

PREPARATION, CHARACTERIZATION AND BIOCOMPATIBILITY EVALUATION
OF POLY(DIOL CITRATE) BLEND POLYETHERSULFONE HEMODIALYSIS
MEMBRANES

MUHAMAD ZULHILMI BIN ZAILANI

UNIVERSITI TEKNOLOGI MALAYSIA

PREPARATION, CHARACTERIZATION AND BIOCOMPATIBILITY EVALUATION
OF POLY(DIOL CITRATE) BLEND POLYETHERSULFONE HEMODIALYSIS
MEMBRANES

MUHAMAD ZULHILMI BIN ZAILANI

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Master of Philosophy

School of Graduate Studies
Universiti Teknologi Malaysia

JANUARY 2017

To my beloved mother and father

ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and the Most Merciful.

Alhamdulillah, Thank you Allah for giving me this opportunity, the strength and the patience to complete my dissertation, despite all the challenges and difficulties. I would like to express my utmost appreciation to my supervisor Prof Dr. Ahmad Fauzi Bin Ismail for his constructive advices, support and time in the completion of this thesis. Not forgotten, my co-supervisor Dr. Siti Hamimah Binti Sheikh Abdul Kadir for her support and knowledge pertaining my area of study. Their guidance had motivated me to push forward and guided me to the right paths.

I also would like to thanks my friends, all staffs and members of Advanced Membrane Technology Research Centre (AMTEC) for their encouragement and helps during my time of needs. Not forget on staff in Laboratories Management Unit (UPMU) UTM and faculty of science who had help me throughout my experimental works.

Special thanks and deepest gratitude go to my beloved parents;, my dad Zailani Bin Mohd Said and my mother Samini Binti Rahmat and my sisters for their endless love, prayers and encouragement. Also not forgetting my wife, Norazurin Binti Amir for her support and care. To those who indirectly contributed in this research, your kindness means a lot to me. Thank you very much.

Also not to be left out, the Ministry of Education (MOE) of Malaysia, for the MyBrain15 (MyMaster) sponsorship and Research University Grant (Q.J130000.2409.01G46). Thank you for the generous financial support which made my pursuit of study possible.

ABSTRACT

Hemodialysis is one of the applied membrane technologies that is regarded as a life-saving therapy for patients with impaired kidneys. It purifies blood toxins outside of the patient's body using a dialyzer as a kidney replacement. Synthetic materials such as polyethersulfone (PES) polymer are currently being used to fabricate the dialyzer membrane for hemodialysis. However, the blood compatibility or hemocompatibility of these materials are still inadequate and administration of an anticoagulant (heparin) is required throughout the dialysis procedure to avoid blood clotting. Therefore this study aimed at developing a biocompatibility membrane for hemodialysis application. In this study, poly (1,8-octanediol citrate) (POC) that synthesized through a simple polycondensation method was used to enhance membrane biocompatibility. Different compositions of POC (0-3%) were added into polyethersulfone (PES) dope solutions to fabricate modified biocompatible PES membranes via the phase-inversion technique. The biocompatibility of the modified PES membranes was evaluated by human serum fibrinogen (FBG) protein adsorption, platelet adhesion, activated partial thromboplastin time (APTT) and prothrombin time (PT), thrombin–antithrombin III (TAT), complement (C3a and C5a) activation and Ca^{2+} absorption on membrane. Results showed that higher POC wt.% caused a 31% reduction of FBG adsorption, less platelets adhesion, prolonged APTT (11.1 seconds) and PT (2.5 seconds), lower TAT activation, suppressed C5a and C3a activation and absorbed 35% more Ca^{2+} ion compared to pristine PES membrane. These results indicated that modified PES blended POC has good biocompatibility properties, suggesting potential application in the field of blood purification, especially in hemodialysis.

ABSTRAK

Hemodialisis merupakan antara aplikasi dalam teknologi membran yang dapat menyelamatkan nyawa pesakit yang mengalami masalah buah pinggang. Ia membersihkan toksin darah diluar badan pesakit menggunakan dialiser sebagai pengganti kepada buah pinggang. Bahan sintetik seperti polimer polietersulfon (PES) digunakan sebagai bahan utama untuk membuat membran dialiser untuk hemodialisis. Walau bagaimanapun, keserasian antara darah dan bahan sintetik ini masih tidak mencukupi dan anti-beku darah (heparin) diperlukan sepanjang prosedur dialisis untuk mengelakkan pembekuan darah berlaku. Objektif kajian ini adalah bertujuan untuk membangunkan satu membran bioserasi yang baru bagi digunakan dalam aplikasi hemodialisis. Dalam kajian ini, poli(1,8-octanediol citrate) (POC) yang disintesis melalui kaedah polikondensasi mudah telah digunakan untuk meningkatkan bioserasian dengan membran. Komposisi POC yang berbeza (0-3%) telah ditambah pada larutan polimer PES untuk membuat membran PES yang diubahsuai melalui teknik fasa penyongsangan. Keserasian-bio membran PES yang diubahsuai telah dinilai dengan penjerapan protein fibrinogen serum (FBG), kebolelekatan platelet, masa separa-aktif tromboplastin (APTT) dan masa prothrombin (PT), pengaktifan thrombin-antithrombin III (TAT), pengaktifan komplimen (C3a dan C5a) dan penyerapan Ca^{2+} ion pada membran. Hasil kajian menunjukkan bahawa dengan wt.% POC yang tinggi, telah mengurangkan 31% penjerapan FBG, mengurangkan platelet melekat, memanjangkan masa APTT (11.2 saat) dan PT (2.5 saat) lebih lama, pengaktifan TAT yang rendah, mengurangkan pengaktifan C5a dan C3a dan menyerap lebih 35% ion Ca^{2+} berbanding membrane asal. Dapatan kajian ini menunjukkan bahawa membran yang diubahsuai dengan campuran POC mempunyai kebioserasian yang lebih baik bagi menyediakan aplikasi membran yang lebih praktikal dalam bidang pembersihan darah, terutamanya bagi aplikasi hemodialisis.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	viii
	LIST OF TABLES	xi
	LIST OF FIGURES	xii
	LIST OF SYMBOLS	xv
	LIST OF ABBREVIATION	xvii
	LIST OF APPENDICES	xix
1	INTRODUCTION	1
	1.1 Background of Research	1
	1.2 Problem Statement	3
	1.3 Objective of Study	4
	1.4 Scope of Study	5
	1.5 Significance of Study	6
2	LITERATURE REVIEW	8
	2.1 Membrane Technology	8
	2.1.1 Introduction	8
	2.1.3 Membrane In Brief	8
	2.1.4 Membrane Process	11
	2.2 Hemodialysis	13
	2.2.1 Introduction	13
	2.2.2 Hemodialysis in Malaysia	13
	2.2.3 Human Kidney	14

