S. C., Chen, C. H., & Lin, C. S. (2009).
 Lipid accumulation and CO 2 utilization of Nannochloropsis oculata in response to CO 2 aeration. *Bioresource technology*, 100(2), 833-838.
- Cho, H. S., Oh, Y. K., Park, S. C., Lee, J. W., & Park, J. Y. (2013). Effects of enzymatic hydrolysis on lipid extraction from Chlorella vulgaris. *Renewable energy*, *54*, 156-160.
- Cho, H. U., Kim, Y. M., Choi, Y. N., Xu, X., Shin, D. Y., & Park, J. M. (2015). Effects of pH control and concentration on microbial oil production from *Chlorella vulgaris* cultivated in the effluent of a low-cost organic waste fermentation system producing volatile fatty acids. *Bioresource technology*, 184, 245-250.
- Christwardana, M., & Soetrisnanto, D. (2013). Phytoremediations of Palm Oil Mill Effluent (POME) by using aquatic plants and microalge for biomass production. *Journal of Environmental Science and Technology*, 6(2), 79.
- Clarens, A. F., Resurreccion, E. P., White, M. A., & Colosi, L. M. (2010). Environmental life cycle comparison of algae to other bioenergy feedstocks. *Environmental science & technology*, 44(5), 1813-1819.
- Converti, A., Casazza, A.A., Ortiz, E.Y., Perego, P., Del Borghi, M., (2009). Effect of temperature and nitrogen concentration on the growth and lipid content of Nannochloropsis oculata and *Chlorella vulgaris* for biodiesel production. *Chemical Engineering and Processing: Process Intensification*, 48, 1146-1151.
- Cooney, M., Young, G., & Nagle, N. (2009). Extraction of bio-oils from microalgae. Separation & Purification Reviews, 38(4), 291-325.
- Costa, J.A.V., de Morais, M.G., (2010). The role of biochemical engineering in the production of biofuels from microalgae. *Bioresource Technology*, 102, 2-9.
- Cuscov, M., & Muller, F. L. (2015). Differentiating humic and algal surface active substances in coastal waters by their pH-dependent adsorption behaviour. *Marine Chemistry*, 174, 35-45.
- D'Oca, M. G. M., Viêgas, C. V., Lemoes, J. S., Miyasaki, E. K., Morón-Villarreyes, J. A., Primel, E. G., & Abreu, P. C. (2011). Production of FAMEs from

- several microalgal lipidic extracts and direct transesterification of the *Chlorella pyrenoidosa. biomass and bioenergy*, 35(4), 1533-1538.
- Darajeh, N., Idris, A., Truong, P., Abdul Aziz, A., Abu Bakar, R., & Che Man, H. (2014). Phytoremediation Potential of Vetiver System Technology for Improving the Quality of Palm Oil Mill Effluent. *Advances in Materials Science and Engineering*.
- Davis, R., Aden, A., & Pienkos, P. T. (2011). Techno-economic analysis of autotrophic microalgae for fuel production. *Applied Energy*, 88(10), 3524-3531.
- de Andrade, G. A., Berenguel, M., Guzmán, J. L., Pagano, D. J., & Acién, F. G. (2016). Optimization of biomass production in outdoor tubular photobioreactors. *Journal of Process Control*, 37, 58-69.
- de Morais, M. G., & Costa, J. A. V. (2007). Isolation and selection of microalgae from coal fired thermoelectric power plant for biofixation of carbon dioxide. *Energy Conversion and Management*, 48(7), 2169-2173.
- Dean, A. P., Sigee, D. C., Estrada, B., & Pittman, J. K. (2010). Using FTIR spectroscopy for rapid determination of lipid accumulation in response to nitrogen limitation in freshwater microalgae. *Bioresource Technology*, 101(12), 4499-4507.
- de-Bashan, L. E., Hernandez, J. P., & Bashan, Y. (2015). Interaction of Azospirillum spp. with Microalgae: A Basic Eukaryotic–Prokaryotic Model and Its Biotechnological Applications. In *Handbook for Azospirillum* (pp. 367-388). Springer International Publishing.
- Del Río, E., Armendáriz, A., García-Gómez, E., García-González, M., & Guerrero, M. G. (2015). Continuous culture methodology for the screening of microalgae for oil. *Journal of biotechnology*, 195, 103-107.
- Demirbas, A. (2000). Mechanisms of liquefaction and pyrolysis reactions of biomass. *Energy Conversion and Management*, 41: 633-646.
- Demirbas, A. (2003). Biodiesel fuels from vegetable oils via catalytic and non-catalytic supercritical alcohol transesterifications and other methods: a survey. *Energy conversion and Management*, 44(13), 2093-2109.
- Demirbas, M. F. (2010). Microalgae as a feedstock for biodiesel. *Energy Education Science and Technology Part* A, 25(1–2): 31–43.

- Dijkstra, A.J. (2006). Revisiting the formation of trans isomers during partial hydrogenation of triacylglycerol oils. *European Journal of Lipid Science and Technology*, 108: 249-264.
- Dismukes, G. C., Carrieri, D., Bennette, N., Ananyev, G. M., & Posewitz, M. C. (2008). Aquatic phototrophs: efficient alternatives to land-based crops for biofuels. *Current opinion in biotechnology*, 19(3), 235-240.
- Dominguez, L., Cases, V., Birek, C., Rodriguez, M., Prats, D., (2012).Influence of organic loading rate on the performance of ultrafiltration and microfiltration membrane bioreactors at high sludge retention time. *Chemical Engineering Journal*, 181-82, 132-143.
- Dortch, Q. (1990). The Interaction Between Ammonium and Nitrate Uptake in Phytoplankton. *Marine Ecology Progress Series*, 61,183-201.
- Doucha, J., & Lívansky, K. (2006). Productivity, CO2/O2 exchange and hydraulics in outdoor open high density microalgal (*Chlorella sp.*) photobioreactors operated in a Middle and Southern European climate. *Journal of Applied Phycology*, 18(6), 811-826.
- Du, Z., Mohr, M., Ma, X., Cheng, Y., Lin, X., Liu, Y.,& Ruan, R. (2012). Hydrothermal pretreatment of microalgae for production of pyrolytic bio-oil with a low nitrogen content. *Bioresource technology*, 120, 13-18.
- Duffy, J. E., Canuel, E. A., Adey, W., et al. (2009). Biofuels: Algae. *Science*, 326(5958): 1345–1345.
- Dunahay, T., Benemann, J., & Roessler, P. (1998). A look back at the US Department of Energy's Aquatic Species Program: Biodiesel from algae (Vol. 328). Golden: National Renewable Energy Laboratory.
- Dunstan, G. A., Volkman, J. K., Jeffrey, S. W., & Barrett, S. M. (1992). Biochemical composition of microalgae from the green algal classes Chlorophyceae and Prasinophyceae. 2. Lipid classes and fatty acids. *Journal of Experimental Marine Biology and Ecology*, 161(1), 115-134.
- Duong, V. T., Li, Y., Nowak, E., & Schenk, P. M. (2012). Microalgae isolation and selection for prospective biodiesel production. *Energies*, *5*(6), 1835-1849.
- Eaton, A. D., Clesceri, L. S., Rice, E. W., Greenberg, A. E., & Franson, M. A. H. (2005). APHA: standard methods for the examination of water and wastewater. *Centennial Edition.*, *APHA*, *AWWA*, *WEF*, *Washington*, *DC*.

- Ehimen, E. A., Sun, Z. F., & Carrington, C. G. (2010). Variables affecting the in situ transesterification of microalgae lipids. *Fuel*, 89(3), 677-684.
- Elsey, D. Jameson, B. Raleigh and M. J. Cooney. (2007). Fluorescent Measurement of Microalgal Neutral Lipids. Journal of Microbiological Methods. 68, 639-642.
- Elumalai, S., Sarvanan, G.K., Ramganesh, S., Sakahivel R., Prakasam, V., (2013,). Phycoremediation of Textile Dye Industrial Effluent from Tripur District, Tamilnadu, India. *International journal of Science Innovation and Discoveries*, 3: 31-37.
- Erata, M., Kubota, M., Takahashi, T., Inouye, I., & Watanabe, M. (1995). Ultrastructure and phototactic action spectra of two genera of cryptophyte flagellate algae, Cryptomonas and Chroomonas. *Protoplasma*, 188(3-4), 258-266.
- Falkowski PG, Raven JA (1997). Aquatic photosynthesis. Blackwater Science, London, p 375.
- Feng, Y., Li, C., Zhang, D. (2011). Lipid production of *Chlorella vulgaris* cultured in artificial wastewater medium. *Bioresource Technology*, 102, 101-105.
- Fernández, F. A., Sevilla, J. F., Pérez, J. S., Grima, E. M., & Chisti, Y. (2001). Airlift-driven external-loop tubular photobioreactors for outdoor production of microalgae: assessment of design and performance. *Chemical Engineering Science*, 56(8), 2721-2732.
- Fernández-Reiriz, M. J., Perez-Camacho, A., Ferreiro, M. J., Blanco, J., Planas, M., Campos, M. J., & Labarta, U. (1989). Biomass production and variation in the biochemical profile (total protein, carbohydrates, RNA, lipids and fatty acids) of seven species of marine microalgae. *Aquaculture*, 83(1), 17-37.
- Fowler, S.D., Brown, W.J., Warfel, J., Greenspan, P. (1979). Use of Nile Red for the Rapid in situ Quantitation of Lipids on Thin-layer Chromatograms. *Journal of Lipid Research* 28, 1225–1232.
- Fukuda, H., Kondo, A., & Noda, H. (2001). Biodiesel fuel production by transesterification of oils. *Journal of bioscience and bioengineering*, 92(5), 405-416.
- Fulton L. (2004). Biomass and Agriculture Sustainability, Markets and Policies. International Energy Agency (IEA) biofuels study–interim report: result and key messages so far. 105–112.

