

DECOLOURISATION AND DEGRADATION OF ACID ORANGE 7 USING AN
ACCLIMATISED BAC-ZS MIXED BACTERIAL CULTURE

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.... *For my parents; Bay Nguang Hong and Chai Nyuk Mui*
.... *For my lovely husband Khoo Kiat Siong*
.... *For my sisters; Wendy, Elaine and Caroline*

You brought colours into my life

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“The roots of all goodness lie in the soil of appreciation for goodness” – Dalai Lama

ABSTRACT

Formation of dark coloured auto-oxidation compounds had resulted in reduced efficiency of the sequential anaerobic-aerobic treatment system to decolourise sulphonated azo dyes. In view of this, a monosulphonated azo dye, Acid Orange 7 (AO7) was selected as a model dye to study the decolourisation of AO7 and its auto-oxidation compounds by a mixed bacterial culture, BAC-ZS. It consisted of three bacteria namely *Brevibacillus panacihumi* strain ZB1, *Lysinibacillus fusiformis* strain ZB2 and *Enterococcus faecalis* strain ZL. The decolourisation and degradation process was performed using the sequential facultative anaerobic-aerobic system. Optimisation of the co-substrate showed that the combination of glucose (5 g/L) and yeast extract (3 g/L) was the best co-substrate for decolourisation; 98% of AO7 colour was removed within 2 h of facultative anaerobic phase. When the decolourised solution was further treated under the aerobic phase, auto-oxidation reaction resulted in heavy browning effect after 24 h of agitation. The browning effect had drastically decreased the decolourisation to 72%. However, continuous agitation up to 48 h successfully decolourised the auto-oxidation compounds as indicated by the increase in decolourisation up to 90%. Consequently, the decolourisation was accompanied by 73% decrease in Chemical Oxygen Demands (COD) and an increase of 94% of bacteria concentration (absorbance at 600 nm). It was also found that the initial pH 6.6 of AO7 solution dropped to pH 4.5 during facultative anaerobic decolourisation and increased to pH 7.7 at the end of aerobic treatment. The degradation of AO7 dye was determined and confirmed using the UV-Vis spectrophotometry and FTIR analysis. In addition, the formation of auto-oxidation compounds, 1,2-naphtholquinone and 1,4-benzoquinone were detected and monitored using HPLC analysis. Further phytotoxicity tests using *Cucumis sativus* confirmed detoxification of the final treated solution by BAC-ZS. Quantification of BAC-ZS using real-time polymerase chain reaction (RT-PCR) showed *E. faecalis* strain ZL was the dominant bacteria in the acclimatised BAC-ZS and throughout the AO7 treatment process. The annotation of the draft genome of each bacteria revealed presence of genes coding for the azoreductases, dioxygenases and monooxygenases which played important roles in degradation and mineralisation of AO7 dye. In conclusion, the acclimatised BAC-ZS mixed bacterial culture has good potential to be used in the biological treatment of textile effluent.

ABSTRAK

Pembentukan sebatian gelap auto-oksidasi mengakibatkan penurunan efikasi sistem rawatan berjujuk anaerobik-aerobik untuk menyahwarna pewarna sulfur azo. Oleh itu, pewarna monosulfur azo, Acid Orange 7 (AO7) digunakan sebagai pewarna model dalam kajian untuk menghasilkan campuran kultur bakteria yang berkeupayaan menyahwarnakan AO7 dan juga sebatian auto-oksidasinya. Tiga jenis bakteria; *Brevibacillus panacihumi* strain ZB1, *Lysinibacillus fusiformis* strain ZB2 dan *Enterococcus faecalis* strain ZL digunakan untuk membentuk kultur bacteria campuran, BAC-ZS. Proses penyahwarna dan degradasi AO7 adalah berpandukan sistem berjjukan fakultatif anaerobik-aerobik. Substrak optimasi menunjukkan gabungan glukosa (5 g/L) dengan ekstrak yis (3 g/L) adalah yang terbaik dan 98% warna AO7 dinyahwarnakan dalam tempoh 2 jam fakultatif anaerobik. Lanjutan rawatan ke fasa aerobik akibatkan oksidasi larutan dalam tempoh 24 jam. Pemerangan larutan akibatkan penurunan drastik peratusan penyahwarna (72%). Akan tetapi, pengoncangan berterusan selepas 48 jam berjaya menyahwarnakan sebatian auto-oksidasi berikutan peningkatan peratusan penyahwarna (90%). Proses penyahwarna diikuti penurunan Keperluan Oksigen Kimia (COD) sebanyak 73% dan peningkatan konsentrasi bakteria sebanyak 94% berdasarkan ketumpatan optik (Abs_{600nm}). Bacaan pH larutan AO7 (pH 6.6) menurun ke pH 4.5 semasa penyahwarna fakultatif anaerobik dan meningkat ke pH 7.7 pada peringkat akhir rawatan aerobik. Degradasi AO7 ditentukan berdasarkan spektroskopi UV-Vis dan analisis FTIR. Malahan, pembentukan sebatian auto-oksidasi; 1,2-naphtholquinone dan 1,4-benzoquinone dikesan menggunakan analisis HPLC. Ujian fitotoksikitas menggunakan *Cucumis sativus* membuktikan larutan AO7 telah dinyahtoksikan selepas rawatan dengan BAC-ZS. Quantifikasi BAC-ZS menggunakan real-time polymerase chain reaction (RT-PCR) menunjukkan *E. faecalis* strain ZL adalah bakteria dominan dalam BAC-ZS dan kekal sebagai spesies dominan sepanjang proses degradasi AO7. Anotasi draf genom bakteria menunjukkan kehadiran gen-gen pengkodan enzim azoreductase, dioxygenase dan monooxygenase yang penting untuk mineralisasi AO7. Kesimpulannya, kultur campuran BAC-ZS mempunyai potensi yang baik untuk digunakan dalam rawatan biologi kumbahan tekstil.

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LIST OF SYMBOLS

% v/v	-	percentage volume per volume
% w/v	-	percentage weight per volume
°C	-	celcius
μl	-	microlitre
μM	-	micromolar
∞	-	infinity
Ct	-	threshold cycle
Da	-	dalton
g	-	gram
g/L	-	gram per litre
Kb	-	kilo base
kPa	-	kilopascal
L	-	litre
mg/L	-	milligram per litre
min	-	minutes
ml	-	millilitre
mM	-	milimolar
mm	-	milimeter
ng	-	nanogram
ng/L	-	nanogram per litre
nm	-	nanometer
pg	-	picogram
pM	-	picamolar
ppm	-	part per million
rpm	-	rotation per minute

sec	-	seconds
T _m	-	melting temperature
V	-	volt

LIST OF ABBREVIATIONS

2-ABS	-	2-aminobenzenesulfonic acid
4-ABS	-	4-aminobenzenesulfonic acid
AO7	-	Acid Orange 7
BLASTn	-	Basic Local Alignment Search Tool - nucleotide
BLASTp	-	Basic Local Alignment Search Tool - protein
bp	-	basepair
COD	-	Chemical Oxygen Demand
dH ₂ O	-	distilled water
DNS	-	3,5-Dinitrosalicylic acid
EC	-	Enzyme code
EDTA	-	Ethylenediaminetetraacetic acid
FADH	-	Flavin adenine dinucleotide
FISH	-	Fluorescence <i>in situ</i> hybridisation
FMN	-	Flavin mononucleotide
FTIR	-	Fourier transform infrared spectroscopy
GO	-	gene ontology
GST	-	gluthione S-transferase
HPLC	-	High-performance liquid chromatography
KEGG	-	Kyoto Encyclopedia of Genes and Genomes
NADH	-	Nicotinamide adenine dinucleotide
NADH-DCIP	-	Nicotinamide adenine dinucleotide-2,6 dichlorophenol indophenol
NADPH	-	Nicotinamide adenine dinucleotide phosphate
NaOH	-	Sodium hydroxide
NCBI	-	National Center for Biotechnology Information

OD	-	Optical density
OFAT	-	One-Factor-A-Time
PCR-DGGE	-	Polymerase chain reaction-denaturing gradient gel electrophoresis
PCR-SSCP	-	Polymerase chain reaction-single stranded conformation polymorphism
QRT-PCR	-	Quantitative real time polymerase chain reaction
Rnase	-	Ribonuclease
rRNA	-	Ribosomal ribonucleic acid
SD	-	Standard deviation
SMILES	-	Simplified Molecular-Input Line-Entry System
SOD	-	Superoxide dismutase
TAE	-	Tris-acetate-EDTA
UM-BDD	-	University of Minnesota-biocatalysis and biodegradation database
UV	-	Ultraviolet

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CHAPTER 1

INTRODUCTION

1.1 Background of the problem

Synthetic dyes such as azo dye are extensively used in the food, pharmaceutical, textile, cosmetics and paper industries because of their colour varieties and fastness in production as compared to natural dyes (Chang and Lin, 2001; Carneiro *et al.*, 2007). Among the total synthetic dyes produced annually, around 60 – 70% consist of azo dyes (Mc Mullan, 2001). Azo dyes that are characterised by double bonds (-N=N-) are the largest and most versatile class of dyes and are the most common chromophore in azo dyes (Stolz, 2001). Most of the azo dyes released into environment originate from the textile and the dyestuff manufacturing industries (Carliell *et al.*, 1995). Pearce *et al.* (2003) explained during dye processing approach, about 2 % of dyes are directly discharged into aqueous effluent. Additionally, an estimation of 2 – 50% of various applied dyes can be lost in the effluent during textile colouring processes (Easton, 1995).

The azo dyes that dissolve in water can cause water stream to become highly coloured even at low concentrations. Dye concentration as low as 0.005 ppm is visible in clear river water (Banat *et al.*, 1996) whereas according to O'Neill *et al.*

(1999), textile processing wastewater usually contains between 10 – 200 mg/L dye concentration. Once the intense coloured wastewater flow into the river as seen in Figure 1.1, the colour will influence the absorbance of light entering the water thus, greatly reduces photosynthesis of aquatic flora (Slokar *et al.*, 1998).