2.2.4	Brief History of Hemodialysis	16
2.2.5	Principle of hemodialysis	17
2.2.6	Hemodialysis System	19
2.2.6.1	Hemodialyzer	19
2.2.6.2	Blood Circuit	20
2.2.6.3	Dialysate	21
2.3	Membrane for Hemodialysis	21
2.4	Biocompatibility of Hemodialysis Membrane	24
2.4.1	Blood Compatibility of Membrane	26
2.4.2	Protein Adsorption on Membrane Surface	27
2.4.3	Coagulation Activation by Membrane	27
2.4.4	Adhesion of Platelet on Membrane Surface	28
2.4.5	Complement Activation by Membrane	29
2.4.6	Other Membrane Induced Biological Reaction	31
2.5	Hemodialysis Membrane Bio-incompatibility	31
	Clinical Consequences	
2.5.1	Heparin Induced Thrombocytopenia	32
2.6	Improvement of Membrane Biocompatibility	32
2.6.1	Modification of Cellulose-Based Membrane	32
2.6.2	Modification of Synthetic Membrane	32
2.7	Poly(diols citrate)	37
2.7.1	Poly(1,8-octanediol citrate)	38
3	RESEARCH METHODOLOGY	41
3.1	Research Design	41
3.2	Material	42
3.2.1	Polymer selection	42
3.2.2	Solvent	43
3.2.3	Additives	43
3.3	Preparation and Characterization of Poly(1,8-octanediol citrate)	43

	3.4 Flat-sheet Membrane Preparation	
	3.5 Membrane Characterization	45
	3.5.1 Membrane Surface Composition	46
	3.5.2 Thermogravimetric Analysis (TGA)	46
	3.5.3 Membrane Morphology Studies	46
	3.5.4 Zeta Potential	47
	3.5.5 Membrane Wettability	47
	3.5.6 Membrane Porosity and Pore Size	47
	3.6 Membrane Performance Evaluation	48
	3.6.1 Pure Water Flux (PWF) and BSA Rejection	48
	3.6.2 Uremic Toxins Clearance	
	3.7 Blood Compatibility Evaluations	48
	3.7.1 Protein Adsorption	50
	3.7.2 Platelets Adhesion	51
	3.7.3 Plasma Coagulation Time	52
	3.7.4 Membrane Ca ²⁺ Absorption	53
	3.7.5 Complement and Coagulation Activation	54
		54
4	RESULT AND DISCUSSION	56
	4.1 Characterization of Synthesized Poly(1,8-octanediol citrate)	56
	4.1.1 ATR-FTIR Analysis	56
	4.1.2 ¹ H NMR Spectra Analysis	57
	4.1.3 MALDI-TOF Analysis	58
	4.2 Characterization of Pristine PES and Modified PES Membranes with Different POC Concentration Loading	59
	4.2.1 Membrane Surface Composition (ATR-FTIR)	59
	4.2.2 Membrane Composition TGA Analysis	60
	4.2.3 Membrane Cross-section Morphology	61
	4.2.4 Membrane Surface Morphology	63
	4.2.5 Membrane Surface Charge	64

4.2.6	Membrane Wettability	65
4.2.7	Membrane Porosity and Pore Size	66
4.3	Effect of Different POC Weight Loading on Membrane Dialysis Performance	67
4.3.1	Pure Water Flux and BSA Rejection	67
4.3.2	Uremic Toxins Clearance	68
4.4	Effect of Different POC Weight Loading on Membrane Biocompatibility	71
4.4.1	Protein Adsorption	71
4.4.2	Platelet Adhesion	72
4.4.3	Plasma Coagulation Time (APTT and PT)	73
4.4.4	Membrane Ca ²⁺ absorption	74
4.4.5	Coagulation Activation	75
4.4.6	Complement Activation	76
5	CONCLUSION AND RECOMMENDATIONS	78
5.1	Conclusion	79
5.2	Recommendation	80
	REFERENCES	81
	APPENDIX A-E	90-99

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Membrane separation process summary	11
2.2	Kidney daily excreted fluid and substances	15
2.3	Hemodialyzer performance parameter	19
2.4	Common dialysis membrane type and examples	21
2.5	Modification of cellulose based hemodialysis membrane	32
2.6	Modification of synthetic membrane to improve biocompatibility	34
2.7	Poly(diols citrates) example and its potential applications	39
3.1	Dope solution composition for membrane preparation	45
3.2	Normal APTT and PT test range value.	52

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Membrane basic and process.	8
2.2	Membrane morphological structure classification.	9
2.3	Dead-end and cross-flow filtration mode.	10
2.4	The number of new dialysis patients and total current dialysis patients for the last 10 years.	13
2.5	Cross-section of human kidney and its functional unit nephron.	14
2.6	Dialysis, ultrafiltration and convection	16
2.7	Hemodialysis delivery system setup.	19
2.8	Hemodialyzer module.	19
2.9	Chemical structure of membrane material	21
2.10	Electron microscopic cross-sectional view of the synthetic membrane with three different morphology	22
2.11	Biological response of blood-dialysis membrane interaction during hemodialysis process.	24
2.12	The overview of blood-membrane interaction showing component that link to each other causing thrombosis and inflammation during hemodialysis.	24
2.13	Illustration of fibrin formation, platelet adhesion and thrombus formation of membrane surface.	25
2.14	Blood coagulation activation cascade pathways.	27
2.15	Complement system activation by hemodialysis membrane through alternative pathway.	29

2.16	SEM images of different membranes with reduce platelet adhesion.	36
2.17	Poly(1,8-octanediol citrates) (POC) formation from two monomer, citric acid and 1,8-octanediol through polycondensation method.	37
3.1	Experimental flow chart	40
3.2	Chemical structure of PES, Veradel® A-301	41
3.3	Dimethylacetamide (DMAc)	42
3.4	Polyvinylpyrrolidone (PVP)	43
3.5	Experimental set-up for poly(1,8-octanediol citrate).	44
3.6	Schematic diagram of bench scale cross-flow ultrafiltration system.	48
3.7	Schematic diagram of bench scale for dialysis set-up.	49
3.8	Platelet adhesion procedure.	51
3.9	Graphical diagram for coagulation assay procedure.	52
3.10	Graphical diagram for complement and coagulation activation procedure.	54
4.1	The ATR-FTIR spectrum of POC polymer (S1 and S2) that synthesized in two different time.	56
4.2	¹ H NMR spectrum of POC polymer.	56
4.3	MALDI-OF-MS spectra of POC polymer (S1 and S2) that synthesized in two different time.	57
4.4	ATR-FTIR spectra of the POC and the M-0, M-1, M-2 and M-3 membrane.	58
4.5	The thermal degradation of PES (M-0) and POC blend PES (M-1, M-2 and M-3) flat-sheet membrane.	59
4.6	SEM micrograph images of (a) cross-sectional and (b) top layer of the M) flat-sheet membranes	61
4.7	AFM topography images of each membrane samples surface.	63
4.8	Membranes zeta-potential at pH 7	64

