

OPTIMIZATION OF MEDIUM AND CULTIVATION CONDITIONS FOR
D-LACTIC ACID PRODUCTION USING CASSVA STARCH

RAMZI ATA ABD ALSAHEB

A dissertation submitted in fulfilment of the
requirements for the award of the degree of
Master of Engineering (Bioprocess)

Faculty of Chemical and Energy Engineering
Universiti Teknologi Malaysia

JUNE 2016

ACKNOWLEDGEMENT

In preparing this dissertation, I was in contact with many people, researchers, academicians, and practitioners. They have contributed towards my understanding and thoughts. In particular, I wish to express my sincere appreciation to my main dissertation supervisor, Professor Dr. Hesham A. El Enshasy, for encouragement, guidance, critics and friendship, Without their continued support and interest, this dissertation would not have been the same as presented here.

My fellow postgraduate students should also be recognized for their support. My sincere appreciation also extends to all my colleagues and others who have provided assistance at various occasions. Their views and tips are useful indeed. Unfortunately, it is not possible to list all of them in this limited space. I am grateful to all my family members.

ABSTRACT

In this research, seven standard strains of lactic acid bacteria will be screened at first to evaluate their capability to produce D-lactic acid. Selected for investigation based on their D-lactic acid production capability with three media (MRS, RAM, BASAL). The results indicated that, all strains could produce lactic acid but at different yields. These isolates provided the concentration of lactic acid ranging from 1.24-2.51, 0.44-3.24-0.49-2.10 [g/l] and total acidity expressed ranging from 0.53-1.28, 0.61-1.97 and 0.69-1.58% in the three medium respectively. The maximum D-lactic acid 4.44 [g/L] was obtained at 48 hours fermentation with an initial cassava starch concentration. Therefore, 10 [g/L] of cassava starch concentration was chosen to be used as the inexpensive carbon source in RAM medium for D-lactic acid production by the isolate (*Lactobacillus delbruekii*). For nitrogen sources effect of different nitrogen sources on D-lactic acid production was investigated during this study (*Lactobacillus delbruekii*) was cultivated with 4-5 [g/L] yeast extract, Peptone. Yeast extract showed the higher D- lactic acid production followed by Peptone from 4.30 [g/L], 4.22 [g/L] respectively. For Dipotassium phosphate (K_2HPO_4) source. The maximum yield was 4.75 [g/L] of D-lactic acid was produced. The process optimization will be started by optimization of medium composition using un-optimized medium and optimized medium, followed by complete growth kinetics studies in shake flask level. Result showed The D-lactic acid produced was in un-optimized medium and optimized medium 3.25 [g/L], 5.47 [g/L] respectively. Shake flask level scaled up in 16-L bioreactor for the production of D- lactic acid by using two strategy cultivations, with controlled pH and without controlled pH. The final results after 48 hours cultivation as follows, 9.12 [g/L], 14.25 [g/L] respectively, for the production of D- lactic acid.

ABSTRAK

Dalam kajian ini, tujuh jenis piawain bakteria asid laktik akan disaring pada mulanya untuk menilai keupayaan mereka untuk menghasilkan D-laktik asid. Dipilih untuk penyelidikan berdasarkan keupayaan pengeluaran asid D-laktik mereka dengan tiga media (MRS, RAM, Basal) Keputusan menunjukkan bahawa, semua plawaian bakteria boleh menghasilkan asid laktik tetapi pada kadar hasil yang berbeza. Pencilan menunjukkan kepekatan asid laktik yang terdiri daripada from 1.24-2.51, 0.44-3.24-0.49-2.10 [g/l] dan jumlah keasidan dikeluarkan antara 0.53-1.28, 0.61-1.97 and 0.69-1.58% dalam tiga media masing-masing. D-laktik asid maksimum 4.44 [g/L] telah diperolehi di penapaian 48h dengan kepekatan kanji ubi kayu awal. Oleh itu, 10 [g/L] kepekatan kanji ubi kayu telah dipilih untuk digunakan sebagai sumber karbon murah dalam media RAM untuk D-laktik pengeluaran asid oleh pencilan (*Lactobacillus delbruekii*). Untuk sumber nitrogen kesan sumber nitrogen yang berbeza pada D- pengeluaran asid laktik disiasat semasa kajian ini (*Lactobacillus delbruekii*) telah ditanam dengan 4-5 [g/l] ekstrak yis, pepton. Ekstrak yis menunjukkan D- laktik asid pengeluaran yang lebih tinggi diikuti oleh pepton dari 4.30 [g/L], 4.22 [g/L] masing-masing. Untuk dipotassium fosfat sumber (K_2HPO_4). Hasil maksimum adalah 4.75 [g/L] D-laktik asid dihasilkan. Pengoptimuman proses akan bermula dengan mengoptimumkan komposisi media dengan media tidak-dioptimumkan dan media dioptimumkan, diikuti dengan pertumbuhan kajian Goncang tahap kelalang ditingkatkan. Keputusan menunjukkan Asid D-laktik dihasilkan adalah masing-masing di media un-dioptimumkan dan media dioptimumkan 3.25 [g/L], 5.47 [g/L]. Goncang tahap kelalang ditingkatkan dalam 16-L bioreaktor untuk pengeluaran D- asid laktik dengan menggunakan dua pengkulturan strategi, dengan pH dikawal dan tanpa pH dikawal. Keputusan akhir selepas penanaman 48 jam seperti berikut, 9.12 [g/L], 14.25 [g/L] masing-masing, untuk pengeluaran D- laktik asid.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	ABSTRACT	iii
	ABSTRAK	v
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xi
	LIST OF FIGURES	xiv
	LIST OF ABBREVIATIONS	xvi
	LIST OF SYMBOLS	xvii
	LIST OF APPENDICES	xx
1	INTRODUCTION	1
	1.1 Background of the research	1
	1.2 Problem Statement	3
	1.3 Research objectives	4
	1.4 Scope and limitation of the study	4
2	LITERATURE REVIEW	6
	2.1 Introduction	6
	2.2 Historical Background of Lactic Acid	6
	2.3 Physical and Chemical Properties	8
	2.4 Chemical Synthesis of Lactic Acid	11
	2.5 Microbial Production via Fermentation	12
	2.6 Applications of Lactic Acid	13
	2.6.1 Food industries	15
	2.6.2 Dairy applications	16
	2.6.3 Vegetables	18
	2.6.4 Meat products	19

