STUDIES ON THE EFFECT OF DIFFERENT BIOPROCESS PARAMETERS ON PECTINASE PRODUCTION BY Aspergillus niger

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To my beloved mother and father

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In the name of Allah, the Beneficent, the Merciful.

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ABSTRACT

Pectinase is heterogeneous group of enzymes that have a polysaccharides substrate and breaks down the pectin component, which found in the plant cell wall, into simple sugar and galacturonic acid. The breaks down of the pectin will cause the plant tissues to undergo some modification on its cell wall, or other activities such as maceration or cell lysis. The pectinases or usually known as pectinolytic enzymes were extensively used in food industries that involve degradation of plant materials to fasten the fruit juice extraction process. Among different biofactories of pectinases, filamentous fungi such as Aspergillus niger were the best known for the production and secretion of pectinase. Therefore, the objective of this research was to develop industrial culture medium and a cultivation strategy for the production and secretion of pectinases in a semi-industrial scale by A. niger. In this study, the effect of medium composition on the production and secretion of pectinase was studied through the classical method where the medium screenings were done initially to find the best medium for the production of pectinase. The optimized medium through classical method were pectin industrial (30 g L^{-1}), ammonium sulphate (3.33 g L^{-1}), di-potassium hydrogen phosphate (1 g L^{-1}), magnesium sulfate heptahydrate (0.05 g L^{-1}), potassium chloride (0.05 g L^{-1}) and iron sulphate heptahydrate (0.1 g L^{-1}). Following this step, medium optimization was carried out using statistical approach and the optimized medium were pectin industrial (32.22 g L⁻¹), ammonium sulphate (4.33 g L^{-1}) , di-potassium hydrogen phosphate (1.36 g L⁻¹), magnesium sulphate heptahydrate (0.05 g L^{-1}), potassium chloride (0.05 g L^{-1}) and iron sulphate heptahydrate (0.1 g L^{-1}). Therefore, the result of pectinase production for optimized medium was 64.83 % higher compared to unoptimized medium. Next, the effect of processing parameters which was pH condition (controlled pH at 5.5 and uncontrolled pH) was studied in the batch fermentation. The pectinase production of controlled pH was 51.35 % higher compared to the uncontrolled pH. However, the selection of the best condition for fed-batch fermentation was uncontrolled pH due to the low costing during the cultivation. Finally, the fed-batch strategies for full media and monocorbon were studied and the best feeding strategies was from the monocarbon feeding due to the higher prectinase yield with 429.95 U mL⁻¹ compared to the full media feeding with 138.27 U mL⁻¹. Thus, these all together lead to the development of industrial process for pectinase production in semi-industrial scale.

