INACTIVATION OF Escherichia coli AND Listeria monocytogenes WITH Nd:YAG LASER AND ITS HARMONICS

MOSES ELISHA KUNDWAL

A thesis submitted in fulfilment of the requirements for the award of the degree of Doctor of Philosophy (Physics)

Faculty of Science Universiti Teknologi Malaysia

JANUARY 2016

To my wife, Dorcas Moses, and daughter, Bwalasom Moses, who endured a lot of inconveniences during the period of this study

ACKNOWLEDGEMENT

All thanks to God, the Most High, for His steadfast love and protection. He is the One who gives life and all the enablement for accomplishment in both little and great things.

This work would not have been possible without the contributions and encouragements of some individuals and institutions. Firstly, I wish to express my profound gratitude to my supervisor, Dr Abd Rahman Bin Tamuri, and cosupervisor, Dr Mohd Nizam Lani of Universiti Malaysia Terengganu. They did not only provide me with academic guidance, but have also been of great support and encouragement to me in some of my blink moments during the period of this study. I am very thankful to them.

I am greatly indebted to the management of Federal College of Education, Yola for sponsoring my study through the Tertiary Education Trust Fund (TET-Fund) of the Federal Government of Nigeria. I also wish to express my appreciation to the management and staff of University Teknologi Malaysia and Universiti Malaysia Terengganu for their collaborative support. All experiments on bacteria inactivation were performed at the Food Microbiology Laboratory of Universiti Malaysia Terengganu. My sincere thanks go to the staff and students of the above laboratory for their tremendous support.

Finally, I wish to thank some of my family members, colleagues, friends and kindred for the various roles they played to facilitate my study. Amongst many of them are Dr Bello Yusuf Idi, Dr Hamman Tukur Gabdo, Mr Markus Linah Ndenah, Pastor Nuhu Ndwakan, Mr Linus Gubbi, Mr Manliura Philemon Datilo, Comrade Jeremiah Ngyakwar and Mr Mohammed Mahfuz. I truly appreciate your support.

ABSTRACT

Ultraviolet (UV) lasers have been employed for many applications such as in the food industry, medicine and dentistry. The use of laser and other pulsed light systems for phototherapy and other microbial disinfections are based on spectral characteristics and configuration of the light used. The objective of this study was to use a flash lamp-pumped passive Q-switched Nd:YAG laser for inactivation two pathogenic bacteria, namely Escherichia coli and Listeria monocytogenes. A 1064 nm pulsed infrared (IR) laser and its 2nd and 3rd harmonics were used to irradiate Escherichia coli at various values of fluence. A 350 nm continuous wave (CW) UV lamp was also used in the inactivation process for the purpose of comparison with the 3rd harmonic (355 nm) pulsed laser. The result indicates that there is a statistical significant difference between mean \log_{10} reductions for the three laser wavelengths. The mean \log_{10} reductions for the 355 nm pulsed (UV) laser are higher than the corresponding mean log₁₀ reductions for the CW UV light. A t-test conducted on the mean log₁₀ reductions obtained for the pulsed UV laser and the CW UV light indicates that there is a significant difference between the two sets of mean log₁₀ reductions. When E. coli and L. monocytogenes samples were irradiated with the pulsed UV laser at three different pulse frequencies, the result shows higher inactivation effect at higher pulse frequency than at lower pulse frequency. Statistical analysis, using two-way ANOVA, shows that the mean log₁₀ reductions for the three pulsed frequencies were significantly different. However, no statistical significant difference was observed between mean log₁₀ reductions obtained for treatment with the pulsed UV laser and the CW UV light on three different sample volumes of E. coli and L. monocytogenes.

ABSTRAK

Laser lampau ungu (UV) telah digunakan untuk pelbagai aplikasi seperti dalam industri makanan, perubatan dan pergigian. Penggunaan laser dan sistem denyutan cahaya lain untuk fototerapi dan disinfeksi mikrob lain adalah berdasarkan ciri-ciri spektrum dan konfigurasi cahaya yang digunakan. Objektif kajian ini adalah untuk menggunakan laser Nd:YAG suis-Q pasif dipam lampu kilat bagi menyahaktifkan dua bakteria patogenik, iaitu Escherichia coli dan Listeria monocytogenes. Laser denyutan inframerah (IR) 1064 nm dan harmoniknya yong ke-2 dan ke-3 telah digunakan untuk menyinari Escherichia coli pada pelbagai nilai dos tenaga. Lampu UV gelombang selanjar (CW) 350 nm juga digunakan dalam proses menyahaktif bagi tujuan perbandingan dengan denyutan laser harmonik ke-3 (355 nm). Keputusan menunjukkan bahawa terdapat perbezaan statistik yang signifikan antara pengurangan log₁₀ purata bagi tiga panjang gelombang laser. Pengurangan log₁₀ untuk 355 nm laser denyut (UV) adalah lebih tinggi daripada pengurangan log₁₀ purata sepadan untuk gelombang cahaya UV CW. *Ujian-t* telah dijalankan ke atas pengurangan log₁₀ purata diperoleh bagi laser denyut UV dan cahaya UV CW menunjukkan bahawa terdapat perbezaan yang signifikan antara kedua-dua set pengurangan log₁₀ purata. Apabila sampel *E. coli* L. monocytogenes telah disinarkan dengan laser denyut UV pada tiga frekuensi denyutan yang berbeza, keputusan menunjukkan kesan menyahaktif lebih tinggi pada frekuensi denyut tinggi daripada frekuensi denyut lebih rendah. Analisis statistik, menggunakan dua-cara ANOVA, menunjukkan bahawa pengurangan log₁₀ purata bagi tiga frekuensi denyutan adalah jauh berbeza. Walau bagaimanapun, tiada perbezaan statistik yang signifikan telah diperhatikan antara pengurangan log₁₀ purata diperoleh bagi rawatan dengan laser denyut UV dan cahaya UV CW pada tiga jilid sampel yang berbeza E. coli dan L. monocytogenes.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xi
	LIST OF FIGURES	xiii
	LIST OF ABBREVIATIONS	xvi
	LIST OF SYMBOLS	xviii
	LIST OF APPENDICES	XX
1	INTRODUCTION	1
	1.1 Background	1
	1.2 Statement of problem	6
	1.3 Objectives	7
	1.4 Scope of study	7
	1.5 Significance of study	8
	1.6 Original contributions of this study	8
	1.7 Thesis structure and organization	9

2	LIT	TERATURE REVIEW	10
	2.1	Overview	10
	2.2	The Nd:YAG laser	10
	2.3	Non-linear optical conversion	12
		2.3.1 Sum frequency generation	12
		2.3.2 Model equations	15
	2.4	Generation of harmonics within the UV range using	
		non-linear optics	17
	2.5	Inactivation of Bacteria	19
	2.6	Pulsed light systems	21
		2.6.1 Source	22
		2.6.2 Target	22
		2.6.3 Chamber	23
	2.7	Principle of pulsed light	23
	2.8	Pulsed light parameters and microbial inactivation	24
		2.8.1 Spectral distribution	25
		2.8.2 Fluence	25
		2.8.3 Pulse duration	28
		2.8.4 Pulse frequency	29
		2.8.5 Number of pulses applied	29
		2.8.6 Target parameters	30
	2.9	Mechanism of bacterial inactivation using pulsed light	30
		2.9.1 Role of the	
		-violet (UV) spectrum	32
		2.9.2 Continuous flow UV light versus pulsed UV light	38
		2.9.3 Pulsed laser	39
		2.9.4 Prospects and challenges	40

