

**ANTI-TYROSINASE AND CYTOTOXIC ACTIVITIES OF SYNTHETIC
COMPOUND 2'-HYDROXY-3,4,3',4'-TETRAMETHOXYCHALCONE**

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Dedicated to:

My beloved parents,

Haji Ahmad bin Haji Ramli and Hajah Awa binti Othman

who give me strength and full support,

my family,

my lecturers,

and my friends.

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ABSTRACT

It has been demonstrated by several studies that synthetic chalcone compound products having antioxidant and anti tyrosinase properties. In this study, the novel synthetic 2'-hydroxy-3,4,3',4'-tetramethoxychalcone (HTMC) was evaluated for their antityrosinase, antioxidant, and cytotoxicity properties. In anti tyrosinase activities, HTMC showed an activity and is postulated as potential tyrosinase inhibitor. The antityrosinase activity of HTMC, however, was lower than Kojic Acid (positive control). It was due to the structure of HTMC that have been modified synthetically at position 4 by inserting methoxy group. For that reason, the potency of HTMC as tyrosinase inhibitor was reduced and this statement supports our finding that is HTMC have less active compared to kojic acid. In contrast, HTMC demonstrate low antioxidant activities for both DPPH and FRAP assays. The highest percentage of inhibition for HTMC was 37% in DPPH assay at the concentration of 242 μM whilst the highest FRAP value of HTMC was 2 351 $\mu\text{mol/L}$. It is due to their chemical structure that contains only 1 group of hydroxyl in A-ring at position 2 that reflected to its ability to scavenge free radicals. HTMC showed negligible toxicity effect on Chinese Hamster Ovary (CHO) cell at the concentration of (0, 0.01, 0.1 and 1) μM . As it considered as preliminary report on cytotoxicity effect of HTMC, this data is valuable to be further investigated on human cell line.

ABSTRAK

Beberapa kajian penyelidikan telah menunjukkan bahawa produk kalkon sintetik mempunyai aktiviti sebagai antioksidan dan anti-tyrosinase. Dalam kajian ini, sebatian sintetik novel iaitu 2'-hydroxy-3,4,3',4'-tetramethoxychalcone (HTMC) telah dinilai untuk sifat-sifat antityrosinase, antioksidan, dan sitotoksiti mereka. Dalam aktiviti anti-tyrosinase, HTMC menunjukkan aktiviti dan diandaikan berpotensi sebagai perencat enzim tyrosinase. Walau bagaimanapun, aktiviti antityrosinase HTMC adalah rendah berbanding asid kojik (kawalan positif). Ini disebabkan oleh struktur kimia HTMC telah diubah suai secara sintetik pada kedudukan 4 dengan memasukkan kumpulan metoksi. Atas sebab itu, potensi HTMC sebagai perencat tyrosinase dikurangkan dan kenyataan ini disokong oleh keputusan yang diperolehi bahawa HTMC adalah kurang aktif berbanding dengan asid kojik. Sementara itu, HTMC menunjukkan aktiviti antioksidan yang rendah untuk kedua-dua jenis assay iaitu DPPH dan FRAP. HTMC mencatatkan peratus tertinggi perencatan sebanyak 37% dalam DPPH assay (pada kepekatan 242 μM) manakala nilai FRAP pula adalah 2 351 $\mu\text{mol/L}$. Ia adalah disebabkan oleh struktur kimia sebatian ini yang mengandungi hanya 1 kumpulan hidroksil dalam cincin A yang mempengaruhi keupayaan untuk mengumpul radikal bebas. HTMC menunjukkan kesan toksik yang kecil terhadap sel Chinese Hamster Ovary (CHO) pada kepekatan (0, 0.01, 0.1 and 1) μM . Memandang ini merupakan laporan pertama untuk kesan toksik oleh HTMC, data ini penting untuk dilanjutkan pada sel manusia.

