BIOLOGICAL ASSESSMENT OF UNCOATED AND HYDROXYAPATITE-COATED FERUM AS BIODEGRADABLE SCAFFOLD FOR HARD TISSUE REGENERATION

NURIZZATI BINTI MOHD DAUD

A thesis submitted in fulfillment of the requirements for the award of the degree of Master of Engineering (Biomedical)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

MAC 2014

TO AYAH, MOHD DAUD OTHMAN TO IBU, NGAMINI MARDIMIN AND TO ALL MY BROTHERS; MUHAMMAD ALIF SULAIMAN AHMAD DANIAL

WITH ALL MY LOVE

ACKNOWLEDGEMENT

In The Name of Allah, The Most Gracious, The Most Merciful

"Say: If the sea were ink for the words of my Lord, the sea would surely be consumed before the words of my lord are exhausted, though we were to bring the like of that (sea) to add" (Surah Al Kahf: 109)

First and foremost, I would like to express my deepest gratitude to Allah S.W.T for His guidance and blessings throughout my life and the gift of strength in completing my thesis.

In particular, I wish to express my highest gratitude to my Master Degree Supervisor, Dr Hendra Hermawan for his critics, encouragement, consultations and funding throughout completing my experiment and thesis. Not forgetting my cosupervisor, Dr Asmah Hamid (UKM) and Prof Ir Dr Rafiq for their time and cooperation.

Special thanks goes to TCERG and MEDITEG group members especially kak Samsiah, for their support and informative discussion. My sincere appreciation extends to those who have provided assistance at various occasions. Their opinion and views are useful indeed.

Last but not least, I am also indebted to all my lovely family members especially my father, Mohd Daud Othman, my mother, Ngamini Mardimin and all my brothers for giving me such a valuable support and motivation.

May Allah gives His blessings to all of us and only Him could repay my debt to all of you. Amin.

And say, Lord increase my knowledge (Surah Taha: 114)

ABSTRACT

Pure iron (Fe) and its alloys have been recently emphasized as potential biodegradable metals due to their good mechanical properties that are close to those of stainless steel 316L. This research was focused more on the study of cell-material interaction and to analyze the effect of corrosion product on cell behavior by performing degradation study. In this study, samples were prepared by coating hydroxyapatite (HA) and hydroxyapatite/poly (E-Caprolactone) (HA/PCL) onto porous iron using dip coating method. Biosafety and biofunctionality of the sample were evaluated by using human skin fibroblast (HSF) and mesenchymal stem (MSC) cells. Analysis by Inductively Coupled Plasma- Mass Spectrometry (ICP-MS) revealed that concentration of ion Fe was decreased in the medium containing HAcoated Fe. However, the weight loss of the sample is high compared to pure porous iron and HA/PCL-coated Fe. A positive cell response to the Fe ions was revealed during the first 21 days of the cell toxicity study using indirect method. After 21 days the HSF cell viability decreased due to acidic eluates and the increase of Fe ions concentration that promoted the formation of the reactive oxygen intermediates (ROI). From the results obtained, it showed that the HSF and MSC cells exhibited higher viability when in contact with the Fe-HA and Fe-PCL/HA than with the Fe specimens. However, there is a significant decrease (p<0.05) of cells when cultured on three different samples after 3 days of incubation. HA-coated porous Fe also provides support for attachment of the cells. Observation under Scanning Electron Microscope (SEM) reveals that the filopodia of the mesenchymal stem cells preferred to develop onto irregular surface of HA-coated Fe. This study provided evidences of a good cell-material interaction on the porous Fe that may confirm the feasibility of using porous biodegradable ferum as hard tissue scaffolds.

ABSTRAK

Ferum tulen dan ferum berasaskan aloi telah dikenal pasti mempunyai potensi sebagai material terbiodegradasi kerana ciri mekanikalnya yang seakan-akan menyerupai stainless steel 316L. Namun begitu, kadar terdegradasi metal ini mengambil tempoh masa yang lama. Dalam kajian ini, sampel ferum yang poros dan disaluti dengan hidroksiapatit (HA) serta campuran hidroksiapatit/poly ɛ-kaprolakton (HA/PCL) menggunakan kaedah celup penyalutan (dip coating) telah disediakan. Hasil analisis menggunakan ICP-MS menunjukkan bahawa kepekatan ion Ferum yang diukur dalam medium yang direndam dengan ferum bersalut HA adalah rendah walaupun mempunyai pengurangan berat yang ketara berbanding sampel Ferum tulen yang poros dan sampel Ferum bersalut HA/PCL. Kajian ketoksisitan sel melalui kaedah tidak langsung ke atas ion ferum memberikan respon yang positif sehingga 21 hari. Selepas itu, viabiliti sel HSF menunjukkan penyerosotan disebabkan oleh elut medium yang berasid dan peningkatan kepekatan ion ferum yang menggalakkan pembentukan reactive oxygen intermediate (ROI). Seterusnya, ujian biokompatibiliti material dilaksanakan ke atas sel dengan menggunakan sel fibroblas manusia (HSF) dan sel stem mesenkimal manusia (MSC). Kedua-dua sel ini memberikan peningkatan dalam peratusan viabiliti apabila diuji menggunakan Febersalut HA dan Fe-bersalut HA/PCL. Namun begitu, terdapat penurunan yang signifikan apabila sel dibiarkan selama tiga hari dengan semua sampel secara langsung. Pemerhatian di bawah SEM menunjukkan filopodia sel stem mesenkimal lebih mudah berkembang ke atas permukaan tidak rata pada ferum bersalut HA. Hal ini membuktikan bahawa sampel tersebut memberikan sokongan yang baik untuk pelekatan sel dan mengesahkan kebolehan material ferum terbiodegradasikan yang poros sebagai templat (scaffold) untuk tisu keras.