	2.10 Inactivation of Escherichia coli and Listeria monocytogenes	
	using pulsed light	42
	2.11 Enumeration of bacteria	45
	2.11.1 Measures of inactivation	46
	2.12 Summary	47
3	METHODOLOGY	49
	3.1 Overview	49
	3.2 Generation of second and third harmonics of 1064 nm laser	
	using non-linear optics	51
	3.2.1 Materials	52
	3.2.2 Procedure	55
	3.3 Inactivation of bacteria	57
	3.3.1 Preparation of media and agar plates	57
	3.3.2 Sample preparation	58
	3.3.3 Sources of radiation	63
	3.3.4 General experimental set up	65
	3.3.5 Serial dilution and colony counting	70
	3.3.6 Statistical analysis	71
	3.4 Summary	72
4	RESULTS AND DISCUSSIONS	73
	4.1 Introduction	73
	4.2 Generation of harmonics	73
	4.3 Inactivation of bacteria	80
	4.3.1 Log ₁₀ reductions for different laser wavelengths and	
	different fluence	81

	4.3.2 Log ₁₀ reductions of Total Viable Count of <i>E. coli</i> and	1
	L. monocytogenes at different pulse frequencies	89
	4.3.3 Log ₁₀ reductions of Total Plate Count of <i>E. coli</i> and	
	L. monocytogenes at for different sample volumes	94
	4.4 Summary	96
5	CONCLUSION	97
	5.1 Conclusion	97
	5.2 Recommendations	98
REFERE	NCES	99
Appendice	es A-D	09 -155

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Some reported non-linear optical conversions for the UV range	18
2.2	Parameters for calculation of output parameters of pulsed laser (modified from Simrock (2013))	27
2.3	Ultraviolet light effects on living cell (Modified from Beltrl and Barbosa (2004))	33
2.4	Typical bond energies of important biological moieties and their corresponding wavelengths (Smith, 2013)	34
2.5	Characteristic wavelengths and corresponding photon energies of radiation within UV and visible portions of the electromagnetic spectrum (Blatchley and Peel, 2001)	
2.6	Summary of UV sources and their basic characteristics (Modified from Blatchley and Peel, 2001)	37
2.7	Some reported inactivation of <i>Escherichia coli</i> using Xenon flash lamp	43
2.8	Some reported inactivation of <i>Listeria monocytogenes</i> using Xenon flash lamp	44
3.1	Some properties of crystals used for generation of harmonics (Castech, 2013)	53
3.2	Media used for experimental study of bacteria inactivation	58

3.3	Laser and UV light parameters used for treatment	65
4.1	Mean values for pulse energies of fundamental $(E_{\omega o})$, second harmonic $(E_{2\omega o})$ and third harmonic $(E_{3\omega o})$ with the corresponding input voltages (V_i) and electrical input energies (E_i)	74
4.2	Mean energy conversion ratios	77
4.3	Log ₁₀ reductions of <i>E. coli</i> treated with PIRLa, PGLa, PUVaLa and CWUVaLi at four values of total fluence	82
4.4	Result of two-way ANOVA for log_{10} reductions of $E.\ coli$ at different values of fluence for three laser wavelengths	84
4.5	Result for t - t es t for log_{10} reductions of E . $coli$ due to treatments with PUVaLa and CWUVaLi	89
4.6	Log ₁₀ reductions for <i>E. coli</i> and <i>L. monocytogenes</i> treated with PUVaLa at pulse frequencies of 1 Hz, 3 Hz and 5 Hz for four values of treatment time	90
4.7	Result for two-way ANOVA for log_{10} reductions of <i>E. coli</i> and <i>L. monocytogenes</i> at three pulse frequencies for different treatment times	93
4.8	Log ₁₀ reductions of <i>E. coli</i> and <i>L. monocytogenes</i> treated with PUVaLa and CWUVaLi for three values of sample volume	94
4.9	Result for two-way ANOVA for log_{10} reductions of <i>E. coli</i> and <i>L. monocytogenes</i> treated with PUVaLa and CWUVaLi	
	for three sample volumes	96

LIST OF FIGURES

FIGURE NO	TITLE	PAGE
1.1	The electromagnetic spectrum (modified from Shapley (2015))	2
1.2	Divisions of the UV portion of the electromagnetic spectrum showing wavelengths in nm	4
2.1	Schematic setup of a passively Q-switched Nd:YAG laser resonator	11
2.2	Schematic of experimental arrangement for third harmonic generation (THG)	15
2.3	Block diagram of a pulsed light system for inactivation of microbial organisms	22
2.4	Block diagram of a pulsed UV system (adapted from Yazdi and Darghahi (2006))	24
2.5	Intercalation of pathogen reduction agent (Bryant, 2007)	31
2.6	Scanning electron photomicrograph of <i>Escherichia coli</i> (A), <i>Listeria monocytogenes</i> (B) and <i>Salmonella typhimurium</i> (C) treated in a UV-assisted TiO ₂ -PCO reaction (Kim, <i>et al.</i> , 2013)	
3.1	Work flow chart for the study	50
3.2	Passive Q-switched Nd:YAG laser panel (500-1000 mJ,	
	6 ns)	52

3.3	Picture of constructed crystal holder fitted with BBO crystal	
	for generation of third harmonic: (a) Posterior view, (b) Anterior view	54
3.4	The Nd: YAG laser fitted with constructed crystal holder	54
3.5	Picture of power meter used (MILLES CRIOT)	55
3.6	Picture of spectrometer used (ASEQ Instruments Model LR1)	55
3.7	Schematic of experimental set-up to measure energies of the fundamental (1064 nm), the second harmonic (532 nm) and the third harmonic (355 nm) of pulsed IR laser	56
3.8	Picture of the experimental setup for generation of harmonics	57
3.9	An overview of sample preparation process	60
3.10	Pictures of some of the equipment used during sample preparation: (a) Incubator (Memmert); (b) Autoclave machine (Hirayama HVE-50); (c) Centrifuge machine (Grozen 1580R); (d) Colony counter (Funke Gerber); (e) Automatic spiral plating machine (Interscience® easy spiral)	61
3.11	Pair colony plates of the 2^{nd} and 3^{rd} decimal dilutions of <i>E. coli</i> sample	62
3.12	Pair colony plates of the 2^{nd} and 3^{rd} decimal dilutions of <i>L. monocytogenes</i> sample	62
3.13	Picture of UV lamp used for the study	64
3.14	Schematics of experimental setup	66
3.15	Picture of actual experimental setup showing the container for (a) laser treatment; (b) continuous wave light treatment; (c) control	67
3.16	Schematic overview of general experimental procedure	68