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LIST OF ABBREVIATIONS AND SYMBOLS

<i>et al.</i> ,	:	and others
EtOH	:	ethanol
NaOH	:	sodium hydroxide
HCl	:	acid chloride
UV	:	Ultraviolet
g	:	gram
μM	:	Micromolar
%	:	percent
s	:	seconds
min	:	minutes
v/v	:	volume per volume
°C	:	degree Celsius
ml	:	millilitre
mg	:	milligram
<i>sp.</i>	:	species
DMSO	:	Dimethyl sulfoxide
HTMC	:	2'-hydroxy-3,4,3',4'-tetramethoxychalcone
DMEM	:	Dulbecco's Modified Eagle Medium
FBS	:	fetal bovine serum
DPPH assay	:	1,1-Diphenyl-2-picrylhydrazyl
FRAP assay	:	Ferric Reducing Ability of Plasma
MTT assay	:	3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
L-DOPA	:	L-3,4- dihydroxyphenylalanine,
ANOVA	:	Analysis Of Variance
IC ₅₀	:	50% inhibitory concentration
nm	:	nanometer
OH	:	Hydroxyl group
UV	:	Ultraviolet
v/v	:	Volume per volume
°C	:	Degree of Celsius

MeOH	:	Methanol
CV	:	Coefficient of Variations
e.g	:	example
mg	:	Miligram
μ l	:	Microliter
μ g	:	Microgram

CHAPTER 1

INTRODUCTION

1.1 Background of Research

Natural antioxidants, mostly in fruits and vegetables have grown a great concern among consumers as epidemiological studies have signified that frequent consumption of natural antioxidant is associated with lower risk of cardiovascular disease and cancer (Renaud *et al.*, 1998; Temple *et al.*, 2000). Three major groups of defensive effects of natural antioxidant in fruits and vegetables are vitamins, phenolics and caratenoids (Halliwell, 1996). Phenolics groups is identified by the presence of a hydroxyl group (-OH) bonded directly to an aromatic hydrocarbon group. Variety of phenolic compounds has been discovered previously to contain secondary structure called flavonoids. One of six skeleton structure of the main classess of flavonoids is chalcone. Avila *et al.*, (2008) reported that hydroxylation at position 2' in A-ring of chalcone is indirectly promoting structural stability.

In chemistry, organic synthesis specifically becomes core of attraction for scientists due to the ability to construct beneficial products artificially for human goods. It have been demonstrated by several studies about synthetic chalcone product including 2',5'-dihydroxy-4-chloro-dihydrochalcone and 2',5'-dihydroxydihydrochalcone that inhibit the iNOS protein expression and the former

compound also inhibits cyclooxygenase-2 (COX-2) activity in RAW 264.7 cells (Ko *et al.*, 2003). The antioxidant activity of some synthetic 2'-hydroxy-chalcone and its derivatives has been investigated by Detsi and her colleagues (2009). They suggested that among tested compounds with methoxy group on positions 2' and 4' of the A-ring, possess combined antioxidant-LOX inhibitory profile (Detsi *et al.*, 2009). Besides, Nerya *et al.* (2004) showed the relatedness between structure of chalcone and tyrosinase inhibition activity. These findings implied that some chalcone may be promising as anti-inflammatory, antioxidant and antityrosinase agents and have potential in medical perspective.

Tyrosinase is a copper-containing enzyme that extensively distributed in nature. This enzyme is also known as polyphenol oxidase (Whitaker, 1995). Two major reactions that catalyzes by tyrosinase is relating molecular oxygen in the melanin biosynthesis pathway: a) the hydroxylation of monophenols to *o*-phenols (monophenolase activity), and b) the oxidation of the *o*-phenols to *o*-quinones (diphenolase activity). These quinones are reactive compound and have a tendency to polymerize spontaneously to form melanin (Seo *et al.*, 2003). This scenario can be determined by various dermatological disorders or the production of abnormal pigmentation, for instance age spot, freckles, melasma and liver spot that can be a serious aesthetic setback (Briganti *et al.*, 2003). Therefore, tyrosinase inhibitors have become progressively vital in medication and in cosmetic as it can block melogenic pathway by inhibiting enzymatic oxidation. On top of that, natural tyrosinase inhibitors are in general believed to be fewer side effects and can be produced at reasonable low cost (Saewan *et al.*, 2011).

