OPTIMIZATION OF *ORTHOSIPHON STAMINEUS*-LOADED NANOSTRUCTURED LIPID CARRIER USING D-OPTIMAL MIXTURE DESIGN FOR IMPROVED LIPOLYSIS ACTIVITY

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This thesis is dedicated to my parents and family

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ABSTRACT

Orthosiphon stamineus (OS) is a Malaysian medicinal herb that was reported to have weight reduction effect and normally prepared as herbal infusion. Effective topical utilization of OS requires a good drug delivery system in order to overcome the stratum corneum to reach the targeted area. Nanostructured lipid carrier (NLC) is a promising carrier for topical drug delivery. The goal of the present study was to optimize the formulation of OS-loaded nanostructured lipid carrier (OS-NLC) for improved lipolysis activity. OS-NLC was prepared using melt emulsification homogenization technique with different types of lipid to obtain good homogeneity and miscibility of the formulation. The result revealed the preferred selection of glyceryl monostearate as solid lipid and triglyceride as liquid lipid since they showed good homogeneity. The formulation of OS-NLC was optimized using D-optimal mixture design which consisted of an amount of active ingredients, solid lipid, and liquid lipid as independent variables while particle size, polydispersity index (PDI) and encapsulation efficiency as dependent variables. From the study, it was found that the optimum formulation of OS-NLC was made of 4% OS, 1% glyceryl monostearate, and 5% triglyceride with 88.57 ± 1.187 nm particle size, 0.135 ± 0.007 PDI and 98.10 \pm 1.101 % encapsulation efficiency. Coefficient of determination (R²) indicated a good fit between predicted values and the experimental data points for particle size, PDI and encapsulation efficiency, which were found to be 0.9404, 0.9138 and 0.8754, respectively. Transmission electron microscopy images exhibited the spherical shape of OS-NLC. Fourier transform infrared spectroscopy analysis demonstrated an interaction between OS extract and NLC system. The zeta potential of OS-NLC was -16.7 ± 0.5033 mV. Storage stability of OS-NLC was conducted at cold and room temperature for one month by measuring the particle size, PDI and zeta potential. The results revealed that there were no significant changes in particle size, PDI and zeta potential within one month. The in vitro penetration study using Franz diffusion cell showed that the penetration flux of OS-NLC (2188.74 µg cm⁻¹h⁻ ¹) was significantly higher than OS (1614.20 μ g cm⁻¹h⁻¹). The result proved that NLC encapsulate OS had better delivery system compared to OS extract. In lipolysis study, OS-NLC was found to stimulate the release of glycerol in 3T3-L1 adipocytes cells. Taken together, the optimal OS-NLC formulation had efficiently enhanced penetration into skin and improved the lipolysis activity in adipocyte cell.

ABSTRAK

Orthosiphon stamineus (OS) adalah herba perubatan Malaysia yang dilaporkan mempunyai kesan penurunan berat badan dan biasanya digunakan sebagai infusi herba. Penggunaan berkesan OS memerlukan sistem penghantaran ubat yang baik untuk mengatasi lapisan stratum corneum. Pembawa lipid berstruktur nano (NLC) adalah pembawa yang berkebolehan untuk penghantaran ubat topikal. Matlamat kajian ini adalah untuk mengoptimumkan formulasi OS dimuatkan ke dalam pembawa lipid berstruktur nano (OS-NLC) bagi meningkatkan meningkatkan aktiviti lipolisis dalam sel lemak. OS-NLC dihasilkan daripada teknik penyeragaman pengemulsian lebur dengan menggunakan pelbagai jenis lipid untuk mendapatkan keseragaman dan kebolehcampuran formulasi yang baik. Hasil kajian mencadangkan pilihan gliseril monostearat sebagai lipid pepejal dan trigliserida sebagai lipid cecair lebih diutamakan kerana mereka menunjukkan keseragaman yang baik. Formulasi OS-NLC dioptimumkan dengan menggunakan reka bentuk campuran D-optimum yang terdiri daripada bahan aktif, lipid pepejal dan lipid cecair sebagai pemboleh ubah tak bersandar manakala pemboleh ubah bersandar terdiri daripada saiz partikel, indeks kepoliserakan (PDI) dan kecekapan pengkapsulan. Berdasarkan keputusan kajian, formulasi optimum OS-NLC terdiri daripada 4% OS, 1% gliseril monostearat, dan 5% trigliserida dengan saiz partikel 88.57 \pm 1.187 nm, 0.135 \pm 0.007 PDI dan 98.10 \pm 1.101% kecekapan enkapsulasi. Pekali penentuan (R²) menunjukkan kesesuaian antara nilai ramalan dan data eksperimen untuk saiz partikel, PDI dan kecekapan enkapsulasi masing-masing adalah 0.9404, 0.9138 dan 0.8754. Imej mikroskop elektron transmisi menunjukkan OS-NLC berbentuk sfera. Spektroskopi inframerah transformasi Fourier menunjukkan interaksi antara ekstrak OS dan sistem NLC. Potensi zeta bagi OS-NLC ialah -16.7 ± 0.5033 mV. Kestabilan penyimpanan OS-NLC telah dijalankan pada suhu sejuk dan bilik selama satu bulan dengan mengukur saiz partikel, PDI dan potensi zeta. Hasil kajian mendapati tiada perubahan signifikan dalam saiz partikel, PDI dan potentsi zeta dalam masa satu bulan. Kajian penembusan transdermal in vitro menggunakan sel penyebaran Franz menunjukkan bahawa fluks penembusan OS-NLC $(2188.74 \ \mu g \ cm^{-1}h^{-1})$ adalah lebih tinggi daripada OS $(1614.20 \ \mu g \ cm^{-1}h^{-1})$. Hasilnya membuktikan bahawa NLC merangkumi OS menunjukkan sistem penghantaran yang lebih baik berbanding dengan ekstrak OS. Dalam kajian lipolisis, OS-NLC didapati boleh melepaskan gliserol dalam sel adiposit 3T3-L1. Secara keseluruhannya, formulasi OS-NLC yang optimum adalah cekap dalam meningkatkan penembusan ke dalam kulit dan meningkatkan aktiviti lipolysis dalam sel adiposit.

