

PRODUCTION OF VALUABLE ENZYMES THROUGH
FERMENTATION OF TROPICAL FRUIT DREGS

HII WANG SING

A dissertation submitted in partial fulfilment of the
requirements for the award of the degree of
Master of Engineering (Bioprocess)

Faculty of Chemical Engineering
Universiti Teknologi Malaysia

FEBRUARY 2014

TO MY BELOVED FAMILY AND SSN

ACKNOWLEDGEMENT

This dissertation has accumulated many debts of thankfulness throughout its completion. First of all, I would like to express my sincere thanks to Faculty of Chemical Engineering, Universiti Teknologi Malaysia which provided me a good learning environment and research opportunity.

I would also like to extend my sincere appreciation to my supervisor, Dr. Roshanida Binti A. Rahman who has provided a great help in constructive criticism on my study, valuable suggestions, continuous encouragement and supervising throughout the completion of dissertation. Without her commitment, advice, and help, this dissertation would not be able to success.

In addition, I would also like to pay special tribute to all the master and PhD students for their patience, valuable advices and guidance in this dissertation. Sincere appreciation is also extended to the laboratory assistants for always being there to guide and share their knowledge with me.

Last but not least, I would like to thank my beloved family especially my mother Wong Sio Kee, my brother and all my sisters for their support, caring, and encouragement.

ABSTRACT

The amount of waste in Malaysia is getting higher due to the increasing of population and development. The statistic showed that Malaysian generated 23,000 tonnes of waste in the year 2008 and the amount is expected to rise until 30,000 tonnes by the year 2020. From the waste generated, there is about 48% of organic and food waste. Therefore, it has been many solutions suggested nowadays to solve the organic waste management problem, especially in the conversion of waste into value added products. In this study, a bioconversion of selected tropical fruit dregs into valuable enzymes was investigated. The tropical fruit dregs used in the fermentation were papaya, pineapple and guava. The optimization of amylase activity, lipase activity, reducing sugar concentration and biomass content were carried out for 120th day of the tropical fruit dregs fermentation by response surface methodology. The fermentation of guava waste performed an optimum amylase activity at 0.05 U/ mg whereas the fermentation of papaya, pineapple and guava waste showed an optimum lipase activity at 111.33 U/mL. For the optimization of multiple responses, the combination of papaya, pineapple and guava waste performed the optimum enzyme activities at 95th day of the fermentation period. The optimum amylase and lipase activities were 0.0289 U/mg and 105.971 U/mL respectively. These valuable enzymes that produced during the fermentation process are believed to be a good cleaning agent.

ABSTRAK

Jumlah sisa di Malaysia semakin meningkat disebabkan oleh peningkatan bilangan penduduk dan pembangunan negara. Statistik menunjukkan Malaysia telah menjana 23,000 tan sisa buangan pada tahun 2008 dan jumlah itu dijangka akan meningkat sehingga 30,000 tan metrik menjelang tahun 2020. Sebanyak 48% daripada sisa buangan itu adalah terdiri daripada sisa organik dan makanan. Oleh itu, banyak cara penyelesaian telah disyorkan pada hari ini untuk menyelesaikan masalah pengurusan bahan buangan organik, terutamanya dengan cara menambah nilai pada sisa buangan sehingga menjadi produk nilai tambah. Proses penukaran dan penambahan nilai dari sesetengah buah-buahan tropika yang dipilih kepada enzim yang berguna akan dikaji dalam kajian ini. Sisa buah-buahan tropika yang digunakan dalam penapaian ialah betik, nanas dan jambu. Satu analisis tentang mengoptimumkan ujian enzim pada assay amilase dan lipase serta kepekatan gula dan kandungan biomas telah dijalankan sepanjang 120 hari dalam tempoh proses penapaian dengan menggunakan kaedah gerak balas permukaan. Penapaian sisa guava menunjukkan aktiviti amylase yang optimum pada 0.05 U/mg manakala penapaian sisa gabungan betik, nanas dan jambu menunjukkan aktiviti lipase yang optimum pada 111.33 U/mL. Bagi pengoptimuman pelbagai factor, sisa gabungan betik, nanas dan jambu telah menghasilkan enzim secara optimum pada hari ke-90 dalam tempoh proses penapaian. Aktiviti amilase dan lipase yang optimum adalah 0,0289 U / mg dan 105,971 U / mL masing-masing. Enzim bernilai yang dihasilkan semasa proses penapaian dipercayai dapat menjadi sebagai ejen pembersihan yang baik.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	x
	LIST OF FIGURES	xii
	LIST OF SYMBOLS	xiv
1	INTRODUCTION	1
	1.1 Background of Study	1
	1.2 Problem Statement	4
	1.3 Objectives	4
	1.4 Scope of Study	5
2	LITERATURE REVIEW	7
	2.1 Organic Waste Managment	7
	2.2 Fermentation	9
	2.2.1 Solid State Fermentation (SSF) and Submerged Fermentation (SmF)	9
	2.2.2 Fermentation of Garbage Enzymes	12
	2.3 Enzyme	14
	2.4 Amylase	16