ABSTRAK

Pektinase adalah kumpulan heterogen enzim yang mempunyai substrat polisakarida dan bertanggungjawab untuk memecahkan komponen pektin, yang ditemui di dinding sel tumbuhan, ke dalam gula ringkas dan asid galacturonik. Pemecahan komponen pektin akan menyebabkan tisu tumbuhan menjalani beberapa pengubahsuaian pada dinding sel, atau aktiviti-aktiviti lain seperti kehabisan tenaga atau sel lysis. Enzim pektinase atau biasanya dikenali sebagai enzim pektinolitik telah digunakan dengan meluas dalam industri makanan yang melibatkan degradasi bahan tumbuhan untuk mengikat proses pengekstrakan jus buah-buahan. Antara pengilangan berbeza pektinase, kulat berfilamen seperti Aspergillus niger adalah yang terbaik dikenali untuk pengeluaran dan rembesan pektinase. Oleh itu, objektif kajian ini adalah untuk membangunkan penghidupan media didalam industri dan strategi menghidupkan Aspergillus niger untuk pengeluaran dan rembesan pektinase dalam skala semi- industri oleh A. niger. Dalam kajian ini, kesan komposisi medium kepada pengeluaran dan rembesan pektinase akan dikaji menggunakan kaedah klasik di mana pemilihan media telah dilakukan pada mulanya untuk mencari medium terbaik untuk pengeluaran pektinase. Medium yang optimum adalah melalui kaedah klasik terdiri daripada pektin perindustrian (30 g L^{-1}), ammonium sulfat (3.33 g L^{-1}), di-kalium hidrogen fosfat (1g L⁻¹), magnesium sulfat heptahydrate (0.05 g L⁻¹), kalium klorida (0.05 g L^{-1}) dan besi sulfat heptahydrate (0.1 g L^{-1}). Berikutan langkah ini, pengoptimuman medium telah dijalankan dengan menggunakan pendekatan statistik dan medium yang telah dioptimumkan terdiri daripada pektin industri (32.22 g L⁻¹), ammonium sulfat (4.33 g L⁻¹), di-kalium hidrogen fosfat (1.36 g L⁻¹), magnesium sulfat heptahydrate (0.05 g L⁻¹), kalium klorida (0.05 g L⁻¹) dan magnesium sulfat heptahydrate (0.1 g L^{-1}). Oleh itu, hasil daripada pengeluaran pektinase daripada medium yang telah dioptimumkan adalah 64.83% lebih tinggi berbanding dengan media dioptimumkan melalui kaedah klasik. Seterusnya, kesan parameter pemprosesan seperti pH (pH dikawal pada 5.5 dan tidak terkawal) telah dikaji didalam fermentasi sesekelompok. Pengeluaran pektinase yang dikawal oleh pH adalah 51.35% lebih tinggi berbanding dengan pH yang tidak terkawal. Walau bagaimanapun, pemilihan keadaan yang terbaik untuk suapan sesekelompok fermentasi adalah pH yang tidak terkawal kerana kerana pengekosan yang rendah semasa penanaman. Akhir sekali, strategi makan-kumpulan untuk media penuh dan monokarbon dikaji dan strategi pemberian makanan terbaik adalah daripada makanan monokarbon yang disebabkan oleh hasil pektinase yang tinggi dengan 429.95 U mL⁻¹ berbanding dengan media penuh makan dengan 138,27 U mL⁻¹. Oleh itu, ini semua bersama-sama membawa kepada pembangunan proses industri untuk pengeluaran pectinase dalam skala semi-industri.

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LIST OF ABBREVIATIONS

A. niger	-	Aspergillus niger
$Y_{P/x}$	-	Yield coefficient
$U_{enzyme} g_{cells}^{-1}$	-	Yield coefficient unit
g L ⁻¹	-	Cell biomass unit
$U mL^{-1}$	-	Pectinase unit
μ	-	Specific growth rate
di	-	Impeller diameter
dt	-	Tank diameter
CDW	-	Cell dry weight
$FeSO_4 \cdot 7H_2O$	-	Ferrous sulphate heptahydrate
HCl	-	Hydrochloric acid
$H_2PO_4^-$	-	Orthophosphate
K_2HPO_4	-	Di-potassium hydrogen phosphate
KNO ₃	-	Potassium nitrate
KCl	-	Potassium chloride
MgSO4.7H2O	-	Magnesium sulphate heptahydrate
NaCl	-	Sodium chloride
NaNO ₃	-	Sodium nitrate
$(NH_4)_2SO_4$	-	Ammonium sulphate

CHAPTER 1

INTRODUCTION

1.1 Introduction of Research

Nowadays, the pectinolytic enzyme by filamentous fungi from industrial food waste like orange peels was used in the food industry for juice and wine production and is extensively done in order to utilize the abundant waste and commercialize in industries (Mantovani *et al.*, 2005). As a result, pectinolytic enzyme becomes one of the upcoming enzymes of the commercial sector (Kashyap *et al.*, 2001).