Figure 1.1: A newspaper article from News Strait Times shows one of the polluted rivers in Malaysia due to textile wastewater

Kodam *et al.* (2006) mentioned most of the azo dyes are either inert or non-toxic, but become toxic, mutagenic and carcinogenic upon their biotransformation. The azoreductases in intestinal bacteria, liver cells and skin surfaces micro flora would reduce azo bonds in azo dyes to colourless aromatic amines (Xu *et al.*, 2007). Several researchers have done studies related to risk assessment of aromatic amines and they have proven carcinogenicity of aromatic amines to the human body (Hildenbrand *et al.*, 1999).

Considering both environmental pollution and serious health-risk factors caused by azo dyes, finding alternative ways to treat synthetic dyes are mandatory particularly for small scale textile industries. This is because in reality, the textile effluents were directly channelled into the main streams of water resources without proper treatment system (Moosvi *et al.*, 2005). This is a major environmental issue as most of the textile dyeing and processing industries are located in developing countries whereby rivers are the main source of drinking water and daily activities. In India for instance, an average mill discharges is about 15 million litres of contaminated effluent per day that causes chronic and acute toxicity (Dave *et al.*, 2009).

While in Malaysia, there are more than 200 textiles factories with majority consisting of small scale textile industries. Textile industry is the major source of wastewater and accounts for 22 % of the total volume of industrial wastewater produced in Malaysia (Idris *et al.*, 2007). Most of these textile industries use conventional treatment methods to treat the textile effluents. The conventional methods are effective in removing the fiber and to reduce COD reading of the wastewater however, it is not effective to treat the colour of the wastewater as the presence of sulpho and azo groups in the azo dyes, make the dyes xenobiotic and recalcitrant to oxidative biodegradation (Killa *et al.*, 1983). As a result, the treated textiles wastewater remains highly coloured as seen in Figure 1.2.

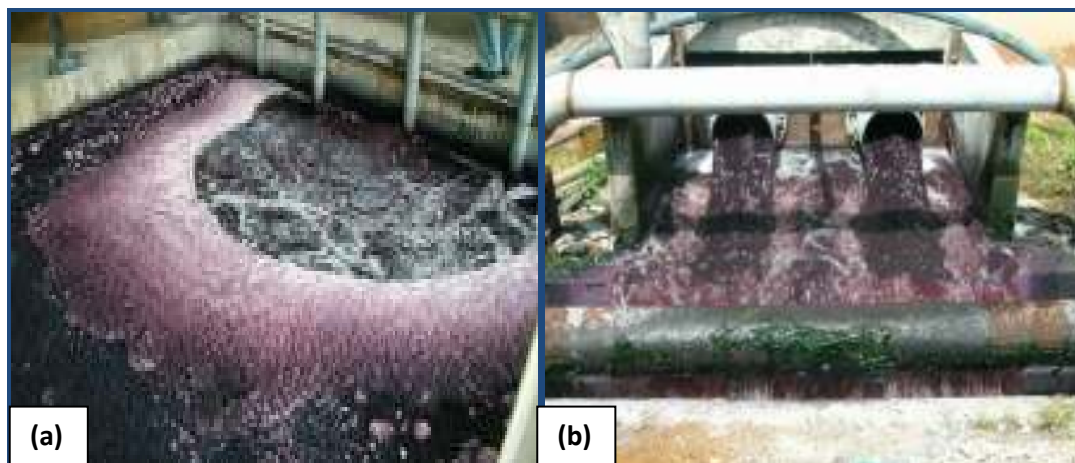


Figure 1.2: Pictures taken in one of the local textiles factories in Malaysia. (a) Textiles wastewater in treatment tank. (b) Treated textiles wastewater discarded to drain

1.2 Problem statement

Several physical and chemical techniques were tested to be effective in dye removal such as physical adsorption using activated carbon (Choi *et al.*, 2008), ozonation (Gharbani *et al.*, 2008), electrolytic treatment using a three dimensional electrode reactor (Xu *et al.*, 2008) as well as Fenton-like reaction (Rahmani *et al.*, 2010). The use of physico-chemical methods however, have their shortcomings due to excess amount of chemical usage, excessive sludge generation with obvious disposal problems, costly plant requirements or operating expenses and lack of effective colour reduction, particularly for sulfonated azo dyes (Banat *et al.*, 1996).

The bioremediation using effective microbial to decolourise and degrade dyes used in textile industry is an environmental-friendly and cost-effective method compared to the physical and chemical decomposition processes. The successful application of bioremediation processes are very dependent on the microorganisms

exploited in the systems. In recent years, attention has been focused on fundamental work in revealing the existence of wide range of microorganisms capable of decolourising a wide range of dyes (Moosvi *et al.*, 2005, 2007; Dafale *et al.*, 2008; Telke *et al.*, 2010). Practically, it has been reported that mixed cultures are useful in decolourisation of synthetic dyes, as some microbial consortia can achieve the biodegradation tasks that no individual pure strain can undertake successfully (Nigam *et al.*, 1996).

Among the biological methods that have been introduced, sequential anaerobic-aerobic treatment system was considered the most reliable method to treat textile wastewater that consists of various dyes (Khan and Banarjee, 2010). Successful application of mixed bacteria cultures in decolourising textile dyes in the sequential anaerobic-aerobic treatment system had been well reported (O'Neill *et al.*, 200; Oh *et al.*, 2004; Ong *et al.*, 2005). Although being a reliable system, this method still has a major drawback; it is not effective in degrading sulphonated azo dyes. The sulphonated azo dyes could be decolourised under anaerobic phase however, once in contact with air under aerobic phase, the colourless aromatic compounds are fast auto-oxidised to form dark and recalcitrant by-products (Supaka *et al.*, 2004; Mendez-Paz *et al.* 2005, 2005a). The formation of dark solution is non-reversible (Coughlin *et al.*, 2002; Wang *et al.*, 2012).

Therefore, developing a mixed bacterial culture with ability to decolourise sulphonated azo dyes as well as to overcome formation of dark solution during auto-oxidation process in a sequential anaerobic-aerobic treatment system is essential. For this reason, three objectives were outlined for depth investigation of the mixed bacteria culture as an alternative approach for decolourisation and degradation of sulphonated textile dyes.

1.3 Objectives of study

- (1) To isolate and develop a mixed bacterial culture for decolourisation of azo dye
- (2) To evaluate the efficiency of acclimatised mixed bacterial culture for dye degradation using sequential facultative anaerobic-aerobic treatment system
- (3) To monitor population dynamics of acclimatised mixed bacterial culture using real-time polymerase chain reaction (RT-PCR).

1.4 Scope of the study

The present study was focused on the development of a mixed bacterial culture with potential use for the decolourisation of textile dyes in a sequential anaerobic-aerobic system. Acid Orange 7(AO7) has wide application in textiles, cosmetics and leather tanning industries. Being a monosulphonated dye, degradation of AO7 in a sequential anaerobic-aerobic treatment system would form dark browning solution at the end of treatment therefore; this dye was used as the model azo dye in this study. The mixed bacterial culture was formed using two bacteria screened and isolated from textile wastewater together with *E. faecalis* strain ZL that earlier has been proven to decolourise the AO7 dye. The mixed bacterial culture was acclimatised in AO7 decolourised solution prior used for treatment and named as BAC-ZS. Optimisation of co-substrate was investigated to increase the decolourisation efficiency. The decolourisation and degradation of AO7 was performed in sequential facultative anaerobic-aerobic system. In general, the degradation performances were observed based on the colour removal, the COD reduction, pH changes and indirect measurement of bacteria concentration during the treatment process. The efficiency of the BAC-ZS to mineralise AO7 was tested by investigating the dye solution before and after treatment using HPLC and FTIR analyses as well as the phytotoxicity test. Further, species-specific primers were

designed and used in determining the population dynamic of BAC-ZS during the dye treatment process via RT-PCR.

solution. The use of microorganisms to degrade polymerised compounds has not been reported before and is worthy for further investigation to elucidate degradation pathway associated with the BAC-ZS mixed bacterial culture.

- ii. The potential application of BAC-ZS mixed culture can be extended to include real textile waste water using the sequential anaerobic-aerobic bioreactor.

- iii. With the advance of bioinformatics, the information derived from the annotated draft genomes of each bacterium can be further exploited to treat other types of pollutants.

REFERENCES

- Abraham, K.J. and John, G.H. (2007). Development of a classification scheme using a secondary and tertiary amino acid analysis of azoreductase gene. *Journal of Medical and Biological Sciences* 1(2): 1 – 5.
- Adedayo, O., Javadpour, S., Taylor, C., Anderson, W.A. and Moo-Young, M. (2004). Decolourization and detoxification of methyl red by aerobic bacteria from a wastewater treatment plant. *World Journal of Microbiology and Biotechnology* 20: 545 – 550.
- Ali, H. (2010). Biodegradation of synthetic dyes – a review. *Water Air Soil and Pollution* 213: 251 – 273.
- Altschul, S.F., Madden, T.L., Schäffer, A.A., Zhang, J., Zhang, Z., Miller, W., Lipman, D.J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic acids research* 25(17): 3389 – 3402.
- Ahmed, I., Yokota, A., Yamazoe, A. and Fujiwara, T. (2007) Proposal of *lysini bacillus boronitolerans* gen. nov. sp. nov., and transfer of *Bacillus fusiformis* to *lysini bacillus fusiformis* comb. nov. and *Bacillus spaericus* to *Lysinibacillus sphaericus* comb. nov. *International Journal of Systematic and Evolutionary Microbiology* 57: 1117 – 1125.
- An, S.Y., Min, S.K., Cha, I.H., Choi, Y.L., Cho, Y.S., Kim C.H. and Lee, Y.C (2002). Decolorization of triphenylmethane and azo dyes by *Citrobacter* sp. *Biotechnology Letters* 24: 1037 – 1040.
- Anastasi, A., Spina, F., Prigione, V, Tigini, T., Giansanti, P. and Varese, G.C. (2010). Scale-up of a bioprocess for textile wastewater treatment using *Bjerkandera adusta*. *Bioresource Technology* 101: 3067 – 3075.