2.7	Applications of lactic acid bacteria in Agriculture	19
2.8	Industrial and probiotic applications of lactic acid bacteria	20
2.8.1	Non-food application	20
2.8.2	Probiotic applications	21
2.9	Animal feed supplement	22
2.10	Supplements	22
2.11	Lactic Acid in Chemical, Pharmaceutical, and cosmetic Industries	23
2.11.1	Pharmaceutical Industry	23
2.11.2	Solvents	23
2.11.3	Cosmetics	24
2.12	Homofermentative lactate fermentation	24
2.12.1	Heterofermentative lactate fermentation	24
2.12.2	Chemical synthesis Vs Fermentation	24
2.12.3	Lactic acid bacteria	25
2.12.4	Pathways Fermentation of Lactic Acid Bacteria	26
2.13	Microorganisms in lactic acid fermentation	27
2.14	Selection of microorganism	29
2.15	Characterization and taxonomy	29
2.16	Nutrient requirement for lactic acid fermentation	30
2.16.1	Carbon sources	30
2.16.2	Alternative carbon sources	31
2.16.3	Nitrogen sources	32
2.17	Lactic acid fermentation processes	33
2.17.1	Batch fermentation	33
2.17.2	Fed-batch fermentation	34
2.18	Factors affecting on lactic acid fermentation	35
2.18.1	Effect of temperature	35
2.18.2	Effect of pH	36
2.18.3	Effect of incubation period	37
2.18.4	Effect of agitation	37
2.19	Inhibition of lactic acid fermentation	37
2.19.1	Substrate inhibition	38
2.19.2	End-product inhibition	38
2.20	Purification techniques of lactic acid	39
2.21	Chemical and nutritional composition of cassava starch	39

3	METHODOLOGY	43
3.1	Microorganism	43
3.2	Preparation of Working Cell Culture	43
3.3	Inoculum Preparation	44
3.4	Medium Screening	44
3.4.1	Bioreactor cultivation	46
3.4.1.1	Batch Cultivation for <i>Lactobacillus</i> sp.	
3.4.1.2	Fed-batch cultivation for high lactic acid Production	47
3.5	Analytical method	48
3.5.1	Determination Concentrations of carbon sources	48
3.5.2	Determination Concentrations of Nitrogen sources	48
3.5.3	Determination of phosphate source	49
3.5.4	Determination pH of D-lactic acid production Medium	49
3.5.5	Determination Cultivation temperatures	49
3.5.6	Determination Inoculum sizes	50
3.5.7	Determination Total Reduce sugar	50
3.5.8	Optical Density Determination	50
3.5.9	Growth kinetic comparison between optimized and un-optimized medium	51
3.5.10	Determination of D-lactic acid concentration	51
4	RESULTS & DUSCUSSION	53
4.1	Screening and selection of lactic acid bacteria for D-lactic acid Production	53
4.1.1	Screening of lactic acid bacteria for D-lactic acid Production	53
4.1.2	Selection of D-lactic acid-producing isolates and test for cassava starch utilization using agar medium	54
4.1.2.1	Selection of D-lactic acid-producing Isolate	54
4.1.2.2	Investigation on cassava starch utilization using agar medium	54
4.2	Optimization of some conditions for D-lactic acid Production	61

4.2.1	Effect of different cassava starch concentrations on lactic acid production	61
4.2.2	Effect of different nitrogen sources on lactic acid production	64
4.2.2.1	Effect of different peptone concentrations on lactic acid production	64
4.2.2.2	Effect of different yeast extract concentrations on lactic acid production	67
4.2.3	Effect of different Dipotassium phosphate concentrations on D-lactic acid production	69
4.3	Initial pH of D-lactic acid production medium	71
4.4	Cultivation temperatures	73
4.5	Inoculum sizes	75
4.6	Growth Kinetics Comparison	77
4.7	Batch cultivation in 16-L Bioreactor	83
5	CONCLUSION	90
	REFERENCES	92
	Appendices	101 - 105

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Thermodynamic and Physical properties of lactic acid.	9
2.2	Lactic Acid Characteristics	10
2.3	Fermented Food, beverages and its relationship with lactic acid bacteria.	15
2.4	Probiotic of Lactic Acid Bacteria Species.	21
2.5	Production lactic acid by organism type, concentration, yield and productivity.	29
2.6	Common Genera of Lactic acid bacteria and their Shape and Type of Lactic Acid they produce.	30
2.7	Lactic acid fermentation from agricultural resources by batch fermentation.	34
2.8	Composition of cassava materials.	40
2.9	Composition of cassava materials compared with other sources of starch	41
2.10	Chemical composition of cassava roots and leaves.	42
3.1	MRS agar (De Man, Rogosa and Sharpe Medium)	45
3.2	RAM Medium added cassava starch.	45
3.3	Basal Medium added cassava starch.	46
4.1	Lactic acid production by 10 selected isolates using MRS medium containing 2% of glucose and incubation at 35°C for 24 h under anaerobic conditions.	55
4.2	Lactic acid production by 7 selected isolates using RAM medium containing 2% of glucose and incubation at 35°C for 24 h under anaerobic conditions.	57
4.3	Lactic acid production by 7 selected isolates using Basal medium containing 2% of glucose and incubation at 35°C for 24 h under anaerobic conditions.	59