Since 1968 until now, many studies done on the pectinolytic enzyme or pectinase enzyme for commercial and widely used in industrial processing either in fruit or vegetables (Solis-Pereyra *et al.*, 1993). Whereas, filamentous fungus is used for commercial enzyme production in food and beverage industries as genera of *Aspergillus* is granted as GRAS (generally regarded as safe) (Iwashita, 2002). Therefore, many studies are focusing on the increasing of the yield of production and secretion of pectinase from *Aspergillus niger* especially in bioprocessing parameter indirectly will help the industrial sector in establishing its productivity.

However, there are other application of pectinase enzyme instead of used in the food industry which are basically used as retting and degumming of fiber crops. Retting is a fermentation process where certain bacteria and fungi decompose the pectin of the bark and release the fiber. The treatment of pectic wastewater also used pectinase enzyme in their process in order to remove pectin substances from wastewater. Next, it is also used in paper making as it can depolymerise polymers of galacturonic acid and will lower the cationic demand of pectin solutions and will filtrate it from peroxide bleaching. Furthermore, oil extraction is basically used pectinase enzyme to extract the oil in aqueous process by liquefying the structural cell wall component. Whereas, in tea fermentation; it helps to improve the foam forming property of instant tea powders by destroying tea pectin (Kashyap *et al.*, 2001).

The advantages of the use of pectinase in the beverage industries include allowing the producer to diversify the type of product in term of its cloudy, clearer juice and concentrates. The enzymes also help to produce the juices and concentrate in a very stable and have a good taste. Interestingly, pectinase can help in reducing production cost in term of higher yield, less equipment and labour especially in a concentration process (Kashyap *et al.*, 2001).

1.2 Problem Statement

The optimization of production medium for pectinase production towards low cost and suitable media composition for industrial purpose was crucially important in order to meet the increasing demand of this enzyme. Therefore, in this study; a filamentous fungus of *Aspergillus niger* was used for the production and secretion of pectinase in a semi industrial scale. In order to optimize the production and secretion of pectinases by *A. niger*, the different bioprocess parameters were studied. The parameters were including the cultivation media at shake flasks level and bioprocessing condition (pH condition) in stirred tank 16-L bioreactor. Furthermore, the limitations studies on feeding strategist with different feeding solution in order to increase the yields of pectinase production in the semi-industrial scale 16-L bioreactor application. Hence, these all together will lead to the development of industrial process for pectinase production in semi-industrial scale.

1.3 Research Objective

The objective of the research was to develop industrial culture medium and a cultivation strategy for the production of pectinase in semi-industrial scale by *A. niger.*

1.4 Research Scope

To accomplish the objective, there are five research scopes were applied:

- Medium screening and optimization for shake flask cultivation using classical method.
- 2) Medium optimization for shake flask cultivation using statistical method.
- Comparison between classical media optimization method and statistical medium optimization method.
- 4) Batch cultivation of *A. niger* in a stirred tank 16-L bioreactor for high production of pectinase.
- 5) Fed-batch cultivation of *A. niger* in a stirred tank 16-L bioreactor for high production of pectinase.

REFERENCES

- Abubakar, A., Suberu, H. A., Bello, I. M., Abdulkadir, R., Daudu, O. A., Lateef, A.
 A. (2013). Effect of pH on Mycelial Growth and Sporulation of Aspergillus Parasiticus. Journal of Plant Science. 1(4):64-67.
- Ahamed, A., and Vermette, P. (2008). Enhanced Enzyme Production from Mixed Cultures of *Trichoderma Reesei* RUT-C30 and *Aspergillus niger* LMA Grown As Fed Batch in Stirred Tank Bioreactor. *Biochemical Engineering Journal*. 42: 41-46.
- Aguilar, G., and Huitrón, C. (1990). Constitutive Exo-Pectinase Produced By Aspergillus sp. CH-Y-1043 on Different Carbon Source. Biotechnology Letters. 12(9): 655-661.1
- Arijit, D., Sourav, B., Naimisha V, R., and Rajan S, S. (2013). Improved Production and Purification of Pectinase from *Streptomyces* sp. GHBA10 Isolated from Valapattanam Mangrove Habitat, Kerala, India. *International Research Journal of Biological Sciences*. 2(3):16-22.
- Askolin, S., Penttilä, M., Wösten, H. A B., and Nakari-Setälä. (2005). The *Trichoderma reesei* Hydrophobin Genes *hfb1* and *hfb2* Have Diverse Functions in Fungal Development. *FEMS Microbiology letters*. 253:281-288.
- Awofolu, O. R., Okonkwo, J. O., Roux-Van Der Merwe, R., Badenhorst, J., and Jordaan, E. (2006). A New Approach to Chemical Modification Protocols of *Aspergillus niger* and Sorption of Lead Ion By Fungal Species. *Electronic Journal* of Biotechnology. 9(4): 1-10.
- Chaudhri, A., and Suneetha, V. (2012). Microbially Derived Pectinases: A Review. *IOSR Journal of Pharmacy and Biological Sciences*. 2(2):1-5.
- Czitrom, V. (1999). One–Factor-At-A-Time versus Designed Experiments. *The American Statistician*. 53(2).

