# SYNTHESIS AND OPTIMIZATION OF MICRO-SIZED BACTERIAL-BASED VIOLET PIGMENT USING RESPONSE SURFACE METHODOLOGY

HARYANI BINTI MOHD YATIM

UNIVERSITI TEKNOLOGI MALAYSIA

# SYNTHESIS AND OPTIMIZATION OF MICRO-SIZED BACTERIAL-BASED VIOLET PIGMENT USING RESPONSE SURFACE METHODOLOGY

HARYANI BINTI MOHD YATIM

A dissertation submitted in partial fulfilment of the requirements for the award of the degree of Master of Science (Chemistry)

> Faculty of Science Universiti Teknologi Malaysia

> > JANUARY 2017

To My Beloved Mak & Ayah

#### ACKNOWLEDGEMENT

In the name of ALLAH S.W.T., The Most Gracious and The Most Merciful.

Upon completion of this project, I would like to express my gratitude to the following people and many parties for all their help and involvement during the accomplishment of this project. First and foremost, a billion gratitude and appreciation to my supervisor, Dr. Siti Aminah binti Setu for .her guidance, encouragement, and kindness. It would be hard to complete this project without her support.

I also would like to express my deepest gratitude to Prof. Dr. Wan Azlina Ahmad for allowing me using the instrument in the biotechnology laboratory and also for her willingness to share her expert opinion with me. I would like to extend my appreciation to all the biotechnology laboratory members for their assistance in ensuring the smoothness of the project. Furthermore, thank you to Universiti Teknologi Malaysia for giving me an opportunity to conduct this project as partial fulfillment of the requirement for the award of the degree of Master of Science Chemistry. I gain new experiences, acquaintances and knowledge from here.

Last but not least, I wish to express my utmost gratitude and appreciation to both my parents, Mohd Yatim bin Asral and Kamariah binti Bachok for all their trust and support in terms of moral and financial towards me throughout this time. Not to forget to all people in my family especially my youngest sister, Khairiah for her willingness to assist me during my hard time, Kak Haryati and Abg Zuhairi as well as my nieces (Aliya, Afrina and Athirah) and also Abang Khairi with his wife, Kak Liza for all their continuous support and motivation for me to complete my research.

#### ABSTRACT

Violet pigments, extracted from a bacteria known as *Chromobacterium* violaceum, has raised the enthusiasm of researchers in conducting comprehensive studies on these pigments due to their diverse biological activities include antibacterial and antioxidant properties. There is, however, a limitation related with the solubility of the violet pigment, by which it is commonly dissolved in toxic solvents such as dimethyl sulfoxide (DMSO) and methanol instead of being soluble in biological fluids and water. This approach did not synchronized with the public demands for products that are both eco-friendly and safe towards the environment and human body. Hence, this study provides a method to synthesise the violet pigment in microscale through an encapsulation technique using chitosan-tripolyphosphate (TPP) microparticles. Owing to the exceptional properties of high surface to volume ratio of microparticles, the solubility of the violet pigment in water and biological fluid could be improved. The synthesis of microparticles in this study involved ionic gelation between chitosan and TPP, in which several parameters were taken into consideration in order to control the dispersion stability of the violet pigment in the suspension. It is well known that particles in microscale will tend to aggregate, thus causing diminution of their biological activities. Therefore, preparation parameters, including the concentration of chitosan, tripolyphosphate (TPP) and pigment as well as the mass ratio of chitosan to TPP, were optimized using Response Surface Methodology (RSM). The aim was to obtain small particles size down to microscale with low range of polydispersity index (PDI) and high zeta potential. Minimum particle size of 149.0 nm with polydispersity index of 0.367 and zeta potential of +23.40 mV was obtained at the optimal formulations of 2.33 mg/mL of Cs, 1.5 mg/mL of TPP and 1 ppm of violet pigment and at mass ratio of chitosan:TPP of 7:1

### ABSTRAK

Pigmen ungu, diekstrak daripada bakteria dikenali sebagai Chromobacterium violaceum, telah menimbulkan semangat para penyelidik dalam menjalankan penyelidikan yang menyeluruh ke atas pigmen ini disebabkan oleh aktiviti-aktiviti biologi mereka termasuk sifat nyahbakteria dan sifat nyahoksida. Walaubagaimanapun, terdapat beberapa masalah berkait dengan kelarutan pigmen ungu di mana kebiasaannya larut di dalam pelarut yang toksik seperti dimethyl sulfoxide (DMSO) dan methanol dimana sepatutnya menjadi larut di dalam cecair biologi dan air. Pendekatan ini tidak selari dengan permintaan awam terhadap barang yang mesra alam dan selamat untuk alam persekitaran dan badan manusia. Oleh itu, kajian ini menyediakan kaedah dalam mensintesis pigmen ungu dalam skala mikro melalui teknik pengkapsulan dengan chitosan-tripolyphosphate (TPP) mikropartikel. Kelarutan pigmen ungu ke dalam air dapat diatasi disebabkan sifat luar biasanya jaitu tinggi nisbah antara permukaan dan isi padu. Sintesis mikropartikel di dalam kajian ini melibatkan penggelan ionik antara chitosan dan TPP dimana beberapa parameter telah dititikberatkan bagi mengawal kestabilan pembubaran pigmen ungu di dalam ampaian. Ianya telah diketahui ramai bahawa partikel di skala mikro cenderung untuk mengalami penggumpalan yang juga mengurangkan aktiviti-aktiviti biologinya. Oleh itu, parameter-parameter penyediaan dalam kajian ini termasuk kepekatan chitosan, tripolyphosphate (TPP) dan pigmen serta nisbah jisim chitosan kepada TPP telah dioptimumkan menggunakan Response Surface Methodology (RSM). Ini bertujuan untuk mendapatkan saiz partikel yang kecil sehingga skala mikro dengan indeks taburan (PDI) yang rendah dan tinggi nilai potensi zeta. Saiz minimum partikel, 149.0 nm bersama PDI, 0.367 dan potensi zeta, +23.40 mV telah diperolehi di formulasi optimum, 2.33 mg/mL chitosan, 1.5 mg/mL TPP dan 1 ppm pigmen ungu pada nisbah jisim chitosan: TPP 7:1.