4.1	Pulse energies of the fundamental wavelength $E_{\omega o}$, $2^{\rm nd}$	
	harmonic $E_{2\omega o}$ and $3^{\rm rd}$ harmonic $E_{3\omega o}$ versus the input electrical energy of the laser E_i	75
4.2	Pulse energies of 2 nd and 3 rd harmonics versus pulse energy of the fundamental wavelength.	76
4.3	Pulse energy of 3 rd harmonic versus pulse energy of the 2 nd harmonic.	77
4.4	Output spectrum after KDP	79
4.5	Output spectrum after BBO	80
4.6	Histogram of \log_{10} reduction versus fluence for <i>E. coli</i> treated with three laser wavelengths and a continuous wave UV-A light	83
4.7	Log ₁₀ reduction of E . $coli$ versus fluence treated with three laser wavelengths and a continuous wave UV-A light	84
4.8	Box plots of log_{10} reductions for of E . $coli$ treated with three laser wavelengths and continuous wave UV-A light	87
4.9	Histogram of \log_{10} reduction versus treatment time for <i>E. coli</i> and <i>L. monocytogenes</i> treated with Pulsed UVaLa at three pulsed frequencies	91
4.10	Inactivation of <i>E. coli</i> and <i>L. monocytogenes</i> at three pulsed frequencies of PUVaLa	92
4.11	Histogram of \log_{10} reduction versus sample volume for <i>E. coli</i> and <i>L. monocytogenes</i> treated with PUVaLa and	
	CWUVaLi	95

LIST OF ABBREVIATIONS

Abbreviation Meaning

AC - Alternating Current

ANOVA - Analysis of Variance

CFU - Colony Forming Units

CW - Continuous Wave

CWUVaLi - Continuous Wave UV-A Light

DC - Direct Current

df - Degree of Freedom

DFG - Difference-Frequency Generation

DNA - Deoxyribonucleic Acid

DVD - Digital Versatile Disc

FWHM - Full-Width Half-Maximum

IPL - Intense Pulsed Light

IR - Infrared

LR - Log₁₀ Reduction

M - Mean

MS - Mean Squared

NA - Nutrient Agar

NB - Nutrient Broth

PGLa - Pulsed Green Laser

PIRLa - Pulsed Infrared Laser

PK - Percentage Kill

PLS - Pulsed Light System

PUVaLa - Pulsed UV-A Laser

SE - Standard Error

SF - Survival Fraction

SFG - Sum-Frequency Generation

SHG - Second Harmonic Generation

SPC - Standard Plate Count

SS - Sum of Squared

STA - Spectrophotometric Turbidimetric Analysis

THG - Third Harmonic Generation

TPC - Total Plate Count

TSA - Tryptic Soy Agar

TVC - Total Viable Count

USFDA - United States Food and Drug Administration

UV - Ultraviolet

VPC - Viable Plate Count

LIST OF SYMBOLS

Symbol		Meaning
A	-	Area
c	-	Speed of light
C	-	Capacitance
D	-	Duty cycle
$d_{\it eff}$	-	Effective nonlinear coefficient
D_{FI}	-	Decimal reduction fluence
\boldsymbol{E}	-	Electric field
E_p	-	Pulse energy
F	-	Fluence
f, ν	-	Frequency
h	-	Planck constant
I	-	Intensity
L	-	Length
n	-	Refractive index
N	-	Number of the pulses
P	-	Power
P_{NL}	-	Non-linear polarization
R	-	Repetition rate
T	-	Time
V	-	Voltage
x_R	-	Rayleigh range
δ	-	Dephasing factor
Δk	-	Phase mismatch

η	-	Conversion efficiency
λ	-	Wavelength

ho - Walk-off angle

au - Pulse duration

 χ^2 - Second order susceptibility

 ω - Angular frequency

LIST OF APPENDIXES

APPENDIX	TITLE	PAGE
A	How total fluence is obtained from other pulse light parameter	rs 110
В	Colony counting method for interscience easy automatic	
	spiral platers	111
C	Details of data obtained from inactivation of bacteria	115
D	Publications from this study	153

CHAPTER 1

INTRODUCTION

1.1 Background

Ever since the first laser was constructed by Theodore Maiman in 1960 (Koechner, 2006), studies aimed at exploring the nature and expanding the applicability of lasers has been continued by scientists across the globe. The ruby (or Maiman) laser was of the red visible light radiation (694.3 nm) with pulse-duration of the order of milliseconds, pulse energy of about 1 J and an average power of the order of kilowatts (Abramczyk, 2005). Progressively, other types of laser were developed and today we have gas lasers, liquid lasers, solid state lasers and semiconductor lasers, among others. Laser lights are now being produced for virtually the entire visible range of the electromagnetic spectrum (Figure 1.1) and even beyond (up to the x-ray region), with pulse-duration reaching the order of attosecond (10⁻¹⁸ s) and an attainable average peak power of zetawatts (10²¹ W) (Koechner, 2006; Milonni and Eberly, 2010).

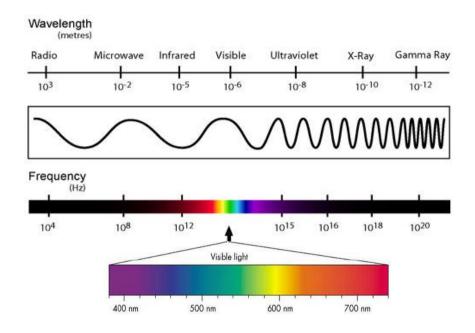


Figure 1.1 The electromagnetic spectrum (modified from Shapley (2015))

The original energy or power of laser lights obtained from an oscillator is usually too low for most applications. Therefore, it needs further amplification for it to be useful for such applications. Output pulse energies from femtosecond lasers typically do not exceed a few nanojoules, and peak powers of megawatts (Abramczyk, 2005). Hence, laser or pulse amplifiers are employed to raise output power to useful levels required for various applications. In seeking methods to shorten pulses and to increase peak powers and peak intensities on intended targets, researchers often trade-off various design and input parameters in order to obtain an optimize state of operation for a particular application. With pre-determined design parameters, the next option for optimization is for a researcher to manipulate the input parameters.

Applications of lasers vary as widely as their range of the wavelengths, pulsewidth or peak power. Applications are found in industries (e.g. laser machining, laser cutting or drilling), medicine (e.g. dentistry, neural network, dermatology, and surgery), home or office appliances (e.g. laser printer, laser scanner and laser pointer), research laboratories (e.g. spectrometry and diffraction experiments), military (e.g. laser guided smart bombs and laser blinders), communication (e.g. laser fibers), etc (Diels and Arissian, 2011). Lasers are also important parts of common devices such as bar-code scanners used in supermarkets, compact disc systems, and digital versatile disc (DVD) players.