TABLE OF CONTENTS

CHAPTER	TITLES	PAGE

DECLARATION OF ORIGINALITY	ii
DEDICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	xi
LIST OF FIGURES	xii
LIST OF SYMBOLS AND ABBREVIATIONS	xiv
LIST OF APPENDICES	XV

1 INTRODUCTION

1.1	Background	1
1.2	Problem Statement	3
1.3	Objectives	3
1.4	Significance of Study	4
1.5	Scope of Study	4

2 Literature Review

2.1	Biomaterials in implant study	
	2.1.1 Biological Response to Biomaterials	6
	2.1.2 Requirements of Scaffold	7
	2.1.2.1 Biocompatibility	7
	2.1.2.2 Biodegradability	8
	2.1.2.3 Mechanical Strength	8
	2.1.2.4 Interconnected Pore Structure	9
	2.1.2.5 Surface Modification: Solution coatings	9
2.2	The Needs for Biodegradable Materials	
	2.2.1 Biodegradable Metal: Iron	11
	2.2.2 Biodegradable Ceramic: Hydroxyapatite	13
	2.2.3 Biodegradable Polymer:	14
	Poly (ε-Caprolactone) (PCL)	
2.3	Biocompatibility Assessment	
	2.3.1 Biosafety	16
	2.3.1.1 Cytotoxicity Evaluation	16
	a) Morphological Evaluation	16
	b) Biochemical Test	17
	2.3.2 Biofunctionality	18
	2.3.2.1 Cell-Surface Interaction	19
2.4	The Importance of Cell Culture	
	2.4.1 Relevant cells	21
	i. Human Skin Fibroblast (HSF)	22
	ii. Human Mesenchymal Stem Cells (HMSCs)	22

Materials and Methods

3

3.1	Prepa	ration of Sample	
	3.1.1	Porous iron samples	24
	3.1.2	Preparation of HA suspension	25
	3.1.3	Preparation of HA/PCL suspension	25
	3.1.4	Dip coating process	25
	3.1.5	Sample characterization	26
3.2	Clean	ing and Sterilization	26
3.3	Cell C	ulture System and Maintenance	
	3.3.1	Method of cytocompatibility testing	27
	3.3	3.1.1 Direct Contact Test	27
	3.3	3.1.2 Indirect Contact Test (Elution Assay)	28
	3.3.2	Maintenance of Cell Used	29
	3.3	3.2.1 Human Skin Fibroblast (HSF 1184) Cell	29
		Line	
	3.3	3.2.2 Human Mesenchymal Stem Cell (hMSC)	29
3.4	Static	Immersion Test	
	3.4.1	Degradation kinetics of all samples	30
	3.4	.1.1 Weight loss rate measurement	30
	3.4	.1.2 Determination of iron release	31
3.5	Cytoco	ompatibility of Pure Porous Iron	31
3.6	Cell A	ttachment Analysis	
	3.6.1	Quantitative	32
	3.6.2	Fluorescent staining (Qualitative)	33
3.7	Cell vi	iability	34
3.8	Cell p	roliferation	34

	3.9	Cell Morphology	35
	3.10	Statistical analysis	35
4	Resul	It and Discussion	
	4.1	Morphology and EDX of the sample	36
	4.2	Biocompatibility of Pure Porous Fe	39
	4.3	Degradation Kinetics of Three Samples	42
	4.4	Cytocompatibility using HSF cells	45
	4.5	Cytocompatibility of hMSC	50
5	CON	CLUSIONS	
	5.1	Summary	56
	5.2	Suggestions for Future Work	57
REFERENC	ES		58
APPENDICE	S A-C		69-81

Х

LIST OF TABLES

TABLE NO.	TITLE	PAGE	
2.1	Comparison of different techniques in coating	10	
2.2	Mechanical properties of different biomaterials	12	
2.3	The initial evaluation test	15	
3.1	Extraction guidance with a little modification as provided by ISO	28	

LIST OF FIGURES

FIGURES NO.	TITLES	PAGE
2.1	Hydrophilic or hydrophobic in vitro surfaces when in contact with a cell culture medium (a) Protein adsorption (b) cell adhesion	20
4.1	SEM and EDX of HA coated-Fe	37
4.2	SEM and EDX of HA/PCL-coated Fe	38
4.3	Human Skin Fibroblast (HSF) cell viability and iron concentration after 28 days immersion in medium cell	39
4.4	Micrograph of HSF seeded in well plate view from optical microscope (a-b) HSF seeded with porous pure Fe (c) HSF seeded without sample (control); after 24 hours	41
4.5	Weight loss rate of three different samples in MEM solution	42
4.6	pH value of MEM solution with three different types of samples after 21 days of incubation	43
4.7	Weight loss and determination of Fe in MEM solution condition with porous Fe, HA/PCL coated Fe and HA- coated Fe	44

