

PENGGUNAAN SISA UDANG UNTUK PENGHASILAN KITINASE OLEH
Trichoderma virens MENGGUNAKAN FERMENTASI KEADAAN PEPEJAL

RACHMAWATY

Tesis ini dikemukakan sebagai memenuhi
Syarat penganugerahan ijazah
Doktor Falsafah (Biosains)

Fakulti Biosains dan Kejuruteraan Perubatan
Universiti Teknologi Malaysia

FEBRUARI 2015

Dikhaskan untuk yang saya sayangi dan sentiasa menyokong saya
Ayah dan mama,
Suami saya Ferry eko cahyono
Anak-anak saya, ahli keluarga dan kawan-kawan
Orang-orang yang memberi inspirasi kepada saya untuk penyelesain pengajian ini

PENGHARGAAN

Dengan nama Allah Yang Maha Pengasih Lagi Maha Penyayang. Setinggi-tinggi kesyukuran dipanjatkan kepada Allah SWT, kerana dengan limpahan rahmat dan karunia serta izin-Nya jualah kajian ini dapat disempurnakan.

Saya ingin merakamkan penghargaan dan terima kasih kepada penyelia saya, PM. Dr. Madidah Md Salleh atas segala ilmu, tunjuk ajar, bimbingan, nasihat dan dorongan yang diberi sepanjang tempoh penyelidikan ini dijalankan.

Penghargaan ini juga saya tujukan buat kaki tangan makmal Fakulti Biosains dan Kejuruteraan Perubatan yang banyak membantu bagi penyelidikan ini iaitu Puan Fatimah dan Cik Sarah. Ucapan terima kasih yang tidak terhingga ditujukan buat rakan-rakan yang telah banyak memberi sokongan dari segi ilmu dan moral iaitu Noratiqah Binti Kamsani, Puan Huszalina Hussin, Anisah Jamil, Nurrazzean Haireen Mohd. Tumpang, Nurashikin Ihsan, Shankar A/L Ramanthan, Ang Siow Kuang, Ahmad Fawwaz Mohd Raji, Hartati dan Halifah Pagarra.

Jutaan terima kasih yang tak terhingga dikirimkan buat ibu saya Sutinah Siradju yang tidak pernah lupa mendoakan kejayan dan kebahagiaan anakanda. Penghargaan ini juga buat suami tersayang Ferry Eko Cahyono serta anak-anak tercinta, Dzaqqiyah dan Muh. Fahmi dan kakak beradik sekalian yang senantiasa memberikan semangat dan kiriman doa.

ABSTRAK

Penghasilan kitinase oleh *Trichoderma virens* menggunakan sisa udang sebagai substrat telah dikaji melalui fermentasi keadaan pepejal dengan kandungan lembapan pada 70%. Enam kaedah prarawatan yang berbeza iaitu prarawatan ketuhar, gelombang mikro, pendidihan dan penghancuran, pengeringan suria, dan kimia telah dilakukan terhadap sisa udang dengan sisa udang tanpa prarawatan dijadikan sebagai kawalan. Aktiviti kitinase tertinggi diperoleh daripada prarawatan gelombang mikro pada hari ketiga fermentasi dengan aktiviti kitinase pada 0.194 U/g IDS, 3.2 kali lebih tinggi berbanding kawalan (0.06 U/g IDS). Kajian terhadap kesan sumber nitrogen terhadap penghasilan kitinase menggunakan reka bentuk faktorial umum menunjukkan yang ammonium sulfat dengan 30.29 mM nitrogen memberi kesan yang signifikan berbanding ekstrak yis dengan 7.43 mM nitrogen. Reka bentuk faktorial 2-peringkat, masa eraman, suhu, dan kelembapan substrat juga memberi kesan yang signifikan kepada penghasilan kitinase. Reka bentuk komposit berpusat (RBKB) digunakan dalam mengoptimumkan keadaan bagi penghasilan kitinase sisa udang melalui fermentasi keadaan pepejal. Penghasilan kitinase didapati meningkat 2.46 kali ganda (0.487 U/g IDS) pada keadaan optimum iaitu pada suhu 27.9 °C, kelembapan substrat 54.5% dan enam hari masa pengeraman. Bagi pencirian separa kitinase, suhu dan pH optimum masing-masing adalah pada 60°C dan pH 3.0. Kitinase mengekalkan 72% aktiviti pada suhu 70°C. Walau bagaimanapun, kehilangan jumlah aktiviti kitinase berlaku selepas 60 minit pengeraman pada suhu 70°C dan 80°C dengan sisa aktiviti masing-masing 48% dan 28%. Kitinase lebih stabil dalam pH berasid daripada pH beralkali. Berat molekul kitinase adalah 50 dan 42 kDa bagi endokitinase, 33 dan 25 kDa bagi eksokitinase dan 18 kDa bagi protease. Pengekstrakan kitinase mentah dari *Trichoderma virens* dapat merencatkan pertumbuhan *Ganoderma boninense*.

ABSTRACT

The chitinase production by *Trichoderma virens* using shrimp waste as a substrate was studied in solid state fermentation with 70% of moisture content. Six different pretreatment methods namely oven pretreatment, microwave pretreatment, boiling and crushing pretreatment, sun-dried pretreatment and chemical pretreatment were conducted on shrimp waste with non-treated shrimp waste as a control. The highest chitinase activity was obtained from microwave pretreatment on the third day of fermentation with chitinase activity of 0.194 U/g IDS, 3.2 fold higher than the untreated shrimp waste (0.06 U/g IDS). Study on the effect of nitrogen source on chitinase production using general factorial design showed that ammonium sulphate with 30.29 mM nitrogen gave significant effect compared to yeast extract with 7.43 mM nitrogen. Two level factorial design, incubation time, temperature, and substrate moisture also have a significant impact on the production of chitinase. Central composite design (CCD) was used in optimizing the conditions for chitinase production of shrimp waste by solid-state fermentation. Chitinase production was found to have increased 2.46 times (0.487 U/g IDS) at optimum condition: temperature of 27.9 °C, 54.5% of substrate moisture and six days of incubation time. The optimal degradation showed an improvement of chitinase production of 2.46 fold as compared to before optimization using CCD. For partial characterization of chitinase, the optimum temperature and pH are at 60 °C and pH 3.0, respectively. Chitinase retained 72% of its activity at 70 °C. However, the loss of the chitinase activity occurred after 60 minutes of incubation at 70 °C and 80 °C with residual activity are 48% and 28%, respectively. Chitinase was more stable in acidic than in alkaline pH. The molecular weight of chitinase was 50 and 42 kDa for endochitinase, 33 and 25 kDa for eksokitinase and 18 kDa for protease. Extraction of crude chitinase from *Trichoderma virens* can inhibit the growth of *Ganoderma boninense*.

KANDUNGAN

BAB	PERKARA	MUKA SURAT
	PENGAKUAN	ii
	DEDIKASI	iii
	PENGHARGAAN	iv
	ABSTRAK	v
	ABSTRACT	vi
	KANDUNGAN	vii
	SENARAI JADUAL	xiv
	SENARAI RAJAH	xvii
	SENARAI SINGKATAN	xxi
	SENARAI SIMBOL	xxiii
	SENARAI LAMPIRAN	xxiv
1	PENDAHULUAN	1
	1.1 Latar belakang kajian	1
	1.2 Objektif kajian	4
	1.3 Skop kajian	4
2	KAJIAN LITERATUR	5
	2.1 Kitinase	5
	2.2 Mekanisme tindak balas kitinase dalam hidrolisis kitin.	6
	2.3 Sumber kitinase	8
	2.3.1 Mikroorganisma penghasil kitinase	9
	2.3.2 <i>Trichoderma virens</i> sebagai penghasil kitinase	11

2.4	Substrat bagi penghasilan kitinase	12
2.4.1	Sisa udang sebagai substrat	12
2.4.2	Kitin sebagai substrat	13
2.4.2.1	Aplikasi kitin	17
2.5	Penggunaan kitinase	18
2.6	Prarawatan sisa udang	20
2.7	Penghasilan kitinase	21
2.7.1	Fermentasi	21
2.7.1.1	Fermentasi Tenggelam (FmT)	22
2.7.1.2	Fermentasi keadaan pepejal (FKP)	23
2.7.2	Perbandingan sistem penghasilan kitinase	27
2.8	Faktor mempengaruhi penghasilan kitinase dalam FKP	28
2.8.1	Kandungan kelembapan substrat	29
2.8.2	Jenis dan saiz inokulum	31
2.8.3	Suhu	33
2.8.4	pH	35
2.8.5	Sumber nitrogen	36
2.8.6	Masa pengeraman	36
2.9	Pengoptimuman parameter menggunakan reka bentuk eksperimen	39
2.9.1	Penggunaan reka bentuk eksperimen dalam penghasilan kitinase	39
2.10	<i>Ganoderma boninense</i>	42
2.10.1	Morfologi <i>Ganoderma boninense</i>	42
2.10.2	Pengawalan <i>Ganoderma boninense</i>	45
2.10.3	Mekanisme tindakan enzim <i>Trichoderma</i> sp. sebagai agen biokawalan	46
3	BAHAN DAN METODOLOGI	48
3.1	Mikroorganisma dan penyelenggaraan	48
3.2	Penyediaan inokulum	48
3.3	Agar Dekstrosa Kentang (PDA)	49

3.4	Reka bentuk kajian dan metodologi	49
3.5	Prarawatan sisa udang	52
3.5.1	Rawatan pengeringan suria	52
3.5.2	Rawatan pendidihan dan penghancuran	52
3.5.3	Rawatan ketuhar	53
3.5.4	Rawatan gelombang mikro	53
3.5.5	Rawatan kimia	53
3.5.6	Sisa udang mentah (tidak dirawat)	53
3.6	Medium penghasilan kitinase	53
3.7	Penghasilan kitinase menggunakan FKP	54
3.8	Kaedah analisis	54
3.8.1	Penentuan kitinase	55
3.8.2	Penyediaan kitin berkoloid	55
3.8.3	Penentuan kepekatan protein	55
4	PENYARINGAN KAEDAH PRARAWATAN SISA UDANG BAGI PENGHASILAN KITINASE OLEH <i>Trichoderma virens</i>	57
4.1	Pengenalan	57
4.2	Bahan dan kaedah	58
4.2.1	Metodologi penyelidikan	58
4.2.2	Prarawatan sisa udang	59
4.2.3	Medium fermentasi dan penghasilan kitinase secara FKP	59
4.2.4	Persampelan	61
4.2.5	Analisis	61
4.3	Keputusan dan perbincangan	61
4.3.1	Kesan prarawatan yang berbeza terhadap penghasilan kitinase	61
4.3.2	Kesan prarawatan yang berbeza terhadap struktur sisa udang	68
4.4	Kesimpulan	77

5	KAJIAN KESAN SUMBER NITROGEN TERHADAP PENGHASILAN KITINASE MENGUNAKAN REKA BENTUK FAKTORIAL UMUM	78
5.1	Pengenalan	78
5.2	Bahan dan kaedah	79
5.2.1	Pengiraan nisbah kandungan nitrogen dan karbon	79
5.2.2	Reka bentuk eksperimen untuk penghasilan kitinase	80
5.2.3	Reka bentuk faktorial umum	80
5.2.4	Persampelan	82
5.2.5	Pencerakinan	82
5.3	Keputusan dan perbincangan	82
5.3.1	Kesan sumber nitrogen terhadap penghasilan kitinase menggunakan reka bentuk faktorial umum	82
5.4	Kesimpulan	86
6	PENENTUAN FAKTOR-FAKTOR YANG MEMPENGARUHI PENGHASILAN KITINASE MENGUNAKAN REKA BENTUK FAKTORIAL DUA PERINGKAT	87
6.1	Pengenalan	87
6.2	Bahan dan kaedah	89
6.2.1	Metodologi kajian	89
6.2.2	Substrat dan prarawatan	89
6.2.3	Medium untuk penghasilan kitinase	89
6.2.4	Keadaan kultur	91
6.2.5	Penghasilan kitinase dalam sistem FKP	91
6.2.6	Persampelan	92
6.2.7	Analisis	92
6.3	Keputusan dan Perbincangan	92