4.9	Contact angle and percentage of water absorbed by each membrane samples.	65
4.10	Porosity and pore size for each membrane samples.	66
4.11	Pure water flux and BSA rejection for each membrane samples.	67
4.12	Uremic toxins (urea and creatinine clearance) for each membrane samples after 2 hours of dialysis.	69
4.13	The concentration ($\mu\text{g}/\text{cm}^2$) of protein (bovine serum albumin and fibrinogen) being absorbed on M-0, M-1, M-2 and M-3 membranes surface.	71
4.14	S.E.M micrographic images of platelet adhesion on the M-0, M-1, M-2 and M-3 membrane surface.	72
4.15	(a) Activated partial thromboplastin times (APTTs) and prothrombin times (PTs) for the membranes, respectively. (b) Percentage of Ca^{2+} being absorb by 100 mg of membrane.	74
4.16	The concentrations of TAT III generated for by each membrane samples	75
4.17	The concentrations of complement C3a and C5a generated for by each membrane samples	76

LIST OF SYMBOLS

γ	-	Gamma
α	-	Alpha
β	-	Beta
μm	-	micrometer
μL	-	microliter
Q_b	-	Blood flow rate
Q_d	-	Dialysate flow rate
IgG	-	Immunoglobulin G (antibody G)
δt	-	Solubility coefficient ($\text{MPa}^{1/2}$)
Δx	-	thickness selectivity of the membrane (μm)
L	-	thickness of the support layer (μm)
K_{uf}	-	ultrafiltration coefficient of the membrane (mL/h/mmHg)
K_oA	-	Clearance efficiency
M_w	-	Molecular weight (g/mol)/Da
Da	-	Dalton
C	-	Carbon
H	-	Hydrogen
N	-	Nitrogen
O	-	Oxygen
t	-	Time taken (h).

R_a	-	mean surface roughness
M_1	-	Dry membrane mass
M_2	-	Wet membrane mass
ε	-	Porosity
A	-	Effective membrane area
ρ	-	water density (998 kg/m ³)
μ	-	water viscosity at 25 °C (8.9×10^{-4} Pa.s)
ΔP	-	pressure (Pascal)
r_m	-	pore size radius
Q	-	flow rate of permeated water (m ³ /s)
J	-	Water flux
V	-	volume of water permeated (L)
Δt	-	permeation time
C_p	-	Permeate concentration
C_f	-	Feed concentration
$CaCl_2$	-	Calcium chloride
$C3a$	-	Complement 3a
$C5a$	-	Complement 5a
CL	-	Clearance
C_i	-	initial concentration
C_s	-	sample concentration

LIST OF ABBREVIATION

PES	-	Polyethersulfone
POC	-	Poly(1,8-octane citrate)
CABE	-	Citric acid based elastomer
HD	-	Hemodialysis
HF	-	Hemofiltration
RRT	-	Renal replacement therapy
PSf	-	Polysulfone
PMMA	-	Poly(methyl methacrylate)
PAN	-	Polyacrylonitrile
ATR-FTIR	-	Attenuated Total Reflectance-Fourier Transform Infrared Spectroscopy
¹ H NMR	-	Proton Nucleus Magnetic Resonance
MALDI-TOF	-	Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry
SEM	-	Scanning electron microscopy
TF	-	Tissue factor
vWF	-	von Willebrand factor
ACE	-	Angitensin converting enzyme
HIT	-	Heparin-induced thrombocytopenia
POC-HA	-	Poly (1,8-octanediol citrates)-hydroxyapatite
CUPE	-	Crosslinked urethane-doped polyester
TGA	-	Thermogravimetric analysis

BSA	-	Bovine serum albumin
FBG	-	Fibrinogen
APTT	-	Activated partial thromboplastin time
PT	-	Prothrombin time
TAT	-	Thombin anti-thrombin III
C3a	-	Complement 3a
C5a	-	Complement 5a
MAC	-	Membrane attack complex

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Poly(1,8-octanediol citrate)	90
B	Membrane dope solution	90
C	QuantiChrom™ Assay Kit (BioAssay Systems, USA) Reagent and Standard Preparation	91
D	Micro BCA™ Protein Assay Kits	93
E	Complement C3a and C5a, (Cusabio Biotech)	94
F	Human TAT Complex (Enzygnost TAT micro, Assay Pro)	96

CHAPTER 1

INTRODUCTION

1.1 Background of Research

Since the breakthrough in 1978 by french cleric, J. Abbé Nollet, membrane technology was only being studied by scientist in 1960s for its commercialization in industrial application (Fane et al., 2011). In 1960s, Loeb and Sourirajan developed asymmetric membrane for industrial applications (Kołtuniewicz, 2005). Then in 1980s, techniques such as reverse osmosis, ultrafiltration, microfiltration and electro dialysis resulted in worldwide membrane commercialize applications in large scale plants (Baker, 2012; Fane et al., 2011). After years of rapid development of the membrane technology, the applications of membranes was used in the development of membranes for medical separation processes, in particular, the artificial kidney (hemodialysis) (Baker, 2012).

Kidney is a vital organ in human composed of millions unit of filter which remove excess waste products and fluid out of the body. This function is essential for normal homeostasis in maintaining body fluid and chemical composition inside the body. Damage to the kidney may lead to the reduction in its performance. Subsequently, the damage result in build-up of toxins and excess fluid that may affect health. A therapy called renal replacement therapy (RRT) or commonly known as hemodialysis is required to overcome the problem.

