

DEPOSITION OF THE HYDROXYAPATITE PARTICLES ON THE  
POLYETHER ETHER KETONE SURFACE FOR ORTHOPAEDIC  
APPLICATIONS

DAVOOD ALMASI

A thesis submitted in fulfilment of the  
requirements for the award of the degree of  
Doctor of Philosophy (Mechanical Engineering)

Faculty of Mechanical Engineering  
Universiti Teknologi Malaysia

AUGUST 2015

*I would like to dedicate this thesis to*

*My Parents*

## **AKNOWLEDGEMENT**

At first, thanks to ALLAH, the most gracious and the most merciful, for providing me the opportunity of doctoral study and ability to accomplish this research.

I would like to express my deepest gratitude towards Assoc. Prof. Dr. Izman Sudin who nurtured me to become a researcher. Without his kind guidance, encouragement and valuable advices during the research and writing, this thesis would not be come to the light. I wish also to express my sincere appreciation to my co-supervisor Prof. Dr. Mohammed Rafiq Bin Dato' Abdul Kadir for his generous time and patience to teach me becoming a researcher.

Special gratitude to Dr. Maliheh Sadeghi, Dr. Naznin Sultana, Dr. Nida Iqbal and Dr. Ali Asadi for their kind helps during the research.

## ABSTRACT

Polyether-ether-ketone (PEEK) has a similar elastic modulus close to bone and as a result it can be a suitable alternative material to metallic implants which generally have higher moduli. However, the bio-inertness of PEEK prevents it from integrating well with the surrounding tissues. Many efforts have been made to overcome this problem including the deposition of hydroxyapatite (HA) on PEEK via plasma spraying. Current issues which have arisen with this method are low bonding strength between the substrate and the coating layer, as well as producing a non-uniform density of the coating. In this study, chemical deposition method was used to deposit HA crystalline particles on a treated PEEK substrate without any subsequent sintering process. The surface of PEEK was first sulphonated to create  $-\text{SO}_3\text{H}$  functional group. It was then immersed in suspension of HA in water where the HA particles were chemically connected to the  $-\text{SO}_3\text{H}$  functional group. The treated layer has a 3D porous property but exhibits very low mechanical properties. A compress force was applied on the coated layer for improving its mechanical properties. EDX and XRD results confirmed the existence of crystalline HA on the treated layer. FT-IR analysis was used to confirm the chemical bonding between HA and the substrate. The bioactivity of the HA treated layer was evaluated in terms of wettability. The water contact angle test showed 50% increase in wettability of the treated samples as compared to PEEK. The scratch and nano-indentation tests were carried out on the treated layer to assess its adhesion strength to the substrate and its mechanical strength respectively. The results showed that the compression caused 142% and 36.9% increment of the elasticity modulus and scratch hardness of the treated layer respectively. However, it caused 59% increase in the water contact angle. These findings indicate that the proposed new method is easy to process and does not require special equipment as compared to the Plasma Spray technique. The proposed method can be an alternative technique to change the bio-inertness of PEEK to become more bioactive PEEK which is good for polymeric implant material.

## ABSTRAK

Polieter - eter – keton (PEEK) mempunyai modulus kekenyalan yang hampir sama dengan tulang dan oleh yang demikian, ia sesuai digunakan sebagai satu bahan alternatif kepada implan logam sedia ada yang secara umumnya mempunyai modulus kekenyalan yang lebih tinggi. Namun, sifat bio-lengai PEEK menghalangnya daripada menyatu dengan baik dengan tisu di sekelilingnya. Banyak usaha telah dilakukan untuk mengatasi masalah ini termasuklah melalui pemendapan hidroksiapatit (HA) ke atas PEEK dengan kaedah penyemburan plasma. Isu-isu terkini yang timbul daripada kaedah ini adalah daya ikatan yang lemah di antara substrat dan lapisan salutan serta penghasilan ketumpatan salutan yang tidak seragam. Dalam kajian ini, kaedah mendapan kimia telah digunakan untuk memendapkan zarah hablur HA ke atas substrat PEEK terawat tanpa memerlukan proses pengsinteran selepasnya. Mula-mula sekali permukaan PEEK menjalani proses pengulfonan untuk membentuk kumpulan berfungsi  $-SO_3H$ . Kemudian, ia direndamkan dalam larutan HA di dalam air yang mana zarah-zarah HA bergabung secara kimia dengan kumpulan berfungsi  $-SO_3H$ . Lapisan yang dirawat mempunyai sifat keliangan 3 dimensi tetapi mempamerkan sifat kekuatan mekanikal yang sangat rendah. Satu daya mampatan telah dikenakan ke atas lapisan salutan tersebut untuk memperbaiki sifat mekanikalnya. Keputusan EDX dan XRD mengesahkan kewujudan HA berhablur pada lapisan salutan yang terawat. Analisa FT-IR telah digunakan untuk mengesahkan ikatan kimia di antara HA dan substrat. Keaktifan bio lapisan terawat HA dinilai dari segi sifat kebolehasahannya. Ujian sudut sentuhan air menunjukkan peningkatan sebanyak 50% sifat kebolehasahan bagi sampel terawat berbanding dengan sampel PEEK. Ujian calar dan ujian lekukan nano masing-masing telah dijalankan ke atas lapisan terawat untuk menilai kekuatan lekatannya kepada substrat dan juga menilai kekuatan mekanikalnya. Keputusan menunjukkan bahawa mampatan telah menyebabkan peningkatan sebanyak 142% dan 36.9% kepada modulus kekenyalan dan kekerasan calar masing-masing ke atas lapisan terawat. Walau bagaimana pun, mampatan ini juga menyebabkan peningkatan sebanyak 59% kepada sudut sentuhan air. Penemuan-penemuan ini menunjukkan kaedah baru yang dicadangkan adalah mudah untuk diproses dan tidak memerlukan peralatan khas jika dibandingkan dengan teknik semburan plasma. Kaedah yang dicadangkan ini boleh menjadi satu kaedah alternatif bagi mengubah sifat bio-lengai PEEK kepada sifat yang lebih bioaktif yang mana ia baik bagi bahan implan berasaskan polimer.

## TABLE OF CONTENTS

<b>CHAPTER</b>	<b>TITLE</b>	<b>PAGE</b>
	<b>DECLARATION</b>	ii
	<b>DEDICATION</b>	iii
	<b>AKNOWLEDGEMENT</b>	iv
	<b>ABSTRACT</b>	v
	<b>ABSTRAK</b>	vi
	<b>TABLE OF CONTENTS</b>	vii
	<b>LIST OF TABLES</b>	xii
	<b>LIST OF FIGURES</b>	xiii
	<b>LIST OF ABBREVIATIONS</b>	xvi
	<b>LIST OF APPENDICES</b>	xix
<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
	1.1 Background of the Research	1
	1.2 Problem Statement	3
	1.3 Objectives of the Research	4
	1.4 Scopes of the Research	4
	1.5 Significance of the Research	5
	1.6 Organization of the Thesis	6
<b>2</b>	<b>LITERATURE REVIEW</b>	<b>7</b>

2.1	Introduction	7
2.2	Overview of Biomaterials and their Applications	7
2.2.1	PEEK's Biomedical Applications	10
2.3	Brief Overview of PEEK	12
2.3.1	Physical and Chemical Properties of PEEK	13
2.3.2	Mechanical Properties of PEEK	16
2.3.3	Sterilization of PEEK	20
2.3.4	Imaging Properties of PEEK	21
2.3.5	Biocompatibility of PEEK	22
2.3.6	PEEK Wettability	23
2.4	Bioactive PEEK Implant	25
2.4.1	Surface Modification of PEEK	25
2.4.1.1	Direct Surface Modification	26
2.4.1.2	Surface Coating	29
2.4.2	Bioactive PEEK Composites	33
2.5	Principle of Nanoindentation and Scratch Tests	38
2.5.1	Nanoindentation Test	38
2.5.2	Scratch Test	40
2.6	Summary	43
<b>3</b>	<b>RESEARCH METHODOLOGY</b>	<b>45</b>
3.1	Introduction	45
3.2	Research Design	45
3.3	Material	46
3.4	Sample Preparation	48
3.4.1	Preparation of PEEK Substrates	48
3.4.2	Sulphonation of PEEK Surface	49
3.4.3	Deposition HA Particles on the SPEEK Treated Layer	50
3.4.4	Compression Test on Treated Layer	51
3.5	Characterization	52
3.5.1	Fourier Transform Infrared Spectroscopy (FTIR) Study	52
3.5.2	X-ray Diffraction Study	52
3.5.3	Surface Morphology Study	52