1.2 Problem statement

Synthetic compounds of chalcone namely 2'-hydroxy-3,4,3',4'-tetramethoxychalcone in this study have been synthesized via classical methods of Claisen-Schmidt condensation reaction. Their physicochemical properties are reflected to their extent bioactivities such as anti-tyrosinase and antioxidant

activities. Previous research supports that this plant-derived polyphenolic compound is believed to possess a potential as whitening agents. It is based on its chemical structure that contains hydroxyl group in A-ring at position 2 and methoxyl groups in A- and B-ring of this compound. Thus far, the 2'-hydroxy-3,4,3',4'-tetramethoxychalcone synthetic compound, however, is not discovered precisely in term of their bioactivities. Therefore, this study is performed to explore pharmacologically effects of this synthetic compound to overcome some of the limitations associated with constraint in the production of natural pure compound.

1.3 Objectives

- a) To investigate an anti-tyrosinase effect of synthetic compound namely 2'-hydroxy-3,4,3',4'-tetramethoxychalcone (HTMC)
- b) To evaluate antioxidant property of HTMC by using DPPH and FRAP assay
- c) To determine cytotoxicity effect of HTMC on the growth of Chinese Hamster Ovary (CHO) cell.

1.4 Scope of work

In this study, synthetic compound namely 2'-hydroxy-3,4,3',4'-tetramethoxychalcone was obtained from Dr. Farediah binti Ahmad, Department of Chemistry, Faculty of Science, UTM. Anti-tyrosinase assay was done in order to determine the ability of this synthetic compound to inhibit tyrosinase enzyme in the synthesis of melanin. The antioxidant activity of this compound was determined in terms of their anti radical power as assessed by DPPH radical scavenging assay and FRAP assay. Furthermore, IC_{50} (half maximal inhibitory concentration) of this synthetic compound is also can be demonstrated from these assays. MTT assay was performed so as to assess the cell viability for evaluation of different concentration of synthetic compound that are introduced to CHO cell line.

1.5 Significant of research

To date, biological activities explicitly anti-tyrosinase, antioxidant and cytotoxicity effects of this synthetic compound is not revealed by previous researcher. Therefore, this study hypothesized that synthetic compound namely 2'-hydroxy-3,4,3',4'-tetramethoxychalcone have antioxidant properties and tyrosinase inhibition activity. The cytotoxicity effect of this compound can serve an overview that reflected to their ability to be toxic to human cell at certain concentration. These 3 indicators will be manipulated for further study to support their applications in modern and traditional medicine.

REFERENCES

- Aljadi, A.M. and Kamaruddin, M.Y. (2004). Evaluation of the phenolic contents and antioxidant capacities of two Malaysian floral honeys. *Food Chemistry*. 85: 513–518.
- Ani, V., Varadaraj, M. C., and Naidu, K. A. (2006). Antioxidant and antibacterial activities of polyphenolic compounds from bitter cummin (*Cuminum nigrum L.*). *European Food Research and Technology*. 224: 109–115.
- Avila, H. P., Smania, F. A., Monache, F. D. and Smania, J. A. (2008). Structure–activity relationship of antibacterial chalcones. *Bioorganic and Medicinal Chemistry*. 16: 9790–9794.
- Bandgar, B. P., Shrikant, S. G., Ragini, G. B., Jalinder, V. T. and Chandrahas, N. K. (2010). Synthesis and biological evaluation of simple methoxylated chalcones as anticancer, anti-inflammatory and antioxidant agents. *Bioorganic and Medicinal Chemistry*. 18: 1364–1370.
- Benzie, I. F. F. and Strain, J. J. (1999). Ferric reducing/antioxidant power assay: Direct measure of the total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. *Methodology in Enzymology*. 299:15-27.
- Benzie, I. F. F. and Strain, J. J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant power”: The FRAP assay. *Analytical Biochemistry*. 239:70-76.

