

SCURRULA FERRUGINEA METHANOL EXTRACT INDUCES REACTIVE
OXYGEN SPECIES-MEDIATED AND MITOCHONDRIAL-DEPENDENT
APOPTOSIS IN BREAST CANCER CELLS

MOHSEN MARVI BAIGI

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*I would like to dedicate this thesis to my beloved wife, my lovely unborn child and
my lovely father and mother
for their endless support and encouragement*

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ABSTRACT

The purpose of this study is to investigate antioxidant and anticancer activities of *Scurrula ferruginea* extracts. The antioxidant activities of the extracts were evaluated using various assays. The extracts were further investigated to examine their cytotoxic activity on human breast cancer cell lines; MDA-MB-231, MDA-MB-468 and MCF-7 using MTT assay. Microscopic examinations of cells were carried out to elucidate the modes of cell death. The effect of the extracts on cancer cells colony formation and migration were determined. Changes in mitochondrial membrane potential and level of reactive oxygen species (ROS) were measured. Western blot and cell cycle analysis were performed to unravel the mechanism of action of extracts against the breast cancer cells. Using GC-MS analysis, chemical composition of extracts were characterized to reveal the presence of anti-cancerous compounds. Our study on stem methanol extract has shown the highest amount of phenolic, flavonoid contents, strong DPPH radical scavenging and metal chelation activity in comparison to other extracts. The stem aqueous and methanol extracts have shown higher cytotoxic effect towards MDA-MB-231 cells compared to other cell lines with IC₅₀ value of 50.35 and 19.27 µg/mL, after 72 h of treatment, respectively. Morphological observations revealed properties of apoptosis in the treated cells. The results displayed that the extracts have the ability to stop migration of cancer cells and also inhibit the colony formation of cancer cells. Moreover, the results have shown that the extracts induced apoptosis in breast cancer cells by ROS generation and mitochondrial depolarization. Furthermore, this study demonstrated that methanol extract inhibited the proliferation of breast cancer cells via induction of cell cycle arrest at G₀/G₁ phase and apoptosis through a mitochondria-dependent apoptosis pathway. The findings of present study revealed the potential antioxidant and anticancer activities of *S. ferruginea* stem methanol extract which may serve as a promising candidate in the search of a new anti-cancer drug.

ABSTRAK

Tujuan kajian penyelidikan ini adalah untuk mengkaji aktiviti antioksidan dan antikanser bagi ekstrak *Scurrula ferruginea*. Aktiviti antioksidan bagi ekstrak dianalisa menggunakan pelbagai kaedah asai. Ekstrak tersebut juga dikaji secara lebih mendalam untuk mengenal pasti aktiviti sitotoksik terhadap garisan sel kanser payudara; MDA-MB-231, MDA-MB-468 dan MCF-7 menggunakan asai MTT. Analisis mikroskopik terhadap sel-sel telah dilaksanakan untuk menghuraikan mod kematian sel. Kesan daripada ekstrak terhadap pembentukan dan migrasi koloni kanser telah ditentukan. Perubahan kepada keupayaan membran mitokondria dan tahap reaktif spesies oksigen (ROS) telah diukur. Western blot dan analisis kitaran sel telah digunapakai untuk menguraikan mekanisme tindakan bagi ekstrak terhadap sel-sel kanser payudara. Dengan menggunakan analisis GC-MS, komposisi kimia bagi ekstrak telah dicirikan dan menunjukkan kehadiran sebatian anti-kanser. Ekstrak methanol batang memberikan kuantiti fenolik dan flavonoid yang sangat tinggi serta aktiviti penyingkiran radikal DPPH dan pengelatan logam yang kukuh berbanding ekstrak yang lain. Ekstrak akueus dan methanol dari batang menunjukkan kesan sitotoksik yang lebih tinggi terhadap sel MDA-MB-231 berbanding garisan sel yang lain dengan nilai IC_{50} masing-masing sebanyak 50.35 dan 19.27 $\mu\text{g/mL}$ setelah 72 jam rawatan. Pemerhatian morfologi mendedahkan ciri-ciri apoptosis dalam sel-sel yang dirawat. Hasil kajian menunjukkan bahawa ekstrak-ekstrak tersebut mempunyai keupayaan untuk memberhentikan migrasi sel-sel kanser di samping menghalang pembentukan koloni sel-sel kanser. Tambahan pula, hasil kajian menunjukkan bahawa ekstrak-ekstrak mencetuskan apoptosis dalam sel-sel kanser payu dara melalui penjanaan ROS dan penyahkutuban mitokondria. Selain itu, kajian ini juga menunjukkan bahawa ekstrak methanol menghalang penyebaran sel-sel kanser payu dara dengan merencatkan kitaran sel pada fasa G₀/G₁ dan apoptosis melalui satu laluan apoptosis yang mempunyai pergantungan terhadap mitokondria. Hasil kajian menunjukkan bahawa aktiviti antioksidan dan antikanser bagi ekstrak methanol dari batang *S. ferruginea* berpotensi menjadi calon kepada pencarian ubat anti-kanser baru.

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LIST OF ABBREVIATION

NCR	-	The National Cancer Registry
DCIS	-	Ductal Carcinoma In Situ
LCIS	-	Lobular Carcinoma In Situ
HER	-	Human Epidermal Growth Factor Receptor
ER	-	Estrogen Receptor
PR	-	progesterone receptor
NCI	-	National Cancer Institute
ROS	-	Reactive Oxygen Species
CAM	-	Complementary and Alternative Medicine
DPPH	-	Diphenyl-2-picryl hydrazine
ABTS	-	2, 2'-azino bis-(3-ethyl benzo thiazoline-6-sulphonic acid)
EDTA	-	Ethylenediaminetetraacetic acid
DMEM	-	Dulbecco's Modified Eagle Medium
FBS	-	Fetal Bovine Serum
PI	-	Propidium Iodide
PBS	-	phosphate buffer saline
DMSO	-	Dimethyl Sulfoxide
TPC	-	Total Phenolic Content
TFC	-	Total Flavonoid Content
GC-MS	-	Gas chromatography-mass spectroscopy
RPMI	-	Roswell Park Memorial Institute

PDT	-	Population Doubling Time
MTT	-	Thiazolyl Blue Tetrazolium Bromide
AO/EB	-	Acridine orange/Ethidium bromide
MMP	-	Mitochondrial Membrane Potential
BCA	-	Bicinchoninic Acid
BSA	-	Bovine Serum Albumin
SDS-PAGE	-	Sodium dodecyl sulfate polyacrylamide gel electrophoresis
AP	-	Alkaline Phosphatase
MOMP	-	Mitochondrial Outer Membrane Permeabilization
DISK	-	Death-Inducing Signaling Complex
ML-I	-	Mistletoe Lectin I
HR-QOL	-	Health-Related Quality Of Life
VA	-	<i>Viscum album</i>
ADCC	-	Antibody-Dependent Cell-mediated Cytotoxicity
TNF α	-	Tumor Necrosis Factor alfa
CRF	-	Cancer Related Fatigue
TCM	-	Traditional Chinese Medicine
LS	-	Life Satisfaction
TEAC	-	Trolox Equivalent Antioxidant Capacity
RT	-	Retention Time

CHAPTER 1

INTRODUCTION

1.1 Research Background

1.1.1 Breast Cancer

Cancer of breast formed due to formation of malignant tumor in the cells of breast. Initially the growth of breast cancer is local which is followed by extension within lymph vessels into regional lymph nodes and invasion of small vein which results in systematic metastatic spread (Spratt & Tobin, 1995). Breast cancer is the most common type of non-skin malignancy among women worldwide. It has been reported that the incidence and mortality of breast cancer have increased during the last two decades (*American Cancer Society Global Cancer Facts & Figures 2nd Edition*, 2011; Jemal *et al.*, 2011; Ferlay *et al.*, 2013). Based on 2006-2010 statistics, the number of deaths in the United States was 22.6 per 100,000 women per year. It is predicted that an estimated 231,840 new cases of breast cancer and 40,730 breast cancer-related deaths will occur among women in 2015 worldwide (“American Cancer Society. Cancer Facts & Figures,” 2015)

The incidence rate of breast cancer is highest in North America with the age standardized rates of 99.4 per 100,000 population, followed by countries in the Eastern Europe, South America, Southern Africa, and western Asia with moderate incidence rates, while the lowest incidence rates are reported in most African countries (Yip *et al.*, 2006; Ferlay *et al.*, 2010).

It is reported that approximately one million female are diagnosed with breast malignancy with an estimated 410,000 deaths every year, worldwide (Coughlin & Ekwueme, 2009). The incidence and mortality of breast cancer was reported lower in low-resource countries compared to high-resource countries (Smith, 2006). In most of the Asian countries, the incidence rate of breast cancer is increasing (Abdullah *et al.*, 2013). An increasing in the prevalence of breast cancer was reported in Malaysia as well (Abdullah *et al.*, 2013). The highest incidence rate for breast cancer in Malaysia was observed at women between 50-60 years old (Dahlui *et al.*, 2011). It is estimated that one out of twenty Malaysian women have chance to get breast cancer at some point of their lives (Dahlui *et al.*, 2011).

Breast cancer is the most common cancer among Malaysian women (Lim *et al.*, 2008). The National Cancer Registry (NCR) 2003-2005 reported an age-standardized rate (ASR) of 47.3 per 100 000. The incidence is highest in Chinese (59.9 per 100 000) followed by Indians (54.2 per 100 000) and Malays (34.9 per 100 000) (Lim *et al.*, 2008). The International Agency for Research in Cancer (GLOBOCAN) 2012 estimated the ASR of breast cancer in Malaysia as 38.7 per 100,000 with 5410 new cases in 2012 (["http://globocan.iarc.fr,"](http://globocan.iarc.fr)).

1.1.2 Breast Cancer Treatment

Different treatment options are currently available including local therapy and systemic therapy. Local therapy includes surgery, radiotherapy or a combination of the two, applied to kill cancer cells from a limited (local) area such as lymph nodes, breast and chest wall. Systemic therapy includes endocrine or hormone therapy and chemotherapy which administered following primary surgery or radiotherapy to kill or inhibit metastases and to improve survival. Table 1.1 represents various methods of breast cancer treatment and their common side effects. Selection of treatment strategies depend on tumor size, metastatic potential, axillary lymph node status and molecular and patient profile (Liao *et al.*, 2013). Systemic therapy with cytotoxic chemotherapy and endocrine therapy were found to be effective in prolonging disease-free and survival time (Peto *et al.*, 2000).

Table 1.1: Summary of various methods of breast cancer treatment and their common side effects.

Methods	Mechanism of action	Side effects	References
Surgery	Conservative and mastectomy	Lymphedema, chronic nerve damage, infection at the incision site, armpit discomfort	(Karen <i>et al.</i> , 2002; Ridner <i>et al.</i> , 2011)
Radiotherapy	Using high dose of radiation	Skin reactions of the area being radiated	(Sjövall <i>et al.</i> , 2010)
Biological targeted therapy	Using monoclonal antibody and medicine Herceptin (Trastuzumab) Tykerb (lapatinib)	Weakness, diarrhea, Pain, fever Itchy and dry skin, diarrhea	(Nahta <i>et al.</i> , 2006)
Endocrine or hormone therapy	Using aromatase inhibitors and tamoxifen by blocking the action of estrogen Tamoxifen: Aromatase inhibitors:	Vaginal discharge, an increase in thromboembolic events and uterine sarcoma Musculoskeletal adverse effect ,hot flashes, increased LDL, loss of libido, vaginal dryness	(Kalidas & Brown, 2005; Connor & Attai, 2013)
Chemotherapy	The most commonly type of treatment using anti-breast cancer drugs Carboplatin, Cisplatin: Cyclophosphamide:	Nephrotoxicity Pulmonary toxicity	(Yao <i>et al.</i> , 2007; Chandwani <i>et al.</i> , 2012; Gianni <i>et al.</i> , 2008)

Despite of varied side effects, using chemotherapy either as a single compound or combination therapy with multiple-agents is still the most commonly used treatment option by breast cancer patients (Ozer *et al.*, 2000). Chemotherapy uses anti-breast cancer drugs and cytotoxic agents for treatment of metastatic breast cancer (ER-negative tumors). Tumor cell response to chemotherapy and cytotoxic agents through an active form of cell death is known as apoptosis or programmed cell death. It is now well established that other modes of cell death such as necrosis and autophagy also take place following chemotherapy in tumor cells (Brown & Attardi, 2005).