3.5.4	Surface Roughness Study	53
3.5.5	Water Contact Angle Study	53
3.5.6	In vitro Bioactivity Tests Procedure	54
3.5.6.1	Apatite Formation Test	54
3.5.6.2	Cell Attachment Analysis	55
3.6	Mechanical Properties Analysis	57
3.6.1	Nanoindentation Test	57
3.6.2	Scratch Test	58
3.7	Summary	60
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>61</b>
4.1	Introduction	61
4.2	Preliminary Results and Discussion	61
4.2.1	Effect of Sulphonation Time on the Treated Layer Thickness	62
4.2.2	Effect of Immersion Time on the Deposition of HA Particles	62
4.2.3	Summary	64
4.3	Detailed Experiment Results and Discussions	64
4.3.1	Fine Tuning of the Sulphonation Time	64
4.3.2	Results and Analysis of HA Deposited Layer on the Treated PEEK	65
4.3.2.1	FT-IR Analysis for HA Chemical Bonding	66
4.3.2.2	XRD Analysis for HA Crystallinity	68
4.3.2.3	Morphology of the Treated Layer	70
4.3.2.4	Effect of Sulphonation Time on the Treated Layer Properties	77
4.3.2.5	Surface Roughness Study	77
4.3.2.6	Water Contact Angle	79
4.3.2.7	Apatite Formation (Bioactivity) Study	81
4.3.2.8	In-vitro Cell Responses	83
4.3.3	Effect of the Compression Load on the Treated Layer Properties	84



4.3.3.1	Comparison of Surface Morphology of Treated Layer Before and After Compression	85
4.3.3.2	Scratch Test Results and Discussion	86
4.3.3.3	Nanoindentation Results and Discussion	97
4.3.3.4	Surface Roughness	101
4.3.3.5	Water Contact Angle Analysis	103
4.4	Summary of the Findings	104
<b>5</b>	<b>CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE WORK</b>	<b>107</b>
5.1	Introduction	107
5.2	Conclusions	107
5.3	Recommendations for Future Work	108
	<b>REFERENCES</b>	<b>110</b>
	Appendices A-C	129-131

**LIST OF TABLES**

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Typical physical properties of PEEK ("Vitrex PEEK," 1999).	15
2.2	Ultimate tensile strength of some biomaterials	17
2.3	Elastic Modulus of Some Biomaterial (GPa)	19
3.1	Reagents for preparation of SBF	55
4.1	Measured parameters from nanoindentation plot	100

## LIST OF FIGURES

<b>FIGURE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Some of the polymer applications in orthopedic area (Ramakrishna <i>et al.</i> , 2001)	9
2.2	Cross section of human vertebra (Sandukas, 2012).	11
2.3	Chemical structure of PEEK (Kurtz, 2012).	14
2.4	Hydrophilic surface (left), normal hydrophilic surface (middle) and hydrophobic surface (right) (Yuan and Lee, 2013).	24
2.5	(a) surface profile during loading with an indenter (b) schematic of indentation force curve (Oliver and Pharr, 1992).	39
2.6	Schematic of scratch test process	41
3.1	Flow chart of the overall research design	47
3.2	Preparation of the PEEK disc samples involves cutting the PEEK rod, grinding, and ultrasonically cleaning via acetone	48
3.3	Sulphonation of the PEEK disc via immersion in sulphoric acid and followed by immersion in water.	49
3.4	Deposition steps of HA particles on SPEEK via immersion in HA suspension followed by ultrasonic cleaning.	50
3.5	Applying of compressive load on the treated layer.	51

3.6	Hysitron TI 750H Ubi nanomechanical test system	58
3.7	Scratch test equipment (Micro Materials Limited).	59
3.8	Geometric of the conical spherical Rockwell stylus.	59
4.1	Treated layer thickness after being sulphonated at 3 and 20 minutes.	62
4.2	HA deposition amount at varied immersion time.	63
4.3	treated layer thickness after being sulphonated at 5, 10, and 15 minutes.	65
4.4	FT-IR spectra of HA, PEEK, SPEEK targets, and SPEEK/HA treated layer.	67
4.5	Schematic of chemical reaction which lead to deposition of HA particle on PEEK.	68
4.6	XRD spectra of HA, PEEK, and SPEEK targets, and SPEEK/HA treated layer.	70
4.7	FESEM images of the surface of (a) PEEK, (b-e) SPEEK with different magnification, and (f) EDX results of SPEEK top surface.	71
4.8	FESEM images of the top surface of (a-d) SPEEK/HA with different magnification, (e) EDX results of SPEEK/HA top surface, and (f) pore size frequency distribution.	73
4.9	FESEM images of different produced morphology of the treated layer top surface.	74
4.10	Surface morphology of the treated layer with (a) 3, (b) 5 and (c) 10 minutes sulphonation time.	75
4.11	Cross section of the treated layer with different magnification.	76
4.12	Micro roughness due to (a) micro scratch on the surface of the PEEK samples, (b) pits and agglomeration on the surface of the treated layer.	77

4.13	AFM 3D height images of (a) PEEK, and PEEK treated with (b) 3, (c) 5, (d) 10 minutes sulphonation time.	78
4.14	The effect of sulphonation time on the arithmetic mean surface roughness of the treated layer.	79
4.15	The effect of sulphonation time on the water contact angle of treated PEEK.	80
4.16	Images of water droplet on the surface of (a) PEEK and (b) SPEEK/HA (treated layer).	81
4.17	Effect of the surface treatment of the PEEK samples on the apatite formation (bioactivity).	82
4.18	SEM image of the morphology of the attached cell on the (a) PEEK, (b) treated layer	83
4.19	hBMSC proliferation on PEEK and PEEK treated layer – Alamar blue assay.	84
4.20	FESEM images of the top surface of SPEEK/HA (a, b) before and (c, d) after applying the compression load at different magnification.	86
4.21	Penetration depth and normal load versus scratch distance for (a) 3,(b) 5 ,(c) 10 minutes sulphonation time without and (d) 3,(e) 5 ,(f) 10 minutes sulphonation time with applying compression load.	88
4.22	SEM scratch images of the treated layers before applying compression load on (a) 3, (b) 5, and (c) 10 minutes sulphonation time samples.	90
4.23	SEM scratch images of the treated layers after applying compression load, on (a) 3 min, (b) 5 min, (c) 10 min sulphonation time samples.	92
4.24	The effect of the applying compression load on the penetration depth/normal load of the samples with 3 minutes, 5 minutes, and 10 minutes sulphonation time.	94

4.25	Scratch hardness of the treated layer for different sulphonation time, without and with applying compression load.	95
4.26	The effect of compression on horizontal load/scratch distance with (a) 3 , (b) 5, and (c) 10 minutes sulphonation time.	97
4.27	Indentation load versus penetration depth curves	99
4.28	The effect of compression on the Elastic modulus of treated layer with different sulphonation time.	100
4.29	AFM 3D height images of the treated PEEK with (a) 3, (b) 5, and (c) 10 minutes sulphonation time after applying the compression load	102
4.30	The effect of compression load on surface roughness of the treated layer at different sulphonation time.	103
4.31	Water contact angle of the samples (untreated and treated PEEK) at different sulphonation times before and after being compressed.	104

## LIST OF ABBREVIATIONS

Ac	-	projected contact area between the sample and indenter
ALP	-	Alkaline phosphatase
ANAB	-	Accelerated Neutral Atom Beam
ASTM	-	American Society for Testing and Materials
A-TiO <sub>2</sub>		Anatase-rich titanium dioxide
BCP	-	Biphasic Calcium Phosphates
BIC	-	Bone - in- contact
BMP-2	-	Bone morphogenetic protein-2
CFR-PEEK	-	Carbon-fiber-reinforced PEEK
CS	-	Calcium oxide and silicon dioxide
CT	-	Computed tomography
CVD	-	Chemical Vapour Deposition
DLC	-	Diamond-like carbon
DSC	-	Differential scanning calorimetry
E	-	Modulus of elasticity
ECM	-	Extracellular matrixes
EDX	-	Energy Dispersive X-ray Spectroscopy
F	-	Load
FDA	-	Food and Drug Administration of united state
FN	-	Fibronectin
GPa	-	Giga Pascal
gr	-	Gram
Gy	-	Gray (SI unit of absorbed radiation)
H	-	Hardness
HA	-	Hydroxyapatite
HBMSC	-	Human Bone Mesenchymal Stem Cells

hr/hrs	-	Hour/Hours
Hs	-	Scratch hardness
ISO	-	International Organization for Standardization
LC	-	Critical load
LC1	-	First cohesive failure in coating layer
LC2	-	Adhesive failure
min	-	Minute
MPa	-	Mega Pascal
MRI	-	Magnetic resonance imaging
MTS	-	Methoxyphenyl tetrazolium salt
NMR	-	Infrared spectroscopy nuclear magnetic resonance
°C	-	Degree of Celsius
PAEK	-	Polyaryl ether ketones
PEEK	-	Polyether ether ketone
PEKEKK	-	Poly-ether-ketone-ether-ketone-ketone
PLIF	-	Posterior lumbar inter-body fusion cage
PLLA	-	Poly-L-lactic acid
PMMA	-	Poly (methyl methacrylate)
ppb	-	Part per billion
PS	-	Polystyrene
PVD	-	Plasma vapor deposition
R-TiO <sub>2</sub>	-	Rutile-rich titanium dioxide
S	-	Stiffness
SBF	-	Simulated Body Fluid
SPEEK	-	Sulphonated Polyether ether ketone
T <sub>g</sub>	-	Glass transition temperature
Ti	-	Titanium
TiO <sub>2</sub>	-	Titanium dioxide
UV	-	Ultraviolet
VPS	-	Vacuum plasma spraying
W	-	Width of the scratch
XRD	-	X-ray diffraction
XRD	-	X-ray Diffraction Spectroscopy
YSZ	-	Ytria-stabilized zirconia