- Ang, S.K. (2014). Biodegradation of untreated oil palm trunk for cellulases and xylanase production by locally isolated fungi using solid state fermentation. Doctor Philosophy, Universiti Teknologi Malaysia, Skudai.
- Anjaneya, O., Souche, S.Y., Santoshkumar, M. and Karegoudar, T.B. (2011). Decolorization of sulfonated azo dye Metanil Yellow by newly isolated bacterial strains: *Bacillus* sp. strain AK1 and *Lysinibacillus* sp. strain AK2. *Journal of Hazardous Materials* 190: 351 – 358.
- Asad, S., Amoozegar, M.A., Pourbabae, A.A., Sarbolouki, M.N. and Dastgheib, S.M.M. (2007). Decolorization of textile azo dyes by newly isolated halophilic and halotolerant bacteria. *Bioresource Technology* 98: 2082 – 2088.
- Ayed, L., Cheriaa, J., Laadhari, N., Cheref, A. and Bakhrouf, A. (2009). Biodegradation of crystal violet by an isolated *Bacillus* sp. *Annals of Microbiology* 59(2): 267 – 272.
- Baek, K.H., Yoon, B.D., Cho, D.H., Kim, B.H., Oh, H.M. and Kim, H.S. (2009). Monitoring bacterial population dynamics using real-time PCR during the bioremediation of crude-oil-contaminated soil. *Journal of Microbiology and Biotechnology* 19(4): 339 – 345.
- Bafana, A., Chakrabarti, T. and Devi, S.S. (2008). Azoreductase and dye detoxification activities of *Bacillus velezensis* strain AB. *Applied Microbiology and Biotechnology* 77: 1139 – 1144.
- Banat, I.M., Nigam, P., Singh, D. and Marchant, R. (1996). Microbial decolorization of textile-dye-containing effluents: A review. *Bioresource Technology*: 217 – 227.
- Bardi, L. and Marzona, M. (2010). Factors affecting the complete mineralisation of azo dyes. *Biodegradation of Azo Dyes*. H. Atacag Erkurt (ed.) *The Handbook of Environmental Chemistry* 9:195– 210, London: Springer – Verlag.
- Barsing, P., Tiwari, A., Joshi, T. and Garg, S.(2011). Application of a novel bacterial consortium for mineralization of sulphonated aromatic amines. *Bioresource Technology* 102: 765 – 771.
- Baud-Grasset., F., Baud-Grasset, S. and Safferman, S.I. (1993). Evaluation of the bioremediation of a contaminated soil with phytotoxicity tests. *Chemosphere* 26(7): 1365 – 1374.
- Bay, H.H., Lim, C.K., Kee, T.C., Ware, I., Chan, G.F., Shahir, S. and Ibrahim, Z. (2014). Decolourisation of Acid Orange 7 recalcitrant auto-oxidation

- coloured by-products using an acclimatised mixed bacterial culture. *Environmental Sciences and Pollution Research* 21(5): 3891 – 3906.
- Ben Mansour, H., Corroler, D., Barillier, D., Ghedira, K., Chekir, L. and Mosrati, R. (2009). Influence of the chemical structure on the biodegradability of acids yellow 17, violet 7 and orange 52 by *Pseudomonas putida*. *Annals of Microbiology* 59(1): 9 – 15.
- Bonakdarpour, B., Vyrides, I. and Stuckey, D.C. (2011). Comparison of the performance of one stage and two stage sequential Anaerobic-aerobic biological processes for the treatment of reactive-azo-dye-containing synthetic wastewaters. *International Biodeterioration and Biodegradation* 65: 591 – 599.
- Bonser, G.M., Bradshaw, L., Clayson, D.B. and Jull, J.W. (1956). A further study on the carcinogenic properties of ortho-hydroxyamines and related compounds by bladder implantation in the mouse. *British Journal of Cancer* 10: 539 – 553.
- Brinkman, N.E., Haugland, R.A., Wymer, L.J., Byappanahalli, M., Whitman, R.L. and Vesper, S.J. (2003). Evaluation of a rapid, quantitative real-time PCR method for enumeration of pathogenic *Candida* cells in water. *Applied and Environmental Microbiology* 69(3): 1775 – 1782.
- Carliell, C.M., Barclay, S.J., Naidoo, N., Buckley, C.A., Mulholland, D.A. and Senior, E. (1996). Treatment of exhausted reactive dye bath effluent using anaerobic digestion: laboratory and full-scale trials. *Water SA* 22(3): 225 – 233.
- Carliell, C.M., Barclay, S.J., Naidoo, N., Buckley, C.A., Mulholland, D.A. and Senior, E. (1995). Microbial decolourisation of a reactive azo dye under anaerobic conditions. *Water SA* 21(1): 61 – 69.
- Carneiro, P.A., Nogueira, R.F.P. and Zanoni, M.V.B. (2007). Homogeneous photodegradation of C.I. Reactive Blue 4 using a Photo-Fenton process under artificial and solar irradiation. *Dyes and Pigments* 74: 127 – 132.
- Carvalho, M.C., Pereira, C., Gonc-alves, I.C., Pinheiro, H.M., Santos, A.R., Lopes, A. and Ferra, M.I. (2008). Assessment of the biodegradability of a monosulfonated azo dye and aromatic amines. *International Biodeterioration and Biodegradation* 62: 96 – 103.

- Cases, I. and de Lorenzo, V. (2001) The black cat/white cat principle of signal integration in bacterial promoters. *EMBO Journal*. 20: 1 – 11.
- Cervantes, F.J. and Dos Santos, A.B. (2011). Reduction of azo dyes by anaerobic bacteria: microbiological and biochemical aspects. *Reviews in Environmental Science and Biotechnology*. 10: 125 – 137.
- Chan, G.F., Rashid, N.A., Koay, L.L., Chang, S.Y. and Tan, W.L. (2011). Identification and Optimization of Novel NAR-1 Bacterial Consortium for the Biodegradation of Orange II. *Insight Biotechnology* 1(1): 7 – 16.
- Chan, G.F., Abdul Rashid, N.A., Chua, L.S., Ab.Illah, N., Nasiri, R. and Mohamad Ikubar, M.R. (2012). Communal microaerophilic-aerobic biodegradation of Amaranth by novel NAR-2 bacterial consortium. *Bioresource Technology* 105: 48 – 59.
- Chan, G.F., Gan, H.M., Rashid, N.A. (2012a). Genome sequence of *Enterococcus* sp. strain C1, an azo dye decolorizer. *Journal of Bacteriology* 194(20): 5716 – 5717.
- Chandrakant, S.K. and Shwetha, S.R. (2012). Role of microbial enzymes in the bioremediation of pollutants: A review. *Enzyme Research* vol. 2011, Article ID 805187, 11 pages, 2011. doi:10.4061/2011/805187
- Chang, J.S., Chou, C. and Chen, S.Y. (2001). Decolorization of azo dyes with immobilized *Pseudomonas luteola*. *Process Biochemistry* 36: 757 – 763.
- Chang, J.S., Chen, B.Y. and Lin, Y.S. (2004). Stimulation of bacterial decolorization of an azo dye by extracellular metabolites from *Escherichia coli* strain NO3. *Bioresource Technology* 91: 243 – 248.
- Chang, S.J. and Lin, Y.C. (2001). Decolorization kinetics of recombinant *Escherichia coli* strain harboring azo dye decolorization determinants for *Rhodococcus* sp. *Biotechnology Letters* 23: 631 – 636.
- Chen, B.Y., Chen, S.Y., Lin, M.Y. and Chang, J.S. (2006). Exploring bioaugmentation strategies for azo-dye decolorization using a mixed consortium of *Pseudomonas luteola* and *Escherichia coli*. *Process Biochemistry* 41: 1574 – 1581.
- Chen, B.Y. and Chang, J.S. (2007). Assessment upon species evolution of mix consortia for azo dye decolorization. *Journal of the Chinese Institute of Chemical Engineers* 38: 259 – 266.

- Chen, H.Z., Wang, R.F. and Cerniglia C.E. (2004). Molecular cloning, overexpression, purification, and characterization of an aerobic FMN-dependent azoreductase from *Enterococcus faecalis*. *Protein Expression and Purification* 34: 302 – 310.
- Chen, H.Z., Hopper, S.L. and Cerniglia, C.E. (2005). Biochemical and molecular characterization of an azoreductase from *Staphylococcus aureus*, a tetrameric NADPH-dependent flavoprotein. *Microbiology* 151: 1433 – 1441.
- Chen, K.C., Wu, J.Y., Liou, D.J. and Hwang, J.S.C. (2003). Decolorization of the textile dyes by newly isolated bacterial strains. *Journal of Biotechnology* 101(1): 57 – 68.
- Chengalroyen, M. D. and Dabbs, E. R. (2013). The microbial degradation of azo dyes: minireview. *World Journal of Microbiology and Biotechnology* 29: 389 – 399.
- Chikere, C.B., Surridge, K.J., Cloete, E.T., Okpokwasili, G.C. (2011). Phylogenetic diversity of dominant bacterial communities during bioremediation of crude oil-polluted soil. *An interdisciplinary Journal of Applied Science* 6(2): 61 – 76.
- Choi, H.D., Shin, M.C., Kim, D.H., Jeon, C.S. and Baek K. (2008). Removal characteristics of reactive black 5 using surfactant-modified activated carbon. *Desalination* 223: 290 – 298.
- Chung, K.T. and Cerniglia, C.E. (1992). Mutagenicity of azo dyes: Structure-activity relationships. *Mutation Research* 277: 201 – 220.
- Conesa, A., Götz, S., García-Gómez, J.M, Terol1, J., Talón, M and Robles, M. (2005). Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics* 21(8): 3674 – 3676.
- Coughlin, M.F., Kinkle, B.K. and Bishop, P.L. (1997). Characterization of aerobic azo dye-degrading bacteria and their activity in biofilms. *Water Science and Technology* 36: 341 – 346.
- Coughlin, M.F., Kinkle, B.K. and Bishop, P.L. (2002). Degradation of acid orange 7 in an aerobic biofilm. *Chemosphere* 46(1): 11 – 19.
- Cui, D., Li, G., Zhao, D., Gu, X., Wang, C. and Zhao, M. (2012). Microbial community structures in mixed bacterial consortia for azo dye treatment under aerobic and anaerobic conditions. *Journal of Hazardous Materials* 221– 222: 185 – 192.