- Blois, M.S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*. 181: 1199-1200.
- Brand-Williams, W., Cuvelier, M. E. and Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. *Lebensmittel-Wissenschaft Technologie*. 28: 25–30.
- Briganti, S., Camera, E. and Picardo, M. (2003). Chemical and instrumental approaches to treat hyperpigmentation. *Pigment Cell Research*. 16: 101-110.
- Cadenas, E. and Packer, L. (1996). Handbook of antioxidants. Marcel Dekker, New York.
- Calvino, V., Picallo, M., Lopez-peinado, A. J., Martin-Aranda, R. M. and Duran Valle, C. (2006). Ultrasound accelerated Claisen–Schmidt condensation: A green route to chalcones. *Journal of Applied Surface Science*. 252 (17): 6071-6074.
- Cao, G., Sofic, E. and Prior, R. L. (1997). Antioxidant and prooxidant behavior of flavonoids: structure-activity relationships. *Free Radical Biology and Medicinal*. 22: 749–760.
- Chang, T. S. (2009). An updated review of tyrosinase inhibitors. *International Journal of Molecular Sciences*. 10: 2440-2475.
- Cheng, Z.J., Lin, C.H., Hwang, T.L. and Teng, C.M.(2001). Brousochalcone A, a potent antioxidant and effective suppressor of inducible nitric oxide synthase in lipopolysacchraide-activated macrophages. *Biochemistry and Pharmacology*. 61: 939–946.
- Climent, M. J., Corma, A., Iborra, S. and Velty, A. (2004). Activated Hydrotalcites as Catalysts for the Synthesis of Chalcones of Pharmaceutical Interest. *Journal of Catalyst*. 221 : 474-482.

- Cushnie, T. P. T. and Lamb, A. J. (2005). Antimicrobial activity of flavonoids. *International Journal of Antimicrobial Agents*. 26: 343–356.
- Denizot, F. and Lang, R. (1986). Rapid colorimetric assay for cell growth and survival. Modifications to the tetrazolium dye procedure giving improved sensitivity and reliability. *Journal of Immunology Methods*. 89(2): 271-277.
- Detsi, A., Majdalani, M., Christos, A. K., Dimitra, H. L. and Panagiotis, K. (2009). Natural and synthetic 2'-hydroxy-chalcones and aurones: Synthesis, characterization and evaluation of the antioxidant and soybean lipoxygenase inhibitory activity. *Bioorganic and Medicinal Chemistry*. 17: 8073–8085.
- Eddarir, S., Catelle, N., Bakkour, Y. and Ronlando, C. (2003). An efficient synthesis of chalcones based on the Suzuki reaction. *C. Tetrahedron Letter*. 44(28): 5359-5363.
- Edwards, M. L., Stemerick, D. M. and Sunkara, P. S. (1990). Chalcone: a new class of antimitotic agents. *Journal of Medicinal Chemistry*. 33: 1948-1954.
- Elzaawely, A. A., Xuan, T. D. and Tawata, S. (2007). Essential oils, kava pyrones and phenolic compounds from leaves and rhizomes of *Alpinia zerumbet* (Pers.) and their antioxidant activity. *Food Chemistry*. 103: 486–494.
- Fernandez, M. J. (2003). Apoptosis induced by different doses of caffeine in CHO cells. *Journal of Applied Toxicology*. 23(4): 221-224.
- Furman, C., Lebeau, J., Fruchart, J. C., Bernier, J. L., Duriez, P., Cotelle, N. and Teissier, E. (2001). Di-tert-butylhydroxylated flavonoids protect endothelial cells against oxidized LDL-induced cytotoxicity. *Journal of Biochemistry and Molecular Toxicology*. 15: 270–278.