$\alpha$	-	Indenter face angle
$\beta$ TCP	-	$\beta$ -tricalciumphosphate
$\delta$	-	Indentation depth
$\delta_c$	-	Contact depth
$\varepsilon$	-	Geometric constant of indenter
$\mu\text{m}$	-	Micro meter
$\nu$	-	Poisson's ratio

**LIST OF APPENDICES**

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
A	List of Publications	134
B	Calculation Details of the HA Crystallinity	135
C	Table of the Reading Data of the Ra in Nanometer	136

## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of the Research**

One of the most costly and wide spread health problems is back pain, resulting in more lost productivity of the patient in comparison to other diseases in the United State (Birkmeyer and Weinstein, 1999; Frymoyer and Ducker, 1991). For instance, lasting back pain was increased in the patient from 3.9% to 10.2% between 1992 and 2006 in North Carolina households (Freburger *et al.*, 2009). In 2014 in the United States, over 200,000 spinal fusion surgeries were performed (Zigler, 2015). Around 25% of back pain patients will undergo spinal surgery. Spinal fusion is one of the methods which can be used in many cases to reduce problems of some segment of the spinal column. In spinal fusion surgery, two or more adjacent vertebrae (bone of spinal column) will be immobilized. The adjacent vertebrae are bonded together and the disc between the vertebrae is removed to stimulate the bone growth in between them forming one solid bone (Wu and Yarin, 2013). Many interbody fusion cages with various materials are made. Some of them are made from titanium such as the Ray, Harms, Pyramesh, Lordotic LT and InterFix cages. Some of them are made from polyether ether ketone (PEEK) such as SynCage, and some are made from Cortical Bone Dowel and carbon fiber such as Brantigan cage (Kurtz and Devine, 2007).

The high elastic modulus of titanium (~100GPa) as compared to the adjacent bone, which is implanted in, for instance cortical bone (~15GPa) can result in stress shielding which cause decline of the bone that is in contact with the titanium implant. In the case of spinal implants, in which a large percentage volume of the vertebrae consists of cancellous bone (the modulus of elasticity ~0.3GPa) the shielding problem becomes one of the main issues. In the design of spinal cage, choosing material with low modulus of elasticity is desired. Cancellous bone is a soft, cellular-structured bone, which absorbs shock in the spinal column. Metal ions release is another issue in metallic implants which can cause failure (Ramakrishna *et al.* 2001; Stadelmann *et al.* 2008). These phenomena increase motivation of research towards polymeric implants. In order to reduce the problem of titanium spinal fusion cage, polymer materials with closer modulus elasticity to the bone and high strength have been utilized (Kurtz and Devine, 2007).

Low modulus of elasticity, excellent chemical stability, resistance to radiation used in sterilization procedures, transparency to radio waves, compatibility with reinforcing agent (such as carbon fiber) which can get wide variety of mechanical strength (4~20GPa, depending on fiber volume fraction), etc. have made PEEK as an ideal choice for load bearing implants (Han *et al.*, 2010; Kurtz and Devine, 2007; Sobieraj *et al.* 2009; Toth *et al.*, 2006; Williams, 2008; Xing *et al.*, 2004). PEEK has been used for load bearing orthopaedic applications such as screws, spinal cage, and dental implant (Schwitalla and Muller, 2011; Williams *et al.* 1987).

Food and Drug Administration of the United States (FDA) have accepted carbon-fiber-reinforced PEEK (CFR-PEEK) for spinal fusion cages for human use since the 1990s. Biomaterial PEEK has twenty years of successful clinical history in spinal fusion cage (Kurtz and Devine, 2007). While titanium is still being used for manufacturing the spinal fusion cages, most of the companies have switched their focus to PEEK instead of titanium. For instance, the designers of the Wallis posterior dynamic stabilization system have changed their titanium inter-spinous component to PEEK (Senegas, 2002).

Despite these excellent properties, PEEK is still categorized as bioinert due to its very low reaction with the surrounding bone tissue, which limits its potential applications (Kurtz and Devine, 2007; Rabiei and Sandukas, 2013). To overcome this problem several methods have been proposed, in which the deposition of the hydroxyapatite is one the most attractive methods for researchers. This coating has attracted the attention of many researchers and is the only commercial method for improving the bioactivity of the PEEK implants. Crystalline HA coating is required to stabilize the layer especially for long term orthopaedic and dental implants. In the present methods, the coated HA on the implants require sintering process in order to transform amorphous structure to crystalline (Rabiei and Sandukas, 2013). Sulphonation of PEEK is one of the activation method which deposit the  $\text{SO}_3\text{H}$  functional group via immersion in sulphuric acid (Zhao *et al.*, 2013). This active surface of sulphonated PEEK (SPEEK) can be use for binding with particles.

## 1.2 Problem Statement

Traditional heat treatment for crystallizing of amorphous HA coating layer has been done via annealing at  $600^\circ\text{C}$  or higher. This temperature is above the melting point of PEEK ( $340^\circ\text{C}$ ). To overcome this problem, three new methods of microwave sintering (Adams *et al.*, 2006), laser-induced crystallization (P. M. Smith, Carey, and Sigmon, 1997), and hydrothermal annealing (Ozeki *et al.* 2003; Tong *et al.*, 1997; Yang *et al.* 2007) have been investigated and reported with some success. PEEK is transparent to microwave and it allows heating on HA layer. Laser method is able to provide localized heating and saturated steam is utilized in hydrothermal annealing. Although these methods can crystallize the HA coating layer without spoiling the bulk of the PEEK substrate, at the interface layer between the HA coating and the substrate is impaired (Rabiei and Sandukas, 2013). This research was guided by the following questions:

1. Can sulphonated PEEK be linked to crystalline HA?

2. What is the effect of the sulphonation time on the wettability (bioactivity) of the treated PEEK?
3. What is the effect of the compression load on the properties of the treated layer?

### **1.3 Objectives of the Research**

The principal objective of this research was to establish a new method for depositing crystalline HA on PEEK with the intention to eliminate subsequent sintering of the HA coating. The specific objectives for this research were as follows:

1. To evaluate the linking process via sulphonation for depositing the HA particles with high percentage of crystallinity.
2. To evaluate the effect of different sulphonation process parameters on the surface wettability of the treated PEEK.
3. To compare the mechanical properties and surface morphology of the treated layer before and after applying compression load.

### **1.4 Scopes of the Research**

The research was conducted within the following limits:

1. Polyether ether ketone (PEEK) was used as the substrate material.
2. Sulphonation process was conducted to activate the surface of the PEEK by depositing  $-\text{SO}_3\text{H}$  polar functional group on the molecular chain of the PEEK. The sulphonation process time was varied between 3 to 20 minutes.

3. Fourier Transfer Infrared Spectroscopy (FTIR) was used for evaluating the chemical bonding; X-Ray Diffraction (XRD) for evaluating the crystallinity of the treated layer; Field Emission Scanning Electron Microscope (FESEM) for observing surface morphology; Energy Dispersive X-Ray Spectroscopy (EDX) for determining elements; and Atomic Force Microscope (AFM) for evaluating surface roughness.
4. Mechanical properties of the treated layer were investigated using nanoindentation and micro scratch test.
5. Wettability property of the treated layer was evaluated using water contact angle analysis.
6. Hydraulic press was employed to apply compression load of 15MPa on the treated layer.

### **1.5 Significance of the Research**

Plasma spray is the most popular method used to deposit HA on the PEEK biomedical implant. It was followed by microwave sintering process treatment for increasing the HA crystallinity. Due to high temperature processing, this method has been reported posing problems at the interface HA - substrate layer though the process is capable to provide the required crystallinity level. The equipment used for depositing HA via plasma spray technique is also expensive and demands a secondary microwave process which adds cost and time for manufacturing an implant. In contrast, the proposed method uses a chemical deposition of crystalline HA particles on sulphonated PEEK that could eliminate these costly equipment and secondary processes to manufacture an implant. It is expected the cost will become cheaper with the new method. This encouraging technique also has a great potential to eliminate problems occurred at the interface layer due to high temperature. It is also expected that the manufactured implant via the proposed method becomes more sustainable in the long run.

## **1.6 Organization of the Thesis**

This thesis consists of five chapters: Chapter 1 briefly discusses the background of problem, problem statement, objectives of the research, scope of the research, contribution and organization of the thesis. Chapter 2 discusses the background of biomaterials, and recent strategies for improving the bioactivity of the PEEK's implant. Chapter 3 describes the research methodology used in conducting this research. Chapter 4 presents the experimental results and discusses the findings from experimental trials. Chapter 5 provides the conclusions of the research.