1.2 Problem Statement

Although many treatment methods are currently established including surgery, radiotherapy, biological therapy, hormone therapy and chemotherapy, these therapies are less effective and recurrence is still occurring in breast cancer patients due to side effects and toxicity of drugs in normal cell and aggressive behaviour of the tumours (Table 1.3). In spite of many improvement in the use of hormonal and adjuvant cytotoxic therapies in breast cancer patients, there is no considerable reduction in mortality of breast cancer today (Eggenschwiler *et al.*, 2007). Costly treatment methods and serious side effects associated with available therapies may cause greater tendencies among people to use herbal medicines for health care.

Complementary and alternative medicine (CAM) as one of the major aspect of cancer therapy has been developed in last few years in order to alleviate drug side effects and relief pain in breast cancer patients (Ostermann *et al.*, 2009). A large proportion of cancer patients (up to 80%) use complementary and alternative medicine (CAM) (Vardy *et al.*, 2013). Breast cancer patients are among the most likely users of CAM (Bennett *et al.*, 2009). Among CAM, herbal supplements (anti-oxidants) is the most commonly used group of cancer treatment. Cancer treatment using herbal medicine has a history of more than 2000 years (Craig, 1999).

Harmful effects of conventional treatment as well as toxicity of chemotherapy create a significant problem in breast cancer therapy. The alternate solution to decrease side effects of chemotherapeutic drugs is the use of medicinal plants. Use of medicinal plants which have fewer side effects as compared to synthetic drugs can provide an alternative to the use of conventional allopathic medicine for treatment of breast cancer. In addition, any practical solution to manage cancer progression is of paramount importance. Therefore, there is a need to evaluate whether medicinal plant extracts are able to act as potent anticancer agent by controlling the cancer progression or arresting the carcinogenic process.

Previous research findings have shown that various European mistletoe extracts from different host trees are capable of inducing apoptosis and cell death in numerous tumor cells and human cancer cell lines (Ramaekers *et al.*, 2007; Harmsma *et al.*, 2006).

Although various studies investigated the effect of European mistletoe on cancer, not many studies focused on other species of mistletoe from other continents. Malaysia's rainforest being part of the world's tropical rainforest is also considered as one of the most evolved and diverse rainforest in the world. *Scurrula ferruginea* is one of the mistletoe species in Malaysia which is used as a folk medicine for treatment of several ailments (Barlow, 1991). It has been reported that a decoction of *S. ferruginea* leaves along with *Millettia sericea* used for bathing malarial patients. In addition, a poultice of the pounded leaves administered as a post-partum protective medicine and also applied for snake bite and wound (Burkill *et al.*, 1966) (Perry, 1978). Moreover, this plant is traditionally employed in the treatment of many diseases including gastrointestinal malfunction, high blood pressure and hypertension (Ameer *et al.*, 2009).

Ethno-medical knowledge plays an important role in selection of plants for discovery of novel drugs. Therefore, *S. ferruginea* was selected for the present study based on its reputation in folk medicine. There is no report on antioxidant capacity, anticancer activity and mechanism of action of *S. ferruginea*. The current study

provide the scientific rational for antioxidant and anti-breast cancer activities of *S. ferruginea*.

1.3 Objectives of Study

Based on the above-mentioned problem statements, the objectives of the present study are as follow:

1. To evaluate potential of *S. ferruginea* crude extracts based on the antioxidant activity and phytochemical analysis
2. To investigate the selective cytotoxic effects of selected extracts on breast cancer cells and study apoptosis-inducing effects of extracts
3. To study the mechanism of growth arrest and unravel apoptotic pathway involve in breast cancer cell death by selected extract

1.4 Scope of Study

Aerial parts of *S. ferruginea* (Jack) Danser including stems, leaves and flowers were used in the present study. Different types of breast cancer cell lines including MCF-7 (luminal A breast carcinoma), MDA-MB-231(Claudin-low breast carcinoma) and MDA-MB-468 (basal-like breast carcinoma) which are differ in molecular markers status and invasiveness have been selected for the present study.

To achieve the listed objectives, the study was confined to the following scopes:

1. Determination of total phenolic and total flavonoid content by Folin-Ciocalteu and aluminum chloride methods, respectively and antioxidant activities of different extracts by assessing DPPH free

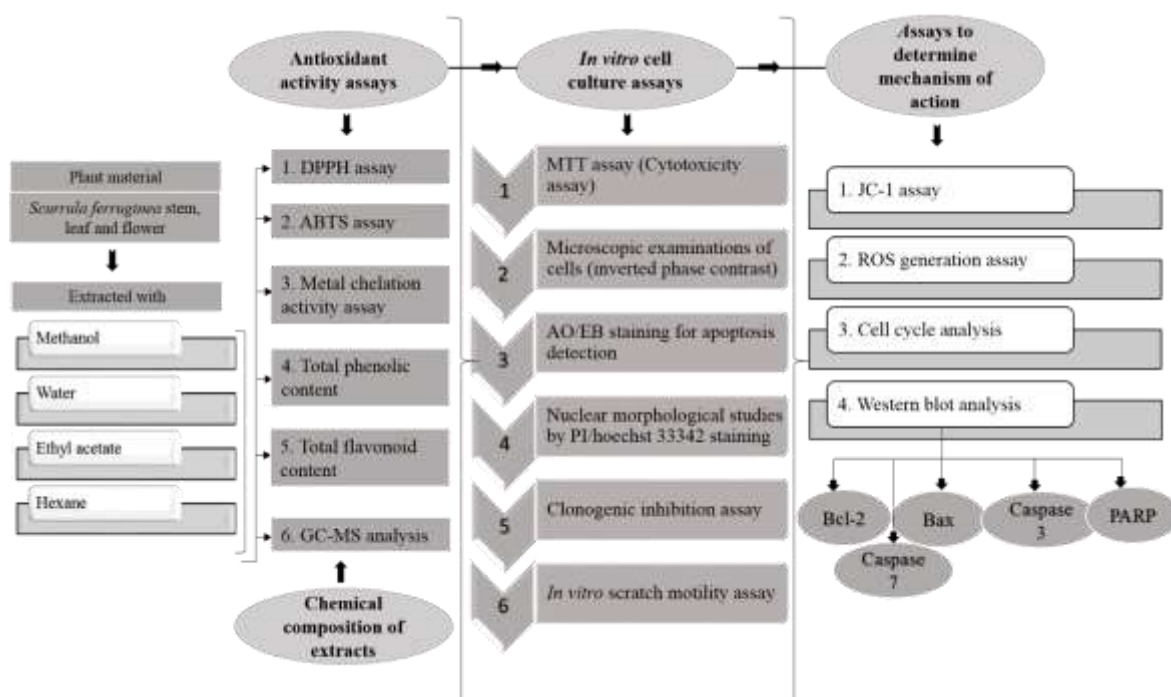
radical scavenging activity, ABTS and metal chelation capacity of *S. ferruginea* extracts.

2. Analysis of chemical composition using GC-MS of *S. ferruginea* extracts.
3. Evaluation of selective cytotoxic activities of selected extracts against breast cancer cell lines and non-cancerous cell line using MTT assay and characterization of the cell death using AO/EB and Hoechst/PI staining methods.
4. Determination of cell migration inhibition efficiency and colony forming ability of treated cancer cells using scratch assay and colony forming assay respectively.
5. Measurement of mitochondrial membrane potential by JC-1 assay and investigation on the potential mechanism of apoptosis as the result of oxidative stress by measuring intracellular ROS level using DCF-DA assay.
6. Determination of cell death mechanism pathway of selected extract against breast cancer cell through the regulation of bcl-2, bax, caspase-3, caspase-7 and PARP proteins using western blot analysis and possible cell cycle arrest using flow cytometric analysis.

1.5 Significant of Study

- i. Growth inhibitory effects on different carcinoma cell types may be crucial for effective control of breast cancer; therefore, the present study is of great importance to introduce a novel candidate in battling breast cancer particularly ER-negative breast carcinoma.
- ii. The present study is also paving the way for further research on *S. ferruginea* in the field of pharmaceutical industry and anti-cancer drug discovery for the development of anticancer agents.
- iii. This study provides an experimental basis for systematic and clinical research of medicines for treatment of breast cancer in the future.

1.6 Methodology



REFERENCES

- Abdullah, N. A., Rozita, W., Mahiyuddin, W., Muhammad, N. A., Ali, Z. M., Ibrahim, L., Kamaluddin, M. A. (2013). Survival rate of breast cancer patients in Malaysia : a population-based study. *Asian Pacific Journal of Cancer Prevention*, 14, 4591–4594.
- Abhyankar, G., Suprasanna, P., Pandey, B. N., Mishra, K. P., Rao, K. V., & Reddy, V. D. (2010). Hairy root extract of *Phyllanthus amarus* induces apoptotic cell death in human breast cancer cells. *Innovative Food Science & Emerging Technologies*, 11(3), 526–532.
- Ahmad, P., Jaleel, C., Salem, M., Nabi, G., & Sharma, S. (2010). Roles of enzymatic and nonenzymatic antioxidants in plants during abiotic stress. *Critical reviews in biotechnology*, 3, 161–75.
- Ameer, O. Z., Salman, I. M., Yam, M. F., Abd Allah, H. H., Abdulla, M. H., Shah, A. M., Asmawi, M. Z. (2009). Vasorelaxant Properties of *Loranthus ferruginea* Roxb. Methanolic Extract. *International Journal of Pharmacology*, 5(1), 44–50.
- American Cancer Society *Global Cancer Facts & Figures 2nd Edition*. (2011). *Global Cancer Facts & Figures 2nd Edition*. Atlanta: American Cancer Society.
- American Cancer Society. *Cancer Facts & Figures*. (2015). *American Cancer Society*.
- Amudha, M., & Rani, S. (2014). GC-MS Analysis of Bioactive components of *Cordia retusa* (Boraginaceae). *Hygeia: journal for drugs and medicines*, 6(April), 12–19.
- Andersson, I. (2005). Invasive Breast Cancer. In *Radiologic-Pathologic Correlations from Head to Toe* (pp. 757–766).

- Anna-Maria, L.-L., Velasco, M. G., & Reinhard, B. (2006). Mistletoe treatments for minimising side effects of anticancer chemotherapy. *GMS Health Technology Assessment*, 2, 1–8.
- Arora, S., Bhardwaj, A., Srivastava, S. K., Singh, S., McClellan, S., Wang, B., & Singh, A. P. (2011). Honokiol arrests cell cycle, induces apoptosis, and potentiates the cytotoxic effect of gemcitabine in human pancreatic cancer cells. *PloS one*, 6(6), e21573.
- Ashraf, M. F., Aziz, M. A., Stanslas, J., Ismail, I., & Kadir, M. A. (2013). Assessment of Antioxidant and Cytotoxicity Activities of Saponin and Crude Extracts of *Chlorophytum borivilianum*. *The Scientific World Journal*, 2013.
- Atasever-Arslan, B., Yilancioglu, K., Bekaroglu, M., Taskin, E., Altinoz, E., & Cetiner, S. (2015). Cytotoxic effect of extract from *Dunaliella salina* against SH-SY5Y neuroblastoma cells. *General physiology and biophysics*, 34(2), 201–207.
- Auerbach, L., Dostal, V., Václavik-Fleck, I., Kubista, E., Rosenberger, A., Rieger, S., Schierholz, J. . (2005). Signifikant höherer Anteil aktivierter NK-Zellen durch additive misteltherapie bei chemotherapierten Mamma-Ca-Patientinnen in einer prospektiv randomisierten doppelblinden Studie. In R. Scheer, R. Bauer, H. Becker, V. Fintelmann, F. Kemper, & H. Schilcher (Eds.), *Fortschritte in der Misteltherapie* (pp. 543–554). Essen: KVC Verlag.
- Bailly, C. (2009). Ready for a comeback of natural products in oncology. *Biochemical pharmacology*, 77(9), 1447–1457.
- Bali, E., Açık, L., Elçi, P., Sarper, M., Avcu, F., & Vural, M. (2015). *In vitro* antioxidant, cytotoxic and pro-apoptotic effects of *Achillea teretifolia* Willd extracts on human prostate cancer cell lines. *Pharmacognosy magazine*, 11, 308–315.
- Balsano, C., & Alisi, A. (2009). Antioxidant effects of natural bioactive compounds. *Current pharmaceutical design*, 15(26), 3063–3073.
- Bantel, H., Engels, I. H., Voelter, W., Klaus, S.-O., & Sebastian, W. (1999). Mistletoe lectin activates caspase-8 / FLICE independently of death receptor signaling and enhances anticancer drug-induced apoptosis. *Cancer Research*, 59, 2083–2090.
- Barlow, B. A. (1991). Provisional key to the genera of Loranthaceae and Viscaceae of the flora Malesiana region. *Flora Malesiana Bull*, 10, 335–338.