Hemodialysis is highly successful lifesaving and life sustaining therapy for renal failure patient (Vilar & Farrington, 2011). In 2012, there were 28,590 patients receiving dialysis in Malaysia, and vast majority (92%) of these patients are on

hemodialysis (Lim et al., 2013). According to Datuk Seri Dr. Hasan Abdul Rahman Malaysian Health director-general, every year a patient in average had to spend about RM 33,000.00 for dialysis treatment and more than RM 700 million for total dialysis patients (Arukesamy, 2011). However, 60% of the cost was government subsidized. Therefore, Malaysian government spent almost RM 420 million for dialysis operational cost only (include staff salaries, consumables e.g., dialyser reuse 3 times, blood lines, CAPD system and dialysate), equipment maintenance and utilities) which do not include associate medical cost (RM 270/patient), capital cost and general hospital cost (RM 360/patient) (Lim et al., 1999).

In general, the process of dialysis involve two way movement of molecules across a semipermeable membrane. Clinically, this movement takes place in and out of blood, across a semipermeable membrane. The blood is exposed to an artificial membrane outside of the body, the process is called hemodialysis (HD) or hemofiltration (HF) (Ahmad, 2009). The dialyzer made of dialysis membrane is the most critical and important part of dialysis set up form separate adjacent paths for blood and dialysate, which flow on opposite sides of the membrane in opposite directions to maximize diffusion gradients. According to Malaysian Hemodilaysis Quality and Standard (2012) published by Ministry of Health Malaysia a dialyzers should be made from biocompatible membrane and approved by regulatory authorities in USA, Europe, Japan or local equivalent.

In selecting the material for hemodialysis, benchmarks such as biocompatibility towards white blood cell and complement system, impermeability toward dialysate impurities, adsorption capacity and pore size must be considered (Boure, 2004). The success of hemodialysis is highly dependent on the membrane used (Gautham et al., 2013). The earlier type of material used in the making of dialysis membrane was the cellulose based membrane (e.g.; cuprophane). Later, due to certain undesired effect such as immunological reaction, the cellulose was substituted by other material like mixed-cellulosic synthetic membrane (e.g.; Hemophan). Currently, the synthetic membrane (e.g; polyethersulfone (PES), polysulfone (PS), polyacrylonitrile (PAN), polycarbonate, polymethylmethacrylate (PMMA), and polyamide) is the latest type of material commonly used to produce dialysis membrane (Ahmad, 2009). Moreover, aside from having better performance in removing middle molecule (500-

15000 Da) and β 2-microglobulin, the synthetic membranes also comprise higher biocompatibility properties (Kerr & Huang, 2010; Macleod et al., 2005).

Here, in this study PES is used as a material to fabricate dialysis membrane. The PES membranes are widely employed in biomedical fields such as artificial organs and medical devices used for blood purification, e.g., hemodialysis, hemodiafiltration, hemofiltration, plasmapheresis and plasma collection (Su et al., 2011). In order to achieve the production of high biocompatibility dialysis membrane a citric acid derived biodegradable elastomers (CABEs) was added as additive. This new class of biomaterials is all synthesized with non-toxic monomers using simple and cost effective procedures. These materials share one common monomer named citric acid, which is a non-toxic metabolic product of the Krebs cycle. In addition to the sodium form of citric acid, sodium citrate, is an anticoagulant currently used in hospitals. Thus, CABEs may also possess suitable hemocompatibility for blood contacting applications (Tran et al., 2009).

A novel biodegradable elastomer, poly (1, 8-octanediol-co-citrate) (POC) is synthesized and used as biocompatible additives to produce biocompatible modified PES flat-sheet membranes. The fabricated membranes are characterized for its physical and chemical properties. The dialysis performance of the modified membrane also been evaluated. The biocompatibility of the modified membranes are analyzed by studying the membrane blood compatible properties.

1.2 Problem Statement

In blood-contacting membrane application such as hemodialysis, the protein adsorption on the membrane surface cause some serious problem to membrane performance. The adsorption of protein on membrane surface eventually lead to membrane fouling resulting in flux declining and reduction of membrane selectivity. Furthermore, the adsorbed proteins plugged membrane pores affecting efficacy, by decreasing the pore size of the membrane and prevent transmembrane crossing for waste and solutes. Membrane protein adsorption may be caused by few factors such

as the membrane surface chemistry, morphology, hydrophilicity and also the adsorbed protein size, charge, shape, pH value, and so on. Besides causing membrane fouling that affect membrane performance, adsorption of protein also leads to membrane biocompatibility problems (Sun et al., 2003).

Biocompatibility of the membrane is one of the important aspect in developing a membrane for hemodialysis. It is the biological reaction between dialysis membrane and the blood. Exposure of dialysis membrane to blood leads to normal physiological response of blood components such as activation of platelets, inflammatory response, complement blood cells, and complement cascade (Ahmad, 2009; Gawaz et al., 1999) which may cause adverse effect to the dialysis patient. Activation and adhesion of the platelet on the membrane surface cause coagulation of blood (Sperling et al., 2009). Therefore, to avoid coagulation an anticoagulant such as heparin is used in the dialysis system. Yet, the usage of heparin causes several complication to dialysis patient like heparin-induced thrombocytopenia (HIT) Type II and bleeding that might lead to severe morbidity and mortality (Suranyi & Chow, 2010). Therefore, effective membrane with higher biocompatibility to perform sufficient hemodialysis is an important key to cope with the rising demands from kidney failure patient.

1.3 Objectives of Study

Based on the problem statement, the aim of this study was to reduce the incidence of membrane compatible reaction and membrane fouling in the dialyzer. Therefore, this study sought to produce a new biocompatible synthetic polymer membrane to perform sufficient hemodialysis and to cope with the rising demands from kidney failure patient in Malaysia. The objectives of this research are:

1. To synthesize poly(1,8-octanediol citrate) (POC) as additive to improve PES membrane biocompatibility
2. To fabricate and characterize physical and chemical properties of the modified PES flat-sheet membrane with different POC wt.% loading (0-3%).

3. To evaluate the biocompatibility of the modified PES/POC flat-sheet membranes different POC wt.% loading (0-3%) for hemodialysis applications.

1.4 Scope of Study

1. Preparation of poly(1,8-octane citrate) (POC) via simple polycondensation method.
2. Characterization of the POC with Attenuated Total Reflectance-Fourier Transform Infrared Spectroscopy (ATR-FTIR), proton Nucleus Magnetic Resonance (^1H NMR), Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry (MALDI-TOF).
3. Dope preparation for different wt.% loading (0-3%) concentration of POC blend PES membranes.
4. Fabrication of modified PES membranes via liquid-liquid phase inversion method.
5. Characterization of modified PES membranes by ATR-FTIR, scanning electron microscope (SEM), atomic-force microscopy (AFM), Thermogravimetric Analysis (TGA), zeta-potential, membrane wettability, membrane porosity and pore size.
6. Evaluation of the modified membranes' performance for pure water flux, solute rejection and uremic toxins clearance.
7. Evaluation of the modified membranes biocompatibility through protein adsorption, platelet adhesion, plasma coagulation time, calcium ion absorption and coagulation and complement activation.