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LIST OF ABBREVATIONS

ANOVA	-	Analysis of variance
ATL	-	Adipose triglyceride lipase
DEX	-	Dexamethasone
DMEM	-	Dulbelcco's modified Eagle's medium
DSC	-	Differential scanning calorimetry
EE	-	Encapsulation efficiency
EUP	-	Eupatorin
FBS	-	Fetal bovine serum
FTIR	-	Fourier transform infrared spectroscopy
GMS	-	Glyceryl monostearate
HPLC	-	High performance liquid chromatography
HSL	-	Hormone sensitive lipase
IBMX	-	3-isobutyl-1-methylxanthine
MAG	-	Monoacylglycerol lipase
MTT	-	3-(4,5-dimethylthiazol-2-Yl)-2,5-diphenyltetrazolium bromide
NLC	-	Nanostructured lipid carrier
OS	-	Orthosiphon stamineus
PBS	-	Phosphate buffered saline
PDI	-	Polydispersity index
PKA	-	Protein kinase A
PKG	-	Protein kinase G
RA	-	Rosmarinic acid
SLN	-	Solid lipid nanoparticles
SN	-	Sinensetin

TMF	-	3'-hydroxy-5,6,7,4'-tetramethoxyflavone
ZP	-	Zeta potential

LIST OF SYMBOLS

cm	-	Centimeter
µg/ml	-	Microgram per mililitre
%	-	Percent
°C	-	Degree celcius
mg/ml	-	Miligram per mililitre
μL	-	Microlitre
nm	-	Nanometer
mM	-	Milimolar
μΜ	-	Micromolar
nmole	-	Nanomole
mm	-	Milimetre
ml	-	Mililitre
rpm	-	Revolutions per minute
g	-	Gram

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

The largest organ in the human body is the skin. The function of skin is to protect the body from any unwanted influences from the environment. The skin consists of the epidermis, dermis and hypodermis. Stratum corneum is the outermost layer of the human skin. It is known that stratum corneum acts as a skin barrier function. A successful transdermal drug delivery must prevail over the stratum corneum. Thus, many approaches have been done to overcome the barrier function and enhance drug transport into the skin (Ghafourian *et al.*, 2004)

Nanotechnology or nanoparticles used as drug delivery vehicle generally have particle size less than 100nm. Nanoparticles are effective transport and delivery system since it improve the bioavaibility of drugs and provide the necessary protection of drugs molecule. Lipid nanoparticles such liposome, nanoemulsion, microemulsion, solid lipid nanoparticles, and nanostructured lipid carrier are gaining a lot of interest as a vehicle for controlled released of active substances and targeting to skin layer. In the beginning of 1990s, solid lipid nanoparticles (SLN) were developed as a carrier system to substitute emulsion, liposomes and polymeric nanoparticles (Muller *et al.*, 2002). SLN is produced by solid lipid only. However, SLN produced a perfect crystal lattice which provides limited space to hold the active, hence causes expulsion of active from the lipid matrix during storage (Bunjes and Koch, 1996).

Nanostructured lipid carrier (NLC) has been introduced to overcome the limitation of solid lipid nanoparticles. Components of NLC are solid lipid, liquid lipid, surfactants and water. NLC is produced by blending both solid and liquid lipids. The combination of solid and liquid lipids alters the formation of perfect crystal thus the matrix is in imperfect form and provides space to hold the active in the lipid matrix. In view of topical administration, NLC has occlusive properties and can reduce transepidermal water loss (TEWL) thus enhance penetration of active ingredient through the stratum corneum by increased hydration (Uprit *et al.*, 2013). NLC offers potential advantages such as protection of active ingredients against degradation, controlled and sustained release of active drug can be achieved, high drug loading capacity, and improve skin permeation (Patel *et al.*, 2013).

Having an excess body fat will lead to several health risk such as being overweight. Adipocytes acts as cells that stored energy as fat. Three different types of adipocytes are white adipocytes, brown adipocytes and beige adipocytes. White adipocytes are responsible to store energy and brown adipocytes to dissipate energy in thermogenesis. While the function of beige adipocytes is still not clear (Stephens, 2012).

Lipolysis is the process of breakdown of triglyceride to free fatty acid and glycerol. This process takes place in white adipose tissue. Enzymes are involved in lipolysis process are adipose triglyceride lipase (ATGL), hormone sensitive lipase (HSL) and monoacylglycerol lipase (MAG) (Zechner *et al.*, 2012). In the first step of lipolysis ATGL is responsible to catalyze the conversion of triglyceride into diacylglycerol whereas HSL will hydrolyze diacylglycerol to monoacylglycerol and MAG hydrolyze monoacylglycerol to glycerol. The main pathway for lipolysis process

is through the activation of cAMP dependent protein kinase A (PKA) (Kim *et al.*, 2014).

Orthosiphon stamineus (OS) or commonly known as *Misai kucing* in malay community is a Lamiacae family. OS has been used to treat diabetes, hypertension, menstrual disoder, epilepsy and rheumatic arthritis. Three phytochemical compounds found in different extract of OS are polymethoxylated flavonoids, phenylpropanoids and terpenoids. Major component of aqueous extract in OS is rosmarinic acid which exhibits antioxidant, immunomodulatory and anti-cancer activity (Scheckel *et al.,* 2008; Yam *et al.,* 2009; Ameer *et al.,* 2012). Combination of OS powder with green tea has been utilized for weight reduction effect (Ameer *et al.,* 2012). In addition, previous study by Son *et al.,* (2011) found OS can reduced visceral fat mass and food intake in Sprague-Dawley rats. Study conducted by Sayedan *et al.* (2016) found that the ethanolic extract of OS leaves significantly reduced a gain in body weight which give weight reduction effect in obese mice induced by a high fat diet. This conclude that OS can be a medicinal food for body weight control.