- Dafale, N., Rao, N.N., Meshram, S.U. and Wate, S.R. (2008). Decolorization of azo dyes and simulated dye bath wastewater using acclimatized microbial consortium – Biostimulation and halo tolerance. *Bioresource Technology* 99: 2552 – 2558.
- Daims, H., Taylor, M.W. and Wagner, M. (2006). Wastewater treatment: a model system for microbial ecology. *Trends Biotechnology* 24(11): 483 – 489.
- Dharmaraj, S. (2006). Real Time-PCR: the basics. Retrieved on August 17, 2013 from <http://www.ambion.com/techlib/basics/rtpcr/index.html>.
- Dave, S.R. and Dave, R.H. (2009). Isolation and characterization of *Bacillus thuringiensis* for Acid red 119 dye decolourisation. *Bioresource Technology* 100: 249–253.
- Davies, L.C., Pedro, I.S., Novais, J.M. and Martins-Dias, S. (2006). Aerobic degradation of Acid Orange 7 in a vertical-flow constructed wetland. *Water Research* 40(10): 2055 – 2063.
- Dawkar, V.V., Jadhav, U.U., Jadhav, S.U., Govindwar, S.P. (2008). Biodegradation of disperse textile dye Brown 3REL by newly isolated *Bacillus* sp. VUS. *Journal of Applied Microbiology* 105(1): 14 – 24.
- Dawkar, V.V., Jadhav, U.U., Ghodake, G.S. and Govindwar, S.P. (2009). Effect of inducers on the decolorization and biodegradation of textile azo dye Navy blue 2GL by *Bacillus* sp. VUS. *Biodegradation* 20: 777 – 787.
- Dedeyan, B., Klonowska, A. and Tagger, S. (2000). Biochemical and molecular characterization of a laccase from *Marasmius quercophilus*. *Applied and Environmental Microbiology* 66(3): 925 – 929.
- Desai, C., Jain, K., Patel, B. and Madamwar, D. (2009). Efficacy of bacterial consortium-AIE2 for contemporaneous Cr(VI) and azo dye bioremediation in batch and continuous bioreactor systems, monitoring steady-state bacterial dynamics using qPCR assays. *Biodegradation* 20: 813 – 826.
- Diwaniyan, S. Kharb, D., Raghukumar, D. and Kuhad, R.C. (2010). Decolorization of synthetic dyes and textile effluents by Basidiomycetous fungi. *Water, Air and Soil Pollution* 210(1 – 4): 409 – 419.
- Dos Santos, A.B., Cervantes, F.J., Yaya-Beas, R.E., Van Lier, J.B. (2003). Effect of redox mediator, AQDS, on the decolourisation of a reactive azo dye containing triazine group in a thermophilic anaerobic EGSB reactor. *Enzyme and Microbial Technology* 33: 942 – 51.

- Dos Santos, A.B., Bisschops, I.A.E., Cervantes, F.J. and van Lier, J.B. (2004). Effect of different redox mediators during thermophilic azo dye reduction by anaerobic granular sludge and comparative study between mesophilic (30°C) and thermophilic (55°C) treatments for decolourisation of textile wastewaters. *Chemosphere* 55: 1149 – 1157.
- Dos Santos, A.B., Cervantes, F.J. and Lier van, J.B.(2007). Review paper on current technologies for decolourisation of textile wastewaters: Perspectives for anaerobic biotechnology. *Bioresource Technology* 98: 2369 – 2385.
- Du, L.D., Wang, S., Li, G., Wang, B., Jia, X.M., Zhao, Y.H. and Chen, Y.L. (2011). Biodegradation of malachite green by *Pseudomonas* sp. strain DY1 under aerobic condition: characteristics, degradation products, enzyme analysis and phytotoxicity. *Ecotoxicology*. 20: 438 – 446.
- Duncan, D.B. (1955). Multiple range and multiple F test. *Biometrics* 11(1): 1 – 42.
- Eisheid, A.C. (2011). SYTO dyes and EvaGreen outperform SYBR Green in real-time PCR. *BMC Research Notes* 4: 263.
- Elisangela, F., Andrea, Z., Fabio, D.G., Cristiano, R., Regina,D.L. and Artur C.P. (2009). Biodegradation of textile azo dyes by a facultative *Staphylococcus arlettae* strain VN-11 using a sequential microaerophilic/aerobic process. *International Biodeterioration and Biodegradation* 63: 280 – 288.
- Field, J.A., Stams, A.J.M., Kato, M. and Schraa, G. (1995). Enhanced biodegradation of aromatic pollutants in cocultures of anaerobic and aerobic bacterial consortia. *Antonie van Leeuwenhoek* 67: 47 – 77.
- Forgacs, E., Cserhádi, T. and Oros, G. (2005). Removal of synthetic dyes from wastewaters: a review. *Environment International* 30: 953 – 971.
- Franciscon, E., Zille, A., Fantinatti-Garboggini, F., Silva, I.S., Cavaco-Paulo, A. and Durrant, L.R. (2009). Microaerophilic–aerobic sequential decolourization/biodegradation of textile azo dyes by a facultative *Klebsiella* sp. strain VN-31. *Process Biochemistry* 44: 446 – 452.
- Franciscon, E., Piubeli, F., Fantinatti-Garboggini, F., Ragagnin de Menezes, C., Silva, I.S., Cavaco-Paulo, A., Grossman, M.J. and Durrant, L.R. (2010). Polymerization study of the aromatic amines generated by the biodegradation of azo dyes using the laccase enzyme. *Enzyme and Microbial Technology* 46: 360 – 365.

- Franciscon, E., Grossman, M.J., Paschoal, J.A.R., Reyes, F.G.R and Durrant, L.R. (2012). Decolorization and biodegradation of reactive sulfonated azo dyes by a newly isolated *Brevibacterium* sp. strain VN-15. *SpringerPlus* (1): 1 –10.
- Fu, C.J., Carter, J.N., Li, Y., Porter, J.H. and Kerley, M.S. (2006). Comparison of agar plate and real-time PCR on enumeration of *Lactobacillus*, *Clostridium perfringens* and total anaerobic bacteria in dog faeces. *Letters in Applied Microbiology* 42(5): 490 – 494.
- Gan, H.M., Shahir, S., Ibrahim, Z. and Yahya, A. (2011). Biodegradation of 4-aminobenzenesulfonate by *Ralstonia* sp. PBA and *Hydrogenophaga* sp. PBC isolated from textile wastewater treatment plant. *Chemosphere* 82: 507 – 513.
- Gan, H.M., Chew, T.H., Tay, Y.L., Lye, S.F., Yahya, A. (2012). Genome sequence of *Hydrogenosphaga* sp. strain PBC, a 4-aminobenzenesulfonate-degrading bacterium. *Journal of Bacteriology* 194(17): 4759 – 4760.
- Gasparic, M.B., Tengs. T., La Paz, J.L., Holst-Jensen, A., Pla, M., Esteve, T, Zel, J. and Gruden, K. (2010). Comparison of nine different real-time PCR chemistries for qualitative and quantitative applications in GMO detection. *Analytical and Bioanalytical Chemistry* 396(6): 2023 – 2029.
- Gharbani, P., Tabatabaie, S. M. and Mehrizad, A. (2008). Removal of Congo Red from textile wastewater by ozonation. *International Journal of Environmental Science and Technology* 5(4): 495 – 500.
- Ghodake, G., Jadhav, U., Tamboli, D., Kagalkar, A. and Govindwar, S. (2011). Decolorization of textile dyes and degradation of mono-azo dye Amaranth by *Acinetobacter calcoaceticus* NCIM 2890. *Indian Journal of Microbiology* 51(4): 501 – 508.
- Gianfreda, L., Xu, F., Bollag, J.M. (1999). Laccases: A useful group of oxidoreductive enzymes. *Bioremediation Journal* 3:1 – 25.
- Golob, V., Vinder, A., Simoni, M (2005). Efficiency of the coagulation/flocculation method for the treatment of dyebath effluents. *Dye Pigment* (67): 93 – 97.

- Gudnason, H., Dufva, M., Bang, D.D. and Wolff, A. (2007). Comparison of multiple DNA dyes for real-time PCR: effects of dye concentration and sequence composition on DNA amplification and melting temperature. *Nucleic Acid Research* 35(9): 1 – 8.
- Guo, J., Kang, L., Wang, X. and Yang, J. (2010). Decolorization and Degradation of Azo Dyes by Redox Mediator System with Bacteria. *Biodegradation of Azo Dyes*. H. Atacag Erkurt (ed.) *The Handbook of Environmental Chemistry* 9: 85 – 100, London: Springer – Verlag.
- Hall (2007). phylogenetic trees made easy: *A how-to manual*, 3rd edition. Sinauer Associates, Sunderland, Massachusetts.
- Hall, S.J., Hugenholtz, P., Siyambalapitiya, N., Keller, J. and Blackall, L.L. (2002). The development and use of real-time PCR for the quantification of nitrifiers in activated sludge. *Water Science and Technology* 46(1 – 2): 267 – 272.
- Handayani, W., Meitiniarti, V.I. and Timotius, K. H. (2007). Decolorization of Acid Red 27 and Reactive Red 2 by *Enterococcus faecalis* under a batch system. *World Journal of Microbiology and Biotechnology* 23: 1239 – 1244.
- Harms, G., Layton, A.C., Dionisi, H.M., Gregory, I.R., Garrett, V.M., Hawkins, S.A., Robinson, K.G. and Sayler, G.S. (2003). Real – time PCR quantification of nitrifying bacteria in a municipal wastewater treatment plant. *Environmental Science and Technology* 37: 343 – 351.
- Haug, W., Schmidt, A., Nörtemann, B., Hempel, D.C., Stolz, A. and Knackmuss, H.J. (1991). Mineralization of the sulfonated azo dye Mordant Yellow 3 by a 6-aminonaphthalene-2-sulfonate-degrading bacterial consortium. *Applied and Environmental Microbiology* 57(11): 3144 – 3149.
- He, J., Ritalahti, K.M., Aiello, M.R. and Löffler, F.E. (2003). Complete detoxification of vinyl chloride by an anaerobic enrichment culture and identification of the reductively dechlorinating population as a *Dehalococcoides* species. *Applied and Environmental Microbiology* 69: 996 – 1003.
- Higuchi, R., Dollinger, G., Walsh, P.S. and Griffith R., (1992). Simultaneous application and detection of specific DNA sequences. *Biotechnology* 10: 413 – 417.