## REFERENCES

- Abu Bakar, M. S., Cheang, P., and Khor, K. A. (1999). Thermal processing of hydroxyapatite reinforced polyetheretherketone composites. *Journal of Materials Processing Technology*, 89–90(0), 462-466.
- Abu Bakar, M. S., Cheang, P., and Khor, K. A. (2003). Tensile properties and microstructural analysis of spheroidized hydroxyapatite–poly (etheretherketone) biocomposites. *Materials Science and Engineering: A*, 345(1–2), 55-63.
- Abu Bakar, M. S., Cheng, M. H., Tang, S. M., Yu, S. C., Liao, K., Tan, C. T., Cheang, P. (2003). Tensile properties, tension-tension fatigue and biological response of polyetheretherketone-hydroxyapatite composites for load-bearing orthopedic implants. *Biomaterials*, 24(13), 2245-2250.
- Adams, D., Malgas, G., Smith, R. D., Massia, S. P., Alford, T. L., and Mayer, J. W. (2006). Microwave annealing for preparation of crystalline hydroxyapatite thin films. *Journal of Materials Science*, 41(21), 7150-7158.
- ASTM, and C1624-05. (2010). Standard Test Method for Adhesion Strength and Mechanical Failure Modes of Ceramic Coatings by Quantitative Single Point Scratch Testing.
- ASTM, and D7027-13. (2013). Standard Test Method for Evaluation of Scratch Resistance of Polymeric Coatings and Plastics Using an Instrumented Scratch Machine. ASTM standard.
- Awaja, F., Bax, D. V., Zhang, S., James, N., and McKenzie, D. R. (2012). Cell Adhesion to PEEK Treated by Plasma Immersion Ion Implantation and Deposition for Active Medical Implants. *Plasma Processes and Polymers*, 9(4), 355-362.
- Baier, R. E., Shafrin, E. G., and Zisman, W. A. (1968). Adhesion: mechanisms that assist or impede it. *Science*, 162(3860), 1360-1368.

- Barkarmo, S., Wennerberg, A., Hoffman, M., Kjellin, P., Breiding, K., Handa, P., and Stenport, V. (2013). Nano-hydroxyapatite-coated PEEK implants: a pilot study in rabbit bone. *J Biomed Mater Res A*, 101(2), 465-471.
- Barletta, M., Gisario, A., and Rubino, G. (2011). Scratch response of high-performance thermoset and thermoplastic powders deposited by the electrostatic spray and 'hot dipping' fluidised bed coating methods: The role of the contact condition. *Surface and Coatings Technology*, 205(21), 5186-5198.
- Batchelor, A. W., and Chandrasekaran, M. (2004). *Service Characteristics of Biomedical Materials and Implants*: Imperial College Press.
- Beauvais, S., and Decaux, O. (2007). *Plasma Sprayed Biocompatible Coatings on PEEK Implants*. Thermal Spray 2007: Global Coating Solutions: Proceedings of the 2007 International Thermal Spray Conference: ASM International.
- Birkmeyer, N. J., and Weinstein, J. N. (1999). Medical versus surgical treatment for low back pain: evidence and clinical practice. *Eff Clin Pract*, 2(5), 218-227.
- Blackwood, D. J. (2003). Biomaterials: Past successes and future problems. *Corrosion Reviews*, 21(2-3), 97-124.
- Blundell, D. J., and Osborn, B. N. (1983). The morphology of poly(aryl-ether-etherketone). *Polymer*, 24(8), 953-958.
- Bombac, D. (2007). Review of materials in medical applications. Pregled materialov v medicinskih aplikacijah. *RMZ - Materials and geoenvironment*, 54(4), 471-499.
- Boyan, B. D., Hummert, T. W., Dean, D. D., and Schwartz, Z. (1996). Role of material surfaces in regulating bone and cartilage cell response. *Biomaterials*, 17(2), 137-146.
- Brantigan, J. W., and Steffee, A. D. (1993). A carbon fiber implant to aid interbody lumbar fusion. Two-year clinical results in the first 26 patients. *Spine (Phila Pa 1976)*, 18(14), 2106-2107.
- Brantigan, J. W., Steffee, A. D., and Geiger, J. M. (1991). A carbon fiber implant to aid interbody lumbar fusion. Mechanical testing. *Spine (Phila Pa 1976)*, 16(6 Suppl), S277-282.
- Briem, D., Strametz, S., Schröder, K., Meenen, N. M., Lehmann, W., Linhart, W., Rueger, J. M. (2005). Response of primary fibroblasts and osteoblasts to

- plasma treated polyetheretherketone (PEEK) surfaces. *Journal of Materials Science: Materials in Medicine*, 16(7), 671-677.
- Chen, Q., and Thouas, G. A. (2015). Metallic implant biomaterials. *Materials Science and Engineering: R: Reports*, 87(0), 1-57.
- Chi, M.H., Tsou, H.K., Chung, C.J., and He, J.-L. (2013). Biomimetic hydroxyapatite grown on biomedical polymer coated with titanium dioxide interlayer to assist osteocompatible performance. *Thin Solid Films*, 549(0), 98-102.
- Cho, D. Y., Liao, W. R., Lee, W. Y., Liu, J. T., Chiu, C. L., and Sheu, P. C. (2002). Preliminary experience using a polyetheretherketone (PEEK) cage in the treatment of cervical disc disease. *Neurosurgery*, 51(6), 1343-1349; discussion 1349-1350.
- Chou, L., Marek, B., and Wagner, W. R. (1999). Effects of hydroxylapatite coating crystallinity on biosolubility, cell attachment efficiency and proliferation in vitro. *Biomaterials*, 20(10), 977-985.
- Cizek, G. R., and Boyd, L. M. (2000). Imaging Pitfalls of Interbody Spinal Implants. *Spine*, 25(20), 2633-2636.
- Clifford, C. A., and Seah, M. P. (2005). Quantification issues in the identification of nanoscale regions of homopolymers using modulus measurement via AFM nanoindentation. *Applied Surface Science*, 252(5), 1915-1933.
- Comesaña, R., Quintero, F., Lusquiños, F., Pascual, M. J., Boutinguiza, M., Durán, A., and Pou, J. (2010). Laser cladding of bioactive glass coatings. *Acta Biomaterialia*, 6(3), 953-961.
- Comyn, J., Mascia, L., Xiao, G., and Parker, B. M. (1996). Plasma-treatment of polyetheretherketone (PEEK) for adhesive bonding. *International Journal of Adhesion and Adhesives*, 16(2), 97-104.
- Converse, G. L., Conrad, T. L., Merrill, C. H., and Roeder, R. K. (2010). Hydroxyapatite whisker-reinforced polyetherketoneketone bone ingrowth scaffolds. *Acta Biomaterialia*, 6(3), 856-863.
- Converse, G. L., Conrad, T. L., and Roeder, R. K. (2009). Fatigue life of hydroxyapatite whisker reinforced polyetherketoneketone. *Trans. Soc. Biomaterials*, 32, 584.
- Converse, G. L., Conrad, T. L., and Roeder, R. K. (2009). Mechanical properties of hydroxyapatite whisker reinforced polyetherketoneketone composite

- scaffolds. *Journal of the Mechanical Behavior of Biomedical Materials*, 2(6), 627-635.
- Converse, G. L., Yue, W., and Roeder, R. K. (2007). Processing and tensile properties of hydroxyapatite-whisker-reinforced polyetheretherketone. *Biomaterials*, 28(6), 927-935.
- Cook, S. D., and Rust-Dawicki, A. M. (1995). Preliminary evaluation of titanium-coated PEEK dental implants. *J Oral Implantol*, 21(3), 176-181.
- Corvelli, A. A., Roberts, J. C., Biermann, P. J., and Cranmer, J. H. (1999). Characterization of a peek composite segmental bone replacement implant. *Journal of Materials Science*, 34(10), 2421-2431.
- D907-12a, A. (2012). Standard Terminology of Adhesives. [www.astm.org](http://www.astm.org).
- D7334-08, A. (2013). Standard Practice for Surface Wettability of Coatings, Substrates and Pigments by Advancing Contact Angle Measurement.
- Dai, Y., Xu, M., Wei, J., Zhang, H., and Chen, Y. (2012). Surface modification of hydroxyapatite nanoparticles by poly(L-phenylalanine) via ROP of L-phenylalanine N-carboxyanhydride (Pha-NCA). *Applied Surface Science*, 258(7), 2850-2855.
- Davis, J. (2003). Overview of biomaterials and their use in medical devices. *Handbook of materials for medical devices. Illustrated edition, Ohio: ASM International*, 1-11.
- Dennes, T. J., and Schwartz, J. (2009). A Nanoscale Adhesion Layer to Promote Cell Attachment on PEEK. *Journal of the American Chemical Society*, 131(10), 3456-3457.
- Devine, D. M., Hahn, J., Richards, R. G., Gruner, H., Wieling, R., and Pearce, S. G. (2013). Coating of carbon fiber-reinforced polyetheretherketone implants with titanium to improve bone apposition. *J Biomed Mater Res B Appl Biomater*, 101(4), 591-598.
- Edited by Basil R. Marple, M. (2007). *Thermal Spray 2007: Global Coating Solutions: Proceedings of the 2007 International Thermal Spray Conference*: ASM International.
- Effect of Silk in Silk/PLGA Hybrid Films on Attachment and Proliferation of Human Aortic Endothelial Cells. (2013). *Polymer Korea*, 37(2), 127.