However, most weight reduction drug has been hardly approved by the Food and Drug Administration (FDA) authority, as compared with transdermal patches and beauty creams. In order to loss fat at specific areas, lipolytic agent such as aminophylline and caffeine can be applied to the skin. Aminophylline and caffeine were found to liberate subcutaneous fat, thinning the fat layer and also reducing cellulite (Caruso *et al.*, 2007). Previous study revealed that external medicinal such as topical cream composed of aminophylline possess weight reduction effect (Petrofsky, *et al.*, 2014). Therefore, it was highly potential to use OS as a topical drug to induce lipolysis in adipocyte which located in subcutaneous layer as well as to achieve systemic weight reduction. Formulation of NLC to encapsulate OS is a challenging task due to the trial and error which can be time consuming and required high cost. Optimization provide significant understanding of develop formulation and their properties can be gained. Experimental design plays bigger role in cosmetic formulation because it provides better understanding about the effect of different product formulation on product properties (Rajin *et al.*, 2007). The desired formulation can be achieved as fast as possible with experimental design because it can reduce number of experiments. Several statistical designs available such as response surface methodology (RSM), factorial design, combined and mixture design. For experiments that have various mixed ingredients, mixture design is employ in order to identify ingredients that give effect on the dependent variables and also to determine the optimal mixing ratio that maximizes or minimizes the dependent variables (Choi, 1998). The D optimal design is suitable for highly constraint design and widely used in drug delivery devices (El-Malah *et al.*, 2006).

The present work was aim to produce cost effective of OS-NLC by optimizing the formulation using D-optimal mixture design in order to improve skin penetration and induce lipolysis activity.

1.2 Problem Statement

Topical drug delivery (TDD) can be described as the application of a drug containing formulation directly to the skin. Once topical formulation was applied to the skin, they must interact with the skin condition which can control the rate of release of the compound. TDD is favoured due to the ease of delivery and most importantly TDD have no intervention with gastric and intestinal fluid (Sharma *et al.*, 2013). However, TDD poses a challenge due to the skin barrier. Besides, the protection of active ingredient is a crucial factor in TDD to protect the active ingredient from degradation and successfully delivered the active ingredient to the targeted area.

Nanostructured lipid carriers (NLC) is a lipid based nanoparticles which normally have a particle size less than 100 nm. NLC is considered as a novel drug delivery system to deliver active ingredients with high solubility, stability, effective skin penetration and low irritation. Many studies have found that NLC can increase the encapsulation efficiency due to the involvement of two type of lipids in NLC which gives the imperfection in their structure thus provide more space to accommodate the active ingredient. The addition of liquid lipid is the main factor that contributes higher encapsulation efficiency because of the higher solubility of drugs in liquid lipid in comparison with solid lipid (Muchow *et al.*, 2008). NLC has the ability to incorporate both hydrophilic and lipophilic drugs. Moreover, the main advantage of NLC over other delivery system is the protection of active ingredient from degradation.

Generally OS has been used as diuretic, to treat rheumatism, kidney and bladder inflammation. Most of the compounds found on OS such as terpenoids, polyphenols, orthosiphols were found to have weight reduction effect (Jayaprakasam et al., 2006). Study by Son *et al.*, (2011) found that the OS can reduce appetite and fat deposition, food intake and visceral fat mass in sprague dawley rats which conclude that OS can act as medicinal food application for body weight control. Collectively, it is postulated that OS may play an important role in the treatment of obesity such as numerous invivo study were conducted by using Sprague dawley rats. However, the ability of OS

to act as anti-obesity medicinal by inducing lipolysis through direct action on adipose tissue is still not clear

The most effective way to reduce weight is by exercise and healthy diet. However, with dieting and exercise it is hard to target where fat loss occurs. Fat loss can occur in any area of the body. Therefore, if fat loss is to be targeted at specific areas, lipolytic agent can be applied to the skin. In addition, due to the topical administration of drug are much safer compared with oral medication, an external topical formulation of OS seem to have high potential to be used as a weight reduction treatment. However, the poor skin permeation of OS was a challenge to develop OS as transdermal delivery system.

Therefore, in this study, NLC is chosen as a carrier to deliver the OS to the targeted area due to the ability of NLC to encapsulate and protect OS from degradation and to overcome the stratum corneum. In addition, this study emphasized the production of OS-NLC for cosmeceutical application and reveal that NLC is an effective delivery system that enhanced penetration over stratum corneum.

1.3 Objective

The objective of this research is to optimize the formulation of *Orthosiphon stamineus* loaded nanostructured lipid carrier to enhance penetration through the skin and induced lipolysis in adipocyte

1.4 Scope of the Research

In order to achieve the objective of this study, the following scopes have been identified as below:

- 1. Extraction of OS using maceration method.
- 2. Formulation of OS-NLC using melt emulsification homogenization technique.
- 3. Optimization of OS-NLC using D-optimal mixture design by design expert software.
- Characterization of optimized formulation of OS-NLC using Transmission Electron Microscopy (TEM), Differential Scanning Calorimetry (DSC), Fourier Transform Infrared Spectroscopy (FTIR)
- 5. In vitro penetration study by Franz type diffusion cell
- 6. In vitro cytotoxicity study of OS-NLC on 3T3-L1 preadipocyte using MTT assay
- 7. Investigation on lipolysis induction of OS-NLC on 3T3-L1 adipocytes