- Bar-Sela, G. (2011). White-Berry Mistletoe (*Viscum album L.*) as complementary treatment in cancer: Does it help? *European Journal of Integrative Medicine*, 3(2), e55–e62.
- Becker, H. (1986). Botany of European Mistletoe (*Viscum Album L.*). *Oncology*, 43(Suppl 1), 2–7.
- Becker, H. (2000). *European mistletoe: Taxonomy, host trees, parts used, physiology.* (A Büssing & Mistletoe The Genus *Viscum*, Eds.) (pp. 31–41). Harwood Academic Publishers.
- Bennett, J. a, Cameron, L. D., Whitehead, L. C., & Porter, D. (2009). Differences between older and younger cancer survivors in seeking cancer information and using complementary/alternative medicine. *Journal of general internal medicine*, 24(10), 1089–94.
- Berger, M., & Schm, D. (1983). Studies on the Tumor-inhibiting Efficacy of Iscador in Experimental Animal Tumors. *J Cancer Res Clin Oncol*, 262–265.
- Beuth, J, Gabius, H., Steuer, M., Geisel, J., Ko, H., & Pulverer, G. (1993). Influence of mistletoe lectin administration on defined acute phase reactants in cancer patients. *Medizinische Klinik*, 88(5), 287–290.
- Beuth, J, Ko, H., Gabius, H., Burrichter, H., Oette, K., & Pulverer, G. (1992). Behavior of lymphocyte subsets and expression of activation markers in response to immunotherapy with galactoside-specific lectin from mistletoe in breast cancer patients. *Clinical Investigator*, 70, 658–661.
- Beuth, J, Ko, H. L., Schneider, H., Tawadros, S., Kasper, H. U., Zimst, H., & Schierholz, J. M. (2006). Intratumoral application of standardized mistletoe extracts down regulates tumor weight via decreased cell proliferation, increased apoptosis and necrosis in a murine model. *Anticancer research*, 26(6B), 4451–456.
- Beuth, J, Ko, H., Tunggal, L., Geisel, J., & Pulverer, G. (1993). Comparative studies on the immunoactive action of galactoside-specific mistletoe lectin. Pure substance compared to the standardized extract. *Arzneimittelforschung*, 43(2), 166–169.
- Beuth, J, Schneider, B., & Schierholz, J. M. (2008). Impact of complementary treatment of breast cancer patients with standardized mistletoe extract during aftercare: a controlled multicenter comparative epidemiological cohort study. *Anticancer research*, 28(1B), 523–527.

- Beuth, J., Stoffel, B., Ko, H., Buss, G., Tunggal, L., & Pulverer, G. (1995). Immunoactive effects of various mistletoe lectin-1 dosages in mammary carcinoma patients. *Arzneimittelforschung*, *45*(4), 505–507.
- Beuth, Josef. (2009). Evidence-Based Complementary Medicine in Breast Cancer Therapy. *Breast care (Basel, Switzerland)*, *4*(1), 8–12.
- Block, K. I., Koch, A. C., Mead, M. N., Tothy, P. K., Newman, R. a, & Gyllenhaal, C. (2007). Impact of antioxidant supplementation on chemotherapeutic efficacy: a systematic review of the evidence from randomized controlled trials. *Cancer treatment reviews*, *33*(5), 407–18.
- Bock, P. R., Friedel, W. E., Karasmann, M., & Schneider, B. (2004). Efficacy and Safety of Long-term Complementary Treatment with Standardised European Mistletoe Extract (*Viscum album L*) in Addition to the Conventional Adjuvant Oncological Therapy in Patients with Primary Non-metastatic Breast Cancer. *Arzneimittelforschung*, *54*(8), 456–466.
- Bogoeva, V., Ivanov, I., Kulina, H., Russev, G., & Atanasova, L. (2013). A novel cytokinin-binding property of mistletoe lectin I from *viscum album*. *Biotechnology & Biotechnological Equipment*, *27*(1), 3583–3585.
- Bont, R. De, & Larebeke, N. Van. (2004). Endogenous DNA damage in humans : a review of quantitative data. *Mutagenesis*, *19*(3), 169–185.
- Borrelli, E. (2001). Evaluation of the quality of life in breast cancer patients undergoing lectin standardized mistletoe therapy. *Minerva Medica*, *92*, 105–107.
- Brandenberger, M., Simões-Wüst, A. P., Rostock, M., Rist, L., & Saller, R. (2012). An exploratory study on the quality of life and individual coping of cancer patients during mistletoe therapy. *Integrative cancer therapies*, *11*(2), 90–100.
- Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995). Use of a Free Radical Method to Evaluate Antioxidant Activity. *Lebensm Wiss Technology*, *28*, 25–30.
- Bras, M., Queenan, B., & Susin, S. A. (2005). Programmed Cell Death via Mitochondria : Different Modes of Dying. *Biochemistry*, *70*(2), 231–239.
- Brien, K. M. O., Cole, S. R., Tse, C., Perou, C. M., Carey, L. A., Foulkes, W. D., Millikan, R. C. (2011). Intrinsic breast tumor subtypes, race, and long-term

- survival in the Carolina Breast Cancer Study. *Clinical cancer research*, 16(24), 6100–6110.
- Brown, J., & Attardi, L. (2005). The role of apoptosis in cancer development and treatment response. *Nature Reviews Cancer*, 5, 231–237.
- Bruno, S., Bino, G. Del, Gorczyca, W., Hotz, M. A., Lassota, P., & Traganos, F. (1992). Features of apoptotic cells measured by flow cytometry. *Cytometry*, 808, 795–808.
- Burger, A., Mengers, U., Kelter, G., Schüler, J., & Fiebig, H. (2003). No evidence of stimulation of human tumor cell proliferation by a standardized aqueous mistletoe extract *in vitro*. *Anticancer Research*, 23(5A), 3801–3806.
- Burger, A., Mengers, U., Schüler, J., & Fiebig, H. (2001). Antiproliferative activity of an aqueous mistletoe extract in human tumor cell lines and xenografts *in vitro*. *Arzneimittelforschung*, 51(9), 748–757.
- Burkill, I. H., Birtwistle, W., Foxworthy, F. W., Scrivenor, J. B., & Watson, J. G. (1966). *A dictionary of the economic products of the Malay peninsula. Vol. II (I-Z)*. Kuala Lumpur, Malaysia: on behalf of the government of Malaysia and Singapore by the ministry of Agriculture and co-operatives.
- Büssing, A. (2000a). *Biological and Pharmacological Properties of Viscum Album L. In Mistletoe. The Genus Viscum*. (A Büssing, Ed.) (pp.123–182). Amsterdam: Harwood Academic Publishers.
- Büssing, A. (2000b). *Mistletoe. The Genus Viscum*. Amsterdam: Harwood Academic Publishers.
- Büssing, A. (2006). Immune Modulation Using Mistletoe (*Viscum Album L.*) Extracts Iscador. *Arzneimittelforschung*, 56(6A), 508–515.
- Büssing, A., Bücknerb, U., Enser-Weisb, U., Schnellec, M., Schumannc, A., Schietzeld, M., Hackmanne, J. (2008). Modulation of chemotherapy-associated immunosuppression by intravenous application of *Viscum album L.* Extract (Iscador): A randomised phase II study. *European Journal of Integrative Medicine*, 1(1), 2–3.
- Büssing, A, Stumpf, C., Tröger, W., & Schietzel, M. (2007). Course of mitogen-stimulated T lymphocytes in cancer patients treated with *Viscum album* extracts. *Anticancer research*, 27(4C), 2903–2910.
- Büssing, A., Troger, W., Stumpf, C., & Schietzel, M. (2008). Local Reactions to Treatments with *Viscum album L.* Extracts and their Association with T-

- Lymphocyte Subsets and Quality of Life. *Anticancer research*, 28, 1893–1898.
- Büssing, Arndt, Bischof, M., Hatzmann, W., Bartsch, F., Soto-Vera, D., Fronk, E.-M., Stein, G. M. (2005). Prevention of surgery-induced suppression of granulocyte function by intravenous application of a fermented extract from *Viscum album L.* in breast cancer patients. *Anticancer research*, 25(6C), 4753–4757.
- Büssing, Arndt, Raak, C., & Ostermann, T. (2012). Quality of life and related dimensions in cancer patients treated with mistletoe extract (iscador): a meta-analysis. *Evidence-based complementary and alternative medicine*, 2012.
- Bussinga, A., Suzartb, K., Bergmann, J., Pfiillefl, U., Schietzek, M., & Schweizerb, K. (1996). Induction of apoptosis in human lymphocytes treated with *Viscum album L.* is mediated by the mistletoe lectins. *Cancer Letters*, 99, 59–72.
- Butler, M. S. (2004). The Role of Natural Product Chemistry in Drug Discovery. *Journal of Natural Products*, 67, 2141–2153.
- Carey, L. A., Perou, C. M., Livasy, C. A., Dressler, L. G., Cowan, D., Conway, K., Earp, H. S. (2006). Race, Breast Cancer Subtypes, and Survival in the Carolina Breast Cancer Study. *Journal of American Medical Association*, 295(21), 2492–2502.
- Chandwani, K. D., Ryan, J. L., Peppone, L. J., Janelsins, M. M., Sprod, L. K., Devine, K., Mustian, K. M. (2012). Cancer-related stress and complementary and alternative medicine: a review. *Evidence-based complementary and alternative medicine : eCAM*, 2012, 979213.
- Chien Lee, A. T., Azimahtol, H. L. P., & Tan, A. N. (2003). Styrylpyrone Derivative (SPD) induces apoptosis in a caspase-7-dependent manner in the human breast cancer cell line MCF-7. *Cancer cell international*, 3(16), 1–8.
- Chinnaiyan, A., O'Rourke, K., Tewari, M., & Dixit, V. (1995). FADD, a novel death domain-containing protein, interacts with the death domain of Fas and initiates apoptosis. *Cell*, 81(4), 505–512.
- Chipuk, J. E., & Green, D. R. (2011). How do BCL-2 proteins induce mitochondrial outer membrane permeabilization? *Trends in cell biology*, 18(4), 157–164.
- Christen, P., & Cuendet, M. (2012). Plants as a source of therapeutic and health products. *Chimia*, 66(5), 320–323.

