

A COMPARITIVE STUDY ON IMMOBILISED MYCELIUM AND SPORES OF  
*P. CHYRISOSPORIUM* IN PVA-ALGINATE-SULFATE BEADS FOR TEXTILE  
DYES EFFLUENT TREATMENT.

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*Specially dedicated to To my beloved family:*

*Espindada and Ami*

*Shah Khalid*

*Shah Hassan*

*Naveed-u-llah khan*

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*Azan*

*And fiancée*

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## ABSTRACT

Effluents discharging from the textile and dye industries to the neighbouring water are causing serious health concern to the environment and are getting attention of water regulatory agencies. Among the recent proposed treatment methods for treatment of textile effluents biological treatment has gain more attention because of its cheap and effective approach. Biotreatment with white rot fungi seems to be a viable option in the existing biological treatment process. This study, investigates the treatment of textile effluents with immobilized spores and mycelium of *Phanerochaete chrysosporium* separately in PVA-alginate-sulfate beads. Screening for the dye discoloration was done by using Design Expert 9.0.3.1. Screening process was conducted by using a two level factorial programmed with three factors namely temperature, number of beads loaded and initial dye concentration. The responses namely colour and COD reduction, enzymatic activities, reusability, storage stability and toxicity were thoroughly investigated to determine the system efficiency. Results revealed that optimum colour reduction and enzyme activity was achieved in immobilized spores compared to immobilize mycelium at 37 °C, 10gm beads and 300 mg/L dye concentration also the enzyme activity was comparatively high in immobilized spores as compared to immobilized mycelium. The reusability test also revealed that the immobilized fungus could be reused for up to 5 times to treat dye effluents. Toxicity test also proved the ability of immobilized cells in reduction of toxicity level. In conclusion, spores and mycelium were successfully immobilized in PVA-alginate-sulfate beads and they both serve as a potential mean and methods for treating textile dye effluents.

## ABSTRAK

Sisa buangan daripada industri tekstil dan pewarna terhadap air persekitaran akan menyebabkan masalah kesihatan yang serius dan ianya telah mendapat perhatian daripada agensi kawal selia air. Antara kaedah rawatan terbaru yang telah dicadangkan ialah melalui rawatan biologi kerana ianya murah dan berkesan. Rawatan menggunakan kulat reput putih merupakan pilihan yang baik bagi proses rawatan secara biologi yang sedia ada. Projek ini mengkaji rawatan sisa buangan tekstil menggunakan spora dan miselium daripada *Phanerochaete chrysosporium* yang telah disekat gerak secara berasingan dalam manik PVA-alginat-sulfat. Saringan bagi perubahan warna sisa buangan pewarna dilakukan dengan menggunakan Design Expert 9.0.3.1. Proses pemeriksaan telah dijalankan dengan menggunakan tahap dua factorial yang diprogramkan bersama tiga faktor iaitu suhu, jumlah manik yang digunakan dan kepekatan pewarna. Tindak balas dikaji melalui warna dan pengurangan COD, aktiviti enzim, penggunaan semula, kestabilan penyimpanan dan ketoksikan. Ianya telah dikaji dengan teliti bagi menentukan kecekapan sistem yang digunakan. Hasil kajian menunjukkan bahawa pengurangan warna dan enzim aktiviti yang optimum telah dicapai menggunakan spora tersekat gerak berbanding miselium tersekat gerak pada keadaan 37 °C, 10 g manik dan kepekatan pewarna pada 300 mg / L dimana aktiviti enzim adalah agak tinggi pada spora tersekat gerak berbanding miselium tersekat gerak. Ujian kebolehgunaan manik juga mendedahkan bahawa kulat tersekat gerak boleh digunakan sehingga 5 kali bagi merawat sisa buangan pewarna. Ujian ketoksikan juga membuktikan keupayaan sel yang tersekat gerak terhadap pengurangan tahap ketoksikan. Kesimpulannya, spora dan miselium telah berjaya disekat gerak pada manik PVA-alginat-sulfat dan ianya berpotensi bagi merawat sisa buangan tekstil dan pewarna.