6.3.1	Analisis Varians (ANOVA)	93
6.3.2	Analisis respons	96
6.3.3	Kesan utama dan interaksi	99
6.3.4	Plot kebarangkalian normal reja	101
6.4	Kesimpulan	102
7	PENGOPTIMUMAN PENGHASILAN KITINASE OLEH <i>Trichoderma virens</i> DALAM FERMENTASI KEADAAN PEPEJAL	104
7.1	Pengenalan	104
7.2	Bahan dan kaedah	105
7.2.1	Metodologi kajian	105
7.2.2	Mikroorganisma	107
7.2.3	Substrat dan prarawatan	107
7.2.4	Medium dan penghasilan kitinase	107
7.2.5	Kaedah analisis	107
7.2.6	Reka bentuk eksperimen	108
7.3	Keputusan dan Perbincangan	111
7.3.1	Reka bentuk Komposit Berpusat (RBKB)	111
7.3.2	Pembentukan model	111
7.3.3	Plot respon permukaan	113
7.3.4	Pengoptimuman dan pengesahan model	118
7.4	Kesimpulan	121
8	PENCIRIAN SEPARA KITINASE MENTAH YANG DIHASILKAN OLEH <i>Trichoderma virens</i> DALAM FERMENTASI FASA PEPEJAL	122
8.1	Pengenalan	122
8.2	Bahan dan kaedah	123
8.2.1	Metodologi kajian	123
8.2.2	Mikroorganisma	125
8.2.3	Medium fermentasi	125

8.2.4	Penghasilan kitinase dalam FKP	125
8.2.5	Persampelan	125
8.2.6	Analisis	126
8.2.7	Penyediaan larutan penimbal yang berbeza	126
8.2.8	Pencirian separa kitinase mentah	126
8.2.8.1	Pengenalpastian suhu optimum dan kestabilan haba	127
8.2.8.2	Pengenalpastian pH optimum dan kestabilan pH	128
8.2.8.3	Aktiviti relatif	128
8.2.8.4	Aktiviti residual	129
8.2.9	Pemendakan Asid Trikloroasetik (TCA)	129
8.2.10	Penentuan berat molekul kitinase menggunakan elektroforesis gel natrium dodesilsulfat poliakrilamid (SDS-PAGE)	129
8.3	Keputusan dan perbincangan	130
8.3.1	Suhu optimum dan kestabilan haba	130
8.3.2	pH optimum dan kestabilan pH	133
8.3.3	Penentuan berat molekul kitinase mentah	136
8.4	Kesimpulan	138
9	POTENSI KITINASE MENTAH DIHASILKAN OLEH <i>Trichoderma virens</i> SEBAGAI ANTIMIKROB KEPADA <i>Ganoderma boninense</i>	140
9.1	Pengenalan	140
9.2	Bahan dan kaedah	141
9.2.1	Mikroorganisma	141
9.2.1.1	Kultur <i>Trichoderma virens</i>	141
9.2.1.2	Kultur <i>Ganoderma boninense</i>	142
9.2.2	Medium fermentasi	142
9.2.3	Penghasilan kitinase melalui FKP	142
9.2.4	Persampelan	142
9.2.5	Analisis	144

9.2.6	Tindakan antikulat oleh antimikrob	144
9.2.6.1	Ujian cerakin kultur berkembar	144
9.2.6.2	Ujian turasan kultur	145
9.3	Keputusan dan perbincangan	145
9.3.1	Pertumbuhan koloni <i>T. virens</i> dan <i>G. boninense</i>	145
9.3.2	Perencatan <i>T. virens</i> terhadap <i>G. boninense</i>	147
9.3.3	Turasan Kultur	149
9.4	Kesimpulan	152
10	KESIMPULAN UMUM DAN KAJIAN LANJUTAN	153
10.1	Kesimpulan umum	153
10.2	Kajian lanjutan	155
	RUJUKAN	157
	Lampiran A – H	181 - 195

SENARAI JADUAL

NO. JADUAL	TAJUK	MUKA SURAT
2.1	Jenis-jenis kitinase dan fungsinya	6
2.2	Mod tindakan menghidrolisi kitinase	8
2.3	Peranan kitinase dalam filum yang berbeza	9
2.4	Mikroorganisma penghasil kitinase	10
2.5	Peratus komposisi proksimat (%) berdasarkan berat kering sisa kulit krustasia.	13
2.6	Jenis struktur kitin	17
2.7	Penggunaan kitin, kitosan dan bahan terbitannya	18
2.8	Penggunaan kitinase	19
2.9	Perawatan fizikal dan kimia terhadap penguraian sisa udang	21
2.10	Penghasilan kitinase melalui fermentasi keadaan pepejal oleh mikroorganisma berbeza.	27
2.11	Perbandingan antara fermentasi keadaan pepejal dan fermentasi tenggelam.	28
2.12	Kandungan kelembapan substrat bagi penghasilan kitinase daripada mikroorganisme berbeza	30
2.13	Saiz inokulum yang berbeza bagi penghasilan kitinase.	32
2.14	Suhu fermentasi yang berbeza bagi penghasilan kitinase	34
2.15	Nilai pH awal yang berbeza bagi penghasilan	

	kitinase.	35
2.16	Sumber nitrogen yang berbeza dalam penghasilan kitinase.	37
2.17	Masa inkubasi yang berbeza bagi penghasilan kitinase.	38
2.18	Penggunaan RSM dalam pelbagai pengoptimuman bagi penghasilan kitinase.	41
4.1	Kesan prarawatan berbeza kepada sisa udang dalam penghasilan kitinase	62
4.2	Ringkasan kandungan mineral dalam kulit udang bagi sebelum dan selepas prarawatan	63
5.1	Reka bentuk eksperimen untuk penyaringan sumber nitrogen bagi penghasilan nitrogen menggunakan sisa udang diprarawatan dengan gelombang mikro sebagai substrat.	82
5.2	Kepekatan sumber nitrogen pada kombinasi berbeza.	82
5.3	Sumber nitrogen terbaik menggunakan reka bentuk faktorial umum bagi penghasilan kitinase menggunakan sisa udang sebagai substrat.	84
6.1	Nilai berkod dan sebenar bagi pemboleh ubah yang digunakan dalam reka bentuk faktorial dua peringkat	92
6.2	Reka bentuk faktorial dua peringkat bagi pemboleh ubah (dengan aras berkod) bagi aktiviti kitinase (U/g IDS) sebagai respon.	95
6.3	Analisis varians (ANOVA) aktiviti kitinase menggunakan reka bentuk faktorial dua peringkat.	96
7.1	Nilai pemboleh ubah berkod dan sebenar bagi pengoptimuman pemboleh ubah dalam penghasilan kitinase daripada sisa udang	

	dalam fermentasi keadaan pepejal.	109
7.2	Matriks reka bentuk komposit pusat pecahan separa 2^3 untuk pengoptimuman penghasilan kitinase daripada sisa udang dalam FKP	110
7.3	Analisi regresi (ANOVA) bagi penghasilan kitinase menggunakan RBKB	112
7.4	Reka bentuk komposit pusat pemboleh ubah (dalam atas berkod) dengan nilai eksperimen dan ramalan aktiviti kitinase.	113
7.5	Perbandingan penghasilan kitinase antara keadaan tidak optimum dan keadaan optimum bagi penghasilan kitinase menggunakan sisa udang di prarawat sebagai substrat.	122
8.1	Penyediaan larutan penimbal yang berbeza.	127
9.1	Kadar perencatan <i>T. virens</i> terhadap <i>G.</i> <i>boninense</i> menggunakan kultur berkembar	148

SENARAI RAJAH

NO. RAJAH	TAJUK	MUKA SURAT
2.1	Mekanisme enzim kitinolitik	7
2.2	Struktur kitin, kitosan dan selulosa	15
2.3	Struktur (a) N-asetilglukosamina dan (b) glukosamina	15
2.4	Ciri-ciri yang mendefinisikan sistem fermentasi keadaan pepejal (FKP)	24
2.5	Skema bagi beberapa proses berskala mikro yang berlaku semasa FKP	25
2.6	Kandul spora <i>Ganoderma boninense</i>	42
2.7	Struktur kimia lignin	43
2.8	Mekanisme degradasi pemisahan lakase bagi model unsur lignin 4,6-dit (t-butil) guaiakol.	44
3.1	Carta aliran reka bentuk eksperimen	51
4.1	Aliran kerja kajian mengenai penghasilan kitinase daripada prarawatan sisa udang oleh <i>Trichoderma virens</i> .	60
4.2	Kandungan mineral pada rawatan kimia Mikrograf SEM bagi prarawatan sisa udang pembesaran x 1,000 dan x 2,000.	62
4.3	Kesan prarawatan gelombang mikro bagi penghasilan kitinase.	65
4.4	Kesan prarawatan ketuhar bagi penghasilan kitinase	66
4.5	Kesan prarawatan pendidihan dan penghancuran bagi penghasilan kitinase.	66

4.6	Kesan prarawatan pengeringan suria bagi penghasilan kitinase.	67
4.7	Kesan sampel kawalan bagi penghasilan kitinase.	67
4.8	Kesan prarawatan kimia bagi penghasilan kitinase.	68
4.9	Kandungan mineral pada sisa udang kawalan	68
4.10	Kandungan mineral pada rawatan pengeringan suria	71
4.11	Kandungan mineral pada rawatan gelombang mikro	71
4.12	Kandungan mineral pada rawatan ketuhar	73
4.13	Kandungan mineral pada rawatan pendidihan dan penghancuran	73
4.14	Kandungan mineral pada sisa udang dengan prarawatan kimia	73
4.15	Gambaran pertumbuhan <i>Trichoderma virens</i> dalam proses fermentasi keadaan pepejal pada substrat sisa udang (pembesaran x40).	76
5.1	Penghasilan kitinase maksimum dengan penambahan sumber nitrogen inorganik dalam plot satu faktor menggunakan analisis statistik perisian <i>Stat-Ease[®] Design Expert</i>	84
6.1	Reka bentuk eksperimen untuk penyaringan faktor-faktor signifikan yang mempengaruhi aktiviti menggunakan <i>T. virens</i> melalui pendekatan reka bentuk faktorial dua peringkat.	90
6.2	Plot kebarangkalian setengah normal bagi kesan masa pengeraman (A), suhu (B), kelembapan substrat (C), pH (D), saiz inokulum (E) dan kepekatan ammonium sulfat (F).	97
6.3	Graf interaksi masa pengeraman – suhu.	99
6.4	Plot kesan utama bagi aktiviti kitinase (U/g	

	IDS).	100
6.5	Plot kebarangkalian normal reja bagi penghasilan kitinase daripada <i>T. virens</i> .	101
7.1	Reka bentuk eksperimen bagi pengoptimuman penghasilan kitinase oleh <i>T. virens</i> menggunakan pendekatan reka bentuk statistik RSM.	105
7.2	Plot respons permukaan aktiviti kitinase daripada persamaan model : kesan masa pengeraman dan suhu.	113
7.3	Plot respons permukaan aktiviti kitinase daripada persamaan model : kesan masa pengeraman dan kelembapan substrat.	115
7.4	Plot respons permukaan aktiviti daripada persamaan model : kesan suhu dan kelembapan substrat.	116
7.5	Kebarangkalian normal reja student bagi penghasilan kitinase daripada sisa udang di prarawat.	119
7.6	Plot reja student melawan respons ramalan.	120
8.1	Metodologi kajian bagi pencirian kitinase mentah yang dihasilkan oleh <i>T. virens</i>	125
8.2	Aktiviti relatif kitinase pada suhu berbeza	132
8.3	Kestabilan haba kitinase mentah pada suhu berbeza.	133
8.4	Aktiviti relatif kitinase mentah pada pH berbeza bagi masa pengeraman selama 60 minit.	134
8.5	Kestabilan bagi kitinase mentah pada pH berbeza.	136
8.6	Analisis SDS-PAGE bagi kitinase mentah yang dihasilkan oleh <i>T. virens</i> .	137
9.1	Metodologi kajian mengenai mekanisme perencatan pertumbuhan <i>Ganoderma boninense</i> oleh <i>Trichoderma virens</i> .	143