- Fan, J. P., Tsui, C. P., Tang, C. Y., and Chow, C. L. (2004). Influence of interphase layer on the overall elasto-plastic behaviors of HA/PEEK biocomposite. *Biomaterials*, 25(23), 5363-5373.
- Freburger, J. K., Holmes, G. M., Agans, R. P., Jackman, A. M., Darter, J. D., Wallace, A. S., Carey, T. S. (2009). The rising prevalence of chronic low back pain. *Arch Intern Med*, 169(3), 251-258.
- Frymoyer, J. W., and Ducker, T. B. (1991). *The Adult Spine: Principles and Practice*: Raven Press.
- Gao, C., Gao, Q., Li, Y., Rahaman, M. N., Teramoto, A., and Abe, K. (2012). Preparation and in vitro characterization of electrospun PVA scaffolds coated with bioactive glass for bone regeneration. *J Biomed Mater Res A*, 100(5), 1324-1334.
- García, A. J., Vega, M. D., and Boettiger, D. (1999). Modulation of cell proliferation and differentiation through substrate- dependent changes in fibronectin conformation. *Molecular Biology of the Cell*, 10(3), 785-798.
- Geetha, M., Singh, A. K., Asokamani, R., and Gogia, A. K. (2009). Ti based biomaterials, the ultimate choice for orthopaedic implants – A review. *Progress in Materials Science*, 54(3), 397-425.
- Green, S., and Devine, J. (2004). PEEK-OPTIMA Polymer: An Alternative Material for the Development of Long-Term Medical Implant Applications. *BONEZone*, 3-5.
- Ha, S. W., Eckert, K. L., Wintermantel, E., Gruner, H., Guecheva, M., and Vonmont, H. (1997). NaOH treatment of vacuum-plasma-sprayed titanium on carbon fibre-reinforced poly(etheretherketone). *J Mater Sci Mater Med*, 8(12), 881-886.
- Ha, S. W., Gisepp, A., Mayer, J., Wintermantel, E., Gruner, H., and Wieland, M. (1997). Topographical characterization and microstructural interface analysis of vacuum-plasma-sprayed titanium and hydroxyapatite coatings on carbon fibre-reinforced poly(etheretherketone). *J Mater Sci Mater Med*, 8(12), 891-896.
- Ha, S. W., Kirch, M., Birchler, F., Eckert, K. L., Mayer, J., Wintermantel, E., Vonmont, H. (1997). Surface activation of polyetheretherketone (PEEK) and formation of calcium phosphate coatings by precipitation. *J Mater Sci Mater Med*, 8(11), 683-690.

- Ha, S. W., Mayer, J., Koch, B., and Wintermantel, E. (1994). Plasma-sprayed hydroxylapatite coating on carbon fibre reinforced thermoplastic composite materials. *Journal of Materials Science: Materials in Medicine*, 5(6-7), 481-484.
- Hahn, B. D., Park, D. S., Choi, J. J., Ryu, J., Yoon, W. H., Choi, J. H., Jung, I. K. (2013). Osteoconductive hydroxyapatite coated PEEK for spinal fusion surgery. *Applied Surface Science*, 283(0), 6-11.
- Han, C. M., Lee, E. J., Kim, H. E., Koh, Y. H., Kim, K. N., Ha, Y., and Kuh, S. U. (2010). The electron beam deposition of titanium on polyetheretherketone (PEEK) and the resulting enhanced biological properties. *Biomaterials*, 31(13), 3465-3470.
- Han, C. M., Jang, T. S., Kim, H. E., and Koh, Y. H. (2014). Creation of nanoporous TiO<sub>2</sub> surface onto polyetheretherketone for effective immobilization and delivery of bone morphogenetic protein. *J Biomed Mater Res A*, 102(3), 793-800.
- Han, C. M., Lee, E. J., Kim, H. E., Koh, Y. H., Kim, K. N., Ha, Y., and Kuh, S. U. (2010). The electron beam deposition of titanium on polyetheretherketone (PEEK) and the resulting enhanced biological properties. *Biomaterials*, 31(13), 3465-3470.
- Hansen, D. C. (2008). Metal corrosion in the human body: the ultimate bio-corrosion scenario. *The Electrochemical Society Interface*, 17(2), 31.
- Heiland, K., Hill, D. J. T., O'Donnell, J. H., and Pomery, P. J. (1994). Radiation degradation of poly(arylene ether ketone)s. *Polymers for Advanced Technologies*, 5(2), 116-121.
- Helsen, J. A., and Breme, H. J. (1998). *Metals as biomaterials*: Wiley.
- Hemlata Garg, Gaurav Bedi, and Garg, A. (2012). Implant Surface Modifications: A Review. *Journal of Clinical and Diagnostic Reseach*, 6(2), 319-324.
- Hench, L. L., and Paschall, H. A. (1973). Direct chemical bond of bioactive glass-ceramic materials to bone and muscle. *Journal of Biomedical Materials Research*, 7(3), 25-42.
- Hengky, C., Kelsen, B., and Saraswati, P. (2009). Mechanical and biological characterization of pressureless sintered hydroxapatite-polyetheretherketone biocomposite. *International Conference on Biomedical Engineering (ICBME) Proceedings*, 23, 261-264.

- Heo, Y., Im, H., and Kim, J. (2013). The effect of sulfonated graphene oxide on Sulfonated Poly (Ether Ether Ketone) membrane for direct methanol fuel cells. *Journal of Membrane Science*, 425–426(0), 11-22.
- Ilze, M., and Janis, M. (2008). Effect of substrate hardness and film structure on indentation depth criteria for film hardness testing. *Journal of Physics D: Applied Physics*, 41(7), 074010.
- Jaafar, J., Ismail, A. F., and Mustafa, A. (2007). Physicochemical study of poly(ether ether ketone) electrolyte membranes sulfonated with mixtures of fuming sulfuric acid and sulfuric acid for direct methanol fuel cell application. *Materials Science and Engineering: A*, 460–461(0), 475-484.
- Jahan, M. S., Wang, C., Schwartz, G., and Davidson, J. A. (1991). Combined chemical and mechanical effects on free radicals in UHMWPE joints during implantation. *J Biomed Mater Res*, 25(8), 1005-1017.
- Janaki, K., Elamathi, S., and Sangeetha, D. (2008). Development and Characterization of Polymer Ceramic Composites for Orthopedic Applications. *Trends Biomater. Artif. Organs*, 22(3), 169-178.
- Jarcho, M., Kay, J. F., Gumaer, K. I., Doremus, R. H., and Drobeck, H. P. (1977). Tissue, cellular and subcellular events at a bone-ceramic hydroxylapatite interface. *J Bioeng*, 1(2), 79-92.
- Jian, S. R., Chen, G. J., and Lin, T. C. (2010). Berkovich nanoindentation on AlN thin films. *Nanoscale Research Letters*, 5(6), 935-940.
- Jiya, T., Smit, T., Deddens, J., and Mullender, M. (2009). Posterior lumbar interbody fusion using nonresorbable poly-ether-ether-ketone versus resorbable poly-L-lactide-co-D,L-lactide fusion devices: a prospective, randomized study to assess fusion and clinical outcome. *Spine (Phila Pa 1976)*, 34(3), 233-237.
- Jockisch, K. A., Brown, S. A., Bauer, T. W., and Merritt, K. (1992). Biological response to chopped-carbon-fiber-reinforced peek. *J Biomed Mater Res*, 26(2), 133-146.
- Jung, H. D., Sun Park, H., Kang, M. H., Lee, S. M., Kim, H. E., Estrin, Y., and Koh, Y.H. (2014). Polyetheretherketone/magnesium composite selectively coated with hydroxyapatite for enhanced in vitro bio-corrosion resistance and biocompatibility. *Materials Letters*, 116(0), 20-22.

- Kasemo, B., and Lausmaa, J. (1988). Biomaterial and implant surfaces: on the role of cleanliness, contamination, and preparation procedures. *J Biomed Mater Res*, 22(A2 Suppl), 145-158.
- Katzer, A., Marquardt, H., Westendorf, J., Wening, J. V., and von Foerster, G. (2002). Polyetheretherketone—cytotoxicity and mutagenicity in vitro. *Biomaterials*, 23(8), 1749-1759.
- Keselowsky, B. G., Collard, D. M., and García, A. J. (2003). Surface chemistry modulates fibronectin conformation and directs integrin binding and specificity to control cell adhesion. *Journal of Biomedical Materials Research - Part A*, 66(2), 247-259.
- Khoury, J., Kirkpatrick, S. R., Maxwell, M., Cherian, R. E., Kirkpatrick, A., and Svrluga, R. C. (2013). Neutral atom beam technique enhances bioactivity of PEEK. *Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms*, 307(0), 630-634.
- Kim, I. Y., Sugino, A., Kikuta, K., Ohtsuki, C., and Cho, S. B. (2009). Bioactive composites consisting of PEEK and calcium silicate powders. *Journal of Biomaterials Applications*, 24(2), 105-118.
- Kirkpatrick, A., Kirkpatrick, S., Walsh, M., Chau, S., Mack, M., Harrison, S., Khoury, J. (2013). Investigation of accelerated neutral atom beams created from gas cluster ion beams. *Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms*, 307(0), 281-289.
- Koenig, A. L., Gambillara, V., and Grainger, D. W. (2003). Correlating fibronectin adsorption with endothelial cell adhesion and signaling on polymer substrates. *Journal of Biomedical Materials Research - Part A*, 64(1), 20-37.
- Kokubo, T., and Takadama, H. (2006). How useful is SBF in predicting in vivo bone bioactivity? *Biomaterials*, 27(15), 2907-2915.
- Krishnamurithy, G., Murali, M. R., Hamdi, M., Abbas, A. A., Raghavendran, H. B., and Kamarul, T. (2014a). Characterization of bovine-derived porous hydroxyapatite scaffold and its potential to support osteogenic differentiation of human bone marrow derived mesenchymal stem cells. *Ceramics International*, 40(1), 771-777.
- Krishnamurithy, G., Murali, M. R., Hamdi, M., Abbas, A. A., Raghavendran, H. B., and Kamarul, T. (2014b). Characterization of bovine-derived porous