2.4.1	Microbial Amylase	17
2.4.2	Production of Microbial Amylase	17
2.5	Lipase	21
2.5.1	Microbial Lipase	22
2.5.2	Production of Enzyme Lipase	25
3	METHODOLOGY	29
3.1	Chemicals	29
3.2	Sample Preparation	30
3.3	Experimental Design	31
3.4	Analytical Methodologies	32
3.4.1	Determination of Biomass Content	33
3.4.2	Determination of Reducing Sugar Concentration	33
3.4.3	Determination of Amylase Activities	34
3.4.4	Determination of Lipase Activity	35
4	RESULTS AND DISCUSSION	37
4.1	Effect of Incubation Time and Sample Variety on Simple Fermentation Process	37
4.2	Regression Analysis for the Responses	39
4.3	Response 1: Amylase Activity	45
4.3.1	Optimization Point for Amylase Activity	51
4.4	Response 2: Lipase Activity	54
4.4.1	Optimization Point for Lipase Activity	59
4.5	Response 3: Reducing Sugar Concentration	60
4.5.1	Optimization Point for Reducing Sugar Concentration	64
4.6	Response 4: Residue Mass	66
4.6.1	Optimization Point for Residue Mass	71
4.7	Optimization Point Prediction and Verification for Both Enzyme Amylase and Lipase	72
5	CONCLUSION	73

REFERENCES

75

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	List of microorganisms involved in solid state fermentation processes.	11
2.2	Applications of solid state fermentation	12
2.3	General comparison between solid state fermentation and submerged fermentation.	13
2.4	Examples of enzymes used in various industrial segments and their applications.	15
2.5	Some amylase-producing microorganisms.	18
2.6	Amylase-producing microorganisms under different carbon sources.	19
2.7	Amylase-producing microorganisms under different nitrogen sources.	19
2.8	Amylase-producing microorganisms under different pH values.	20
2.9	Amylase-producing microorganisms under different temperature.	21
2.10	Some lipase-producing microorganisms.	23
2.11	Commercially available microbial lipases.	24
2.12	Lipase-producing microorganisms under different pH and temperature.	25
2.13	Lipase-producing microorganisms under different substrates.	26
2.14	Lipase-producing microorganisms under different growth media.	27
2.15	Lipase-producing microorganisms under different sources of nitrogen and carbon.	28
3.1	Chemicals used in the experiments	29

3.2	Contents of tropical fruit dregs in different plastic container.	30
3.3	Experiment design matrix for fermentation of tropical fruit dregs.	31
4.1	Responses of the fermentation process with two categorical factors.	40
4.2	Regression equations for investigated responses along with ANOVA results.	43
4.3	Regression analysis for amylase activity in fermentation process.	47
4.4	Optimum amylase activities under different substrate and incubation period.	53
4.5	Regression analysis for lipase activity in fermentation process.	56
4.6	Regression analysis for reducing sugar concentration in fermentation process.	61
4.7	Regression analysis for biomass content in fermentation process.	67
4.8	Numerical optimization of RSM for enzyme activities.	72

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
4.1	Diagnostic plot of response (amylase activity) for actual, predicted and their residuals.	48
4.2	Interaction graph of amylase activities versus incubation time for all the sample variety.	50
4.3	The highest amylase activity for guava waste at 110 th day of the fermentation process.	52
4.4	Diagnostic plot of response (lipase activity) for actual, predicted and their residuals.	57
4.5	Interaction graph of lipase activities versus incubation time for all the sample variety.	58
4.6	The highest lipase activity for the combination of papaya, pineapple and guava waste at the 90 th day of the fermentation process.	60
4.7	Diagnostic plot of response (reducing sugar concentration) for actual, predicted and their residuals.	62
4.8	Interaction graph of reducing sugar concentration versus incubation time for all the sample variety.	63
4.9	The highest reducing sugar concentration for the pineapple waste at the 80 th day of the fermentation process.	64
4.10	Diagnostic plot of response (residue mass) for actual, predicted and their residuals.	68
4.11	Interaction graph of residue mass versus incubation time for	

	all the sample variety.	69
4.12	The highest residue mass for the combination of papaya and guava waste at the 120 th day of the fermentation process.	71

LIST OF SYMBOLS

°C	Degree Celcius
%	Percentage
w/v	Weight/Volume
v/v	Volume/Volume
g	Gram
mL	Milliliter
µL	Microliter
cm	Centimeter
M	Molar
mM	Millimolar
µg/mL	Microgram per milliliter
rpm	Revolutions per minute
V _e	Elution volume
V _o	Void volume
kDa	Kilo Dalton
U/mL	Unit of lipase per milliliter

CHAPTER 1

INTRODUCTION

1.1 Background of Study

The issue of the environmental quality is enthusiastic in these recent years (Li *et al.*, 2009). These may related to the continuous development of the countries in order to improve the lives of citizens. The development is either directly or indirectly affected the environment which causes climate change and global warming to the earth. As the residents on this planet, human beings are suffering from the adverse effects of the development instead of enjoying the life. Therefore, many countries started to promote the concept of sustainable development. With this concept, all the processes of development are assurance for not destroying the environment or ecological balance.

There are many factors that contribute to the degradation of environmental quality. Among these factors, the management of household and industrial waste is the most serious issue (Jalil, 2010). In Malaysia, an average of 3500 tonnes of wastes has been generated by the citizens per day (Tarmudi *et al.*, 2009). All the wastes will end up as the landfill and around 50% of the wastes are organic wastes (Behzad *et al.*, 2011). Landfill is traditional waste management in Malaysia. There are about 230

operating landfill sites in Malaysia, however, majority of the landfill are poorly managed (Masirin *et al.*, 2008). The situation is definitely against the concept of sustainable development and further brings down the development of a country.