- Hildenbrand, S., Schmahl, F.W., Wodarz, R., Kimmel, R. and Dartsch, P.C. (1999). Azo dyes and carcinogenic aromatic amines in cell cultures. *International Archives of Occupational and Environmental Health* 72(3): 52 – 56.
- Horitsu, H., Takada, M., Idaka, E., Tomoyeda, M and Ogawa, T. (1977). Aminoazobenzene by *Bacillus subtilis*. *European Journal of Applied Microbiology* (4): 217 – 224.
- Hughes, B.S., Cullum, A.J. and Bennett, A.F. (2007). Evolutionary adaptation to environmental pH in experimental lineages of *Escherichia coli*. *Evolution* 61(7): 1725 – 1734.
- Ibrahim, Z., Amin, M.F.M., Yahya, A., Aris, A. and Muda, K. (2010). Characteristics of developed granules containing selected decolourising bacteria for the degradation of textile wastewater. *Water Science and Technology* 61(5): 1279 – 1288.
- Idris, A., Hashim, R. Abdul Rahman, R., Ahmad, W.A., Ibrahim, Z., Abdul Razak, P.R., Mohd Zin, H. and Bakar, I. (2007). Application of bioremediation process for textile wastewater treatment using pilot plant. *International Journal of Engineering and Technology* 4(2): 228 – 243.
- Idaka, E. and Ogawa, Y. (1978). Degradation of azo compounds by *Aeromonas hydrophila* var. 2413. *Journal of the Society of Dyers and Colourists*, 94: 91 – 94.
- Işik, M. and Sponza, D.T. (2003). Effect of oxygen on decolorization of azo dyes by *Escherichia coli* and *Pseudomonas* sp. and fate of aromatic amines. *Process Biochemistry* 38: 1183 – 1192.
- Işik, M. and Sponza, D.T. (2007). Fate and toxicity of azo dye metabolites under batch long-term anaerobic incubations. *Enzyme and Microbial Technology* 40: 934 – 939.
- Jadhav, J.P., Kalyani, D.C., Telke, A.A., Phugare, S.S. and Govindwar, S.P. (2010). Evaluation of the efficacy of a bacterial consortium for the removal of color, reduction of heavy metals, and toxicity from textile dye effluent. *Bioresource Technology* 101: 165 – 173.
- Janda, J.M. and Abbott, S.L. (2007). 16S rRNA gene sequencing for bacterial identification in the diagnostic laboratory: Pluses, Perils and Pitfalls. *Journal of Clinical Microbiology* 45: 2761 – 2764.

- Joe, M.H., Lim, S.Y., Kim, D.H. and Lee, I.S. (2008). Decolorization of reactive dyes by *Clostridium bifermentans* SL186 isolated from contaminated soil. *World Journal of Microbiology and Biotechnology* 24: 2221 – 2226.
- Jonstrup, M., Kumar, N., Murto, M. and Mattiasson, B. (2011). Sequential anaerobic-aerobic treatment of azo dyes: Decolourisation and amine degradability. *Desalination* 280(1): 229 – 246.
- Joshi, T., Iyengar, L., Singh, K. and Garg, S. (2008). Isolation, identification and application of novel bacterial consortium TJ-1 for the decolourization of structurally different azo dyes. *Bioresource Technology* 99: 7115 – 7121.
- Jung, R., Steinle, D. and Anliker, R. (1992). A compilation of genotoxicity and carcinogenicity data on aromatic aminosulphonic acids. *Food and Chemical Toxicology* 30: 635 – 660.
- Kalyani, D.C., Patil, P.S., Jadhav, J.P. and Govindwar, S.P. (2008). Biodegradation of reactive textile dye Red BLI by an isolated bacterium *Pseudomonas* sp. SUK1. *Bioresource Technology* 99: 4635–4641.
- Kalyani, D.C., Telke, A.A., Dhanve, R.S. and Jadhav, J.P. (2009). Ecofriendly biodegradation and detoxification of Reactive Red 2 textile dye by newly isolated *Pseudomonas* sp. SUK1. *Journal of Hazardous Materials* 2(3): 735 – 742.
- Kalyani, D.C., Phugare, S.S., Shedbalkar, U.U. and Jadhav, J.P. (2011). Purification and characterization of a bacterial peroxidase from the isolated strain *Pseudomonas* sp. SUK1 and its application for textile dye decolorization. *Annals of Microbiology* 61(3): 483 – 491.
- Kandelbauer, A. and Guebitz, G.M. (2005). Bioremediation for the decolorization of textile dyes – A review. In *Environmental Chemistry* (pp 269 – 288). Springer Berlin Heidelberg.
- Kapdan, I.K. and Oztekin, R. (2003). Decolorization of textile dyestuff Reactive Orange 16 in fed-batch reactor under anaerobic condition. *Enzyme and Microbial Technology* 33: 231 – 235.
- Keck, A., Klein, J., Kudlich, M., Stolz, A., Knackmuss, H.J., and Mattes, R. (1997). Reduction of azo dyes by redox mediators originating in the naphthalenesulfonic acid degradation pathway of *Sphingomonas* sp. strain BN6. *Applied Environmental Microbiology* 63: 3684 – 3690.

- Khalid, A., Arshad, M. and Crowley, D.E. (2008). Decolorization of azo dyes by *Shewanella* sp. under saline conditions. *Applied Microbiology and Biotechnology* 79: 1053 – 1059.
- Khalid, A., Arshad, M. and Crowley, D.E. (2008a). Accelerated decolorization of structurally different azo dyes by newly isolated bacterial strains. *Environmental Biotechnology* 78: 361 – 369.
- Khalid, A., Arshad, M. and Crowley, D. (2009). Biodegradation potential of pure and mixed bacterial cultures for removal of 4-nitroaniline from textile dye wastewater. *Water Research* 43(3): 1110 – 1116.
- Khan, R. and Banarjee, U.C. (2010). Decolorization of azo dyes by immobilized bacteria. *Biodegradation of Azo Dyes* 9: 73 – 84.
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S. and Chimni, S.S. (2005). Decolorization of various azo dyes by bacterial consortium. *Dyes and Pigments* 67: 55 – 61.
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S. and Chimni, S.S. (2006). Biodegradation of azo dye C.I. Acid Red 88 by an anoxic-aerobic sequential bioreactor. *Dyes and Pigments* 70: 1 – 7.
- Khelifi, E., Bouallagui, H., Touhami, Y., Godon, J.J. and Hamdi, M. (2009). Bacterial monitoring by molecular tools of a continuous stirred tank reactor treating textile wastewater. *Bioresource Technology* 100: 629 – 633.
- Kim, M.K., Sathiyaraj, S., Pulla, R.K. and Yang, D.C. *Brevibacillus panacihumi* sp. nov., a β -glucosidase producing bacterium. *International Journal of Systematic and Evolutionary Microbiology* 59: 1227 – 1231.
- Knapp, J.S. and Newby, P.S. (1995). The microbiological decolorization of an industrial effluent containing a diazo-linked chromophore. *Water Research* 29: 1807 – 1809.C
- Kodam, K.M. and Gawai, K.R. (2006). Decolorisation of Reactive Red 11 and 152 azo dyes under aerobic conditions. *Indian Journal of Biotechnology* 5: 422 – 424.
- Kolekar, Y.M., Pawar, S.P., Gawai, K.R., Lokhande, P.D., Shouche, Y.S. and Kodam, K.M. (2008). Decolorization and degradation of Disperse Blue 79 and Acid Orange 10 by *Bacillus fusiformis* KMK5 isolated from the textile dye contaminated soil. *Bioresource Technology* 99: 8999 – 9003.

- Kolekar, Y.M., Nemade, H.N., Markad, V.L., Adav, S.S., Patole, M.S. and Kodam, K.M. (2012). Decolorization and biodegradation of azo dye, reactive blue 59 by aerobic granules. *Bioresource Technology* 104: 818 – 822.
- Kulla, G.H., Klausener, F., Meyer, U., Lüdeke, B. and Leisinger, T. (1983). Interference of aromatic sulfo groups in the microbial degradation of the azo dyes Orange I and Orange II. *Archives of Microbiology* 135: 1 – 7.
- Kumar, K., Devi, S.S., Krishnamurthi, K., Gampawar, S., Mishra, N., Pandya, G.H and Chakrabarti, T. (2005). Decolorisation, biodegradation and detoxification of benzidine based azo dye. *Bioresource Technology* 97(3): 407 – 413.
- Lim, C.K., Bay, H.H., Aris, A., Abdul Majid, Z. and Ibrahim, Z. (2013). Biosorption and biodegradation of Acid Orange 7 by *Enterococcus faecalis* strain ZL: optimization by response surface methodological approach. *Environmental Sciences and Pollution Research* 20(7): 5056 – 5066.
- Lim, C.K. (2014). Biofilm-based macrocomposite for the decolourisation and degradation of Acid Orange 7 azo dye. Doctor Philosophy, Universiti Teknologi Malaysia, Skudai.
- Lim, J., Lee, S., and Hwang, S. (2008). Use of quantitative real-time PCR to monitor population dynamics of ammonia-oxidizing bacteria in batch process. *Journal of Industrial Microbiology and Biotechnology* 35: 1339 – 1344.
- Lin, C., Gan, L. and Chen, Z.L. (2010). Biodegradation of naphthalene by strain *Bacillus fusiformis* (BFN). *Journal of Hazardous Materials* 182: 771 – 777.
- Liu, Z.J., Chen, H.Z., Shaw, N., Hopper, S.L., Chen, L.R., Chen, S.W., Cerniglia, C.E. and Wang, B.C. (2007). Crystal structure of an aerobic FMN-dependent azoreductase(AzoA) from *Enterococcus faecalis*. *Archives of Biochemistry and Biophysics* 463: 68 – 77.
- Lodato, A., Alfieri, F., Olivieri, G., Di Donato, A., Marzocchella, A. and Salatino, P. (2007). Azo-dye conversion by means of *Pseudomonas* sp. OX1. *Enzyme and Microbial Technology* 41: 646 – 652.
- Lopes de Oliveira, P., Duarte, M.C.T, Ponezi, A.N. and Durrant, L.R. purification and partial characterization of manganase peroxidase from *Bacillus pumilus* and *Paenibacillus* sp. *Brazilian Journal of Microbiology* 40: 818 – 826.
- Ma, H.B., Shieh, K.J., Chen, G., Tracy Qiao, X. and Chuang, M.Y. (2006). Application of Real-time Polymerase Chain Reaction (RT-PCR). *The Journal of American Science* 2(2): 1 – 92.