## **TABLE OF CONTENT**

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	V
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	X
	LIST OF FIGURES	xi
	LIST OF ABBREVIATIONS	xiii
1	INTRODUCTION	1
	1.1 Background of study	1
	1.2 Problem Statement	2
	1.3 Objectives	3
	1.4 Scope of Study	3
	1.5 Significance of Study	3
2	LITERATURE REVIEW	5
	2.1 Pigments	5
	2.1.1 Denotation of pigment	5
	2.1.2 Synthetic pigments	7
	2.1.3 Natural pigments	8
	2.1.3.1 Bacterial pigments	9
	2.1.3.2 Violet pigment: Violacein	10

2.2 Nanoparticles	
2.2.1 Definition and application	
2.2.2 Chitosan nanoparticles	
ion method of chitosan nanoparticles	15
Emulsion and cross-linking	16
Emulsion-droplet coalescence	17
Emulsion solvent diffusion	18
Reverse micellar method	19
Desolvation	20
Ionic gelation and polyelectrolyte	21
complexation	
nking agent	22
Glutaraldehyde	23
Tripolyphosphate	24
nt as stabilizer in formation of	25
ticles	
rface Methodology	27
Composite Design (CCD)	28
nnken Design (BBD)	29
optimization of chitosan nanoparticles	30
ND METHODS	32
gents	32
	32
micro-sized violet pigment	33
3.4 Optimization of micro-sized violet pigment	
3.4.1 Experimental design	
alysis and optimization of the applied	34
tion of the applied models	35
3.5 Characterization of micro-sized violet pigment	
	s on and application an anoparticles ion method of chitosan nanoparticles ion method of chitosan nanoparticles Emulsion and cross-linking Emulsion-droplet coalescence Emulsion solvent diffusion Reverse micellar method Desolvation Ionic gelation and polyelectrolyte complexation on thing agent Glutaraldehyde Tripolyphosphate Int as stabilizer in formation of ticles rface Methodology Composite Design (CCD) Inken Design (BBD) Ion optimization of chitosan nanoparticles IND METHODS gents Imicro-sized violet pigment Iof micro-sized violet pigment Iof micro-si

3

	3.5.1 Particle size, PDI and zeta potential	36
	3.5.2 Field emission scanning electron microscopy	36
	(FESEM)	
	3.6 Operational framework	37
4	<b>RESULTS AND DISCUSSION</b>	38
	4.1 Formation of micro-sized violet pigment	38
	4.2 Data analysis and optimization	40
	4.3 Validation of the models	48
	4.4 Morphological analysis using FESEM	49
5	CONCLUSIONS AND RECOMMENDATION	51
	5.1 Conclusion	51
	5.2 Recommendations	52

# REFERENCES

53

## LIST OF TABLES

TABLE	TITLE	PAGE
2.1	Pigment producing bacterial species	9
2.2	Parameters and response involved in optimization of	31
	chitosan nanoparticles and the significancy of the each term	
3.1	Variables in Box Behnken design (BBD) used for	35
	optimization of micro-sized bacterial pigment	
4.1	Measured responses of dependent variables in BBD	41
4.2	ANOVA for the response surface model of responses	43
	particle size, PDI and zeta potential.	
4.3	Quadratic model and the regression coefficients for particle	44
	size (R1), PDI (R2)and zeta potential (R3) of micro-sized	
	violet pigment.	
4.4	Optimized formulation of micro-sized violet pigment, the	48
	predicted and actualvalues of the responses	

## LIST OF FIGURES

FIGURE	TITLE	
2.1	Schematic diagram of absorption and reflection of light on flat surface with pigment particles	6
2.2	Important groups for appearance of color of 4- Hydroxyazobenzene	10
2.3	Chemical structure of violacein	11
2.4	Protonation of chitosan dissolved in acetic acid solution	
2.5	Schematic representations of cross-linked chitosan nanoparticles with anionic molecules (polyanion), polyelectrolyte and water-soluble neutral polymers	15
2.6	Schematic representations of preparation of chitosan nanoparticles by emulsion and cross-linking	16
2.7	Schematic representations on preparation of chitosan nanoparticles by emulsion-droplet coalescence	17
2.8	Schematic representations on preparation of chitosan nanoparticles by emulsion solvent diffusion	18
2.9	Schematic representations on preparation of chitosan nanoparticles by reverse micellar	20
2.10	Schematic representations on preparation of chitosan nanoparticles by desolvation	21
2.11	Schematic representations on preparation of chitosan nanoparticles by ionic gelation	22
2.12	Function of cross-linking in formation of chitosan nanoparticles	23