Many researches focused on the study of organic wastes as it occupies the large portion of the total wastes. Generally, organic wastes are anything that comes from biological origins that are biodegradable, such as fruits, vegetables, meats, crop wastes and so on. As the population keeps rising, the rate of producing organic wastes is also getting increase. Inadequate technologies in managing the large amount of wastes cause severe pollution to the environment. In this situation, the government is not the only one to blame on, as the residents should also pay the responsibility on the problems.

Tan (2009) had introduced the simple fermentation method of organic wastes to the world. With this simple fermentation of organic wastes, everyone can make their wastes into useful products (garbage enzymes) and not simply dispose the wastes into landfills or bring into the incinerators. The raw materials that used in fermentation are organic waste such as vegetables and fruit dregs, sugar and water (Bhavani, 2011). The fermentation process is considered as anaerobic process as there is no oxygen supply to the medium and the container will be closed tightly throughout the fermentation process.

Enzymes will be produced throughout the fermentation process, which is termed as “garbage enzyme” (Tan, 2009). The garbage enzymes are believed to act as cleaning agents, fertilizer and even also insecticide. In general, enzymes also well-known as biocatalysts which are important in living organisms as it can increase the rate of chemical reaction. Therefore, the production of garbage enzymes will indirectly replace non-biodegradable cleaning materials and responsible in decomposing toxic chemicals into harmless types.

Organic wastes such as fruit dregs can be obtained from every household as fruits are good source of vitamins, antioxidants, fibre and minerals that required by everyone on the Earth. Therefore, fruit dregs such as fruit peels, cuttings and bits are the common organic wastes that generated by us. If every household is able to utilize the simple fermentation method in fermenting fruit dregs at home, the effort will not only reduce the wastes that going to dispose in the landfills, but also save the environment.

In this study, tropical fruit dregs are being fermented using the simple fermentation method. The tropical fruit dregs that are going to use is from papaya, pineapple, and guava. The reason of choosing these tropical fruits is based on its availability in Malaysia. As theoretically, those fruits are basically from tropical country and Malaysia is one of the major exporters. Besides that, there is no season for the fruits to grow and the fruits can be obtained throughout the year. Second reason is that the demand of domestic food products is getting increase. The introduction of tropical fruits in the market is successful as it is safer to consume if compared to the imported fruits.

There is still no scientific research on the garbage enzymes that fermentated using simple fermentation method. Therefore, the reason of why the garbage enzymes are useful in cleaning purposes needs to be further investigated scientifically. Bhavani (2011) assumed that the end product which is useful in cleaning purpose is basically acetic acid which is known as non-toxic cleaner based on its acidic properties. Anyway, the garbage enzymes contributed with its low cost production, no addition of synthetic chemicals and production of high energy. If everyone can make simple fermentation of organic wastes instead of disposing or incinerating the wastes improperly, environmental pollution can be reduced.

1.2 Problem Statement

The increasing amount of organic wastes and the waste management system in Malaysia had always become an issue of the environmental problem. Many researches are done on bioconversion of the wastes into other valuable products rather than disposed the wastes into landfills. The bioconversion of organic wastes is typically referred to fermentation of the wastes that collected from industrial area. However, the environmental problem still cannot be solved as the organic wastes that produced by household are difficult to categorize and further process the wastes into valuable products.

Simple fermentation method is introduced where every household is able to put their responsibility on the organic wastes that had been generated. The fermentation will produce valuable enzymes which also term as “garbage enzyme”. Garbage enzyme is excellent in the cleaning activities as it is believed that enzymes are excreted after three months of fermentation (Tan, 2009). However, there is still lack of scientific evidence to prove the duration for the enzymes to be produced and the amount of enzyme that can be produced by using different raw materials in the fermentation process.

1.3 Objectives

The objective of this study is to optimize the enzyme production of fermented tropical fruit dregs by using response surface methodology.

1.4 Scope of Study

In this study, the selected tropical fruit dregs are fermented only with molasses and water. The selected tropical fruit dregs are included papaya, pineapple, and guava. The reasons of choosing these tropical fruit dregs are based on its availability in Malaysia and there are non-seasonal fruits. Molasses with the lower cost are being chosen instead of brown sugars. This is due to the higher potential of un-processing sugars as carbon source in the fermentation. Dark brown liquid with unpleasant smell will be produced after complete fermentation step. It is believed that microorganisms that present in the fruit dregs utilize molasses as their carbon source and further produce useful enzymes. These useful enzymes can be used as cleaning agent in the routine household activities.

The optimization of the tropical fruit dregs fermentation is performed by response surface methodology (RSM). RSM is useful in assembling a functional relationship between the response values obtained from experiments and a set of design variables. In this study, the design variables are the incubation time and sample varieties. There are seven fermentation samples that will be fermented with certain ratio of pineapple, papaya and guava wastes. Each of the fermentation samples will be tested on several enzyme activities at 30th, 60th, 80th, 90th, 100th, 110th and 120th days.

These enzyme activities included lipase and amylase. Lipase and amylase are chosen instead of protease because protease is the promising enzyme in cleaning purpose and many researches have been done on it. Study of lipase and amylase are under highly concern nowadays based on their potential of biodegradation in the environment. Besides that, these enzymes loss their enzymatic activity under washing and cleaning conditions which reduces pollution problems.