- Maier, J. Kandelbauer, A., Erlacher, A., Cavaco – Paulo, A. and Gübitz, M. (2004). A new alkali – thermostable azoreductase from *Bacillus* sp. strain SF. *Applied and Environmental Microbiology* 70(2): 837 – 844.
- Malinen, E., Kassinen, A., Rinttila, T. and Palva, A. (2003). Comparison of real-time PCR with SYBR Green I or 59-nuclease assays and dot-blot hybridization with rDNA-targeted oligonucleotide probes in quantification of selected faecal bacteria. *Microbiology* 149: 269 – 277.
- Mane, U.V. Gurav, P.N., Deshmukh, A.M. and Govindwar, S.P. (2008). Degradation of textile dye reactive navy – blue Rx (Reactive blue–59) by an isolated Actinomycete *Streptomyces krainskii* SUK – 5. *Malaysian Journal of Microbiology*, 4(2): 1 – 5.
- Mao, F., Leung, W.Y. and Xin, X. (2007). Characterization of EvaGreen and the implication of its physicochemical properties for qPCR applications. *BMC Biotechnology*. DOI:10.1186/1472-6750-7-76.
- Martins, M.A.M., Ferreira, I.C., Santos, I.M., Queiroz, M.J. and Lima, N. (2001). Biodegradation of bioaccessible textile azo dyes by *Phanerochaete chrysosporium*, *Journal of Biotechnology* 89: 91 – 98.
- Mate, M.S. and Pathade, G. (2012). Biodegradation of C.I. Reactive Red 195 by *Enterococcus faecalis* strain YZ66. *World Journal of Microbiology and Biotechnology* 28:815 – 826.
- McMullan, G., Meehan, C., Conneely, A., Kirby, N., Robinson, T., Nigam, P., Banat, I.M., Marchant, R. and Smyth, W.F. (2001). Microbial decolourisation and degradation of textile dyes. *Applied Microbiology and Biotechnology* 56: 81 – 87.
- Melgoza, R.M., Cruz, A. and Buitrón, G. (2004). Anaerobic/aerobic treatment of colorants present in textile effluents. *Water Science and Technology* 50(2): 149 – 155.
- Mendes, S., Farinha, A., Ramos, C.G., Leitao, J.H., Viegas C.A. and Martins, L.O. (2011). Synergistic action of azoreductase and laccase leads to maximal decolourization and detoxification of model dye-containing wastewaters. *Bioresource Technology* 102: 9852 – 9859.
- Méndez-Paz, D., Omil, F. and Lema, J.M. (2003). Modeling of the Acid Orange 7 anaerobic biodegradation. *Water Science and Technology* 48(6): 133 – 139.

- Méndez-Paz, D., Omil, F. and Lema, J.M. (2005). Anaerobic treatment of azo dye Acid Orange 7 under fed-batch and continuous conditions. *Water Research* 39: 771 – 778.
- Méndez-Paz, D., Omil, F. and Lema, J.M. (2005a). Anaerobic treatment of azo dye Acid Orange 7 under batch conditions. *Enzyme and Microbial Technology* 36: 264 – 272.
- Miller, G.L., Blum, R., Glennon, W.E. and Blurton, A.L., (1960). Measurement of carboxymethylcellulase activity. *Analytical Biochemistry* 1: 127 – 132.
- Misal, S.A., Lingojar, D.P., Shinde, R.M. and Gawai, K.R. (2011). Purification and characterization of azoreductase from alkaliphilic strain *Bacillus badius*. *Process Biochemistry* 46: 1264 – 1269.
- Modi, H.A., Rajput, G. and Ambasana, C. (2010). Decolorization of water soluble azo dyes by bacterial cultures, isolated from dye house effluent. *Bioresource Technology* 101: 6580 – 6583.
- Mohd RamLan, M.A., Azizan, N.A., Bay, H.H, Lim, C.K., Mohamad, S.E. and Ibrahim, Z. (2013). Decolourisation of Reactive Black 5 by Azoreductase Produced by *Brevibacillus panacihumi* ZBI. *Journal Teknologi* 59: 11 – 16.
- Moosvi, S., Keharia, H. and Madamwar, D. (2005). Decolourization of textile dye Reactive Violet 5 by a newly isolated bacterial consortium RVM 11.1. *World Journal of Microbiology and Biotechnology* 21: 667 – 672.
- Moosvi, S., Kher, X. and Madamwar, D. (2007). Isolation, characterization and decolorization of textile dyes by a mixed bacterial consortium JW-2. *Dyes and Pigments* 74(3): 723 – 729.
- Moutaouakkil, A., Zeroual, Y., Dzayri, F.Z., Talbi, M., Lee, K. and Blaghen, M. (2003). Bacterial decolorization of the azo dye methyl red by *Enterobacter agglomerans*. *Annals of Microbiology* 53: 161 – 169.
- Moutaouakkil, A., Zeroual, Y., Dzayri, F.Z., Talbi, M., Lee, K. and Blaghen, M. (2003a). Purification and partial characterization of azoreductase from *Enterobacter agglomerans*. *Archives of Biochemistry and Biophysics* 413: 139 – 146.
- Morrison, J.M. and John, G.H. (2013). The non-enzymatic reduction of azo dyes by flavin and nicotinamide cofactors under varying conditions. *Anaerobe* 23: 87 – 96.

- Muda, K., Aris, A., Salim, M.R., Ibrahim, Z., Yahya, A., van Loosdrecht, M.C.M., Ahmad, A. and Nawahwi, M.Z. (2010). Development of granular sludge for textile wastewater treatment. *Water Research* 44: 4341 – 4350.
- Nachiyar, C.V. and Rajkumar, G.S. (2003). Degradation of a tannery and textile dye, Navitan Fast Blue S5R by *Pseudomonas aeruginosa*. *World Journal of Microbiology and Biotechnology* 19: 609 – 614.
- Nachiyar, C.V. and Rajakumar, G.S. (2005). Purification and characterization of an oxygen insensitive azoreductase from *Pseudomonas aeruginosa*. *Enzyme and Microbial Technology* 36: 503 – 509.
- Nakamiya, K., Ooi, S. and Kinoshita, T. (1997). Non-haem hydroquinone peroxidase from *Azotobacter beijerinckii* HM121. *Journal of Fermentation and Bioengineering* 84: 14 – 21.
- Neoh, C.H., Yahya, A., Adnan, R., Abdul Majid, Z. and Ibrahim, Z. (2013). Optimization of decolorization of palm oil mill effluent (POME) by growing cultures of *Aspergillus fumigatus* using response surface methodology. *Environmental Sciences and Pollution Research*. 20(5): 2912 – 2923.
- Nicholas, K.B. and Nicholas, H.J. (1997). GeneDoc: a tool for editing and annotating multiple sequence alignments. Retrieved from www.psc.edu/biomed/genedoc
- Nigam, P., Mc Mullan, G., Banat, I.M. and Marchant, R. (1996). Decolourisation of effluent from the textile industry by a microbial consortium. *Biotechnology letters* 18(1): 117 – 120.
- Novotny, C., Svobodova, K., Erbanova, P., CajthamL, T., Kasinath, A., Lang, E. and Sasek, V. (2004) Ligninolytic fungi in bioremediation: extracellular enzyme production and degradation rate. *Soil Biology and Biochemistry* 36: 1545 – 1551.
- Oh, Y.K., Kim, Y.J., Ahn, Y., Song, S.K. and Park, S. (2004). Color removal of real textile wastewater by sequential anaerobic and aerobic reactors. *Biotechnology and Bioprocess Engineering* 9: 419 – 422.
- O'Neill, C., Hawkes, F. R., Hawkes, D. L., Lourenco, N. D., Pinheiro, H. M. and Delee, W. (1999). Color in textile effluents-sources, measurement, discharge consents and simulation: a review. *Journal of Chemical Technology and Biotechnology* 74: 1009 – 1018.
- O'Neill, C., Lopez, A., Esteves, S., Hawkes, F.R., Hawkes, D.L. and Wilcox, S. (2000). Azo-dye degradation in an anaerobic-aerobic treatment system

- operating on simulated textile effluent. *Applied Microbiology and Biotechnology* 53: 249 – 254.
- Ong, S.A., Toorisaka, E., Hirata, M. and Hano, T. (2005). Treatment of azo dye Orange II in a sequential anaerobic and aerobic sequencing batch reactor system. *Environmental Chemistry Letters* 2(4): 203 – 207.
- Ong, S.A., Toorisaka, E., Hirata, M. and Hano, T. (2005a). Treatment of azo dye Orange II in aerobic and anaerobic-SBR systems. *Process Biochemistry* 40: 2907 – 2914.
- Organization for Economic Cooperation and Development (1984). Terrestrial Plants: Growth test, OECD guidance for testing chemicals, No.208, Paris.
- Oturkar, C.C., Nemade, H.N., Mulik, P.M., Patole, M.S., Hawaldar, R.R. and Gawai, K.R. (2011). Mechanistic investigation of decolorization and degradation of Reactive Red 120 by *Bacillus lentus* BI377. *Bioresource Technology* 102: 758 – 764.
- Özcan, A., Oturan, M.A., Oturan, N. and Şahin, Y. (2009). Removal of Acid Orange 7 from water by electrochemically generated Fenton's reagent. *Journal of Hazardous Materials* 163(2 – 3): 1213 – 1220.
- Pandey, A., Singh, P. and Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *International Biodeterioration and Biodegradation* 59(2): 73 – 84.
- Passardi, F., Bakalovic, N., Teixeira, F.K., Margis-Pinheiro, M., Penel, C. and Dunand, C. (2007). Prokaryotic origins of the non-animal peroxidase superfamily and organelle-mediated transmission to eukaryotes. *Genomics* 3: 567 – 579.
- Parshetti, G., Kalme, S., Saratale, G. and Govindwar S. (2006). Biodegradation of Malachite Green by *Kocuria rosea* MTCC 1532. *Acta Chimica Slovenica* 53: 492 – 498.
- Patil, P.S., Shedbalkar, U.U. Kalyani, D.C. and Jadhav, J.P. (2008). Biodegradation of Reactive Blue 59 by isolated bacterial consortium PMB11. *Journal of Industrial Microbiology and Biotechnology* 35: 1181 – 1190.
- Pearce, C.I., Lloyd, J.R. and Guthrie, J.T. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments* 58(3): 179 – 196.