REFERENCES

- Adnyana, K., Setiawan, F., and Insanu, M. (2013). From ethnopharmacology to clinical study of Orthosiphon stamineus Benth. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 66–73
- Akowuah, G., Zhari, I., Norhayati, I., Sadikun, A. and Khamsah, S. (2004). Sinensetin, euparotin, 3'-hydroxy-5,6,7,4'-tetramethoxyflavone and rosmarinic acid contents and oxidative effect of *Orthosiphon stamineus* from Malaysia. *Food Chemistry*. 87, 559-566
- Alkalin, O., Akay, K., U., Sennaroglu, B., and Tez, M. (2010). Optimization of chemical admixture for concrete on mortar performance tests using mixture experiments. *Chemometrics and Intelligent Laboratory System*. 104, 233-242
- Almatar, M., Ekal, H. and Rahmat, Z. (2014). A glance on medical applications of Orthosiphon stamineus and some of its oxidative compounds. International Journal of Pharmaceutical Sciences Review and Research. 24 (2), 83-88
- Aisha, A., F., A., Majid, A., M., S., A., & Ismail, Z. (2014). Preparation and characterization of nano liposomes of Orthosiphon stamineus ethanolic extract in soybean phospholipids. *BMC Biotechnology*, 14, 23.
- Ameer, O., Z., Salman, I., M., Asmawi, Z., M., Ibraheem, Z., O. and Yam, M., F. (2012). Orthosiphon stamineus: traditional uses, phytochemistry, pharmacology, and toxicology: a review. Journal of Medicinal Food. 15(8), 1-13.
- Barras, A., Boussekey, L., Courtade, C. and Boukherroub, R. (2013). Hypericinloaded lipid nanocapsules for photodynamic cancer theraphy in vitro. *The Royal Society of Chemistry*. 5, 10562-10572

- Bezaira, V. and Langin, D. (2009). Regulation of adipose tissue lipolysis revisited. *Proceedings of The Nutrition Society*.68 (4), 350-360
- Branno, H. (2016). Subcutaneous tissues. Retrieved from the website: www.verywell.com
- Bhaskar, K., Anbu, J, Ravichandiran, V., Venkateswarlu, V. and Rao, Y.M. (2009). Lipid nanoparticles for transdermal delivery of flurbiprofen: formulation, in vitro, ex vivo and in vivo studies. *Lipids in Health and Disease*. 8, 1-15
- Bolourchian, N., Hadidi, N., Foroutan, S.M., and Shafaghi, B. (2009). Development and optimization of sublingual tablet formulation for physostigmine salicylate. *Acta Pharmaceutica*. 59, 301-312.
- Bolsoni-Lopes, A. and Alonso-Vale, M., I., C. (2015). Lipolysis and lipases in white adipose tissue - An update. *Archives of Endocrinology and Metabolism*. 59(4), 335-342
- Bunjes, H., Westesen, K., and Koch, M. H. J. (1996). Crystallization tendency and polymorphic transitions in triglyceride nanoparticles. *International Journal of Pharmaceutics*, 129(1-2), 159–173.
- Bunjes, H., Koch, M. H. J. and Westesen, K. (2002). Effects of surfactants on the crystallization and polymorphism of lipid nanoparticles. *Progress in Colloid and Polymer Science*. 121, 7-10.
- Caruso, M. K., Pekarvoic, S., Raum, W. J. and Greenway, F. (2007). Topical weight reduction from the waist. *Diabetes, Obesity and Metabolism.* 9(3), 300-303
- Choi, J., Y., Park, Y., S., Kim, Y., J., Won, C., K., Kim, B., R., Son, K., J., Lee, S., H. and Kim, W., Y. (2013). Combined treatment of betullinic acid, a PTP1B inhibitor, with *Orthosiphon stamineus* extract decreases body weight in high-fatfed mice. *Journal of Medicinal Food*. 16(1), 2-8.
- Collier, S. W., Sheikh, N. M., Sakr, A., Lichtin, J. L., Stewart, R. F. and Bronaugh, R. L. (1989). Maintenance of skin viability during in vitro percutaneous absorption/metabolism studies. *Toxicology and Applied Pharmacology*. 99 (3), 522-533
- Dandagi, P. M., Dessai, G. A., Gadad, A., P. and Desai, V. B. (2014). Formulation and evaluation of nanostructured lipid carrier (NLC) of lornoxicam. *International Journal of Pharmacy and Pharmaceutical Sciences*. 6(2): 73-77
- Dingler, A., Blum, R., P., Niehus, Muller, R., H. and Gohla, S. (1999). Solid lipid nanoparticles (SLNTM/LipopearlsTM) a pharmaceutical and cosmetic carrier for

the application of vitamin E in dermal products. *Journal of Microencapsulation*. 16(6), 751-767

- Dijkstra, M. (2001). Computer simulations of charge and steric stabilised colloidal suspension. *Currnet Opinion in Colloid & interface Science*, 6(4), 372-382
- Dubey, A., Prabhu, P., and Kamath, J. V. (2012). Nano Structured lipid carriers : A Novel Topical drug delivery system. *International Journal of PharmTech Research*. 4(2), 705–714.
- El-Malah, Y., Nazzal, S. and Khanfar, N., M. (2006). D-optimal mixture design: optimization of ternary matrix blends for controlled zero-order drug release from oral dosage forms, *Drug Development and Industrial Pharmacy*. 32, 1207–1218
- Emami, J., Rezazadeh, M., Varshosaz, J., Tabbakhian, M., and Aslani, A. (2012). Formulation of LDL targeted nanostructured lipid carrier loaded with paclitaxel: a detailed study of preparation, freeze drying condition and in vivo cytotoxicity. *Journal of Nanomaterials*. 12, 1-10.
- Fang, C.L., Al-Suwayeh, S. A, and Fang, J. Y. (2013). Nanostructured lipid carriers (NLCs) for drug delivery and targeting. *Recent Patents on Nanotechnology*. 7(1), 41–55.
- Gaba, B., Fazli, M., Khan, S., Ali, A., Baboota, S., and Ali, J. (2015). Nanostructured lipid carrier system for topical delivery of terbinafine hydrochloride. *Bulletin of Faulty of Pharmacy, Cairo University.* 52(2), 147-159
- Gadad, A. P., Tigadi, S.G., Dandagi, P. M., Mastiholimath, S. V., and Bolmal, U. B. (2016). Rosuvastatin loaded nanostructured lipid carrier: for enhancement of oral bioavailability. *Indian Journal of Pharmaceutical Education and Research*. 50(4), 605-611
- Ghafourian, T., Zandasrar, P., Hamishekar, H., and Nokhodchi, A. (2004). The effect of penetration enhancers on drug delivery through skin: A QSAR study. *Journal* of Controlled Release, 99(1), 113–125.
- Ghate, V. M., Lewis, S. A., Prabhu, P., Dubey, A., and Patel, N. (2016). Nanostructured lipid carriers for topical delivery of tretinoin. *Europian Journal* of Pharmaceutics and Biopharmaceutics. 108, 253-261
- Girousse, A. and Langin, D. (2012). Adipocytes lipases and lipid droplet-associated proteins: insight from transgenic mouse models. *International Journal of Obesity*. 36 (4), 581-594