- Criddle, C. S. (1993). The Kinetics of Cometabolism. *Biotechnology and Bioengineering*. 41:1048-1056.
- Debing, J., Peijun, L., Stagnitti, F., Xianzhe, X., and Li, L. (2006). Pectinase Production by Solid Fermentation from Aspergillus niger by a New Prescription Experiment. Ecotoxicology and Environment Safety. 64:244-250.
- Davis, D. J., Burlak, C., Money, N. P. (2000). Osmotic Pressure of Fungal Compatible Osmolytes. *Mycology Series*. 104(7):800-804.
- Díaz, A. B., Caro, I., Ory, I. D., and Blandino, A. (2007). Evaluation of the Condition for the Extraction of Hydrolytic Enzymes Obtained by Solid State Fermentation from Grape Pomace. *Enzyme and Microbial Technology*. 41:302-306.
- Dogan, N (2008). Production of Pectinase Enzyme From Aspergillus Sojae In Batch and Fed-Batch System. Master of Science. İzmir Institute of Technology.
- Dunn, I. J., Heinzle, E., Ingham, J., and Přenosil, J. E. (2000). Biological ReactionEngineering. Dynamic Modelling Fundamentals with Simulation Examples.Second, completely revised edition. Germany: Wiley-VCH GmbH & Co.KGaA.
- Esawy, M. A., Gamal, A. A., Kamel, Z., Ismail, A. S., and Abdel-Fattah, A. F. (2013). Evaluation of Free and Immobolized Aspergillus niger NRC1ami Pectinase Applicable in Industrial Processes. *Carbohydrate Polymers*. 92: 1463-1469.
- El enshasy, H. A., Kleine, J., Rinas, U. (2006). Agitation Effects on Morphology and Protein Productive Fractions of Filamentous and Palleted Growth Forms of Recombinant Aspergillus niger. Process Biochemistry. 41:2103-2112.
- El enshasy, H. A. (2007). Chapter 9, Filamentous Fungal cultures-Process Characteristics, Products, and Applications. Bioprocessing. *Bioprocessing For Value-Added Products from Renewable Resources*. Elsevier Science and Technology Books.
- Elshafei, A. M., and Abdel-Fatah, O. M. (2001). Evidence for a Non-Phosphorylated Route of Galactose Breakdown in Cell-Free Extracts of *Aspergillus niger*. Enzyme and Microbial Technology. 29:76-83.
- Favela-Torres, E., Cordova-López, J., García-Rivro, M., and Gutiérrez-Rojas, M. (1997). Kinetics of Growth of Aspergillus niger during Submerged, Agar Surface and Solid State Fermentations. Process Biochemistry. 33(2): 103-107.