1.5 Significance of Study

The number of dialysis patient in Malaysia is rapidly increasing each and every year. Data from 22nd Report of the Malaysian Dialysis 2015 and Transplant Registry by the National Renal Registry showed the number of dialysis patient in Malaysia was steeply increase for the last 10 years from 13000 in 2004 and at least 35000 people in 2015. Each year the amount of new dialysis patient increase with 6107 new people need dialysis in 2014 (Goh & Ong, 2015). Herein Malaysia, there are few researchers that currently doing research on dialysis membrane but not to be commercialize. None of them focuses on improving biocompatibility of hemodialysis membrane and only study on membrane performances. Nowadays, local dialysis center and hospital still use an imported dialyzer which can be costly. Therefore, this study aims to produce locally produced high performance biocompatible hemodialysis membrane which eventually can reduce current dialyzer cost in Malaysia.

Dialyzer or artificial kidney is the heart of the dialysis which contain a semi-permeable membrane that function as kidney replacement. Nowadays, synthetic polymer such as polyethersulfone (PES) is being used to make dialysis membrane due to excellent physical and chemical properties. However, the hydrophobic nature of the polymer tends to adsorb protein and cause membrane bio-incompatibility that lead to thrombosis. Several modification have been made to improve membrane biocompatibility with anti-coagulant properties, for example by using a heparin (anti-coagulant) to modified membrane through coating (AN69ST) or grafting it on membrane surface (Morena et al., 2010; Tang et al., 2012). These modified membrane allows heparin-free dialysis which can reduce adverse events from prolong usage of heparin such as heparin induced thrombocytopenia (HIT).

The significance of this current research is by studying the development of modified PES dialysis membranes that blended with POC to improve it biocompatibility. Previous study had found that POC exhibit a biocompatible properties for blood contacting application, hence modification of the PES dialysis membrane with POC can improve biocompatibility of membrane and prevent

thrombosis formation on membrane surface. Therefore, by using this PES blend POC dialysis membrane, an anticoagulant free dialysis can be perform.

REFERENCES

- Ahmad, S. (2009). *Manual of clinical dialysis* (2nd Edition, Vol. 40). New York: Springer.
- Alaei Shahmirzadi, M. A., Hosseini, S. S., Ruan, G., & Tan, N. R. (2015). Tailoring PES nanofiltration membranes through systematic investigations of prominent design, fabrication and operational parameters. *RSC Adv.*, 5(61), 49080–49097.
- Ameer, G., Yang, J., & Webb, A. R. (2012). Novel Biodegradable Elastomeric Scaffold For Tissue Engineering and Light Scattering Finerprinting Method for Testing The Same. Evanston, IL, USA: U.S. Patent and Trademark Office.
- Amore, A., & Coppo, R. (2002). Immunological basis of inflammation in dialysis. *Nephrology Dialysis Transplantation*, 17 Suppl 8, 16–24.
- Arepally, G. M., & Ortel, T. L. (2006). Heparin-Induced Thrombocytopenia. *The New England Journal of Medicine*, 355, 809–817.
- Arukesamy, K. (2011, December 26). Government funds almost 60% of all haemodialysis funding. *The Sun Daily*. Kuala Lumpur.
- Azar, A. T. (2013). *Modeling and Control of Dialysis Systems*. (A. T. Azar, Ed.) *Journal of Chemical Information and Modeling* (Vol. 53). Springer-Verlag Berlin Heidelberg.
- Baker, R. W. (2012). *Membrane Technology and Applications*. *Membrane Technology* (Third Edition). John Wiley & Sons, Ltd.
- Barzin, J., Feng, C., Khulbe, K. C., Matsuura, T., Madaeni, S. S., & Mirzadeh, H. (2004). Characterization of polyethersulfone hemodialysis membrane by ultrafiltration and atomic force microscopy. *Journal of Membrane Science*, 237(1-2), 77–85.
- Barzin, J., Madaeni, S. S., Mirzadeh, H., & Mehrabzadeh, M. (2004). Effect of polyvinylpyrrolidone on morphology and performance of hemodialysis membranes prepared from polyethersulfone. *Journal of Applied Polymer Science*, 92, 3804–3813.
- Barzin, J. & Sadatnia, B. (2007). Theoretical phase diagram calculation and membrane morphology evaluation for water/solvent/polyethersulfone systems. *Polymer*,

48(6), 1620–1631.

- Baura, G. D. (2012). Hemodialysis Delivery Systems. In *Medical Device Technologies: A Systems Based Overview Using Engineering Standards* (pp. 193–216). Massachusetts: Academic Press.
- Boure, T. (2004). Which dialyser membrane to choose? *Nephrology Dialysis Transplantation*, 19(2), 293–296.
- Chanard, J., Lavaud, S., Randoux, C., & Rieu, P. (2003). New insights in dialysis membrane biocompatibility: relevance of adsorption properties and heparin binding. *Nephrology Dialysis Transplantation*, 18, 252–257.
- Cheung, A. K. (1990). Biocompatibility of Hemodialysis Membranes. *Journal of the American Society of Nephrology*, 1(2), 150–161.
- Clark, W. R., Hamburger, R. J., & Lysaght, M. J. (1999). Effect of membrane composition and structure on solute removal and biocompatibility in hemodialysis. *Kidney International*, 56(6), 2005–2015.
- Dahe, G. J., Teotia, R. S., Kadam, S. S., & Bellare, J. R. (2011). The biocompatibility and separation performance of antioxidative polysulfone/vitamin E TPGS composite hollow fiber membranes. *Biomaterials*, 32(2), 352–365.
- Ding, T., Liu, Q., Shi, R., Tian, M., Yang, J., & Zhang, L. (2006). Synthesis, characterization and in vitro degradation study of a novel and rapidly degradable elastomer. *Polymer Degradation and Stability*, 91, 733–739.
- Drüeke, T. B. (2000). β 2-Microglobulin and amyloidosis. *Nephrology Dialysis Transplantation*, 15(suppl 1), 17–24.
- Eknoyan, G. (2009). The wonderful apparatus of John Jacob Abel called the “artificial kidney”. *Seminars in Dialysis*, 22(3), 287–96.
- Fane, A. G., Wang, R., & Jia, Y. (2011). Membrane Technology: Past, Present and Future. In L. K. Wang (Ed.), *Handbook of Environmental Engineering: Membrane and Desalination Technologies* (Vol. 13, pp. 691–692).
- Fleming, G. M. (2014). Renal replacement therapy review. *Organogenesis*, 7(1), 2–12.
- Galli, F., Rovidati, S., Chiarantini, L., Campus, G., Canestrari, F., & Buoncrisiani, U. (1998). Bioreactivity and biocompatibility of a vitamin E-modified multilayer hemodialysis filter. *Kidney International*, 54(2), 580–589.
- Gao, A., Liu, F., & Xue, L. (2014). Preparation and evaluation of heparin-immobilized poly (lactic acid) (PLA) membrane for hemodialysis. *Journal of Membrane Science*, 452, 390–399.