In the health sector, laser irradiation has been adopted as a technique for curing or controlling various bacteria-related diseases in medicine and dentistry (Andrade *et al.*, 2008; Franzen *et al.*, 2011; Nandakumar *et al.*, 2003; Schoenly *et al.*, 2012). The use of lasers in these and other health-related fields depends on the absorption ability of interacting targets or media. The power and wavelength of beam, in addition to the absorption properties of the interacting biological tissue determine the depth of penetration of a laser beam (Abramczyk, 2005; Diels and Arissian, 2011). For bacteria and similar microbes, absorption of radiation is said to depend on the wavelength of the incident radiation, with the ultraviolet (UV) region providing the most effective absorption when compared to other regions of the electromagnetic spectrum.

The UV laser finds applications in diverse fields such as in material processing (Ya et al., 2009b; Zhai et al., 2013), lithography (Ya, et al., 2009b; Yang et al., 2009), various spectroscopic techniques (Franzke et al., 1998; Suzuki et al., 2008; Ya, et al., 2009b; Yang, et al., 2009), medicine (Ya, et al., 2009b), and others. One area in which lasers are put into applications that are more closely related to human systems is in the interaction of lasers with bio-samples. Studies in this area are often fashioned out in such a way that most of the resulting applications are tailored towards medicine, surgery or dentistry. Practically all of these applications are for curative purpose.

The UV spectrum is the portion of the electromagnetic spectrum between visible light and X-rays, with wavelength ranging from about 10 nm to about 400

nm. Depending on its spectral properties and applications, the UV spectrum is further subdivided into four regions as shown in Figure 1.2. Another classification mode of the UV spectrum refers to the longer wavelengths greater than 200 nm as near-UV while those below 200 nm are called far-UV.

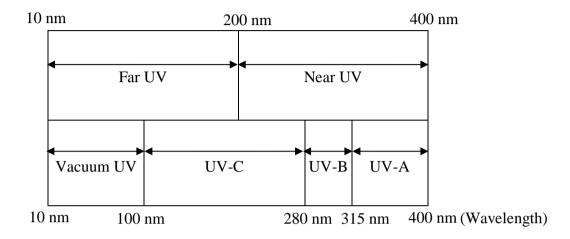


Figure 1.2 Divisions of the UV portion of the electromagnetic spectrum showing wavelengths in nm

Although the broad spectrum from infrared to UV can be used for microbial inactivation, most of the germicidal effects have been attributed to the UV region, particularly the UV-C. Several documented studies (Daryany *et al.*, 2009; Bohrerova *et al.*, 2008; Farrell *et al.*, 2011; Marquenie *et al.*, 2003; Matafonova and Batoev, 2012; Villarroel *et al.*, 2012a) indicate that UV light can be used for various decontamination and sterilization processes. Specifically, pulsed UV light has been reported to be more efficient in inactivation of bacteria and other microbial organisms than continuous wave UV light (Bohrerova, *et al.*, 2008; Cheigh *et al.*, 2012; Farrell, *et al.*, 2011; Hierro *et al.*, 2011). The transmission electron micrographs of intense pulsed light (IPL) and UV-C-induced cell damage observed by Cheigh, *et al.* (2012) indicate that bacterial cell structures were destroyed by IPL treatment but not by UV-C treatment. Lethal effects of pulsed light on microbes is said to also depend on the fluence incident on the sample, the composition of the

emitted light spectrum and the distance of the sample from the light source (Artíguez *et al.*, 2011). Other factors that affect microbial inactivation in liquid samples are thickness, colour, opacity, viscosity, product flow conditions and presence of particulate material (Pataro *et al.*, 2011).

By the nature of pulsed laser, it should have more power than an ordinary continuous wave light because of its coherence and directionality. Hence, it is premised that the interaction of pulsed UV laser with bacteria should have more lethal effect than continuous wave UV light of similar output configuration. However, it was stated (López *et al.*, 2007) that there are no independent experimental reports to confirm some claims of pulse light having more penetrating power than continuous wave light. Studying the effects of parameters of pulsed laser, such as, fluence, spectral distribution and pulse frequency on inactivation of bacteria in comparison to result obtained for continuous wave UV light could verify the veracity of this supposition. So far, most of the studies conducted using laser for either direct (Dinu *et al.*, 2002; Nandakumar, *et al.*, 2003) or indirect (Andrade, *et al.*, 2008; Baudelet *et al.*, 2009; Franzen, *et al.*, 2011; Schoenly, *et al.*, 2012) microbial inactivation were aimed at applications in dentistry and medicine, with emphasis on roles played by laser wavelength, pulse power and pulse width.

Apart from wavelength, other parameters of pulsed laser, like pulse width and pulse frequency, could as well play some roles in bacteria inactivation process for a particular wavelength of a pulsed laser. This may have some implications for optimization of lasers or other pulsed light systems to maximise performance. This study is designed to further explore the potential of pulsed UV laser light as a means of inactivation of bacteria in liquid medium by looking into roles played by laser fluence and pulse frequency. The two pathogenic bacteria chosen for this study have been known to cause either food poisoning or food spoilage. Certain strains of *Escherichia coli* are known to cause diarrhoea, resulting from intake of contaminated water or food. In addition, *Listeria monocytogenes* is the cause of listeriosis, also resulting from eating contaminated food. The food poisoning resulting from the actions of these bacteria could lead to illnesses of epidemic proportion. The result of

food spoilage could lead to economic losses from both production companies and retailers.

1.2 Statement of problem

Non-thermal methods of inactivating bacteria are being sought because such methods leave less damaging effects on processed materials such as food and surgical materials. Heat treatment of foods at high temperature can affect texture, flavour and appearance of the product whereas less severe thermal treatment may result to inadequate decontamination (Maktabi et al., 2011). Pasteurization of food materials and other methods of thermal sterilization also results into rise in temperature in the bulk of the material, which may be undesirable. Using chemical disinfection is effective, but it may also leave behind residual by-products, which may be toxic or be mutagenic (Daryany et al., 2008). Previous studies conducted indicate the potentiality of pulsed light, especially of the UV type (Bohrerova, et al., 2008; Cheigh, et al., 2012; Farrell, et al., 2011; Hierro, et al., 2011), for inactivation of bacteria. However, a basic limitation in the use of pulsed light is in the depth of penetration of light in the materials being processed, especially coloured liquids. Depth of penetration is usually limited by the power of the pulsed light in addition to the level of transparency of the sample material being processed. Therefore, pulsed UV lasers, which are narrow-banded and provide high-intensity emissions, with desirable penetration depth in water and other fluids may be potentially more effective (Daryany, et al., 2008). This study was designed to look into the effectiveness of pulsed UV laser in decontamination and sterilization, especially with regards to any possible roles by the laser wavelength, pulse frequency and sample depth, in liquid sample. Determination of prime values for laser parameters, at which inactivation is optimal, may lead to efficient use of laser systems for the purpose of bacteria inactivation in liquid food items.