The residue mass of the sample and the concentration of reducing sugar (glucose) are also being tested besides the enzyme activities. These are to ensure the completion of the fermentation process. The residue mass of the sample and reducing sugar concentration are increased following the raising of enzyme activities.

REFERENCES

- Afolabi, I. S., Marcus, G. D., Olanrewaju, T. O. and Chizea, Vivian. (2011). Biochemical effect of some food processing methods on the health promoting properties of under-utilized *Carica papaya* seed. *Journal of Natural Products*. 4: 17-24.
- Agamuthu, P., and Fauziah, S. H. (2006). MSW disposal in Malaysia: landfill management. In: Proceedings of the 2nd Expert Meeting on Solid Waste Management in Asia and the Pacific Islands, Kitakyushu, November 23-24, 2006.
- Alkan, H., Baysal, Z., Uyar, F. and Dogru, M. (2007). Production of lipase by a newly isolated *Bacillus coagulans* under solid-state fermentation using melon wastes. *Appl Biochem Biotechnol*. 136(2): 183-192.
- Arpigny, J.L. & Jaeger, K.-E. (1999). Bacterial lipolytic enzymes: classification and properties. *Biochemical Engineering Journal*, 343:177-183.
- Arrizon, J. and Gschaedler, A. (2002). Increasing fermentation efficiency at high sugar concentrations by supplementing an additional source of nitrogen during the exponential phase of the tequila fermentation process. *Canada Journal of Microbiology*. 48:965-970.
- Balkan, B., Balkan, S. and Ertan, F. (2011). Optimization of parameters for α -amylase production under solid state fermentation by *Trichothecium roseum*. *Romanian Biotechnological Letters*. 16(5):6591-6600.
- Barrios-Gonzales, J., Tomasini, A., Viniegra-Gonzalez, G., and Lopez, L. (1988). Penicillin production by solid state fermentation. In: Solid State Fermentation in Bioconversion of Agro-industrial Raw Materials, Ed. M. Raimbault, ORSSOM, Montpellier Fr., pp 39-51.

- Behzad, N., Ahmad, R., Saied, P., Elmira, S. and Mokhtar, M. B. (2011). Challenges of solid waste management in Malaysia. *Research Journal of Chemistry and Environment*. 15(2):1-4.
- Bhargav, S., Panda, B. P., Ali, M. and Javed, S. (2008). Solid-state fermentation: An overview. *Chem. Biochem. Eng. Q.* 22(1)-49-70
- Bhavani, P. (2011). Turning garbage into gold. *VegVibe*, 10-12.
- Bottino, N.R., Vandenburg, G.A. & Reiser, R. 1967. Resistance of certain long-chain polyunsaturated fatty acids of marine oils to pancreatic lipase hydrolysis. *Chemistry and material science*, 2:489-493.
- Cinthia, A. A. S., Maria, P. F. L. and Gustavo, G. F. (2013). Biotransformation of pegui and guavira fruit wastes via solid state bioprocess using *Pleurotus Sajor-Caju*. *International Journal of Bioscience, Biochemistry and Bioinformatics*. 3(2):88-92
- Chen, J.Y., C.M. Wen and T.L. Chen. (1999). Effect of oxygen transfer on lipase production by *Acinetobacter radioresistens* *Biotechnol. Bioeng.* 62: 311-316.
- Couto, S. R. and Sanroman, M. A. (2006). Application of solid-state fermentation to food industry- A review. *J. Food Eng.* 76:291-302.
- Demirkan, E. (2011). Production, purification and characterization of α -amylase by *Bacillus subtilis* and its mutant derivatives. *Turkish Journal Biology*. 35:705-712.
- Diaz, A., Sieiro, C. and Villa, T. G. (2003). Production and partial characterization of a β -amylase by *Xanthophyllomyces dendrorhous*. *Letters in Applied Microbiology*. 36(4):203-207.
- Drouault, S., Corthier, G., Ehrlich, D. and Renault, P. (2000). Expression of the *Staphylococcus hyicus* lipase in *Lactococcus lactis*. *Appl. Environ. Microbiol.* 66(2): 588-598.
- Elibol, M. (2004). Optimization of medium composition for actinorhodin production by *Streptomyces coelicolor* A3(2) with response surface methodology. *Process Biochemistry*. 39(9):1057-1062.
- Ellaiah, P., Srinivasulu, B. Adinarayana, K. (2002). A review on microbial alkaline proteases. *Journal of Science and Industrial Research*. 61: 690-704.
- Fadiloğlu, S. and Erkmen, O. (2002). Effects of carbon and nitrogen sources on lipase production by *Candida rugosa*. *Turkish Journal of Engineering and Environmental Science*. 26:249-254.