- Gautham, A., Muhammed Javad, M., Manavalan, M., & Najeeb, M. A. (2013). Hemodialysis Membranes: Past, Present and Future Trends. *International Research Journal of Pharmacy*, 4(5), 16–19. 8407.04505.
- Gawaz, M. P., Mujais, S. K., Schmidt, B., Blumenstein, M., & Gurland, H. J. (1999). Platelet-Leukocyte Aggregates During Hemodialysis: Effect of Membrane Type. *Artificial Organs*, 23(1), 29–36.
- Goh, B., & Ong, L. (2015). *22nd Report of The Malaysian Dialysis and Transplant Registry*. Kuala Lumpur.
- Golper, T. A., Fissell, R., Fissell, W. H., Hartle, P. M., Sanders, M. L., & Schulman, G. (2014). Hemodialysis: Core Curriculum 2014. *American Journal of Kidney Diseases*, 63(1), 153–63.
- Gorbet, M. B., & Sefton, M. V. (2004). Biomaterial-associated thrombosis: roles of coagulation factors, complement, platelets and leukocytes. *Biomaterials*, 25(26), 5681–703.
- Grooteman, M. P. C., & Nubé, M. J. (2004). Impact of the type of dialyser on the clinical outcome in chronic haemodialysis patients: does it really matter? *Nephrology Dialysis Transplantation*, 19(12), 2965–70.
- Hakim, R. M. (1993). Clinical implications of hemodialysis membrane biocompatibility. *Kidney International*, 44, 484–494.
- Hakim, R. M. (2000). Clinical implications of biocompatibility in blood purification membranes. *Nephrol Dial Transplant*, 15 Suppl 2, 16–20.
- Hakim, R. M., Fearon, D. T., & Lazarus, J. M. (1984). Biocompatibility of dialysis membranes: effects of chronic complement activation. *Kidney International*, 26(2), 194–200.
- Hayama, M., Yamamoto, K. I., Kohori, F., Uesaka, T., Ueno, Y., Sugaya, H., ... Sakai, K. (2004). Nanoscopic behavior of polyvinylpyrrolidone particles on polysulfone/polyvinylpyrrolidone film. *Biomaterials*, 25(6), 1019–1028.
- Higuchi, A., Shirano, K., Harashima, M., Yoon, B. O., Hara, M., Hattori, M., & Imamura, K. (2002). Chemically modified polysulfone hollow fibers with vinylpyrrolidone having improved blood compatibility. *Biomaterials*, 23(13), 2659–2666.
- Hoenich, N. A. (2004). Update on the Biocompatibility of Hemodialysis Membranes. *Hong Kong Journal of Nephrology*, 6(2), 74–78. [http://doi.org/10.1016/S1561-5413\(09\)60162-9](http://doi.org/10.1016/S1561-5413(09)60162-9).

- Hoenich, N. A. (2007). Cellulose for Medical Applications: Past, Present, and Future. *BioResources*, 1(2), 270–280.
- Hoenich, N. A., Woffindin, C., Turnbulls, J., & Sarah, J. (1997). Synthetically modified cellulose: an alternative to synthetic membranes for use in haemodialysis?, *18*(19), 1299–1303.
- Huang, X.-J., Guduru, D., Xu, Z.-K., Vienken, J., & Groth, T. (2011). Blood Compatibility and Permeability of Heparin-Modified Polysulfone as Potential Membrane for Simultaneous Hemodialysis and LDL Removal. *Macromolecular Bioscience*, 11(1), 131–140.
- Irfan, M., & Idris, A. (2015). Overview of PES biocompatible / hemodialysis membranes: PES – blood interactions and modification techniques. *Materials Science and Engineering C*, 56, 574–592.
- Irfan, M., Idris, A., Yusof, N. M., Khairuddin, N. F. M., & Akhmal, H. (2014). Surface modification and performance enhancement of nano-Hybrid f-MWCNT /PVP90/PES hemodialysis membranes. *Journal of Membrane Science*, 467, 73–84.
- Ishihara, K., Fukumoto, K., Iwasaki, Y., & Nakabayashi, N. (1999). Modification of polysulfone with phospholipid polymer for improvement of the blood compatibility . Part 2: Protein adsorption and platelet adhesion, *20*, 1553-1559.
- Jacobs, C. (2009). [Renal replacement therapy by hemodialysis: an overview]. *Néphrologie & Thérapeutique*, 5(4), 306–12.
- Kaleekkal, N. J., Thanigaivelan, A., Tarun, M., & Mohan, D. (2015). A functional PES membrane for hemodialysis - Preparation, Characterization and Biocompatibility. *Chinese Journal of Chemical Engineering*, 23(7), 1236–1244.
- Kerr, P. G., & Huang, L. (2010). Review: membranes for haemodialysis. *Asian Pacific Society of Nephrology*, 15(4), 381–5.
- Kim, K. J., Fane, A. G., Fell, C. J. D., & Joy, D. C. (1992). Fouling mechanisms of membranes during protein ultrafiltration. *Journal of Membrane Science*, 68(1-2), 79–91.
- Kokubo, K., Kurihara, Y., Kobayashi, K., Tsukao, H., & Kobayashi, H. (2015). Evaluation of the Biocompatibility of Dialysis Membranes. *Blood Purification*, 40(4), 293–297.
- Kołtuniewicz, A. B. (2005). The history and state of arts in membrane technologies. Retrieved November 26, 2015, from

http://www.etseq.urv.es/assignatures/ops/presentacio_membranes.pdf.