- Gao, W., Xiong, Y., Zhang, W. Yuan and Q. Wu. (2008). Rapid quantitation of lipid in microalgae by time-domain nuclear magnetic resonance. *Microbiological Methods*: 75,437-440.
- Ge, S., & Champagne, P. (2016). Nutrient removal, microalgal biomass growth, harvesting and lipid yield in response to centrate wastewater loadings. *Water research*, 88, 604-612.
- Gerpen JV. (2005). Biodiesel Processing and Production. *Fuel Process Technol*. 86:1097–107.
- Ghirardi, M. L., Zhang, L., Lee, J. W., Flynn, T., Seibert, M., Greenbaum, E., & Melis, A. (2000). Microalgae: a green source of renewable H 2. Trends in biotechnology, 18(12), 506-511.
- Giannelli, L., Yamada, H., Katsuda, T., & Yamaji, H. (2015). Effects of temperature on the astaxanthin productivity and light harvesting characteristics of the green alga Haematococcus pluvialis. *Journal of bioscience and bioengineering*, 119(3), 345-350.
- Giordano, M., Kansiz, M., Heraud, P., Beardall, J., Wood, B., & McNaughton, D. (2001). Fourier transform infrared spectroscopy as a novel tool to investigate changes in intracellular macromolecular pools in the marine microalga Chaetoceros muellerii (Bacillariophyceae). *Journal of Phycology*, 37(2), 271-279.
- Gnansounou, E., & Raman, J. K. (2016). Life cycle assessment of algae biodiesel and its co-products. *Applied Energy*, 161, 300-308.
- Gonçalves, A. L., Pires, J. C., & Simões, M. (2016). Biotechnological potential of Synechocystis salina co-cultures with selected microalgae and cyanobacteria: Nutrients removal, biomass and lipid production. *Bioresource technology*, 200, 279-286.
- Graham LE, Graham JM, Wilcox LW (2009). Algae. Benjamin Cummings, University of California, California, U.S.
- Griffiths, M.J., Harrison, S.T.L. (2009). Lipid productivity as a key characteristic for choosing algal species for biodiesel production. *Journal of Applied Phycology*, 21, 493-507.
- Grobbelaar, J. U. (2006). Photosynthetic response and acclimation of microalgae to light fluctuations. *Algal Cultures Analogues of Blooms and Applications*, *Science Publishers*, *Enfield*, USA, 671-683.

- Guihéneuf, F., Schmid, M., & Stengel, D. B. (2015). Lipids and Fatty Acids in Algae: Extraction, Fractionation into Lipid Classes, and Analysis by Gas Chromatography Coupled with Flame Ionization Detector (GC-FID). *Natural Products From Marine Algae: Methods and Protocols*, 173-190.
- Guil-Guerrero, J.L., Navarro-Juarez, R., Lopez-Martinez, J.C., Campra-Madrid, P., Rebolloso-Fuentes, M.M. (2004). Functional properties of the biomassof three microalgal species. *Journal of Food Engineering*, 65,511-517.
- Guiry, M. D., & Guiry, G. M. (2014). AlgaeBase. World-wide electronic publication. Galway: National University of Ireland.
- Guo, Z., Liu, Y., Guo, H., Yan, S., & Mu, J. (2013). Microalgae cultivation using an aquaculture wastewater as growth medium for biomass and biofuel production. *Journal of Environmental Sciences*, 25, S85-S88.
- Gurr, M.I., Harwood, J.L., Frayn, K.N. (2002). *Lipid Biochemistry: An Introduction*, (5th ed.). Blackwell: Oxford, UK, p. 320.
- Hadiyanto, M. C., & Soetrisnanto, D. (2013). Phytoremecliations of Palm Oil Mill Effluent (POME) by Using Aquatic Plants and Microalge for Biomass Production. *Journal of Environmental Science and Technology*, 6(2), 79-90.
- Hadiyanto, M. M. A. N., & Hartanto, G. D. (2012, September). Enhancement of biomass production from Spirulina sp. cultivated in POME medium. In Proceedings of the International Conference on Chemical and Material Engineering (1-6).
- Halim, R., Harun, R., Danquah, M. K., & Webley, P. A. (2012). Microalgal cell disruption for biofuel development. *Applied Energy*, 91(1), 116-121.
- Han, F., Huang, J., Li, Y., Wang, W., Wang, J., Fan, J., & Shen, G. (2012). Enhancement of microalgal biomass and lipid productivities by a model of photoautotrophic culture with heterotrophic cells as seed. *Bioresource* technology, 118, 431-437.
- Hansen, S. (2007). Feasibility study of performing an life cycle assessment on crude palm oil production in Malaysia (9 pp). *The International Journal of Life Cycle Assessment*, 12(1), 50-58.
- Heckman, C. W. (1985). The development of vertical migration patterns in the sediments of estuaries as a strategy for algae to resist drift with tidal currents. Internationale Revue der gesamten Hydrobiologie und Hydrographie, 70(1), 151-164.

- Henderson, R., Parsons, S. A., & Jefferson, B. (2008). The impact of algal properties and pre-oxidation on solid–liquid separation of algae. *Water Research*, 42(8), 1827-1845.
- Hernandez, J. P., de-Bashan, L. E., & Bashan, Y. (2006). Starvation enhances phosphorus removal from wastewater by the microalga Chlorella spp. co-immobilized with Azospirillum brasilense. *Enzyme and Microbial Technology*, 38(1), 190-198.
- Hirano, A., Ueda, R., Hirayama, S., & Ogushi, Y. (1997). CO 2 fixation and ethanol production with microalgal photosynthesis and intracellular anaerobic fermentation. *Energy*, 22(2), 137-142.
- Hoh, D., Watson, S., & Kan, E. (2016). Algal biofilm reactors for integrated wastewater treatment and biofuel production: A review. *Chemical Engineering Journal*, 287, 466-473.
- Ho, S. H., Chen, C. Y., & Chang, J. S. (2012). Effect of light intensity and nitrogen starvation on CO₂ fixation and lipid/carbohydrate production of an indigenous microalga Scenedesmus obliquus CNW-N. *Bioresource Technology*, 113, 244-252.
- Hossain, A.B.M.S., Salleh, A., Boyce, A.N., Chowdhury, P., Naqiuddin, M. (2008). Biodiesel fuel production from algae as renewable energy. *American Journal of Biochemistry and Biotechnology*, 4(3):250-4.
- Hosub, K., Soonjin, H., Haeki, S., Jaeki, S., Kwangguk, A., Chun G. (2007). Effects of Limiting Nutrients and N: P Ratios on the Phytoplankton Growth in a Shallow Hypertrophic Reservoir. *Hydrobiologia*, 589 (1), 255-267.
- Hsieh, C. H., & Wu, W. T. (2009). Cultivation of microalgae for oil production with a cultivation strategy of urea limitation. *Bioresource technology*, 100(17), 3921-3926.
- Hsieh, C.-H., Wu, W.-T. (2009). A novel photobioreactor with transparent rectangular chambers for cultivation of microalgae. *Biochemical Engineering Journal* 300–305.
- Hu, Q., Sommerfeld, M., Jarvis, E., Ghirardi, M., Posewitz, M., Seibert, M., & Darzins, A. (2008). Microalgal triacylglycerols as feedstocks for biofuel production: perspectives and adances. *The Plant Journal*, 54(4), 621-639.
- Huang, G., Chen, F., Wei, D., Zhang, X., Chen, G. (2010). Biodiesel production by microalgal biotechnology. *Applied Energy*, 87, 38-46.