- Farid, M. A. F. and Shata, H. M. A. H. (2011). Amylase production from *Aspergillus oryzae* LS1 by solid-state fermentation and its use for the hydrolysis of wheat flour. *Iranian Journal of Biotechnology*. 9(4):267-274.
- Fnides, B., Yallese, M. A., Mabrouki, T. and Rigal, J-F. (2011). Application of response surface methodology for determining cutting force model in turning hardened AISI H11 hot work tool steel. *Indian Academy of Sciences*. 36(1):109-123.
- Gajdoš, R. (1998). Bioconversion of organic waste by the year 2010: to recycle elements and save energy. *Resources, Conservation and Recycling*. 23: 67-86.
- Ghosh, S., Chakraborty, R., Chatterjee, G. and Raychaudhuri, U. (2012). Study on fermentation conditions of palm juice vinegar by response surface methodology and development of a kinetic model. *Brazilian Journal of Chemical Engineering*. 29(3): 461-472.
- Gupta, R., Gigras, P., Mohapatra, H., Goswami, V. K. and Chauhan, B. (2003). Microbial α -amylases: a biotechnological perspective. *Process Biochem*. 38:1599-1616.
- Hajar, N., Zainal, S., Atikah, O., and Tengku Elida, T. (2012). Optimization of ethanol fermentation from pineapple peel extract using Response Surface Methodology (RSM). *World Academy of Science, Engineering and Technology*. 72:641-647.
- Hemalatha, R. and Anbuselvi, S. (2013). Physicochemical constituents of pineapple pulp and waste. *Journal of Chemical and Pharmaceutical Research*. 5(2):240-242.
- Hun, C. J., Zaliha, R. N. A. R., Salleh, A. B. and Basri, M. (2003). A newly isolated organic solvent tolerant *Bacillus sphaericus* 205y producing organic solvent-stable lipase. *Biochemical Engineering Journal*. 15:147-151.
- Ibrahim, A. D., Aisha, I. S., Alhassan, S., Danladi, M. S., Saadatu, A. S., Adamu, A. A. and Gambo, A. (2011). Bioutilization of *Adansonia Digitata* fruit pulp by *Bacillus* species for amylase production. *International Journal of Plant, Animal and Environmental Science*. 1(1):35-41.
- Itelima, J. Onwuliri, F. Onyimba, I. and Oforji, S. (2013). Bio-ethanol production from banana, plantain and pineapple peels by simultaneous saccharification and fermentation process. *International Journal of Environmental Science and Development*. 4(2): 213-216.

- Jaeger, K.-E. & Reetz, M.T. (1998). Microbial lipases form versatile tools for biotechnology. *Trends in Biotechnology*, 16(9):396-403.
- Jaeger, K.E. & Wohlfarth, S. 1993. Bacterial lipases: Biochemistry, molecular genetics, and applications in biotechnology. *Bioengineering*, 9:39-46.
- Jakson, R. B., Cook, C. W., Phippen, J. S. and Palmer, S. M. (2009). Increased below ground biomass and soil CO₂ fluxes after a decade of carbon dioxide enrichment in a warm-temperate forest. *Ecology*. 90:3352-3366.
- Jalil, M. A. (2010). Sustainable development in Malaysia: A case study on household waste management. *Journal of Sustainable Development*. 3(3):91-102.
- Jamrath, T., Lindner, C., Popovic, M. K. and Bajpai, R. (2012). Production of amylases and proteases by *Bacillus caldolyticus* from food industry wastes. *Journal of Food Technology and Biotechnology*. 50(3):355-361.
- José, G. P. M., Ernani, P., Cristina, B. C., Severino, M. A., Eduardo, M. G., Ingridy, S. R. C. and Lígia, M. A. (2012). Antimicrobial potential and chemical composition of agro-industrial wastes. *Journal of Natural Products*. 5(2012): 27-36.
- Juwon, A. D. and Emmanuel, O. F. (2012). Experimental investigations on the effects of carbon and nitrogen sources on concomitant amylase and polygalacturonase production by *Trichoderma viride* BITRS-1001 in submerged fermentation. *Biotechnology Research International*. 2012:1-8.
- Kachensuwan, C., Komolkittikan, K. and Laloknam, S. Detection of Lipase Activity from Fruits. 34th Congress on Science and Technology of Thailand. November 10, 2008. Bangkok. P.1-6.
- Kandari, V. and Gupta, S. (2012). Bioconversion of vegetable and fruit peel wastes in viable product. *Journal of Microbiology and Biotechnology Research*. 2(2):308-312.
- Karp, G. (2009). *Cell and molecular biology: Concepts and experiments*. New York: John Wiley & Son.
- Kim, S. and Dale, B. E. (2004). Global potential bioethanol production from wasted crops and crop residues. *Biomass Bioenergy*. 26:361-375.
- Kokab, S., Asghar, M., Rehman, K., Asad, M. J. and Adedyo, O. (2003). Bio-processing of banana peel for α -amylase production by *Bacillus subtilis*. *International Journal of Agriculture and Biology*. 5(1):36-39.
- Kishore, K. A. and Reddy, G. V. (2011). Optimization of operating variables in the