- Geissman, T.A. and Crout, D. H. G. (1969). *Organic Chemistry of Secondary Plant Metabolism*. California: Freeman, Cooper and Company.
- Ghiselle, A., Serafini, M., Maiane, G., Azzini, E. and Ferro-Luzzi, A. (1995). A fluorescence-based method for measuring total plasma antioxidant capacity. *Free Radical Biology and Medicine*. 18: 29-36.
- Gohari, A. R., Hajimehdipoor, H., Saeidnia, S., Ajani, Y. and Hadjiakhoondi, A. (2010). Antioxidant Activity of some medicinal species using FRAP Assay. *Journal of Medicinal Plants*. 10 (37): 54-60.
- Guo, J. T., Lee, H. L., Chiang, S. H., Lin, F. I. and Chang, C. Y. (2001). Antioxidant properties of the extracts from different parts of broccoli in Taiwan. *Journal of Food Drug Analysis*. 9(2): 96-101.
- Habsah, M., Faridah, A., Permana, D., Lajis, N.H., Alib, A. M., Sukaric, M.A., Taufiq Y. Y. Hinc, Kikuzakid, H. and Nakatanid, N. (2004). DPPH free radical scavenger components from the fruits of *Alpinia rafflesiana* Wall. ex. Bak. (Zingiberaceae). *Zeitschrift fur Naturforschung C: Journal of Biosciences*. 59: 811-815.
- Hadi, A., Javid, S. M., Hadi, A. and Gholamreza, Z. (2010). Synthesis, in vitro antimicrobial and antioxidant activities of chalcone and flavone derivatives holding allylic substitutions. *Medicinal Chemistry Research*. 20: 1318-1324.
- Halliwell, B. (1996). Antioxidants in human health and disease. *Annual Review of Nutrition*. 16: 33-50.
- Halliwell, C. B. and Gutteridge, J. M. (2000). Free radicals and antioxidants in the year 2000 - A historical look to the future. *Annals of the New York Academy of Sciences*. 899: 136-147.

- Hsieh, H. K., Tsao, L. T., Wang, J. P. (2000). Synthesis and anti-inflammatory effect of chalcones. *Journal of Pharmacy and Pharmacology*. 52: 163–171.
- Hsu, Y.L., Kuo, P.L., Tzeng, W.S., Lin, C.C. (2006). Chalcone inhibits the proliferation of human breast cancer cell by blocking cell cycle progression and inducing apoptosis. *Food and Chemical Toxicology*. 44: 704–713.
- Hua, Z. S., Luo, J. G., Wang, X. B., Wang, J. S., Yi, K. L. (2009). Two novel monoterpene–chalcone conjugates isolated from the seeds of *Alpinia Katsumadai*. *Bioorganic and Medicinal Chemistry Letters*. 19: 2728–2730.
- Jayapal, M. R., Sreenivasa, P. K. and Sreedhar, N. Y. (2010). Synthesis and characterization of 2,6-Dihydroxy substituted chalcones using PEG-400 as a recyclable solvent. *Journal of Pharmaceutical Sciences*. 2(8):450-458.
- Jin, C., Yong, J., He, H. and Fu, L. (2011). Synthesis and antitumor activity of novel chalcone derivatives. *Biomedicine and Pharmacotherapy*. 3019: 1–3.
- Jing, D., Wangyuan, C. and Guangzhong, Y. (2011). Analytical Methods: A novel antioxidant activity index (AAU) for natural products using the DPPH assay. *Food Chemistry*. 125(4): 1430-1435.
- Khatib, S., Nerya, O., Musa, R., Shmuel, M., Tamir, S. and Jacob, V. (2005). Chalcones as potent tyrosinase inhibitors: the importance of a 2,4-substituted resorcinol moiety. *Bioorganic and Medicinal Chemistry*. 13: 433–441.
- Kim, T. H., Seo, W. D., Ryu, H. W., Seo, H. R., Jin, Y. B., Lee, M., Ji, Y. H., Ki, H. P. and Lee, Y. S. (2010). Anti-tumor effects by a synthetic chalcone compound is mediated by c-Myc-mediated reactive oxygen species production *Chemico-Biological Interactions*. 188: 111–118.