4.8	The number of attached cells on three types of sample; porous Fe, HA/PCL-coated Fe, HA-coated Fe	46
4.9	Proliferation of HSF 1184 cultured on different types of scaffold ($n=3$) after 2, 3 and 5 days	47
4.10	HSF cell attachment (%) in three different types of samples after 4 hour and 24 hour	48
4.11	Detachment of HSF cells at surface of (a) Porous Fe, (b) HA/PCL-coated Fe (c) HA-coated Fe	49
4.12	Micrograph of MSC seeded in well plate view from optical microscope (a-b) MSC seeded with porous pure Fe (c) MSC seeded without sample (control); after 24 hours	51
4.13	Proliferation of hMSC cultured on different types of scaffold (n=3) after certain days	52
4.14	Morphologies of MSC cells observed by SEM	54

xiii

LIST OF SYMBOLS AND ABBREVIATIONS

SEM	-	Scanning Electron Microscope
EDX	-	Energy Dispersive X-ray spectroscopy
μm	-	micrometer
DNA	-	Deoxyribo Nucleic acid
°C	-	Degree celcius
Mm	-	milimeter
w/v	-	Weight per volume
PBS	-	Phosphate Buffer Saline
Rpm	-	Rotate Per Minute
CO_2	-	Carbon Dioxide
μl	-	microliter
Nm	-	nanometer
Ppm	-	Part Per Million

LIST OF APPENDICES

APPENDIX

TITLE

PAGE

A	Analysis of Metal Ions by ICP-MS and AAS	69
В	Toxicity Testing of Medical Devices	79
С	Journal	81

CHAPTER 1

INTRODUCTION

1.1 Background

In a case of organ function failure, biomedical implant made as a scaffold is needed to augment, repair and replace the function of affected tissue for reconstruction. The three-dimensional (3-D) scaffold should be porous, have a good mechanical strength and able to provide a necessary support for cells to perform its function [1]. Recently, biodegradable biomaterials are becoming interesting research topics due to its ability to support the healing process of a tissue and subsequently degrade on the site of implantation. Thus, degradable material represents a promising future in implant studies as they eliminate the risk of secondary surgeries. Furthermore, it will reduce the risk of refracture and stress shielding.

Studies on degradable materials such as polymer have shown excellent biocompatibility and an optimum degradation rate. However, polymer could not withstand a high strength application [2] which makes it less competence to use in the orthopaedic sector. Currently, metallic materials such as iron (Fe) and magnesium (Mg) have been introduced into the biomedical field and have received incredible interest. Iron has mechanical properties similar to bone and its degradation product is non-toxic to a human body [3].

Hydroxyapatite (HA) is a well known bioactive ceramics with nearly same chemical composition as a human bone. It has excellent biocompatibility and great bonding ability with bone structure [4]. However, shaping and implanting HA are difficult because of its weak and brittle mechanical properties. The use of HA coatings on metallic implants have been reported to stimulate bone healing. During the early stage of implantation, the coating shows improvement in the aspect of rate and strength of the metallic implant. It would speed up the rehabilitation of patients by decreasing the insertion time of implant to final reconstruction [1, 4-7]. To overcome the brittleness issue, double coating of HA with poly (ε-caprolactone) (PCL) were introduced by previous research [8]. Reportedly, the coating will become stable and flexible without experiencing crack or delamination compared to the hydroxyapatite single coating [8].

The relevant test is necessary to study the biosafety and biofunctionality of new devices by biomaterialists. Cell attachment is one of a vital part in determining the biocompatibility of a material upon implantation into the human body. This role will further gives information on the cell migration, differentiation and proliferation, thus making the iron based implant is suitable to be used in bone repair and regeneration.

So far, new interest of HA coatings on biodegradable metallic material was increasing in recent years. However, the rapid corrosion rate of Mg is the main restriction for biomedical application. HA has been shown to have the ability to decrease the corrosion rate and improve the bioactivity of Mg alloy [9-10]. Previous research demonstrates that coating of iron foam with calcium phosphate/chitosan gives mechanically remarkable results as it mimics human bone, which can minimize the stress-shielding effects [11]. Therefore, this research is going to be focused more

on the study of cell-material interaction in porous iron. The porous iron was also coated with HA to see its effect on the cell behavior and its degradation study.

1.2 Problem Statement

Iron has been proposed as the potential biodegradable implant due to its mechanical properties that are similar to stainless steel. However, even it is considered as biocompatible, the range of application is limited by its toxicity at maximum concentration [3, 12]. Hydroxyapatite often stated as an osteoconductive material because of its ability to support bone tissue progression surrounding the implants and to induce fixation via chemical bonding [13]. At this point, this research explored the possibility of using iron for bone applications, whereas its bulk was minimized by forming porous structure. Hydroxyapatite (HA) and poly (ε-caprolactone)/hydroxyapatite (PCL/HA) coating was then applied to improve cell attachment and growth. HA-coated degradable implant with interconnected pores is expected to promote osseointegration without reducing its mechanical properties.

1.3 **Objectives**

In this study, porous Fe was coated with HA using dip coating, a simple and cost effective method. PCL was also used as polymeric binder to improve HA coating on the surface of the porous Fe. The cytocompatibility and degradation behaviour of those materials were investigated *in vitro*. Therefore, the objectives of this research are as follow:

- To develop HA-coated porous Fe, HA/PCL-coated porous Fe and evaluate their degradation behaviour.
- To analyse the cell-material interactions on the developed material by a series of cytocompatibility testing using two types of different cells; human skin fibroblast and human mesenchymal stem cells.