- Huang, W., Li, B., Zhang, C., Zhang, Z., Lei, Z., Lu, B., & Zhou, B. (2015). Effect of algae growth on aerobic granulation and nutrients removal from synthetic wastewater by using sequencing batch reactors. *Bioresource technology*, 179, 187-192.
- Huang, Y., Chen, M., Liu, D., et al. (2008). Effect of Nitrogen, Phosphorus, Light Formation and Disappearance and Water Temperature on the of Blue green Algae Bloom. Journal of Northwest Science Technology University of Agriculture and Forest (Nature Science Edition), 36(9), 93-100.
- Huntley, M. E., & Redalje, D. G. (2007). CO2 mitigation and renewable oil from photosynthetic microbes: a new appraisal. *Mitigation and adaptation strategies for global change*, 12(4), 573-608.
- Huntley, M., Redalje, D.G. (2007). CO₂ Mitigation and Renewable Oil from Photosynthetic Microbes: A New Appraisal. *Mitigation and Adaptation Strategies for Global Change*, 12: 573-608.
- Ibrahim, R. I., Wong, Z. H., & Mohammad, A. W. (2015). Optimization and performance evaluation for nutrient removal from palm oil mill effluent wastewater using microalgae. In *IOP Conference Series: Materials Science and Engineering* (Vol. 78, No. 1, p. 012006). IOP Publishing.
- Illman, A. M., Scragg, A. H., & Shales, S. W. (2000). Increase in Chlorella strains calorific values when grown in low nitrogen medium. *Enzyme and microbial technology*, 27(8), 631-635.
- Im, H., Kim, B., & Lee, J. W. (2015). Concurrent production of biodiesel and chemicals through wet in situ transesterification of microalgae. *Bioresource* technology, 193, 386-392.
- Jacob-Lopes, E., Scoparo, C.H.G., Lacerda, L.M.C.F., Franco, T.T. (2009). Effect of light cycles (night/day) on CO2 fixation and biomass production by microalgae in photobioreactors. *Chemical Engineering and Processing: Process Intensification* 48(1), 306-310.
- James, A. T., & Martin, A. J. P. (1952). Gas-liquid partition chromatography: the separation and micro-estimation of volatile fatty acids from formic acid to dodecanoic acid. *Biochemical Journal*, 50(5), 679.
- Jang, E.S., Jung, M.Y., Min, D.B. (2005). Hydrogenation for low trans and high conjugated fatty acids. *Comprehensive Reviews in Food Science and Food Safety*, 4: 22-30.

- Janssen, M., Janssen, M., de Winter, M., Tramper, J., Mur, L.R., Snel, J., Wijffels, R.H. (2000). Efficiency of light utilization of *Chlamydomonas reinhardtii* under medium-duration light/dark cycles. *Journal of Biotechnology*, 78, 123-137.
- Janssen, M., Kuijpers, T.C., Veldhoen, B., Ternbach, M.B., Tramper, J., Mur, L.R., Wijffels, R.H. (1999). Specific growth rate of *Chlamydomonas reinhardtii* and *Chlorella sorokiniana* under medium duration light/dark cycles: 13-87 s. *Journal of Biotechnology*, 70, 323-333.
- Janssen, M., Tramper, J., Mur, L. R., & Wijffels, R. H. (2003). Enclosed outdoor photobioreactors: Light regime, photosynthetic efficiency, scale-up, and future prospects. *Biotechnology and bioengineering*, 81(2), 193-210.
- Jena, U., & Das, K. C. (2011). Comparative evaluation of thermochemical liquefaction and pyrolysis for bio-oil production from microalgae. *Energy & fuels*, 25(11), 5472-5482.
- Jimenez, C., Cossío, B. R., Labella, D., & Niell, F. X. (2003). The feasibility of industrial production of Spirulina (Arthrospira) in Southern Spain. *Aquaculture*, 217(1), 179-190.
- John, D. M., Whitton, B. A., & Brook, A. J. (2002). The freshwater algal flora of the British Isles: an identification guide to freshwater and terrestrial algae (Vol. 1). Cambridge University Press.
- Kaewpintong, K. (2004). Cultivation of Haematococcus pluvialis in airlift bioreactor. *Chulalongkorn University, Chulalongkorn*, Thailand.
- Kamarudin, K. F., Tao, D. G., Yaakob, Z., Takriff, M. S., Rahaman, M. S. A., & Salihon, J. (2015). A review on wastewater treatment and microalgal by-product production with a prospect of palm oil mill effluent (POME) utilization for algae. *Der Pharma Chemica*, 7(7), 73-89.
- Kamyab H., Md Din, M.F, Tin Lee, C., Keyvanfar A., Shafaghat A., Abd Majid, M. Z., Ponraj M, Xiao, Yun T. (2015b). Lipid Production by microalgae *Chlorella pyrenoidosa* cultivated in Palm Oil Mill Effluent (POME) using Hybrid Photo Bioreactor (HPBR). *Desalination and Water Treatment*, 55, 3737–3749.
- Kamyab, H., Fadhil, M., Lee, C., Ponraj, M., Soltani, M., Eva, S. (2014b). Micro-Macro Algal Mixture as a Promising Agent for Treating POME Discharge

- and its Potential Use as Animal Feed Stock Enhancer. *Jurnal Teknologi* 5, 1–4.
- Kamyab, H., Lee, C.T., Md Din, M.F., Ponraj, M., Mohamad, S.E., Sohrabi, M. (2014a). Effects of nitrogen source on enhancing growth conditions of green algae to produce higher lipid. *Desalination and Water Treatment* 52, 3579–3584.
- Kamyab, H., Md Din, M. F., Keyvanfar, A., Abd Majid, M.Z., Talaiekhozani, A., Shafaghat, A., Lee, C.T., Jeng Shiun, L., Ismail, H.H. (2015a). Efficiency of Microalgae *Chlamydomonas* on the Removal of Pollutants from Palm Oil Mill Effluent (POME). *Energy Procedia*, 75 -2400 2408.
- Kamyab, H., Ponraj, M., Fadhil, M., Abd Majid, M. Z.,*, Keyvanfar A., Rezania,S.,Shazwin., M.T.(2016). Isolation and screening of microalgae from agro-industrial wastewater (POME) for biomass and biodiesel sources. *Desalination and Water Treatment*.DOI:
- Karpagam, R., Raj, K. J., Ashokkumar, B., & Varalakshmi, P. (2015). Characterization and fatty acid profiling in two fresh water microalgae for biodiesel production: Lipid enhancement methods and media optimization using response surface methodology. *Bioresource technology*, 188, 177-184.
- Kayombo, S., Mbwette, T.S.A., Katima, J.H.Y., and Jorgensen, S.E. (2003). Effects of Substrate Concentrations on The Growth of Heterotrophic Bacteria and Algae in Secondary Facultative Ponds. *Water Research*, 37: 2937-2943.
- Kebelmann, K., Hornung, A., Karsten, U., & Griffiths, G. (2013). Intermediate pyrolysis and product identification by TGA and Py-GC/MS of green microalgae and their extracted protein and lipid components. *Biomass and bioenergy*, 49, 38-48.
- Kessler, J. O., Nedelcu, A. M., Solari, C. A., & Shelton, D. E. (2015). Cells Acting as Lenses: A Possible Role for Light in the Evolution of Morphological Asymmetry in Multicellular Volvocine Algae. In *Evolutionary Transitions to Multicellular Life* (pp. 225-243). Springer Netherlands.
- Khalid, R., and Wan Mustafa, W.A. (1992). External benefits of environmental regulation: Resource recovery and the utilisation of effluents. *The Environmentalist*, 12: 277-285.

- Khan, S. A., Hussain, M. Z., Prasad, S., & Banerjee, U. C. (2009). Prospects of biodiesel production from microalgae in India. *Renewable and Sustainable Energy Reviews*, 13(9), 2361-2372.
- Khozin-Goldberg, I., Cohen, Z. (2006). The effect of phosphate starvation on the lipid and fatty acid composition of the fresh water eustigmatophyte Monodus subterraneus. *Phytochemistry*, 67, 696–701.
- Kim, B. H., Ramanan, R., Kang, Z., Cho, D. H., Oh, H. M., & Kim, H. S. (2016). Chlorella sorokiniana HS1, a novel freshwater green algal strain, grows and hyperaccumulates lipid droplets in seawater salinity. Biomass and Bioenergy, 85, 300-305.
- Kimura K., Yamaoka M., Kamisaka Y. (2004). Rapid Estimation of Lipids in Oleaginous Fungi and Yeasts Using Nile Red Fluorescence. *Microbiological Methods*. 56:331–338.
- Kittikun, A.H., Prasertsan, P., Srisuwan, G., and Krause, A. (2000). Environmental Management of Palm Oil Mill. Internet Conference on Material Flow Analysis of Integrated Bio-Systems.
- Knothe, G. (2008). "Designer" biodiesel: Optimizing Fatty Ester Composition to Improve Fuel Properties. *Energy Fuel*. 22:1358–1364.
- Kong, W. B., Yang, H., Cao, Y. T., Song, H., Hua, S. F., & Xia, C. G. (2013). Effect of glycerol and glucose on the enhancement of biomass, lipid and soluble carbohydrate production by Chlorella vulgaris in mixotrophic culture. *Food Technology and Biotechnology*, 51(1), 62-69.
- Krienitz, L., Ustinova, I., Friedl, T., & Huss, V. A. (2001). Traditional generic concepts versus 18S rRNA gene phylogeny in the green algal family Selenastraceae (Chlorophyceae, *Chlorophyta*). *Journal of Phycology*, 37(5), 852-865.
- Krohn, B. J., McNeff, C. V., Yan, B., & Nowlan, D. (2011). Production of algae-based biodiesel using the continuous catalytic Mcgyan process. *Bioresource technology*, 102(1), 94-100.
- Lam, M. K., Lee, K. T., & Mohamed, A. R. (2009). Life cycle assessment for the production of biodiesel: a case study in Malaysia for palm oil versus jatropha oil. *Biofuels, Bioproducts and Biorefining*, 3(6), 601-612.