9.2	Graf pertumbuhan koloni <i>T. virens</i> dan <i>G. boninense</i> 6 hari selepas diinokulasi.	146
9.3	Koloni <i>T. virens</i> dan <i>G. boninense</i> selepas 6 hari.	147
9.4	Kultur berkembar <i>T. virens</i> sebagai ejen biokawalan dan <i>G. boninense</i> dalam PDA selepas 7 hari masa pegeraman	148
9.5	Turasan kultur , (A) kawalan (+) dengan pentakloronitrobenzena, (B) kitinase mentah, (C) kawalan (-) dengan air suling steril.	150

SENARAI SINGKATAN

(GlcNAc) ₂	-	Diasetilkitobiosa
(NH ₄) ₂ SO ₄	-	Ammonium sulfat
Al	-	Aluminium
ANOVA	-	Analisis varian
ATCC	-	Jenis kultur koleksi Amerika
BCA	-	Agen Biokawalan
BGK	-	Bran gandum komersial
BSA	-	Albumin serum bovin
BSR	-	Pereputan pangkal batang
Ca	-	Kalsium
CaCl ₂ .2H ₂ O	-	Kalsium klorid dehidrat
Cl	-	Klor
Co	-	Kobalt
CoA	-	Koenzim A
Cu	-	Tembaga
DNS	-	Asid dinitrosalisilik
Fe	-	Ferum
FKP	-	Fermentasi keadaan pepejal
FmT	-	Fermentasi tenggelam
GlcNAc	-	N-asetil-glukosamin
HCl	-	Hidroklorik asid
K	-	Kalium
KCl	-	Kalium klorida
KH ₂ PO ₄	-	Kalium dihidrogen fosfat
Mg	-	Magnesium
MgSO ₄ .7H ₂ O	-	Magnesium Sulfat heptahidrat
Mn	-	Mangan

Mo	-	Molibdenum
N ₂	-	Nitrogen
Na	-	Natrium
NAG	-	N-asetil-glukosamin
NaOH	-	Natrium hidroksida
NH ₃	-	Ammonia
NO ₃	-	Nitrat
OFAT	-	Satu faktor dalam satu masa
P	-	Fosfor
PAGE	-	Elektroforesis gel poliakrilamid
PDA	-	Agar dekstroza kentang
PIRG	-	Peratusan perencatan jejari pertumbuhan
RBKP	-	Reka bentuk komposit pusat
RSM	-	Kaedah gerak balas permukaan
S	-	Sulfur
SDS	-	Natrium dodesil sulfat
SEM	-	Mikroskop elektron penskanan
Si	-	Silikon
SKKUP	-	Sisa kitin kulit udang pepejal
T	-	Titanium
TCA	-	Trikloroasetik
Zn	-	Zink

SENARAI SIMBOL

μg	-	Mikrogram
μL	-	Mikroliter
μmol	-	Mikromol
$^{\circ}\text{C}$	-	Darjah selsius
g	-	Gram
g/L	-	Gram per liter
h	-	Hari
i/i	-	Isi padu per isi padu
j/j	-	Jisim per jisim
kDa	-	Kilo dalton
kg	-	Kilogram
min	-	Minit
mL	-	Mililiter
mm	-	Milimeter
mM	-	Milimolar
nm	-	Nanometer
OD	-	Ketumpatan optik
psm	-	Putaran seminit
U/g IDS	-	Unit per gram substrat kering awal

SENARAI LAMPIRAN

LAMPIRAN	TAJUK	MUKA SURAT
A	Penghitungan spora menggunakan haemocytometer	181
B	Kaedah DNS	183
C	Kaedah Lowry	185
D	Penukaran Faktor	187
E	Komposisi Penimbal	189
F	Penyediaan untuk SDS-PAGE	191
G	Prosedur Pewarnaan Perak	193
H	Penerbitan	195

BAB 1

PENGENALAN

1.1 Latar Belakang Kajian

Setiap tahun, Malaysia menghasilkan antara 60-70% sisa udang berdasarkan berat kering daripada pemprosesan makanan bagi eksport makanan laut. Sisa udang kaya dengan kitin iaitu homopolimer N-asetil-D glukosamina (GlcNAc) yang dihubungkan oleh ikatan β -1,4. Kitin terkandung sebanyak 30-40% dalam sisa udang selain dua lagi kandungan utama sisa udang iaitu protein (10-30% (j/j)) dan kalsium karbonat (10-30% (j/j)). Terbitan kitin mempunyai nilai ekonomi yang tinggi berdasarkan aktiviti biologi dan aplikasi agrokimianya (Muzarelli *et al.*, 2012). Walau bagaimanapun, sisa udang sukar diurai secara semulajadi dan menjadi antara penyumbang utama kepada pencemaran alam sekitar.

Kini, penulenan dan pengubahsuaian kitin daripada sisa udang kepada produk karbohidrat bernilai tambah melibatkan rawatan yang menggunakan bahan kimia selain hidrolisis tak terkawal. Cara ini hanya membawa kepada penghasilan produk sampingan yang tidak diinginkan serta kos penulenan yang tinggi untuk menyingkirkan protein dan kalsium karbonat (Chaiharn *et al.*, 2013).

Banyak bakteria dan kulat yang dapat menghasilkan enzim kitinolitik luar sel yang dikenali sebagai kitinase (E.C. 3.2.1.14). Enzim kitinase mampu menukarkan kitin kepada sebatian yang bermanfaat kepada industri seperti glukosamina (Das *et al.*, 2012). Kitinase mempunyai peranan yang penting dalam kawalan biologi perosak

dan penyakit (Kumar *et al.*, 2012). Selain itu, kitinase juga digunakan dalam penyelidikan biologi untuk menghasilkan protoplas kulat bagi menguraikan dinding sel kulat, dan dalam penjagaan kesihatan manusia untuk menghasilkan penyediaan oftalmik (Narayana dan Vijayalakshimi, 2009). Penggunaan kitinase untuk mengawal penyakit pada tumbuhan yang disebabkan oleh pelbagai kulat fitopatogen, serangga, nematod, dan juga penghasilan pelbagai oligomer kitin juga semakin mendapat sambutan (Huang *et al.*, 2005; De la vega *et al.*, 2006; Chang *et al.*, 2007). Namun, kos pengeluaran kitinase yang tinggi telah meningkatkan keperluan untuk mencari strain yang dapat menghasilkan kitinase dalam dengan banyak secara kos efektif (Mabuchi *et al.*, 2000).

Penyelidikan terdahulu telah menunjukkan keupayaan beberapa spesies kulat seperti *Trichoderma* sp. dan *Aspergillus* sp. (Felse dan Panda, 2000; Noppakarn *et al.*, 2002), dan bakteria serta aktinomiset seperti *Bacillus subtilis* (Wang *et al.*, 2006), *B. cereus* (Chang *et al.*, 2007) dan *Streptomyces* (Akagi *et al.*, 2006) dalam menghasilkan kitinase. Keupayaan ini menjadikan sisa udang yang kaya dengan kitin sebagai sumber substrat (sumber karbon) terbaik kepada mikroorganisma-mikroorganisma ini untuk menghasilkan kitinase (Green *et al.*, 2005; Chang *et al.*, 2010), sekaligus membantu dalam menyelesaikan masalah pengurusan sisa selain memberi manfaat komersial (Chaiharn *et al.*, 2013).

Fermentasi keadaan pepejal (FKP) adalah teknologi yang membolehkan pertumbuhan mikroorganisma tanpa kehadiran air bebas (Digankumar *et al.*, 2010). FKP merupakan kaedah penghasilan enzim yang menarik kerana kaedah ini mempunyai kelebihan dari segi penjanaan efluen yang rendah, penggunaan alatan fermentasi yang ringkas, dan produk terhasil boleh terus digunakan. Tambahan pula, kaedah kultur tenggelam sebelum ini lebih digemari untuk penghasilan beberapa enzim industri seperti amilase, selulase, hemiselulase, protease, dan xilanase (Chang *et al.*, 2007).

Pada tahun-tahun kebelakangan ini, penggunaan kaedah FKP semakin menarik minat ramai penyelidik kerana beberapa kajian yang melibatkan enzim (Wang dan Yang, 2007), perasa (Ferron *et al.*, 1996), pewarna (Nimnoi dan Lumyong, 2011),

dan beberapa bahan lain yang penting kepada industri makanan telah menunjukkan yang FKP dapat memberikan hasil yang lebih tinggi (Couto dan Sanroman, 2006) berbanding fermentasi tenggelam (FmT). Tambahan pula, FKP lebih kos efektif kerana menggunakan sisa sebagai sumber (Robinson and Nigam, 2003).

Kajian tentang kos bagi kedua-dua kaedah FKP dan FmT telah dibuat oleh Castilho *et al.* (2000). Dalam kajiannya, Castillo *et al.* melakukan analisis ekonomi yang terperinci terhadap penghasilan lipase oleh *Penicillium restrictum* dan mendapati jumlah pelaburan modal yang diperlukan FmT adalah 78% lebih tinggi daripada FKP untuk skala penghasilan 100 m³ lipase setiap tahun. Kos keseluruhan produk bagi FKP juga adalah 47% lebih rendah daripada harga jualan.

Kajian-kajian ini menunjukkan bahawa kelebihan utama bagi proses FKP ialah penggunaan bahan mentah yang sangat murah sebagai substrat utama. Penggunaan sisa bahan laut bukan sahaja menyelesaikan masalah alam sekitar malah mengurangkan kos penghasilan kitinase oleh mikroba. Oleh itu, FKP sememangnya merupakan kaedah yang berkesan dalam penggunaan sisa pepejal kaya nutrien sebagai substrat. Memandangkan sisa makanan dan agrikultur mengandungi karbohidrat dan nutrien lain yang tinggi sisa-sisa ini boleh digunakan sebagai substrat untuk menghasilkan bahan kimia dan enzim secara pukal menggunakan teknik FKP (Singhania *et al.*, 2009).

Dalam kajian ini, *Trichoderma virens* digunakan sebagai penghasil kitinase dengan menguraikan kitin daripada sisa udang sebagai substrat dalam proses fermentasi keadaan pepejal.

1.2. **Objektif Kajian**

1. Memilih kaedah prarawatan terbaik bagi penghasilan kitinase yang tinggi oleh *Trichoderma virens* menggunakan sisa udang secara fermentasi keadaan pepejal (FKP).
2. Mengkaji kesan sumber nitrogen terhadap penghasilan kitinase oleh *Trichoderma virens* menggunakan reka bentuk faktorial umum (FU) dalam proses FKP.
3. Menyaring faktor-faktor yang mempengaruhi penghasilan kitinase oleh *Trichoderma virens* menggunakan faktorial dua peringkat (FDP) dalam proses FKP.
4. Mengoptimumkan faktor-faktor yang mempengaruhi penghasilan kitinase oleh *Trichoderma virens* menggunakan reka bentuk komposit berpusat (RBKB) dalam proses FKP.
5. Pencirian kitinase mentah yang dihasilkan oleh *Trichoderma virens*.
6. Mengkaji potensi penggunaan kitinase mentah yang dihasilkan oleh *Trichoderma virens* sebagai agen antikulat kepada *Ganoderma boninense*.

1.2 **Skop Kajian**

Skop kajian ini memfokuskan kepada prarawatan sisa udang bagi penghasilan kitinase oleh *Trichoderma virens* secara FKP. Pengaruh fizikal dan faktor persekitaran telah dikenal pasti menggunakan reka bentuk FU dan reka bentuk FDP, manakala pengoptimuman penghasilan kitinase secara FKP telah dijalankan menggunakan RBKP. Pencirian kitinase dilakukan dengan mengkaji keoptimuman dan kestabilan kitinase mentah pada suhu dan pH yang berbeza, dan potensi kitinase mentah dalam merencat *Ganoderma boninense*.