4.4	D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>) after cultivation in RAM broth containing various concentrations of cassava starch at 35°C for 48 h.	62
4.5	Effect of initial pH of the optimized RAM broth on growth and D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>). When cultivated in the medium at 35°C for 48 h.	65
4.6	Effect of cultivation temperatures on growth and D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>) when cultivated in RAM broth containing 10 g/l of cassava starch with the optimum initial pH for 48 h.	67
4.7	Effect of inoculum sizes on growth and D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>). When cultivated in RAM broth containing 10 g/l of cassava starch at the optimum temperature for 48 h.	69
4.8	Effect of initial pH of the optimized RAM broth on growth and D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>). When cultivated in the medium at 35°C for 48 h.	71
4.9	Effect of cultivation temperatures on growth and D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>) when cultivated in RAM broth containing 10 [g/l] of cassava starch When cultivated in the medium at 35°C for 48 h	73
4.10	Effect of inoculum sizes on growth and D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>). When cultivated in RAM broth containing 10 [g/l] of cassava starch when cultivated in the medium at 35°C for 48 h.	75
4.11	Medium composition for un-optimized and optimized media.	77
4.12	Growth kinetics in shake flask cultures using the un-optimized medium.	78
4.13	Growth kinetics in shake flask cultures using the optimized medium.	80
4.14	Cultivation parameters for batch cultivations of (<i>Lactobacillus delbrueckii</i>) in 16-L bioreactor.	83
4.15	D-Lactic acid production, cell dry weight, Total Reducing sugar, and dissolve oxygen changes in batch bioreactor 16-L cultivation with controlled pH.	84

4.16	D-Lactic acid production, cell dry weight, Total Reducing sugar, and dissolve oxygen changes in batch bioreactor 16-L cultivation with un-controlled	87
4.17	Kinetics of D-lactic acid and cell growth production by (<i>Lactobacillus delbrueckii</i>) in shake flask and bioreactor 16-L using different media and different cultivation conditions.	89

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	The Lactic acid isomer forms	9
2.2	Degree of dissociation of lactic acid as function of pH	11
2.3	Comparison between chemical synthesis method (a) and microbial Fermentation method (b) to produce of lactic acid (SSF: represents simultaneous scarification and fermentation)	13
2.4	Diagram of the applications of lactic acid	14
2.5	The pathways of heterofermentative (dotted line) and homofermentative (solid line) lactic acid bacteria	27
2.6	Cassava starch conversion	42
4.1	lactic acid productions by 7 selected isolates using MRS medium incubation at 35°C for 48 h under anaerobic conditions.	56
4.2	lactic acid productions by 7 selected isolates using RAM medium incubation at 35°C for 48 h under anaerobic conditions.	58
4.3	lactic acid production by 7 selected isolates using Basal medium incubation at 35°C for 48 h under anaerobic conditions.	60
4.4	D-lactic acid production of bacterial isolate (<i>Lactobacillus delbruekii</i>) cultivation in RAM broth containing various concentrations of cassava starch at 35°C for 48 h.	63
4.5	D-lactic acid production of bacterial isolate (<i>Lactobacillus delbruekii</i>) after cultivation in RAM broth containing various concentrations of peptone at 35°C for 48 h.	66
4.6	D-lactic acid production of bacterial isolate (<i>Lactobacillus delbruekii</i>) after cultivation in RAM broth containing various concentrations of yeast extract at 35°C for 48 h.	68
4.7	D-lactic acid production of bacterial isolate (<i>Lactobacillus delbruekii</i>) after cultivation in RAM broth containing various concentrations of Dipotassium phosphate at 35°C for 48h.	69

4.8	Effect of initial pH of the optimized RAM broth on D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>). When cultivated in the medium at 35°C for 48 h.	72
4.9	Effect of cultivation temperatures on D-lactic acid production of bacterial isolate (<i>Lactobacillus delbrueckii</i>) when cultivated in RAM broth containing 10 [g/l] of cassava starch When cultivated in the medium at 35°C for 48 h.	74
4.10	Effect of inoculum sizes on D-lactic acid production of bacterial isolaten (<i>Lactobacillus delbrueckii</i>). When cultivated in RAM broth contains10 [g/l] of cassava starch when cultivated in the medium at 35°C for 48 h.	76
4.11	Growth curve kinetic of (<i>Lactobacillus delbrueckii</i>) in un-optimized media in shake flask.	79
4.12	Growth curve kinetic of (<i>Lactobacillus delbrueckii</i>) in un-optimized media in shake flask.	81
4.13	D-Lactic acid production, cell dry weight, Total Reducing sugar, and dissolve oxygen changes in batch bioreactor 16-L cultivation with controlled pH.	85
4.14	D-Lactic acid production, cell dry weight, Total Reducing sugar, and dissolve oxygen changes in batch bioreactor 16-L cultivation with un-controlled pH.	88

LIST OF ABBREVIATIONS

AOAC	-	Association of Official Analytical Chemists
MC	-	MRS containing 0.5% CaCO ₃
MRS	-	De Man Rogosa and Shapes medium
RAM	-	Rogasa Agar Modified medium
<i>et al.</i> ,	-	Ahmed (and others)
PLA	-	Poly lactic acid
LAB	-	lactic acid bacteria
RAM	-	Rogasa Agar Modified medium
rDNA	-	Ribosomal deoxyribonucleic acid
UV	-	Ultraviolet
CDW	-	Cell dry weight
DO	-	Dissolved Oxygen
OD	-	Optical Density
OD ₅₄₀	-	Optical Density at 540 nm
OD ₆₀₀	-	Optical Density at 600 nm
sp.	-	Species

LIST OF SYMBOLS

%	-	Percent
>	-	Greater than
F	-	Feed rate ($\text{g L}^{-1}\text{h}^{-1}$)
KS	-	Substrate utilization constant
S	-	Substrate concentration (g L^{-1})
So	-	Feed substrate concentration ($\text{g substrate L}^{-1}$)
t	-	Time
t0	-	Initial time (h)
V	-	Volume
v/v	-	Volume per volume
vvm	-	Volume per volume per minute
X	-	Biomass concentration (g L^{-1})
X0	-	Original biomass concentration (g L^{-1})
%	-	Percent
>	-	Greater than
F	-	Feed rate ($\text{g L}^{-1}\text{h}^{-1}$)
KS	-	Substrate utilization constant
So	-	Feed substrate concentration ($\text{g substrate L}^{-1}$)
t	-	Time
t0	-	Initial time (h)
V	-	Volume
v/v	-	Volume per volume

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Equipments	101-105

CHAPTER 1

INTRODUCTION

1.1 Research Background

The Cassava (*Manihotesculenta crantz*) is imperishable bush with a massive storage root (El-Sharkawy, 2003). The dry substance of roughly 35-40% is for the most part comprised of starch and the protein substance being low. Both roots and leaves are used in food preparation. All parts of the plant contain cyanogenic glycosides. Upon processing, these are hydrolyzed to release cyanohydrins, which in turn release the toxic HCN (Brimer and Preedy, 2014).