2.13	Postulated cross-linking structure by glutaraldehyde	24
2.14	Structure of TPP	24
2.15	General structure of surfactant	25
2.16	Hydrophilic part and hydrophobic tail of Polysorbate 80	27
2.17	Schematic models of central composite design for evaluating	28
	three factors	
2.18	Graphical representation of a) cube for BBD b) three	29
	interlocking of 2 <sup>2</sup> factorial design	
3.1	Operational framework of the present study	37
4.1	Particle size distribution of micro-sized violet pigment by intensity	39
4.2	Zeta potential distribution of micro-sized violet pigment.	40
4.3	Predicted versus (vs.) actual plots of (a) particle size and (b)	42
	zeta potential of micro-sized violet pigment	
4.4	Perturbation plots for (a) particle size (b) zeta potential	45
4.5	(a) 3D response surface graphs and (b) 2D contour plots	46
	illustrating the interaction effect of concentration of chitosan	
	(Cs), A and mass ratio of Cs:TPP, on the particle size.	
4.6	(a) 3D response surface graphs and (b) 2D contour plots	47
	showing the interaction effect of concentration of chitosan	
	(Cs), A and mass ratio of Cs:TPP, C on the zeta potential.	
4.7	FESEM images of (a) Blank (chitosan-tripolyphosphate	49
	microparticles) (b) Optimized micro-sized violet pigment at	
	100,000 magnifications	

### LIST OF ABBREVIATIONS

BBD	Box- Behnken design
Cs	Chitosan
°C	Degree Celcius
DMSO	Dimethyl sulfoxide
DLS	Dynamic Light Scattering
FESEM	Field Emission Scanning Electron Microscope
h	hours
L	Liter
μL	Microliter
μm	Micrometer
mg	Milligram
min	minutes
mL	Milliliter
mV	Millivolt
М	Molar
nm	Micrometer
ppm	Part per million
PDI	Polydispersity index
RSM	Response surface methodology
$R^2$	R squared
rpm	Revolutions per minute
TPP	Tripolyphosphate
% v/v	Volume/volume percent

### **CHAPTER 1**

### INTRODUCTION

### **1.1 Background of study**

Natural and synthetic pigments have been applied in food, clothes, cosmetics, inks, papers and many other materials whereby they imparted beauty in all the materials by transmitting color. Nowadays, people around the world have begun to reset and change their perspectives and concerns toward health issues and environmental conservation. This resulted into an upsurge in demand for natural pigments from the industries due to their non-toxicity, non-carcinogenicity and biodegradable properties (Aruldass, 2015). Pigments have become essential and although they are present in all organisms due to their primary function as a colorant, only natural pigments have specific functions. Some of those functions include photosynthesis in plants, oxygen and carbon dioxide transportation in organisms and as a protective screen in humans and other vertebrates. Moreover, there are a many pigments that possess biological activities such as antibacterial, antioxidant, anticancer and many others (Delgado Vargas *et al.*, 2000; Kumar *et al.*, 2015). Microorganism is recognized as a potential source of natural pigment over plants due to their advantages of high yield, stability, cost efficiency, accessibility and easy scale-up production of pigment (Tuli *et al.*, 2014).

In nature, numerous microorganisms, including yeast, fungi, algae and bacteria, have been known to produce pigments. However, bacteria offers several exceptional advantages owing to their short half life cycle, invulnerable toward season and climate, ability to create various pigments with different colors and shades as well as having an easier scale-up production process and many others (Sutthiwong *et al.*, 2014). In this study, violet pigment produced by *Chromobacterium violaceum* UTM5 was utilized due to its interesting pharmacological properties including antibacterial activity and their facile growth in common laboratory media such as nutrient agar (Aruldass, 2015).

### **1.2 Problem Statement**

Demands on natural pigment, primarily on bacterial pigment, have been rising for decades due to its peculiar features that are independent toward environmental changes to provide various shades and colors of pigment, as well as exhibiting an easy scale-up production of pigment. Violet pigment produced from Chromobacterium violaceum UTM5 has great potential in pharmaceutical applications due to its many properties such as antimicrobial, antiviral, antiprotozoal and antioxidant properties. Violet pigment is composed of two derivatives which are violacein and deoxyviolacein. However, violacein derivatives of the violet pigment have a limitation, in where they can only dissolved in toxic solvents such as dimethyl sulfoxide (DMSO) and methanol. Hence, encapsulation of this bacterial-based violet pigments with the polymeric chitosan-tripolyphosphate (chitosan-TPP) microparticles is expected to improve its solubility owing to the distinct features of moderately large activated surface area to volume ratio. This allows the interaction between the pigment and water molecules to occur more easily. Thus, this could improve the dissolving process of the violet pigment without the use of toxic solvents. Nevertheless, chitosan-tripolyphosphate (Cs-TPP) microparticles prepared by ionotropic gelation method have been known to aggregate or fuse directly after preparation and has a limited stability during storage. Therefore, optimization of preparation parameters such as Cs, TPP and pigment concentration as well as mass ratio of Cs to TPP was carried out using Response Surface Methodology. The purpose of the optimization was to acquire the optimum conditions for attaining micro-sized violet pigment with low polydispersity index and high zeta potential.