- Lai, C. H., Rao, Y. K., Fang, S. H., Sing, Y. T. and Tzeng, Y. M. (2010). Identification of 3',4',5'-trimethoxychalcone analogues as potent inhibitors of *Helicobacter pylori*-induced inflammation in human gastric epithelial cells. *Bioorganic and Medicinal Chemistry Letters*. 20: 5462–5465.
- Lee, Y. S., Lim, S. S., Shin, K. H., Kim, Y. S., Ohuchi, K. and Jung, S. H. (2009). Anti-angiogenic and Anti-tumor Activities of 2'-Hydroxy-4'-methoxychalcone. *Biological and Pharmaceutical Bulletin*. 29: 1028–1031.
- Lim, T. Y., Lin, Y. Y. and Yule, C. M. (2009) Evaluation of antioxidant, antibacterial and antityrosinase activities of four *Macaranga* species. *Food Chemistry*. 114: 594-599.
- Linn, Y. M., Zhou, Y, Flavin, M. T., Zhou, L. M., Nie, W. and Chen, F. C. (2002) Chalcones and flavonoids as anti-tuberculosis agents. *Bioorganic and Medicinal Chemistry*. 10: 2795–2802.
- Mandge, S., Singh, H. P., Dutta, G. S. and Hari, N. M. N. S. (2007). Synthesis and characterization of some chalcone derivatives. *Trends in Applied Sciences Research*. 1: 52-56.
- Mária, T., Klára, S., Ágnes, S. and Ilona, S. V. (2003). Examination on antioxidant activity in the greater celandine (*Chelidonium majus* L.) extracts by FRAP method. *Symposium Acta Biologica Szegediensis*. 47(1-4): 115-117.
- Mickisch, G., Fajta S., Keilhauer G., Schlick E., Tschada, R. and Alken P. (1990). Chemosensitivity testing of primary human renal cell carcinoma by a tetrazolium based microculture assay (MTT). *Journal of Urological Research*. 18(2): 131-6.
- Mohamad, H., Abas, F., Permana, D., Lajis, N. H., Ali, A. M., Sukari, M. A., Hin, T. T. Y., Kikuzaki, H. and Nakatani, N. (2004). DPPH free radical scavenger components from the fruits of *Alpinia rafflesiana* Wall. *Ex Bak*.

(Zingiberaceae). *Zeitschrift fur Naturforschung C: Journal of Biosciences*. 59: 811-815.

Molyneux, P. (2004). The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin Journal of Science and Technology*. 26(2): 211-219.

Nerya, O., Ramadan, M., Khatib, S., Tamir, S. and Vaya, J. (2004). Chalcones as potent tyrosinase inhibitors: the effect of hydroxyl positions and numbers. *Phytochemistry*. 65: 1389–1395.

Ozcelik, O., Lee, J. H. and Min, D. B. (2003). Effects of light, oxygen and pH on the absorbance of 2,2-diphenyl-1-picrylhydrazyl. *Journal of Food Science*. 68: 487–490.

Peng, C., Chen, S., Lin, F. and Lin, Z. (2000). Detection of antioxidative capacity in plants by scavenging organic free radical DPPH. *Progress in Biochemistry and Biophysics*. 27(6): 578–661.