RUJUKAN

- Aam, B.B., Heggset, E.B., Norberg, A.L., Sorlie, M., Varum, K.M. and Eijsink, V.G.H. (2010). Production of chitooligosaccharides and their potential applications in medicine. *Marine Drugs*, 8: 1482-1517.
- Abdul-Ameer, Z. W. (2008). *Production and Characterization of Inulinase Produced By Aspergillus niger using solid state fermentation*. University of Baghdad: Thesis Master Science Biotechnology.
- Afrasa, M., Megersa, N. and Alemu, T. (2014). Characterization of fungal extracts from *Trichoderma* isolates : their effects against coffee wilt pathogen *Gibberella xyloporae*. *Ethiopian Journal of Science* , 2: 82-85.
- Akagi, K, Watanabe, J., Hara, M., Kezuka, Y, Chikaishi, E, Yamaguchi, T, Akutsu, H, Nonaka, T., Watanabe, T. and Ikegami, T. (2006). Identification of the substrate interaction region of the chitin-binding domain of *Streptomyces griseus* chitinase C. *Journal of Biochemistry*, 139: 483-493.
- Al-Ahmadi, K.J., Yazdi, M.T., Najafi, M.F., Shahverdi, A.R. (2008). Optimization of medium and cultivation conditions for chitinase production by newly isolated *Aeromonas* sp. *Biotechnology*, 7 : 266-272.
- Almeida, R.S., Wilson, D. and Hube, B. (2009). *Candida albicans* iron acquisition within the host. *FEMS Yeast Research*, 9 : 1000-1012.
- Andronopoulo, E. and Vorgias, C.E. (2003). Purification and characterization of a new hyperthermostable, allosamidin-insensitive and denaturation-resistant chitinase from the hyperthermophilic archaeon *Thermococcus chitonophagus*. *Extremophiles*, 7: 1-12.
- Annamalai N., Rajeswari M. V., Vijayalakshmi S. and Balasubramanian T. (2011). Purification and characterization of chitinase from *Alcaligenes faecalis* AU02 by utilizing marine wastes and its antioxidant activity. *Annual Microbiology*, 61: 801-807.

- Aoyagi, S, Onishi, H. and Machida, Y. (2007). Novel chitosan wound dressing loaded with minocycline for the treatment of severe burn wounds. *International Journal of Pharmaceutics*, 330 : 138-145.
- Ariffin, D., Idris, A.S. and Singh, G. (2000). *Status of Ganoderma in oil 3 palm. In Ganoderma diseases of penennial crops*. CABI Publishing. 49-52.
- Arst, H.N. and Penalva, M.A. (2003). pH regulation in *Aspergillus* and parallels with higher eukaryotic regulatory systems. *Trends in Genetics*, 19: 224–231.
- Aye, K.N. and Stevens, W. F. (2004). Improved chitin production by pretreatment of shrimp shells. *Chemical Tehnology and Biotechnology*,79: 421-425.
- Azaliza Safarida Binti Wasli (2007). *Prawn waste degradation using crude chitinase produced by Trichoderma virens UKM-1*. Univesiti Teknologi Malaysia: Thesis Master of Science.
- Barbosa, C., Jorge, J.A. and Guimaraes, L.H.S. (2012). Optimization of the chitinase production by different *Metarhizium anisopliae* strains under solid-state fermentation with silkworm chrysalis as substrate using CCRD. *Advance in Microbiology*, 2: 268-276.
- Barbosa, M. A. G., Rehn, K. G., Menezes, M. and Mariano, R. L. R. (2001). Antagonism of *Trichoderma* species on *Cladosporium herbarum* and their enzymatic characterization. *Brazilian Journal of Microbiology*, 32:32-98.
- Bautista-Bañosa, S., Herna'ndez-Lauzardoa, A. N., Vela'zquez-del Vallea, M.G., Herna'ndez-Lo'peza, M., Ait Barkab, E., Bosquez-Molinac and Wilsond, C.L. (2006). Chitosan as a potential natural compound to control pre and postharvest diseases of horticultural commodities. *Crop Protectio*, 25: 108-118.
- Bautista-Bañosa, S., Hernandez-Lopez, M., Bosquez-Molinab, E and Wilson, C.L. (2003). Effects of chitosan and plant extracts on growth of *Colletotrichum gloeosporioides*, anthracnose levels and quality of papaya fruit. *Crop Protection*, 22: 1087-1092.
- Bellaaja, O.G., Hajjia, S., Younesa, I., Chaabounib, M., Nasria, M and Jelloulia, K. (2013). Optimization of chitin extraction from shrimp waste with *Bacillus pumilus* A1 using response surface methodology. *International Journal of Biological Macromolecules*, 61: 243-250.

- Bezerra, M. A., Santelli, R. E., Oliveira, E.P., Villiar, L.S. and Escaleira, L.A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76: 965-977.
- Bezerra, R.P., Matsudo, M.C., Converti, A., Sato, S. and de Carvalho, J.C.M. (2008). Influence of ammonium chloride feeding time and light intensity on the cultivation of *Spirulina (Arthrospira) platensis*. *Biotechnology and Bioengineering*, 100: 297-304.
- Bhushan, B. and Hoondal, G. S. (1998). Isolation, purification and properties of a thermostable chitinase from an alkalophilic *Bacillus* sp. BG-11. *Biotechnology Letters*, 20: 157-159.
- Biesebeke, R., Ruijter, G., Rahardjo, Y.S.P., Hoogschagen, M.J., Heerikhuisen, M., Levin, A., van Driel, K.G.A., Schutyser, M.A.I., Dijksterhuis, J., Zhu, Y., Weber, F.J., de Vos, W.M., van den Hondel, K.M.J.J., Rinzema, A. and Punt, P.J. (2002). *Aspergillus oryzae* in solid-state and submerged fermentations. *FEMS Yeast Research*, 2: 245-248.
- Binod, P., Pusztahelyi, T., Nagy, V., Sandhya, C., Szakacs, G., Istvan P ´ocsi and Pandey, A. (2005). Production and purification of extracellular chitinases from *Penicillium aculeatum* NRRL 2129 under solid-state fermentation. *Enzyme and Microbial Technology*, 36: 880-887.
- Binod, P., Sandhya, C., Suma, P., Szakacs, G. and Pandey, A. (2007). Fungal biosynthesis of endochitinase and chitobiase in solid state fermentation and their application for the production of N-acetyl-D-glucosamine from colloidal chitin. *Bioresource Technology*, 98: 2742-2748.
- Bivi, M. R., Farhana, S. N., Khairulmazmi, A. and Idris, A. (2010). Control of *Ganoderma boninense*: a causal agent of basal stemrot disease in oil palm with endophyte bacteria in vitro. *International Journal of Agriculture and Biology*, 12: 833-839
- Bobelmann, F., Romano, P., Fabritius, H., Raabe, D. and Epple, M. (2007) The composition of the exoskeleton of two Crustacea: The American lobster *Homarus americanus* and the edible crab *Cancer pagurus*. *Thermochim Acta*, 463: 65–68.
- Brandl, F., Sommer, F. and Achim, G. (2007). Rational design of hydrogels for tissue engineering : Impact of physical factors on cell behavior. *Biomaterials*, 28: 134-146.

- Brunner, E., Ehrlich, H., Schupp, P., Hedrich, R., Hunoldt, S., Kammer, M., Machill, S., Paasch, S., Bazhenov, V.V., Kurek, D.V., Arnold, T., Brockmann, S., Ruhnow, M. and Born, R. (2009). Chitin-based scaffolds are an integral part of the skeleton of the marine demosponge *Ianthella basta*. *Journal Structure Biology*, 168: 539-547.
- Buchholz, K., Kasche, V. and Bornscheuer, U.T. (2005). *Biocatalysts and enzyme technology*. Weinheim, Wiley-VCH.
- Calabrese, V., Lodi, R., Tonon, C., D'Agata, V., Sapienza, M., Scapagnini, G., Mangiameli, A., Pennisi, G., Giuffrida, S. A. M. and Butterfield, D. A (2005). Oxidative stress, mitochondrial dysfunction and cellular stress response in Friedreich's ataxia. *Journal of the Neurological Sciences*, 233: 145-162.
- Castilho, L.R., Mendronho, R. A. and Alves, T. L. M. (2000). Production and extraction of pectinases obtained by solid state fermentation of agroindustrial residues with *Aspergillus niger*. *Bioresouces Technology*, 71: 45-50.
- Castillo, F.D.H., Padilla, A.M.B., Morales, G.G., Siller, M.C., Herrera, R.R., Gonzales, C.N.A. and Reyes, F.C. (2011). In vitro antagonist action of *Trichoderma* strains against *Sclerotinia sclerotiorum* and *Sclerotium cepivorum*. *American Journal of Agricultural and Biological Sciences*, 6: 410-417.
- Chaiarn, M., Lumyong, S., Hasan, N. and Plikomol, A. (2013). Solid-state cultivation of *Bacillus thuringiensis* R 176 with shrimp shells and rice straw as a substrate for chitinase production. *Annual Microbiology*, 63: 443-450.
- Chang, W. T., Chen, C. S. and Wang, S. L. (2003). An antifungal chitinase produced by *Bacillus cereus* with shrimp and crab shell powder as a carbon source. *Current Microbiology*, 47: 102-108.
- Chang, W. T., Chen, M. L. and Wang, S. L. (2010). An antifungal chitinase produced by *Bacillus subtilis* using chitin waste as a carbon source. *World journal Microbiology and Biotechnology*, 26: 945-960.
- Chang, W.T., Chen, Y. and Jao, C.L. (2007). Antifungal activity and enhancement of plant growth by *Bacillus cereus* grown on shellfish chitin wastes. *Bioresouces Technology*, 98: 1224-1230.
- Chang, Y. N., Huang, J.C., Lee, C.C., Shih, I.L. and Tzeng, Y.M. (2002). Use of response surface methodology to optimize culture medium for production of

- lovastatin by *Monascus ruber*. *Enzyme Microbiology and Technology*, 30: 889-894.
- Cheison, S. C., Schmitt, M., Leeb, E., Letzel, T. and Kulozik, U. (2010). Influence of temperature and degree of hydrolysis on the peptide composition of trypsin hydrolysates of β -lactoglobulin analysis by LC-MS. *Food Chemistry*, 121: 457-467.
- Chmielowski, R.A., Wu, H. S. and Wang, S. S. (2007). Scale-up of upstream and downstream operations for the production of glucosamine using microbial fermentation. *Biotechnology Journal*, 2: 996-1006.
- Chong, K.P., Rossall, S. and Markus, A. (2009). In vitro synergy effect of syringic acid, caffeic acid and 4-hydroxybenzoic acid against *Ganoderma boninense*. *International Journal of Engineering and Technology*, 1: 282-284.
- Christians, N. (2011). *Fundamentals of turfgrass management*. 4th edition. New York : John Wiley and Sons Publishers.
- Chupp, G. L., Lee, C.G., Jarjour, N., Shim, Y.M. and Holm, C.T. (2007). A chitinase like protein in the lung and circulation of patients with severe asthma. *New England Medical Journal*, 357: 2016-2027.
- Contreras-Cornejo, H.A., Macias-Rodriguez, L., Cortes-Penagos, C. and Lopez-Bucio, J. (2009). *Trichoderma virens*, a plant beneficial fungus, enhances biomass production and promotes lateral root growth through an auxin-dependent mechanism in arabidopsis. *Plant Physiology*, 149 : 1579-1592.
- Couto, S. R. (2008). Exploitation of biological wastes for the production of value-added products under solid-state fermentation conditions. *Biotechnology Journal*, 3: 859-870.
- Couto, S.R. and Sanroman, M.A. (2006). Application of solid-state fermentation to food industry – A Review. *Journal of Food Engineering*, 76: 291-302.
- Crespo, M. O. P, Martinez, M.V., Hernandez, J.L. and Yusty, M.A.L. (2006). High-performance liquid chromatographic determination of chitin in the snow crab. *Chionoecetes opilio*. *Journal of Chromatography A*, 1116 : 189-192.
- Cristian, F. A., Nathalia, K.A., Maria, G.B.P., Marcia, R.S.P., Gorete, R.M. and Everaldo, S.S. (2010). Chitooligosaccharides enzymatic production by *Metarhizium anisopliae*. *Bioprocess Biosystems Engineering*, 33: 893-899.