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## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of Study**

Textile, leather, paper, and food industries are widely using dyes as a colouring agent (Ali and El-Mohamedy, 2012). The textile industry is a diverse sector in terms of production of raw materials, operating processes, product development, and equipment. Textile industries are well-characterized for consuming large quantities of water, energy, and discharging high volumes of waste into public sewage treatment plants. Textile industry uses high volumes of water from the beginning of washing of fibbers until washing of finished products. Subsequently, produces enormous volumes of wastewater which also contaminated with variety of chemicals including Azo dyes, formaldehyde, dioxins and heavy metals such as copper and chromium that might harm human and environment (Le Marechal *et al.*, 2012; Solís *et al.*, 2012). So without prior treatment of dyes effluents before discharging to water would effects the sunlight penetration negatively which in turn will slow down the photosynthesis process and will alter the gas solubility in water (Egli, 2007).

To treat the textiles effluents various techniques had been proposed (Rodríguez Couto, 2009). Biological method gain the attention in textile effluent treatment because of its no deteriorating effect on the environment (Sani *et al.*, 1998). Basidiomycetes fungi is commonly use in biological method to degrade wide range of different synthetic dyes.

The ability of fungi to rapidly adapt their metabolism to varying carbon and nitrogen sources is important for their survival. This metabolic activity is achieved through the production of a large set of intra and extracellular enzymes able to degrade various complex organic pollutants such as polyaromatic hydrocarbons, organic waste, dye effluents and steroid compounds (Pointing, 2001) (Glenn and Gold, 1983) (Cripps *et al.*, 1990). In the past few decades white-rot fungi has been evaluated for their ability to degrade polyaromatic hydrocarbons and discoloration of textile synthetic dyes (Nozaki *et al.*, 2008; Reddy, 1995; Wesenberg *et al.*, 2003).

The dye discolorization rate and effectiveness through microorganisms can be enhanced by cell immobilization (Nicell, 2003). Cell in Immobilized form gives better operational stability and higher efficiency (Iqbal and Saeed, 2007). The immobilization matrix stimulate ligninolytic enzymes production by providing a surrounding that mimics cell natural habitat, depending on the nature of support matrix (Suhaimi, 2010)

Polyvinyl alcohol (PVA) can be used for cell immobilization because recent researches had showed superior mechanical strength and chemical stability of the PVA- immobilized biomass (Idris *et al.*, 2008; Iqbal and Saeed, 2007) . PVA has these unique properties that it is highly stable, non-toxic to organism and can be produced cheaply at industrial scale (Lozinsky and Plieva, 1998). This study aims to investigate and compare the combine advantages offered by PVA, spores and mycelium of white rot fungi to treat the textile effluents.

## 1.2 Problem Statement

The textile industries uses dyes which are designed to be recalcitrant and resistant when expose to light, sweat, and water, chemical and microbial attack for a better quality in order to fulfill the needs of humans. Dye color found in effluents would directly give rise to water pollution, and discharged of such highly concentrated contaminant can cause toxic, mutagenic and carcinogenic effects (Chung *et al.*, 1992). In textile industries during processing, about 40% of used dyes are released into the water (Faraco *et al.*, 2009). The dyes which are used in textile might contain grease, wax, heavy metal, surfactants and suspended solid (Ahmad *et al.*, 2012). So without prior treatment of dyes effluents before discharging to water would effects the sunlight penetration negatively which in turn will slow down the photosynthesis process and growth of aquatic biota and will alter the gas solubility in water (Egli, 2007).

Alternative chemical and physical technologies have been reported for cleanup of contaminated textile waste water. But these techniques have certain limitations such as most of them are expensive and produce a huge amount of sludge (Robinson *et al.*, 2001). Thus, to overcome these limitations and problems a quick and cost effective approach is needed.