- Godin, B. and Touitou, E. (2007). Transdermal skin delivery: predictions for humans from *in vivo*, *ex vivo* and animal models. *Advanced Drug Delivery Reviews*. 59(11), 1152-1161
- Gomes, M. J., Martins, S., Ferreira, D. Segundo, M. A. and Reis, S. (2014). Lipid nanoparticles for topical and transdermal application of alopecia treatment: development, physicochemical characterization, an in vitro release and penetration studies. *International Journal of Nanomedicine*. 8, 2769-2781
- Gopalakannan, S. and Senthilvelan, T. (2013). Application of response surface method on machining of Al-SiC nano-composites. *Measurement*. 46, 2705-2715
- Gramdorfa, S., Hermanna, S., Hentschela, A., Schraderb, K., Muller, R. H., and Kraumed, M. (2008). Crystallized Miniemulsions: Influence of operating parameters during high pressure homogenization on size and shape of particles. *Colloids and Surfaces a-Physicochemical and Engineering Aspects*. 331(1-2), 108-113.
- Haaland, P.D. (1989). "Experimental Design in Biotechnology". Marcel Dekker, New York. 259
- Hadgraft, J. (2004). Skin deep. European Journal of Pharmaceutics and Biopharmaceutics. 58, 291–299.
- Hamishehkar, H., Shokri, J., Fallahi, S., Jahangiri, A., Ghanbarzadeh, S. and Kouhsoltani, M. (2015). Histopathological evaluation of caffeine-loaded solid lipid nanoparticles in efficient treatment of cellulite. *Drug Development and Industrial Pharmacy*. 4 (10), 1640-1646
- Han, F., Li, S., Yin, R., Liu, H. and Xu, L. (2008). Effects of surfactants on the formation and characterization of a new type of colloidal drug delivery system: nanostructured lipid carriers. *Colloids and Surfaces*. 315, 210-126.
- Hasenhuettl, G. L., and Hartel, R. W. (2008). Food Emulsifiers and Their Applications. (2nd ed.) New York: Springer.
- Hejri, A., Khosravi, A., Gharanjiq, K., and Hejazi, M. (2013). Optimization of the formulation of β-carotene loaded nanostructured lipid carriers prepared by solvent diffusion method. *Food Chem.* 141(1), 117-123.
- Hoffman, M. (2014). Picture of the skin. Retrieved from the website: http://www.webmd.com/skin-problems-and-treatments/picture-of-the-skin#1

- Hosny, K. M. (2016). Alendronate sodium as enteric coated solid lipid nanoparticles; preparation, optimization, and in vivo evaluation to enhance its oral bioavailability. *PLoS ONE*, 11 (5), 1-14
- Hossain, M., A. and Ismail, Z. (2012). Quantification and enrichment of sinensetin in the leaves of Orthosiphon stamineus. Arabian Journal of Chemistry. 9, 1338-1341
- How, C. W., Abdullah, R. and Abbasalipourkabir, R. (2011). Physicochemical properties of nanostructured lipid carriers as colloidal carrier system stabilized with polysorbate 20 and polysorbate 80. *African Journal of Biotechnology*. 10(9),1684-1689
- Hua, S., and Wu, S. Y. (2013). The use of lipid-based nanocarriers for targeted pain therapies. *Front Pharmacol.* 4, 143.
- Jang, M. K., Kim, C. H., Seong, J., K. and Jung, M. H. (2012). ATF 3 inhibits adipocytes differentiation on 3T3-L1 cells. *Biochemical and Biophysical Research Communications*. 421, 38-43.
- Jayaprakasam, B., Olson, L., K., Schitzki, R., E., Tai, M., H.and Nair, M., G. (2006). Amelioration of obesity and glucose intolerance in high fat-fed C5BL/6 mice by anthocyanins and ursolic acid in cornelian cherry (cornus mas). *Journal of Agriculture anf Food Chemistry*. 54, 243-248
- Jain, N. K. and Ram, A. (2011). Development and characterization of nanostructured lipid carrier of oral hypoglycemic agent. Selection of surfactants. *International Journal of Pharmaceutical Sciences Review and Research*. 7(2), 125-130
- Jia, L. J., Zhang, D. R., Li, Z. Y., Feng, F. F., Wang, Y. C., Dai, W. T., Duan, C. X., Zhang, Q. (2010). Preparation and characterization of silybin-loaded nanostructured lipid carriers. *Drug Delivery*. 17(1), 11-8.
- Joshi, M., and Patravale, V. (2008). Nanostructured lipid carrier (NLC) based gel celecoxib. *International Journal of Pharmaceutics*. 346,124-132.
- Kamoun A., Chaabouni, M., Sargent, M., Roger PTL (2002). Mixture design applied to the formulation of hydrotropes for liquid detergents. *Chemometrics and Intelligent Laboratory System.* 63, 69-79.
- Kim, H. K., Della-Fera, M., Lin, J., and Baile, C. A. (2006). Docosahexaenoic acid inhibits adipocyte differentiation and induces apoptosis in 3T3-L1 preadipocytes. *The Journal of Nutrition. 136*, 2965–2969.