1.4 Significance of Study

Previous studies on iron underlining its degradation behaviour, mechanical properties and its biocompatibility have demonstrated its potential to be developed as degradable metallic biomaterials. Most of them were performed on solid Fe samples. This work was done on porous Fe samples with surface modification. The aims were to explore the biocompatibility of porous Fe, the way cells react and attach to the surface of the porous Fe structure. In addition, the effect of HA coating was investigated based on the behaviour of two different cells.

1.5 Scope of Study

Four scopes have been drawn to achieve the objectives of the research, that is:

- 1) Preparation of sample, where porous Fe coated with HA and PCL/HA by using dip coating method.
- 2) Characterization of material and coating by using SEM and EDX.
- Determination of the degradation behaviour by measuring weight change and concentration of Fe release during immersion tests in cell culture medium.
- 4) Investigation of the effect of degradation towards cell viability, proliferation and morphology by using several assays and microscopic observation.

REFERENCE

- Wang, Y., L. Liu, and S. Guo, *Characterization of biodegradable and cytocompatible nano-hydroxyapatite/polycaprolactone porous scaffolds in degradation in vitro*. Polymer Degradation and Stability, 2010. 95(2): p. 207-213.
- 2. Middleton, J.C. and A.J. Tipton, *Synthetic biodegradable polymers as orthopedic devices*. Biomaterials, 2000. **21**(23): p. 2335-46.
- Zhu, S., et al., Biocompatibility of pure iron: In vitro assessment of degradation kinetics and cytotoxicity on endothelial cells. Materials Science and Engineering: C, 2009. 29(5): p. 1589-1592.
- Miao, X., et al., *Hydroxyapatite coating on porous zirconia*. Materials Science and Engineering: C, 2007. 27(2): p. 257-261.
- 5. Ong, J.L. and D.C.N. Chan, *Hydroxyapatite and Their Use As Coatings in Dental Implants: A Review.* 2000. **28**(5&6): p. 667-707.
- 6. Hung, K.-Y., et al., *Titanium surface modified by hydroxyapatite coating for dental implants*. Surface and Coatings Technology, 2013. **231**(0): p. 337-345.
- Li, T., et al., *Hydroxyapatite coating by dipping method, and bone bonding strength.* Journal of Materials Science: Materials in Medicine, 1996. 7(6): p. 355-357.
- Jo, J.-H., et al., Hydroxyapatite/Poly (ε-Caprolactone) double coating on magnesium for enhanced corrosion resistance and coating flexibility. Journal of biomaterials applications, 2012.
- Salman, S.A., K. Kuroda, and M. Okido, Preparation and Characterization of Hydroxyapatite Coating on AZ31 Mg Alloy for Implant Applications. Bioinorganic Chemistry and Applications, 2013. 2013: p. 6.
- 10. Noorakma, A.W., et al., *Hydroxyapatite-Coated Magnesium-Based Biodegradable Alloy: Cold Spray Deposition and Simulated Body Fluid Studies.* Journal of Materials Engineering and Performance, 2013: p. 1-8.
- Wen, Z., et al., A construction of novel iron-foam-based calcium phosphate/chitosan coating biodegradable scaffold material. Materials Science and Engineering: C, 2013. 33(3): p. 1022-1031.

- Zhang, E., H. Chen, and F. Shen, *Biocorrosion properties and blood and cell compatibility of pure iron as a biodegradable biomaterial*. J Mater Sci Mater Med, 2010. 21(7): p. 2151-63.
- Weng, W., et al., Sol-gel preparation of bioactive apatite films. Surface and Coatings Technology, 2003. 167(2-3): p. 292-296.
- 14. Patrick, C.W., A.G. Mikos, and L.V. McIntire, *Frontiers in tissue engineering*. 1998: Elsevier.
- Chen, G., T. Ushida, and T. Tateishi, *Scaffold Design for Tissue Engineering*. Macromolecular Bioscience, 2002. 2(2): p. 67-77.
- O'Brien, F.J., Biomaterials & scaffolds for tissue engineering. Materials Today, 2011. 14(3): p. 88-95.
- Williams, D.F., *The Williams Dictionary of Biomaterials*. 1999: Liverpool University Press.
- Lim, I.A.L., *Biocompatibility of stent materials*. MIT Undergraduate Research Journal (MURJ), 2004. 11(1): p. 33-37.
- 19. Peters, K., R. Unger, and C.J. Kirkpatrick, *Biocompatibility Testing*, in *Biomedical Materials*, R. Narayan, Editor. 2009, Springer US. p. 261-292.
- Morais, J.M., F. Papadimitrakopoulos, and D.J. Burgess, *Biomaterials/tissue interactions: possible solutions to overcome foreign body response*. AAPS J, 2010. 12(2): p. 188-96.
- 21. Kirkpatrick, C.J., et al., *Tissue response and biomaterial integration: the efficacy of in vitro methods*. Biomol Eng, 2002. **19**(2-6): p. 211-7.
- Michaelis, S., R. Robelek, and J. Wegener, *Studying cell-surface interactions in vitro: a survey of experimental approaches and techniques*. Adv Biochem Eng Biotechnol, 2012. **126**: p. 33-66.
- 23. Temenoff, J.S. and A.G. Mikos, *Biomaterials The Intersection of Biology and Materials Science*. 2008, Houston, Texas: Pearson Education International.
- 24. Anselme, K., Osteoblast adhesion on biomaterials. Biomaterials, 2000.
 21(7): p. 667-681.
- Kumari, T.V., et al., *Cell surface interactions in the study of biocompatibility*.
 Trends Biomaterials of Artificial Organs, 2002. 15(2): p. 37-41.
- 26. Vats, A., et al., *Scaffolds and biomaterials for tissue engineering: a review of clinical applications*. Clin Otolaryngol Allied Sci, 2003. **28**(3): p. 165-72.