## 1.3 Objectives

The specific objectives of this research are as follows:

- (a) To immobilize spore and mycelium of *P. chrysosporium* in PVA-alginate-sulfate beads.

- (b) To treat textile dyes effluent using immobilized spore and mycelium of *P. chrysosporium* and to compare the degradability ability of both.
- (c) To optimize the physical parameter for textile effluents treatment using designs expert method.

#### **1.4 Scope of Research**

This research study investigates the *Phanerochaete chrysosporium* spores and mycelium's ability to decolorize textile dye effluents. PVA-alginate-sulfate beads used were a modified version from previous works (Idris *et al.*, 2008)

To test and compare the decolorizing and degrading ability of immobilized mycelium and spores different test were performed such as American Dyes Manufacturer's Institute (ADMI) reduction, Chemical Oxygen Demand (COD) and enzymatic activities. To optimize the physical parameters for the immobilized spores and mycelium a statistical tool Design Expert 9.0.3 was used. For examining the morphology of immobilized mycelium and spores of *Phanerochaete chrysosporium* Scanning Electron Microscope (SEM) was used. For ensuring the effectiveness of the immobilized matrix we also performed the reusability, toxicity storage stability and cell leakage tests.

#### **1.5 Significant of Research**

This study highlights the immobilization of spores and mycelium of *Phanerochaete chrysosporium* in PVA-alginate-sulfate beads in treating textile

effluents. Previously researchers have used this immobilization matrix, successfully for immobilizing an enzymes (Idris *et al.*, 2008), and *Phanerochaete chrysosporium* (Idris *et al.*, 2013) and it enhanced the beads' shapes, resulting the best surface area for the cells, reduce cell leakage and cells agglomeration. This study will ascertain between immobilized spores and mycelium of *Phanerochaete chrysosporium* which one will be more able to increase the discolorization rate compared to one another.

The research if successfully accomplished will provide a better understanding of treating the dye effluent of textile waste water and possibly can be add catalogue of the already identified biological treatment methods for dye effluent can serve as seeds for bio-augmentation in biodegradation of textile waste..

## REFERENCES

- Ahmad, A. L., Harris, W. A., and Ooi, B. S. (2012). Removal of dye from wastewater of textile industry using membrane technology. *Jurnal Teknologi*, 36(1), 31–44.
- Aksu, Z. (2003). Reactive dye bioaccumulation by *Saccharomyces cerevisiae*. *Process Biochemistry*, 38(10), 1437-1444.
- Aksu, Z., Kılıç, N. K., Ertuğrul, S., and Dönmez, G. (2007). Inhibitory effects of chromium(VI) and Remazol Black B on chromium(VI) and dyestuff removals by *Trametes versicolor*. *Enzyme and Microbial Technology*, 40(5), 1167-1174.
- Ali, N., and El-Mohamedy, R. (2012). Microbial decolourization of textile waste water. *Journal of Saudi Chemical Society*, 16(2), 117-123.
- Archibald, F. S., Bourbonnais, R., Jurasek, L., Paice, M. G., and Reid, I. D. (1997). Kraft pulp bleaching and delignification by *Trametes versicolor*. *Journal of Biotechnology*, 53(2–3), 215-236.
- Asgher, M., Kausar, S., Bhatti, H. N., Hassan Shah, S. A., and Ali, M. (2008a). Optimization of medium for decolorization of Solar golden yellow R direct textile dye by *Schizophyllum commune* IBL-06. *International Biodegradation and Biodegradation*, 61(2), 189-193.
- Asgher, M., Kausar, S., Bhatti, H. N., Shah, S. A. H., and Ali, M. (2008b). Optimization of medium for decolorization of Solar golden yellow R direct