- hydroxyapatite scaffold and its potential to support osteogenic differentiation of human bone marrow derived mesenchymal stem cells. *Ceramics International*, 40(1, Part A), 771-777.
- Kumar, S., Anderson, D. P., and Adams, W. W. (1986). Crystallization and morphology of poly(aryl-ether-ether-ketone). *Polymer*, 27(3), 329-336.
- Kurtz, S. M. (2012). Chapter 1 - An Overview of PEEK Biomaterials *PEEK Biomaterials Handbook* (pp. 1-7). Oxford: William Andrew Publishing.
- Kurtz, S. M., and Devine, J. N. (2007). PEEK biomaterials in trauma, orthopedic, and spinal implants. *Biomaterials*, 28(32), 4845-4869.
- Landi, E., Tampieri, A., Celotti, G., and Sprio, S. (2000). Densification behaviour and mechanisms of synthetic hydroxyapatites. *Journal of the European Ceramic Society*, 20(14), 2377-2387.
- Laurens, P., Ould Bouali, M., Meducin, F., and Sadras, B. (2000). Characterization of modifications of polymer surfaces after excimer laser treatments below the ablation threshold. *Applied Surface Science*, 154-155(0), 211-216.
- Laurens, P., Sadras, B., Decobert, F., Arefi-Khonsari, F., and Amouroux, J. (1998). Enhancement of the adhesive bonding properties of PEEK by excimer laser treatment. *International Journal of Adhesion and Adhesives*, 18(1), 19-27.
- Laurens, P., Sadras, B., Décobert, F., Aréfi-Khonsari, F., and Amouroux, J. (1999). Laser-induced surface modifications of poly(ether ether ketone): Influence of the excimer laser wavelength. *Journal of Adhesion Science and Technology*, 13(9), 983-997.
- Le Guéhennec, L., Soueidan, A., Layrolle, P., and Amouriq, Y. (2007). Surface treatments of titanium dental implants for rapid osseointegration. *Dental Materials*, 23(7), 844-854.
- Lee, J., Lee, S., Kim, S., Kim, K., Kim, Y., Song, J., Khang, G. (2013). Effect of Silk in Silk/PLGA Hybrid Films on Attachment and Proliferation of Human Aortic Endothelial Cells. *POLYMER-KOREA*, 37(2), 127-134.
- Lee, J. H., Jang, H. L., Lee, K. M., Baek, H.-R., Jin, K., Hong, K. S., Lee, H. K. (2013). In vitro and in vivo evaluation of the bioactivity of hydroxyapatite-coated polyetheretherketone biocomposites created by cold spray technology. *Acta Biomaterialia*, 9(4), 6177-6187.
- Lehnert, D., Wehrle-Haller, B., David, C., Weiland, U., Ballestrem, C., Imhof, B. A., and Bastmeyer, M. (2004). Cell behaviour on micropatterned substrata:

- Limits of extracellular matrix geometry for spreading and adhesion. *Journal of Cell Science*, 117(1), 41-52.
- Li, H. M., Fouracre, R. A., Given, M. J., Banford, H. M., Wysocki, S., and Karolczak, S. (1999). The effects on polyetheretherketone and polyethersulfone of electron and andgamma; irradiation. *Dielectrics and Electrical Insulation, IEEE Transactions on*, 6(3), 295-303.
- Li, K., Yeung, C. Y., Yeung, K. W. K., and Tjong, S. C. (2012). Sintered Hydroxyapatite/Polyetheretherketone Nanocomposites: Mechanical Behavior and Biocompatibility. *Advanced Engineering Materials*, 14(4), B155-B165.
- Lin, T. W., Corvelli, A. A., Frondoza, C. G., Roberts, J. C., and Hungerford, D. S. (1997). Glass peek composite promotes proliferation and osteocalcin production of human osteoblastic cells. *J Biomed Mater Res*, 36(2), 137-144.
- Liu, F., Yang, F., Gao, Y., Jiang, W., Guan, Y., Rack, P., Liaw, P. K. (2009). Micro-scratch study of a magnetron-sputtered Zr-based metallic-glass film. *Surface and Coatings Technology*, 203(22), 3480-3484.
- Liu, X., Chu, P. K., and Ding, C. (2004). Surface modification of titanium, titanium alloys, and related materials for biomedical applications. *Materials Science and Engineering: R: Reports*, 47(3-4), 49-121.
- Ma, R., Weng, L., Bao, X., Ni, Z., Song, S., and Cai, W. (2012). Characterization of in situ synthesized hydroxyapatite/polyetheretherketone composite materials. *Materials Letters*, 71(0), 117-119.
- Ma, R., Weng, L., Bao, X., Song, S., and Zhang, Y. (2013). In vivo biocompatibility and bioactivity of in situ synthesized hydroxyapatite/polyetheretherketone composite materials. *Journal of Applied Polymer Science*, 127(4), 2581-2587.
- Ma, R., Weng, L., Fang, L., Luo, Z., and Song, S. (2012). Structure and mechanical performance of in situ synthesized hydroxyapatite/polyetheretherketone nanocomposite materials. *Journal of Sol-Gel Science and Technology*, 62(1), 52-56.
- Marchand-Brynaert, J., Pantano, G., and Noiset, O. (1997). Surface fluorination of PEEK film by selective wet-chemistry. *Polymer*, 38(6), 1387-1394.
- Mathieson, I., and Bradley, R. H. (1996). Improved adhesion to polymers by UV/ozone surface oxidation. *International Journal of Adhesion and Adhesives*, 16(1), 29-31.

- McAfee, P. C. (1999). Interbody fusion cages in reconstructive operations on the spine. *J Bone Joint Surg Am*, 81(6), 859-880.
- McMillin, C. (1993). *Evaluation of PEKEKK composites for spine implants*. Paper presented at the 38th international SAMPE symposium.
- Melnyk, A. D., Chak, J. D., Cripton, P. A., Dvorak, M. F., and Oxland, T. R. (2012). Shear force measurements on low- and high-stiffness posterior fusion devices. *Med Eng Phys*, 34(9), 1260-1267.
- Michael, K. E., Vernekar, V. N., Keselowsky, B. G., Meredith, J. C., Latour, R. A., and García, A. J. (2003). Adsorption-induced conformational changes in fibronectin due to interactions with well-defined surface chemistries. *Langmuir*, 19(19), 8033-8040.
- Mittal, K. L. (1995). *Adhesion Measurement of Films and Coatings*: Taylor and Francis.
- Modic, M. T., Steinberg, P. M., Ross, J. S., Masaryk, T. J., and Carter, J. R. (1988). Degenerative disk disease: assessment of changes in vertebral body marrow with MR imaging. *Radiology*, 166(1 Pt 1), 193-199.
- Morrison, C., Macnair, R., MacDonald, C., Wykman, A., Goldie, I., and Grant, M. H. (1995). In vitro biocompatibility testing of polymers for orthopaedic implants using cultured fibroblasts and osteoblasts. *Biomaterials*, 16(13), 987-992.
- Nieminen, T., Kallela, I., Wuolijoki, E., Kainulainen, H., Hiidenheimo, I., and Rantala, I. (2008). Amorphous and crystalline polyetheretherketone: Mechanical properties and tissue reactions during a 3-year follow-up. *J Biomed Mater Res A*, 84(2), 377-383.
- Noiset, O., Schneider, Y.-J., and Marchand-Brynaert, J. (1997). Surface modification of poly(aryl ether ether ketone) (PEEK) film by covalent coupling of amines and amino acids through a spacer arm. *Journal of Polymer Science Part A: Polymer Chemistry*, 35(17), 3779-3790.
- Noiset, O., Schneider, Y. J., and Marchand-Brynaert, J. (1997). Surface modification of poly(aryl ether ether ketone) (PEEK) film by covalent coupling of amines and amino acids through a spacer arm. *Journal of Polymer Science, Part A: Polymer Chemistry*, 35(17), 3779-3790.