- Cruz, J., Pintor-Toro, J.A., Benitez, T., Liobell, A. and Romero, L.C. (1995). A novel endo-beta-1,3-glucanase, BGN13.1, involved in the mycoparasitism of *Trichoderma harzianum*. *Journal Bacteriology*, 177: 6937–6945.
- Dahiya, N., Rupinder, T. and Gurinder, S. H. (2006). Biotechnological aspects of chitinolytic enzymes : a review. *Applied Microbiology and Biotechnology*, 71: 773-782.
- Dahiya, N., Rupinder, T., Ram Prakash, T. and Gurinder Singh, H. (2005). Chitinase production in solid-state fermentation by *Enterobacter* sp. NRG4 using statistical experimental design. *Current Microbiology*, 51: 222-228.
- Daniel, M.B., Michael, D.R. and Stuart, J.E. (1996). *Protein methods*. 2nd Edition. New York : John Wiley and Sons, Inc., Publication.
- Das, S. N., Neeraja, C., Sarma, P., Prakash, J.M, Purushotham, P., Kaur, M., Dutta, S and Podile, A. R. (2012). Microbial chitinases for chitin waste management. *Microorganism in Environmental Management : Microbes and Environment*, 6: 135-145.
- de la Vega, L.M., Barboza-Corona, J. E., Aguilar-Uscanga, M.G., Ramírez-Lepe, M. (2006). Purification and characterization of an exochitinase from *Bacillus thuringiensis* subsp. aizawai and its action against phytopathogenic fungi. *Canadian Journal of Microbiology*, 52: 651-657.
- De Smedt, S.C., Demeester, J. and Hennink, W. E. (2000). Cationic polymer based gene delivery systems. *Pharmaceutical Resources*, 17: 113-126.
- Demirel, M. and Kayan, B. (2012). Application of response surface methodology and central composite design for the optimization of textile dye degradation by wet air oxidation. *International Journal of Industrial Chemistry*, 3: 1-10.
- Digantkumar, C., Jyoti, D., Datta, M. and Amita, S. (2010). Utilization of agro-industrial waste for xylanase production by *Aspergillus foetidus* MTCC 4898 under solid state fermentation and its application in saccharification. *Biochemical Engineering Journal*, 49: 361-369.
- Dutta, P.K. (2005), *Chitin and Chitosan : Opportunities and Challenges*. Midnapore, India : SSM International Publication.
- Dutta, P.K., Tripathi, S., Mehrotra, G.K. and Dutta, J. (2009). Perspectives for chitosan based antimicrobial films in food applications. *Food Chemistry*, 114: 1173-1182.

- Elad, Y. and Kapat, A. (1999). The role of *Trichoderma harzianum* protease in the biocontrol of *Botrytis cinerea*. *European Journal of Plant Pathology*, 105: 177-189.
- El-Dein, A., Hosny, M.S, El-Shayeb, N.A., Abood, A. and Abdel-Fattah, A.M. (2010). A potent chitinolytic activity of marine *Actinomyces* sp. and enzymatic production of chitooligosaccharides. *Australian Journal of Basic and Applied Sciences*, 4: 615-623.
- El-Katatny, M.H., Gudelj, M., Robra, K. H., Elnaghy, M. A. and Gubitza, G. M. (2001). Characterization of a chitinase and an endo- β -1,3-glucanase from *Trichoderma harzianum* Rifai T24 involved in control of the phytopathogen *Sclerotium rolfsii*. *Applied Microbiology and Biotechnology*, 56: 137-143.
- Emani, C., Garcia, J.M., Finch, E.L., Pozo, M.J., Uribe, P., Kim, D.J., Sunilkumar, G., Cook, D.R., Kenerley, C.M. and Rathore, K.S. (2003). Enhanced fungal resistance in transgenic cotton expressing an endochitinase gene from *Trichoderma virens*. *Plant Biotechnology Journal*, 1: 321-336.
- Fang, H., Zhao, C. and Song, X.Y (2010). Optimization of enzymatic hydrolysis of steam exploded corn stover by two approaches: response surface methodology or using cellulase from mixed cultures of *Trichoderma reesei* RUT-C30 and *Aspergillus niger* NL02. *Bioresource Technology*, 101: 4111–4119.
- Felse P. A. and Panda T. (2000). Submerge culture production of chitinase by *Trichoderma harzianum* in stirred tank bioreactors - the influences by agitation speed. *Biochemical Engineering*, 4: 115-120.
- Felse, P.A. and Panda, T. (2000). Production of microbial chitinases - a revisit. *Bioprocess Engineering*, 23: 127-134.
- Ferreira, S., Duarte, A.P., Ribeiro, M.H.L., Queiroz, J.A. and Domingues, F.C. (2009). Response surface optimization of enzymatic hydrolysis of *Cistus ladanifer* and *Cytisus striatus* for bioethanol production. *Biochemical Engineering Journal*, 45: 192-200.
- Ferron, G., Bonnarne, P. and Durand, A. (1996). Prospect for the microbial production of food flavours. *Trends in Food Science and Technology*, 7: 285-293.
- Franco, L. O., De Cassia, R., Portoll, A. L. F., Messias, A. S., Fukushima, K. and De Campos, G. M. (2004). Heavy metal biosorption by chitin and chitosan

- isolated from *Cunninghamella elegans* (IFM 46109). *Brazilian Journal of Microbiology*, 35: 247-247.
- Gajera, H., Domadiya, R., Patel, S., Kapopara, M. and Golakiya, B. (2013). Molecular mechanism of *Trichoderma* as bio-control agents against phytopathogen system – a review. *Current Research in Microbiology and Biotechnology*, 1: 133-142
- Galbe, M. and Zacchi, G.. (2002). A review of the production of ethanol from softwood. *Applied Microbiology and Biotechnology*, 59: 618-628.
- Gallaher, C.M., Munion, J., Hesslink, R., Wise, J. and Gallaher, D.D. (2000). Cholesterol reduction by glucomannan and chitosan is mediated by changes in cholesterol absorption and bile acid and fat excretion in rats. *Journal of Nutrition*, 130: 2753-2759.
- Gan, Z., Yang, J., Tao, N., Yu, Z. and Zhang, K. (2007). Cloning and expression analysis of a chitinase gene *crchi1* from the mycoparasitic fungus *Clonostachys rosea* (syn. *Gliocladium roseum*). *The Journal of Microbiology*, 45: 422-430.
- Geisseler, D., William, R. H., Rainer, G. J. and Bernard, L. (2010). Pathways of nitrogen utilization by soil microorganisms - A review. *Soil Biology and Biochemistry*, 42 : 2058 - 2057.
- Gemma, E. S., Veronica, M.H. and David, J.A (1998). Inducible chitinolytic system of *Aspergillus fumigatus*. *Microbiology*, 144 : 1575-1581.
- George, A. and Roberts, F. (1992). *Chitin Chemistry*. London. Macmilland Press
- Ghanem, K. M., Fahad, A., Al-Fassi and Reem, M. F. (2011). Statistical optimization of cultural conditions for chitinase production from shrimp shellfish waste by *Alternaria alternata*. *African Journal Microbiology*, 5 : 1649-1659.
- Ghanem, K. M, Al-Ghani, S. M. and Al-Makisha, N. H. (2010). Statistical optimization of cultural conditions for chitinase production from fish scales waste by *Aspergillus terreus*. *African Journal Biotechnology*, 9: 5135-5146.
- Ghufran, M., Kordi, H.K, and Tancung A.B. 2007. *Pengelolaan kualitas air dalam budidaya perairan*. Jakarta. RinekaCipta.
- Gkargkas, K., Mamma, D., Nedev, D., Topakas, E., Christakopoulos, P., Kekos, D., Macris, B.J. (2004). Studies on a N-acetyl- β -d-glucosaminidase produced by *Fusarium oxysporum* F3 grown in solid-state fermentation. *Process Biochemistry*, 39 : 1599-1605.

- Gohel, V., Singh, A., Vimal, M., Ashwini, P. and Chhatpar, H. S. (2006). Bioprospecting and antifungal potential of chitinolytic microorganisms. *African Journal Biotechnology*, 5 : 54-72.
- Gokul, B., Lee, J.H, Song, K.B., Rhee, S.K., Kim, C.H. and Panda, T. (2000). Characterization and applications of chitinases from *Trichoderma harzianum* - A review. *Bioprocess Engineering*, 23: 691-694.
- Gonvalces, A. R. and Schuchardt, U. (2002). Hydrogenolysis lignin. *Applied Biochemistry and Biotechnology*, 98: 1213-1219.
- Gottipati, R. and Mishra, S. (2010). Process optimization of adsorption of Cr(VI) on activated carbons prepared from plant precursors by a two-level full factorial design. *Chemical Engineering Journal*, 160 : 99-107.
- Gowthaman, M.K., Krishna, C. and Moo.-Young, M. (2001). Fungal solid state fermentation- an overview. *Applied Mycology and Biotechnology*, 1: 305-352
- Green, A.T., Healy, M. and Healy, A. (2005). Production of chitinolytic enzymes by *Serratia marcescens* QMB1466 using various chitinous substrates. *Journal of Chemical Technology and Biotechnology*, 80 : 28-34.
- Guerarda, F., Sumaya-Martinez, M.T., Laroque, D., Chabeauda, A. and Dufosse, L. (2007). Optimization of free radical scavenging activity by response surface methodology in the hydrolysis of shrimp processing discards. *Process Biochemistry*, 42 : 1486-1491.
- Guo, S. H., Chen, J. K., and Lee, W. C. (2004). Purification and characterization of extracellular chitinase from *Aeromonas schubertii*. *Enzyme and Microbial Technology*, 35: 550-556.
- Hamid, R., Khan, M. A., Ahmad, M., Ahmad, M.M., Abdin, M.Z., Musarrat, J., and Javed, S. (2013). Chitinases : An update. *Journal Pharmacy Bioallied Science*, 5: 21-29.
- Han, Y., Li, Z., Mio, X. and Zhang, F. (2008). Statistical optimization of medium components to improve the chitinase activity of *Streptomyces* sp. Da11 associated with the South China Sea sponge *Craniella australiensis*. *Process Biochemistry*, 43:1088-1093.
- Haran, S., Schickler, H., Oppenheim, A. and Chet, I. (1995). New component of chitinolytic system of *Trichoderma harzianum*. *Mycology Resource*, 99: 441-446.

- Harman, G.E, Howell, C.R., Viterbo, A., Chet, I. and Lorito, M. (2004). *Trichoderma* species - opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, 2: 43-56.
- Hasan, S., Gupta, G., Shreya, A. and Kaur, H. (2014). Lytic enzymes of *Trichoderma*: their role in plant defense. *International Journal of Applied Research and Studies (IJARS)*, 3: 1-5.
- Holker, U. and Lenz, J. (2005). Solid-state fermentation — are there any biotechnological advantages? *Current Opinion in Microbiology*, 8: 301-306.
- Horn, S.J., Sorlie, M., Vaaje-Kolstad, G., Norberg, A.L., Synstad, B., Vårum, K.M. and Eijsink, V.G.H. (2006). Comparative studies of chitinases A, B and C from *Serratia marcescens*. *FEBS Journal*, 24 : 39-53.
- Howard, M.B. (2004). *Complex Polysaccharide degradation by Microbulbifer degradans strain 2-40: Studies of the chitinolytic system and carbohydrate architecture*. University of Maryland: Thesis Doctor Philosophy
- Howell, C. R. (2006). *Understanding the mechanisms employed by Trichoderma virens to effect biological control of cotton diseases*. Symposium The Nature and Application of Biocontrol Microbes II : *Trichoderma* spp. August 3, 2004. Presented at the Annual Meeting of The American Phytopathological Society. 174-180.
- Howell, C. R (2003). Mechanisms employed by *Trichoderma* species in the biological control of plant diseases : the history and evolution of current concepts. *Plant disease*, 87: 4-10.
- Huang, C. J., Wang, T. K., Chung, S.C. and Chen, C.Y. (2005). Identification of an antifungal chitinase from a potential biocontrol agent *Bacillus cereus*. *Journal Biochemical Molecular and Biology Science*, 38: 82-88.
- Hushiarian, R., Yusof, N.A. and Dutse, S.W. (2013). Detection and control of *Ganoderma boninense*: strategies and perspectives. *Springer Plus*, 2: 1-12.
- Inbar, J. and Chet, I. (1995). The role of recognition in the induction of specific chitinases during mycoparasitism by *Trichoderma harzianum*. *Microbiology*, 141: 2823-2829.
- Ishida, H., Hata, Y., Kawato, A., Abe, Y., Suginami, K. and Imayasu, S. (2000). Identification of functional elements that regulate the glucoamylase-encoding gene (glaB) expressed in solid-state culture of *Aspergillus oryzae*. *Current Genetic*, 37: 373-379.