- Hutmacher, D.W., Scaffolds in tissue engineering bone and cartilage.
 Biomaterials, 2000. 21(24): p. 2529-2543.
- Kretlow, J.D. and A.G. Mikos, From material to tissue: Biomaterial development, scaffold fabrication, and tissue engineering. AIChE Journal, 2008. 54(12): p. 3048-3067.
- 29. Huber, M., et al., Presence of corrosion products and hypersensitivityassociated reactions in periprosthetic tissue after aseptic loosening of total hip replacements with metal bearing surfaces. Acta Biomater, 2009. 5(1): p. 172-80.
- Julio San Román and R.L. Reis, eds. *Biodegradable Systems in Tissue Engineering and Regenerative Medicine*. Cytotoxicity Screening of Biodegradable Polymeric Systems, ed. Gabriela A. Silva, et al. 2004, CRC Press.
- Mitragotri, S. and J. Lahann, *Physical approaches to biomaterial design*. Nature materials, 2009. 8(1): p. 15-23.
- Karageorgiou, V. and D. Kaplan, *Porosity of 3D biomaterial scaffolds and osteogenesis*. Biomaterials, 2005. 26(27): p. 5474-91.
- Nishiguchi, S., et al., *Alkali- and heat-treated porous titanium for orthopedic implants*. Journal of Biomedical Materials Research, 2001. 54(2): p. 198-208.
- 34. van den Dolder, J., et al., Bone tissue reconstruction using titanium fiber mesh combined with rat bone marrow stromal cells. Biomaterials, 2003.
 24(10): p. 1745-1750.
- Filiaggi, M.J., N.A. Coombs, and R.M. Pilliar, Characterization of the interface in the plasma-sprayed HA coating/Ti-6Al-4V implant system. J Biomed Mater Res, 1991. 25(10): p. 1211-29.
- Fu, Y. and A. Batchelor, *Hot isostatic pressing of hydroxyapatite coating for improved fretting wear resistance*. Journal of materials science letters, 1998. 17(20): p. 1695-1696.
- 37. Herø, H., et al., *Hydroxyapatite coatings on Ti produced by hot isostatic pressing*. Journal of Biomedical Materials Research, 1994. **28**(3): p. 343-348.
- Guo, L. and H. Li, Fabrication and characterization of thin nanohydroxyapatite coatings on titanium. Surface and Coatings Technology, 2004. 185(2–3): p. 268-274.

- Mavis, B. and A.C. Taş, *Dip Coating of Calcium Hydroxyapatite on Ti-6Al-*4V Substrates. Journal of the American Ceramic Society, 2000. 83(4): p. 989-991.
- Sridhar, T., et al., *Electrophoretic deposition of hydroxyapatite coatings and corrosion aspects of metallic implants*. Corrosion reviews, 2002. 20(4-5): p. 255-294.
- Chen, F., et al., Biocompatibility of electrophoretical deposition of nanostructured hydroxyapatite coating on roughen titanium surface: in vitro evaluation using mesenchymal stem cells. J Biomed Mater Res B Appl Biomater, 2007. 82(1): p. 183-91.
- Choudhury, P. and D. Agrawal, Sol-gel derived hydroxyapatite coatings on titanium substrates. Surface and Coatings Technology, 2011. 206(2): p. 360-365.
- 43. Varma, H., S. Kalkura, and R. Sivakumar, *Polymeric precursor route for the preparation of calcium phosphate compounds*. Ceramics international, 1998.
 24(6): p. 467-470.
- 44. Erne, P., M. Schier, and T.J. Resink, *The road to bioabsorbable stents:* reaching clinical reality? Cardiovascular and interventional radiology, 2006.
 29(1): p. 11-16.
- 45. Purnama, A., et al., Assessing the biocompatibility of degradable metallic materials: State-of-the-art and focus on the potential of genetic regulation. Acta Biomaterialia, 2010. 6(5): p. 1800-1807.
- Meyer U., et al., Basic Reactions Of Osteoblasts On Structured Material Surfaces. European Cells and Materials, 2005. 9.
- 47. Athanasiou, K.A., et al., Orthopaedic applications for PLA-PGA biodegradable polymers. Arthroscopy: The Journal of Arthroscopic & Related Surgery, 1998. 14(7): p. 726-737.
- Hermawan, H., Biodegradable Metals: State of the Art, in Biodegradable Metals. 2012, Springer Berlin Heidelberg. p. 13-22.
- 49. Hermawan, H., et al., *Fe-Mn alloys for metallic biodegradable stents: degradation and cell viability studies.* Acta Biomater, 2010. **6**(5): p. 1852-60.
- 50. Staiger, M.P., et al., *Magnesium and its alloys as orthopedic biomaterials: A review*. Biomaterials, 2006. **27**(9): p. 1728-1734.