- textile dye by *Schizophyllum commune* IBL-06. *International Biodegradation & Biodegradation*, 61(2), 189-193.
- Azmi, W., Sani, R. K., and Banerjee, U. C. (1998). Biodegradation of triphenylmethane dyes. *Enzyme and Microbial Technology*, 22(3), 185-191.
- Babu, B. R., Parande, A. K., Raghu, S., and Prem Kumar, T. (2007). Cotton textile processing: Waste generation and effluent treatment. *Journal of Cotton Science*, 11(3), 141-153.
- Banat, I. M., Nigam, P., Singh, D., and Marchant, R. (1996). Microbial decolorization of textile-dyecontaining effluents: A review. *Bioresource Technology*, 58(3), 217-227.
- Bishnoi, K., Kumar, R., and Bishnoi, N. R. (2008). Biodegradation of polycyclic aromatic hydrocarbons by white rot fungi *Phanerochaete chrysosporium* in sterile and unsterile soil. *J Sci Ind Res India*, 67, 538-542.
- Cerdeira, A., Russo, C., and Marques, M. (2009). Electroflocculation for textile wastewater treatment. *Brazilian Journal of Chemical Engineering*, 26(4), 659-668.
- Cestari, A. R., Vieira, E. F. S., Nascimento, A. J. P., de Oliveira, F. J. R., Bruns, R. E., and Airolidi, C. (2001). New Factorial Designs to Evaluate Chemisorption of Divalent Metals on Aminated Silicas. *Journal of Colloid and Interface Science*, 241(1), 45-51.
- Chen, C., Chen, J. K., Ni, W., Tian, X. J., and Huang, F. (2008). Biodegradation of orange G by wood-rot fungi *Phanerochaete sordida* TXJ-1302A and *Tyromyces lauteus* TXJ-1302B. *Bioresource Technology*, 99(9), 3926-3929.
- Chen, K.-C., and Lin, Y.-F. (1994). Immobilization of microorganisms with phosphorylated polyvinyl alcohol (PVA) gel. *Enzyme and Microbial Technology*, 16(1), 79-83.

- Chung, K.-T., Stevens, S. E., and Cerniglia, C. E. (1992). The reduction of azo dyes by the intestinal microflora. *Critical reviews in microbiology*, 18(3), 175-190.
- Coulibaly, L., Gourene, G., and Agathos, N. S. (2004). Utilization of fungi for biotreatment of raw wastewaters. *African Journal of Biotechnology*, 2(12), 620-630.
- Cripps, C., Bumpus, J. A., and Aust, S. D. (1990). Biodegradation of azo and heterocyclic dyes by *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*, 56(4), 1114-1118.
- Cui, F., and Dolphin, D. (1990). The role of manganese in model systems related to lignin biodegradation.
- Cullen, D. (1997). Recent advances on the molecular genetics of ligninolytic fungi. *Journal of Biotechnology*, 53(2), 273-289.
- Daniel, G., Pettersson, B., Nilsson, T., and Volc, J. (1990). Use of immunogold cytochemistry to detect Mn(II)-dependent and lignin peroxidases in wood degraded by the white rot fungi *Phanerochaete chrysosporium* and *Lentinula edodes*. *Canadian Journal of Botany*, 68(4), 920-933.
- Dittmer, J. K., Patel, N. J., Dhawale, S. W., and Dhawale, S. S. (1997). Production of multiple laccase isoforms by *Phanerochaete chrysosporium* grown under nutrient sufficiency. *FEMS Microbiology Letters*, 149(1), 65-70.
- Egli, J. (2007). Wastewater treatment in the textile industry. *Dyeing Printing Finishing*, 10(2007), 60-66.
- Eichlerova, I., Homolka, L., and Nerud, F. (2006). Synthetic dye decolorization capacity of white rot fungus *Dichomitus squalens*. *Bioresource Technology*, 97(16), 2153-2159.