- Fawole, O. B., and Odunfa, S. A.(2003). Some Factors Affecting Production of Pectic Enzymes by Aspergillus niger. International Biodeterioration & Biodegradation. 52:223-227.
- Friedrich, J., Cimerman, A., and Steiner, W. (1989). Submerged Production of Pectinolystic Enzymes By Aspergillus niger: Effect of Different Aeration/Agitation Regimes. Applied Microbiology and Biotechnology. 31:490-494.
- Gerlach, S. R., Siendenberg, D., Gerlach, D., Schügerl, K., Giuseppin, M. L. F., and Hunik, J. (1998). Influence of Reactor Systems on the Morphology of Aspergillus awamori. Application of Neural Network and Cluster Analysis for Characterization of Fungal Morphology. Process Biochemistry. 33(6):601-615.
- Gummadi, S. N., and Kumar, D. S. (2008). Batch and Fed Batch Production of Pectin Lyase and Pectate Lyase by Novel Strain *Debaryomyces nepalensis* in Bioreactor. *Bioresource Technology*. 99:874-881.
- Griffith, G. W., Easton, G. L., Detheridge, A., Roderick, K., Edwards, A., Worgan, H. J., Nicholson, J., and Perkins, W. T. (2007). Copper deficiency in potato dextrose agar causes reduced pigmentation in cultures of various fungi. *FEMS Microbiology letters*. 276: 165-171.
- Grimm, L. H., Kelly, S., Krull, R., and Hempel, D. C. (2005). Morphology and Productivity of Filamentous Fungi. *Applied Microbial Biotechnology*. 69:375-384.
- Gyamerah, M., Merichetti, G., Adedayo, O., Scharer, J. M., and Moo-Young, M. (2002). Bioprocessing Strategies for Improving Hen Egg-White Lysozyme (HEWL) Production by Recombinant *Aspergillus niger* HEWL WT-13-16. *Applied Microbiology Biotechnology*. 60:403-407.
- Harvey, L. M., and Mcneil, B. (1994). Liquid Fermentation Systems and Product Recovery of Aspergillus. Biotechnology Handbook. 7:141-176.
- Heerd, D., Yegin, S., Tari, C., and Fernandez-Lahore, M. (2012). Pectinase Enzyme-Complex Production by Aspergillus spp. in Solid-State Fermentation: A Comparative Study. Food and Bioproduct Processing. 90:102-110.
- Hibbert, D., B. (2012). Experimental Design in Chromatography: A Tutorial Review. *Journal of Chromatography B*. 910:2-13.
- Hogema, B. M., Arents, J. C., Inada, T., Aiba, H., Dam, K. V., and Postma, P. W. (1997). Catabolite Repression by Glucose 6-Phosphate, Gluconate and Lactose in Escherichia Coli. *Molecular Microbiology*. 24(4):857-867.