Billions huge amounts of cassava are delivered yearly around the world. Notwithstanding this exclusive half of generation is prepared into consumable one leaving a huge amount of starch cassava that can be harmful to environment because release of HCN (Petrov *et al.*, 2008). The financial obligation to the modern cassava processor depends just on this amount and this let us consider how we can get advantage however much as could reasonably be expected from this colossal measure of cassava.

Starch can be a suitable substrate for producing biochemical materials due to its rich degradability, providing a beneficial approach of cassava (Sommart *et al.*, 2000). Microbial biomass protein and natural acid can move out as profitable final items by the biotechnological treatment of starch. Changing starch into esteemed reflection and fitting activity of starch stream are among the by and large examined viewpoints in cassava research (Adnan and Tan, 2007).

Lactic acid is a standout amongst the most imperative natural acids which could be created from cassava. This natural acid is generally utilized as a part of numerous applications. These days, 80% of lactic acid is utilized as a part of food, and sustenance related commercial enterprises. The business creation of lactic acid is extremely regular and it is conceivable either by chemical synthetic or by fermentation process (Xiaodong *et al.*, 1997).

Acetaldehyde and hydrogen cyanide derived lacto nitrile hydrolysis is a frequently used technique for its synthesis. However bacterial fermentation of simple sugars is mostly used for biotechnological production of the acid, accountable for almost 50% of the total lactic acid capacity (Huang *et al.*, 2003).

Lactic acid, mixture of two isomers dependably comes about while delivering synthetically. Though, a solitary or a blend of two isomers in different extents is yielded, contingent on substrate microorganism and growth conditions utilized as a part of fermentation process. Use of cheap crude materials, for example, starch, sugarcane beet-sugar, molasses and other carb rich materials make natural generation favorable over chemical synthesis (Tsao *et al.*, 1999).

Various vast organizations are worried in the advancement of procedure and creation of lactic acid on account of the colossal potential interest of lactic acid as a segment and feedstock in different commercial industrial, for example, nourishment handling drinks and pharmaceutical. For delivering lactic acid biologically starch and sugars are to a great extent utilized as substrates as a part of the exchange forms (Huang *et al.*, 2005).

The production of lactic acid using microbial resources has been broadly researched and several lactic acid producing bacteria *Streptococcus*, *Tetragenococcus*, *Vagococcus*, *Lactobacillus*, *Lactococcus*, *Leuconostoc*, *Oenococcus*, *Pediococcus*, *Carnobacterium*, *Enterococcus* *Clostridium* and *Weissella* have been reported (Stiles and Holzapfel 1997). Producing lactic acid biologically is mostly completed by the bacterial fermentation of simple sugars. Due to their large production yield and growth rate bacterial strains like *Lactobacillus* and *Lactococcus* have received worldwide profit (Richter and Berthold, 1998).

However, the (1) high costs for substrate pretreatment, hydrolysis (2) particular nutrients supplementation (3) controlling pH through fermentation (4) lactic acid recovery and purification are the major cases with the processes of bacterial fermentation. For example, producing lactic acid by traditional means from starch demands pretreatment by gelatinization and liquefaction performed for 10-15 minutes between 100-130 °C temperature after that enzyme scarification to glucose and following its processing to lactic acid by fermentation. The two-step process includes of successive enzymatic hydrolysis and microbial fermentation makes unappealing from economic perspectives. Generally the composition of cassava is 800 g kg⁻¹ water, 20 g kg⁻¹ protein, 1 g kg⁻¹ fat, 170 g kg⁻¹ carbohydrate and 9g kg⁻¹ other components like inorganic minerals ,vitamins and metals (Alves *et al.*, 2007).

The modulation of all the desired nutrients for various fermentation is the advantage of such a feedstock as compare to other agro industrial wastes (sugar beet, or extract of wheat bran, peel of citrus fruits). All others require collection with carbon or nitrogen sources and other trivial nutrients. Therefore least preprocessing and supplementation are the advantages of using cassava as a feedstock. The production cost of lactic acid would be decreased by producing using starch.

1.2 Problem Statement

This Research includes two problem statements, economical and environmental Several years of intensive research has been done for the production of lactic acid based on the importance of this acid in food, and chemical industries. However, most of previous studies were focused on using standard medium for lactic acid bacteria cultivations such as MRS broth contain expensive carbon and nitrogen sources.

Billions huge amounts of cassava are delivered yearly around the world. Notwithstanding this exclusive half of generation is prepared into consumable one leaving a huge amount of starch cassava that can be harmful to environment because release of HCN. The financial obligation to the modern cassava processor depends just on this amount and this let us consider how we can get advantage however much as could reasonably be expected from this colossal measure of cassava.

Therefore, there was a need to develop cheap culture medium to improve the process economy of lactic acid production. Thus, in the present work the potential use of cassava as cheap and available carbon source for the development of more efficient and cheap process for lactic acid production will be investigated.

This work included investigation of lactic acid production beginning, from screening of possibility of Lactic acid bacteria (LAB) for lactic acid production. At least 7 standard strains of lactic acid bacteria obtained from Wellness Industries Culture Collection (WICC) at the Institute of Bio product Development (IBD), University Technology Malaysia, of will be screened and selected for D-lactic acid production. Several optimum production parameters (particularly nutrient sources using cassava starch as a carbon source, nitrogen sources, phosphate source, optimum temperature, pH, cassava concentration and inoculums size) for D-lactic acid production will be determined.

1.3 Research objectives

The main objective of this study the production of D-lactic acid through culture media optimization and cultivation strategy in semi-industrial scale 16-L bioreactor by using cassava starch as a main carbon source.

1.4 Scope and limitation of the study

The scope of the research will be as follows:

1.4.1 Screening and selection of the possibility strain of (LAB) for D-lactic acid production using starch of cassava as feedstock.