- fermentation of citric acid using Response Surface Methodology. *2011 2nd International Conference on Chemical Engineering and Applications*. IACSIT Press, Singapore, 108-113.
- Kishore, K. A. and Reddy, G. V. (2012). Optimization of incubation time, fermentation temperature & O₂ flow rate in citric acid fermentation using Response Surface Methodology (RSM). *International Journal of Advanced Engineering Technology*. 3(1): 64-67.
- Krishna, P. R., Srivastava, A. K., Ramaswamy, N. K., Suprasanna, P. and Souza, S. F. D. (2012). Banana peel as substrate for α -amylase production using *Aspergillus niger* NCIM 616 and process optimization. *Indian Journal of Biotechnology*. 11:314-319.
- Kumar, P. K. R. *Microbial production of gibberellic acid*. PhD. Thesis. Mysore University, Mysore, India; 1987.
- Kumar, S., Kikon, K., Upadhyay, A., Kanwar, S. S. and Gupta, R. (2005). Production, purification and characterization of lipase from thermophilic and alkaliphilic *Bacillus coagulans* BTS-3. *Protein Expression and Purification*. 41:38-44.
- Kunamneni, A., Permaul, K. and Singh, S. (2005). Amylase production in solid state fermentation by the thermophilic fungus *thermomyces lanuginosus*. *Journal of Bioscience and Bioengineering*. 100(2):168-171.
- Lakshmi, B. S., Kanguane, P., Abraham, B. and Pennathur, G. (1999). Effect of vegetable oils in the secretion of lipase from *candida rugosa* (DSM 2031). *Letters in Applied Microbiology*. 29:66-70.
- Lee, D. W., Kim, H. W., Lee, K. W., Kim, B. C., Choe, E. A., Lee, H. S., Kim, D. S. and Pyun, Y. R. (2001). Purification and characterization of two distinct thermostable lipases from the gram-positive thermophilic bacterium *Bacillus thermoleovorans* ID-1. *Enzyme and Microbial Technology*. 29: 363-371.
- Li, P. J., Stagnitti, F., Gong, Z. Q., Xiong, X. Z., Li, X. T., Hu, Z. Z., Sun, Y., Wang, Z. J., Gao, K., Kong, C. J. and Li, P. (2009). Environmental quality: issues and causes of deterioration- A survey on environmental awareness among the public in Liaoning Province, China. *International Journal of Sustainable Development & World Ecology*. 16(3): 143-150.
- Li, S., Yang, X., Yang, S., Zhu, M. and Wang, X. (2012). Technology prospecting on

- enzymes: application, marketing and engineering. *Computational and Structural Biotechnology Journal*. 2(3):1-11.
- Li, X., Zhang, Z. and Song, J. N. (2012). Computational enzyme design approaches with significant biological outcomes: progress and challenges. *Computational and Structural Biotechnology Journal*. 2(3).
- Liu, B. L. and Tzeng, Y. M. (1998). Optimization of growth medium for production of spores from *Bacillus thuringiensis* using response surface methodology. *Bioprocess Engineering*. 18: 413-418.
- Liu, J. Z., Weng, L. P., Zhang, Q. L., Xu, H. and Ji, L. N. (2003). Optimization of glucose oxidase production by *Aspergillus niger* in a benchtop bioreactor using response surface methodology. *World Journal of Microbiology & Biotechnology*. 19:317-323.
- Long, Z. D., Xu, J. H. and Pan, J. (2007). Significant improvement of *Serratia marcescens* lipase fermentation, by optimizing medium, induction and oxygen supply. *Application of Biochemistry and Biotechnology*. 142(2):148-157.
- Macrae, A.R. & Hammond, R.C. (1985). Present and Future Applications of Lipases. *Biotechnology and Genetic Engineering*, 3:193-217.
- Marie, Z. (2012). Olive oil as inductor of microbial lipase, olive oil-constituents, quality, health properties and bioconversion, Dr. Dimitriou (Ed.), ISBN:978-953-307-921-9, In Tech, Available from <http://www.intechopen.com/books/olive-oil-constituents-quality-health-properties-and-bioconversions/olive-oil-as-inductor-of-microbial-lipase>
- Marshall, A. G. (2013). An anarchistic understanding of the social order: Environmental degradation, indigenous resistance and a place for the sciences. *Spanda Journal*. 4(1):53-67.
- Masirin, M. I. H. M., Ridzuan, M. B. Mustapha, S. and Adon, R. (2008). An overview of landfill management and technologies: A Malaysian case study at Ampar Tenang. *1st National Seminar on Environment, Development & Sustainability (PSI Senviro 2008): Ecological, Economical and Social Aspects*. 28-29 July 2008. Selangor, Malaysia, 157-165.
- Massadeh, M., Sabra, F., Dajani, R. and Arafat, A. (2012). Purification of lipase

- enzyme produced by *Bacillus Streatothermophilus* HU1. *International Conference on Ecosystems and Biological Sciences (ICEBS2010)*. 19-20 May 2012. Penang, Malaysia, 34-37.
- McAuliffe, J. C., Aehle, W. Whited, G. M. and Ward, D. E. (2007). Industrial enzymes and biocatalysts. In Kent, J. A. and Riegel's, K., (eds) *Handbook of industrial chemistry and biotechnology Vol. 1, 11th ed.* (p. 1375-1420) Springer science + Business media, city of publication.
- Mienda, B. S., Idi, A. and Umar, A. (2011). Microbiological features of solid state fermentation and its applications- an overview. *Research in Biotechnology*. 2(6):21-26.
- Mobarak-Qamsari, E., Kasra-Kermanshahi, R. and Moosavi-nejad, Z. (2011). Isolation and identification of a novel, lipase-producing bacterium, *Pseudomonas aeruginosa* KM110. *Iranian Journal of Microbiology*. 3(2):92-98.
- Montgomery, D. C. (1997). *Design and Analysis of Experiments*. New York: John Wiley and Sons.
- Murakoshi, Y., Makita, T., Kato, M. and Kobayashi, T. (2012). Comparison and characterization of α -amylase inducers in *Aspergillus nidulans* based on nuclear localization of AmyR. *Applied Microbial and Cell Physiology*. 94:1629-1635.
- Niu, D., Zuo, Z., Shi, G. Y. and Wang, Z. X. (2009). High yield recombinant thermostable α -amylase production using an improved *Bacillus licheniformis* system. *Microbial Cell Factories*. 8:58.
- Pandey, A. (1992). Recent process developments in solid-state fermentation. *Process Biochemistry*. 27(2):109-117.
- Pandey, A., Nigam, P., Soccol, V. T., Singh, D., Mohan, R. (2000). Advances in microbial amylases. *Biotechnology Applied Biochemistry*. 31, 135-52.
- Pandey, A. (2003). Solid- state fermentation. *Biochemical Engineering Journal*. 13(2):81-84.
- Pandey, A., Gangadharan, D., Sivaramakrishnan, S. and Nampoothiri, K. M. (2006). Solid culturing of *Bacillus amyloliquefaciens* for alpha amylase production. *Journal of Food Technology and Biotechnology*. 44(2):269-274.
- Papargyropoulou, E. (2010, October). For the love of food. Retrieved June 19, 2012, from <http://www.academia.edu>