- Papanikolaou, G. and K. Pantopoulos, *Iron metabolism and toxicity*. Toxicol Appl Pharmacol, 2005. 202(2): p. 199-211.
- 52. Peuster, M., et al., Long-term biocompatibility of a corrodible peripheral iron stent in the porcine descending aorta. Biomaterials, 2006. 27(28): p. 4955-4962.
- 53. Stohs, S. and D. Bagchi, Oxidative mechanisms in the toxicity of metal ions.
 Free Radical Biology and Medicine, 1995. 18(2): p. 321-336.
- 54. Chuenjitkuntaworn, B., et al., Polycaprolactone/hydroxyapatite composite scaffolds: Preparation, characterization, and in vitro and in vivo biological responses of human primary bone cells. Journal of Biomedical Materials Research Part A, 2010. 94A(1): p. 241-251.
- 55. Denissen, H.W., et al., *Tissue response to dense apatite implants in rats.* Journal of Biomedical Materials Research, 1980. **14**(6): p. 713-721.
- 56. Petit, R., *The use of hydroxyapatite in orthopaedic surgery: A ten-year review*. European Journal of Orthopaedic Surgery & Traumatology, 1999.
 9(2): p. 71-74.
- Jaffe, W. and D. Scott, *Total Hip Arthroplasty with Hydroxyapatite-Coated Prostheses*, in *Joint Arthroplasty*, S. Imura, M. Wada, and H. Omori, Editors. 1999, Springer Japan. p. 159-187.
- Shen, Z., et al., Dense hydroxyapatite-zirconia ceramic composites with high strength for biological applications. Advanced Materials, 2001. 13(3): p. 214-216.
- 59. Khor, H.L., et al., Poly(ε-caprolactone) films as a potential substrate for tissue engineering an epidermal equivalent. Materials Science and Engineering: C, 2002. 20(1-2): p. 71-75.
- Hutmacher, D.W., et al., Mechanical properties and cell cultural response of polycaprolactone scaffolds designed and fabricated via fused deposition modeling. Journal of Biomedical Materials Research, 2001. 55(2): p. 203-216.
- 61. Ng, K.W., et al., *Evaluation of ultra-thin poly (ε-caprolactone) films for tissue-engineered skin.* Tissue engineering, 2001. 7(4): p. 441-455.
- Yoon, C. and D. Ji, Effects of In Vitro degradation on the weight loss and tensile properties of PLA/LPCL/HPCL blend fibers. Fibers and Polymers, 2005. 6(1): p. 13-18.

- Rohner, D., et al., *In vivo efficacy of bone-marrow-coated polycaprolactone* scaffolds for the reconstruction of orbital defects in the pig. Journal of Biomedical Materials Research Part B: Applied Biomaterials, 2003. 66(2): p. 574-580.
- 64. Ciapetti, G., et al., Osteoblast growth and function in porous poly epsilon caprolactone matrices for bone repair: a preliminary study. Biomaterials, 2003. 24(21): p. 3815-24.
- 65. Coombes, A.G.A., et al., Precipitation casting of polycaprolactone for applications in tissue engineering and drug delivery. Biomaterials, 2004.
 25(2): p. 315-325.
- 66. Causa, F., et al., Poly-ε-caprolactone/hydroxyapatite composites for bone regeneration: In vitro characterization and human osteoblast response. Journal of Biomedical Materials Research Part A, 2006. 76(1): p. 151-162.
- Kim, H.-W., J.C. Knowles, and H.-E. Kim, *Hydroxyapatite/poly* (< i> ε</i> caprolactone) composite coatings on hydroxyapatite porous bone scaffold for drug delivery. Biomaterials, 2004. 25(7): p. 1279-1287.
- 68. Gu, X., et al., *In vitro corrosion and biocompatibility of binary magnesium alloys.* Biomaterials, 2009. **30**(4): p. 484-498.
- 69. Bruinink, A. and R. Luginbuehl, *Evaluation of biocompatibility using in vitro methods: interpretation and limitations*. Adv Biochem Eng Biotechnol, 2012.
 126: p. 117-52.
- Wazen, R.M., et al., *Initial evaluation of bone ingrowth into a novel porous titanium coating*. J Biomed Mater Res B Appl Biomater, 2010. 94(1): p. 64-71.
- Mueller, P.P., et al., Control of smooth muscle cell proliferation by ferrous iron. Biomaterials, 2006. 27(10): p. 2193-2200.
- 72. ISO-10993-5, *Biological Evaluation of medical devices-part 5: Test for cytotoxicity:in vitro methods.* 1992, ANSI/AAMI: Arlington.
- 73. 07, A.-F.-. Standard Practice for Direct Contact Cell Culture Evaluation of Materials for Medical Devices. 2007.
- Kirkpatrick, C.J., et al., *Current trends in biocompatibility testing*. Proc Inst Mech Eng H, 1998. 212(2): p. 75-84.
- Cheung, S., et al., *Fibroblastic interactions with high-porosity Ti-6Al-4V* metal foam. J Biomed Mater Res B Appl Biomater, 2007. 82(2): p. 440-9.