- Pereira, L., Coelho, A.V., Viegas, C.A., Santos, M.M., Robalo, M.P. and Martins, L.O. (2009). Enzymatic biotransformation of the azo dye Sudan Orange G with bacterial CotA-laccase. *Journal of Biotechnology* 139: 68 – 77.
- Phugare, S.S., Kalyani, D.C., Surwase, S.N. and Jadhav, J.P. (2011). Ecofriendly degradation, decolorization and detoxification of textile effluent by a developed bacterial consortium. *Ecotoxicology and Environmental Safety* 74: 1288 – 1296.
- Pinheiro, H.M., Touraud, E. and Thomas, O. (2004). Aromatic amines from azo dye reduction: status review with emphasis on direct UV spectrophotometric detection in textile industry wastewaters. *Dyes and Pigments* 61: 121 – 139.
- Plumb, J.J., Bell, J., and Stuckey, D.C. (2001). Microbial populations associated with treatment of an industrial dye effluent in an anaerobic baffled reactor. *Applied and Environmental Microbiology* 67: 3226 – 3235.
- Pourbabaee, A.A. and Malekzadeh, F. (2005). Decolorization of Methyl Orange (as a model azo dye) by the newly discovered *Bacillus Sp.* *Iranian Journal of Chemistry and Chemical Engineering* 24(3): 41 – 45.
- Pourbabaee, A.A., Malekzadeh, F., Sarbolouki, M.N. and Najafi, F. (2006). Aerobic decolorization and detoxification of a disperse dye in textile effluent by a new isolate of *Bacillus sp.* *Biotechnology and Bioengineering* 93(4): 631 – 635.
- Rahmani, A.R., Zarrabi, M., Samarghandi, M. R., Afkhami, A. and Ghaffari, H. R. (2010). Degradation of azo dye Reactive Black 5 and Acid Orange 7 by Fenton-like mechanism. *Iranian Journal of Chemical Engineering* 7(1): 87 – 94.
- Ramalho, P.A., Cardoso, M.H., Cavaco-Paulo, A. and Ramalho, T. (2004). Characterization of azo reduction activity in a novel Ascomycete Yeast strain. *Applied and Environmental Microbiology* 70(4): 2279 – 2288.
- Rajaguru, P., Kalaiselvi, K., Palanivel, M. and Subburam, V. (2000). Biodegradation of azo dyes in a sequential anaerobic–aerobic system. *Applied Microbiology and Biotechnology* 54: 268 – 273.
- Rajee, O. and Patterson, J. (2011). Decolorization of azo dye (Orange MR) by an autochthonous bacterium, *Micrococcus sp.* DBS 2. *Indian Journal of Microbiology* 51(2): 159 – 163.
- Rau, J., Maris, B., Kinget, R., Samyn, C., van den Mooter, G. and Stolz A. (2002). Enhanced anaerobic degradation of polymeric azo compounds by *Escherichia*

- coli* in the presence of low-molecular-weight redox mediators. *Journal of Pharmacy and Pharmacology* 54: 1471 – 1479.
- Razo – Flores, E., Luijten, M., Donlon, B.A., Lettinga, G. and Field, J.A. (1997). Complete biodegradation of the azo dye azodisalicylate under anaerobic conditions. *Environmental Science and Technology* 31: 2098 – 2103.
- Rehorek, A. and Plum, A. (2006). Online LC-MS-MS process monitoring for optimization of biological treatment of wastewater containing azo dye concentrates. *Analytical and Bioanalytical Chemistry* 384: 1123 – 1128.
- Ren, S.Z., Guo, J., Zeng, G.Q. and Sun, G.P. (2006). Decolorization of triphenylmethane, azo, and anthraquinone dyes by a newly isolated *Aeromonas hydrophila* strain. *Environmental Biotechnology* 72: 1316 – 1321.
- Renganathan, S., Thilagaraj, W.R., Miranda, L.R., Gautam, P. and Velan, M. (2006). Accumulation of Acid Orange 7, Acid Red 18 and Reactive Black 5 by growing *Schizophyllum commune*. *Bioresource Technology* 97: 2189 – 2193.
- Rozen, S. and Skaletshy, H.J. (2000). *Primer 3 on the WWW for general users and for biologist programmers*. In: Krawetz S, Misener S (eds) *Bioinformatic Methods and Protocols: Methods in Molecular Biology* pg: 365 – 386. Humana Press, Totowa, NJ.
- Sahasrabudhe, M. M. and Pathade, G. R. (2011). Decolourization of C.I. Reactive Yellow 145 by *Enterococcus faecalis* strain YZ66. *Archives of Applied Science Research* 3 (3): 403 – 414.
- Sandhya, S. (2010). Biodegradation of azo dyes under anaerobic condition: Role of azoreductase. *Biodegradation of Azo dyes*. H. Atacag Erkurt (ed.). *The Handbook of Environmental Chemistry* 9: 39 – 57, London: Springer – Verlag.
- Saraswathi, K. and Balakumar, S. (2009). Biodecolourization of azo dye (pigmented red 208) using *Bacillus firmus* and *Bacillus laterosporus*. *Journal of Bioscience and Technology* 1(1): 1 – 7.
- Saratale, R.G., Saratale, G.D., Kalyani, D.C., Chang, J.S. and Govindwar, S.P. (2009). Enhanced decolorization and biodegradation of textile azo dye Scarlet R by using developed microbial consortium-GR. *Bioresource Technology* 100: 2493 – 2500.
- Saratale, R.G., Saratale, G.D., Chang, J.S., and Govindwar, S.P. (2009a). Decolorization and biodegradation of textile dye Navy Blue HER by

- Trichosporon beigelii* NCIM-3326. *Journal of Hazardous Materials* (166): 1421 – 1428.
- Saratale, R. G., Saratale, G. D., Chang, J. S. and Govindwar, S. P. (2010). Decolorization and biodegradation of reactive dyes and dye wastewater by a developed bacterial consortium. *Biodegradation* 21: 999 – 1015.
- Saratale, R.G., Saratale, G.D., Chang, J.S. and Govindwar S.P. (2011). Bacterial decolorization and degradation of azo dyes: A review. *Journal of the Taiwan Institute of Chemical Engineers* 42: 138 – 157.
- Saratale, R.G., Gandhi, S. S., Purankar, M.V., Kurade, M.B., Govindwar, S.P., Oh, S.E. and Saratale, G.D. (2013). Decolorization and detoxification of sulfonated azo dye C.I Remazol Red and Textiel effluent by isolated *Lysinibacillus* sp. RGS. *Journal of Bioscience and Bioengineering* 115(6): 658 – 667.
- Sarrau, B.D., Clavel, T., Clerte, C., Carlin, F., Ginies, C. and Nguyen, C. (2012) Influence of anaerobiosis and low temperature on *Bacillus cereus* growth, metabolism and membrane properties. *Applied and Environmental Microbiology* 78(6): 1715 – 1723.
- Sharma, D. K., Saini, H. S., Singh, M., Chimni S. S. and Chadha B.S. (2004). Biodegradation of acid blue-15, a textile dye, by an up-flow immobilized cell bioreactor. *Journal of Industrial Microbiology and Biotechnology* 31: 109 – 114.
- Sheth, N. T. and Dave, S. R. (2009). Optimisation for enhanced decolourization and degradation of Reactive Red BS C.I. 111 by *Pseudomonas aeruginosa* NGKCTS. *Biodegradation* 20:827 – 836.
- Shida, O., Takagi, H., Kadowaki, K. and Komagata, K. (1996). Proposal for two new genera, *Brevibacillus* gen. nov. and *Aneurinibacillus* gen. nov. *International Journal of Systematic and Evolutionary Microbiology* 46(4): 939 – 46.
- Shin, S.G., Lee, S., Lee, C., Hwang, K. and Hwang, S. (2010). Qualitative and quantitative assessment of microbial community in batch anaerobic digestion of secondary sludge. *Bioresource Technology* 101: 9461 – 9470.
- Singh, P., Iyengar, L. and Pandey, A. (2012). Bacterial decolorization and degradation of azo dyes. *Microbial Degradation of Xenobiotics*. S.N. Singh

- (ed.), in *Environmental Science and Engineering* (pp 101 – 133). Springer Berlin Heidelberg.
- Sinha, S., Chattopadhyay, P., Pan, I, Chatterjee, S., Chanda, P., Bandyopadhyay, D., Das, K. and Sen, S.K. (2009). Microbial transformation of xenobiotics for environmental bioremediation. *African Journal of Biotechnology* 8(22): 6016 – 6027.
- Slokar, Y.M. and Le Marechal, A.M. (1998). Methods of decoloration of textile wastewater. *Dyes and Pigments* 37(4): 355 – 356.
- Song, M., Shin, S.G. and Hwang, S. (2010). Methanogenic population dynamics assessed by real-time quantitative PCR in sludge granule in upflow anaerobic sludge blanket treating swine wastewater. *Bioresource Technology* 101: 523 – 528.
- Spagni, A., Grilli, S., Casu, S. and Mattioli, D. (2010). Treatment of a simulated textile wastewater containing the azo-dye reactive orange 16 in an anaerobic-biofilm anoxic-aerobic membrane bioreactor. *International Biodeterioration & Biodegradation* 64: 676 – 681.
- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of azo dyes. *Applied Microbiology and Biotechnology* 56(1 – 2): 69 – 80.
- Sugumar, R.W. and Sadanandan, S. (2010). Combined anaerobic-aerobic bacterial degradation of dyes. *E-Journal of Chemistry* 7(3): 739 – 744.
- Supaka, N., Juntongjin, K., Damronglerd, S., Delia, M.L. and Strehaiano, P. (2004). Microbial decolorization of reactive azo dyes in a sequential anaerobic-aerobic system. *Chemical Engineering Journal* 99: 169 – 176.
- Suzuki, Y., Yoda, T., Ruhul, A. and Sugiura, W. (2001). Molecular cloning and characterization of the gene coding for Aazoreductase from *Bacillus* sp. OY1-2 isolated from soil. *The Journal of Biological Chemistry* 276(12): 9059 – 9065.
- Tamboli, D.P., Gomare, S.S., Kalme, S.S., Jadhav, U.U. and Govindwar, S.P. (2010). Degradation of Orange 3R, mixture of dyes and textile effluent and production of polyhydroxyalkanoates from biomass obtained after degradation. *International Biodeterioration and Biodegradation* 64: 755 – 763.
- Tan, N.C.G., van Leeuwen, A., van Voorthuizen, E.M., Slenders, P., Prenafeta-Boldu, F.X., Temmink, H., Lettinga, G. and Field, J.A. (2005). Fate and