- Kim, S. O., Sakchaisri, K., Asami, Y., Ryoo, I. J., Choo, S. J., Yoo, I. D., Soung, N. K., Kim, Y. S., Jang, J. H., Kim, B. Y. and Ahn, J. S. (2014). Illudins c2 and C3 stimulate lipolysis in 3T3-L1 adipocytes and supress adipogenesis in 3T3-L1 preadipocytes. *Journal of Natural Products*. 77, 744-750
- Khalil, M., R., Abd-Elbary, A., Kassem, A., M., Ghorab, M., M. and Basha, M. (2014).
 Solid lipid nanoparticles for topical delivery of meloxicam: development and in vitro characterization. 1st Annual International Interdisciplinary Conference, AIIC. April 24-26, 2013. Azores, Portugal. 2013. 779-798
- Krishnaiah, D., Sarbatly, R., Anisuzzaman, S. M., and Madais, E. (2012). Study on car shampoo formulation using D-optimal statistical design. *International Journal of Industrial Chemistry*. 3(31), 1-8.
- Kumari, S., Pandita, D., Poonia. N., and Lather, V. (2016). Nanostructured lipid carriers for topical delivery of an anti-acne drug: characterization and ex vivo evaluation. *Pharmaceutical Nanotechnology*. 3(2), 122-133
- Lafontan, M., and Langin, D. (2009). Lipolysis and lipid mobilization in human adipose tissue. *Progress in Lipid Research*. 48(5), 275–297.
- Loo, C.H., Basri, M., Ismail, R., Lau, H.L.N., Tejo, B.A., Kanthimathi, M.S., Hassan, H.A., and Choo, Y.M. (2013). Effect of composition in nanostructured lipid carrier (NLC) on skin hydration and occlusion. *International Journal of Nanomedicine*. 8, 13-22.
- Lopes, C. P. A. L. (2014). Development and characterization of lipid nanoparticles prepared by miniemulsion technique. Master of Science Degree in Biotechnology, Universidade de Lisboa, Portugal.
- Mader, K., and Menhert, W. (2005). Solid Lipid Nanoparticles: Concepts, Procedures, and Physicochemical Aspects. Lipospheres Drug Targets Delivery: Approaches, methods, and applications. New York: CRC Press.
- El Maghraby, G.M., Barry, B. W. and Williams, A. C. (2008). Liposome and skin: from drug delivery to model membranes. *European Journal of Pharmaeutical Sciences*. 34(4), 203-222.
- Marina, A. M., Man, Y. B.C., Nazimah, S. A. H. and Amin, I. (2009). Chemical properties of virgin coconut oil. *Journal of American Oil Chemists's Society*. 86, 301-307

- Mehnert, W and Mader, K. (2001). Solid lipid nanoparticles: production, characterization and applications. *Advanced Drug Delivery Reviews*. 47(2-3), 165–96.
- Miller, M.A. and Pisani, E. (1999). The cost of unsafe injections. *Bulletin of the World Health Organization*. 77(10), 808-811
- Mishra, S., Kesharwani, R., Tiwari, A. K. and Patel, D. K. (2016). Improvement of drug penetration through the skin by using nanostructured lipid carries (NLC). *International Journal of Pharmacy & Pharmaceutical Research*. 6(3), 481-496.
- Montgomery, D. C. (2009). Design and Analysis of Experiments: Model Adequacy Checking. 7th edition. John Wiley and Sons (Asia) Inc.111 River Street, Hoboken. 75-79
- Moser, K., Kriwet, K., Naik, A., Kalia, Y. N., & Guy, R. H. (2001). Passive skin penetration enhancement and its quantification in vitro. *European Journal of Pharmaceutics and Biopharmaceutics*. *52*, 103–112.
- Muhammad, A. A., Abas, F., Mohammed, A. S. and Ghazali, M. (2013). Anti- and pro-lipase activity of selected medicinal, herbal and aquatic plants and structure elucidation of an anti-lipase compound. *Molecules*. 18, 14651-14669
- Muller, R. H., Radtke, M., and Wissing, S. A. (2002). Solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) in cosmetic and dermatological preparations. *Advanced Drug Delivery Reviews*. 54, S131-S155.
- Muller, R. H., Petersen, R. D., Hommoss, A., and Pardeike, J. (2007). Nanostructured lipid carriers (NLC) in cosmetic dermal products. *Advanced Drug Delivery Reviews*. 59(6), 522–530.
- Muchow, M., Maincent, P., and Muller, R., H. (2008). Lipid nanoparticles with a solid matrix (SLN, NLC, LDC) for oral drug delivery. *Drug Development and Industrial Pharmacy*. 34(12), 1394-1405
- Ng, W., K. Yazan, L. S., Yap, L., H, Hafiza, W., A., G., W., N., How, C., W., and Abdullah, R. (2015). Thymoquinone-loaded nanostructured lipid carrier exhibited cytotoxicity towards breast cancer cell lines (MDA-MB-231 and MCF-7) and cervical cancer cell lines (HeLa and SiHa). *BioMed Research International*. 1-10
- Nik Nurul Najihah binti Nik Mat Daud (2016). *Optimization of process extraction and preparation of anticancer formulation from Annona Muricata leaves*. Master of Engineering, Universiti Teknologi Malaysia, Skudai.