Pietta P. G. (2000). Flavonoids as Antioxidants. *Journal of Natural Product*. 63: 1035-1042.

Polkowski, K., Skierski, J.S. and Mazurek, A.P. (2000). Anticancer activity of genistein-piperazine complex. *In vitro* study with HL-60 cells. *Reviews*. 57: 223–232.

Pulido, R., Bravo, L. and Saura-Calixto, F. (2000). Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural and Food Chemistry*. 48: 3396–3402.

- Rajendra, P. Y., Rao, L. A., Rambabu R. and Ravi Kumar P. (2007). Synthesis and biological evaluation of some novel chalcone derivatives. *Oriental Journal of Chemistry*. 23(3): 927-937.
- Rao, Y. K., Fang, S. H. and Tzeng, Y. M. (2009). Synthesis and biological evaluation of 3',4',5'-trimethoxychalcone analogues as inhibitors of nitric oxide production and tumor cell proliferation. *Bioorganic and Medicinal Chemistry*. 17: 7909–7914.
- Rao, Y. K., Kao, T.Y., Ko, J. L., Tzeng, Y. M. (2010). Chalcone HTMC causes *in vitro* selective cytotoxicity, cell-cycle G1 phase arrest through p53-dependent pathway in human lung adenocarcinoma A549 cells, and *in vivo* tumor growth suppression. *Bioorganic and Medicinal Chemistry Letters*. 20: 6508–6512.
- Rao, Y. K., Fang, S. H. and Tzeng, Y. M. (2004). Differential effects of synthesized 2'-oxygenated chalcone derivatives: modulation of human cell cycle phase distribution. *Bioorganic and Medicinal Chemistry*. 12: 2679-2686.
- Rattanachitthawat, S., Suwannalert, P., Riengrojpitak, S., Chaiyasut, C. and Pantuwatana, S. (2010). Phenolic content and antioxidant activities in red unpolished Thai rice prevents oxidative stress in rats. *Journal of Medicinal Plants Research*. 4 (9): 796-801.
- Renaud, S. C., Gueguen, R., Schenker, J. and d'Houtaud, A. (1998). Alcohol and mortality in middle-aged men from eastern France. *Epidemiology*. 9: 184–188.
- Rice Evans, C. A., Miller, N. J. and Paganga, G. (1996). Antioxidant Properties of Phenolic Compounds. *Trends in Plants. Science Reviews*. 2: 152-159.

- Rishton, G. M. (2008). Natural products as a robust source of new drugs and drug leads: past successes and present day issues. *American Journal of Cardiology*. 101(10A): 43-49.
- Rosa M. S., Juan, Mayo, C., Tan, D. X., Silvia, L. B., Mohan, N. and Russel J. (2003). Antioxidant activity of melatonin in Chinese hamster ovarian cells: changes in cellular proliferation and differentiation *Reiter Biochemical and Biophysical Research Communications*. 302: 625–634.
- Saewan, N., Koyomboon, S. and Chantrapromma, K. (2011). Antityrosinase and anticancer activities of flavanoids from *Blumea balsamifera* DC. *Journal of Medicinal Plants Research*. 5(6): 1018-1025.
- Satyanarayana, M., Tiwari, P., Tripathi, B. K., Srivastava, A. K. and Pratab, R. (2004). Synthesis and antihyperglycemic activity of chalcone based aryloxypropanolamines. *Bioorganic and Medicinal Chemistry*. 12: 883–889.
- Schwarz, K., Bertelsen, G., Nissen, L. R., Gardner, P. T., Heinonen, M. I., Hopia, A., Huynh-Ba, T., Lambelet, P., McPhail, D., Skibsted, L. H. and Tijburg, L. (2001). Investigation of plant extracts for the protection of processed foods against lipid oxidation. Comparison of antioxidant assays based on radical scavenging, lipid oxidation and analysis of the principal antioxidant compounds. *European Food Research Technology*. 212: 319-328.
- Seo, S. Y., Sharma, V. K., Sharma, N. (2003). Mushroom tyrosinase: recent prospects. *Journal of Agricultural and Food Chemistry*. 51: 2837–2853.
- Shohaib, T., Shafique, M., Dhanya, N. and Madhu, C. D. (2011) Importance of Flavonoides in therapeutics. *Hygeia: Journal for Drugs and Medicines*. 3: 1-18.