- Eichlerova, I., Homolka, L., and Nerud, F. (2007). Decolorization of high concentrations of synthetic dyes by the white rot fungus *Bjerkandera adusta* strain CCBAS 232. *Dyes and Pigments*, 75(1), 38-44.
- Faraco, V., Pezzella, C., Miele, A., Giardina, P., and Sannia, G. (2009). Bio-remediation of colored industrial wastewaters by the white-rot fungi *Phanerochaete chrysosporium* and *Pleurotus ostreatus* and their enzymes. *Biodegradation*, 20(2), 209-220.
- Fitzgerald, S. W., and Bishop, P. L. (1995). Two stage anaerobic/aerobic treatment of sulfonated azo dyes. *Journal of Environmental Science & Health Part A*, 30(6), 1251-1276.
- Forgacs, E., Cserháti, T., and Oros, G. (2004). Removal of synthetic dyes from wastewaters: a review. *Environment International*, 30(7), 953-971.
- Fu, Y., and Viraraghavan, T. (2001). Removal of C.I. acid blue 29 from an aqueous solution by *Aspergillus niger*. *AATCC Magazine*, 1(1), 36-40.
- Galhaup, C., Wagner, H., Hinterstoisser, B., and Haltrich, D. (2002). Increased production of laccase by the wood-degrading basidiomycete *Trametes pubescens*. *Enzyme and Microbial Technology*, 30(4), 529-536.
- Galli, E., Di Mario, F., Rapanà, P., Lorenzoni, P., and Angelini, R. (2003). Copper biosorption by *Auricularia polytricha*. *Letters in Applied Microbiology*, 37(2), 133-137.
- Ghoreishi, S. M., and Haghghi, R. (2003). Chemical catalytic reaction and biological oxidation for treatment of non-biodegradable textile effluent. *Chemical Engineering Journal*, 95(1), 163-169.
- Glenn, J. K., and Gold, M. H. (1983). Decolorization of several polymeric dyes by the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*, 45(6), 1741-1747.

- Glenn, J. K., and Gold, M. H. (1985). Purification and characterization of an extracellular Mn (II)-dependent peroxidase from the lignin-degrading basidiomycete, *Phanerochaete chrysosporium*. *Archives of Biochemistry and Biophysics*, 242(2), 329-341.
- Gold, M. H., and Alic, M. (1993). Molecular biology of the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Microbiological Reviews*, 57(3), 605-622.
- Gold, M. H., Kuwahara, M., Chiu, A. A., and Glenn, J. K. (1984). Purification and characterization of an extracellular H<sub>2</sub>O<sub>2</sub>-requiring diarylpropane oxygenase from the white rot basidiomycete, *Phanerochaete chrysosporium*. *Archives of Biochemistry and Biophysics*, 234(2), 353-362.
- Gold, M. H., Wariishi, H., and Valli, K. (1989). *Extracellular peroxidases involved in lignin degradation by the white rot: Basidiomycete Phanerochate chrysosporium*. Paper presented at the ACS Symposium Series, 127-140.
- Gold, M. H., Youngs, H. L., and Gelpke, M. D. (2000). Manganese peroxidase. *Metal ions in biological systems*, 37, 559-586.
- Gomes, A., Brás, R., Ferra, M., Amorim, M., and Porter, R. (2000). Biological treatment of effluent containing textile dyes. *Coloration Technology*, 116(12), 393-397.
- Haemmerli, S. D., Schoemaker, H. E., Schmidt, H. W. H., and Leisola, M. S. A. (1987). Oxidation of veratryl alcohol by the lignin peroxidase of *Phanerochaete chrysosporium* Involvement of activated oxygen. *FEBS Letters*, 220(1), 149-154.
- Haglund, C. (1999). Biodegradation of xenobiotic compounds by the white-rot fungus *Trametes trogii*. *Molecular Biotechnology Programme*, 30.