- Ishihara, M., Fujita, M., Obara, K., Hattori, H., Nakamura, S., Nambu, M., Kiyosawa, T., Kanatani, Y., Takase, B., Kikuchi, M. and Maehara, T. (2006). Controlled releases of FGF-2 and paclitaxel from chitosan hydrogels and their subsequent effects on wound repair, angiogenesis, and tumor growth. *Current Drug Delivery*, 3: 351-358.
- Jami Al Ahmadi, K., Tabatabaei Yazdi, M., Fathi Najafi, M., Shahverdi, A.R., Faramarzi, M.A., Zarrini, G. and Behravan, J. (2008). Optimization of medium and cultivation conditions for chitinase production by the newly isolated: *Aeromonas* sp. *Biotechnology*, 2: 266-272.
- Jiang, Y. and Li, Y. (2001). Effects of chitosan coating on postharvest life and quality of longan fruit. *Food Chemistry*, 73: 139-149.
- Jinantana, J. and M. Sariah, 1997. Antagonistic effect of Malaysian isolates of *Trichoderma harzianum* and *Gliocladium viren* on *Sclerotium rolfsii*. *Pertanika: Journal Tropical Agricultural Science*, 20: 35-41
- John, R.P., Tyagi, R. D., Prevost, D., Brar, S. K., Pouleur, S. and Surampalli, R.Y. (2010). Mycoparasitic *Trichoderma viride* as a biocontrol agent against *Fusarium oxysporum* f. sp. *adzuki* and *Pythium arrhenomanes* and as a growth promoter of soybean. *Crop Protection*, 29 : 1452-1459.
- Jubina, P. A., Girija, V. K. (1998): Antagonistic rhizobacteria for management of *Phytophthora capsici*, the incitant of foot rot of black pepper. *Journal Mycology of Plant Pathology*, 28 : 147-153.
- Kamal, K.P. and Brian, M.G. (2006). Biological control of plant pathogens. *The Plant Health Instructor*, 1:1-20.
- Kapat, A., Rakshit, S. and Pandey, T. (1996). Optimization of carbon and nitrogen sources and the environmental conditions for the production of chitinase using *Trichoderma harzianum*. *Bioprocess Engineering*, 15: 13-20.
- Khoushab, F. and Yamabhai, M. (2010). Chitin research revisited. Review. *Marine Drugs*, 8: 1988-2012.
- Kim, H.S., Timmis, K.N. and Golyshin, P.N. (2007). Characterization of a chitinolytic enzyme from *Serratia* sp. KCK isolated from kimchi juice. *Applied Microbiology and Biotechnology*, 75: 1275-1283.
- Kim, J.S and Je, Y. H. (2010). A novel biopesticide production: attagel-mediated precipitation of chitinase from *Beauveria bassiana* SFB-205 supernatant for thermotolerance. *Applied Microbiology and Biotechnology*, 87: 1639-1648.

- Kim, Y.C., Jung H., Kim, K.Y. and Park, S.K. (2008). An effective biocontrol bioformulation against Phytophthora blight of pepper using growth mixtures of combined chitinolytic bacteria under different field conditions. *European Journal of Plant Pathology*, 120 : 373-382.
- Knorr, D. 1982. Functional properties of chitin and chitosan. *Journal Food Science*, 47:593-595.
- Kong, Q., Chen, F., Wang, X., Li, J., Guan, B. and Lou, X (2011). Optimization of conditions for enzymatic production of ACE inhibitory peptides from collagen. *Food Bioprocess Technology*, 4: 1205-1211.
- Kubicek, C.P., Marc, R. L., Peterbauer, C.K. and Lorito, M. (2001). *Trichoderma*: from genes to biocontrol. *Journal of Plant Pathology*, 83: 11-23.
- Kumar, D. P., Singh, R. K., Anupama, P.D., Solanki, M. K., Kumar, S., Srivastava, A. K., Singhal, P. K. and Arora, A. K. (2012). Studies on exo-chitinase production from *Trichoderma asperellum* UTP-16 and its characterization. *Indian Journal Microbiology*, 52 : 388-395.
- Kumar, K., Amaresan, N., Bhagat, S., Madhuri, K. and Srivastava, R. C. (2012). Isolation and characterization of *Trichoderma* spp. for antagonistic activity against root rot and foliar pathogens. *Indian Journal Microbiology*, 52: 137-144.
- Kumirska, J., Czerwicka, M., Kaczynski, Z., Bychowska, A., Brzozowski, K., Thöming, J. and Stepnowski, P. (2010). Application of spectroscopic methods for structural analysis of chitin and chitosan. *Marine Drugs*, 8 : 1567-1636.
- Kurita, K. (2008). Chitin and Chitosan : Functional biopolymers from marine crustaceans. *Marine Biotechnology*, 8: 203-226.
- Lee, H., Song, M. and Hwang, S. (2003). Optimizing bioconversion of deproteinated cheese whey to mycelia of *Ganoderma lucidum*. *Process Biochemistry*, 38: 1685-1693.
- Lima, L. H., Ulhoa, C.J., Fernandes, A.P. and Felix, C.R (1997). Purification of a chitinase from *Trichoderma* sp. and its action on *Sclerotium rolfsii* and *Rhizoctonia solani* cell wall. *Folia Microbiologica*, 44 : 45-49.
- Liu, B.L., Kao, P.M., Tzeng, Y. M. and Feng, K.C. (2003). Production of chitinase from *Verticillium lecanii* F091 using submerged fermentation. *Enzyme and Microbial Technology*, 33: 410-415.

- Liu, M., Cai, Q. X., Liu, H.Z., Zhang, B.H., Yan, J.P. and Yuan, Z.M. (2002). Chitinolytic activities in *Bacillus thuringiensis* and their synergistic effects on larvicidal activity. *Journal of Applied Microbiology*, 93: 374-379.
- Lowry, O.H., Rousenbrought, N.J., Farr, A.L. and Randall, J.R. (1951). Protein measurements with the folin reagent. *Journal Biology and Chemistry*, 193: 265-275.
- Lumsden, R. D., Locke, J. C., Adkins, S. T., Walter, J. F., and Ridout, C. J. 1992. Isolation and localization of the antibiotic gliotoxin produced by *Gliocladium virens* from alginateprill in soil and soilless media. *Phytopathology*, 82 : 230-235.
- Mabuchi, N., Hashizumi, I. and Araki, Y (2000). Characterization of chitinases excreted by *Bacillus cereus* CH. *Canadian Journal of Microbiology*, 46 : 370-375.
- Madigan, M. T., Martinko, J. M. and Parker, J. (2000). *Brock Biology of Microorganisms*. New York, Prentice Hall. 324-360
- Manghsoudi, V. and Yaghmaei, S. (2010). Comparison of solid substrate and submerged fermentation for chitosan production by *Aspergillus niger*. *Transaction C. Chemistry and Chemical Engineering*, 17 : 153-157.
- Matsumoto, K. S. (2006). *Fungal chitinases*. book : *advances in agricultural and food biotechnology*, Trivandrum, India.
- Matsumoto, Y., Gerardo, S.C., Sergio, R. and Keiko, S (2004). Production of β -N-acetylhexosaminidase of *Verticillium lecanii* by solid state and submerged fermentation utilizing shrimp waste silage as substrate and inducer. *Process Biochemistry*, 39 : 665-671.
- Mazutii, M., Bender, J. P., Treichel, H. and Di Luccio, M. (2006). Optimization of inulinase production by solid state fermentation using sugarcane bagasse as substrate. *Journal Biotechnology*, 12 : 1123-1131.
- Meija-Saules, J.E., Waliszewski, K. N., Garcia, M.A. and Cruz-Camarillo, R. (2006). The use of crude shrimp shell powder for chitinase production by *Serratia marcescens* WF. *Food Technology and Biotechnology*, 44 : 95-100.
- Metcalf, T. L., Dillon, P. J. and Metcalfe, C. D. (2008). Effects of formulations of the fungicide, pentachloronitrobenzene on early life stage development of the Japanese medaka (*Oryzias latipes*). *Chemosphere*, 71: 1957-1962.

- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, 31: 426-428.
- Mitchel, D. A., Krieger, N., Stuart, D.M. and Pandey, A. (2000). New development in solid-state fermentation. II. rational approaches to the design, operation and scale-up of bioreactors. *Process Biochemistry*, 35: 1211-1225.
- Mitchell, D.A., Berovic, M. and Krieger, N. (2006). *Bioreactors: Fundamentals of Design and Operation*. London, Spriger. 30-59
- Monteiro, V. N. and Ulhoa, C. J. (2006). Biochemical characterization of a b-1,3-glucanase from *Trichoderma koningii* induced by cell wall of *Rhizoctonia solani*. *Current Microbiology*, 52: 92-96.
- Montgomery, D. C. (1997). *Design and analysis of experiments*. 4th edition. New York: John Wiley and Sons.97-120
- Montgomery, D. C. (2003). *Design and analysis of experiments*. 5th edition. New York: John Wiley and Sons. 122-135
- Mrudula, S. and Anitharaj, R. (2011). Pectinase production in solid state fermentation by *Aspergillus niger* using orange peel as substrate. *Global Journal of Biotechnology and Biochemistry*, 6: 64-71
- Mulatu, A. (2010). *Characterization and testing of antifungal extracts from Trichoderma Isolates against Fusarium xylarioides, the causative agent of coffee wilt disease. Microbial, Cellular and Molecular Biology*.Addis Abab University: Thesis Doctoral Philosophy
- Muzzarelli, R. A. A. (2010). Chitins and chitosans as immunoadjuvants and non-allergenic drug carriers. *Marine Drugs*, 8 : 292-312.
- Muzzarelli, R. A. A. (2011). Potential of chitin/chitosan-bearing materials for uranium recovery : An interdisciplinary review. *Carbohydrate Polymers*, 84: 54-63.
- Muzzarelli, R. A. A. and Muzarelli C. (2006). *Chitosan, a dietary supplement and a food technology commodity. Functional food carbohydrates*. Francis and Taylor. 80-120
- Muzzarelli, R. A. A., Boudrant, J., Meyer, D., Manno, N., DeMarchis, M. and Paoletti, M.G. (2012). Current views on fungal chitin/chitosan, human chitinases, food preservation, glucans, pectins and inulin: A tribute to Henri Braconnot, precursor of the carbohydrate polymers science, on the chitin bicentennial. *Carbohydrate Polymers*, 87: 995-1012.