1.3 Objectives

The main objective of the study was to use a flash lamp-pumped passive Q-switched Nd:YAG laser for inactivation of selected pathogenic bacteria. This main objective was pursued through the following sub-objectives:

- (i) To modulate the output of the Nd:YAG laser from IR to its 3rd harmonic in the UV region by using non-linear crystals.
- (ii) To examine the effectiveness of the 3rd harmonic of the Nd:YAG laser in inactivation of two selected pathogenic bacteria (*Escherichia coli* and *Listeria monocytogenes*).
- (iii) To observe the effect of pulse frequency (or pulse repetition rate) of the 3rd harmonic of the Nd:YAG laser in the process of the bacteria inactivation.
- (iv) To observe the effect of sample volume in the process of inactivation of bacteria with the 3rd harmonic of the Nd:YAG laser.

1.4 Scope of study

A flash lamp-pump Q-switched Nd:YAG laser with pulse duration of 6 ns was employed for the study. The pulse energy of the laser can be varied in the range 0-1000 mJ. The pulse repetition is in the range of 1-5 Hz, in steps 1 Hz. The laser wavelengths used for the study are 1064 nm, 532 nm and 355 nm. A 350 nm continuous wave UV lamp, rated 4 W, was also used for the study for the purpose of

comparison. Two species of pathogenic bacteria, *Escherichia coli* (ATCC 11775) and *Listeria monocytogenes* (ATCC 7645), obtained from Institute of Bio-Information Technology, Universiti Selangor, were the samples used for the study. The bacteria samples used are common pathogenic bacteria associated with food poisoning. All experiments were conducted under ambient conditions of room temperature and atmospheric pressure.

1.5 Significance of study

Study of the interaction of lasers with microbial organisms (particularly the pathogenic types) leads to applications such as food preservation, food safety, decontamination of immediate environment, sterilization of equipment, etc; which are preventive rather than curative. Hence, possible application from this study is in the development of efficient point-of-use UV laser devices for decontamination and sterilization for use at homes, hospitals, industries and work places.

1.6 Original contributions of this study

The following are some of the academic contributions derivable from this study:

➤ The study compares the effects of three pulsed laser wavelengths in inactivation of *Escherichia coli* and it was found that 355 nm pulsed laser was more efficient in the inactivation process than 532 nm and 1064 nm pulsed lasers.

- ➤ The effectiveness of inactivation of the 355 nm pulsed laser was compared with that of a 350 nm continuous wave UV light of close output configuration and it was found that the 355 nm laser was more efficient in the inactivation of *Escherichia coli*.
- ➤ The study also reveals that inactivation of both *Escherichia coli* and *Listeria monocytogenes* is more efficient at higher laser pulse frequency than at lower frequency.

1.7 Thesis structure and organization

This thesis is composed of five chapters. Chapter 1 gives a general background of the study followed by problem statement, objectives of study, scope of study, significance of study and contributions of study, in that order. In chapter 2, literature related to the study was reviewed. This includes review of some past studies done on harmonic generation using non-linear optics and microbial inactivation with pulsed light. Relevant formulae for the study are also highlighted. Chapter 3 describes the methodology employed for the study which includes list of materials used, a brief description of experimental procedures and mode of data analysis. Details of the experimental results obtained as well as analysis of results for harmonic generation by non-linear optical conversion and interaction of laser with samples studied are presented in chapter 4. Chapter 5 is the conclusion and recommendations.

REFERENCES

- Abramczyk, H. (2005). *Introduction to Laser Spectroscopy*. (10.1016/b978-044451662-6/50007-1) Amsterdam: Elsevier Science.
- Andrade, A., Feist, I., Pannuti, C., Cai, S., Zezell, D. and Micheli, G. (2008).
 Nd:YAG laser clinical assisted in class II furcation treatment. *Lasers in Medical Science*. 23(4), 341-347.
- Artíguez, M. L., Lasagabaster, A. and Marañón, I. M. d. (2011). Factors affecting microbial inactivation by Pulsed Light in a continuous flow-through unit for liquid products treatment. *Procedia Food Science*. 1(0), 786-791.
- Artíguez, M. L. and Martínez de Marañón, I. (2015). Improved process for decontamination of whey by a continuous flow-through pulsed light system. Food Control. 47(0), 599-605.
- Asnaashari, M. and Safavi, N. (2013). Disinfection of contaminated canals by different laser wavelengths, while performing root canal therapy. *Journal of Lasers in Medical Sciences*. 4(1), 8-16.
- Bandla, S., Choudhary, R., Watson, D. G. and Haddock, J. (2012). UV-C treatment of soymilk in coiled tube UV reactors for inactivation of Escherichia coli W1485 and Bacillus cereus endospores. *LWT Food Science and Technology*. 46(1), 71-76.
- Bank, H. L., John, J., Schmehl, M. K. and Dratch, R. J. (1990). Bactericidal effectiveness of modulated UV light. Applied and Environmental Microbiology. 56(12), 3888-3889.
- Baudelet, M., Boueri, M., Yu, J., Mao, X., Mao, S. S. and Russo, R. (2009). Laser ablation of organic materials for discrimination of bacteria in an inorganic background. 10.1117/12.808485, 72140J-72140J.

- Beltrl, J. A. G. and Barbosa, G. V. C. (2004). Advantages and Limitations on Processing Foods by UV Light. Food Science and Technology International. 10(3), 137-147.
- Bennett, R. and Lancette, G. (2001). Chapter 15 Bacillus cereus Diarrheal Enterotoxin *Bacteriological analytical manual* USFDA.
- Biswas, R., Kuar, A., Sarkar, S. and Mitra, S. (2010). A parametric study of pulsed Nd: YAG laser micro-drilling of gamma-titanium aluminide. *Optics & Laser Technology*. 42(1), 23-31.
- Blatchley, E. R. and Peel, M. M. (2001). Disinfection by ultraviolet irradiation. In Block, S. S. (Ed.) *Disinfection, sterilization, and preservation* (pp. 823 829). Philadealphia, USA: Lippincott Williams and Wilkins.
- Bohrerova, Z., Shemer, H., Lantis, R., Impellitteri, C. A. and Linden, K. G. (2008). Comparative disinfection efficiency of pulsed and continuous-wave UV irradiation technologies. *Water Research*. 42(12), 2975-2982.
- Botschafter, E. M., Schiffrin, A., Yakovlev, V. S., Azzeer, A. M., Krausz, F., Ernstorfer, R. and Kienberger, R. (2010). Collinear generation of ultrashort UV and XUV pulses. *Optics Express*. 18(9), 9173-9180.
- Brinkmann, M., Hayden, J., Letz, M., Reichel, S., Click, C., Mannstadt, W., Schreder, B., Wolff, S., Ritter, S., Davis, M. J., Bauer, T. E., Ren, H., Fan, Y.-H., Menke, Y., Wu, S.-T., Bonrad, K., Krätzig, E., Buse, K. and Paquin, R. A. (2012). Optical Materials and Their Properties. In Träger, F. (Ed.) *Springer Handbook of Lasers and Optics* (10.1007/978-3-642-19409-2_5pp. 253-399)Springer Berlin Heidelberg.
- Bryant, B. J. and Klein, H. G. (2007). Pathogen inactivation: the definitive safeguard for the blood supply. *Archives of pathology & laboratory medicine*. 131(5), 719-733.
- Brygo, F., Dutouquet, C., Le Guern, F., Oltra, R., Semerok, A. and Weulersse, J. M. (2006). Laser fluence, repetition rate and pulse duration effects on paint ablation. *Applied Surface Science*. 252(6), 2131-2138.
- Buchovec, I., Paskeviciute, E. and Luksiene, Z. (2010). Photosensitization-based inactivation of food pathogen Listeria monocytogenes in vitro and on the surface of packaging material. *Journal of Photochemistry and Photobiology B: Biology*. 99(1), 9-14.