- Noiset, O., Schneider, Y. J., and Marchand-Brynaert, J. (1999). Fibronectin adsorption or/and covalent grafting on chemically modified PEEK film surfaces. *J Biomater Sci Polym Ed*, 10(6), 657-677.
- Noiset, O., Schneider, Y. J., and Marchand-Brynaert, J. (2000). Adhesion and growth of CaCo2 cells on surface-modified PEEK substrata. *J Biomater Sci Polym Ed*, 11(7), 767-786.
- Occhiello, E., Morra, M., Guerrini, G. L., and Garbassi, F. (1992). Adhesion properties of plasma-treated carbon/PEEK composites. *Composites*, 23(3), 193-200.
- Oliver, W. C., and Pharr, G. M. (1992). An improved technique for determining hardness and elastic modulus using load and displacement sensing indentation experiments. *Journal of Materials Research*, 7, 1564-1583.
- Ozeki, K., Mishima, A., Yuhta, T., Fukui, Y., and Aoki, H. (2003). Bone bonding strength of sputtered hydroxyapatite films subjected to a low temperature hydrothermal treatment. *Biomed Mater Eng*, 13(4), 451-463.
- Parsons, J. R., Bhayani, S., Alexander, H., and Weiss, A. B. (1985). Carbon fiber debris within the synovial joint. A time-dependent mechanical and histologic study. *Clin Orthop Relat Res*(196), 69-76.
- Petrovic, L., Pohle, D., Münstedt, H., Rechtenwald, T., Schlegel, K. A., and Rupprecht, S. (2006). Effect of  $\beta$ TCP filled polyetheretherketone on osteoblast cell proliferation in vitro. *Journal of Biomedical Science*, 13(1), 41-46.
- Pharr, G. M., Oliver, W. C., Brotzen, F. R., and Others. (1992a). On the generality of the relationship among contact stiffness, contact area, and elastic modulus during indentation. *Journal of Materials Research*, 7(3), 612-617.
- Pharr, G. M., Oliver, W. C., Brotzen, F. R., and Others. (1992b). On the generality of the relationship among contact stiffness, contact area, and elastic modulus during indentation. *Journal of Materials Research*, 7(3), -617.
- Pino, M., Stingelin, N., and Tanner, K. E. (2008). Nucleation and growth of apatite on NaOH-treated PEEK, HDPE and UHMWPE for artificial cornea materials. *Acta Biomaterialia*, 4(6), 1827-1836.
- Pohle, D., Ponader, S., Rechtenwald, T., Schmidt, M., Schlegel, K. A., Münstedt, H., Von Wilmowsky, C. (2007). Processing of three-dimensional laser sintered

- polyetheretherketone composites and testing of osteoblast proliferation in vitro. *Macromolecular Symposia*, 253, 65-70.
- Poullsson, A. H., Eglin, D., Zeiter, S., Camenisch, K., Sprecher, C., Agarwal, Y., Richards, R. G. (2014). Osseointegration of machined, injection moulded and oxygen plasma modified PEEK implants in a sheep model. *Biomaterials*, 35(12), 3717-3728. doi: 10.1016/j.biomaterials.2013.12.056
- R Wieling, A. G. (2009). Osteointegrative surfaces for CF/PEEK implants. *European Cells and Materials*, 17(1), 10.
- Rabiei, A., and Sandukas, S. (2013). Processing and evaluation of bioactive coatings on polymeric implants. *Journal of Biomedical Materials Research Part A*, 101A(9), 2621-2629.
- Ramakrishna, S., Mayer, J., Wintermantel, E., and Leong, K. W. (2001). Biomedical applications of polymer-composite materials: a review. *Composites Science and Technology*, 61(9), 1189-1224.
- Recent progress in interfacial toughening and damage self-healing of polymer composites based on electrospun and solution-blown nanofibers: An overview. (2013). *Journal of Applied Polymer Science*.
- Reisch, M. S. (2007). Medical polymers renaissance. *Chemical and Engineering News*, 85(45), 14-17.
- Rickerby, D. S. (1988). A review of the methods for the measurement of coating-substrate adhesion. *Surface and Coatings Technology*, 36(1-2), 541-557.
- Rivard, C. H., Rhalmi, S., and Coillard, C. (2002). In vivo biocompatibility testing of peek polymer for a spinal implant system: A study in rabbits. *Journal of Biomedical Materials Research*, 62(4), 488-498.
- Riveiro, A., Soto, R., Comesaña, R., Boutinguiza, M., del Val, J., Quintero, F., Pou, J. (2012). Laser surface modification of PEEK. *Applied Surface Science*, 258(23), 9437-9442.
- Roach, P., Farrar, D., and Perry, C. C. (2005). Interpretation of protein adsorption: Surface-induced conformational changes. *Journal of the American Chemical Society*, 127(22), 8168-8173.
- Roeder, R. K., Smith, S. M., Conrad, T. L., Yanchak, N. J., Merrill, C. H., and Converse, G. L. (2009). Porous and bioactive PEEK implants for interbody spinal fusion. *Adv Mater Process*, 167(10), 46-48.

- Sáenz, A., Rivera, E., Brostow, W., and Castano, V. M. (1999). Ceramic biomaterials: an introductory overview. *Journal of Materials Education*, 21(5/6), 267-276.
- Sagomonyants, K. B., Jarman-Smith, M. L., Devine, J. N., Aronow, M. S., and Gronowicz, G. A. (2008). The in vitro response of human osteoblasts to polyetheretherketone (PEEK) substrates compared to commercially pure titanium. *Biomaterials*, 29(11), 1563-1572.
- Sandukas, S. (2012). *Development and Analysis of Bioactive CaP Coatings for Biomedical Implants*. (Doctor of Philosophy), North Carolina State University.
- Scholes, S. C., and Unsworth, A. (2009). Wear studies on the likely performance of CFR-PEEK/CoCrMo for use as artificial joint bearing materials. *J Mater Sci Mater Med*, 20(1), 163-170.
- Schröder, K., Meyer-Plath, A., Keller, D., and Ohl, A. (2002). On the Applicability of Plasma Assisted Chemical Micropatterning to Different Polymeric Biomaterials. *Plasmas and Polymers*, 7(2), 103-125.
- Schwitalla, A. D., and Muller, W. D. (2011). PEEK dental implants: A Review of the Literature. *J Oral Implantol*.
- Scotchford, C. A., Ball, M., Winkelmann, M., Vörös, J., Csucs, C., Brunette, D. M., Textor, M. (2003). Chemically patterned, metal-oxide-based surfaces produced by photolithographic techniques for studying protein- and cell-interactions. II: Protein adsorption and early cell interactions. *Biomaterials*, 24(7), 1147-1158.
- Senegas, J. (2002). Mechanical supplementation by non-rigid fixation in degenerative intervertebral lumbar segments: the Wallis system. *Eur Spine J*, 11 Suppl 2, S164-169.
- Smith, D. C. (1993). Dental implants: materials and design considerations. *Int J Prosthodont*, 6(2), 106-117.
- Smith, D. C., Pilliar, R. M., and Chernecky, R. (1991). Dental implant materials. I. Some effects of preparative procedures on surface topography. *J Biomed Mater Res*, 25(9), 1045-1068.
- Smith, P. M., Carey, P. G., and Sigmon, T. W. (1997). Excimer laser crystallization and doping of silicon films on plastic substrates. *Applied Physics Letters*, 70(3), 342-344.

- Sobieraj, M. C., Kurtz, S. M., and Rimnac, C. M. (2009). Notch sensitivity of PEEK in monotonic tension. *Biomaterials*, 30(33), 6485-6494.
- Spruit, M., Falk, R. G., Beckmann, L., Steffen, T., and Castelein, R. M. (2005). The in vitro stabilising effect of polyetheretherketone cages versus a titanium cage of similar design for anterior lumbar interbody fusion. *European Spine Journal*, 14(8), 752-758.
- Stadelmann, V. A., Terrier, A., and Pioletti, D. P. (2008). Microstimulation at the bone-implant interface upregulates osteoclast activation pathways. *Bone*, 42(2), 358-364.
- Staff, P. D. L. (1994). Effect of Sterilization Methods on Plastics and Elastomers (pp. 173): William Andrew Publishing/Plastics Design Library.
- Steinmann, P. A., and Hintermann, H. E. (1989). A review of the mechanical tests for assessment of thin-film adhesion. *Journal of Vacuum Science andamp; Technology A*, 7(3), 2267-2272.
- Strnad, Z., Strnad, J., Povysil, C., and Urban, K. (2000). Effect of plasma-sprayed hydroxyapatite coating on the osteoconductivity of commercially pure titanium implants. *Int J Oral Maxillofac Implants*, 15(4), 483-490.
- Suture-reinforced electrospun polydioxanone–elastin small-diameter tubes for use in vascular tissue engineering: A feasibility study. (2008). *Acta Biomaterialia*, 4(1), 58.
- Talbott, M. F., Springer, G. S., and Berglund, L. A. (1987). The Effects of Crystallinity on the Mechanical Properties of PEEK Polymer and Graphite Fiber Reinforced PEEK. *Journal of Composite Materials*, 21(11), 1056-1081.
- Tan, K. H., Chua, C. K., Leong, K. F., Cheah, C. M., Cheang, P., Abu Bakar, M. S., and Cha, S. W. (2003). Scaffold development using selective laser sintering of polyetheretherketone-hydroxyapatite biocomposite blends. *Biomaterials*, 24(18), 3115-3123.
- Tan, K. H., Chua, C. K., Leong, K. F., Cheah, C. M., Gui, W. S., Tan, W. S., and Wiria, F. E. (2005). Selective laser sintering of biocompatible polymers for applications in tissue engineering. *Bio-Medical Materials and Engineering*, 15(1-2), 113-124.
- Tan, K. H., Chua, C. K., Leong, K. F., Naing, M. W., and Cheah, C. M. (2005). Fabrication and characterization of three-dimensional poly(ether-etherketone)/-hydroxyapatite biocomposite scaffolds using laser sintering.

*Proceedings of the Institution of Mechanical Engineers, Part H: Journal of Engineering in Medicine*, 219(3), 183-194.