- Paranthaman, R., Vidyalakshmi, R., Muruges, S. and Singaravadisel, K. (2009). Optimization of various culture media for Tannase production in submerged fermentation by *Aspergillus flavus*. *Advances in Biological Research*. 3(1-2):34-39.
- Patel, R. K., Dodia, M. S., Joshi, R. H. and Singh, S. P. (2006). Purification and characterization of alkaline protease from an newly isolated haloalkaliphilic *Bacillus sp.* *Process Biochemistry*. 41:2002-2009.
- Potumarthi, R., Subhakar, C., Vanajakshi, J. and Jetty, A. (2008). Effect of aeration and agitation regimes on lipase production by newly isolated *Rhodotorula mucilaginosa*-MTCC 8737 in stirred tank reactor using molasses as sole production medium. *Application Biochemistry Biotechnology*. 151(2-3):700-710.
- Qazi, J. I. (2005). Microbiological conversion of waste fruits and vegetables into ethanol. *Pakistan Research Repository*.
- Raimbault, M. and Alazard, D. (1980). Culture method to study fungal growth in solid fermentation. *European Journal of Applied Microbiology and Biotechnology*. 9, 199-209.
- Raimbault, M., Soccol, C. R., and Chuzel, G. (1998). International training course on solid state fermentation. Document ORSTOM, Montpellier France, n°1: oo. 204.
- Rashad, M. M., Abdou, H. M., Mahmoud, A. E. and Nooman, M. U. (2009). Nutritional analysis and enzyme activities of *Pleurotus Ostreatus* cultivated on *Citrus Limonium* and *Carica Papaya* wastes. *Australian Journal of Basic and Applied Sciences*. 3(4): 3352-3360.
- Rosenau, F. & Jaeger, K.-E. 2000. Bacterial lipases from *Pseudomonas*: Regulation of gene expression and mechanisms of secretion. *Biochimica et Biophysica Acta*, **82**(11):1023-1032.
- Rushton, L. (2003). Health hazards and waste management. *British Medical Bulletin*. 68:183-197.
- Samsudin, M. D. M and Don, M. M. (2013). Municipal solid waste management in Malaysia: Current practices, challenges and prospect. *Journal Technology*. 62(1): 95-101.

- Saxena, R.K., Ghosh, P.K., Gupta, R., Davidson, W.S., Bradoo, S. & Gulati, R. (1999). Microbial lipases: Potential biocatalysts for the future industry. *Current Science*, 77:101-115.
- Senthilkumar, R. and Selvakumar, G. (2008). Isolation and characterization of an extracellular lipase producing *Bacillus* sp. SS-1 from slaughterhouse soil. *Advanced Biotechnology*. 24-25.
- Sevda, S., Singh, A., Joshi, C. and Rodrigues, L. (2012). Extraction and optimization of guava juice by using Response Surface Methodology. *American Journal of Food Technology*. 7(6):326-339.
- Shafaghat, H., Najafpour, G. D., Rezaei, P. S. and Sharifzadeh, M. (2009). Growth kinetics and ethanol productivity of *Saccharomyces cerevisiae* PTCC 24860 on various carbon sources. *World Applied Sciences Journal*. 7(2):140-144.
- Sharanappa, A., Wani, K. S. and Patil, P. (2011). Bioprocessing of food industrial waste for α -amylase production by solid state fermentation. *International Journal of Advanced Biotechnology and Research ISSN 0976-2612*. 2 (4): 473-480.
- Sharma, R., Chisti, Y. and Banerjee, U. C. (2001). Production, purification, characterization and applications of lipases. *Biotechnology Advances*. 19:627-662.
- Shobhana, J. Dhanashree, C. and Shirish, R. (2013). Lipase production from banana peel extract and potato peel extract. *International Journal of Research in Pure and Applied Microbiology*. 3(1): 11-13.
- Shu, Z., Lin, R., Jiang, H., Zhang, Y., Wang, M. & Huang, J. (2009). A rapid and efficient method for directed screening of lipase-producing *Burkholderia cepacia* complex strains with organic solvent tolerance from rhizosphere. *Journal of Bioscience and Bioengineering*, 107(6):658-661.
- Shukla, P., Bhagat, J. and Shrivastava, S. (2011). Oil cakes as substrate for improved lipase production in solid state fermentation. *International Journal of Microbiology Research*. 3(1):71-73.
- Silva, C. J. S. M. and Roberto, I. C. (2001). Optimization of xylitol production by *Candida guilliermondii* FTI 20037 using response surface methodology. *Process Biochemistry*. 36:1119-1124.
- Sindhu, R., Suprabha, G. N. and Shashidhar, S. (2009). Optimization of process