- Lansing, S., Botero, R. B., and Martin, J. F. (2008). Waste Treatment and Biogas Quality in Small-Scale Agricultural Digesters. *Bioresource Technology*. 99: 5881-5890.
- Laughton, C. (1986). Measurement of The Specific Lipid Content Of Attached Cells In Microtitre Cultures. *Analytical Biochemistry*. 156:307–314.
- Laurens, L. M., & Wolfrum, E. J. (2011). Feasibility of spectroscopic characterization of algal lipids: chemometric correlation of NIR and FTIR spectra with exogenous lipids in algal biomass. *BioEnergy Research*, 4(1), 22-35.
- Lavens, P., and Sorgeloos, P. (1996). Manual on the Production and Use of Live Food for Aquaculture. Manual on the Production and Use of Live Food for Aquaculture. Fisheries and Aquaculture Department, Chapter 2 Algal production.
- Lee, H. W., Roh, S. W., Cho, K., Kim, K. N., Cha, I. T., Yim, K. J., & Kim, S. J. (2015). Phylogenetic analysis of microalgae based on highly abundant proteins using mass spectrometry. *Talanta*, 132, 630-634.
- Lee, K., and Lee, C.-G. (2001). Effect Of Light/Dark Cycles on Wastewater Treatments by Microalgae. *Biotechnology and Bioprocess Engineering*. 6: 194-199.
- Lee, K.Y., Lee, G.C. (2009). Effect of light/dark cycles on wastewater treatments by microalga. *Biotechnology and Bioprocess Engineering* 6, 194-199.
- Li, X., Xu, H., Wu, Q. (2007). Large-scale biodiesel production from microalga Chlorella protothecoids through heterotrophic cultivation in bioreactors. *Biotechnology and Bioengineering*, DOI 10.1002/bit. 21489.
- Li, Y., Chen, Y.F., Chen, P., Min, M., Zhou, W., Martinez, B. (2011). Characterization of a microalga *Chlorella sp.* well adapted to highly concentrated municipal wastewater for nutrient removal and biodiesel production. Bioresource Technology. 102:5138-44.
- Li, Y., Horsman, M., Wu, N., Lan, C.Q., Dubois-Calero, N. (2008a). Biofuels from microalgae. *Biotechnology Progress*, 24(4), 815–20.
- Li, Y., Wang, B., Wu, N., Lan, C.Q. (2008b). Effects of nitrogen sources on cell growth and lipid production of Neochloris oleoabundans. *Applied Microbiology and Biotechnology*, 81(4),629–36.

- Liang, Y., Sarkany, N., Cui, Y. (2009). Biomass and lipid productivities of Chlorella vulgaris under autotrophic, heterotrophic and mixotrophic growth conditions. Biotechnology Letters 31(7), 1043-1049.
- Lim, S. L., Chu, W. L., & Phang, S. M. (2010). Use of *Chlorella vulgaris* for bioremediation of textile wastewater. Bioresource Technology, 101(19), 7314-7322.
- Liu, C., Jin, X., Sun, L., et al. (2005). Effects of pH Oil Growth and Species Changes of Algae in Freshwater. *Journal of Agro-Environment Science*, 24(2), 294-298.
- Liu, C., Sun, H., Zhu, L., et al. (2006). Effects of Salinity Formed with Two Inorganic Salts on Freshwater Algae Growth. Acta Scientiae Circumstantiae, 26(1), 157-161.
- Liu, T., Li, Y., Liu, F., & Wang, C. (2016). The enhanced lipid accumulation in oleaginous microalga by the potential continuous nitrogen-limitation (CNL) strategy. *Bioresource Technology*, 203, 150-159.
- Liu, Z.Y., Wang, G.C., and Zhou, B.C. (2008). Effect of Iron on Growth and Lipid Accumulation in *Chlorella vulgaris*. *Bioresource Technolog*, 99(11),4717-4722.
- Lordan, S., Ross, R. P., & Stanton, C. (2011). Marine bioactives as functional food ingredients: potential to reduce the incidence of chronic diseases. *Marine drugs*, 9(6), 1056-1100.
- Lu, Q., Zhou, W., Min, M., Ma, X., Ma, Y., Chen, P., & Urriola, P. E. (2016). Mitigating ammonia nitrogen deficiency in dairy wastewaters for algae cultivation. *Bioresource technology*, 201, 33-40.
- Lu, W., Wang, Z., Wang, X., & Yuan, Z. (2015). Cultivation of Chlorella sp. Using Raw Diary Wastewater for Nutrient Removal and Biodiesel Production: Characteristics Comparison of Indoor Bench-scale and Outdoor Pilot-scale Cultures. *Bioresource Technology*.
- Lucas, I. A. N. (1970). Observations on the fine structure of the *Cryptophyceae*. the genus *Cryptomonas1*, 2. *Journal of Phycology*, 6(1), 30-38.
- Lyon, S. R., Ahmadzadeh, H., & Murry, M. A. (2015). Algae-Based Wastewater Treatment for Biofuel Production: Processes, Species, and Extraction Methods. In *Biomass and Biofuels from Microalgae* 95-115. Springer International Publishing.

- Ma, A. N., Cheah, S. C., Chow, M. C., & Yeoh, B. G. (1993). Current status of palm oil processing wastes management. *Waste management in Malaysia: current status and prospects for bioremediation.*, 111-136.
- Ma, A. N., Chow, C. S., John, C. K., Ibrahim, A., & Isa, Z. (2001). Palm oil mill effluent—a survey. In *Proceeding PORIM regional workshop on palm oil mill technology and effluent treatment, Palm Oil Research Institute of Malaysia (PORIM) Serdang, Malaysia* 233-269.
- Ma, A. N., Seiji, A., Yoshio, T., & Hanif, J. (1994). Environmental Seminar on Palm Oil and Rubber Effluent Management.
- Ma, A.N. (1999). Treatment of palm oil mill effluent. In Singh, G., Lim, K.H., Leng,T., and David, L.K. (eds). *Oil palm and the environment: a Malaysian perspective*. Malaysian Oil Palm Growers' Council Kuala Lampur, 113-126.
- Ma, A.N. (2000). Environmental Management for the Oil Palm Industry. *Palm Oil Dev.*, 30:1-10.
- Ma, G., Hu, W., Pei, H., Jiang, L., Song, M., & Mu, R. (2015). In situ heterogeneous transesterification of microalgae using combined ultrasound and microwave irradiation. *Energy Conversion and Management*, 90, 41-46.
- Makewicz, A., Gribi, C., Eichenberger, W. (1997). Lipids of Ectocarpus fasciculatus (phaeophyceae). Incorporation of (1-14C) oleate and the role of TAG and MGDG in lipid metabolism. *Plant Cell Physiol.*, 38, 952–962.
- Malaysian Palm Oil Board (MPOB). (2012). http://www.mpob.gov.my/.
- Mallick, N. (2002). Biotechnological Potential of Immobilized Algae for Wastewater N, P and Metal Removal: A Review. *BioMetals*. 15: 377-390.
- Mansour, M. P., Frampton, D. M., Nichols, P. D., Volkman, J. K., & Blackburn, S. I. (2005). Lipid and fatty acid yield of nine stationary-phase microalgae: applications and unusual C24–C28 polyunsaturated fatty acids. *Journal of Applied Phycology*, 17(4), 287-300.
- Markou, G., & Georgakakis, D. (2011). Cultivation of filamentous cyanobacteria (blue-green algae) in agro-industrial wastes and wastewaters: a review. *Applied Energy*, 88(10), 3389-3401.
- Mata, T. M., Martins, A. A., & Caetano, N. S. (2010). Microalgae for biodiesel production and other applications: a review. *Renewable and sustainable energy reviews*, 14(1), 217-232.