1.4.2 Optimization of medium composition by using one factor at time (OFAT) using cassava based medium.

1.4.3 Growth kinetics in shake flask cultures using the un-optimized medium.

1.4.4 Growth kinetics in shake flask cultures using the optimized medium

1.4.5 Study the effect of bioprocessing conditions on D-lactic acid production in semi industrial scale 16-L bioreactor.

1.4.6 Batch cultivation of *Lactobacillus* sp. in semi-industrial scale 16-L bioreactor.

REFERENCES

- Malek, R.A., Hamdan, S., El Enshasy, H., Othman, N.Z., Zainol, N.A., Sarmidi, M.R. and Aziz, R. (2010). Production of *Lactobacillus salivarius*, a new probiotic strain isolated from human breast milk, in semi-industrial scale and studies on its functional characterization. *Current Research, Technology and Education Topics in Applied Microbiology and Microbial Biotechnology*, 2, 1196-1204.
- Adnan, A.F.M. and Tan, I.K. (2007). Isolation of lactic acid bacteria from Malaysian foods and assessment of the isolates for industrial potential. *Bioresource Technology*, 98(7), 1380-1385.
- Albenzio, M., Corbo, M.R., Rehman, S.U., Fox, P.F., De Angelis, M., Corsetti, A., Sevi, A. and Gobetti, M. (2001). Microbiological and biochemical characteristics of Canestrato Pugliese cheese made from raw milk, pasteurized milk or by heating the curd in hot whey. *International journal of food microbiology*, 67(1), 35-48.
- Alves, V.D., Mali, S., Beleia, A. and Grossmann, M.V.E. (2007). Effect of glycerol and amylose enrichment on cassava starch film properties. *Journal of Food Engineering*, 78(3), 941-946.
- Alves, A.A.C. (2002). Cassava botany and physiology. *Cassava: biology, production and utilization*, 67-89.
- Aristidou, A. and Penttila, M. (2000). Metabolic engineering applications to renewable resource utilization. *Current Opinion in Biotechnology*, 11(2), 187-198.
- Armstrong, M. (1996). *A handbook of personnel management practice*. Kogan Page.
- Axelsson, L. (2004). Lactic acid bacteria: classification and physiology. *Food scines and technology new york*, 139, 1-66.
- Badel, S., Bernardi, T. and Michaud, P. (2011). New perspectives for *Lactobacilli exopolysaccharides*. *Biotechnology advances*, 29(1), 54-66.
- Bomrungnok, W., Sonomoto, K., Pinitglang, S. and Wongwicharn, A. (2012). Single Step Lactic Acid Production from Cassava Starch by *Laactobacillus plantarum* SW14 in Conventional Continuous and Continuous with High Cell Density. *APCBEE Procedia*, 2, 97-103.

- Balcazar, J.L., De Blas, I., Ruiz-Zarzuola, I., Cunningham, D., Vendrell, D. and Muzquiz, J.L. (2006). The role of probiotics in aquaculture. *Veterinary microbiology*, 114(3), 173-186.
- Bashan, Y. and De-Bashan, L.E. (2010). Chapter two-how the plant growth-promoting bacterium *Azospirillum* promotes plant growth—a critical assessment. *Advances in agronomy*, 108, 77-136.
- Berg, G. (2009). Plant–microbe interactions promoting plant growth and health: perspectives for controlled use of microorganisms in agriculture. *Applied microbiology and biotechnology*, 84(1), 11-18.
- Bloemberg, G.V. and Lugtenberg, B.J. (2001). Molecular basis of plant growth promotion and biocontrol by *rhizobacteria*. *Current opinion in plant biology*, 4(4), 343-350.
- Preedy, V.R. ed. (2014). *Processing and impact on antioxidants in beverages*. Elsevier.
- Caplice, E. and Fitzgerald, G.F. (1999). Food fermentations: role of microorganisms in food production and preservation. *International journal of food microbiology*, 50(1) .131-149.
- Chiarini, L., Mara, L. and Tabacchioni, S. (1992). Influence of growth supplements on lactic acid production in whey ultrafiltrate by *Lactobacillus helveticus*. *Applied microbiology and biotechnology*, 36(4), 461-464.
- Chen, Y., Krol, J., Huang, W., Mirro, R. and Gossain, V. (2008). Anaerobic yeast fermentation for the production of ethanol in a versatile lab fermentor. *Nature Methods*, 5(12).
- Gonçalves, L.M.D., Ramos, A., Almeida, J.S., Xavier, A.M.R.B. and Carrondo, M.J.T. (1999). Elucidation of the mechanism of lactic acid growth inhibition and production in batch cultures of *Lactobacillus rhamnosus*. *Applied Microbiology and Biotechnology*, 48(3), 346-350.
- Gatje, G. and Gottschalk, G. (1991). Limitation of growth and lactic acid production in batch and continuous cultures of *Lactobacillus helveticus*. *Applied microbiology and biotechnology*, 34(4), 446-449.
- Casida, J.E. (1964). Esterase inhibitors as pesticides. *Science*, 146(3647), 1011-1017.
- Charles, A.L., Sriroth, K. and Huang, T.C. (2005). Proximate composition, mineral contents, hydrogen cyanide and phytic acid of 5 cassava genotypes. *Food Chemistry*, 92(4), 615-620.
- Cordell, J.L., Falini, B.R., Erber, W.N., Ghosh, A.K., Abdulaziz, Z., MacDonald, S., Pulford, K.A., Stein, H.A. and Mason, D.Y. (1984). Immunoenzymatic labeling of monoclonal antibodies using immune complexes of alkaline phosphatase and