- Nam, T., Park, S., Lee, S.Y., Park, K., Choi, K., Song, I.C., Han, M.H., Leary, J.J., Yuk, S.A., Kwon, I.C., Kim, K. and Jeong, S.Y. (2010). Tumor targeting chitosan nanoparticles for dual-modality optical/MR cancer imaging. *Bioconjugate Chemistry*, 21 : 578-582.
- Nampoothiri, K. M., Baiju, T. V., Sandhya, C, Sabu, A., Szakacs, G., Pandey, A. (2004). Process optimization for antifungal chitinase production by *Trichoderma harzianum*. *Process Biochemistry*, 39 : 1583-1590.
- Narayana, K. J. P. and Vijayalaksimi, M. (2009). Chitinase production by *Streptomyces* sp. ANU 6277. *Brazilian Journal Microbiology*, 4: 36-40.
- Narayanan, K., Chopade, N., Raj, P.V., Subrahmanyam, V.M. dan Rao, J. V. (2013). Fungal chitinase production and its application in biowaste management. *Journal of Science and Industrial Research*, 72: 393-399.
- Nawani, N. N. and Kapadnis B. P. (2005). Optimization of chitinase production using statistics based experimental designs. *Process Biochemistry*, 40: 651-660.
- Nawani, N. N. and Kapadnis, B. P. (2004). Production dynamics and characterization of chitinolytic system of *Streptomyces* sp. NK1057, a well equipped chitin degrader. *World Journal of Microbiology and Biotechnology*, 20: 487-494.
- Nelson, D. L. and Cox, M. M. (2000). *Lehninger, Principles of Biochemistry*. 3rd edition. New York : Worth Publishing. 230-320
- Ngo, D.N., Lee, S. H., Kim, M.M. and Kim, S.K. (2009). Production of chitin oligosaccharides with different molecular weights and their antioxidant effect in RAW 264.7 cells. *Journal of Functional Foods*, 1: 188-198.
- Nimnoi, P. and Lumyong, S. (2011). Improving solid state fermentation of *Monascus purpureus* on agricultural product for pigment production. *Food and Bioprocess Technology*, 4: 1384-1390.
- Nopakarn, R., Plikomol, A., Yano, S., Wakayama, M. and Tachiki, T. (2002). Utilization of shrimp shellfish waste a substrate for solid state cultivation of *Aspergillus* sp.S1-13: evaluation of a culture based on chitinase formation which is necessary for chitin assimilation. *Journal Bioscience and Bioengineering*, 93: 550-556.
- Nurrazean, H. T., Madihah M. S., and Suraini Abd-Aziz (2011). Screening of factors influencing exopolymer production by *Bacillus licheniformis* strain t221a using

- 2-level factorial design. *Progress in Molecular and Environmental Bioengineering*,1: 395-404.
- Oh, Y. S., Shih, I. L., Tzeng, Y.M. and Wang, S. L. (2000). Protease produced by *Pseudomonas aeruginosa* K-187 and its application in the deproteinization of shrimp and crab shell wastes. *Enzyme and Microbial and Technology*,21: 3-10.
- Okamoto, Y., Yano, R., Miyatake, K.,Tomohiro, I., Shigemasa, Y., Minami, S. (2003). Effects of chitin and chitosan on blood coagulation. *Carbohydrate Polymers*, 53: 337–342.
- Okigbo, R.N. and Ikediugwu, F. E. O. (2000). Studies on biological control of postharvest rot in yams (*Dioscorea* spp) using *Trichoderma viride*. *Journal Phytopathology*, 148: 351-355.
- Ouattara, B., Simard, R.E., Piette, G., Begin, A. and Holley R.A. (2000). Inhibition of surface spoilage bacteria in processed meats by application of antimicrobial films prepared with chitosan. *International Journal of Food Microbiology*,62: 139-148.
- Ozogul, Y. (2000). The possibility of using crustacean wastes products (CWP) on Rainbow trout (*Oncorhynchus mykiss*) feeding. *Turki Journal Biology*, 24: 845-854.
- Pal, K. K. and Gardener, B.M.S. (2006). Biological Control of Plant Pathogens *The Plant Health Instructor*, 1: 1 – 25.
- Pandey, A. (2003). Solid-State fermentation. *Biochemical Engineering Journal*, 13: 81-84.
- Pandey, A., Ashakumari, L., Selvakumar, P. and Vijayalakshmi, K.S (1994). Influence of water activity on growth and activity of *Aspergillus niger* for glucoamylase production in solid state fermentation. *World Journal Microbiology and Biotechnology*, 10 : 458-486.
- Paoletti, M. G., Lorenzo, N., Roberta, D. and Salvatore, M (2007). Human gastric juice contains chitinases that can degrade chitin. *Annual Nutrition Metabolism*, 51: 244-251.
- Park, I. K., Kim, T. H., Park, Y. H., Shin, B.A., Choi, E.S, Chowdhury, E.H., Akaike, T. and Cho, C.S. (2001). Galactosylated chitosan-graft-poly (ethylene glycol) as hepatocyte-targeting DNA carrier. *Journal of Controlled Release*, 76: 349-362.

- Paterson, R.R.M. (2007). *Ganoderma* disease of oil palm—A white rot perspective necessary for integrated control. *Crop Protection*, 26 : 1369-1376.
- Patidar, P., Agrawal, D., Banerjee, T. and Patil, S. (2005). Optimisation of process parameters for chitinase production by soil isolates of *Penicillium chrysogenum* under solid substrate fermentation. *Process Biochemistry*, 40: 2962-2967.
- Patil, N. S., Shailesh, R. W. and Jyoti, P. J. (2005). Purification and characterization of an extracellular antifungal chitinase from *Penicillium ochrochloron* MTCC 517 and its application in protoplast formation. *Process Biochemistry*, 48 : 176-183.
- Patil, R.S., Ghormade, V. and Desphande, M. V. (2000). Chitinolytic enzymes: an exploration. *Enzyme and Microbial Technology*, 26: 473-483
- Pitson, S. M., Seviou, R. J. and Mc Dougall, B.M. (1993). Noncellulolytic fungal β -glucanases: their physiology and regulation. *Enzyme and Microbial and Technology*, 15: 178-192.
- Qazi, S. S. and Khachatourisans, G.G. (2008). Addition of exogenous carbon and nitrogen sources to aphid exuviae modulates synthesis of proteases and chitinase by germinating conidia of *Beauveria bassiana*. *Archives Microbiology*, 189: 589-596.
- Qi, B., Chen, X., Shen, F., Su, Y. and Wan, Y. (2009). Optimization of enzymatic hydrolysis of wheat straw pretreated by alkaline peroxide using response surface methodology. *Industrial and Engineering Chemical Resource*, 48: 7346-7353.
- Raabe, D., Romano, P., Sachs, C., Al-Sawalmiha, A., Brokmeier, H.G., Yib, S.B., Servos, G. and Hartwig, H.G. (2005). Discovery of a honey comb structure in the twisted plywood patterns of fibrous biological nano composite tissue. *Journal of Crystal Growth*, 283 : 1-7.
- Raabe, D., Romano, P., Sachs, C., Fabritius, H., Al-Sawalmih, A., Yi, S.B., Servos, G. and Hartwig, H.G. (2006). Microstructure and crystallographic texture of the chitin-protein network in the biological composite material of the exoskeleton of the lobster *Homarus americanus*. *Material Science and Engineering*, 421: 143-153.
- Radjacommar, R., Venkatesan, S. and Samiyappan, R. (2010). Biological control of phytopathogenic fungi of vanilla through lytic action of *Trichoderma* sp. and

- Pseudomonas fluorescens*. *Archives of Phytopathology and Plant Protection*,43: 1-17.
- Rajulu, M. B. G., Thirunavukkarasu, N., Suryanarayanan, T. S., Ravishankar, J. P., El Gueddari, N. E. and Moerschbacher, B. M. (2011). Chitinolytic enzymes from endophytic fungi. *Fungal Diversity*, 47: 43-53.
- Rattanakit, N., Yano, S., Plikomol, A., Wakayama, M. and Tachiki, T. (2007). Purification of *Aspergillus* sp. S1-13 chitinases and their role in saccharification of chitin in mash of solid state culture with shellfish waste. *Journal Bioscience and Bioengineering*,103: 535-541.
- Rattanakit, N., Yano, S., Wakayama, M., Plikomol, A. and Tachiki, T. (2003). saccharification of chitin using solid-state culture of *Aspergillus* sp. S 1-1 3 with shellfish waste as a substrate. *Journal of Bioscience and Bioengineering*, 95: 391-396.
- Rees, R. W., Flood, J., Hasan, Y., Potter, U. and Cooper, R.M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*); mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Pathology*, 58 : 982-989.
- Rita, S. and Durgeshwer, S. (2010). Chitin and Chitosan : Promising biopolymers for wound dressing application. *Journal of Clinical Dermatology*, 11 : 264-268
- Roberts, G.A.F. (1992).*Solubility and solution behaviour of chitin and chitosan*, in: G.A.F. Roberts (Ed.), *Chitin Chemistry*,MacMillan, Houndmills. 274–329.
- Robinson, T. and Nigam, P. (2003). Bioreactor design for protein enrichment of agricultural residues by solid state fermentation. *Biochemical Engineering Journal*,13: 197-203.
- Rosgaard, L., Pedersen, S., Cherry J.R., Harris P. and Meyer A.S. (2006). Efficiency of new fungal cellulose systems in boosting enzymatic degradation of barley straw lignocellulose. *Biotechnological Progress*,22: 493-498.
- Roy, I. and Gupta, M. N. (2003). Applications of microwaves in biological sciences. *Current Science*,85: 1685-1693.
- Roy, I., Mondal, K. and Gupta, M. N (2003). Accelerating enzymatic hydrolysis of chitin by microwave pre-treatment. *Biotechnological Progress*, 19: 1648-1653.
- Sandhya, C., Adapa, L.K., Nampoothiri, M. K., Binod, P., Szakacs, G. and Pandey, A. (2004). Extracellular chitinase production by *Trichoderma harzianum* in submerged fermentation. *Journal of Basic Microbiology*, 44: 49-58.

- Sazwani Binti Suhaimi (2007). *Induction, Purification and Characterization of Chitinase Produced by Trichoderma virens UKM-1*. Universiti Teknologi Malaysia : Thesis Master Biosains.
- Seidl, V. (2008). Chitinases of filamentous fungi: a large group of diverse protein with multiple physiological functions. *Fungal Biology Reviews*, 22 : 36-42.
- Sen, R. and Swaminathan, T (1997). Application of response-surface methodology to evaluate the optimum environmental conditions for the enhanced production of surfactin. *Applied Microbiology and Biotechnology*, 47 : 358-363.
- Shahidi, F., Kamil, J. V. A. and You-Jin, J. (1999). Food applications of chitin and chitosans. *Food Science and Technology*, 10 : 37-51.
- Shailes, R. W., Swaroop, S.K. and Jai, S.G. (2011). Chitinase production in solid-state fermentation from *Oerskovia xanthineo lytica* NCIM 2839 and its application in fungal protoplast formation. *Current Microbiology*, 63: 295-299.
- Sharma, N., Sharma, K.P., Gaur, R. K. and Gupta, V. K. (2011). Role of chitinase of plant defense. *Asian Journal of Biochemistry*, 6 : 29-37.
- Sharon, E., Chet, H. and Spiegel, Y. (2011). *Trichoderma* as a biological control agent. biological control of plant-parasitic nematodes : building coherence between microbial ecology and molecular mechanism, *Springer Science*, 12: 183-194.
- Shelma, R., Paul, W., Sharma, C.P. (2010). Development and characterization of self-aggregated nanoparticles from anacardoylated chitosan as a carrier for insulin. *Carbohydrate Polymers*, 80: 285–290.
- Shi, Y., Xi, X. and Yang Zu (2009). Optimization of *Verticillium lecanii* spore production in solid-state fermentation on sugarcane bagasse. *Applied Microbiology and Biotechnology*, 82: 921-927.
- Shuler, M. L. and Kargi, F. (2002). *Bioprocess engineering: basic concepts*. New Jersey, Prentice Hall. 58-75.
- Singh, A.K., Mehta, G. and Chhaptar, H.S. (2009). Optimization of medium constituents for improved chitinase production by *Paenibacillus* sp. D1 using statistical approach. *Letters in Applied Microbiology*, 49:708-714.
- Singh, V., Khan, M., Saif, K. and Tripathi, C. K. M. (2009). Optimization of actinomycin V production by *Streptomyces triostinicus* using artificial neural network and genetic algorithm. *Applied Microbiology and Biotechnology*, 82: 379-385.