- Ismail, A. M. S. (1996). Utilization of Orange Peels for the Production of Multienzyme Complex by Some Fungal Strains. *Process biochemistry*. 31(7): 645-650.
- Iwashita, K. (2002). Review: Recent Studies of Protein Secretion by Filamentous Fungi. Journal of Bioscience and Bioengineering. 94(6):530-535.
- Jonathan, S. G., and Fasidi, I. O. (2001). Effect of Carbon, Nitrogen and Mineral Sources on Growth of *Psathyerella Atroumbonata* (Pegler), a Nigerian Edible Mushroom. Food Chemistry. 72:479-483.
- Kashyap, D. R., Vohra, P. K., Chopra, S., and Tewari, R. (2001). Application of Pectinases in the Commercial Sector: A Review. *Bioresources Technology*. 77: 215-227.
- Katoh, S., and Yoshida, F. (2009). Biochemical Engineering. Federal republic of Germany: WILEY-VCH Verlag GmbH & Co. KGaA
- Kavanagh, K. (2005). Fungi Biology and Applications. England: John Wiley & Sons, Ltd.
- Kortz, D. J., Rinas, U., Hellmuth, K., Sanders, E. A., and Deckwer, W. D.(1995). Simple Fed-Batch Technique for High Cell Density Cultivation of *Escherichia coli*. *Journal of Biotechnology*. 39:59-65.
- Kovarova-kovar, K., and Egli, T. (1998). Growth Kinetics of Suspended Microbial
 Cells: From Single-Substrate-Controlled Growth to Mixed-Substrate Kinetics.
 Microbiology and Molecular Biology Review. 62(3): 646-666.
- Levenspiel, O. (1999). Chemical Reaction Engineering. Third edition . United States of America: John Wiley & Sons, Inc.
- Lim, H. C., and Shin, S. H. (2013). Fed-batch Cultures. Principles and Applications of Semi-Batch Bioreactor. United States of America: Cambridge University Press.
- Mantovani, C. F., Geimba, M. P., and Brandelli, A. (2005). Enzymatic Clarification of Fruit Juices by Fungal Pectin Lyase. *Food Biotechnology*. 19:173-181.
- Maheshwari, R. (2005). Fungi: Experimental Models in Biology. (Mycology Series;V.24). United States of America: Taylor & Francis Group.
- Melzer, G., Dalpiaz, A., Grote, A., Kucklick, M., Göcke, Y., Jonas, R., Dersch, P., Franco-Lara, E., Nörtemann, B., and Hempel, D.C. (2007). Metabolix Flux Analysis using Stoichiometric Models for *Aspergillus Niger*: Comparison under Glucoamylase-Producing and Non-Producing Conditions. *Journal of Biotechnology*. 132:405-417.

- Mitard, A., and Riba, J. P. (1988). Morphology and Growth of Aspergillus niger ATCC 26036 Cultivated at Several Shear Rates. *Biotechnology and Bioengineering*. 32:835-840.
- Mojsov, K. (2010) (a). The Effect of Different Carbon Sources on Biosynthesis of Pectinolytic Enzymes by Aspergillus niger. Applied Technologies and Innovations. 3(3): 23-29.
- Mojsov, K. (2010) (b). The Effect of Inorganic Salts on Biosynthesis of Pectinolytic Enzymes by Aspergillus niger. Prospective of Innovations, Economics & Business. 4(1): 109-112.
- Nair, S. R., and Panda, T. (1997). Statistical Optimization of Medium Components for Improved Synthesis of Pectinase by Aspergillus niger. Bioprocess engineering.16: 169-173.
- Nevalainen, K. M. H., Te'o, V. S.J., and Bergquist, P. L. (2005). Heterologous Protein Expression in Filamentous Fungi. *TRENDS in Biotechnology*. 23(9): 468-474.
- Ohta, K., Hamada, S., and Nakamura, T. (1993). Production of High Concentrations of Ethanol from Inulin by Simultaneous Saccharification and Fermentation Using *Aspergillus niger* and *Saccharomyces cerevisiae*. *Applied and Environmental Microbiology*. 59(3):729-733.
- Okabe, M., Lies, D., Kanamasa, S., and Park, E. Y.(2009). Biotechnological Production of Itaconic Acid and Its Biosynthesis in Aspergillus terreus. Applied Microbiology Biotechnology. 84:597-606.
- Palaniyappan, M., Vijayagopal, V., Viswanathan, R., and Viruthagiri, T. (2009). Screening of Natural Substrates and Optimization of Operating Variables on the Production of Pectinase by Submerged Fermentation Using Aspergillus niger MTCC 281. African Journal of Biotechnology. 8(4):682-686.
- Palmqvist, E., and Hägerdal, B. H. (2000). Fermentation of Lignocellulosic Hydrolysates.I: Inhabition and Detoxification. *Bioresource Technology*. 74: 17-24.
- Papagianni, M. (2004). Review: Fungal Morphology and Metabolite Production in Submerged Mycelia Processes. *Biotechnology Advances*. 22:189-259.
- Park, E. Y., Koike, Y., Higashiyama, K., Fujikawa, S., and Okabe, M. (1999). Effect of Nitrogen Source on Mycelial Morphology and Arachidonic Acid Production in

Culture of *Mortierella Alpine*. Journal of Bioscience and Bioengineering. 88(1):61-67.