- monoclonal anti-alkaline phosphatase (APAAP complexes). *Journal of Histochemistry & Cytochemistry*, 32(2), 219-229.
- Corma, A., Iborra, S. and Velly, A. (2007). Chemical routes for the transformation of biomass into chemicals. *Chemical reviews*, 107(6), 2411-2502.
- Corroler, D., Mangin, I., Desmasures, N. and Gueguen, M. (1998). An ecological study of *lactococci* isolated from raw milk in the Camembert cheese registered designation of origin area. *Applied and Environmental Microbiology*, 64(12), 4729-4735.
- Datta, R. and Henry, M. (2006). Lactic acid: recent advances in products, processes and technologies-a review. *Journal of Chemical Technology and Biotechnology*, 81(7), 1119-1129.
- de Vrese, M. and Schrezenmeir, J.(2008). Probiotics, prebiotics, and synbiotics. *Food biotechnology*, (1-66). Springer Berlin Heidelberg.
- Dien, B.S., Nichols, N.N. and Bothast, R.J. (2002). Fermentation of sugar mixtures using *Escherichia coli* catabolite repression mutants engineered for production of L-lactic acid. *Journal of industrial Microbiology and Biotechnology*, 29(5), 221-227.
- De Vuyst, L. and Vancanneyt, M. (2007). Biodiversity and identification of sourdough lactic acid bacteria. *Food Microbiology*, 24(2), 120-127.
- Dubois, M., Gilles, K.A., Hamilton, J.K., Rebers, P.A.T. and Smith, F. (1956). Colorimetric method for determination of sugars and related substances. *Analytical chemistry*, 28(3), 350-356.
- Ding, S. and Tan, T. (2006). L-lactic acid production by *Lactobacillus casei* fermentation using different fed-batch feeding strategies. *Process Biochemistry*, 41(6), 1451-1454.
- Elmarzugi, N., Enshasy, H.E., Abd Malek, R., Othman, Z., Sarmidi, M.R. and Abdel Aziz, R. (2010). Optimization of cell mass production of the probiotic strain *Lactococcus lactis* in batch and fed-batch culture in pilot scale levels. *Current Research, Technology and Education Topics in Applied Microbiology and Microbial Technology*, 2, 873-879.
- El-Enshasy, H.A. and El-Shereef, A.A. (2008). *Saccharomyces boulardii* Adapted to Dryness Stress: Optimization of High Cell Density Cultivation of Yeast. *Deutsche Lebensmittel-Rundschau*, 104(9), 423-428.
- Elsayed, E.A., Othman, N.Z., Malek, R., Tang, T. and Enshasy, H.A. (2014). Improvement of cell mass production of *Lactobacillus delbrueckii* sp. *bulgaricus* WICC-B-02: A newly isolated probiotic strain from mother's milk.
- Vieira, E.A., Fialho, J.D.F., Faleiro, F.G., Bellon, G., Fonseca, K.G.D., Carvalho, L.J.C.B., Silva, M.S., Paula-Moraes, S.V.D., Oliveira, C.M.D. and Denke, M.L.

- (2011). Characterization of sweet cassava accessions based on molecular, quantitative and qualitative data. *Crop Breeding and Applied Biotechnology*, 11(3), 232-240.
- Fontana, C., Fadda, S., Cocconcelli, P.S. and Vignolo, G. (2011). Lactic acid bacteria in meat fermentations. *Lactic Acid Bacteria: Microbiological and Functional Aspects*, 247. (22-32)
- Gal, J. (2008). The discovery of biological enantioselectivity: Louis Pasteur and the fermentation of tartaric acid, 1857-a review and analysis 150 yr later. *Chirality*, 20(1), 5-19.
- Guilloux-Benatier, M. and Chassagne, D. (2003). Comparison of components released by fermented or active dried yeasts after aging on lees in a model wine. *Journal of Agricultural and Food Chemistry*, 51(3), 746-751.
- Garriga, M., Aymerich, M.T., Costa, S., Monfort, J.M. and Hugas, M. (2002). Bactericidal synergism through bacteriocins and high pressure in a meat model system during storage. *Food Microbiology*, 19(5), 509-518.
- Ginzinger, W., Jaros, D., Lavanchy, P. and Rohm, H. (1999). Raw milk flora affects composition and quality of Bergkase. Physical and sensory properties and conclusions. *Le Lait*, 79(4), 411-421.
- Glick, B.R. (1995). The enhancement of plant growth by free-living bacteria. *Canadian Journal of Microbiology*, 41(2), 109-117.
- Gottschalk, C.W. (1979). Renal nerves and sodium excretion. *Annual review of physiology*, 41(1), 229-240.
- Gueimonde, M. and Salminen, S. (2006). New methods for selecting and evaluating probiotics. *Digestive and Liver Disease*, 38, S242-S247.
- Hashemipour, H., Khaksar, V. and Kermanshahi, H. (2011). Application of probiotic on egg production and egg quality of chukar partridge. *African Journal of Biotechnology*, 10(82), pp.19244-19248.
- Huang, L.P., Dong, T., Chen, J.W. and Li, N. (2000). Biotechnological production of lactic acid integrated with fishmeal wastewater treatment by *Rhizopus oryzae*. *Bioprocess and biosystems engineering*, 30(2), 135-140.
- Huang, C.F., Lin, T.H., Guo, G.L. and Hwang, W.S. (2009). Enhanced ethanol production by fermentation of rice straw hydrolysate without detoxification using a newly adapted strain of *Pichia stipitis*. *Bioresource Technology*, 100(17), 3914-3920.
- Hall, M.B. and Herejk, C. (2001). Differences in yields of microbial crude protein from in vitro fermentation of carbohydrates. *Journal of Dairy Science*, 84(11), 2486-2493.

- Hujanen, M. and Linko, Y.Y.(1996). Effect of temperature and various nitrogen sources on L (+)-lactic acid production by *Lactobacillus casei*. *Applied microbiology and biotechnology*, 45(3), 307-313.
- Hugas, M. and Monfort, J.M. (1997). Bacterial starter cultures for meat fermentation. *Food chemistry*, 59(4), 547-554.
- Ibrahim, F., Ruvio, S., Granlund, L., Salminen, S., Viitanen, M. and Ouwehand, A.C. (2010). Probiotics and immunosenescence: cheese as a carrier. *Fesm Immunology & Medical Microbiology*, 59(1), 53-59.
- Isolauri, E. (2001). Probiotics in human disease. *The American journal of clinical nutrition*, 73(6), 1142S-1146S.
- Jem, K.J., van der Pol, J.F. and de Vos, S. (2010). Microbial lactic acid, its polymer poly lactic acid, and their industrial applications. In *Plastics from bacteria*, (323-346). Springer Berlin Heidelberg.
- John, R.P., Nampoothiri, K.M. and Pandey, A. (2007). Fermentative production of lactic acid from biomass: an overview on process developments and future perspectives. *Applied Microbiology and Biotechnology*, 74(3), 524-534.
- Jones, A.M. and Ingledew, W.M. (1994). Fermentation of very high gravity wheat mash prepared using fresh yeast autolysate. *Bioresource Technology*, 50(2), 97-101.
- Khalaf, S.A. (2001). Lactic acid Production by interspecific hybrids of *rhizopus* strain from potato processing peel waste. *Egyptian Journal of Microbiology (Egypt)*.
- Kharas, G.B., Sanchez-Riera, F. and Severson, D.K. (1994). Polymers of lactic acid. *Plastics from microbes: microbial synthesis of polymers and polymer precursors*, (93-137).
- Law, B.A. and Kolstad, J. (1983). Proteolytic systems in lactic acid bacteria. *Antonie van Leeuwenhoek*, 49(3), 225-245.
- Lu, Z., He, F., Shi, Y., Lu, M. and Yu, L. (2010). Fermentative production of L (+)-lactic acid using hydrolyzed acorn starch, persimmon juice and wheat bran hydrolysate as nutrients. *Bioresource technology*, 101(10), 3642-3648.
- Leroy, F., Verluyten, J. and De Vuyst, L. (2006). Functional meat starter cultures for improved sausage fermentation. *International journal of food microbiology*, 106(3), pp.270-285.
- Linko, Y.Y. and Javanainen, P. (1996). Simultaneous liquefaction, saccharification, and lactic acid fermentation on barley starch. *Enzyme and Microbial Technology*, 19(2), pp.118-123.