- 76. Crawford, J.M., et al., Vital fluorescent staining of human endothelial cells, fibroblasts, and monocytes: assessment of surface morphology. Ann Thorac Surg, 1989. 48(3 Suppl): p. S100-1.
- 77. Ciapetti, G., et al., *In vitro evaluation of cell/biomaterial interaction by MTT assay.* Biomaterials, 1993. **14**(5): p. 359-364.
- Sjogren, G., G. Sletten, and J.E. Dahl, *Cytotoxicity of dental alloys, metals, and ceramics assessed by millipore filter, agar overlay, and MTT tests.* J Prosthet Dent, 2000. 84(2): p. 229-36.
- 79. Fotakis, G. and J.A. Timbrell, *In vitro cytotoxicity assays: comparison of LDH, neutral red, MTT and protein assay in hepatoma cell lines following exposure to cadmium chloride*. Toxicol Lett, 2006. **160**(2): p. 171-7.
- 80. Yun, Y., et al., *Biodegradable Mg corrosion and osteoblast cell culture studies*. Materials Science and Engineering: C, 2009. **29**(6): p. 1814-1821.
- Mosmann, T., Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. Journal of Immunological Methods, 1983. 65(1–2): p. 55-63.
- Weyermann, J., D. Lochmann, and A. Zimmer, *A practical note on the use of cytotoxicity assays*. Int J Pharm, 2005. 288(2): p. 369-76.
- 83. Baxter, L.C., et al., *Fibroblast and osteoblast adhesion and morphology on calcium phosphate surfaces*. Eur Cell Mater, 2002. **4**: p. 1-17.
- 84. Hacking, S.A., et al., *Fibrous tissue ingrowth and attachment to porous tantalum*. Journal of Biomedical Materials Research, 2000. **52**(4): p. 631-638.
- 85. Chen, R.-S., et al., *Cell-surface interactions of rat tooth germ cells on various biomaterials*. Journal of Biomedical Materials Research Part A, 2007.
 83A(1): p. 241-248.
- 86. Kirkpatrick, C.J., et al., *In vitro methodologies to evaluate biocompatibility: status quo and perspective*. ITBM-RBM, 2005. **26**(3): p. 192-199.
- 87. Pizzoferrato, A., et al., Cell culture methods for testing Biocompatibility. Clinical Materials, 1994. 15(3): p. 173-190.
- Sultana, N. and M. Wang, PHBV/PLLA-based composite scaffolds fabricated using an emulsion freezing/freeze-drying technique for bone tissue engineering: surface modification and in vitro biological evaluation. Biofabrication, 2012. 4(1): p. 015003.

- 89. Farack, J., et al., The effect of perfusion culture on proliferation and differentiation of human mesenchymal stem cells on biocorrodible bone replacement material. Materials Science and Engineering: B, 2011. 176(20): p. 1767-1772.
- Schinhammer, M., et al., On the cytocompatibility of biodegradable Fe-based alloys. Materials Science and Engineering: C, 2013. 33(2): p. 782-789.
- Rittié, L. and G. Fisher, *Isolation and Culture of Skin Fibroblasts*, in *Fibrosis Research*, J. Varga, D. Brenner, and S. Phan, Editors. 2005, Humana Press. p. 83-98.
- 92. Kassem, M., Mesenchymal stem cells: biological characteristics and potential clinical applications. Cloning Stem Cells, 2004. 6(4): p. 369-74.
- 93. Eslaminejad, M.B. and F. Faghihi, Mesenchymal Stem Cell-Based Bone Engineering for Bone Regeneration. Regenerative Medicine and Tissue Engineering - Cells and Biomaterials. 2011.
- 94. Bruder, S.P., N. Jaiswal, and S.E. Haynesworth, Growth kinetics, selfrenewal, and the osteogenic potential of purified human mesenchymal stem cells during extensive subcultivation and following cryopreservation. J Cell Biochem, 1997. 64(2): p. 278-94.
- 95. Abdallah, B., H. Saeed, and M. Kassem, Human Mesenchymal Stem Cells: Basic Biology and Clinical Applications for Bone Tissue Regeneration, in Trends in Stem Cell Biology and Technology, H. Baharvand, Editor. 2009, Humana Press. p. 177-190.
- 96. Augello, A., T.B. Kurth, and C. De Bari, Mesenchymal stem cells: a perspective from in vitro cultures to in vivo migration and niches. Eur Cell Mater, 2010. 20: p. 121-33.
- 97. Zhang, D. and K.A. Kilian, *The effect of mesenchymal stem cell shape on the maintenance of multipotency*. Biomaterials, 2013. **34**(16): p. 3962-3969.
- 98. Kon, E., et al., Autologous bone marrow stromal cells loaded onto porous hydroxyapatite ceramic accelerate bone repair in critical-size defects of sheep long bones. J Biomed Mater Res, 2000. 49(3): p. 328-37.
- 99. Tang, Z.G., et al., Surface properties and biocompatibility of solvent-cast poly[-caprolactone] films. Biomaterials, 2004. 25(19): p. 4741-8.