- Castech (2013). Crystal catalog (2012-2013). Available from http://www.castech.com/ downloadRepository/3c74413d-f890-49db-9f75a9fb49193cf5.pdf>. [2 April 2015].
- Cheigh, C. I., Hwang, H. J. and Chung, M. S. (2013). Intense pulsed light (IPL) and UV-C treatments for inactivating Listeria monocytogenes on solid medium and seafoods. *Food Research International*. 54(1), 745-752.
- Cheigh, C. I., Park, M. H., Chung, M.-S., Shin, J. K. and Park, Y. S. (2012). Comparison of intense pulsed light- and ultraviolet (UVC)-induced cell damage in Listeria monocytogenes and Escherichia coli O157:H7. *Food Control*. 25(2), 654-659.
- Chen, C. (2004). Recent advances in deep and vacuum-UV harmonic generation with KBBF crystal. *Optical Materials*. 26(4), 425-429.
- Chen, W.-B. and Zhang, C. (2009). An automated bacterial colony counting and classification system. *Information Systems Frontiers*. 11(4), 349-368.
- Choi, M. S., Cheigh, C. I., Jeong, E. A., Shin, J. K. and Chung, M.-S. (2010). Nonthermal sterilization of Listeria monocytogenes in infant foods by intense pulsed-light treatment. *Journal of Food Engineering*. 97(4), 504-509.
- Daryany, M. K. A., Hosseini, S. M., Raie, M., Fakharie, J. and Zareh, A. (2009). Study on continuous (254 nm) and pulsed UV (266 and 355 nm) lights on BVD virus inactivation and its effects on biological properties of fetal bovine serum. *Journal of Photochemistry and Photobiology B: Biology*. 94(2), 120-124.
- Daryany, M. K. A., Massudi, R. and Hosseini, M. (2008). Photoinactivation of Escherichia coli and Saccharomyces cerevisiae suspended in phosphate-buffered saline-A using 266- and 355-nm pulsed ultraviolet light. *Current Microbiology*. 56(5), 423-428.
- Diels, J. C. and Arissian, L. (2011). Lasers: The Power and Precision of Light. (10.1002/9783527640034.ch3)Wiley-VCH Verlag GmbH & Co. KGaA.
- Dinu, C. Z., Grigoriu, C., Dinescu, M., Pascale, F., Popovici, A., Gheorghescu, L., Cismileanu, A. and Avram, E. (2002). Laser radiation effects on Mycoplasma agalactiae. 10.1117/12.478657, 343-348.
- Falkenstein, Z. (2001). Development of an excimer UV light source system for water treatment. *Ushio America*. 1-6

- Farkas, B. and Geretovszky, Z. (2006). On determining the spot size for laser fluence measurements. *Applied Surface Science*. 252(13), 4728-4732.
- Farrell, H., Hayes, J., Laffey, J. and Rowan, N. (2011). Studies on the relationship between pulsed UV light irradiation and the simultaneous occurrence of molecular and cellular damage in clinically-relevant Candida albicans. *Journal of Microbiological Methods*. 84(2), 317-326.
- Franz, C. M. A. P., Specht, I., Cho, G.-S., Graef, V. and Stahl, M. R. (2009). UV-C-inactivation of microorganisms in naturally cloudy apple juice using novel inactivation equipment based on Dean vortex technology. *Food Control*. 20(12), 1103-1107.
- Franzen, R., Gutknecht, N., Falken, S., Heussen, N. and Meister, J. (2011). Bactericidal effect of a Nd:YAG laser on Enterococcus faecalis at pulse durations of 15 and 25 ms in dentine depths of 500 and 1,000 μm. *Lasers in Medical Science*. 26(1), 95-101.
- Franzke, J., Fox, R. W. and Hollberg, L. (1998). Tunable UV generation at 283 nm by frequency doubling and sum frequency mixing of two semiconductor lasers for the detection of Pb. *Spectrochimica Acta Part B: Atomic Spectroscopy*. 53(14), 1951-1955.
- Gayán, E., Monfort, S., Álvarez, I. and Condón, S. (2011). UV-C inactivation of Escherichia coli at different temperatures. *Innovative Food Science & Emerging Technologies*. 12(4), 531-541.
- Geeraerd, A. H., Valdramidis, V. P. and Van Impe, J. F. (2005). GInaFiT, a freeware tool to assess non-log-linear microbial survivor curves. *International Journal of Food Microbiology*. 102(1), 95-105.
- Grubsky, V. and Feinberg, J. (2007). Phase-matched third-harmonic UV generation using low-order modes in a glass micro-fiber. *Optics Communications*. 274(2), 447-450.
- Guffey, J. S., Payne, W., Jones, T. and Martin, K. (2013). Evidence of resistance development by Staphylococcus aureus to an in vitro, multiple stage application of 405 nm light from a supraluminous diode array. *Photomedicine and Laser Surgery*. 31(4), 179-182.
- Gyu, A. D. and Won, J. G. (2009). Influence of process parameters on drilling characteristics of Al 1050 sheet with thickness of 0.2 mm using pulsed Nd:

- YAG laser. Transactions of Nonferrous Metals Society of China. 19, s157-s163.
- Hamilton, M. A. (2010). The Log-Reduction Measure of Disinfectant Efficacy *KSA SM-07*. Available from Centre for Biofilm Engineering, Montana University website: http://www.biofilm.montana.edu/files/CBE/ documents/KSA-SM-07.pdf>. [9 October 2013].
- Hamilton, M. A. (2011). Testing surface disinfectants: quantitative, semi-quantitative, quantal and alternative methods *KSA-SM-02*. Available from Centre for Biofilm Engineering, Montana University website: http://www.biofilm.montana.edu/files/CBE/documents/KSA-SM-07.pdf>.[10 April 2015]
- Hierro, E., Barroso, E., la Hoz, L. d., Ordóñez, J. A., Manzano, S. and Fernández, M. (2011). Efficacy of pulsed light for shelf-life extension and inactivation of Listeria monocytogenes on ready-to-eat cooked meat products. *Innovative Food Science & Emerging Technologies*. 12(3), 275-281.
- Hijnen, W., Beerendonk, E. and Medema, G. J. (2006). Inactivation credit of UV radiation for viruses, bacteria and protozoan (oo) cysts in water: a review. *Water research*. 40(1), 3-22.
- Hiramoto, T. (1984). *United States Patent No.* 4,464,336. Patent, U. S. Retrieved from http://www.patentbuddy.com/Patent/4464336
- Hong, F.-L., Ishikawa, J., Zhang, Y., Guo, R., Onae, A. and Matsumoto, H. (2004).
 Frequency reproducibility of an iodine-stabilized Nd: YAG laser at 532 nm.
 Optics communications. 235(4), 377-385.
- Hsu, L. and Moraru, C. I. (2011). Quantifying and mapping the spatial distribution of fluence inside a pulsed light treatment chamber and various liquid substrates. *Journal of Food Engineering*. 103(1), 84-91.
- Huang, F., Lou, Q., Yu, T., Dong, J., Lei, B. and Wei, Y. (2001). Tunable solid state UV laser. *Optics & Laser Technology*. 33(2), 111-115.
- Humphreys, C. (1939). Second spectrum of Xenon. J. Res. Natl. Bur. Stand.(US). 22, 19-53.
- Kim, S., Ghafoor, K., Lee, J., Feng, M., Hong, J., Lee, D. U. and Park, J. (2013). Bacterial inactivation in water, DNA strand breaking, and membrane damage

- induced by ultraviolet-assisted titanium dioxide photocatalysis. *Water Research*. 47(13), 4403-4411.
- Kimmelma, O. and Tittonen, I. (2009). Passively Q-switched Nd:YAG pumped UV lasers at 280 and 374 nm. *Optics Communications*. 282(14), 2930-2933.
- Koechner, W. (2006). *Solid-State Laser Engineering (Sixth Revised and Updated Edition*). (10.1007/0-387-29338-8_1)Springer Berlin / Heidelberg.
- Lani, M. N. (2007). Inactivation of listeria monocytogenes by Pulsed UV Illumination and Photorepair Recovery of UV-Damaged Cells. Ph D, University of Strathclyde, Glaogow, Scotland.
- Li, P., Li, D., Li, C. and Zhang, Z. (2004). Simultaneous dual-wavelength continuous wave laser operation at 1.06 μm and 946 nm in Nd: YAG and their frequency doubling. *Optics communications*. 235(1), 169-174.
- López, V. M. G., Ragaert, P., Debevere, J. and Devlieghere, F. (2007). Pulsed light for food decontamination: a review. *Trends in Food Science & Technology*. 18(9), 464-473.
- Luksiene, Z., Buchovec, I. and Paskeviciute, E. (2010). Inactivation of several strains of Listeria monocytogenes attached to the surface of packaging material by Na-Chlorophyllin-based photosensitization. *Journal of Photochemistry and Photobiology B: Biology*. 101(3), 326-331.
- Maden, M., Görgül, G., Sultan, M. N., Akça, G. and Er, Ö. (2013). Determination of the effect of Nd:YAG laser irradiation through dentinal tubules on several oral pathogens. *Lasers in Medical Science*. 28(1), 281-286.
- Maktabi, S., Watson, I. and Parton, R. (2011). Synergistic effect of UV, laser and microwave radiation or conventional heating on E. coli and on some spoilage and pathogenic bacteria. *Innovative Food Science & Emerging Technologies*. 12(2), 129-134.
- Marquenie, D., Geeraerd, A. H., Lammertyn, J., Soontjens, C., Van Impe, J. F., Michiels, C. W. and Nicolaï, B. M. (2003). Combinations of pulsed white light and UV-C or mild heat treatment to inactivate conidia of Botrytis cinerea and Monilia fructigena. *International Journal of Food Microbiology*. 85(1–2), 185-196.

- Matafonova, G. and Batoev, V. (2012). Recent progress on application of UV excilamps for degradation of organic pollutants and microbial inactivation. Chemosphere. 89(6), 637-647.
- McKenzie, K., Maclean, M., Timoshkin, I. V., MacGregor, S. J. and Anderson, J. G. (2014). Enhanced inactivation of Escherichia coli and Listeria monocytogenes by exposure to 405 nm light under sub-lethal temperature, salt and acid stress conditions. *International Journal of Food Microbiology*. 170, 91-98.
- Meggers, W. F., De Bruin, T. L. and Humphreys, C. J. (1929). The first spectrum of xenon. *Science (New York, N.Y.)*. 69(1789), 406.
- Milonni, P. W. and Eberly, J. H. (2010). *Laser Physics*. New Jersey: John Willey & Sons, Inc.
- Mironov, S., Lozhkarev, V., Ginzburg, V. and Khazanov, E. (2009). High-efficiency second-harmonic generation of superintense ultrashort laser pulses. *Applied Optics*. 48(11), 2051-2057.
- Mukhopadhyay, S., Ukuku, D. O., Juneja, V. and Fan, X. (2014). Effects of UV-C treatment on inactivation of Salmonella enterica and Escherichia coli O157:H7 on grape tomato surface and stem scars, microbial loads, and quality. *Food Control*. 44(0), 110-117.
- Muñoz, A., Caminiti, I. M., Palgan, I., Pataro, G., Noci, F., Morgan, D. J., Cronin, D. A., Whyte, P., Ferrari, G. and Lyng, J. G. (2012). Effects on Escherichia coli inactivation and quality attributes in apple juice treated by combinations of pulsed light and thermosonication. *Food Research International*. 45(1), 299-305.
- Murphy, H., Payne, S. and Gagnon, G. (2008). Sequential UV-and chlorine-based disinfection to mitigate *Escherichia coli* in drinking water biofilms. *Water research*. 42(8), 2083-2092.
- Nandakumar, K., Obika, H., Shinozaki, T., Ooie, T., Utsumi, A. and Yano, T. (2003). Laser Impact on Bacterial ATP: Insights into the Mechanism of Laser-Bacteria Interactions. *Biofouling*. 19(2), 109-114.
- Nims, R. (2010). Riboflavin plus UVA irradiation: another inactivation approach to consider. 10 June 2010. *Ray Nim*: *Blog*. Available from: <