- McArdell, C. S., Molnar, E., Suter, M. J. F., & Giger, W. (2003). Occurrence and fate of macrolide antibiotics in wastewater treatment plants and in the Glatt Valley Watershed, Switzerland. *Environmental Science & Technology*, 37(24), 5479-5486.
- McClure, G., Allen, S., Ponseti, R., Sisco, B., Lonjin, T., Carney, B., & Allen, T. M. (2015). Gas Chromatography Mass Spectrometry Analysis of Algae: The Fuel of Our Future.
- McMullan, C. R., & Vaeth, C. L. (2015). Lipid Content of Various Microalgal Species found in Lake Lanier.
- Meher, L. C., Sagar, D. V., & Naik, S. N. (2006). Technical aspects of biodiesel production by transesterification—a review. *Renewable and sustainable energy reviews*, 10(3), 248-268.
- Melack, M.J. (1981). Photosynthetic Activity of Phytoplankton in Tropical African Soda Lakes. *Hydrobiology*, 81,71-85.
- Meng, X., Yang, J., Xu, X., Zhang, L., Nie, Q., & Xian, M. (2009). Biodiesel production from oleaginous microorganisms. *Renewable energy*, 34(1), 1-5.
- Metzger, P., & Largeau, C. (2005). Botryococcus braunii: a rich source for hydrocarbons and related ether lipids. *Applied microbiology and biotechnology*, 66(5), 486-496.
- Miao, X., Wu, Q. (2006). Biodiesel production from heterotrophic microalgal oil. *Bioresource Technology* 97(6), 841-846.
- Minowa, T., Yokoyama, S. Y., Kishimoto, M., & Okakura, T. (1995). Oil production from algal cells of Dunaliella tertiolecta by direct thermochemical liquefaction. *Fuel*, 74(12), 1735-1738.
- Mohsenpour, S. F., & Willoughby, N. (2016). Effect of CO ₂ aeration on cultivation of microalgae in luminescent photobioreactors. *Biomass and Bioenergy*, 85, 168-177.
- Molles, M.C. Jr. (2008). Ecology Concepts and Applications Fourth Edition.

 McGraw Hill, Boston. 604 pp
- Monari, C., Righi, S., & Olsen, S. I. (2016). Greenhouse gas emissions and energy balance of biodiesel production from microalgae cultivated in photobioreactors in Denmark: a life-cycle modeling. *Journal of Cleaner Production*, 112, 4084-4092.

- Morais, M. G. D., Radmann, E. M., Andrade, M. R., Teixeira, G. G., Brusch, L. R.
 D. F., & Costa, J. A. V. (2009). Pilot scale semicontinuous production of Spirulina biomass in southern Brazil. *Aquaculture*, 294(1), 60-64.
- Morais, S., Mata, T. M., Martins, A. A., Pinto, G. A., & Costa, C. A. (2010). Simulation and life cycle assessment of process design alternatives for biodiesel production from waste vegetable oils. *Journal of Cleaner Production*, 18(13), 1251-1259.
- Moreno, J., Vargas, M. Á., & Guerrero, M. G. (2003). Outdoor cultivation of a nitrogen-fixing marine cyanobacterium, Anabaena sp. ATCC 33047. Biomolecular Engineering, 20(4), 191-197.
- MOSTE (Ministry of Science, Technology and the Environment (Malaysia)) (1999).

 Industrial Processes & The Environment (Handbook No.3.) Crude Palm Oil Industry.
- Mubarak, M., Shaija, A., & Suchithra, T. V. (2015). A review on the extraction of lipid from microalgae for biodiesel production. *Algal Research*, 7, 117-123.
- Muller-Feuga, A., Le Guédes, R., Hervé, A., & Durand, P. (1998). Comparison of artificial light photobioreactors and other production systems using Porphyridium cruentum. *Journal of applied phycology*, 10(1), 83-90.
- Munoz, R., & Guieysse, B. (2006). Algal–bacterial processes for the treatment of hazardous contaminants: a review. *Water research*, 40(15), 2799-2815.
- Murdock, J. N., & Wetzel, D. L. (2009). FT-IR microspectroscopy enhances biological and ecological analysis of algae. *Applied Spectroscopy Reviews*, 44(4), 335-361.
- Nan, Y., Liu, J., Lin, R., & Tavlarides, L. L. (2015). Production of biodiesel from microalgae oil (*Chlorella protothecoides*) by non-catalytic transesterification in supercritical methanol and ethanol: Process optimization. *The Journal of Supercritical Fluids*, 97, 174-182.
- Negoro, M., Shioji, N., Miyamoto, K., & Micira, Y. (1991). Growth of microalgae in high CO₂ gas and effects of SOx and NOx. *Applied biochemistry and biotechnology*, 28(1), 877-886.
- Nor Aini, I. Che Maiman.C.H..Hanirah.H..Zawiah.S.and Che Man.Y.B (1999). Trans free vanspati containing tenery blends of Palm oil-Palm stearin- Palm oil-Palm stearikn –Palm Kernel olein, *Journal of the American oil Chemists society*, 76(5),643-548.

- Norris, J. N. (2010). Marine Algae of the Northern Gulf of California: Chlorophyta and Phaeophyceae.
- Olguín, E. J., Galicia, S., Mercado, G., & Pérez, T. (2003). Annual productivity of Spirulina (Arthrospira) and nutrient removal in a pig wastewater recycling process under tropical conditions. *Journal of Applied Phycology*, 15(2-3), 249-257.
- Ono, E., & Cuello, J. L. (2006). Feasibility assessment of microalgal carbon dioxide sequestration technology with photobioreactor and solar collector. *Biosystems engineering*, 95(4), 597-606.
- Orpez, R., Martínez, M.E., Hodaifa, G., El Yousfi, F., Jbari, N. and Sanchez, S. (2009). Growth of The Microalga *Botryococcus braunii* in Secondarily Treated Sewage. *Desalination*. 246:625-630.
- Osundeko, O., & Pittman, J. K. (2014). Implications of sludge liquor addition for wastewater-based open pond cultivation of microalgae for biofuel generation and pollutant remediation. *Bioresource technology*, 152, 355-363.
- Oswal, N., Sarma, P. M., Zinjarde, S. S., & Pant, A. (2002). Palm oil mill effluent treatment by a tropical marine yeast. *Bioresource Technology*, 85(1), 35-37.
- Oswald, W.J., Gotaas, H.B. (1957). Photosynthesis in Sewage Treatment. *Trans. Am. Soc. Civil Eng.*, 122, 73-75.
- Palińska, K. A., Abed, R. M., Charpy, L., Langlade, M. J., Beltrán-Magos, Y., & Golubic, S. (2015). Morphological, genetic and physiological characterization of Hydrocoleum, the most common benthic cyanobacterium in tropical oceans. *European Journal of Phycology*, 50(2), 139-154.
- Palmer, C. M. (1969). A composite rating of algae tolerating organic pollution. *Journal of Phycology*, 5(1), 78-82.
- Pancha, I., Chokshi, K., Maurya, R., Trivedi, K., Patidar, S. K., Ghosh, A., & Mishra, S. (2015). Salinity induced oxidative stress enhanced biofuel production potential of microalgae Scenedesmus sp. CCNM 1077. Bioresource technology, 189, 341-348.
- Park, J.B.K., Craggs, R.J., Shilton, A.N. (2011). Wastewater treatment high rate algal ponds for biofuel production. *Bioresource Technology*, 102(1), 35–42.
- Park, J. W., Na, S. C., Nguyen, T. Q., Paik, S. M., Kang, M., Hong, D., & Jeon, N. L. (2015). Live cell imaging compatible immobilization of *Chlamydomonas*

- reinhardtii in microfluidic platform for biodiesel research. *Biotechnology and bioengineering*, 112(3), 494-501.
- Park, K. C., Whitney, C. G. E., Kozera, C., O'Leary, S. J., & McGinn, P. J. (2015). Seasonal isolation of microalgae from municipal wastewater for remediation and biofuel applications. *Journal of applied microbiology*.
- Parke, M., Green, J. C., & Manton, I. (1971). Observations on the fine structure of zoids of the genus Phaeocystis (Haptophyceae). *Journal of the Marine Biological Association of the United Kingdom*, 51(04), 927-941.
- Parthasarathy, S., Gomes, R. L., & Manickam, S. (2016). Process intensification of anaerobically digested palm oil mill effluent (AAD-POME) treatment using combined chitosan coagulation, hydrogen peroxide (H2O2) and Fenton's oxidation. *Clean Technologies and Environmental Policy*, 18(1), 219-230.
- Parthasarathy, S., Mohammed, R. R., Fong, C. M., Gomes, R. L., & Manickam, S. (2016). A novel hybrid approach of activated carbon and ultrasound cavitation for the intensification of palm oil mill effluent (POME) polishing. *Journal of Cleaner Production*, 112, 1218-1226.
- Patel, A. K., Huang, E. L., Low-Décarie, E., & Lefsrud, M. G. (2015). Comparative shotgun proteomic analysis of wastewater cultured microalgae: Nitrogen sensing and carbon fixation for growth and nutrient removal in *Chlamydomonas reinhardtii*. *Journal of proteome research*.
- Paudel, A., Jessop, M. J., Stubbins, S. H., Champagne, P., & Jessop, P. G. (2015).
 Extraction of lipids from microalgae using CO 2-expanded methanol and liquid CO₂. Bioresource technology, 184, 286-290.
- Perez-Garcia, O., Escalante, F. M., de-Bashan, L. E., & Bashan, Y. (2011). Heterotrophic cultures of microalgae: metabolism and potential products. *Water research*, 45(1), 11-36.
- Perumal, S., Thirunavukkarasu, A. R., & Pachiappan, P. (Eds.). (2015). *Advances in Marine and Brackishwater Aquaculture*. Springer.
- Pflaster, E. L., Schwabe, M. J., Becker, J., Wilkinson, M. S., Parmer, A., Clemente, T. E., & Riekhof, W. R. (2014). A high-throughput fatty acid profiling screen reveals novel variations in fatty acid biosynthesis in *Chlamydomonas reinhardtii* and related algae. *Eukaryotic cell*, 13(11), 1431-1438.
- Pienkos, P.T., Darzins, A. (2009). The promise and challenges of microalgal-derived biofuels. *Biofuels Bioprod Biorefining*, 3(4),431–440.