- Singhania, R.R., Patelbo, A. K., Soccol, C.R. and Pandey, A. (2009). Recent advances in solid-state fermentation. *Biochemical Engineering Journal*, 44: 13-18.
- Soetan, K.O., Olaiya, C. O. and Oyewole, O. E. (2010). The importance of mineral elements for humans, domestic animals and plants: A review. *African Journal of Food Science*, 2 : 200-222.
- Souza, C.P., Almeida, B.C., Colwell, R.R. and Rivera, I.N.G. (2011). The importance of chitin in the marine environment. *Marine Biotechnology*, 13: 823-830.
- Souza, R.F., Gomes, R. C., Coelho,R.R.R., Alviano, C.S. and. Soares, R.M.A. (2003). Purification and characterization of an endochitinase produced by *Colletotrichum gloeosporioides*. *FEMS Microbiology Letters*, 222 : 45-50.
- Sudhakar, P. and Nagarajan, P. (2011). Production of chitinase by solid state fermentation from *Serratia marcescens*. *International Journal of Chemical and Technology Research*, 3 : 590-595.
- Suraini, A. A., Teoh, L.S., Noorjahan, A., Shahab, N. and Kamarulzaman, K. (2008). Microbial degradation of chitin materials by *Trichoderma virens* UKM1. *Journal of Biological Science*, 8: 52-59.
- Suresh, P. V. (2012). Biodegradation of shrimp processing bio-waste and concomitant production of chitinase enzyme and N-acetyl-D-glucosamine by marine bacteria: production and process optimization. *World Journal Microbiology and Biotechnology*, 28 : 2945-2962.
- Suresh, P. V. and Anil Kumar, P. K. (2012). Enhanced degradation of α -chitin materials prepared from shrimp processing byproduct and production of N-acetyl-D-glucosamine by thermoactive chitinases from soil mesophilic fungi. *Biodegradation*, 23: 597-607.
- Suresh, P. V., Anil Kumar, P.K. and Sachindra, N.M (2011). Thermoactive β -N-acetylhexosaminidase production by a soil isolate of *Penicillium monoverticillium* CFR2 under solid state fermentation: parameter optimization and application for N-acetyl chitooligosaccharides preparation from chitin. *World Journal Microbiology and Biotechnology*, 27 : 1435-1447.
- Suresh, P. V., Sachindra, N. M. and Bhaskar, N. (2011). Solid state fermentation production of chitin deacetylase by *Colletotrichum lindemuthianum* ATCC

- 56676 using different substrates. *Journal Food Science and Technology*, 48: 348-356.
- Suresh, P.V. and Chandrasekaran, M. (1999). Impact of process parameters on chitinase production by an alkalophilic marine *Beauveria bassiana* in solid state fermentation. *Process Biochemistry*, 34 : 257-267.
- Suresh, P.V. and Chandrasekaran, M., (1998). Utilization of prawn waste for chitinase production by the marine fungus *Beauveria bassiana* by solid state fermentation. *World Journal of Microbiology and Biotechnology*, 14 : 655-660.
- Suresh, P.V., Sakhare, P. Z., Sachindra, N.M. and Halami, P.M. (2012). Extracellular chitin deacetylase production in solid state fermentation by native soil isolates of *Penicillium monoverticillium* and *Fusarium oxysporum*. *Journal Food Science and Technology*, 51: 1594-1599.
- Susana, R.C. (2008). Exploitation of biological wastes for the production of value-added products under solid-state fermentation conditions. *Biotechnology Journal*,3: 859-870.
- Susanto, A., Prasetyo, A.E. and Wening, S. (2009). Infection rate of *Ganoderma* at four soil texture classes. *Jurnal Fitopatologi Indonesia*, 9 : 39-46.
- Synowiek, J. and Al-Khateeb, N.A. (2003). Production, properties, and some new applications of chitin and its derivatives. *Critical Reviews in Food Science and Nutrition*, 43: 145-171.
- Szekeres, A., Leitgeb, B., Kredics, L., Zsuzsanna, A., Hatvani, L., Manczinger, L. and Vagvolgyi, C.S. (2005). Peptaibols and related peptaibiotics of *Trichoderma*. *Acta Microbiologica et Immunologica Hungaria*, 52 : 137-168.
- Tahira, K., Hasan, C., Madiha, H., Kanwal, S. and Aisha, N. (2012). *Fermentor system*. *Food Biotechnology*. Paper work, 1-37.
- Tanaka, T., Fukui, T., Atomi, H. and Imanaka, T. (2003). Characterization of an exo- β -D-glucosaminidase involved in a novel chitinolytic pathway from the hyperthermophilic archaeon *Thermococcus kodakaraensis* KOD1. *Journal of Bacteriology*, 185 : 5175-5181.
- Tarley, C. R. T., Silveira, G., Lopes dos Santos, W., Matos, G.D., Erik G., Bezerra, M. A., Manuel M. and Sérgio, L.C.F. (2009). Chemometric tools in electroanalytical chemistry: Methods for optimization based on factorial design and response surface methodology. *Microchemical Journal*, 92 : 58-67.

- Thamthiankul, S., Suan-Ngay, S., Tantimavanich, S and Panbangred, W. (2001). Chitinase from *Bacillus thuringiensis* subsp. pakistani. *Applied Microbiology and Biotechnology*, 56 : 394-401.
- Tikhonov, V. E., Lopez,L.V., Salinas, J. and Jansson, H.B. (2002). Purification and characterization of chitinases from the nematophagous fungi *Verticillium chlamydosporium* and *Verticillium suchlasporium*. *Fungal Genetics and Biology*, 35 : 67-78.
- Toharisman, A., Suhartono, M.T., Splinder-Barth, M., Hwang, J.K. and Pyun, Y.R. (2005). Purification and characterization of a thermostable chitinase from *Bacillus licheniformis* Mb-2. *World Journal of Microbiology and Biotechnology*, 21: 733-738.
- Tortora, G.J., Funke, B. R. and Case, C. L. (2004). *Microbiology – An Introduction*. 8th Edition. San Francisco, Pearson Education Inc. 130-240.
- Tran, N.H. (2010). Using *Trichoderma* species for biological control of plant pathogens in Vietnam. *Journal Issaas*, 16: 17-21.
- Tsioptsia, C., Tsivintzelis, I, Papadopoulou, L. and Panayiotou, C. (2009). A novel method for producing tissue engineering scaffolds from chitin, chitin–hydroxyapatite, and cellulose. *Material Science and Engineering*, 29: 159-164.
- Ulhoa, C. J. and Peberdy, J.F. (1992). Purification and some properties to the extracellular chitinase produced by *Trichoderma harzianum*. *Enzyme and Microbial Technology*,14: 236-240.
- Vaidya, R., Roy, S., Macmil, S., Gandhi, S.,Vyas, P. and Chhatpa, H. S. (2003). Purification and characterization of chitinase from *Alcaligenes xylosoxydans*. *Biotechnology Letters*, 25: 715-717.
- Vaidya, R.J., Macmil, S. L. A., Vyas, P.R. and Chhatpa, H.S. (2003). The novel method for isolating chitinolytic bacteria and its application in screening for hyperchitinase producing mutant of *Alcaligenes xylosoxydan*. *Letters in Applied Microbiology*, 36 : 129-134.
- Verma, M., Satinder, K. B., Tyagi, R.D., Surampalli, R.Y. and Valero, J.R (2007). Antagonistic fungi, *Trichoderma* spp.: Panoply of biological control. *Biochemical Engineering Journal*, 37: 1-20.
- Vinale, F., Sivashitamparam, K., Ghisalberti, E.L, Marraa, R., Woo, S.L. and Lorito, M. (2008). *Trichoderma*–plant–pathogen interactions. *Soil Biology and Biochemistry*, 40:1-10.

- Vincent, J.F.V. (2002). Insect cuticle: a natural composite shell system. *Composites Part A: Applied Science and Manufacturing*, 33: 1311-1315.
- Vyas, P. and Desphande, M.V (1989). Chitinase production by *Myrothecium verrucaria* and its significance for fungal mycelial degradation. *Journal of General and Applied Microbiology*, 35; 343-350.
- Wang, L. and Yang, S. T. (2007). *Solid state fermentation and its applications*. In : *Bioprocessing for Value-Added Products from Renewable Resources*. Elsevier. 465-474.
- Wang, S. L. and Hwang (2001). Microbial reclamation of shellfish wastes for the production of chitinases. *Enzyme Microbiology and Technology*, 28: 376-382.
- Wang, S. L., Yen, Y. H., Tsiao, W. J., Changa, W. T. and Wang, C. L. (2002). Production of antimicrobial compounds by *Monascus purpureus* CCRC31499 using shrimp and crab shell powder as a carbon source. *Enzyme and Microbial Technology*, 31: 337-344.
- Wang, S. L., Liang, T. W. and Yen, Y. H. (2011). Bioconversion of chitin-containing wastes for the production of enzymes and bioactive materials. *Carbohydrate Polymers*, 84: 732-742.
- Wang, S. L., Lin T. Y., Yen, Y. H, Liao, H. F. and Chen, Y.J. (2006). Bioconversion of shellfish chitin wastes for the production of *Bacillus subtilis* W-118 chitinase. *Carbohydrate Research*, 341: 2507-2515.
- Wang, S.L. and Hwang, J. R. (2001). Microbial reclamation of shellfish wastes for the production of chitinases. *Enzyme and Microbial Technology*, 28: 376-382.
- Wang, S.L., Yena, Y.H., Tsiaoa, W. J., Changa, W. T. and Wang, C. L. (2002). Production of antimicrobial compounds by *Monascus purpureus* CCRC 31499 using shrimp and crab shell powder as a carbon source. *Enzyme and Microbial Technology*, 31: 337-344.
- Wasli, A. S., Salleh, M. M., Abd-Aziz, S., Hassan, O. and Mahadi, N. M. (2009). medium optimization for chitinase production from *Trichoderma virens* using central composite design. *Biotechnology and Bioprocess Engineering*, 14: 781-787.
- Weir, E. (2000). Production of industrial enzymes in fermentation. *Journal Biology*, 5: 213-224.

- Widyastuti, S.M. (2006). The biological control of *Ganoderma* root by *Trichoderma*. *Proceedings of a workshop held in Yogyakarta, Indonesia, 7–9 February 2006*. Canberra, ACIAR Proceedings No. 124. 67-74.
- Wilhite, S. E., Lumsden, R. D., and Straney, D. C. 1994. Mutational analysis of gliotoxin production by the biocontrol fungus *Gliocladium virens* in relation to suppression of Pythium damping-off. *Phytopathology*, 84:816-821
- Win, N. N. and Stevens, W. F. (2001). Shrimp chitin as substrate for fungal chitin deacetylase. *Applied Microbiology and Biotechnology*, 57: 334-341.
- Wojtasz-Pajak, A., Kolodziejaska, I., Debogorska, A. and Malesa-Cieciewicz, M. (1998). Enzymatic, physical and chemical modifications of krill chitin. *Bulletin of the Sea Fisheries Institute (Gdynia)*, 143 : 29-39.
- Xia, G., Jin, C., Zhou, J., Yang, S., Zhang, S. and Jin, C. (2001). A novel chitinase having a unique mode of action from *Aspergillus fumigatus* YJ-407. *European Journal of Biochemistry*, 268 : 4079-4085.
- Yang, J.K., Shih, I.L., Tzeng, Y.M. and Wang, S.L. (2006). Production and purification of protease from a *Bacillus subtilis* that can deproteinize crustacean wastes. *Enzyme and Microbial Technology*, 26 : 406-413.
- Yano, K. S. and Poulos, T. L. (2003). New understandings of thermostable, peizostable enzymes. *Current Opinion in Microbiology*, 14: 360-365.
- Yuli, P.E., Suhartono, M. T., Rukayadi, Y., Hwang, J.K. and Pyun, Y.R. (2004). Characteristics of thermostable chitinase enzymes from the Indonesian *Bacillus* sp.13.26. *Enzyme and Microbial Technology*, 35 : 147-153.
- Yunus, R., Salleh, S. F., Nurhafidzah and Radiah, A.D. (2010). Effect of ultrasonic pre-treatment on low temperature acid hydrolysis of oil palm empty fruit bunch. *Bioresource Technology*, 101 : 9792-9796.
- Zainab Waleed Abdul-Ameer (2008). *Production and Characterization of Inulinase produced by Aspergillus niger using solid state fermentation*. Al-Nahrain University: Thesis B.Sc Bioscience.
- Zhao, L., Xu, L., Mitomo, H. and Yoshii, F. (2006). Synthesis of pH-sensitive PVP/CM-chitosan hydrogels with improved surface property by irradiation. *Carbohydrate Polymers*, 64: 473–480.
- Zhihui, B., Bo, J., Yuejie, L., Jian, C. and Zuming, L. (2008). Utilization of winery wastes for *Trichoderma viride* biocontrol agent production by solid state fermentation. *Journal of Environmental Sciences*, 20 : 353-358.