### **1.3** Objectives of study

The objectives of this study are:

- i. To synthesize micro-sized bacterial-based violet pigment *via* encapsulation technique.
- To optimize the preparation and stabilization parameters using Response Surface Methodology (RSM)
- iii. To characterize the optimized formulated micro-sized violet pigment

### **1.4** Scope of study

Micro-sized violet pigment was synthesized *via* encapsulation technique with chitosan-tripolyphosphate (Cs-TPP) microparticles. Ionic gelation method was applied as method to prepare Cs-TPP microparticles. The synthesized micro-sized violet pigment were characterized for their physico-chemical properties which included average particle size distribution, polydispersity index (PDI) and zeta potential using Zetasizer Nano (Malvern, UK.). Optimization of the preparation parameters of micro-sized violet pigment was analysed using Response Surface Methodology (RSM) using the Box-Behnken design. Then, the optimized micro-sized violet pigment obtained using the optimal RSM formulation was characterized for their morphology using Field Emission Scanning Electron Microscope (FESEM).

### **1.5** Significance of study

The development of micro-sized violet pigment from this study could improve the solubility of violet pigment which is known as poorly water-soluble pigment. Additionally, the reduction in size of violet pigment into micro-scale could minimize its toxicity towards human in order to be applied as an additive and a colorant in various materials such as foods, cosmetics, clothes, ink, paper and so forth. The synthesized micro-sized violet pigment can also possibly act as therapeutic agents in the pharmaceutical fields. Moreover, the optimization studies could give ideas on the important preparation parameters to be considered when synthesizing violet pigment in the micro-scale.

#### REFERENCES

- Abul Kalam, M., Khan, A. A., Khan, S., Almalik, A., and Alshamsan, A. (2016). Optimizing indomethacin-loaded chitosan nanoparticle size, encapsulation, and release using Box-Behnken experimental design. *International Journal of Biological Macromolecules*, 87, 329–40.
- Abdel-Hafez, S. M., Hathout, R. M., and Sammour, O. A. (2014). Towards better modeling of chitosan nanoparticles production: Screening different factors and comparing two experimental designs. *International Journal of Biological Macromolecules*, 64, 334–340.
- Adegoke, O. A., Kyu, J. K., and Mukherjee, A. (2012). In vitro genotoxicity evaluation of 4-carboxyl-2,6-dinitrophenylazohydroxynaphthalenes using human lymphocytes. *Food and Chemical Toxicology*, 50(3–4), 936–941.
- Agnihotri, S. A., Mallikarjuna, N. N., and Aminabhavi, T. M. (2004). Recent advances on chitosan-based micro- and nanoparticles in drug delivery. *Journal of Controlled Release*, *100*(1), 5–28.
- Ahmad, W. A., Ahmad, W. Y. W., Zakaria, Z. A., and Yusof, N. Z. (2012a). *Application* of *Bacterial Pigments as Colorant*. Springer Berlin Heidelberg.
- Ahmad, W. A., Yusof, N. Z., Nordin, N., Zakaria, Z. A., and Rezali, M. F. (2012b). Production and characterization of violacein by locally isolated chromobacterium violaceum grown in agricultural wastes. *Applied Biochemistry and Biotechnology*, 167(5), 1220–1234.

- Alishahi, A., Mirvaghefi, A., Tehrani, M. R., Farahmand, H., Shojaosadati, S. A., Dorkoosh, F. A., and Elsabee, M. Z. (2011). Shelf life and delivery enhancement of vitamin C using chitosan nanoparticles. *Food Chemistry*, 126(3), 935–940.
- Alshatwi, A. A., Subash-Babu, P., and Antonisamy, P. (2016). Violacein induces apoptosis in human breast cancer cells through up regulation of BAX, p53 and down regulation of MDM2. *Experimental and Toxicologic Pathology*, 68(1), 89– 97.
- Andrighetti-Fröhner, C. R., Antonio, R. V., Creczynski-Pasa, T. B., Barardi, C. R. M., and Simões, C. M. O. (2003). Cytotoxicity and Potential Antiviral Evaluation of Violacein Produced by Chromobacterium violaceum. *Memorias Do Instituto Oswaldo Cruz*, 98(6), 843–848.
- Antoniou, J., Liu, F., Majeed, H., Qi, J., Yokoyama, W., and Zhong, F. (2015). Physicochemical and morphological properties of size-controlled chitosan– tripolyphosphate nanoparticles. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 465, 137–146.
- Aruldass, C. A., Rubiyatno, R., Venil, C. K., and Ahmad, W. A. (2015). Violet pigment production from liquid pineapple waste by Chromobacterium violaceum UTM5 and evaluation of its bioactivity. *Royal Society of Chemistry Advances*, 5(64), 51524– 51536.
- Aruldass, C. A. (2015) Violet pigment from Chromobacterium violaceum UTM5 grown in liquid pineapple waste and its antibacterial and cytotoxicity activities. Ph.D Thesis, Universiti Teknologi Malaysia, Skudai.
- Bao, H., Li, L., and Zhang, H. (2008). Influence of cetyltrimethylammonium bromide on physicochemical properties and microstructures of chitosan-TPP nanoparticles in aqueous solutions. *Journal of Colloid and Interface Science*, 328(2), 270–277.
- Barnett, J. R., Miller, S., and Pearce, E. (2006). Colour and art: A brief history of pigments. *Optics and Laser Technology*, 38(4–6), 445–453.