- Pittman, J. K., Dean, A. P., and Osundeko, O. (2011). The Potential of Sustainable Algal Biofuel Production Using Wastewater Resources. *Bioresource Technology*. 102: 17-25.
- Poh, P. E., & Chong, M. F. (2009). Development of anaerobic digestion methods for palm oil mill effluent (POME) treatment. *Bioresource Technology*, 100(1), 1-9.
- Ponraj, M., & Din, M. F. M. (2013). Effect of light/dark cycle on biomass and lipid productivity by Chlorella pyrenoidosa using palm oil mill effluent (POME). *J. Sci. Ind. Res*, 72(11), 703-706.
- Prinz, D., Juliani, A., & Brontowiyono, W. (2015). Future water management problems in Asian megacities. *Jurnal Sains Dan Teknologi Lingkungan*.
- Pulz, O. (2001). Photobioreactors: production systems for phototrophic microorganisms. *Applied microbiology and biotechnology*, 57(3), 287-293.
- Pulz, O., & Gross, W. (2004). Valuable products from biotechnology of microalgae. *Applied microbiology and biotechnology*, 65(6), 635-648.
- Pushparaj, B., Pelosi, E., Tredici, M. R., Pinzani, E., & Materassi, R. (1997). As integrated culture system for outdoor production of microalgae and cyanobacteria. *Journal of Applied Phycology*, 9(2), 113-119.
- Putri EV, Din MFM, Ahmed Z, Jamaluddin H, Chelliapan S Investigation of microalgae for high lipid content using palm oil mill effluent (Pome) as carbon source. In: International Conference on Environment and Industrial Innovation. IPCBEE, 2011.
- Qin, J. (2005). Bio-hydrocarbons from algae: impacts of temperature, light and salinity on algae growth. *Rirdc*.
- Rajkumar, R., Yaakob, Z., & Takriff, M. S. (2013). Potential of micro and macro algae for biofuel production: a brief review. *Bioresources*, 9(1), 1606-1633.
- Rao, A. R., Dayananda, C., Sarada, R., Shamala, T. R., & Ravishankar, G. A. (2007). Effect of salinity on growth of green alga Botryococcus braunii and its constituents. *Bioresource technology*, 98(3), 560-564.
- Ratledge, C. (1982). Microbial oils and fats: an assessment of their commercial potential algae, yeasts, fungi. *Progress in industrial microbiology*.
- Rawat, I., Kumar, R. R., Mutanda, T., & Bux, F. (2011). Dual role of microalgae: phycoremediation of domestic wastewater and biomass production for sustainable biofuels production. *Applied Energy*, 88(10), 3411-3424.

- Redfield, A.C., Ketchum, B.H., Richards, F.A. (1963). The Influence of Organisms on the Composition of Seawater. New York: Interscience Publication.26-77.
- Reimann, B. E., & Lewin, J. C. (1964). The diatom genus Cylindrotheca Rabenhorst. *Journal of the Royal Microscopical Society*, 83(3), 283-296.
- Ren, H. Y., Liu, B. F., Kong, F., Zhao, L., & Ren, N. (2015). Hydrogen and lipid production from starch wastewater by co-culture of anaerobic sludge and oleaginous microalgae with simultaneous COD, nitrogen and phosphorus removal. *Water Research*, 85, 404-412.
- Richmond, A. (2004). Handbook of microalgal culture: biotechnology and applied phycology. Blackwell Science Ltd.
- Richmond, A., Lichtenberg, E., Stahl, B., & Vonshak, A. (1990). Quantitative assessment of the major limitations on productivity of Spirulina platensis in open raceways. *Journal of Applied Phycology*, 2(3), 195-206.
- Risgaard-Petersen, N. (2003). Coupled nitrification-denitrification in autotrophic and heterotrophic estuarine sediments: On the influence of benthic microalgae. *Limnology and Oceanography*, 48(1), 93-105.
- Rizkytata, B. T., Gumelar, M. T., & Abdullah, T. H. (2014). Industrial Tofu Wastewater as a Cultivation Medium of Microalgae Chlorella vulgaris. *Energy Procedia*, 47, 56-61.
- Rodolfi, L., Chini Zittelli, G., Bassi, N., Padovani, G., Biondi, N., Bonini, G., & Tredici, M. R. (2009). Microalgae for oil: Strain selection, induction of lipid synthesis and outdoor mass cultivation in a low-cost photobioreactor. *Biotechnology and bioengineering*, 102(1), 100-112.
- Romano, I., Bellitti, M. R., Nicolaus, B., Lama, L., Manca, M. C., Pagnotta, E., & Gambacorta, A. (2000). Lipid profile: a useful chemotaxonomic marker for classification of a new cyanobacterium in Spirulina genus. *Phytochemistry*, 54(3), 289-294.
- Rosenberg, J. N., Oyler, G. A., Wilkinson, L., & Betenbaugh, M. J. (2008). A green light for engineered algae: redirecting metabolism to fuel a biotechnology revolution. *Current opinion in Biotechnology*, 19(5), 430-436.
- Rupani, P.F., Singh, R. P., Ibrahim, M. H., and Esa N. (2010). Review of Current Palm Oil Mill Effluent (POME) Treatment Methods: Vermicomposting as a Sustainable Practice. *World Applied Sciences Journal*, 11(1),70-81.

- Salihu, A., & Alam, M. Z. (2012). Palm oil mill effluent: a waste or a raw material?. *Journal of Applied Sciences Research*, (January), 466-473.
- Sanches-Silva A, de Quirós AR, López-Hernández J, Paseiro-Losada P. (2004). Determination of hexanal as indicator of the lipid oxidation state in potato crisps using gas chromatography and high-performance liquid chromatography, *Journal of Chromatography* A 1046(1-2):75-81.
- Sarno, D., Kooistra, W. H., Medlin, L. K., Percopo, I., & Zingone, A. (2005). Diversity in the Genus Skeletonema (*Bacillariophyceae*). An assessment of The taxonomy of S. Costatum-like species with the description of four new species. *Journal of phycology*, 41(1), 151-176.
- Schenk, P. M., Thomas-Hall, S. R., Stephens, E., Marx, U. C., Mussgnug, J. H., Posten, C. & Hankamer, B. (2008). Second generation biofuels: highefficiency microalgae for biodiesel production. *Bioenergy research*, 1(1), 20-43.
- Searchinger, T., Heimlich, R., Houghton, R. A., Dong, F., Elobeid, A., Fabiosa, J., ... & Yu, T. H. (2008). Use of US croplands for biofuels increases greenhouse gases through emissions from land-use change. *Science*, 319(5867), 1238-1240.
- Selvarajan, R., Felföldi, T., Tauber, T., Sanniyasi, E., Sibanda, T., & Tekere, M. (2015). Screening and evaluation of some green algal strains (*Chlorophyceae*) isolated from freshwater and soda lakes for biofuel production. *Energies*, 8(7), 7502-7521.
- Sethupathi, S. (2004). Removal Of Residue Oil From Palm Oil Mill Effluent (Pome) Using Chitosan (Doctoral dissertation, Universiti Sains Malaysia).
- Setlik, I., Sust, V., & Malek, I. (1970). Dual purpose open circulation units for large scale culture of algae in temperate zones. I. Basic design considerations and scheme of a pilot plant. *Algological Studies/Archiv für Hydrobiologie, Supplement Volumes*, 111-164.
- Shafik, H. M., Saad, M. G., & El-Serehy, H. A. (2015). Impact of nitrogen regime on fatty acid profiles of Desmodesmus quadricaudatus and Chlorella sp. and ability to produce biofuel. *Acta Botanica Hungarica*, 57(1), 205-218.
- Shafiqah, N., & Nasir, M. (2013). Development of membrane anaerobic system (MAS) for palm oil mill effluent (POME) treatment.