- Connor, C., & Attai, D. (2013). Adjuvant endocrine therapy for the surgeon: Options, side Effects, and their management. *Annals of surgical oncology*, 20(10), 3188–3193.
- Coriell, L. (1979). Preservation, storage, and shipment. *Methods in enzymology*, 58, 29–36.
- Cory, S., & Adams, J. (2002). The Bcl2 family: regulators of the cellular life-or-death switch. *Nature Reviews Cancer*, 2(9), 647–656.
- Cotter, T. G. (2009). Apoptosis and cancer: the genesis of a research field. *Nature Reviews Cancer*, 9, 501–507.
- Coughlin, S., & Ekwueme, D. (2009). Breast cancer as a global health concern. *Cancer Epidemiology*, 33, 315–318.
- Cragg, G. M., Boyd, M. R., Khanna, R., Kneller, R., Mays, T. D., Mazan, K. D., Sausville, E. A. (1999). International collaboration in drug discovery and development: the NCI experience. *Pured and applied chemistry*, 71(9), 1619–1633.
- Cragg, G. M., Newman, D. J., & Yang, S. S. (2006). Natural Product Extracts of Plant and Marine Origin Having Antileukemia Potential. *Journal of Natural Prodct*, 69, 488–498.
- Cragg, G., Newman, D., & Weiss, R. (1997). Coral reefs, forests, and thermal vents: the worldwide exploration of nature for novel antitumor agents. *Semin Oncology*, 24, 156–163.
- Craig, W. J. (1999). Health-promoting properties of common herbs. *The american journal of clinical nutrition*, 70, 491–499.
- Dahlui, M., Ramli, S., & Bulgiba, A. M. (2011). Breast Cancer Prevention and Control Programs in Malaysia. *Asian Pacific Journal of Cancer Prevention*, 12, 1–4.
- Danial, N., & Korsmeyer, S. (2004). Cell death: critical control points. *Cell*, 116, 205–219.
- Danial, N. N., & Korsmeyer, S. J. (2004). Cell Death: Critical Control Points Review. *Cell*, 116, 205–219.
- Dashora, N., Sodde, V., Prabhu, K. S., & Lobo, R. (2011a). *In vitro* cytotoxic activity of *Dendrophthoe falcata* on human breast adenocarcinoma cells-MCF-7. *International Journal of Cancer Research*, 7(1), 47–54.

- Dashora, N., Sodde, V., Prabhu, K. S., & Lobo, R. (2011b). Antioxidant Activities of *Dendrophthoe falcata* (L.f) Etting. *Pharmaceutical Crops*, 24–27.
- De Vries, E. G. E., Gietema, J. a, & de Jong, S. (2006). Tumor necrosis factor-related apoptosis-inducing ligand pathway and its therapeutic implications. *Clinical cancer research : an official journal of the American Association for Cancer Research*, 12(8), 2390–3.
- Decaudin, D., Marzo, I., Brenner, C., & Kroemer, G. (1998). Mitochondria in chemotherapy-induced apoptosis: a prospective novel target of cancer therapy. *International journal of oncology*, 12(1), 141–152.
- Decker, E. A., & Welch, B. (1990). Role of Ferritin as a Lipid Oxidation Catalyst in Muscle Food. *Journal of agricultural and food chemistry*, 38, 674–677.
- Desagher, S., & Martinou, J. (2000). Mitochondria as the central control point of apoptosis. *Trends in cell biology*, 10(9), 369–377.
- Donepudi, M., Mac, S., Briand, C., & Grütter, M. (2003). Insights into the regulatory mechanism for caspase-8 activation. *MolecularCell*, 11(2), 543–549.
- Drees, M., Berger, D. P., Dengler, W. A., & Fiebig, G. H. (1996). Direct cytotoxic effects of preparations used as unconventional methods in cancer therapy in human tumor xenografts in the clonogenic assay and in nude mice. *Immunodeficient Animals: Models for Cancer Research (Arnold W, Köpf-Maier P, Micheel B, eds)*. Basel, Karger Verlag, 51, 115–122.
- Du, C., Fang, M., Li, Y., Li, L., & Wang, X. (2000). Smac, a mitochondrial protein that promotes cytochrome c-dependent caspase activation by eliminating IAP inhibition. *Cell*, 102(1), 33–42.
- Eggenschwiler, J., von Balthazar, L., Stritt, B., Pruntsch, D., Ramos, M., Urech, K., Viviani, A. (2007). Mistletoe lectin is not the only cytotoxic component in fermented preparations of *Viscum album* from white fir (*Abies pectinata*). *BMC complementary and alternative medicine*, 7(14).
- Eisenbraun, J., Scheer, R., Kröz, M., Schad, F., & Huber, R. (2011). Quality of life in breast cancer patients during chemotherapy and concurrent therapy with a mistletoe extract. *Phytomedicine : international journal of phytotherapy and phytopharmacology*, 18(2-3), 151–157.
- Endo, Y., Tsurugi, K., & Franz, H. (1988). The site of action of the A-chain of mistletoe lectin I on eukaryotic ribosomes. The RNA N-glycosidase activity of the protein. *FEBS letters*, 231(2), 378–380.

- Evan, G., Brown, L., Whyte, M., & Harrington, E. (1995). Apoptosis and the cell cycle. *Current opinion in cell biology*, 7(6), 825–834.
- Fasching, P. a, Thiel, F., Nicolaisen-Murmann, K., Rauh, C., Engel, J., Lux, M. P., Bani, M. R. (2007). Association of complementary methods with quality of life and life satisfaction in patients with gynecologic and breast malignancies. *Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer*, 15(11), 1277–84.
- Favaloro, B., Allocati, N., Graziano, V., Ilio, C. Di, & Laurenzi, V. De. (2012). role of apoptosis in disease. *AGING*, 4(5), 330–349.
- Ferlay, J., Héry, C., Autier, P., & Sankaranarayanan, R. (2010). *Global burden of breast cancer. Breast Cancer*.
- Ferlay, J., Soerjomataram, I., Ervik, M., Dikshit, R., Eser, S., Mathers, C., Bray, F. (2013). Cancer Incidence and Mortality Worldwide: IARC Cancer Base. *Globocan 2012*, 1(11).
- Figueiredo, C. R., Matsuo, A. L., Pereira, F. V, Rabaça, A. N., Farias, C. F., & Girola, N. (2015). Pyrostegia venusta heptane extract containing saturated aliphatic hydrocarbons induces apoptosis on B16F10 Nex2 melanoma cells and displays antitumor activity *in vivo*. *Pharmacognosy magazine*, 10(Suppl 2), 1–20.
- Fisher, E., Gregorio, R., Fisher, B., Redmond, C., Vellios, F., & Sommers, S. (1975). The pathology of invasive breast cancer. A syllabus derived from findings of the National Surgical Adjuvant Breast Project (protocol no. 4). *Cancer*, 1, 1–85.
- Franken, N. A. P., Rodermond, H. M., Stap, J., Haveman, J., & van Bree, C. (2006). Clonogenic Assay of Cells *In Vitro*. *Nature protocols*, 1(5), 2315–2319.
- Freshney, R. I. (2000). *Culture of animal cells: A manual of basic technique* (4th ed.). New York: Wiley Liss.
- Freshney, R. I. (2010). *Culture of animal cells: A manual of basic technique and spacialized application* (Sixth.). New Jersey: Wiley Liss.
- Fulda, S, & Debatin, K.-M. (2006). Extrinsic versus intrinsic apoptosis pathways in anticancer chemotherapy. *Oncogene*, 25(34), 4798–811.
- Fulda, Simone. (2010). Evasion of apoptosis as a cellular stress response in cancer. *International journal of cell biology*, 2010, 370835.

- Ganie, S. A., Dar, T. A., Hamid, R., Zargar, O., Abeer, S. U., Masood, A., Zargar, M. A. (2014). *In Vitro* Antioxidant and Cytotoxic Activities of *Arnebia benthamii* (Wall ex.G.Don): A Critically Endangered Medicinal Plant of Kashmir Valley. *Oxidative Medicine and Cellular Longevity*, 2014.
- Gardin, N. E. (2009). Immunological response to mistletoe (*Viscum album L*) in cancer patients : A four-case series. *Phytotherapy Research*, 23, 407–411.
- Gardner, P. T., White, T. A. C., Mcphail, D. B., & Duthie, G. G. (2000). The Relative Contributions of Vitamin C , Carotenoids and Phenolics to the Antioxidant Potential of Fruit Juices. *Food Chemistry*, 68, 471–474.
- Gerber, B., Scholz, C., Reimer, T., Briese, V., & Janni, W. (2006). Complementary and alternative therapeutic approaches in patients with early breast cancer: a systematic review. *Breast cancer research and treatment*, 95(3), 199–209.
- Gianni, L., Cole, B. F., Panzini, I., Snyder, R., Holmberg, S. B., Byrne, M., Ravaoli, A. (2008). Anemia during adjuvant non-taxane chemotherapy for early breast cancer: Incidence and risk factors from two trials of the International Breast Cancer Study Group. *Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer*, 16(1), 67–74.
- Gomes, A., Fernandes, E., & Lima, J. (2005). Fluorescence probes used for detection of reactive oxygen species. *Journal of Biochemical and Biophysical Methods*, 65(2), 45–80.
- Goossens, M. E., Buntinx, F., & Zeegers, M. P. (2009). Re: Selenium and vitamin E: interesting biology and dashed hope. *Journal of the National Cancer Institute*, 101(19), 1363–4; author reply 1364.
- Govindappa M, C. (2015). Gc-MS Study of Two Column Fractions from Methanol Extracts of *Loranthus Micranthus* and Their *In Vivo* Antidiabetic Activity on Alloxan Induced Diabetic Rats. *Journal of Diabetes & Metabolism*, 06(05).
- Green, D., & Kroemer, G. (2004). The pathophysiology of mitochondrial cell death. *Science*, 305(5684), 626–629.
- Grieve, M. (1994). *A modern herbal* (pp. 547–548.). London: Penguin Books.
- Grossarth-maticek, R., Kiene, H., Baumgartner, S. M., & Ziegler, R. (2001). Use of Iscador, an extract of European mistletoe (*Viscum album*), in cancer treatment: prospective nonrandomized and randomized matched-pair studies nested within a cohort study. *Alternative Therapies*, 7(3), 57–78.

- Grossarth-Maticek, R., & Ziegler, R. (2006). Prospective controlled cohort studies on long-term therapy of breast cancer patients with a mistletoe preparation (Iscador). *Forschende Komplementärmedizin*, *13*(5), 285–292.
- Gutsch, J., Berger, H., Scholz, G., & Denck, H. (1988). Prospective study on radically operated breast cancer with polychemotherapy, HELIXOR and untreated control. *Deutsche Zeitschrift für Onkologie*, *21*(4), 94–101.
- Hagens, C., Giese, T., Staudt, A., Glenz, A., Reinhard-Hennch, B., Loewe-MeschKuehn, A., Strowitzki, T. (2011). Does a treatment with *Viscum album* (Iscador[R] P) in patients with breast cancer influence the expression of the T-cell receptor (TCR)-zeta chains of T-and NK-cells?(Report). *Phytomedicine*, *18*, S24–S24.
- Hajto, T. (1986). Immunomodulatory effects of iscador: a *Viscum album* preparation. *Oncology*, *43*(1), 51–65.
- Hajto, T., Hostanska, K., Fischer, J., & R, S. (1997). Immunomodulatory effects of *Viscum album* agglutinin-I on natural immunity. *Anticancer Drugs*. 1997 Apr;8 Suppl 1:S43-6, 8(Suppl 1), S43–S46.
- Hajto, T., Hostanska, K., Weber, K., Zinke, H., Fischer, J., Mengs, U., Saller, R. (1998). Effect of a recombinant lectin, *Viscum album* agglutinin on the secretion of interleukin-12 in cultured human peripheral blood mononuclear cells and on NK-cell-mediated cytotoxicity of rat splenocytes *in vitro* and *in vivo*. *Natural Immunity*, *16*(1), 34–46.
- Hall, A., Spoerke, D., & Rumack, B. (1986). Assessing mistletoe toxicity. *Annals of emergency medicine*, *15*(11), 1320–1323.
- Halliwell, B. (1994). Free radicals, antioxidants, and human disease: curiosity, cause, or consequence? *Lancet*, *344*, 721–724.
- Halliwell, B. (1996). Antioxidants in human health and disease. *Annual Review of Nutrition*, *16*, 33–50.
- Hanahan, D., & Weinberg, R. (2000). The hallmarks of cancer. *Cell*, *100*(1), 57–70.
- Harmsma, M., Ummelen, M., Dignef, W., Tussenius, K., & Ramaekers, F. (2006). Effects of mistletoe (*Viscum album L.*) extracts Iscador on cell cycle and survival of tumor cells. *Arzneimittelforschung*, *56*(6A), 474–482.
- Harris, J., Morrow, M., & Norton, L. (1997). Malignant tumors of the breast. In V. DeVita, S. Hellman, & S. Rosenberg (Eds.), *Cancer: Principles and Practice of Oncology* (5th editio., pp. 1557–1616). Lippincott-Raven, Philadelphia.