- Hakala, T., Hildén, K., Maijala, P., Olsson, C., and Hatakka, A. (2006). Differential regulation of manganese peroxidases and characterization of two variable MnP encoding genes in the white-rot fungus *Physisporinus rivulosus*. *Applied Microbiology and Biotechnology*, 73(4), 839-849.
- Hatakka, A. (1994). Lignin-modifying enzymes from selected white-rot fungi: production and role from in lignin degradation. *FEMS Microbiology Reviews*, 13(2–3), 125-135.
- Heinfling, A., Martinez, M., Martinez, A., Bergbauer, M., and Szewzyk, U. (1998). Transformation of industrial dyes by manganese peroxidases from *Bjerkandera adusta* and *Pleurotus eryngii* in a manganese-independent reaction. *Applied and environmental microbiology*, 64(8), 2788-2793.
- Higson, F. K. (1991). Degradation of xenobiotics by white rot fungi. *Reviews of Environmental Contamination and Toxicology*, 122, 111-152.
- Hofrichter, M. (2002). Review: lignin conversion by manganese peroxidase (MnP). *Enzyme and Microbial Technology*, 30(4), 454-466.
- Hofrichter, M., Scheibner, K., Schneegass, I., and Fritsche, W. (1998). Enzymatic combustion of aromatic and aliphatic compounds by manganese peroxidase from *Nematoloma frowardii*. *Applied and Environmental Microbiology*, 64(2), 399-404.
- Idris, A., Suhaimi, M. S., Zain, N. A. M., Rashid, R., and Othman, N. (2013). Discoloration of aqueous textile dyes solution by *Phanerochaete chrysosporium* immobilized in modified PVA matrix. *Desalination and Water Treatment*(ahead-of-print), 1-9.
- Idris, A., Zain, N. A. M., and Suhaimi, M. S. (2008). Immobilization of Baker's yeast invertase in PVA-alginate matrix using innovative immobilization technique. *Process Biochemistry*, 43(4), 331-338.

- Ikehata, K., Buchanan, I. D., and Smith, D. W. (2004). Recent developments in the production of extracellular fungal peroxidases and laccases for waste treatment. *Journal of Environmental Engineering and Science*, 3(1), 1-19.
- Iqbal, M., and Saeed, A. (2007). Biosorption of reactive dye by loofa sponge-immobilized fungal biomass of *Phanerochaete chrysosporium*. *Process Biochemistry*, 42(7), 1160-1164.
- Jin, X.-C., Liu, G.-Q., Xu, Z.-H., and Tao, W.-Y. (2007). Decolorization of a dye industry effluent by *Aspergillus fumigatus* XC6. *Applied Microbiology and Biotechnology*, 74(1), 239-243.
- Johansson, T., and Nyman, P. O. (1996). A cluster of genes encoding major isozymes of lignin peroxidase and manganese peroxidase from the white-rot fungus *Trametes versicolor*. *Gene*, 170(1), 31-38.
- Kamei, I., Daikoku, C., Tsutsumi, Y., and Kondo, R. (2008). Saline-Dependent Regulation of Manganese Peroxidase Genes in the Hypersaline-Tolerant White Rot Fungus *Phlebia sp.* Strain MG-60. *Applied and Environmental Microbiology*, 74(9), 2709-2716.
- Kapdan, I. K., Kargia, F., McMullan, G., and Marchant, R. (2000). Effect of environmental conditions on biological decolorization of textile dyestuff by *C. versicolor*. *Enzyme and Microbial Technology*, 26(5-6), 381-387.
- Kashyap, P., Sabu, A., Pandey, A., Szakacs, G., and Soccol, C. R. (2002). Extracellular l-glutaminase production by *Zygosaccharomyces rouxii* under solid-state fermentation. *Process Biochemistry*, 38(3), 307-312.
- Kaushik, P., and Malik, A. (2009). Fungal dye decolourization: recent advances and future potential. *Environment International*, 35(1), 127-141.
- Kent Kirk, T., Tien, M., Kersten, P. J., Kalyanaraman, B., Hammel, K. E., and Farrell, R. L. (1990). [27] Lignin peroxidase from fungi: *Phanerochaete*