- biodegradability of sulfonated aromatic amines. *Biodegradation* 16: 527 – 537.
- Telke, A.A., Kalyani, D.C., Jadhav, J.P. and Govindwar, S.P. (2008). Kinetics and mechanism of reactive red 141 degradation by a bacterial isolate *Rhizobium radiobacter* MTCC 8161, *Acta Chimica Slovenica* 55: 320 – 329.
- Telke, A.A., Kalyani, D.C., Dawkar, V.V. and Govindwar, S.P. (2009). Influence of organic and inorganic compounds on oxidoreductive decolorization of sulfonated azo dye C.I. Reactive Orange 16. *Journal of Hazardous Materials* 172(1): 298 – 309.
- Telke, A.A., Joshi, S.M., Jadhav, S.U., Tamboli, D.P. and Govindwar, S.P. (2010). Decolorization and detoxification of Congo Red and textile industry effluent by an isolated bacterium *Pseudomonas* sp. SU-EBT. *Biodegradation* (21): 283 – 296.
- Tessonnière, H., Vidal, S., Barnavon, L., Alexandre, H. and Remize, F. (2009). Design and performance testing of a real-time PCR assay for sensitive and reliable direct quantification of *Brettanomyces* in wine. *International Journal of Food Microbiology* 129: 237 – 243.
- Tony, B.D., Goyal, D. and Khanna, S. (2009). Decolorization of Direct Red 28 by mixed bacterial culture in an up-flow immobilized bioreactor. *Journal of Industrial Microbiology and Biotechnology* 36: 955 – 960.
- US Environmental Protection Agency (1982). Seed germination/root elongation toxicity tests EC12, Office of Toxic Substances, Washington, DC.
- US Food and Drug Administration (1987). Seed germination and root elongation, environmental assessment technical assistance document 406, Center for Food Safety and Applied nutrition, Center for Veterinary Medicine, Washington, DC.
- Velegraki, T., Poullos, I., Charalabaki, M., Kalogerakis, N., Samaras, P. and Mantzavinos, D. (2006). Photocatalytic and sonolytic oxidation of Acid Orange 7 in aqueous solution. *Applied Catalysis B: Environmental* 62: 159 – 168.
- Vijaykumar, M.H., Vaishampayan, P.A., Shouche, Y.S. and Karegoudar, T.B. (2007). Decolourization of naphthalene-containing sulfonated azo dyes by *Kerstersia* sp. strain VKY1. *Enzyme and Microbial Technology* 40: 204 – 211.

- Viliesid F. and Lily, M.D. (1992). Influence of dissolved oxygen tension on the synthesis of catechol 1,2-dioxygenase by *Pseudomonas putida*. *Enzyme and Microbial Technology* 14: 561 – 565.
- Vinas, M., Sabate, J., Espuny, M.J. and Solanas, A.M. (2005). Bacterial community dynamics and polycyclic aromatic hydrocarbon degradation during bioremediation of heavily creosote-contaminated soil. *Applied and Environmental Microbiology* 71(11): 7008 – 7018.
- Wang, J., Liu, G.F., Lu, H., Jin, R.F., Zhou, J.T. and Lei, T.M. (2012). Biodegradation of Acid Orange 7 and its auto-oxidative decolourization product in membrane-aerated biofilm reactor. *International Biodeterioration and Biodegradation* 67: 73 – 77.
- Wang, W.J., Chen, K.S. and Xu, C.J. (2006). DNA quantification using EvaGreen and a real-time PCR instrument. *Analytical Biochemistry* 356: 303 – 305.
- Wang, X., Cheng, X., Sun, D. and Qi, H. (2008). Biodecolorization and partial mineralization of Reactive Black 5 by a strain of *Rhodopseudomonas palustris*. *Journal of Environmental Sciences* 20: 1218 – 1225.
- Wang, X.D., Sun, C., Gao, S.X., Wang, L.S. and Shuokui, H. (2001). Validation of germination rate and root elongation as indicator to assess phytotoxicity with *Cucumis sativus*. *Chemosphere* 44(8): 1711 – 1721.
- Wesenberg, D., Buchon, F and Agathos, S.N. (2002). Degradation of dye-containing textile effluent by the agaric white-rot fungus *Clitocybula dusenii*. *Biotechnology Letters* 24: 989 – 993.
- Wong, P.K. and Yuen, P.Y. (1998). Decolourization and biodegradation of N,N-dimethyl-p-phenylenediamine by *Klebsiella pneumonia* RS-13 and *Acetobacter liquefaciens* S-1. *Journal of Applied Microbiology* 85: 79 – 87.
- Wuhrmann, K., Mechsner, K. I. and Kappeler, T. (1980). Investigation on rate determining factors in the microbial reduction of azo dyes. *European Journal of Applied Microbiology and Biotechnology* 9: 325 – 338.
- Xu, H., Heinze, T.M., Chen, S., Cerniglia, C.E. and Chen, H. (2007). Anaerobic metabolism of 1-amino-2-naphthol-based azo dyes (Sudan Dyes) by human intestinal microflora. *Applied and Environmental Microbiology* 73(23): 7759 – 7762.

- Xu, L., Zhao, H.Z., Shi, S.Y., Zhang, G.Z. and Ni, J.R. (2008). Electrolytic treatment of CI Acid Orange 7 in aqueous solution using a three-dimensional electrode reactor. *Dyes and Pigments* 77: 158 – 164.
- Xu, M.Y., Guo, J. and Sun, G.P. (2007). Biodegradation of textile azo dye by *Shewanella decolorationis* S12 under microaerophilic conditions. *Applied Microbiology and Biotechnology* 76: 719 – 726.
- Yatome, C., Ogawa, T., Koga, D. and Idaka, E. (1981). Biodegradation of Azo and triphenylmethane dyes by *Pseudomonas pseudomallei* 13na. *Journal of the Society of Dyers and Colourists* 97: 166 – 169.
- Yemashova, N., Kalyuzhnyi, S. (2006). Microbial conversion of selected azo dyes and their breakdown products. *Water Science and Technology* 53 (11): 163 – 171.
- Yesiladal, S.K., Pekin, G., Bermek, H., Arslan-Alaton, I., Orhon, D. and Tamerler, C. (2006). Bioremediation of textile azo dyes by *Trichophyton rubrum* LSK-27. *World Journal of Microbiology & Biotechnology* 22: 1027 – 1031.
- Yoo, E.S., Libra, J. and Adrian, L. (2001). Mechanism of decolorization of azo dyes in anaerobic mixed culture. *Journal of Environmental Engineering* 127(9): 844 – 849.
- Zee van der, F.P., Lettinga, G. and Field, J.A. (2000). The role of (auto)catalysis in the mechanism of anaerobic azo reduction. *Water Science and Technology* 42: 301 – 308.
- Zee van der, F.P., Lettinga, G. and Field, J.A. (2001). Azo dye decolourisation by anaerobic granular sludge. *Chemosphere* 44: 1169 – 1176.
- Zee van der, F.P., Bisschops, I.A.E., Blanchard, V.G., Bouwman, R.H.M., Letting, G. and Field, J.A. (2003). The contribution of biotic and abiotic processes during azo dye reduction in anaerobic sludge. *Water Research* 37: 3098 – 3109.
- Zee van der, F.P. and Villaverde, S. (2005). Combined anaerobic–aerobic treatment of azo dyes – A short review of bioreactor studies. *Water Research* 39: 1425 – 1440.
- Zhang, F., Knapp, J.S. and Tapley, K.N. (1999). Development of bioreactor systems for decolorization of Orange II using white rot fungus. *Enzyme and Microbial Technology* 24(1 – 2): 48 – 53.
- Zhang, R., Wei, M., Ji, H., Chen, X., Qiu, G. and Zhou, H. (2009). Application of real-time PCR to monitor population dynamics of defined mixed cultures of

- moderate thermophiles involved in bioleaching of chalcopyrite. *Applied Microbiology and Biotechnology* 81: 1161 – 1168.
- Zhao, D.F., Wu, W.L., Zhang, Y.B., Liu, Q.Y., Yang, H.B. and Zhao, C.C. (2011). Degrading Bacterium *Bacillus fusiformis* sp. and Influence of Environmental Factors on Degradation Efficiency. *China Petroleum Processing and Petrochemical Technology* 13(4): 74 – 82.
- Zhao, H.Z., Sun, Y., Xu, L.N. and Ni, J.R. (2010). Removal of Acid Orange 7 in simulated wastewater using a three-dimensional electrode reactor: removal mechanisms and dye degradation pathway. *Chemosphere* 78(1): 46 – 51.
- Zhao, X.Q., Yang, L.Y. and Yu, Z.Y. (2008). Characterization of depth-related microbial communities in lake sediments by denaturing gradient gel electrophoresis of amplified 16S rRNA fragments. *Journal of Environmental Sciences* 20: 224 – 230.
- Zollinger, H. (2003). Color Chemistry. Syntheses, properties and applications of organic dyes and pigments. Weinheim: VCH Publishers.
- Zumstein, E., Moletta, R. and Godon, J.J. (2000). Examination of two years of community dynamics in an anaerobic bioreactor using fluorescence polymerase chain reaction (PCR) single-strand conformation polymorphism analysis. *Environmental Microbiology* 2(1): 69 – 78.