- Nugent, C., Prins. J., Whitehead, J. P., Savage, D., Wenthworth, J.M., Charterjee, V. K., and O'Rahilly, S. (2001). Potentiation of glucose uptake in 3T3-L1 adipocytes by PPAR agonists is maintained in cell expressing a PPAR dominant negative mutant: evidence for selectivity in the downstream responses to PPAR activation. *Journal of Molecular Endocrinol.* 15(10), 1729-1738.
- Otto, T., C., and Lane, M. D. (2005). Adipose development from stem cell to adipocytes. *Critcals Reviews in Biochemistry and Molecular Biology*. 40(4), 229-242.
- Pamudji, J. S., Mauludin, R. and Indriani, N (2016). Development of nanostructured lipid carrier containing of retinyl palmitate. *International Journal of Pharmacy* and Pharmaceutical Sciences. 8(2), 256-260
- Patel, D. K., Tripathy, S., Nair, S.K., and Kesharwani, R. (2013). Nanostructured lipid carrier (NLC) a modern approach for topical delivery: a review. *World Journal* of Pharmacy and Pharmaceutical Sciences. 2(3), 921-938
- Petrofsky, J. S., Layman, M. and Khowailed, I. A. (2014). The effect of slimming system including a lipolytic cream and a isometric abdominal exercised regime and a garment on skin temperature. *The Journal of Applied Research*. 13 (2), 7-11
- Rahman, H. S., Rasedee, A., How, C. W., Abdul, A. B., Zeenathul, N. A., Othman, H. H., Saeed, M. I. and Yeap, S. K. (2013). Zerumbone-loaded nanostructured lipid carriers: preparation, characterization, and antileukemic effect. *International Journal of Nanomedicine*. 8, 2769-2781
- Rahimpour, Y., and Hamishehkar, H. (2012). Liposomes in cosmeceutics. *Expert Opinion on Drug Delivery*. *9*(4), 443–55. doi:10.1517/17425247.2012.666968
- Rajin, M., Bono, A. and Mun. H. C. (2007). Optimization of natural ingredient based lipstick formulation by using mixture design. *Journal of Applied Sciences*. 7, 2099-2103
- Rosli, N. A., Noor, N., M., Aziz, R., A., Aziz, A., A., and Hasham, R. Proceeding of 2nd International Science Postgraduate Conference 2014. Faculty of Science, Universiti Teknologi, Malaysia. 2014. 1-11
- Ruela, A. L. M., Perissinato, A. G., Lino, M. E. S., Mudrik. P. S. and Pereira, G. R. (2016). Evaluation of skin absorption of drugs from topical and transdermal formulation. *Brazilian Journal of Pharmaceutical Sciences*. 52(3), 527-544

- Sangsen, Y., Laochai, P., Chotsathidchai, P. and Wiwattanapatapee, R. (2014). Effect of solid lipid and liquid oil ratios on properties of nanostructured lipid carriers for oral curcumin delivery. *Advanced Materials Research*. 1060, 62-65
- Sayedan, A., Alshwsh, M. A., Alshagga, M. A. and Mohamed, Z. (2016). Antiobesity and lipid lowering effects of *Orthosiphon stamineus* in high-fat diet-induced obese mice. *Planta Medica*. 1-9
- Scheckel, K. A., Degner, S. C., and Romagnolo, D. F. (2008). Rosmarinic acid antagonizes activator protein-1-dependent activation of cyclooxygenase-2 expression in human cancer and nonmalignant cell lines. *The Journal of Nutrition*, 138(August), 2098–2105.
- Selvamuthukumar, S., and Velmurugan, R. (2012). Nanostructured lipid carriers: a potential drug carrier for cancer chemotherapy. *Lipids in Health and Disease*. 11(1), 159.
- Silpa, N., Chakravarthi, N. R., Chandramouli, Y. and Kumar, H. K. P. (2012). Moxifloxacin loaded solid lipid nanoparticles (SLNs). Asian Journal of Pharmaceutical Research. 2(2), 105-112.
- Shah, R., Eldridge, D., Palombo, E. and Harding, I. (2014). Optimisation and stability assessment of solid lipid nanoparticles using particle size and zeta potential. *Journal of Physical Science*. 25(1), 59-75.
- Sharma, A., Saini, S., and Rana, C. (2013). Transdermal Drug Delivery System : A Review. International Journal of Research in Pharmaceutical and Biomedical Sciences. 4(1), 286–292.
- Son, J. Y. (2011). Orthosiphon stamineus Reduces Appetite and Visceral Fat in Rats. Journal of the Korean Society for Applied Biological Chemistry. 54, 200–205.
- Sri Janani, N., Bharath Kumar, G., Balakrishna, P., Sruthi, D., Srujan Kumar, M. and Mantry, S. (2014). A review on Strat-M membrane. *International Journal of Innovative Pharmaceutical Sciences and Research*. 2(4), 962-977
- Susanti, D., Amiroudine, M. Z. A. M., Rezali, M. F. and Taher, M. (2012). Friedelin and lanosterol from *Garcinia prainiana* stimulated glucose uptake and adipocytes differentiation in 3T3-L1 adipocytes. *Natural Product Research*. 27, 1-8.
- Suto, B., Berko, S., Kozma, G., Kukovecz, A., Budai-Szucs, M., Eros, G., Kemeny, L., Sztojkov-Ivanov, A., Gaspar, R., and Csanyi, E. (2016). Development of ibuprofen-loaded nanostructured lipid carrier-based gels: characterization and

investigation of in vitro and in vivo penetration through the skin. *International Journal of Nanomedcine*. 11, 1201-1212