- Barnett, J. C. (2007). Synthetic organic dyes, 1856-1901: an introductory literature review of their use and related issues in textile conservation. *Reviews in Conservation*, 3630(8), 67–77.
- Berthold, A., Cremer, K., and Kreuter, J. (1996). Preparation and characterization of chitosan microspheres as drug carrier for prednisolone sodium phosphate as model for antiinflammatory drugs. *Journal of Controlled Release*, 39(1), 17–25.
- Bharmoria, P., Singh, T., and Kumar, A. (2013). Complexation of chitosan with surfactant like ionic liquids: molecular interactions and preparation of chitosan nanoparticles. *Journal of Colloid and Interface Science*, 407, 361–9.
- Bodnar, M., Hartmann, J. F., and Borbely, J. (2005). Preparation and characterization of chitosan-based nanoparticles. *Biomacromolecules*, 6(5), 2521–2527.
- Calvo P, Remuñan-Lopez C., Vila-Jato J. L. and Alonso M. J. (1997). Novel hydrophilic chitosan polyethylene oxide nanoparticles as protein carriers. *Journal of Applied Polymer Science*, 63, 125-132.(Cota-Arriola et al., 2013)
- Chronopoulou, L., Massimi, M., Giardi, M. F., Cametti, C., Devirgiliis, L. C., Dentini, M., and Palocci, C. (2013). Chitosan-coated PLGA nanoparticles: A sustained drug release strategy for cell cultures. *Colloids and Surfaces B: Biointerfaces*, 103, 310– 317.
- Cota-Arriola, O., Cortez-Rocha, M. O., Ezquerra-Brauer, J. M., Lizardi-Mendoza, J., Burgos-Hernández, A., Robles-Sánchez, R. M., and Plascencia-Jatomea, M. (2013).
  Ultrastructural, Morphological, and Antifungal Properties of Micro and Nanoparticles of Chitosan Crosslinked with Sodium Tripolyphosphate. *Journal of Polymers and the Environment*, 21(4), 971–980.
- Dash, M., Chiellini, F., Ottenbrite, R. M., and Chiellini, E. (2011). Chitosan A versatile semi-synthetic polymer in biomedical applications. *Progress in Polymer Science* (*Oxford*), 36(8), 981–1014.

- Delgado-Vargas, F., Jiménez, a R., and Paredes-López, O. (2000). Natural pigments:carotenoids, anthocyanins, and betalain characteristics, biosynthesis, processing, and stability. *Critical Reviews in Food Science and Nutrition*, 40, 173– 289.
- Duan, Y., Xu, S., Wang, Q., Liu, J., and Zhang, Z. (2006). Optimization of preparation of DHAQ-loaded PEG-PLGA-PEG nonaparticles using central composite design. *Journal of Materials Science: Materials in Medicine*, 17(6), 559–563.
- Durán, N., and Menck, C. F. (2001). Chromobacterium violaceum: A Review of Pharmacological and Industiral Perspectives. Critical Reviews in Microbiology, 27(3), 201–22.
- Durán, M., Ponezi, A. N., Faljoni-Alario, A., Teixeira, M. F. S., Justo, G. Z., and Durán, N. (2011). Potential applications of violacein: a microbial pigment. *Medicinal Chemistry Research*, 21(7), 1524–1532.
- Fàbregas, A., Miñarro, M., García-Montoya, E., Pérez-Lozano, P., Carrillo, C., Sarrate, R., and Suñé-Negre, J. M. (2013). Impact of physical parameters on particle size and reaction yield when using the ionic gelation method to obtain cationic polymeric chitosan-tripolyphosphate nanoparticles. *International Journal of Pharmaceutics*, 446(1–2), 199–204.
- Fan, W., Yan, W., Xu, Z., and Ni, H. (2012). Colloids and Surfaces B : Biointerfaces Formation mechanism of monodisperse, low molecular weight chitosan nanoparticles by ionic gelation technique. *Colloids and Surfaces B: Biointerfaces*, 90, 21–27.
- Ferreira, S. L. C., Bruns, R. E., Ferreira, H. S., Matos, G. D., David, J. M., Brandão, G. C., and dos Santos, W. N. L. (2007). Box-Behnken design: An alternative for the optimization of analytical methods. *Analytica Chimica Acta*, 597(2), 179–186.
- Feyzioglu, G. C., and Tornuk, F. (2016). Development of chitosan nanoparticles loaded with summer savory (Satureja hortensis L.) essential oil for antimicrobial and

antioxidant delivery applications. LWT - *Food Science and Technology*, 70, 104–110.