- Pedrolli, D. B., Monteiro, A. C., Gomes, E., and Carmona, E. C. (2009). Pectin and Pectinases: Production, Characterization and Industrial Application of Microbial Pectinolytic Enzymes. *The Open Biotechnology Journal*. 3:9-18.
- Pel, H. J., Winde, J. H. D., Archer, D. B., and Dyer, P. S. (2007). Genome Sequencing and Analysis of the Versatile Cell Factory Aspergillus niger CBS 513.88. Nature Biotechnology. 25(2):221-232.
- Pereira,J.G., Vieira de Queiroz,M., Gomes,E.A., Muro-Abad,J.I and Fernandes de Araújo,E. (2002). Molecular Characterization and Evaluation of Pectinase and Cellulase Production of *Penicilluim* spp. *Biotechnology Letters*. 24: 831-838.
- Punt, P. J., Biezen, N. V., Conesa .A., Albers, A., Mangnus, J., and Hondel, C. V. D. (2002). Filamentous Fungi as Cell Factories for Heterologous Protein Production. *TRENDS in Biotechnology*. 20(5) :200-2007.
- Puri, M., Banerjee, A., and Banerjee, U. C. (2005). Optimization of Process Parameters for the Production of Naringinase by *Aspergillus niger* MTCC 1344. *Process Biochemistry*. 40:195-201.
- Rao, D. G. (2010). Introduction to Biochemical Engineering. Second Edition. New Delhi: Tata McGraw Hill Education Private Limited.
- Rehman, H. U., Aman, A., Silipo, A., Qader, S. A. U., Molinaro, A., and Ansari, A. (2013). Degradation of Complex Carbohydrate: Immobilization of Pectinase from Bacillus Licheniformis KIBGE-IB21 Using Calcium Alginate as a Support. *Food Chemistry*. 139:1081-1086.
- Ridley, B. L., O'Neill, M. A., and Mohnen, D. (2001). Pectin: Structure, Biosynthesis, and Oligoglacturonide-Related Signalling. *Phytochemistry*. 57:929-967.
- Rinas, U., El- Enshasy, H., Emmler, M., Hille, A., Hempel,D. C., and Horn, H. (2005). Model –Based Prediction of Substrate Conversion and Protein Synthesis and Excretion in Recombinant *Aspergillus niger* Biopellets. Chemical Engineering Science.60:2729-2739.
- Salo, V., Niini, S. S., Virtanen, I., and Raudaskoski, M. (1989). Comparative immunocytochemistry of the cytoskeleton in filamentous fungi with dikaryotic and multinucleate hypae. *Journal of Cell Science*. 94: 11-24.