- Sukmana, I. and P. Vermette, *Polymer fibers as contact guidance to orient microvascularization in a 3D environment*. J Biomed Mater Res A, 2010.
 92(4): p. 1587-97.
- 101. Schaffer, J.E., E.A. Nauman, and L.A. Stanciu, Cold drawn bioabsorbable ferrous and ferrous composite wires: an evaluation of in vitro vascular cytocompatibility. Acta Biomater, 2013. 9(10): p. 8574-84.
- 102. Kirkland, N.T., et al., Buffer-regulated biocorrosion of pure magnesium. J
 Mater Sci Mater Med, 2012. 23(2): p. 283-91.
- 103. Elshahawy, W.M., I. Watanabe, and P. Kramer, < i> In vitro</i> cytotoxicity evaluation of elemental ions released from different prosthodontic materials. Dental materials, 2009. 25(12): p. 1551-1555.
- Hänzi, A.C., et al., On the in vitro and in vivo degradation performance and biological response of new biodegradable Mg-Y-Zn alloys. Acta Biomaterialia, 2010. 6(5): p. 1824-1833.
- 105. Lee, S.K., et al., Differential expression of cell surface proteins in human bone marrow mesenchymal stem cells cultured with or without basic fibroblast growth factor containing medium. PROTEOMICS, 2009. 9(18): p. 4389-4405.
- Jansen, J.A., J.P.C.M. van der Waerden, and K. de Groot, *Fibroblast and epithelial cell interactions with, surface-treated implant materials*. Biomaterials, 1991. 12(1): p. 25-31.
- Mascotti, K., J. McCullough, and S.R. Burger, *HPC viability measurement:* trypan blue versus acridine orange and propidium iodide. Transfusion, 2000.
 40(6): p. 693-6.
- 108. Peuster, M., C. Fink, and C. von Schnakenburg, Biocompatibility of corroding tungsten coils: in vitro assessment of degradation kinetics and cytotoxicity on human cells. Biomaterials, 2003. 24(22): p. 4057-4061.
- Bodhak, S., S. Bose, and A. Bandyopadhyay, *Electrically polarized HAp-coated Ti: in vitro bone cell-material interactions*. Acta Biomater, 2010. 6(2): p. 641-51.
- 110. Jo, J.H., et al., *Hydroxyapatite/ Poly ({varepsilon}-Caprolactone) double coating on magnesium for enhanced corrosion resistance and coating flexibility.* J Biomater Appl, 2012.

- 111. Dabrowski, B., et al., *Highly porous titanium scaffolds for orthopaedic applications*. J Biomed Mater Res B Appl Biomater, 2010. **95**(1): p. 53-61.
- Song, L., et al., Antibacterial hydroxyapatite/chitosan complex coatings with superior osteoblastic cell response. Materials Letters, 2011. 65(6): p. 974-977.
- Moravej, M. and D. Mantovani, *Biodegradable metals for cardiovascular* stent application: interests and new opportunities. International journal of molecular sciences, 2011. 12(7): p. 4250-4270.
- 114. Moravej, M., et al., *Electroformed pure iron as a new biomaterial for degradable stents: In vitro degradation and preliminary cell viability studies.*Acta Biomaterialia, 2010. 6(5): p. 1843-1851.
- 115. Cheng, J., et al., Comparative in vitro Study on Pure Metals (Fe, Mn, Mg, Zn and W) as Biodegradable Metals. Journal of Materials Science & Technology, 2013. 29(7): p. 619-627.
- 116. Xu, L., E. Zhang, and K. Yang, *Phosphating treatment and corrosion properties of Mg–Mn–Zn alloy for biomedical application*. Journal of Materials Science: Materials in Medicine, 2009. 20(4): p. 859-867.
- 117. Song, G., Control of biodegradation of biocompatable magnesium alloys. Corrosion Science, 2007. 49(4): p. 1696-1701.
- Song, G. and S. Song, A Possible Biodegradable Magnesium Implant Material. Advanced Engineering Materials, 2007. 9(4): p. 298-302.
- 119. Liu, B. and Y.F. Zheng, Effects of alloying elements (Mn, Co, Al, W, Sn, B, C and S) on biodegradability and in vitro biocompatibility of pure iron. Acta Biomaterialia, 2011. 7(3): p. 1407-1420.
- 120. Li, H., Y. Chen, and Y. Xie, *Photo-crosslinking polymerization to prepare polyanhydride/needle-like hydroxyapatite biodegradable nanocomposite for orthopedic application*. Materials Letters, 2003. 57(19): p. 2848-2854.
- Bogdanski, D., et al., *Biocompatibility of calcium phosphate-coated and of geometrically structured nickel-titanium (NiTi) by in vitro testing methods*. Materials Science and Engineering: A, 2004. 378(1–2): p. 527-531.
- Rastegar, F., et al., Mesenchymal stem cells: Molecular characteristics and clinical applications. World J Stem Cells, 2010. 2(4): p. 67-80.

- 123. Vacanti, V., et al., *Phenotypic changes of adult porcine mesenchymal stem cells induced by prolonged passaging in culture*. J Cell Physiol, 2005. 205(2): p. 194-201.
- 124. Li, Y., et al., Synthesis and cytocompatibility of manganese (II) and iron (III) substituted hydroxyapatite nanoparticles. Journal of Materials Science, 2012.
 47(2): p. 754-763.
- 125. Yu, S., et al., Biocompatibility and osteoconduction of active porous calciumphosphate films on a novel Ti-3Zr-2Sn-3Mo-25Nb biomedical alloy. Colloids Surf B Biointerfaces, 2011. 85(2): p. 103-15.
- 126. Xu, L.-p., E.-I. Zhang, and K. Yang, *Biocorrosion property and cytocompatibility of calcium phosphate coated Mg alloy*. Transactions of Nonferrous Metals Society of China, 2012. 22(8): p. 2014-2020.
- 127. Abdal-hay, A., M. Dewidar, and J.K. Lim, Biocorrosion behavior and cell viability of adhesive polymer coated magnesium based alloys for medical implants. Applied Surface Science, 2012. 261(0): p. 536-546.