- http://rmcpharmanews.blogspot.com/2010/06/riboflavin-plus-uva-irradiation-another.html>. [4 May 2015].
- Palmieri, L. and Cacace, D. (2005). High Intensity Pulsed Light Technology. In Da-Wen, S. (Ed.) *Emerging Technologies for Food Processing* (10.1016/b978-012676757-5/50013-xpp. 279-306). London: Academic Press.
- Pataro, G., Muñoz, A., Palgan, I., Noci, F., Ferrari, G. and Lyng, J. G. (2011). Bacterial inactivation in fruit juices using a continuous flow Pulsed Light (PL) system. *Food Research International*. 44(6), 1642-1648.
- Petrov, V., Marchev, G., Tyazhev, A., Starikova, M., Esteban-Martin, A., Panyutin, V., Badikov, V., Shevyrdyaeva, G., Badikov, D., Reza, M., Sheina, S. and Fintisova, A. (2013). Optical damage limits in chalcogenide nonlinear crystals used in 1064nm pumped nanosecond optical parametric oscillators. *Proceedings of the 2013*, 878603-878603-878620.
- Popruzhenko, S. V., Zaretsky, D. F. and Becker, W. (2010). High-order harmonic generation by an intense infrared laser pulse in the presence of a weak UV pulse. *Physical Review A Atomic, Molecular, and Optical Physics*. 81(6).
- Reynolds, J. (2011). Bacterial Colony Morphology. Available from: < http://delrio.dcccd.edu/jreynolds/microbiology/2421/lab_manual/colony_morph. pdf>. [9 October 2013].
- Reynolds, J. and Farinha, M. (2005). Counting Bacteria. Available from: < http://www.biotech.univ.gda.pl/odl/doc/numbers.pdf>. [8 October 2013].
- Saloman, E. B. (2004). Energy Levels and Observed Spectral Lines of Xenon, XeI through XeIIV. *Journal of Physical and Chemical Reference Data*. 33(3), 765-921.
- Schoenly, J. E., Seka, W., Featherstone, J. D. B. and Rechmann, P. (2012). Near-UV laser treatment of extrinsic dental enamel stains. *Lasers in Surgery and Medicine*. 44(4), 339-345.
- Shapley, P. (2015). Light and the Electromagnetic Spectrum. Available from: http://butane.chem.uiuc.edu/pshapley/GenChem2/A3/3.html. [31 March 2015].
- Simrock (2013). Laser Damage Threshold. Available from: http://www.laser2000.co.uk/technotes/TN_LaserDamageThreshold.pdf>. [17 October 2013].

- Smith, M. B. (2013). March's Advanced Organic Chemistry: Reactions, Mechanisms, and Structure, 7th Edition. Wiley.
- Sutton, S. (2011). Accuracy of Plate Counts. *Journal of Validation Technology*. 17(3), 42 46.
- Suzuki, M., Ganeev, R. A., Baba, M. and Kuroda, H. (2008). Characteristics of high-order harmonic spectrum by using laser-ablated two targets combination. *Physics Letters A*. 372(24), 4480-4483.
- Takeshita, K., Shibato, J., Sameshima, T., Fukunaga, S., Isobe, S., Arihara, K. and Itoh, M. (2003). Damage of yeast cells induced by pulsed light irradiation. *International Journal of Food Microbiology*. 85(1–2), 151-158.
- Tamuri, A., Bidin, N. and Daud, Y. (2008). Quality of the beam produced a pulsed Nd: YAG laser. *Laser physics*. 18(1), 18-21.
- Tessman, I. and Bank, H. (1992). Documentation of the claim that modulation of UV light pulses increases the bactericidal effectiveness of the light [2]. *Applied and Environmental Microbiology*. 58(2), 778-779.
- Turtoi, M. and Nicolau, A. (2007). Intense light pulse treatment as alternative method for mould spores destruction on paper—polyethylene packaging material. *Journal of Food Engineering*. 83(1), 47-53.
- USFDA (2001). BAM: Aerobic Plate Count. Available from: < http://www.fda. gov/Food/FoodScienceResearch/LaboratoryMethods/ucm063346.htm>. [28 October 2013].
- Villarroel, A. Y. R., Aron-Maftei, N., Martín-Belloso, O. and Soliva-Fortuny, R. (2012a). Influence of spectral distribution on bacterial inactivation and quality changes of fresh-cut watermelon treated with intense light pulses. Postharvest Biology and Technology. 69(0), 32-39.
- Villarroel, A. Y. R, Aron-Maftei, N., Martín-Belloso, O. and Soliva-Fortuny, R. (2012b). The role of pulsed light spectral distribution in the inactivation of Escherichia coli and Listeria innocua on fresh-cut mushrooms. *Food Control*. 24(1–2), 206-213.
- Wang, B., Dai, G., Zhang, H., Ni, X., Shen, Z. and Lu, J. (2011). Damage performance of TiO2/SiO2 thin film components induced by a long-pulsed laser. *Applied Surface Science*. 257(23), 9977-9981.

- Wang, G., Geng, A., Bo, Y., Li, H., Sun, Z., Bi, Y., Cui, D., Xu, Z., Yuan, X., Wang, X., Shen, G. and Shen, D. (2006). 28.4 W 266 nm ultraviolet-beam generation by fourth-harmonic generation of an all-solid-state laser *Optics Communications*. 259(2), 820-822.
- Wang, T., MacGregor, S. J., Anderson, J. G. and Woolsey, G. A. (2005). Pulsed ultraviolet inactivation spectrum of Escherichia coli. *Water Research*. 39(13), 2921-2925.
- Wesche, A. M., Gurtler, J. B., Marks, B. P. and Ryser, E. T. (2009). Stress, sublethal injury, resuscitation, and virulence of bacterial foodborne pathogens. *Journal of Food Protection*®. 72(5), 1121-1138.
- Ya, X., Liu, Q., Gong, M., Fu, X. and Wang, D. (2009a). High-repetition-rate high-beam-quality 43 W ultraviolet laser with extra-cavity third harmonic generation. *Applied Physics B: Lasers and Optics*. 95(2), 323-328.
- Ya, X., Liu, Q., Gong, M., Fu, X. and Wang, D. (2009b). High-repetition-rate high-beam-quality 43 W ultraviolet laser with extra-cavity third harmonic generation. *Applied Physics B*. 95(2), 323-328.
- Yazdi, M. K. S. and Darghahi, H. (2006). Inactivation of Pathogenic Bacteria Using Pulsed UV-Light and its Application in Water Disinfection and Quality Control. *Acta Medica Iranica*. 44(5), 305-308.
- Yamauti, M., Senawongse, P., Hamakawa, T., Otsuki, M., Tagami, J., Sato, S., Sato, Y. and Eguchi, T. (2003). Effect of pulse duration of Er:YAG laser on the dentin surface morphology. *International Congress Series*. 1248(0), 139-142.
- Yang, F., Wang, Z., Zhou, Y., Li, F., Xu, J., Xu, Y., Cheng, X., Lu, Y., Bo, Y., Peng, Q., Cui, D., Zhang, X., Wang, X., Zhu, Y. and Xu, Z. (2009). Theoretical and experimental investigations of nanosecond 177.3 nm deep-ultraviolet light by second harmonic generation in KBBF. *Applied Physics B*. 96(2-3), 415-422.
- Yong, H. I., Kim, H. J., Nam, K., Kwon, J. and Jo, C. (2015). Radiation sensitivity of foodborne pathogens in meat byproducts with different packaging. *Radiation Physics and Chemistry*. 115, 153-157.
- Zhai, S. Y., Wang, X. L., Wei, Y., Chen, W. D., Zhuang, F. J., Xu, S., Li, B. X., Fu, J. J., Chen, Z. Q., Wang, H. W., Huang, C. H. and Zhang, G. (2013). A compact efficient deep ultraviolet laser at 266 nm. *Laser Physics Letters*. 10(4), 045402.