- Harvey, A. L. (2008). Natural products in drug discovery. *Drug discovery today*, 13(19-20), 894–901.
- Heiny, B. M., Albrecht, V., & Beuth, J. (1998). Correlation of immune cell activities and beta-endorphin release in breast carcinoma patients treated with galactose-specific lectin standardized mistletoe extract. *Anticancer research*, 18(1B), 583–586.
- Heiny, B. M., & Beuth, J. (1994). Mistletoe extract standardized for the galactoside-specific lectin (ML-1) induces beta-endorphin release and immunopotential in breast cancer patients. *Anticancer research*, 14(3B), 1339–1342.
- Heiny, B.-M. (1991). Additive therapie mit standardisiertem Mistelextrakt reduziert die Leukopenie und verbessert die Lebensqualität von Patientinnen mit fortgeschrittenem Mammakarzinom unter palliativer Chemotherapie (VEC-Schema). *Krebsmedizin*, 12, 1–14.
- Heinzerling, L., von Baehr, V., Liebenthal, C., von Baehr, R., & Volk, H.-D. (2006). Immunologic Effector Mechanisms of a Standardized Mistletoe Extract on the Function of Human Monocytes and Lymphocytes *In Vitro*, *Ex Vivo*, and *In Vivo*. *Journal of Clinical Immunology*, 26(4), 347–359.
- Hengartner, M. (2000). The biochemistry of apoptosis. *Nature*, 12(407), 770–776.
- Herschkowitz, J. I., Simin, K., Weigman, V. J., Mikaelian, I., Usary, J., Hu, Z., Perou, C. M. (2007). Identification of conserved gene expression features between murine mammary carcinoma models and human breast tumors. *Genome biology*, 8(5), R76.
- Hofmann, D. K., Fitt, W. K., & Fleck, J. (1996). Checkpoints in the life-cycle of *Cassiopea* spp : control of metagenesis and metamorphosis in a tropical jellyfish. *The international journal of developmental biology*, 40(1), 331–338.
- Hojjat, S.-A., Nasrollah, G., & Ali, F. (2006). Cytotoxic Effects of Iranian Mistletoe Extract on a Panel of Cancer Cells. *Iranian Journal of Pharmaceutical Sciences*, 2(3), 157–162.
- Hong, C.-E., & Lyu, S.-Y. (2012). The antimutagenic effect of mistletoe lectin (*Viscum album* L. var. *coloratum* agglutinin). *Phytotherapy research*, 26(5), 787–790.

- Horneber, M., Bueschel, G., Huber, R., Linde, K., & Rostock, M. (2008). Mistletoe therapy in oncology (Review). *Cochrane Database of Systematic Reviews*, (2).
- Hou, X., Yuan, X., Zhang, B., Wang, S., & Chen, Q. (2013). Screening active anti-breast cancer compounds from Cortex Magnolia officinalis by 2D LC-MS. *Journal of separation science*, 36(4), 706–12.
- Hsu, Y., Chen, C., Lin, I., Tsai, E., Kuo, P., & Hou, M. (2012). 4-Shogaol, an Active Constituent of Dietary Ginger, Inhibits Metastasis of MDA-MB-231 Human Breast Adenocarcinoma Cells by Decreasing the Repression of NF- κ B/Snail on RKIP. *Journal of Agricultural and Food Chemistry*, 60, 852–861.
<http://globocan.iarc.fr>.
- Hu, H., Ahn, N.-S., Yang, X., Lee, Y.-S., & Kang, K.-S. (2002). Ganoderma lucidum extract induces cell cycle arrest and apoptosis in MCF-7 human breast cancer cell. *International journal of cancer. Journal international du cancer*, 102(3), 250–3.
- Hu, W., & Kavanagh, J. (2003). Anticancer therapy targeting the apoptotic pathway. *The Lancet Oncology*, 4(12), 721–729.
- Huang, Z.-R., Lin, Y.-K., & Fang, J.-Y. (2009). Biological and pharmacological activities of squalene and related compounds: potential uses in cosmetic dermatology. *Molecules (Basel, Switzerland)*, 14(1), 540–54.
- Hugo, F., Schwitalla, S., Niggemann, B., Zänker, K. S., & Dittmar, T. (2007). *Viscum album* extracts Iscador® P and Iscador® M counteract the growth factor induced effects in human follicular B-NHL cells and breast cancer cells. *MEDICINA*, 67(2), 90–96.
- Hutt, N., Kopferschmitt-Kubler, M., Cabalion, J., Purohit, A., Alt, M., & Pauli, G. (2001). Anaphylactic reactions after therapeutic injection of mistletoe (*Viscum album L.*). *Allergol Immunopathol (Madr)*, 29(5), 201–203.
- Ioana, S., Dumitri, N., & Socaciu, C. (2011). Comparative study about antioxidant activities of *Viscum album* from different host trees, harvested in different seasons. *Journal of Medicinal Plants Research*, 5(11), 2237–2244.
- Isa, N. M., Abdul, A. B., Abdelwahab, S. I., Abdullah, R., Sukari, M. A., Kamalidehghan, B., Mohan, S. (2013). Boesenbergin A, a chalcone from *Boesenbergia rotunda* induces apoptosis via mitochondrial dysregulation and

- cytochrome c release in A549 cells *in vitro*: Involvement of HSP70 and Bcl2/Bax signalling pathways. *Journal of Functional Foods*, 5(1), 87–97.
- Jacob, E. (2009). Natural Products-Based Drug Discovery: Some Bottlenecks and Considerations. *Current Science*, 96(6), 753–754.
- Jacobo-Salcedo, M. R., Alonso-Castro, A., Salazar-Olivo, L., Carranza-Alvarez, C., González-Espíndola, L., Domínguez, F., García-Carrancá, A. (2011). Antimicrobial and cytotoxic effects of Mexican medicinal plants. *Natural product communications*.
- Jäger, S., Winkler, K., Pfüller, U., & Scheffler, A. (2007). Solubility studies of oleanolic acid and betulinic acid in aqueous solutions and plant extracts of *Viscum album* L. *Planta medica*, 73(2), 157–162.
- Jantan, I., Abbas Bukhari, S. N., Mohamed Ali, S. M., Wai, L. K., & Ahmed, M. (2015). The evolving role of natural products from the tropical rainforests as a replenishable source of new drug leads. In *Molecules to medicine*.
- Jemal, A., Bray, F., & Ferlay, J. (2011). Global Cancer Statistics. *A Cancer Journal for Clinicians*, 61(2), 69–90.
- Jie, H., Tao, S., Jun, H., Shuangyang, C., Xiaoqiang, C., & Guolin, Z. (2007). Chemical composition, cytotoxic and antioxidant activity of the leaf essential oil of *Photinia serrulata*. *Food chemistry*, 103(2), 355–358.
- Johansson, S., Gullbo, J., Lindholm, P., Ek, B., Thunberg, E., Samuelsson, G., Claeson, P. (2003). Small, Novel Proteins from the Mistletoe *Phoradendron Tomentosum* Exhibit Highly Selective Cytotoxicity to Human Breast Cancer Cells. *Cellular and molecular life sciences*, 60(1), 165–175.
- Johnstone, R. W., Ruefli, A. A., Lowe, S. W., & Victoria, E. M. (2002). Apoptosis : A Link between Cancer Genetics and Chemotherapy Defects in apoptosis underpin both tumorigenesis and. *Cell*, 108, 153–164.
- Kalidas, M., & Brown, P. (2005). Aromatase inhibitors for the treatment and prevention of breast cancer. *Clinical breast cancer*, 6(1), 27–37.
- Kamesaki, H. (1995). Mechanisms involved in chemotherapy-induced apoptosis and their implications in cancer chemotherapy. *International journal of hematology*, 68(1), 29–43.
- Kang, M. R., Kim, H. M., Kang, J. S., Lee, K., Lee, S. D., Hyun, D.-H., Kim, D. C. (2011). Lipid-soluble ginseng extract induces apoptosis and G0/G1 cell cycle

- arrest in NCI-H460 human lung cancer cells. *Plant foods for human nutrition (Dordrecht, Netherlands)*, 66(2), 101–6.
- Kang, Y., Park, H. J., Chung, H., Min, H., Park, E. J., Lee, M. A., Lee, S. K. (2012). Wnt /B -Catenin Signaling Mediates the Antitumor Activity of Magnolol in Colorectal Cancer Cells . *Molecular pharmacology*, 82, 168–177.
- Karen K, S., Mary J, N., Carolyn, C., Lindsey, S., Martin, W. L., & Todd M, T. (2002). Comparison of Side Effects Between Sentinel Lymph Node and Axillary Lymph Node Dissection for Breast Cancer. *Annals of Surgical Oncology*, 9(8), 745–753.
- Katsarou, A., Rhizopoulou, S., & Kefalas, P. (2012a). Antioxidant Potential of the Aerial Tissues of the Mistletoe *Loranthus europaeus* Jacq . *Records of Natural Products*, 6(4), 394–397.
- Katsarou, A., Rhizopoulou, S., & Kefalas, P. (2012b). Antioxidant Potential of the Aerial Tissues of the Mistletoe *Loranthus europaeus* Jacq. *Records of Natural Products, Vol. 6* (4), 394.
- Kelter, G., Schierholz, J. M., Fischer, I. U., & Fiebig, H.-H. (2007). Cytotoxic Activity and Absence of Tumor Growth Stimulation of Standardized Mistletoe Extracts in Human Tumor Models *In Vitro*. *Anticancer Research*, 27(1A), 223–233.
- Kienle, G S, Berrino, F., Büssing, A., Portalupi, E., Rosenzweig, S., & Kiene, H. (2003). Mistletoe in cancer-a systematic review on controlled clinical trials. *European journal of medical research*, 8(3), 109–119.
- Kienle, G S, & Kiene, H. (2007). Complementary cancer therapy: A systematic review of prospective clinical trials on anthroposophic mistletoe extracts. *European Journal of Medical Research*, 12, 103–119.
- Kienle, G., & Kiene, H. (2003). *Die Mistel in der Onkologie-Fakten und konzeptionelle Grundlagen*. Stuttgart, New York: Schattauer Verlag.
- Kienle, Gunver S, Glockmann, A., Schink, M., & Kiene, H. (2009). *Viscum album L.* extracts in breast and gynaecological cancers: a systematic review of clinical and preclinical research. *Journal of experimental & clinical cancer research*, 28, 79.
- Kienle, Gunver S, & Kiene, H. (2010). Influence of *Viscum album L* (European mistletoe) extracts on quality of life in cancer patients: a systematic review of controlled clinical studies. *Integrative cancer therapies*, 9(2), 142–157.