- Gan, Q., Wang, T., Cochrane, C., and McCarron, P. (2005). Modulation of surface charge, particle size and morphological properties of chitosan-TPP nanoparticles intended for gene delivery. *Colloids and Surfaces B: Biointerfaces*, 44(2–3), 65–73.
- Gan, Q., and Wang, T. (2007). Chitosan nanoparticle as protein delivery carrier-Systematic examination of fabrication conditions for efficient loading and release. *Colloids and Surfaces B: Biointerfaces*, 59(1), 24–34.
- Gendy, T. S., El-Temtamy, S. A., Ghoneim, S. A., El-Salamony, R. A., El-Naggar, A. Y., and El-Morsi, A. K. (2016). Response surface methodology for carbon dioxide reforming of natural gas. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 38(9), 1236–1245.
- Gokce, Y., Cengiz, B., Yildiz, N., Calimli, A., and Aktas, Z. (2014). Ultrasonication of chitosan nanoparticle suspension: Influence on particle size. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 462, 75–81.
- Green, P., and MacDonald, L. (2011) Colour Engineering: Achieving Device Independent Colour Volume 30 of Wiley Series in Display Technology. John Wiley & Sons.
- Grenha, A. (2012). Chitosan nanoparticles: A survey of preparation methods, *Journal of Drug Targeting*, 20 (4), 291-300.
- Gulati, N., Nagaich, U., and Saraf, S. A. (2013). Intranasal Delivery of Chitosan Nanoparticles for Migraine Therapy. *Scientia Pharmaceutica*, 81, 843–854.
- Gürses, A., Açıkyıldız, M., Güneş, K., and Gürses, M. S. (2016). Dyes and Pigments: Their Structure and Properties. In *Dyes and Pigments*, 13-29,. Springer International Publishing.

- Hamidi, M., Azadi, A., and Rafiei, P. (2008). Hydrogel nanoparticles in drug delivery. *Advanced Drug Delivery Reviews*, 60(15), 1638–1649.
- Hashad, R. A, Ishak, R. a H., Geneidi, A. S., and Mansour, S. (2016). Methotrexate loading in chitosan nanoparticles at a novel pH: Response surface modeling, optimization and characterization. *International Journal of Biological Macromolecules*, 91, 630–639.
- Honary, S., Ebrahimi, P., and Hadianamrei, R. (2014). Optimization of Particle Size and Encapsulation Efficiency of Vancomycin Nanoparticles by Response Surface Methodology. *Pharmaceutical Development and Technology*, 19(8), 987–998.
- Hussain, Z., and Sahudin, S. (2016). Preparation, characterisation and colloidal stability of chitosan- tripolyphosphate nanoparticles: Optimisation of formulation and process parameters. *International Journal of Pharmacy and Pharmaceutical Sciences*, 8(3).
- Jong, W. H. D.and Borm, P. J. A. (2008). Drug Delivery and Nanoparticles: Applications and Hazards. *International Journal of Nanomedicine*. 3(2), 133-149
- Kaialy, W., and Al Shafiee, M. (2016). Recent advances in the engineering of nanosized active pharmaceutical ingredients: Promises and challenges. *Advances in Colloid* and Interface Science, 228, 71–91.
- Konzen, M., De Marco, D., Cordova, C. A. S., Vieira, T. O., Antônio, R. V., and Creczynski-Pasa, T. B. (2006). Antioxidant properties of violacein: Possible relation on its biological function. *Bioorganic and Medicinal Chemistry*, 14(24), 8307–8313.
- Kumar, A., Vishwakarma, H. S., Singh, J., Dwivedi, S., and Kumar, M. (2015). Microbial pigments: production and their applications in various industries. *International Journal of Pharmaceutical, Chemical & Biological Sciences*, 5(1), 203-12.

- Kunjachan, S., and Jose, S. (2010). Understanding the mechanism of ionic gelation for synthesis of chitosan nanoparticles using qualitative techniques. *Asian Journal of Pharmaceutics*, 4(2), 148.
- Li, B. Z., Wang, L. J., Li, D., Bhandari, B., Li, S. J., Lan, Y., and Mao, Z. H. (2009). Fabrication of starch-based microparticles by an emulsification-crosslinking method. *Journal of Food Engineering*, 92(3), 250–254.
- Li, H.-J., Zhang, A.-Q., Hu, Y., Sui, L., Qian, D.-J., and Chen, M. (2012). Large-scale synthesis and self-organization of silver nanoparticles with Tween 80 as a reductant and stabilizer. *Nanoscale Research Letters*, 7(1), 612.
- Li, X., Qin, Y., Liu, C., Jiang, S., Xiong, L., and Sun, Q. (2016). Size-controlled starch nanoparticles prepared by self-assembly with different green surfactant: The effect of electrostatic repulsion or steric hindrance. Food Chemistry, 199, 356–363.
- Liyanapathirana, C., and Shahidi, F. (2005). Optimization of extraction of phenolic compounds from wheat using response surface methodology. *Food Chemistry*, 93(1), 47–56.
- Lomax, S. Q., and Learner, T. (2006). A Review of the Classes, Structures, and Methods of Analysis of Synthetic Organic Pigments. *Journal of the American Institute for Conservation*, 45(2), 107–125.
- López-León, T., Carvalho, E. L. S., Seijo, B., Ortega-Vinuesa, J. L., and Bastos-González, D. (2005). Physicochemical characterization of chitosan nanoparticles: electrokinetic and stability behavior. *Journal of Colloid and Interface Science*, 283(2), 344–351.
- Martins, D., Costa, F. T. M., Brocchi, M., and Durán, N. (2011). Evaluation of the antibacterial activity of poly-(d,l-lactide-co-glycolide) nanoparticles containing violacein. *Journal of Nanoparticle Research*, 13(1), 355–363.