- Shaker, S., Nemati, A., Montazeri-Najafabady, N., Mobasher, M. A., Morowvat, M. H., & Ghasemi, Y. (2015). Treating Urban Wastewater: Nutrient Removal by Using Immobilized Green Algae in Batch Cultures. *International Journal of Phytoremediation*, (just-accepted).
- Shantha, N. C., & Napolitano, G. E. (1992). Gas chromatography of fatty acids. *Journal of Chromatography* A, 624(1), 37-51.
- Sharon-Gojman, R., Maimon, E., Leu, S., Zarka, A., & Boussiba, S. (2015). Advanced methods for genetic engineering of Haematococcus pluvialis (Chlorophyceae, Volvocales). *Algal Research*, 10, 8-15.
- Sheehan, J., Camobreco, V., Duffield, J., Shapouri, H., Graboski, M., & Tyson, K. S. (2000). *An overview of biodiesel and petroleum diesel life cycles* (No. NREL/TP-580-24772). National Renewable Energy Lab., Golden, CO (US).
- Sheehan, J., Dunahay, T., Benemann, J., & Roessler, P. (1998). A look back at the US Department of Energy's Aquatic Species Program: Biodiesel from algae (Vol. 328). Golden: National Renewable Energy Laboratory.
- Shen, Y., Yuan, W., Pei, Z., Mao, E. (2008). Culture of microalga *Botryococcus* in livestock wastewater. *Transactions of the ASABE* 51 (4), 1395–1400.
- Shuping, Z., Yulong, W., Mingde, Y., Chun, L., & Junmao, T. (2010). Pyrolysis characteristics and kinetics of the marine microalgae Dunaliella tertiolecta using thermogravimetric analyzer. *Bioresource Technology*, 101(1), 359-365.
- Sialve, B., Bernet, N., & Bernard, O. (2009). Anaerobic digestion of microalgae as a necessary step to make microalgal biodiesel sustainable. *Biotechnology advances*, 27(4), 409-416.
- Sicko-Goad, L., & Andresen, N. A. (1991). Effect of growth and light/dark cycles on diatom lipid content and composition1. *Journal of Phycology*, 27(6), 710-718.
- Sigee, D. C., Bahrami, F., Estrada, B., Webster, R. E., & Dean, A. P. (2007). The influence of phosphorus availability on carbon allocation and P quota in Scenedesmus subspicatus: a synchrotron-based FTIR analysis. *Phycologia*, 46(5), 583-592.
- Singh, R.P., Ibrahim, M.H., Esa N. (2010). Composting of waste from palm oil mill: A sustainable waste management practice. *Review in Environmental Science and Biotechnology*, DOI: 10.1007/s11157-010-9199-2.

- Singh, S.P. and Singh, P. (2015). Effect of temperature and light on the growth of algae species: A review. *Renewable and Sustainable Energy Reviews*, 50:431-444.
- Solovchenko, A., et al. (2008). Effects of light intensity and nitrogen starvation on growth, total fatty acids and arachidonic acid in the green microalga Parietochloris incisa. *Journal of Applied Phycology*, 20(3),245-251.
- Sorokin, C., & Krauss, R. W. (1958). The Effects of Light Intensity on the Growth Rates of Green Algae. *Plant physiology*, 33(2), 109.
- Spolaore, P., Joannis-Cassan, C., Duran, E., & Isambert, A. (2006). Commercial applications of microalgae. *Journal of bioscience and bioengineering*, 101(2), 87-96.
- Stehfest, K., Toepel, J., & Wilhelm, C. (2005). The application of micro-FTIR spectroscopy to analyze nutrient stress-related changes in biomass composition of phytoplankton algae. *Plant Physiology and Biochemistry*, 43(7), 717-726.
- Stephenson, A. L., Dennis, J. S., Howe, C. J., Scott, S. A., & Smith, A. G. (2010). Influence of nitrogen-limitation regime on the production by Chlorella vulgaris of lipids for biodiesel feedstocks. *Biofuels*, 1(1), 47-58.
- Stief, P., Poulsen, M., Nielsen, L. P., Brix, H., & Schramm, A. (2009). Nitrous oxide emission by aquatic macrofauna. *Proceedings of the National Academy of Sciences*, 106(11), 4296-4300.
- Stuart, B. (1997). Biological applications of Infrared Spectroscopy. *willey: chichester*, 25-180.
- Suárez-Garcia, F., Martinez-Alonso, A., Llorente, M. F., & Tascon, J. M. D. (2002). Inorganic matter characterization in vegetable biomass feedstocks. *Fuel*, 81(9), 1161-1169.
- Suganya, T., Varman, M., Masjuki, H. H., & Renganathan, S. (2016). Macroalgae and microalgae as a potential source for commercial applications along with biofuels production: A biorefinery approach. *Renewable and Sustainable Energy Reviews*, 55, 909-941.
- Suh, I. S., & Lee, C. G. (2003). Photobioreactor engineering: design and performance. *Biotechnology and Bioprocess Engineering*, 8(6), 313-321.

- Sumathi, S., Chai, S. P., & Mohamed, A. R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, 12(9), 2404-2421.
- Sun, L., Jin, X., Zhong, Y. et al (2006). Changes of Algal Communities in Water Body with Different Proportions of Nitrogen and Phosphorus. *Chinese Journal of Applies Ecology*, 17(7), 1218-1223.
- Sydney, E. B., Sturm, W., de Carvalho, J. C., Thomaz-Soccol, V., Larroche, C., Pandey, A., & Soccol, C. R. (2010). Potential carbon dioxide fixation by industrially important microalgae. *Bioresource Technology*, 101(15), 5892-5896.
- Takemura, N., Iwkume, T., Rusuno, M. (1985). Photosynthesis and Primary Production of Microcystis agruginosa in Lake Kasumigaura. *Journal of Plankton Research*, 7(3), 303-312.
- Tan, X., Kong, F., Yu, Y., et al. (2009). Effects of Enhanced Temperature on Algae Recruitment and Phytoplankton Community Succession. *China Environmental Science*, 29(6), 578-582.
- Tarlan, E., Dilek, F. B., and Yetis, U. (2002). Effectiveness of Algae in The Treatment of A Wood-Based Pulp and Paper Industry Wastewater. *Bioresource Technology*. 84: 1-5.
- Tilman, D. (1982). Resource Competition and Community Structure. New Jersey: *Princeton University Press.* P.98-99.
- Tindall, B. J., Kämpfer, P., Euzéby, J. P., & Oren, A. (2006). Valid publication of names of prokaryotes according to the rules of nomenclature: past history and current practice. *International Journal of Systematic and Evolutionary Microbiology*, 56(11), 2715-2720.
- Tomas, C. R. (1980). Olisthodiscus Luteus (*Chrysophyceae*). V. Its Occurrence, Abundance and Dynamics in Narragansett Bay, Rhode Island. *Journal of Phycology*, 16(2), 157-166.
- Travieso Córdoba, L., Domínguez Bocanegra, A. R., Rincón Llorente, B., Sánchez Hernández, E., Benítez Echegoyen, F., Borja, R., & Colmenarejo Morcillo, M. F. (2008). Batch culture growth of *Chlorella zofingiensis* on effluent derived from two-stage anaerobic digestion of two-phase olive mill solid waste. *Electronic Journal of Biotechnology*, 11(2), 12-19.

- Turon, V., Baroukh, C., Trably, E., Latrille, E., Fouilland, E., & Steyer, J. P. (2015). Use of fermentative metabolites for heterotrophic microalgae growth: yields and kinetics. *Bioresource technology*, 175, 342-349.
- Vairappan, C. S., & Yen, A. M. (2008). Palm oil mill effluent (POME) cultured marine microalgae as supplementary diet for rotifer culture. *Journal of applied phycology*, 20(5), 603-608.
- Valderrama, L.T., Del Campo, C.M., Rodriguez, C.M., de-Bashan, L.E., and Bashan, Y. (2002). Treatment of recalcitrant wastewater from ethanol and citric acid production using the microalga *Chlorella vulgaris* and the macrophyte Lemna minuscula. *Water Res.*, 36, 4185–92.
- Valenzuela-Espinoza, E., Millán-Núñez, R., & Núñez-Cebrero, F. (2002). Protein, carbohydrate, lipid and chlorophyll a content in Isochrysis aff. galbana (clone T-Iso) cultured with a low cost alternative to the f/2 medium. *Aquacultural Engineering*, 25(4), 207-216.
- Vigani, M., Parisi, C., Rodríguez-Cerezo, E., Barbosa, M. J., Sijtsma, L., Ploeg, M., & Enzing, C. (2015). Food and feed products from micro-algae: market opportunities and challenges for the EU. *Trends in Food Science & Technology*, 42(1), 81-92.
- Vijayaraghavan, K., Hemanathan, K. (2009). Biodiesel production from freshwater algae. *Energy Fuels*, 23,5448-5453.
- Wagner, J., Bransgrove, R., Beacham, T. A., Allen, M. J., Meixner, K., Drosg, B., & Chuck, C. J. (2016). Co-production of bio-oil and propylene through the hydrothermal liquefaction of polyhydroxybutyrate producing cyanobacteria. *Bioresource technology*, 207, 166-174.
- Wah, W. P., Sulaiman, N. M., Nachiappan, M., & Varadaraj, B. (2002). Pretreatment and membrane ultrafiltration using treated palm oil mill effluent (POME). Songklanakarin J. Sei. Technol, 24, 891-898.
- Walne, P. R. (1970). Studies on the food value of nineteen genera of algae to juvenile bivalves of the genera Ostrea, Crassostrea, Mercenaria and Mytilus. *Fish. Invest. Ser.* 2, 26(5).
- Wan, C., Alam, M. A., Zhao, X. Q., Zhang, X. Y., Guo, S. L., Ho, S. H., & Bai, F. W. (2015). Current progress and future prospect of microalgal biomass harvest using various flocculation technologies. *Bioresource technology*, 184, 251-257.