- Kischkel, F. C., Hellbardt, S., Behrmann, I., Germer, M., Pawlita, M., Krammer, P. H., & Peter, M. E. (1995). Cytotoxicity-dependent APO-1 (Fas/CD95) - associated proteins form a death-inducing signaling complex (DISC) with the receptor. *EMBO journal*, *14*(22), 5579–5588.
- Klett, C., & Anderer, F. (1989). Activation of natural killer cell cytotoxicity of human blood monocytes by a low molecular weight component from *viscum album* extract. *Arzneimittelforschung*, *39*(12), 1580–1585.
- Knöpfel-Sidler, F., Viviani, A., Rist, L., & Hensel, A. (2005). Human cancer cells exhibit *in vitro* individual receptiveness towards different mistletoe extracts. *Pharmazie*, *60*(6), 448–454.
- Koehn, F. E., & Carter, G. T. (2005). The evolving role of natural products in drug discovery. *nature reviews drug discovery*, *4*, 206–220.
- Kohno, Y., Egawa, Y., Itoh, S., Nagaoka, S., Takahashi, M., & Mukai, K. (1995). Kinetic study of quenching reaction of singlet oxygen and scavenging reaction of free radical by squalene in n-butanol. *Biochim Biophys Acta*. 1995 Apr 28; *1256*(1):52-6., *1256*(1), 52–56.
- Koizumi, T., Shirakura, H., Kumagai, H., Tatsumoto, H., & Suzuki, K. (1996). Mechanism of cadmium-induced cytotoxicity in rat hepatocytes: cadmium-induced active oxygen-related permeability changes of the plasma membrane. *Toxicology*, *114*(2), 125–134.
- Konrad, U., & Renatus, Z. (2006). *Documentation of published clinical trials and observational studies with Iscador®*. (U. Konrad & Z. Renatus, Eds). Arlesheim: Verein für Krebsforschung.
- Kovacs, E. (2000). Serum levels of IL-12 and the production of IFN-gamma , IL-2 and IL-4 by peripheral blood mononuclear cells (PBMC) in cancer patients treated with *Viscum album* extract. *Biomed Pharmacother*, *54*, 305–310.
- Kroemer, G., Galluzzi, L., & Brenner, C. (2007). Mitochondrial Membrane Permeabilization in Cell Death. *Physiological reviews*, *87*(1), 99–163.
- Krohn, R. (2002). The colorimetric detection and quantitation of total protein. *Current protocol in cell biology*, Appendix 3:Appendix 3H.
- Kröz, M., Schad, F., Matthes, B., Pickartz, H., & Girke, M. (2002). Blut- und Gewebseosinophilie, Mistlektin-Antikörper und Lebensqualität bei einer Mammakarzinom-Patientin unter intratumoraler und subkutaner Misteltherapie. *Forsch Komplementärmed Klass Naturheilkd*, *9*, 160–167.

- Kuete, V., Wiench, B., Hegazy, M., Mohamed, T., Fankam, A., Shahat, A., & Efferth, T. (2012). Antibacterial activity and cytotoxicity of selected Egyptian medicinal plants. *Planta Medica*, 78(2), 193–199.
- Kumar, J., Dhar, P., Tayade, A. B., Gupta, D., Chaurasia, O. P., Upreti, D. K., Srivastava, R. B. (2014). Antioxidant Capacities, Phenolic Profile and Cytotoxic Effects of Saxicolous Lichens from Trans-Himalayan Cold Desert of Ladakh. (G. Pintus, Ed.) *PLoS ONE*, 9(6), e98696.
- Kuttan, G., & Kuttan, R. (1993). Reduction of leukopenia in mice by “*viscum album*” administration during radiation and chemotherapy. *Tumori*, 79(1), 74–76.
- Laemmli, U. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227(5259), 680–685.
- Lam, K. S. (2007). New aspects of natural products in drug discovery. *Trends in Microbiology*, 15(6), 279–289.
- Lebel, C. P., Ischiropoulos, H., & Bondys, S. C. (1992). Evaluation of the probe 2',7'-dichlorofluorescein as an indicator of reactive oxygen species formation and oxidative stress. *Chemical research in toxicology*, 5, 227–231.
- Lee, C.-H., Kim, J.-K., Kim, H.-Y., Park, S.-M., & Lee, S.-M. (2009). Immunomodulating effects of Korean mistletoe lectin *in vitro* and *in vivo*. *International immunopharmacology*, 9(13-14), 1555–1561.
- Lev, E., Ephraim, M., & Ben-Arye, E. (2011). European and Oriental mistletoe: From mythology to contemporary integrative cancer care. *European Journal of Integrative Medicine*, 3(3), e133–e137.
- Levine, A. J. (1997). p53 , the Cellular Gatekeeper for Growth and Division. *Cell*, 88(3), 323–331.
- Li, J. W.-H., & Vederas, J. C. (2009). Drug discovery and natural products: end of an era or an endless frontier? *Science*, 325, 161–165.
- Liang, C.-C., Park, A. Y., & Guan, J.-L. (2007). *In Vitro* Scratch Assay: A Convenient and Inexpensive Method for Analysis of Cell Migration *In Vitro*. *Nature protocols*, 2(2), 329–33.
- Liao, G., Apaya, M. K., & Shyur, L. (2013). Herbal Medicine and Acupuncture for Breast Cancer Palliative Care and Adjuvant Therapy. *Evidence-based complementary and alternative medicine*, 2013.
- Lim Chin Chye, G., Rampal, S., & Yahaya, H. (2008). *Cancer Incidence in Peninsular Malaysia 2003-2005*. Kuala Lumpur: National Cancer Registry.

- Lin, K.-L., Tsai, P.-C., Hsieh, C.-Y., Chang, L.-S., & Lin, S.-R. (2011). Antimetastatic effect and mechanism of ovatodiolide in MDA-MB-231 human breast cancer cells. *Chemico-biological interactions*, *194*(2-3), 148–58.
- Linell, F., & Ljungberg, O. (1984). *Atlas of breast pathology*. Copenhagen, Munksgaard.
- Liu, C., Hung, M., Wang, D., Chu, P., Su, J., Teng, T., Tseng, L. (2014). Tamoxifen induces apoptosis through cancerous inhibitor of protein phosphatase 2A–dependent phospho-Akt inactivation in estrogen receptor–negative human breast cancer cells. *Breast Cancer Research*, *16*, 1–15.
- Locksley, R., Killeen, N., & Lenardo, M. (2001). The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell*, *104*(4), 487–501.
- Loewe-Mesch, A., Kuehn, J. J., Borho, K., Abel, U., Bauer, C., Gerhard, I., v Hagens, C. (2008). Adjuvant simultaneous mistletoe chemotherapy in breast cancer--influence on immunological parameters, quality of life and tolerability. *Forschende Komplementärmedizin*, *15*(1), 22–30.
- Lohézic-Le Dévéhat, F., Tomasi, S., Fontanel, D., & Boustie, J. (2002). Flavonols from *Scurrula Ferruginea* Danser (Loranthaceae). *Z. Naturforsch*, *57 C*, 1092–1095.
- MacLachlan, T., Sang, N., & Giordano, A. (1995). Cyclins, cyclin-dependent kinases and cdk inhibitors: implications in cell cycle control and cancer. *Critical Reviews in Eukaryotic Gene Expression*, *5*(2), 127–156.
- Maier, G., & Fiebig, H.-H. (2002). Absence of Tumor Growth Stimulation in a Panel of 16 Human Tumor Cell Lines by Mistletoe Extracts *In Vitro*. *Anti-Cancer Drugs*, *13*(4), 373–379.
- Maletzki, C., Linnebacher, M., Savai, R., & Hobohm, U. (2013). Mistletoe lectin has a shiga toxin-like structure and should be combined with other Toll-like receptor ligands in cancer therapy. *Cancer immunology, immunotherapy*, *62*(8), 1283–1292.
- Mantena, S. K., Sharma, S. D., & Katiyar, S. K. (2006). Berberine, a natural product, induces G1-phase cell cycle arrest and caspase-3-dependent apoptosis in human prostate carcinoma cells. *Molecular cancer therapeutics*, *5*(2), 296–308.

- Martin, E. J., & Forkert, P. (2004). Evidence That 1,1-Dichloroethylene Induces Apoptotic Cell Death in Murine Liver. *The journal of pharmacology and experimental therapeutics*, 310(1), 33–42.
- Massaoka, M. H., Matsuo, A. L., Figueiredo, C. R., Farias, C. F., Girola, N., Arruda, D. C., Travassos, L. R. (2012). Jacaranone Induces Apoptosis in Melanoma Cells via ROS-Mediated Downregulation of Akt and p38 MAPK Activation and Displays Antitumor Activity *In Vivo*. (S.V.Pizzo, Ed.) *PLoS ONE*, 7(6), e38698.
- Mcguire, M., Li, R., Wang, R., & Yu, F. (2005). Isolation and characterization of methyl esters and derivatives from *Euphorbia kansui* and their inhibitory effects on the human. *Journal of pharmacy and pharmaceutical sciences*, 8(3), 528–535.
- Melzer, J., Iten, F., Hostanska, K., & Saller, R. (2009). Efficacy and safety of mistletoe preparations (*Viscum album*) for patients with cancer diseases. A systematic review. *Forschende Komplementärmedizin (2006)*, 16(4), 217–226.
- Merril, C. (1990). Gel-staining techniques. *Methods in enzymology*, 182, 477–488.
- Mihara, M., Erster, S., Zaika, A., Petrenko, O., Chittenden, T., Pancoska, P., Street, S. (2003). p53 has a direct apoptogenic role at the mitochondria. *Molecular cell*, 11, 577–590.
- Moore, G., Gerner, R., & Franklin, H. (1967). Culture of normal human leukocytes. *JAMA*, 199(8), 519–524.
- Morris, C. (2007). Cryopreservation of animal and human cell lines. *Methods in molecular biology*, 368, 227–236.
- Morrow, M., & Schnitt, S. J. (1996). In situ carcinomas. In J. . Harris, M. . Lippman, M. Morrow, & S. Hellman (Eds.), *Diseases of the breast*. Philadelphia: Lippincott-Raven.
- Mueller, E. A., & Anderer, F. A. (1990). A *Viscum album* oligosaccharide activating human natural cytotoxicity is an interferon γ inducer. *Cancer Immunol Immunother*, 32, 221–227.
- Nahta, R., Yu, D., Hung, M.-C., Hortobagyi, G. N., & Esteva, F. J. (2006). Mechanisms of disease: understanding resistance to HER2-targeted therapy in human breast cancer. *Nature clinical practice. Oncology*, 3(5), 269–80.

- Newman, D. J., & Cragg, G. M. (2007). Natural Products as Sources of New Drugs over the Last 25 Years. *Journal of Natural Products*, *70*, 461–477.
- Newman, D. J., Cragg, G. M., & Snader, K. M. (2003). Natural Products as Sources of New Drugs over the Period 1981-2002. *Journal of Natural Products*, *66*, 1022–1037.
- Nielsen, T. O., Hsu, F. D., Jensen, K., Cheang, M., Karaca, G., Hu, Z., Perou, C. M. (2004). Immunohistochemical and Clinical Characterization of the Basal-Like Subtype of Invasive Breast Carcinoma. *Clinical cancer research*, *10*(919), 5367–5374.
- Nik Aina Syazana Nik, Z., & Mohd Dasuki Sul, A. (2015). Phytochemical Analysis, Toxicity and Cytotoxicity Evaluation of Dendrophtoe Pentandra Leaves Extracts. *International journal of applied biology and pharmaceutical technology*, *6*(1), 108–117.
- O'Neill, M., & Lewis, J. (1993). Human Medicinal Agents from Plants. In A. Kinghorn & M. Balandrin (Eds.), *ACS Symposium Series 534* (p.48). Washington.
- Oluwaseun, A. A., & Ganiyu, O. (2008). Antioxidant Properties of Methanolic Extracts of Mistletoes (*Viscum album*) from Cocoa and Cashew Trees in Nigeria. *African Journal of Biotechnology*, *7*(17), 3138–3142.
- Onay-Uçar, E., Karagöz, A., & Arda, N. (2006). Antioxidant activity of *Viscum album* ssp. *Fitoterapia*, *77*(7-8), 556–60.
- Orhan, D. D., Küpeli, E., Yesilada, E., & Ergun, F. (2006). Anti-inflammatory and antinociceptive activity of flavonoids isolated from *Viscum album* ssp. *Zeitschrift für Naturforschung. C, Journal of biosciences*, *61*(1-2), 26–30.
- Osadebe, P. O., Okide, G. B., & Akabogu, I. C. (2004). Study on anti-diabetic activities of crude methanolic extracts of *Loranthus micranthus* (Linn.) sourced from five different host trees. *Journal of Ethnopharmacology*, *95*(2-3), 133–138.
- Osadebe, P., Omeje, E., Kawamura, A., & Okoye, F. (2010). Lupeol derivatives from the Eastern Nigeria mistletoe, *Loranthus micranthus* Linn. (Loranthaceae) with enhanced cell proliferative potentials. *Planta Medica*, *91*.
- Ostermann, T., & Büssing, A. (2012). Retrolective studies on the survival of cancer patients treated with mistletoe extracts: a meta-analysis. *Explore (New York, N.Y.)*, *8*(5), 277–281.