- Mi, F. L., Huang, C. T., Liang, H. F., Chen, M. C., Chiu, Y. L., Chen, C. H., and Sung,
  H. W. (2006). Physicochemical, antimicrobial, and cytotoxic characteristics of a chitosan film cross-linked by a naturally occurring cross-linking agent, aglycone geniposidic acid. *Journal of Agricultural and Food Chemistry*, 54(9), 3290–3296.
- Mitra, S., Gaur, U., Ghosh, P. C., and Maitra, A. N. (2001). Tumour targeted delivery of encapsulated dextran-doxorubicin conjugate using chitosan nanoparticles as carrier. Journal of Controlled Release, 74(1–3), 317–323. https://doi.org/10.1016/S0168-3659(01)00342-X
- Myers, R. H., Montgomery, D. C., and Anderson-Cook, C. M. (2016). Response surface methodology: process and product optimization using designed experiments. John Wiley & Sons.
- Nipun Babu, V., and Kannan, S. (2012). Enhanced delivery of baicalein using cinnamaldehyde cross-linked chitosan nanoparticle inducing apoptosis. *International Journal of Biological Macromolecules*, 51(5), 1103–1108.
- Niwa T, Takeuchi H, Hino T, Kunou N, Kawashima Y. 1993. Preparation of biodegradable nanospheres of water soluble and insoluble drugs with D,L-lactide/glycolide copolymer by a novel spontaneous emulsification solvent diffusion method, and the drug release behavior. *Journal Control Release*, 2, 89-98.
- Ohya, Y., Shiratani, M., Kobayashi, H., and Ouchi, T. (1994). Release behavior of 5fluorouracil from chitosan-gel nanospheres immobilizing 5-fluorouracil coated with polysaccharides and their cell specific cytotoxicity. *Journal of Macromolecular Science—Pure and Applied Chemistry*, 31(5), 629-642.
- Orna, M. V. (2012). The Chemical History of Color, 167. Springer Berlin Heidelberg
- Pan, H., Feng, J., He, G.X., Cerniglia, C. E., and Chen, H. (2012). Evaluation of impact of exposure of Sudan azo dyes and their metabolites on human intestinal bacteria. *Anaerobe*, 18(4), 445–53.

- Patel, N. V., Sheth, N. R., and Mohddesi, B. (2015). Formulation and Evaluation of Genistein – A Novel Isoflavone Loaded Chitosan and Eudragit® Nanoparticles for Cancer Therapy. *Materials Today: Proceedings*, 2(9), 4477–4482.
- Pereira, D. M., Valentão, P., and Andrade, P. B. (2014). Marine natural pigments: Chemistry, distribution and analysis. *Dyes and Pigments*, 111, 124–134.
- Picone, C. S. F., and Cunha, R. L. (2013). Chitosan-gellan electrostatic complexes: Influence of preparation conditions and surfactant presence. *Carbohydrate Polymers*, 94(1), 695–703.
- Pillai, S. K., and Ray, S. S. (2012). Chitosan-based Nanocomposites. Maya, J. Sabu, T. (Eds.) *Natural Polymers*, 2, 33-45. The Royal Society of Chemistry.
- de Pinho Neves, A. L., Milioli, C. C., Müller, L., Riella, H. G., Kuhnen, N. C., and Stulzer, H. K. (2014). Factorial design as tool in chitosan nanoparticles development by ionic gelation technique. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 445, 34–39.
- Rajalakshmi, R., Indira Muzib, Y., Aruna, U., Vinesha, V., Rupangada, V., and Krishna Moorthy, S. B. (2014). Chitosan nanoparticles -an emerging trend in nanotechnology. *International Journal of Drug Delivery*, 6(3), 204–229.
- Rampino, A., Borgogna, M., Blasi, P., Bellich, B., and Cesàro, A. (2013). Chitosan nanoparticles: preparation, size evolution and stability. *International journal of pharmaceutics*, 455(1-2), 219–28.
- Rapp, G. (2009). Pigments and Colorants. Archaeomineralogy (pp. 201–221). Berlin, Heidelberg: Springer Berlin Heidelberg.
- Rath, A., Mathesan, S., and Ghosh, P. (2015). Nanomechanical characterization and molecular mechanism study of nanoparticle reinforced and cross-linked chitosan biopolymer. *Journal of the Mechanical Behavior of Biomedical Materials*, 55, 42– 52.