- Sharma, D. C., and Satyanarayana, T. (2006). A Marked Enhancement in The Production of Highly Alkaline and Thermostable Pectinase By *Bacillus pumilus* dcsr1 in Submerged Fermentation by Using Statistical Methods. *Bioresourece Technology*. 97:727-733.
- Sharma, N., Tripathi, A., (2008). Effect of *Citrus Sinensis* (L.) Osbeck Epicarp Essiential Oil on Growth and Morphobenensis of *Aspergillus niger* (L.) Van Tieghem. *Microbiological Research*. 163:337-344.
- Solis–Pereyra, S., Favela-Torres, E., Viniegra-Gonzalez, G., and Gutierrez-Rojas, M. (1993). Effect of Different Carbon Sources on the Synthesis of Pectinases by *Aspergillus niger* in Submerged and Solid State Fermentations. *Applied Microbiology and Biotechnology*. 39:36-41.
- Sphor, A., Carlsen, M., Nielsen, J., and Villadsen, J. (1998). α-Amylase Production in Recombinant Aspergillus oryzae During Fed-Batch and Continuous Cultivations. Journal of Fermentation and Bioengineering. 86(1):49-56.
- Suresh, B., and Viruthagiri, T. (2010). Optimization and Kinetics of Pectinase Enzyme Using Aspergillus niger by Solid-State Fermentation. Indian Journal of Science and Technology. 3(8):867-870.
- Tari, C., Gögus, N., and Tokatli, F. (2007). Optimization of Biomass, Pallet Size and Polygalacturonase Production by Aspergillus Sojae ATCC 20235 Using Response Surface Methodology. *Enzyme and Microbial Technology*. 40:1108-1116.
- Trejo-Aguilar, B. A., Visser, J., and Aguilar, G. (1996). Pectinase Secretion by Aspergillus FP-180 and Aspergillus niger N-402 Growing Under Stress Induced by the pH of Culture Medium. Pectins and Pectinases : 915-920.
- Torres, E. F., López, J. C., Rivero, M. G., and Rojas, M. G.(1997). Kinetics of Growth of Aspergillus niger during Submerged, Agar Surface And Solid State Fermentation. Process Biochemistry. 3(2): 103-107.
- Vahidi, H., Mojab, F., and Taghavi, N. (2006). Effect of Carbon Sources on Growth and Production of Antifungal Agents by *Gymnopilus Spectabilis*. *Iranian Journal* of Pharmaceutical Research. 3:219-222.
- Vasanthi, and Meenakshisundaram, M. (2012). Optimization of Pectinase Enzyme Production by Using Sour Orange Peel as Substrate in Solid State Fermentation. *Asian Journal of Biochemical and Pharmaceutical Research*. 1(2):16-26

- Vorwerk, S., Somerville, S., and Somerville, C. (2004). The Role of Plant Cell Wall Polysaccharide Composition in Disease Resistance. *TRENDS in Plant Science* . 9(4):203-209.
- Wang, J., and Wan, W. (2008). Factors Influencing Fermentative Hydrogen Production: A Review. *International Journal of Hydrogen Energy*. 1-13.
- Wang, J., and Wan, W. (2009). Experimental Design Methods for Fermentative Hydrogen Production: A Review. *International Journal of Hydrogen Energy*. 34:235-244.
- Wang, L., Ridgway, D., Gu, T., and Moo-Young, M. (2005). Bioprocessing Strategies to Improve Heterologous Protein Production in Filamentous Fungal Fermentations. *Biotechnology Advances*. 23:115-129.
- Wang, L., Ridgway, D., Gu, T., and Moo-Young, M. (2009). Kinetic Modeling of Cell Growth and Product Formation in Submerged Culture of Recombinant Aspergillus niger. Chemical Engineering Communications. 196:481-490
- Weihong, Z., and Peilin, C. (2005). Pectinase Production by Aspergillus niger P-6021 on Citrus changshan – huyou peel in slurry – state fermentation. Chinese Journal Chemical Engineering. 13 (4):510-515.
- Wucherpfenning, T., Hestler, T., and Krull, R. (2011). Morphology Engineering-Osmolality and Its Effect on Aspergillus niger Morphology and Productivity. Microbial Cell Factories. 10 (58):1-15.
- Wucherfenning, T., Lakowitz, A., and Krull, R. (2013). Comprehension of Viscous Morphology-Evaluation of Fractal and Conventional Parameters for Rheological Characterization of Aspergillus niger Culture Broth. Journal of Biotechnology. 163:124-132.
- Wainwright, M. (1992). An Introduction to Fungal Biotechnology. University of Sheffield,UK. John Wiley & Son.
- Yadav, S., Yadav, P. K., Yadav, D., and Yadav, K. D. S. (2009). Pectin Lyase: A Review. *Process Biochemistry*. 44:1-10.
- Yugandhar, N. M., Kumar, D. V. R. R., Prasanthi, V., Kumar, N. K., and Reddy, D. S. R. (2008). Optimization of Pectinase Production from *Manihot utilissima* by *Aspergillus niger* NCIM 548 Using Statistical Experimental Design. *Research Journal of Microbiology*. 3(1):9-16.