- Ostermann, T., Raak, C., & Büssing, A. (2009). Survival of cancer patients treated with mistletoe extract (Iscador): a systematic literature review. *BMC cancer*, 9, 451.
- Ozer, H., Armitage, J. O., Bennett, C. L., Crawford, J., Demetri, G. D., Pizzo, P. A., Winn, R. J. (2000). Update of recommendations for the use of hematopoietic colony-stimulating factors: evidence-based, clinical practice guidelines. *Journal of Clinical Oncology*, 18(20), 3558–3585.
- Pae, H. O., Seo, W. G., Oh, G. S., Shin, M. K., Lee, H. S., Kim, S. B., & Chung, H. T. (2000). Potentiation of tumor necrosis factor-alpha-induced apoptosis by mistletoe lectin. *Immunopharmacology and immunotoxicology*, 22(4), 697–709.
- Pan, L., Chai, H.-B., & Kinghorn, A. D. (2013). Discovery of new anticancer agents from higher plants. *Front Biosci (Schol Ed)*, 4, 142–156.
- Patel, S., & Panda, S. (2013). Emerging roles of mistletoes in malignancy management. *3 Biotech*, 4(1), 13–20.
- Perou, C. M., Sùrlie, T., Eisen, M. B., Rijn, M. Van De, Jeffrey, S. S., Rees, C. A., Grant, S. (2000). Molecular portraits of human breast tumours. *Nature*, 533(May), 747–752.
- Perry, L. . (1978). *Medicinal plants of east and southeast Asia: Attributed properties and uses* (p. 2138). Cambridge, Massachusetts, USA: MIT press.
- Peto, R., Boreham, J., Clarke, M., Davies, C., & Beral, V. (2000). UK and USA breast cancer deaths down 25% in year 2000 at ages 20-69 years. *Lancet*, 355, 1822.
- Phang, C.-W., Malek, S. N. A., & Ibrahim, H. (2013). Antioxidant potential, cytotoxic activity and total phenolic content of *Alpinia pahangensis* rhizomes. *BMC complementary and alternative medicine*, 13(1), 243.
- Piao, B. K., Wang, Y. X., Xie, G. R., Mansmann, U., Matthes, H., Beuth, J., & Lin, H. S. (2004). Impact of complementary mistletoe extract treatment on quality of life in breast, ovarian and non-small cell lung cancer patients. A prospective randomized controlled clinical trial. *Anticancer research*, 24(1), 303–309.
- Pieme, A., Jeanne, N., & Marietta, C. (2012). *In Vitro* Antiproliferative and Anti-Oxidant Activities of Methanol Extracts of *Urena Lobata* and *Viscum Album*

- Against Breast Cancer Cell Lines. *Toxicological & Environmental Chemistry*, 94(5), 987–999.
- Poli, G., Leonarduzzi, G., Biasi, F., & Chiarpotto, E. (2004). Oxidative stress and cell signalling. *Current Medicinal Chemistry*, 11, 1163–1182.
- Pontiki, E., Hadjipavlou-Litina, D., Litinas, K., & Geromichalos, G. (2014). Novel cinnamic acid derivatives as antioxidant and anticancer agents: design, synthesis and modeling studies. *Molecules (Basel, Switzerland)*, 19(7), 9655–74.
- Poulsen, H., Prieme, H., & Loft, S. (1998). Role of oxidative DNA damage in cancer initiation and promotion. *European Journal of Cancer Prevention*, 7, 9–16.
- Prasad, K. N., & Edwards-prasad, J. (1982). Effects of Tocopherol (Vitamin E) Acid Succinate on Morphological Alterations and Growth Inhibition in Melanoma Cells in Culture1. *Cancer Research*, 42(February), 550–555.
- Prat, A., Parker, J. S., Karginova, O., Fan, C., Livasy, C., Herschkowitz, J. I., Perou, C. M. (2010). Phenotypic and molecular characterization of the claudin-low intrinsic subtype of breast cancer. *Breast Cancer Research*, 12(R68).
- Pryme, I. F., Bardocz, S., Pusztai, A., & Ewen, S. W. B. (2006). Suppression of growth of tumour cell lines *in vitro* and tumours *in vivo* by mistletoe lectins. *Histology and histopathology*, 21(3), 285–299.
- Pucci, B., Kasten, M., & Giordano, A. (2000). Cell Cycle and Apoptosis. *Neoplasia*, 2(4), 291–299.
- Rafehi, H., Orłowski, C., Georgiadis, G. T., Ververis, K., El-Osta, A., & Karagiannis, T. C. (2011). Clonogenic assay: adherent cells. *Journal of visualized experiments : JoVE*, (49), 15–17.
- Ramaekers, F., Harmsma, M., Karel, T., Schutte, B., Werner, M., & Ramos, M. (2007). Mistletoe Extracts (*Viscum Album L*) Iscador® Interact with the Cell Cycle Machinery and Target Survival Mechanisms in Cancer Cells. *Medicina*, 67(2), 79–84.
- Rani, M. J., & Chandramohan, N. (2013). Identification of Sesquiterpenes from Lantana Camara Leaves. *International journal of drug development and research*, 5(1), 179–184.
- Re, R., Pellegrini, N., Proteggente, A., Ananth, P., Yang, M., & Rice-Evan, C. (1999). Antioxidant Activity Applying an Improved ABTS Radical Cation Decolorization Assay. *Free Radical Biology and Medicine*, 26, 1231–1237.

- Reed, J. (2000). Apoptosis-based therapies. *nature reviews drug discovery*, 1(2), 111–121.
- Ridner, S. H., Dietrich, M. S., & Kidd, N. (2011). Breast cancer treatment-related lymphedema self-care: education, practices, symptoms, and quality of life. *Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer*, 19(5), 631–7.
- Saelens, X., Festjens, N., Vande Walle, L., van Gorp, M., van Loo, G., & Vandenabeele, P. (2004). Toxic proteins released from mitochondria in cell death. *Oncogene*, 23(16), 2861–74.
- Sahaya Sathish, S., Janakiraman, N., & Johnson, M. (2012). Phytochemical Analysis of *Vitex altissima L.* using UV-VIS and FTIR. *International Journal of Pharmaceutical Sciences and Drug Research*, 4(1), 56–62.
- Sahreem, S., Khan, M. R., & Khan, R. A. (2010). Evaluation of antioxidant activities of various solvent extracts of *Carissa opaca* fruits. *Food Chemistry*, 122(4), 1205–1211.
- Sakarkar, D. M., & Deshmukh, V. N. (2011). Ethnopharmacological Review of Traditional Medicinal Plants for Anticancer Activity. *International Journal of PharmTech Research*, 3(1), 298–308.
- Sanli, T., Steinberg, G. R., Singh, G., & Tsakiridis, T. (2014). AMP-activated protein kinase (AMPK) beyond metabolism A novel genomic stress sensor participating in the DNA damage response pathway. *Cancer biology and therapy*, 15(2), 156–169.
- Saxe, T. (1987). Toxicity of medicinal herbal preparations. *Am Fam Physician*, 35, 135–142.
- Schaller, G., & Urech, K. (1996). Cytotoxicity of Different Viscotoxins and Extracts from the European Subspecies of *Viscum album L.* *Phytotherapy research*, 10, 473–477.
- Schumacher, K., Schneider, B., Reich, G., Stiefel, T., Stoll, G., Bock, P., Beuth, J. (2003). Influence of postoperative complementary treatment with lectin-standardized mistletoe extract on breast cancer patients. *Anticancer research*, 26(6D), 5081–5088.
- Schwartz, J., & Shklar, G. (1992). The selective cytotoxic effect of carotenoids and alpha-tocopherol on human cancer cell lines *in vitro*. *Journal of oral and maxillofacial surgery*, 50(4), 367–373.

- Scudiere, D. A., Shoemaker, R. H., Paul, K. D., Monks, A., Tierney, S., Nofziger, T. H., Boyd, M. R. (1988). Evaluation of a Soluble Tetrazolium/Formazan Assay for Cell Growth and Drug Sensitivity in Culture Using Human and Other Tumor Cell Lines. *Cancer Research*, 48, 4827–4833.
- Semiglasov, V. F., Stepula, V. V, Dudov, A., Lehmacher, W., & Mengs, U. (2004). The standardised mistletoe extract PS76A2 improves QoL in patients with breast cancer receiving adjuvant CMF chemotherapy: a randomised, placebo-controlled, double-blind, multicentre clinical trial. *Anticancer research*, 24(2C), 1293–1302.
- Semiglasov, V., Stepula, V., Dudov, A., Schnitker, J., & Mengs, U. (2006). Quality of life is improved in breast cancer patients by Standardised Mistletoe Extract PS76A2 during chemotherapy and follow-up: a randomised, placebo-controlled,. *Anticancer research*, 26, 1519–1529.
- Sermakkani, M., & Thangapandian, V. (2012). GC-MS Analysis of Cassia Italica Leaf Methanol Extract. *Asian Journal of Pharmaceutical and Clinical Research*, 5(2), 90–94.
- Shall, S., & DeMurcia, G. (2000). Poly(ADP-ribose) polymerase-1: what have we learned from the deficient mouse model? *Mutation Research*, 460(1), 1–15.
- Sharma, P. (2011). Cinnamic acid derivatives: A new chapter of various pharmacological activities. *Journal of Chemical and Pharmaceutical Research*, 3(2), 403–423.
- Shim, H., Park, J., Paik, H., Nah, S., Kim, D. S. H. L., & Han, Y. S. (2007). Acacetin-induced Apoptosis of Human Breast Cancer MCF-7 Cells Involves Caspase Cascade , Mitochondria-mediated Death Signaling and SAPK/JNK1/2-c-Jun Activation. *Molecules and cells*, 24(1), 95–104.
- Shoeb, M. (2008). Anticancer agents from medicinal plants. *Bangladesh Journal of Pharmacology*, 1(2).
- Sigounas, G., Anagnostou, a, & Steiner, M. (1997). D1-Alpha-Tocopherol Induces Apoptosis in Erythroleukemia, Prostate, and Breast Cancer Cells. *Nutrition and cancer*, 28(1), 30–5.
- Simona, V., Rugină, D., & Socaciu, C. (2008). Antioxidant activities of *Viscum album*'s leaves from various host trees. *Bulletin UASVM, Agriculture*, 65(1), 327–332.