- Tanaka-Kamioka, K., Kamioka, H., Ris, H., and Lim, S.-S. (1998). Osteocyte Shape Is Dependent on Actin Filaments and Osteocyte Processes Are Unique Actin-Rich Projections. *Journal of Bone and Mineral Research*, 13(10), 1555-1568.
- Tang, S. M., Cheang, P., AbuBakar, M. S., Khor, K. A., and Liao, K. (2004). Tension-tension fatigue behavior of hydroxyapatite reinforced polyetheretherketone composites. *International Journal of Fatigue*, 26(1), 49-57.
- Tong, W., Chen, J., Cao, Y., Lu, L., Feng, J., and Zhang, X. (1997). Effect of water vapor pressure and temperature on the amorphous-to-crystalline HA conversion during heat treatment of HA coatings. *J Biomed Mater Res*, 36(2), 242-245.
- Toth, J. M., Wang, M., Estes, B. T., Scifert, J. L., Seim Iii, H. B., and Turner, A. S. (2006). Polyetheretherketone as a biomaterial for spinal applications. *Biomaterials*, 27(3), 324-334.
- Tsou, H.K., Hsieh, P.Y., Chung, C.J., Tang, C.H., Shyr, T.W., and He, J.L. (2009). Low-temperature deposition of anatase TiO<sub>2</sub> on medical grade polyetheretherketone to assist osseous integration. *Surface and Coatings Technology*, 204(6-7), 1121-1125.
- Underwood, P. A., Steele, J. G., and Dalton, B. A. (1993). Effects of polystyrene surface chemistry on the biological activity of solid phase fibronectin and vitronectin, analysed with monoclonal antibodies. *Journal of Cell Science*, 104(3), 793-803.
- Valli, J. (1986). A review of adhesion test methods for thin hard coatings. *Journal of Vacuum Science andamp; Technology A*, 4(6), 3007-3014.
- van Dijk, M., Tunc, D. C., Smit, T. H., Higham, P., Burger, E. H., and Wuisman, P. I. (2002). In vitro and in vivo degradation of bioabsorbable PLLA spinal fusion cages. *J Biomed Mater Res*, 63(6), 752-759. doi: 10.1002/jbm.10466
- van Kooten, T. G., Spijker, H. T., and Busscher, H. J. (2004). Plasma-treated polystyrene surfaces: model surfaces for studying cell-biomaterial interactions. *Biomaterials*, 25(10), 1735-1747.
- . Victrex PEEK. (1999). Product Guide Medical.



- Von Wilmonsky, C., Lutz, R., Meisel, U., Srour, S., Rupprecht, S., Toyoshima, T., Schmidt, M. (2009). In Vivo Evaluation of  $\beta$ -TCP Containing 3D Laser Sintered Poly(ether ether ketone) Composites in Pigs. *Journal of Bioactive and Compatible Polymers*, 24(2), 169-184.
- Von Wilmowsky, C., Vairaktaris, E., Pohle, D., Rechtenwald, T., Lutz, R., Münstedt, H., Nkenke, E. (2008). Effects of bioactive glass and  $\beta$ -TCP containing three-dimensional laser sintered polyetheretherketone composites on osteoblasts in vitro. *Journal of Biomedical Materials Research - Part A*, 87(4), 896-902.
- Wang, H., Eliaz, N., Xiang, Z., Hsu, H. P., Spector, M., and Hobbs, L. W. (2006). Early bone apposition in vivo on plasma-sprayed and electrochemically deposited hydroxyapatite coatings on titanium alloy. *Biomaterials*, 27(23), 4192-4203.
- Wang, H., Lu, T., Meng, F., Zhu, H., and Liu, X. (2014). Enhanced osteoblast responses to poly ether ether ketone surface modified by water plasma immersion ion implantation. *Colloids Surf B Biointerfaces*, 117C, 89-97.
- Wang, H., Xu, M., Zhang, W., Kwok, D. T., Jiang, J., Wu, Z., and Chu, P. K. (2010). Mechanical and biological characteristics of diamond-like carbon coated poly aryl-ether-ether-ketone. *Biomaterials*, 31(32), 8181-8187.
- Wang, L., Weng, L., Song, S., and Sun, Q. (2010). Mechanical properties and microstructure of polyetheretherketone–hydroxyapatite nanocomposite materials. *Materials Letters*, 64(20), 2201-2204.
- Wang, L., Weng, L., Song, S., Zhang, Z., Tian, S., and Ma, R. (2011). Characterization of polyetheretherketone–hydroxyapatite nanocomposite materials. *Materials Science and Engineering: A*, 528(10–11), 3689-3696.
- Waser-Althaus, J., Salamon, A., Waser, M., Padeste, C., Kreutzer, M., Pieleles, U., Peters, K. (2014). Differentiation of human mesenchymal stem cells on plasma-treated polyetheretherketone. *Journal of Materials Science: Materials in Medicine*, 25(2), 515-525.
- Wenz, L. M., Merritt, K., Brown, S. A., Moet, A., and Steffee, A. D. (1990). In vitro biocompatibility of polyetheretherketone and polysulfone composites. *Journal of Biomedical Materials Research*, 24(2), 207-215.
- Williams, D. (2008). Polyetheretherketone for long-term implantable devices. *Medical device technology*, 19(1), 8, 10-11.

- Williams, D. F., McNamara, A., and Turner, R. M. (1987). Potential of polyetheretherketone (PEEK) and carbon-fibre-reinforced PEEK in medical applications. *Journal of Materials Science Letters*, 6(2), 188-190.
- Wilson, C. J., Clegg, R. E., Leavesley, D. I., and Percy, M. J. (2005). Mediation of biomaterial-cell interactions by adsorbed proteins: A review. *Tissue Engineering*, 11(1-2), 1-18.
- Wong, K. L., Wong, C. T., Liu, W. C., Pan, H. B., Fong, M. K., Lam, W. M., Lu, W. W. (2009). Mechanical properties and in vitro response of strontium-containing hydroxyapatite/polyetheretherketone composites. *Biomaterials*, 30(23-24), 3810-3817.
- Wu, X.-F., and Yarin, A. L. (2013). Recent progress in interfacial toughening and damage self-healing of polymer composites based on electrospun and solution-blown nanofibers: An overview. *Journal of Applied Polymer Science*, 130(4), 2225-2237.
- Wu, X., Liu, X., Wei, J., Ma, J., Deng, F., and Wei, S. (2012). Nano-TiO<sub>2</sub>/PEEK bioactive composite as a bone substitute material: in vitro and in vivo studies. *Int J Nanomedicine*, 7, 1215-1225.
- Xing, P., Robertson, G. P., Guiver, M. D., Mikhailenko, S. D., Wang, K., and Kaliaguine, S. (2004). Synthesis and characterization of sulfonated poly(ether ether ketone) for proton exchange membranes. *Journal of Membrane Science*, 229(1-2), 95-106.
- Xu, S., Ma, X., Wen, H., Tang, G., and Li, C. (2014). Effect of annealing on the mechanical and scratch properties of BCN films obtained by magnetron sputtering deposition. *Applied Surface Science*.
- Xue, W., Tao, S., Liu, X., Zheng, X., and Ding, C. (2004). In vivo evaluation of plasma sprayed hydroxyapatite coatings having different crystallinity. *Biomaterials*, 25(3), 415-421.
- Yang, C. W., Lui, T. S., and Chang, E. (2007). Low temperature crystallization and structural modification of plasma-sprayed hydroxyapatite coating with hydrothermal treatment. *Advanced Materials Research*, 15, 147-152.
- Yu, S., Hariram, K. P., Kumar, R., Cheang, P., and Aik, K. K. (2005). In vitro apatite formation and its growth kinetics on hydroxyapatite/polyetheretherketone biocomposites. *Biomaterials*, 26(15), 2343-2352.

- Yuan, Y., and Lee, T. R. (2013). Contact Angle and Wetting Properties. In G. Bracco and B. Holst (Eds.), *Surface Science Techniques* (Vol. 51, pp. 3-34): Springer Berlin Heidelberg.
- Zhang, G., Leparoux, S., Liao, H., and Coddet, C. (2006). Microwave sintering of poly-ether-ether-ketone (PEEK) based coatings deposited on metallic substrate. *Scripta Materialia*, 55(7), 621-624.
- Zhang, Y., Hao, L., Savalani, M. M., Harris, R. A., Di Silvio, L., and Tanner, K. E. (2009). In vitro biocompatibility of hydroxyapatite-reinforced polymeric composites manufactured by selective laser sintering. *J Biomed Mater Res A*, 91(4), 1018-1027.
- Zhao, Y., Wong, H. M., Wang, W., Li, P., Xu, Z., Chong, E. Y., Chu, P. K. (2013). Cytocompatibility, osseointegration, and bioactivity of three-dimensional porous and nanostructured network on polyetheretherketone. *Biomaterials*, 34(37), 9264-9277.
- Ziats, N. P., Miller, K. M., and Anderson, J. M. (1988). In vitro and in vivo interactions of cells with biomaterials. *Biomaterials*, 9(1), 5-13.
- Zigler, J. (2015). <http://www.spine-health.com/treatment/artificial-disc-replacement/lumbar-artificial-disc-surgery-chronic-back-pain>.