- parameters for the production of α -amylase from *Penicilium janthinellum* (NCIM 4960) under solid state fermentation. *African Journal of Microbiology Research*. 3(9):498-503.
- Sivaramakrishnan, S., Gangadharan, D., Nampoothiri, K. M., Soccol, C. R. and Pandey, A. (2007). Alpha amylase production by *Aspergillus oryzae* employing solid-state fermentation. *Journal of Scientific & Industrial Research*. 66:621-626.
- Sokolovska, I., Albasi, C. Riba, J. P. and Bales, V. (1998). Production of extracellular lipase by *Candida cylindracea* CBS 6330. *Bioprocess Engineering*. 19(3):179-186.
- Solange I. Mussatto, Lina F. Ballesteros, Silvia Martins and José A. Teixeira (2012). Use of Agro-Industrial Wastes in Solid-State Fermentation Processes, Industrial Waste, Prof. Kuan-Yeow Show (Ed.), ISBN: 978-953-51-0253-3, InTech, Available from: <http://www.intechopen.com/books/industrial-waste/use-of-agro-industrialwastes-in-solid-state-fermentation-processes>
- Sousa, B. A. A. and Correia, R. T. P. (2010). Biotechnological reuse of fruit residues as a rational strategy for agro-industrial resources. *Journal of Technology Management and Innovation*. 5(2):104-112.
- Stanbury, P. F., Whitaker, A. and Hall, S. J. (1995). *Principles of fermentation technology*. Oxford: Pergamon Press.
- Stocklein, W., Sztajer, H., Menge, U. & Schmid, R.D. 1993. Purification and properties of lipase from *Penicillium expansum*. *Biochimica et Biophysica Acta* **1168**:181-189.
- Suganthi, R., Benazir, J. F., Santhi, R., Ramesh, K. V., Anjana, H., Nitya, M., Nidhiya, K. A., Kavitha, G. and Lakshimi, R. (2011). Amylase production by *Aspergillus niger* under solid state fermentation using agroindustrial wastes. *International Journal of Engineering Science and Technology*. 3:1753-1756.
- Tabachnick, B. and Fidell, L. (2007). *Experimental Design using ANOVA*. United States: Thomson Higher Education.
- Tan, C. L. 2009. Wonder Cleaner. *The star*, 7 April: 5.
- Tanyildizi, M. S., Özer, D. and Elibol, M. (2005). Optimization of α - amylase production by *Bacillus* sp. using response surface methodology. *Process Biochemistry*. 40: 2291-2296.
- Tarmudi, Z., Abdullah, M. L. and Tap, A. O. M. (2009). An overview of municipal

- solid wastes generation in Malaysia. *Journal Technology*. 51(F):1-15.
- Thanapimmetha, A., Vuttibunchon, K. and Titapiwatanakun, B. and Srinophakun, P. (2012). Optimization of solid state fermentation for reducing sugar production from agricultural residues of sweet sorghum by *Trichoderma harzianum*. *Chiang Mai J. Sci.* 39(2):270-280.
- Thurairatnam, S., Arasaratnam, V. and Senthuran, A. (2006). Effect of different carbon sources and endogenous nitrogen on simultaneous saccharification and fermentation. *Malaysian Journal of Biochemistry and Molecular Biology*. 13:1-10.
- Unakal, C., Kallur, R. I. and Kaliwal, B. B. (2012). Production of α -amylase using banana waste by *Bacillus subtilis* under solid state fermentation. *European Journal of Experimental Biology*. 2(4):1044-1052.
- Vishwakarma, M., Parashar, V. and Khare, V. K. (2012). Optimization and regression analysis of surface roughness for electric discharge machining of EN-19 alloy steel using tungsten copper electrode with design of experiments. *International Journal of Advanced Science, Engineering and Technology*. 1(2): 43-50.
- Wang, Y., Srivastava, K.C., Shen, G.-J. and Wang, H.Y. (1995). Thermostable alkaline lipase from a newly isolated thermophilic *Bacillus*, strain A30-1 (ATCC 53841). *Journal of Fermentation and Bioengineering*, 79(5):433-438.
- Wilke, D. (1999). Chemicals from biotechnology: molecular plant genetics will challenge the chemical and fermentation industry. *Appl. Microbiol. Biotechnol.* 51:401-404.
- Windish, W. W. and Mhatre, N. S. (1965). Microbial amylases. In Wayne W. U. (ed). *Advances in Applied Microbiology*. 7:273-304.
- Woolley, P. & Petersen, S.B. 1994. *Lipases, their structure, biochemistry and application*. Cambridge University Press, United Kingdom.
- Wong, E. (2013, March 29). Cost of environmental damage in China growing rapidly amid industrialization. *New York Times*, p. 1.
- Zambare, V. (2010). Solid state fermentation of *Aspergillus oryzae* for glucoamylase production on agro residues. *International Journal of Life Sciences*. 4:16-25.