- Singh, R., Avliyakov, N. K., Braga, M., Haykinson, M. J., Martinez, L., Singh, V., Pervin, S. (2013). Proteomic identification of mitochondrial targets of arginase in human breast cancer. *Plos one*, 8(11), e79242.
- Singleton, V. L., & Rossi, A. J. (1965). Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents. *American Journal of Enology and Viticulture*, 16, 144–158.
- Sjövall, K., Strömbeck, G., Löfgren, A., Bendahl, P.-O., & Gunnars, B. (2010). Adjuvant radiotherapy of women with breast cancer-information, support and side-effects. *European journal of oncology nursing : the official journal of European Oncology Nursing Society*, 14(2), 147–153.
- Smith, P., Krohn, R., Hermanson, G., Mallia, A., Gartner, F., Provenzano, M., Klenk, D. (1985). Measurement of protein using bicinchoninic acid. *Analytical biochemistry*, 150(1), 76–85.
- Smith, R. (2006). Breast cancer in limited-resource countries: early detection and access to care. *Breast Journal*, 12(Suppl. 1), S16–S26.
- Smith, T. (2000). Squalene: potential chemopreventive agent. *Expert Opinion on Investigational Drugs*, 9(8), 1841–1848.
- Son, G. S., Ryu, W. S., Kim, H. Y., Woo, S. U., Park, K. H., & Bae, J. W. (2010). Immunologic response to mistletoe extract (*Viscum album* L.) after conventional treatment in patients with operable breast cancer. *Journal of Breast Cancer*, 13(1), 14–18.
- Sova, M. (2012). Antioxidant and antimicrobial activities of cinnamic acid derivatives. *Mini reviews in medicinal chemistry*, 12(8), 749–767.
- Spratt, J. ., & Tobin, G. . (1995). *Gross anatomy of the breast*. Philadelphia: W.B. Saunders.
- Stacey Ricci, M., & Zong, W. (2006). Chemotherapeutic Approaches for Targeting Cell Death Pathways. *Oncologist*, 11(4), 342–357.
- Stauder, H., & Kreuser, E. (2002). Mistletoe extracts standardised in terms of mistletoe lectins (ML I) in oncology: current state of clinical research. *Onkologie*, 25(4), 374–380.
- Steeg, P. S. (2006). Tumor metastasis: mechanistic insights and clinical challenges. *Nature Medicine*, 12, 895–904.
- Steiner, R. (1961). Vortrag Vom 2. 4. 1920; 312 (Vol. 312, pp. 242–262.). Dornach.

- Stewart, B. (1994). Mechanisms of apoptosis: integration of genetic, biochemical, and cellular indicators. *Journal of the National Cancer Institute*, 86(17), 1286–1296.
- Storz, P. (2005). Reactive oxygen species in tumor progression. *Frontiers in Bioscience. a journal and virtual library*, 1(10), 1881–1896.
- Sulaiman, G. M., & Hussien, N. (2013). Phenolic content, antioxidant, antimicrobial and cytotoxic activities of ethanolic extract of *Salix Alba*. *American Journal of Biochemistry and Biotechnology*, 9(1), 41–46.
- Suzuki, M., Youle, R., & Tjandra, N. (2000). Structure of Bax: coregulation of dimer formation and intracellular localization. *Cell*, 103(4), 645–654.
- Suzuki, Y., Imai, Y., Nakayama, H., Takahashi, K., Takio, K., & Takahashi, R. (2001). A serine protease, HtrA2, is released from the mitochondria and interacts with XIAP, inducing cell death. *Molecular cell*, 8(3), 613–621.
- Szatrowski, T. P., & Nathan, C. F. (1991). Production of Large Amounts of Hydrogen Peroxide by Human Tumor Cells¹. *Cancer research*, 51(3), 794–799.
- Tabiasco, J., Pont, F., Fournié, J.-J., & Vercellone, A. (2002). Mistletoe viscotoxins increase natural killer cell-mediated cytotoxicity. *European Journal of Biochemistry*, 269(10), 2591–2600.
- Tatyana, Ç., Slavica, Ç., & Mira popovi, Ç. (2008). Cytotoxic Effects of the *Viscum album L* . Extract on Ehrlich Tumour Cells *In Vivo*. *Phytoterapy Research*, 1103(June), 1097–1103.
- Tian, L., Yin, D., Ren, Y. E., Gong, C., Chen, A., & Guo, F. (2012). Plumbagin induces apoptosis via the p53 pathway and generation of reactive oxygen species in human osteosarcoma cells. *Molecular medicine reports*, 126–132.
- Tong, W., Cortes, U., & Wang, Z. (2001). Poly(ADP-ribose) polymerase: a guardian angel protecting the genome and suppressing tumorigenesis. *Biochimica et biophysica acta*, 1552(1), 27–37.
- Troger, W. (2011). Connection between quality of life and neutropenia in breast cancer patients who were solely treated with chemotherapy or additionally with mistletoe therapy: Results of a randomized study [Zusammenhang von Lebensqualitt und Neutropenie bei Brustkrebspati. *Deutsche Zeitschrift fur Onkologie*, 43, 58–67.

- Tröger, W., Jezdić, S., Ždrale, Z., Tišma, N., & Hamre, H. J. (2009). Quality of life and neutropenia in patients with early stage breast cancer : A randomized pilot study comparing additional treatment with mistletoe extract to chemotherapy alone. *Breast Cancer : Basic and Clinical Research*, 3, 35–45.
- Tröger, W., Zdrale, Z., Stanković, N., & Matijašević, M. (2012). Five-year follow-up of patients with early stage breast cancer after a randomized study comparing additional treatment with *viscum album* (L.) extract to chemotherapy alone. *Breast cancer : basic and clinical research*, 6, 173–180.
- Tröger, W., Zdrale, Z., Tišma, N., & Matijašević, M. (2014). Additional Therapy with a Mistletoe Product during Adjuvant Chemotherapy of Breast Cancer Patients Improves Quality of Life: An Open Randomized Clinical Pilot Trial. *Evidence-based complementary and alternative medicine*, 2014.
- Uçar, E. Ö., Karagöz, A., & Arda, N. (2006). Antioxidant Activity of *Viscum Album ssp. Album*. *Fitoterapia*, 77(July 2001), 556–560.
- Uma, B., & Parvathavarthini, R. (2010). Antibacterial Effect of Hexane Extract of Sea Urchin , *Temnopleurus alexandri*. *International Journal of PharmTech Research*, 2(3), 1677–1680.
- Van Cruchten, S., & Van Den Broeck, W. (2002). Morphological and Biochemical Aspects of Apoptosis, Oncosis and Necrosis. *Anatomia, Histologia, Embryologia*, 223, 214–223.
- Vardy, J., Dhillon, H. M., Clarke, S. J., Olesen, I., Leslie, F., Warby, A., Mclachlan, A. J. (2013). Investigation of herb-drug interactions with ginkgo biloba in women receiving hormonal treatment for early breast cancer. *Springerplus*, 2(Schwabe 2004), 1–5.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant Activity and Total Phenolics in Selected Fruits, Vegetables, and Grain Products. *Journal of Agricultural and Food Chemistry*, 46, 4113–4117.
- Vicaș, S. I., Rugină, D., Leopold, L., Pinte, A., & Socaciu, C. (2011). HPLC Fingerprint of Bioactive Compounds and Antioxidant Activities of *Viscum album* from Different Host Trees. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 39(1), 48–57.
- Vicas, S. I., Rugină, D., & Socaciu, C. (2011). Comparative Study about Antioxidant Activities of *Viscum Album* from Different Host Trees, Harvested in Different Seasons. *Journal of medicinal plants research*, 5(11), 2237–2244.

- Vicas, S., Prokisch, J., Rugina, D., & Socaciu, C. (2009). Hydrophilic and Lipophilic Antioxidant Activities of Mistletoe (*Viscum album*) as Determined by FRAP Method. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 37(2), 112–116.
- Vicas, S., Ruginã, D., & Socaciu, C. (2008). Antioxidant Activities of *Viscum Album*'s Leaves from Various Host Trees. *Bulletin UASVM, Agriculture Cluj-Napoca*, 65(1), 327–332.
- Voelter, W., Wacker, R., Stoeva, S., Tsitsilonis, R., & Betzel, C. (2005). Mistletoe Lectins, Structure and Function. *Frontiers in Natural Product Chemistry*, 1(1), 149–162.
- Wajant, H., Pfizenmaier, K., & Scheurich, P. (2003). Tumor necrosis factor signaling. *Cell death and differentiation*, 10(1), 45–65.
- Walker, J. M. (1994). The Bicinchoninic Acid (BCA) Assay for Protein Quantitation. *Methods in molecular biology*, 32, 5–8.
- Warren, S. (2001). Trypan blue exclusion test of cell viability. *Current protocol in immunology*, 2001, Appendix 3: Appendix 3B., Appendix 3: Appendix 3B.
- Werner, M., Bock, P. R., Hanisch, J., & Stauder, G. (2011). Supportive therapy with mistletoe extract in tumor patients—Results of four controlled pharmacoepidemiological cohort studies as basis for prospective studies (Clinical report). *Phytomedicine*, 18(1), S12–S13.
- Willis, S. N., Chen, L., Dewson, G., Wei, A., Naik, E., Fletcher, J. I., Huang, D. C. S. (2005). Proapoptotic Bak is sequestered by Mcl-1 and Bcl-x L , but not Bcl-2 , until displaced by BH3-only proteins. *Genes and development*, 19(11), 1294–1305.
- Winkler, K., Leneweit, G., & Schubert, R. (2005). Characterization of membrane vesicles in plant extracts. *Colloids and surfaces. B, Biointerfaces*, 45(2), 57–65.
- Wode, K., Schneider, T., Lundberg, I., & Kienle, G. S. (2009). Mistletoe treatment in cancer-related fatigue: a case report. *Cases journal*, 2(1), 77.
- Wollenweber, E., Wieland, A., & Haas, K. (2000). Epicuticular waxes and flavonol aglycones of the European mistletoe, *Viscum album L.* *Z Naturforsch C*, 55(5-6), 314–317.
- Yang, Y., Chen, M., & Sha, C. (2011). Triterpenoids and triterpenoid saponins of *Viscum liquidambaricolum*. *Zhongguo Zhong Yao Za Zhi*, 36(2), 162–165.

- Yao, X. I. N., Panichpisal, K., & Kurtzman, N. (2007). Cisplatin Nephrotoxicity : A Review. *The American Journal of the Medical Sciences*, 334(2), 115–124.
- Yip, C. H., Aishah, N., Taib, M., & Mohamed, I. (2006). Epidemiology of Breast Cancer in Malaysia. *Asian Pacific Journal of Cancer Prevention*, 7, 369–374.
- Yoo, K. M., Lee, C. H., Lee, H., Moon, B., & Lee, C. Y. (2008). Relative Antioxidant and Cytoprotective Activities of Common Herbs. *Food Chemistry*, 106(3), 929–993.
- Yoon, T. J., Yoo, Y. C., Kang, T. B., Shimazaki, K., Song, S. K., Lee, K. H., Kim, J. B. (1999). Lectins isolated from Korean mistletoe (*Viscum album coloratum*) induce apoptosis in tumor cells. *Cancer letters*, 136(1), 33–40.
- Zakaria, Z., Rofiee, M., Mohamed, A., Teh, L., & Salleh, M. (2011). *In vitro* antiproliferative and antioxidant activities and total phenolic contents of the extracts of *Melastoma malabathricum* leaves. *Journal of acupuncture and meridian studies*, 4(4), 248–256.
- Zee-Cheng, R. (1997). Anticancer research on Loranthaceae plants. *Drug Future*, 22, 519–530.
- Zhang, G., Kimijima, I., Onda, M., Kanno, M., Sato, H., Watanabe, T., Abe, R. (1999). Tamoxifen-induced Apoptosis in Breast Cancer Cells Relates to Down-Regulation of bcl-2 , but not bax and bcl-X L , without Alteration of p53 Protein Levels 1. *Clinical Cancer Research*, 5, 2971–2977.
- Zhao, Y., Wang, X., Sun, L., Fan, R., Bi, K., & Yu, Z. (2012). Cytotoxic Constituents of *Viscum Coloratum*. *Zeitschrift fur Naturforschung C*, 67(3-4), 129–134.
- Ziegler, R., & Grossarth-Maticek, R. (2010). Individual patient data meta-analysis of survival and psychosomatic self-regulation from published prospective controlled cohort studies for long-term therapy of breast cancer patients with a mistletoe preparation (Iscador). *Evidence-based complementary and alternative medicine : eCAM*, 7(2), 157–166.
- Zijl F, V. F., Krupitza, G., & Mikulits, W. (2011). Initial steps of metastasis: cell invasion and endothelial transmigration. *Mutation Research*, 728(1-2), 23–34.
- Zorov, D., Juhaszova, M., & Sollott, S. (2006). Mitochondrial ROS-induced ROS release: an update and review. *Biochim Biophys Acta*, 1757(5-6), 509–517.