- Wang, B., Li, Y., Wu, N., & Lan, C. Q. (2008). CO2 bio-mitigation using microalgae. *Applied Microbiology and Biotechnology*, 79(5), 707-718.
- Wang, H. T., Yao, C. H., Liu, Y. N., Meng, Y. Y., Wang, W. L., Cao, X. P., & Xue, S. (2015). Identification of fatty acid biomarkers for quantification of neutral lipids in marine microalgae Isochrysis zhangjiangensis. *Journal of Applied Phycology*, 27(1), 249-255.
- Wang, L., Li, Y., Chen, P., Min, M., Chen, Y., Zhu, J., Ruan, R.R. (2010a). Anaerobic digested dairy manure as a nutrient supplement for cultivation of oil-rich green microalgae *Chlorella sp. Bioresource Technology*, 101, 2623-2628.
- Wang, L., Min, M., Li, Y., Chen, P., Chen, Y., Liu, Y., & Ruan, R. (2010b).
 Cultivation of green algae *Chlorella sp.* in different wastewaters from municipal wastewater treatment plant. *Applied biochemistry and biotechnology*, 162(4), 1174-1186.
- Wehr, J. D., & Sheath, R. G. (2003). Freshwater habitats of algae. *Freshwater Algae of North America*. *Academic Press*, San Diego, CA, 918, 308.
- Wehr, J. D., Sheath, R. G., & Kociolek, J. P. (Eds.). (2015). Freshwater algae of North America: ecology and classification. Elsevier.
- Weissman, J. C., & Tillett, D. T. (1992). Design and operation of an outdoor microalgae test facility: large-scale system results. *Aquatic Species Project Report*, FY 1989, 90, 32-56.
- Weldy, C. S., & Huesemann, M. I. C. H. A. E. L. (2007). Lipid production by Dunaliella salina in batch culture: effects of nitrogen limitation and light intensity. *US Department of Energy Journal of Undergraduate Research*, 7(1), 115-122.
- Welsh, D. T., Bartoli, M., Nizzoli, D., Castaldelli, G., Riou, S. A., & Viaroli, P. (2000). Denitrification, nitrogen fixation, community primary productivity and inorganic-N and oxygen fluxes in an intertidal Zostera noltii meadow. *Marine Ecology Progress Series*, 208(5).
- Widjaja, A., Chien, C. C., & Ju, Y. H. (2009). Study of increasing lipid production from fresh water microalgae Chlorella vulgaris. *Journal of the Taiwan Institute of Chemical Engineers*, 40(1), 13-20.
- Woelkerling, W. J. (1988). The coralline red algae: an analysis of the genera and subfamilies of non *geniculate Corallinaceae*. Oxford University Press.

- Wu Q.Y., Yin S., Sheng G. and Fu J. (1994). New Discoveries In Study On Hydrocarbons From Thermal Degradation Of Heterotrophically Yellowing Algae. *Sci China* (*B*). 37:326–351.
- Wu, T.Y., Abdul Wahab, M., Jahim, J.M., Anuar, N. (2007). A holistic approach to managing palm oil mill effluent (POME) Biotechnological advances in the sustainable reuse of POME. *Biotechnology Advances.*, 27, 40-52.
- Wu, W. T., & Hsieh, C. H. (2008). Cultivation of microalgae for optimal oil production. *Journal of Biotechnology*, 136, S521.
- Wu, Z., & Shi, X. (2007). Optimization for high-density cultivation of heterotrophic *Chlorella* based on a hybrid neural network model. *Letters in applied microbiology*, 44(1), 13-18.
- Xin, L., Hong-ying, H., Ke, G., Ying-xue, S. (2010). Effects of different nitrogen and phosphorus concentrations on the growth, nutrient uptake, and lipid accumulation of a freshwater microalga Scenedesmus sp. *Bioresource Technology*, 101, 5494-5500.
- Xu, H., Yang, L., Mao, H. et al. (2006). Dynamic Studies on the Effect of Phosphorus on the Growth of Microcysis aeruginosa and Scendesmus oblique. *Ecology and Environmental Sciences*, 15(5), 921-924.
- Yacob, S., Hassan, M. A., Shirai, Y., Wakisaka, M., & Subash, S. (2005). Baseline study of methane emission from open digesting tanks of palm oil mill effluent treatment. *Chemosphere*, 59(11), 1575-1581.
- Yacob, S., Shirai, Y., Hassan, M.A., Wakisaka M., and Subash, S. (2006). Start-up operation of semi-commercial closed anaerobic digester for palm oil mill effluent treatment. *Process Biochemistry*, 41(4), 962-964.
- Yadavalli, R., Rao, C. S., Rao, R. S., & Potumarthi, R. (2014). Dairy effluent treatment and lipids production by Chlorella pyrenoidosa and Euglena gracilis: study on open and closed systems. *Asia-Pacific Journal of Chemical Engineering*, 9(3), 368-373.
- Yan, H., & Pan, G. (2002). Toxicity and bioaccumulation of copper in three green microalgal species. *Chemosphere*, 49(5), 471-476.
- Yang, J., Cao, J., Xing, G., & Yuan, H. (2015). Lipid production combined with biosorption and bioaccumulation of cadmium, copper, manganese and zinc by oleaginous microalgae *Chlorella minutissima* UTEX2341. *Bioresource technology*, 175, 537-544.

- Yang, X., Liu, P., Hao, Z., Shi, J., & Zhang, S. (2011). Characterization and identification of freshwater microalgal strains toward biofuel production. *BioResources*, 7(1), 0686-0695.
- Yee, W. (2015). Feasibility of various carbon sources and plant materials in enhancing the growth and biomass productivity of the freshwater microalgae Monoraphidium griffithii NS16. *Bioresource technology*, 196, 1-8.
- Yoo, C., Jun, S. Y., Lee, J. Y., Ahn, C. Y., & Oh, H. M. (2010). Selection of microalgae for lipid production under high levels carbon dioxide. *Bioresource technology*, 101(1), S71-S74.
- Yue, L.C., W. (2005). Isolation and determination of cultural characteristics of a new highly tolerant fresh water microalgae. . *Energy Conversion and Management*, 46, 1868-1878.
- Yusoff, S., Hansen, S.B., (2007). Feasibility study of performing of life cycle assessment on crude palm oil production in Malaysia, *The International J. Life Cycle Assessment*, 12(1): 50-58.
- Yvon-Durocher, G., Dossena, M., Trimmer, M., Woodward, G., & Allen, A. P. (2015). Temperature and the biogeography of algal stoichiometry. *Global Ecology and Biogeography*, 24(5), 562-570.
- Zang, C., Huang, S., Wu, M., et al. (2011). Comparison of Relationships Between pH, Dissolved Oxygen and Chlorophyll a for Aquaculture and Nonaquaculture Waters. *Water Air and Soil Pollution*, 219(1-4), 157-174.
- Zhang, X. X., Zhang, T., & Fang, H. H. (2009). Antibiotic resistance genes in water environment. *Applied microbiology and biotechnology*, 82(3), 397-414.
- Zhang, Y. M., Chen, H., He, C. L., & Wang, Q. (2013). Nitrogen starvation induced oxidative stress in an oil-producing green alga *Chlorella sorokiniana* C3. *PloS one*, 8(7), e69225.
- Zhu, L., Wang, Z., Shu, Q., Takala, J., Hiltunen, E., Feng, P., & Yuan, Z. (2013). Nutrient removal and biodiesel production by integration of freshwater algae cultivation with piggery wastewater treatment. *Water research*, 47(13), 4294-4302.
- Zhu, M., Zhang, X. C., & Mao, Y. X. (2003). Effects of Tempearture, Salinity and Illumination on the Growth of *Thalassiosira sp. MARINE SCIENCES-QINGDAO-CHINESE EDITION-*, 27(12), 61-69.