- Sarvaiya, J., and Agrawal, Y. K. (2015). Chitosan as a suitable nanocarrier material for anti-Alzheimer drug delivery. *International Journal of Biological Macromolecules*, 72, 454–65.
- Sasidharan, A., Sasidharan, N. K., Amma, D. B. N. S., Vasu, R. K., Nataraja, A. V., and Bhaskaran, K. (2015). Antifungal activity of violacein purified from a novel strain of Chromobacterium sp. NIIST (MTCC 5522). *Journal of Microbiology*, 53(10), 694–701.
- Shah, B., Khunt, D., Misra, M., and Padh, H. (2016). Application of Box-Behnken design for optimization and development of quetiapine fumarate loaded chitosan nanoparticles for brain delivery via intranasal route *International Journal of Biological Macromolecules*, 89, 206–218.
- Sugashini, S., and Begum, K. M. M. S. (2013). Optimization using central composite design (CCD) for the biosorption of Cr (VI) ions by cross linked chitosan carbonized rice husk (CCACR). *Clean Technologies and Environmental Policy*, 15(2), 293–302.
- Sutthiwong, N., Fouillaud, M., Valla, A., Caro, Y., and Dufossé, L. (2014). Bacteria belonging to the extremely versatile genus Arthrobacter as novel source of natural pigments with extended hue range. *Food Research International*, 65, 156–162.
- Tang, Z. X., Qian, J. Q., and Shi, L. E. (2007). Preparation of chitosan nanoparticles as carrier for immobilized enzyme. *Applied Biochemistry and Biotechnology*, 136(1), 77–96.
- Tokumitsu, H., Ichikawa, H., and Fukumori, Y. (1999). Chitosan-gadopentetic acid complex nanoparticles for gadolinium neutron-capture therapy of cancer: preparation by novel emulsion-droplet coalescence technique and characterization. *Pharmaceutical Research*, 16(12), 1830-1835.

- Tuli, H. S., Chaudhary, P., Beniwal, V., and Sharma, A. K. (2015). Microbial pigments as natural color sources: current trends and future perspectives. *Journal of Food Science and Technology*, 52(8), 4669–78.
- Vaezifar, S., Razavi, S., Golozar, M. A., Karbasi, S., Morshed, M., and Kamali, M. (2013). Effects of Some Parameters on Particle Size Distribution of Chitosan Nanoparticles Prepared by Ionic Gelation Method. *Journal of Cluster Science*, 24(3), 891–903.
- Vauthier, C., and Bouchemal, K. (2009). Methods for the Preparation and Manufacture of Polymeric Nanoparticles. *Pharmaceutical Research*, 26(5), 1025–1058.
- Venil, C. K., Zakaria, Z. A. and Ahmad, W. A. (2013) Bacterial Pigments and Their Applications. *Process Biochemistry*. 48, 1065-1079.
- Venil, C. K., Aruldass, C. A., Abd Halim, M. H., Khasim, A. R., Zakaria, Z. A., and Ahmad, W. A. (2015). Spray drying of violet pigment from Chromobacterium violaceum UTM 5 and its application in food model systems. *International Biodeterioration & Biodegradation*, 102, 324–329.
- Wijesena, R. N., Tissera, N., Kannangara, Y. Y., Lin, Y., Amaratunga, G. a J., and De Silva, K. M. N. (2015). A method for top down preparation of chitosan nanoparticles and nanofibers. *Carbohydrate polymers*, 117, 731–8.
- Yuan, Y., Chesnutt, B. M., Utturkar, G., Haggard, W. O., Yang, Y., Ong, J. L., and Bumgardner, J. D. (2007). The effect of cross-linking of chitosan microspheres with genipin on protein release. *Carbohydrate Polymers*, 68(3), 561–567.
- Yusof, N. Z. (2010) Isolation and application of violet pigment extracted from Chromobacterium Violaceum. MSc. Thesis, Universiti Teknologi Malaysia, Skudai.

- Zhang, W., Qiao, X., Chen, J., and Chen, Q. (2008). Self-assembly and controlled synthesis of silver nanoparticles in SDS quaternary microemulsion. *Materials Letters*, 62(10–11), 1689–1692.
- Zhao, Y., Wang, Z., Zhang, W., and Jiang, X. (2010). Adsorbed Tween 80 is unique in its ability to improve the stability of gold nanoparticles in solutions of biomolecules. *Nanoscale*, 2(10), 2114-2119.
- Zhang, Q., Wu, Q., Lin, D., and Yao, S. (2013). Effect and mechanism of sodium chloride on the formation of chitosan–cellulose sulfate–tripolyphosphate crosslinked beads. *Soft Matter*, 9, 10354.
- Zolgharnein, J., Shahmoradi, A., and Ghasemi, J. B. (2013). Comparative study of Box-Behnken, central composite, and Doehlert matrix for multivariate optimization of Pb (II) adsorption onto Robinia tree leaves. *Journal of Chemometrics*, 27(1–2), 12–20.