- Stat-Ease, Inc. (2002). "Design-Expert user's guide". Version 6.0.8. Design Expert Software.
- Stephens, J., M. (2012). The fat controller : adipocyte development. *PLoS Biology*. 10(11), 1-3.
- Tengku Muhamad Faris Syafiq bin Tengku Zakaria (2015). Antihyperglycaemic activities of xanthone rich extract of mangosteen (Garcinia Mangostana). Master of Phamaceutical Sciences, International Islamic University, Malaysia.
- Tetyczka, C., Griesbacher, M., Novak, M., Frohlich, E. and Roblegg, E. (2017). Development of nanostructured lipid carriers for intraoral delivery of domperidone. *International Journal of Pharmaceutics*. 526 (1-2), 188-198.
- Thatipamula R. P., Palem C. R., Gannu R., Mudragada S., Yamsani M. R. (2011). Formulation and in vitro characterization of domperidone loaded solid lipid nanoparticles and nanostructured lipid carriers. *DARU Journal of Pharmaceutical Sciences*. 19(1):23–32
- Tiwari, R. and Pathak, K. (2011). Nanostructured lipid carrier versus solid lipid nanoparticles of simvastatin: comparative analysis of characteristics, pharmacokinetics and tissue uptake. *International Journal of Pharmaceutics*. 415, 232-243
- Tortora G, Grabowski S. The Integumentary system. In: Principles of Anatomy and Physiology. 9th edition. John Wiley and son Inc. 2006.
- Triplett M. D., and Rathman J. F. (2008). Optimization of β-carotene loaded solid lipid nanoparticles preparation using a high shear homogenization technique. *Journal of Nanoparticle Research*. 11(3):601–614.
- Trommer, H., and Neubert, R. H. H. (2006). Overcoming the stratum corneum: the modulation of skin penetration. A review. *Skin Pharmacology and Physiology*. 19(2), 106–21.
- Twentyman, P. and Luscombe, M. (1987). A study of some variables in a tetrazolium dye (MTT) based assay for cell growth and chemosensitivity. *British Journal of. Cancer*. 56(3), 279-285
- Uchida, T., Kadhum, W.R., Kanai, S., Todo, H., Oshizaka, T. and Suqibayashi, K. (2015). Prediction of skin permeation by chemical compunds using the artificial

membrane, Strat-MTM. *Europian Journal of Pharmaceutical Sciences*. 67, 113-118

- Uner, M. (2005). Preparation, characterization and physico-chemical properties of solid lipid nanoparticles (SLN) and nanostructured lipid carrier (NLC). Their benefit as collodial drug carrier system. *Instabul University, Faculty of Pharmacy, Department of Pharmaceutical Technology, Instabul Turkey.* 61, 375-386.
- Uprit, S., Sahu., R.K., Roy, M. and Pare, A. (2013). Preparation and characterization of minoxidil loaded nanostructured lipid carrier gel for effective treatment of alopecia. *Saudi Pharmaceutical Journal*. 21(4), 379-385.
- Wastuti Hidayati Suriyah (2012). In vitro study on glucose uptake and insulin stimulating properties of Pluchea Indica (L.) Less. Master of Phamaceutical Sciences, International Islamic University, Malaysia.
- Wiart, C. (2000). Medicinal plant of southeast asia. *Pelanduk Publication Sdn Bhd, Selangor, Malaysia.*
- Wilson, K., J., W., and Waugh A. Ross : Anatomy and Physiology in Health and Illness. 8th Edition. Churchill Livingstone:1996
- Wu, Z., Rosen. E.D., Brun, R., Hauser, S., Adelmant, G., Troy, A. E., McKeon, C., Darlington, G. J., and Spiegelman, B. M. (1999). Cross regulation of C/EBPα and PPARγ controls the transcriptional pathway of adipogenesis and insulin sensitivity. *Molecular Cell*. 3, 151-1589
- Venus, M., Waterman, J., and McNab, I. (2011). Basic physiology of the skin. *Surgery* (*Oxford*). 29(10), 471–474. doi:10.1016/j.mpsur.2011.06.010
- Yam, M. F., Ang, L. F., Basir, R., Salman, I. M., Ameer, O. Z., and Asmawi, M. Z. (2009). Evaluation of the anti-pyretic potential of Orthosiphon stamineus Benth standardized extract. *Inflammopharmacology*, 17(1), 50–54.
- Yuan, H., Wang, L., Du, Y., Hu, F. and Zeng, S. (2007). Preparation and characteristic of nanostructured lipid carriers for control releasing progesterone by melt emulsification. *Colloids and Surfaces B. Biointerfaces*. 60, 174-179.
- Yuniarto, A., Purwani, H., Juanda, D., Setiawan, F. and Kurnia, I. (2015). Kumis kucing (*Orthosiphon stamineus* [Benth]) leaves ethanol extract as anti-obesity agent in high-fat diet-induced obese mice. *Asian Journal of Pharmaceutical and Clinical Research.* 8(6), 234-236

- Yunus, M. A. C., Yaw, L .C. and Idham, Z. (2011). Effects of variables on the production of red-fleshed pitaya powder using response surface methodology. *Jurnal Teknologi*. 56, 15-29.
- Zesbisch, K., Voigt, V., Wabitsch, M., Brandsch, M. (2012). Protocol for effective differentiation on 3T3-L1 cells to adipocytes. *Analytical of Biochemistry*. 425, 88-90
- Zechner, R., Zimmermann, R., Eichman, T. O., Kohlwein, S. D., Haemmerle, G., Lass, A., and Madeo, F. (2012). FAT SIGNALS - Lipases and lipolysis in lipid metabolism and signaling. *Cell Metabolism*. 15(3), 279-291.
- Zhuang, C. Y., Li, N., Wang, M., Zhang, X. N., Pan, W. S. and Peng, J. J. (2010). Preparation and characterization of vinpocetine loaded nanostructured lipid carriers (NLC) for improved oral bioavailability. *International Journal of Pharmaceutics*. 394, 179-185
- Zirak, M. B. and Pezeshki, A. (2015). Effect of surfactant Concentration on the particle size, stability, and potential zeta of beta carotene nano lipid carrier. *International Journal of Current Microbiology and Applied Sciences*. 4(9), 924-932