- Love, P.E.D., Holt, G.D., Shen, L.Y., Li, H. and Irani, Z. (2002). Using systems dynamics to better understand change and rework in construction project management systems. *International Journal of Project Management*, 20(6), 425-436.
- Malek, R.A., bin Hamdan, S., El Enshasy, H.A. and Zalina, N., High cell mass production and characterization of *Lactobacillus salivarius*, a new probiotic strain isolated from human breast milk.
- Macklaim, J.M., Gloor, G.B., Anukam, K.C., Cribby, S. and Reid, G. (2011). At the crossroads of vaginal health and disease, the genome sequence of *Lactobacillus iners AB-1*. *Proceedings of the National Academy of Sciences*, 108, 4688-4695.
- Moo-Young, M., Chisti, Y. and Vlach, D. (1993). Fermentation of cellulosic materials to mycoprotein foods. *Biotechnology advances*, 11(3), 469-479.
- Musa, D., Schulz, R., Harris, R., Silverman, M. and Thomas, S.B. (2009). Trust in the health care system and the use of preventive health services by older black and white adults. *American journal of public health*, 99(7), 1293-1299.
- Mercier, P., Yerushalmi, L., Rouleau, D. and Dochain, D. (1992). Kinetics of lactic acid fermentation on glucose and corn by *Lactobacillus amylophilus*. *Journal of Chemical Technology and Biotechnology*, 55(2), 111-121.
- Musa, H.H. and Seri, H. (2016). The potential benefits of probiotics in animal production and health.
- Narayanan, R. and El-Sayed, M.A. (2004). Shape-dependent catalytic activity of platinum nanoparticles in colloidal solution. *Nano letters*, 4(7), 1343-1348.
- Naveena, B.J., Altaf, M., Bhadrappa, K. and Reddy, G. (2004). Production of L (+) lactic acid by *Lactobacillus amylophilus* GV6 in semi-solid state fermentation using wheat bran. *Food Technology and Biotechnology*, 42(3), 147-152.
- Nduele, M., Ludwig, A. and Van Ooteghem, M. (1993). Physicochemical properties of cassavata starch stored at different conditions of temperature and humidity. *STP pharma sciences*, 3(5), 362-368.
- Perepelkin, K.E. (2002). Polylactide fibres: Fabrication, properties, uses, prospects. review. *Fibre Chemistry*, 34(2), 85-100.
- Pandey, A., Soccol, C.R., Nigam, P., Soccol, V.T., Vandenberghe, L.P. and Mohan, R. (2000). Biotechnological potential of agro-industrial residues. II: cassava bagasse. *Bioresource Technology*, 74(1), pp.81-87.
- Panesar, P.S., Kennedy, J.F., Knill, C.J. and Kosseva, M. (2010). Production of L (+) lactic acid using *Lactobacillus casei* from whey. *Brazilian archives of Biology and Technology*, 53(1), 219-226.

- Petrov, K., Urshev, Z. and Petrova, P. (2008). L (+)-Lactic acid production from starch by a novel *amylolytic Lactococcus lactis subsp. lactis B84*. *Food Microbiology*, 25(4), 550-557.
- Ramzi, A. (2015). Recent applications of poly lactic acid in pharmaceutical and medical industries. *Journal of Chemical and Pharmaceutical Research*, 2015, 7(12):51-63
- Richardson, P.H., Jeffcoat, R. and Shi, Y.C. (2000). High-amylose starches: From biosynthesis to their use as food ingredients. *MRS Bulletin*, 25(12), 20-24.
- Randolph, T.W., Marison, I.W., Martens, D.E. and Von Stockar, U. (1990). Calorimetric control of fed-batch fermentations. *Biotechnology and bioengineering*, 36(7), 678-684.
- Richter, K. and Berthold, C. (1998). Biotechnological conversion of sugar and starchy crops into lactic acid. *Journal of agricultural engineering research*, 71(2), 181-191.
- Ross, R.P., Morgan, S. and Hill, C. (2002). Preservation and fermentation: past, present and future. *International journal of food microbiology*, 79(1), 3-16.
- Roukas, T. and Kotzekidou, P. (1991). Production of lactic acid from deproteinized whey by coimmobilized *Lactobacillus casei* and *Lactococcus lactis* cells. *Enzyme and microbial technology*, 13(1), 33-38.
- Rodriguez-Sandoval, E., Fernandez-uintero, A., Cuvelier, G., Relkin, P. and Bello-Perez, L.A. (2008). Starch retrogradation in cassava flour from cooked parenchyma. *Starch-Starke*, 60(3-4), 174-180.
- Roychoudhury, P.K., Srivastava, A. and Sahai, V. (1995). Extractive bioconversion of lactic acid. In *Downstream Processing Biosurfactants Carotenoids*, (61-87). Springer Berlin Heidelberg.
- Saleem, M., Arshad, M., Hussain, S. and Bhatti, A.S. (2007). Perspective of plant growth promoting *rhizobacteria* (PGPR) containing ACC deaminase in stress agriculture. *Journal of industrial microbiology & biotechnology*, 34(10), 635-648.
- Salovaara, H. and Ganzle, M. (2011). Lactic acid bacteria in cereal-based products. *Lactic Acid Bacteria: Microbiological and Functional Aspects*, 2, (227).
- Sanchez, C., Belleville, P., Popall, M. and Nicole, L. (2011). Applications of advanced hybrid organic–inorganic nanomaterials: from laboratory to market. *Chemical Society Reviews*, 40(2), 696-753.
- Shah, N.P. (2000). Probiotic bacteria: selective enumeration and survival in dairy foods. *Journal of dairy science*, 83(4), 894-907.