- Kourtzelis, I., Markiewski, M. M., Doumas, M., Rafail, S., Kambas, K., Mitroulis, I., Lambris, J. D. (2010). Complement anaphylatoxin C5a contributes to hemodialysis-associated thrombosis. *Blood*, *116*(4), 631–639.
- Krieter, D. H., & Wanner, C. (2010). Membranes for Dialysis and Hemofiltration. *JAMA: The Journal of the American Medical Association*, *304*(13), 1497.
- Li, L., Cheng, C., Xiang, T., Tang, M., Zhao, W., Sun, S., & Zhao, C. (2012). Modification of polyethersulfone hemodialysis membrane by blending citric acid grafted polyurethane and its anticoagulant activity. *Journal of Membrane Science*, *405-406*, 261–274.
- Li, S. S., Xie, Y., Xiang, T., Ma, L., He, C., Sun, S. dong, & Zhao, C. S. (2016). Heparin-mimicking polyethersulfone membranes - hemocompatibility, cytocompatibility, antifouling and antibacterial properties. *Journal of Membrane Science*, *498*, 135–146.
- Lim, T. O., Lim, Y. N., Wong, H. S., Ahmad, G., Singam, T. S., Morad, Z. Loo, C. S. (1999). Cost effectiveness evaluation of the Ministry of Health Malaysia dialysis programme. *Medical Journal of Malaysia*, *54*, 442–452.
- Lim, Y. N, Goh, B., & Ong, L. (2013). *20th Report of The Malaysian Dialysis & Transplant Registry 2012*. (G. B. Leong, O. L. Meng, & L. Y. Ngo, Eds.). Kuala Lumpur: The National Renal Registry.
- Lin, W.-C., Liu, T.-Y., & Yang, M.-C. (2004). Hemocompatibility of polyacrylonitrile dialysis membrane immobilized with chitosan and heparin conjugate. *Biomaterials*, *25*(10), 1947–1957.
- Liu, T. Y., Lin, W. C., Huang, L. Y., Chen, S. Y., & Yang, M. C. (2005). Surface characteristics and hemocompatibility of PAN/PVDF blend membranes. *Polymers for Advanced Technologies*, *16*(5), 413–419.
- Liu, T.-Y., Lin, W.-C., Huang, L.-Y., Chen, S.-Y., & Yang, M.-C. (2005). Hemocompatibility and anaphylatoxin formation of protein-immobilizing polyacrylonitrile hemodialysis membrane. *Biomaterials*, *26*(12), 1437–44.
- Loh, F. F., & Shagar, L. k. (2013, November 7). Dialysis Subsidy Drying Up. *The Star*. Kuala Lumpur.
- Lonsdale, H. . (1982). The Growth of Membrane Technology. *Journal of Membrane Science*, *10*, 81–181.
- Ma, L., Su, B., Cheng, C., Yin, Z., Qin, H., Zhao, J., Zhao, C. (2014). Toward highly

- blood compatible hemodialysis membranes via blending with heparin-mimicking polyurethane: Study in vitro and in vivo. *Journal of Membrane Science*, 470, 90–101.
- Madaeni, S. S., & Rahimpour, A. (2005). Preparation of Polyethersulfone Ultrafiltration Membranes for Milk Concentration and Effects of Additives on Their Morphology and Performance. *Chinese Journal of Polymer Science*, 23(5), 539–548.
- Mader, S. S. (2010). Urinary System in Humans. In *Biology* (Tenth Edit, pp. 670–676). New York: McGraw-Hill.
- Markiewski, M. M., Nilsson, B., Ekdahl, K. N., Mollnes, T. E., & Lambris, J. D. (2007). Complement and coagulation: strangers or partners in crime? *Trends in Immunology*, 28(4), 184–92.
- Motlagh, D., Allen, J., Hoshi, R., Yang, J., Lui, K., & Ameer, G. (2007). Hemocompatibility evaluation of poly(diols citrate) in vitro for vascular tissue engineering. *Journal of Biomedical Materials Research. Part A*, 82(4), 907–16.
- Mudler, M. (1996). *Basic principles of membrane technology*. Kluwer Academic Publishers, Dordrecht, Netherlands.
- Nie, C., Ma, L., Xia, Y., He, C., Deng, J., Wang, L., ... Zhao, C. (2015). Novel heparin-mimicking polymer brush grafted carbon nanotube/PES composite membranes for safe and efficient blood purification. *Journal of Membrane Science*, 475, 455–468.
- Nie, S., Xue, J., Lu, Y., Liu, Y., Wang, D., Sun, S., Zhao, C. (2012). Improved blood compatibility of polyethersulfone membrane with a hydrophilic and anionic surface. *Colloids and Surfaces B: Biointerfaces*, 100, 116–125.
- Nilsson, B., Ekdahl, K. N., Mollnes, T. E., & Lambris, J. D. (2007). The role of complement in biomaterial-induced inflammation. *Molecular Immunology*, 44(1-3), 82–94.
- Ran, F., Nie, S., Zhao, W., Li, J., Su, B., Sun, S., & Zhao, C. (2011). Biocompatibility of modified polyethersulfone membranes by blending an amphiphilic triblock copolymer of poly(vinyl pyrrolidone)-b-poly(methyl methacrylate)-b-poly(vinyl pyrrolidone). *Acta Biomaterialia*, 7(9), 3370–3381.
- Sefton, M. V., Gemmell, C. H., & Gorbet, M. B. (2000). What really is blood compatibility? *Journal of Biomaterials Science. Polymer Edition*, 11(11), 1165–1182.