- Smit, N., Vicanova, J. and Pavel, S., (2009). The hunt for natural skin whitening agents. *International Journal of Molecular Sciences*. 10: 5326–5349.
- Soloneski S. (2002). Effect of dithiocarbamate pesticide zineb and its commercial formulation, azzuro. III. Genotoxic evaluation of CHO cells. *Mutational Research*. 514 (1-2): 201-212.
- Srinivas, K.V.N.S., Koteswara, Y., Rao, I., Mahender, Biswanath, D., Rama Krishna K.V.S., Hara Kishore, K. and Murty, U. S. N. (2003). Flavanoids from *Caesalpinia pulcherrima*. *Phytochemistry*. 63: 789–793.
- Stampfer, M. J., and Rimm, E. B. (1995). Epidemiologic evidence for vitamin E in prevention of cardiovascular disease. *American Journal of Clinical Nutrition*. 62: 1365S-1369S.
- Szollosi, R. and, Varga, I. S. (2002). Total antioxidant power in some species of Labiatae (Adaptation of FRAP method). *Acta Biologica Szeged*. 46 (3 - 4): 125-127.
- Temple, N. J. (2000). Antioxidants and disease: more questions than answers. *Nutrition Research*. 20: 449–459.
- Tepe, B., Sokmen, M., Akpulat, H. A. and Sokmen, A. (2005). In vitro antioxidant activities of the methanol extracts of four *Helichrysum* species from Turkey. *Food Chemistry*. 90: 685–689.
- Vinson, J. A., Dabbagh, Y. A., Serry, M. M. and Jang, J. (1995). Plant flavonoids, especially tea flavonols, are powerful antioxidants using an in vitro oxidation model for heart disease. *Journal of Agricultural and Food Chemistry*. 43: 2800–2802.

- Wang, F., Ma, R. and Yu, L. (2006). Role of mitochondria and mitochondrial cytochrome c in tubeimoside I-mediated apoptosis of human cervical carcinoma HeLa cell line. *Cancer Chemotherapy and Pharmacology*. 57: 389–399.
- Wangyuan, C., Guangzhong, Y. and Jing, D. (2011) A novel antioxidant activity index (AAU) for natural products using the DPPH assay. *Food Chemistry*. 125: 1430–1435.
- Ward, F. E., Garling, D. L., Buckler, R. T., Lawler, D. M. and Cummings, D. P. (1981). Antimicrobial 3-methylene flavanones. *Journal of Medicinal Chemistry*. 24: 1073–1077.
- Whitaker, J.R. (1995). Polyphenol oxidase. In: Wong, D.W.S. (Ed.). *Food Enzymes, Structure and Mechanism*. New York: Chapman and Hall.
- William, R. S. (2000). The role of natural products in a modern drug discovery program. *Drug Discovery Today*. 5: 1-3.
- Won, S. J., Liu, C. T., Tsao, L. T., Weng, J. R., Ko, H. H., Wang, J. P. and Lin, C. N. (2005). Synthetic chalcones as potential anti-inflammatory and cancer chemopreventive agents. *European Journal of Medicinal Chemistry*. 40(1): 103-112.
- Xia, Y., Yang, Z. Y., Xia, P., Bastow, K. I, Nakanishi, Y. and Lee, K. H. (2000) Antitumor agents. Part 202: novel 20-amino chalcones: design, synthesis and biological evaluation. *Bioorganic and Medicinal Chemistry Letters*. 10: 699–701.