- Schuler, D. (2004). Molecular analysis of a subcellular compartment: the magnetosome membrane in *Magnetospirillum gryphiswaldense*. *Archives of microbiology*, 181(1), pp.1-7.
- Sturr, M.G. and Marquis, R.E. (1992). Comparative acid tolerances and inhibitor sensitivities of isolated F-ATPases of oral lactic acid bacteria. *Applied and environmental microbiology*, 58(7), 2287-2291.
- Singh, S.K., Ahmed, S.U. and Pandey, A. (2006). Metabolic engineering approaches for lactic acid production. *Process Biochemistry*, 41(5), 991-1000.
- Sommart, K., Parker, D.S., Rowlinson, P. and Wanapat, M. (2000). Fermentation characteristics and microbial protein synthesis in an in vitro system using cassava, rice straw and dried ruzi grass as substrates. *Asian Australian journal of animal sciences*, 13(8), 1084-1093.
- Severson, D.K. (1998). Lactic acid fermentations. *Nutritional Requirements of Commercially Important Microorganisms*. EsteeKay Associates, Milwaukee, 258-297.
- Spaepen, S., Vanderleyden, J. and Remans, R. (2007). Indole-3-acetic acid in microbial and microorganism-plant signaling. *Fesm microbiology reviews*, 31(4), 425-448.
- Stiles, M.E. and Holzapel, W.H.(1997). Lactic acid bacteria of foods and their current taxonomy. *International journal of food microbiology*, 36(1), pp.1-29.
- Sun, S.Y., Xu, Y. and Wang, D. (2009). Novel minor lipase from *Rhizopus chinensis* during solid-state fermentation: Biochemical characterization and its esterification potential for ester synthesis. *Bioresource technology*, 100(9), 2607-2612.
- Tanguler, H. and Erten, H. (2008). Utilisation of spent brewer's yeast for yeast extract production by autolysis: The effect of temperature. *Food and Bioproducts Processing*, 86(4), 317-321.
- Tango, M.S.A. and Ghaly, A.E. (1999). Effect of temperature on lactic acid production from cheese whey using *Lactobacillus helveticus* under batch conditions. *Biomass and bioenergy*, 16(1), 61-78.
- Tonkuari.(2004). Cassava and the future of starch. *Electronic journal of biotechnology*, 7, (5-8).
- Tong, W.Y., Fu, X.Y., Lee, S.M., Yu, J., Liu, J.W., Wei, D.Z. and Koo, Y.M. (2004). Purification of L (+)-lactic acid from fermentation broth with paper sludge as a cellulosic feedstock using weak anion exchanger Amberlite IRA-92. *Biochemical Engineering Journal*, 18(2), 89-96.
- Tsao, G.T., Cao, N.J., Du, J.C.S.G. and Gong, C.S. (1999). Production of multifunctional organic acids from renewable resources. In *Recent progress in bioconversion of lignocellulosics*, (243-280). Springer Berlin Heidelberg.

- Vasiljevic, T. and Shah, N.P. (2008). Probiotics-from Metchnikoff to bioactives. *International Dairy Journal*, 18(7), 714-728.
- Varun, B., Jones, R.T., Kandasamy, S.K.J., Vijaykumar, P. and Anant, A. (2008). Optimization of production of subtilisin in solid substrate fermentation using response surface methodology. *African Journal of Biotechnology*, 7(13).
- Vancanneyt, M., Neysens, P., De Wachter, M., Engelbeen, K., Snauwaert, C., Cleenwerck, I., Van der Meulen, R., Hoste, B., Tsakalidou, E., De Vuyst, L. and Swings, J. (2005). *Lactobacillus acidifarinae* sp. nov. and *Lactobacillus zymae* sp. nov., from wheat sourdoughs. *International journal of systematic and evolutionary microbiology*, 55(2), 615-620.
- Wouters, J.T., Ayad, E.H., Hugenholtz, J. and Smit, G. (2002). Microbes from raw milk for fermented dairy products. *International Dairy Journal*, 12(2), 91-109.
- Xiaodong, W., Xuan, G. and Rakshit, S.K. (1997). Direct fermentative production of lactic acid on cassava and other starch substrates. *Biotechnology letters*, 19(9), 841-843.
- Yin, P., Nishina, N., Kosakai, Y., Yahiro, K., Pakr, Y. and Okabe, M. (1997). Enhanced production of L (+)-lactic acid from corn starch in a culture of *Rhizopus oryzae* using an air-lift bioreactor. *Journal of fermentation and bioengineering*, 84(3), 249-253.
- Yin, P., Yahiro, K., Ishigaki, T., Park, Y. and Okabe, M. (1998). L (+)-Lactic acid production by repeated batch culture of *Rhizopus oryzae* in air-lift bioreactor. *Journal of Fermentation and Bioengineering*, 85(1), 96-100.
- Oh, H., Wee, Y.J., Yun, J.S., Han, S.H., Jung, S. and Ryu, H.W. (2005). Lactic acid production from agricultural resources as cheap raw materials. *Bioresource technology*, 96(13), 1492-1498.
- Nordmark, T.S. (1993). A filter paper assay for low cellulase activities and the cultivation of *Trichoderma reesei* on acid whey and sweet whey permeate.
- Zheng, Y., Chen, F. and Wang, M. (2013). Use of Bacillus-Based Biocontrol Agents for Promoting Plant Growth and Health. In *Bacteria in Agrobiolgy: Disease Management*, (243-258). Springer Berlin Heidelberg.
- Zhang, D.X. and Cheryan, M. (1991). Direct fermentation of starch to lactic acid by *Lactobacillus amylovorus*. *Biotechnology letters*, 13(10), 733-738.