- Senthilkumar, S., Rajesh, S., Jayalakshmi, a., & Mohan, D. (2013a). Biocompatibility and separation performance of carboxylated poly (ether-imide) incorporated polyacrylonitrile membranes. *Separation and Purification Technology*, *107*, 297–309.
- Senthilkumar, S., Rajesh, S., Jayalakshmi, a., & Mohan, D. (2013b). Biocompatibility studies of polyacrylonitrile membranes modified with carboxylated polyetherimide. *Materials Science and Engineering: C*, *33*(7), 3615–3626.
- Senthilkumar, S., Rajesh, S., Mohan, D., & Soundararajan, P. (2013). Preparation, Characterization, and Performance Evaluation of Poly(Ether-imide) Incorporated Cellulose Acetate Ultrafiltration Membrane for Hemodialysis. *Separation Science and Technology*, *48*(1), 66–75.
- Shakaib, M., Ahmed, I., Yunus, R. M., Idris, A., & Hussain, A. (2013). Influence of monosodium glutamate additive on the morphology and permeability characteristics of polyamide dialysis membranes. *Journal of Applied Polymer Science*, *128*(5), 3346–3355.
- Sivaraman, B., & Latour, R. a. (2010). The relationship between platelet adhesion on surfaces and the structure versus the amount of adsorbed fibrinogen. *Biomaterials*, *31*(5), 832–839.
- Song, H., Ran, F., Fan, H., Niu, X., Kang, L., & Zhao, C. (2014). Hemocompatibility and ultrafiltration performance of surface-functionalized polyethersulfone membrane by blending comb-like amphiphilic block copolymer. *Journal of Membrane Science*, *471*, 319–327.
- Sperling, C., Fischer, M., Maitz, M. F., & Werner, C. (2009). Blood coagulation on biomaterials requires the combination of distinct activation processes. *Biomaterials*, *30*(27), 4447–56.
- Su, B. H., Fu, P., Li, Q., Tao, Y., Li, Z., Zao, H. S., & Zhao, C. S. (2008). Evaluation of polyethersulfone highflux hemodialysis membrane in vitro and in vivo. *Journal of Materials Science: Materials in Medicine*, *19*, 745–751.
- Su, B., Sun, S., & Zhao, C. (2011). Polyethersulfone Hollow Fiber Membranes for Hemodialysis. In A. Carpi (Ed.), *Progress in Hemodialysis - From Emergent Biotechnology to Clinical Practice*. INtech.
- Su, L. C., Xie, Z., Zhang, Y., Nguyen, K. T., & Yang, J. (2014). Study on the Antimicrobial Properties of Citrate-Based Biodegradable Polymers. *Frontiers in Bioengineering and Biotechnology*, *2*(July), 1–9.

- Sun, S., Yue, Y., Huang, X., & Meng, D. (2003). Protein adsorption on blood-contact membranes. *Journal of Membrane Science*, 222(1-2), 3–18.
- Suranyi, M., & Chow, J. S. F. (2010). Review: anticoagulation for haemodialysis. *Asian Pacific Society of Nephrology*, 15(4), 386–392.
- Tang, M., Xue, J., Yan, K., Xiang, T., Sun, S., & Zhao, C. (2012). Heparin-like surface modification of polyethersulfone membrane and its biocompatibility. *Journal of Colloid and Interface Science*, 386(1), 428–40.
- Thevenot, P., Hu, W., & Tang, L. (2008). Surface chemistry influences implant biocompatibility. *Current Topics in Medicinal Chemistry*, 8(4), 270–280.
- Tortora, G. J., & Derrickson, B. H. (2009). Urinary System. In *Principle Anatomy and Physiology* (Twelve Edi, pp. 1020–1080). Massachusetts: Wiley.
- Tran, R., Zhang, Y., Gyawali, D., & Yang, J. (2009). Recent Developments on Citric Acid Derived Biodegradable Elastomers. *Recent Patents on Biomedical Engineering*, 2(3), 216–227.
- Tsunoda, N., Kokubo, K., Sakai, K., Fukuda, M., Miyazaki, M., & Hiyoshi, T. (1999). Surface Roughness of Cellulose Hollow Fiber Dialysis Membranes and Platelet Adhesion. *ASAIJ Journal*, 45(5), 418–423.
- Vilar, E., & Farrington, K. (2011). Haemodialysis. *Medicine*, 39(7), 429–433.
- Vogler, E. a, & Siedlecki, C. a. (2009). Contact activation of blood-plasma coagulation. *Biomaterials*, 30(10), 1857–69.
- Wang, H., Yang, L., Zhao, X., Yu, T., & Du, Q. (2009). Improvement of Hydrophilicity and Blood Compatibility on Polyethersulfone Membrane by Adding Polyvinylpyrrolidone. *Chinese Journal of Chemical Engineering*, 17(2), 324–329.
- Wang, X., Yang, N., Xu, Q., Mao, C., Hou, X., & Shen, J. (2012). Preparation of a novel superhydrophobic PMMA surface with nanostructure and its blood compatibility. *E-Polymers*, (081), 1–8.
- Xiang, T., Xie, Y., Wang, R., Wu, M. B., Sun, S. D., & Zhao, C. S. (2014). Facile chemical modification of polysulfone membrane with improved hydrophilicity and blood compatibility. *Materials Letters*, 137, 192–195.
- Yamashita, A. C., & Sakurai, K. (2015). Dialysis Membranes- Physicochemical Structures and Features. In *Updates in Hemodialysis* (pp. 159–187). INTECH.
- Yang, J., Webb, A. R., & Ameer, G. a. (2004). Novel Citric Acid-Based Biodegradable Elastomers for Tissue Engineering. *Advanced Materials*, 16(6), 511–516.

- Yang, J., Webb, A. R., Pickerill, S. J., Hageman, G., & Ameer, G. a. (2006). Synthesis and evaluation of poly(diols citrate) biodegradable elastomers. *Biomaterials*, 27(9), 1889–98. <http://doi.org/10.1016/j.biomaterials.2005.05.106>
- Yin, Z., Cheng, C., Qin, H., Nie, C., He, C., & Zhao, C. (2015). Hemocompatible polyethersulfone/polyurethane composite membrane for high-performance antifouling and antithrombotic dialyzer. *Journal of Biomedical Materials Research - Part B* 2015, 103B, 97–105.
- Yue, W. W., Li, H. J., Xiang, T., Qin, H., Sun, S. D., & Zhao, C. S. (2013). Grafting of zwitterion from polysulfone membrane via surface-initiated ATRP with enhanced antifouling property and biocompatibility. *Journal of Membrane Science*, 446, 79–91.
- Zhang, X.-Q., Tang, H., Hoshi, R., De Laporte, L., Qiu, H., Xu, X., Ameer, G. a. (2009). Sustained transgene expression via citric acid-based polyester elastomers. *Biomaterials*, 30(13), 2632–41.
- Zhao, C., Liu, X., Nomizu, M., & Nishi, N. (2003). Blood compatible aspects of DNA-modified polysulfone membrane—protein adsorption and platelet adhesion. *Biomaterials*, 24(21), 3747–3755.
- Zhao, C., Xue, J., Ran, F., & Sun, S. (2013). Modification of polyethersulfone membranes – A review of methods. *Progress in Materials Science*, 58(1), 76–150.
- Zhao, H., Serrano, M. C., Popowich, D. a, Kibbe, M. R., & Ameer, G. a. (2010). Biodegradable nitric oxide-releasing poly(diols citrate) elastomers. *Journal of Biomedical Materials Research. Part A*, 93(1), 356–63.
- Zou, W., Qin, H., Shi, W., Sun, S., & Zhao, C. (2014). Surface modification of poly(ether sulfone) membrane with a synthesized negatively charged copolymer. *Langmuir*, 30(45), 13622–13630.