- chrysosporium*. In E. L. Mary (Ed.), *Methods in Enzymology* (Vol. Volume 188, pp. 159-171): Academic Press.
- Khalaf, M. A. (2008). Biosorption of reactive dye from textile wastewater by non-viable biomass of *Aspergillus niger* and *Spirogyra sp.* *Bioresource Technology*, 99(14), 6631-6634.
- Khindaria, A., Yamazaki, I., and Aust, S. D. (1995). Veratryl alcohol oxidation by lignin peroxidase. *Biochemistry*, 34(51), 16860-16869.
- Khoo, K.-M., and Ting, Y.-P. (2001a). Biosorption of gold by immobilized fungal biomass. *Biochemical engineering journal*, 8(1), 51-59.
- Khoo, K. M., and Ting, Y. P. (2001b). Biosorption of gold by immobilized fungal biomass. *Biochemical Engineering Journal*, 8(1), 51-59.
- Kiernan, J. (2001). Classification and naming of dyes, stains and fluorochromes. *Biotechnic & histochemistry*, 76(5-6), 261-278.
- Kirk, T. K., and Farrell, R. L. (1987). Enzymatic "Combustion": The Microbial Degradation of Lignin. *Annual Review of Microbiology*, 41(1), 465-501.
- Knapp, J., Newby, P., and Reece, L. (1995). Decolorization of dyes by wood-rotting basidiomycete fungi. *Enzyme and Microbial Technology*, 17(7), 664-668.
- Kuan, I. C., Johnson, K. A., and Tien, M. (1993). Kinetic analysis of manganese peroxidase: The reaction with manganese complexes. *Journal of Biological Chemistry*, 268(27), 20064-20070.
- Kumar, S. S., Balasubramanian, P., and Swaminathan, G. (2013). Degradation potential of free and immobilized cells of white rot fungus *Phanerochaete chrysosporium* on synthetic dyes. *Intl J Chem Tech Res*, 5, 565-571.

- Kumari, K., and Abraham, T. E. (2007). Biosorption of anionic textile dyes by nonviable biomass of fungi and yeast. *Bioresource Technology*, 98(9), 1704-1710.
- Kunamneni, A., Ballesteros, A., Plou, F. J., and Alcalde, M. (2007). Fungal laccase—a versatile enzyme for biotechnological applications. *Communicating current research and educational topics and trends in applied microbiology*, 1, 233-245.
- Le Marechal, A. M., Križanec, B., Vajnhandl, S., and Valh, J. V. (2012). *Textile finishing industry as an important source of organic pollutants*. Paper presented at the Organic Pollutants Ten Years After the Stockholm Convention-Environmental and Analytical Update.
- Liers, C., Ullrich, R., Steffen, K., Hatakka, A., and Hofrichter, M. (2006). Mineralization of <sup>14</sup>C-labelled synthetic lignin and extracellular enzyme activities of the wood-colonizing ascomycetes *Xylaria hypoxylon* and *Xylaria polymorpha*. *Applied microbiology and biotechnology*, 69(5), 573-579.
- Linko, S. (1992). Production of *Phanerochaete chrysosporium* lignin peroxidase. *Biotechnology Advances*, 10(2), 191-236.
- Liversidge, R. M., Lloyd, G. J., Wase, D. A. J., and Forster, C. F. (1997). Removal of Basic Blue 41 from aqueous solution by linseed cake. *Process Biochem.*, 32, 257-264.
- Lozinsky, V. I., and Plieva, F. M. (1998). Poly(vinyl alcohol) cryogels employed as matrices for cell immobilization. 3. Overview of recent research and developments. *Enzyme and Microbial Technology*, 23(3–4), 227-242.
- Maijala, P., Kleen, M., Westin, C., Poppius-Levlin, K., Herranen, K., Lehto, J. H., et al. (2008). Biomechanical pulping of softwood with enzymes and white-rot fungus *Physisporinus rivulosus*. *Enzyme and Microbial Technology*, 43(2), 169-177.

- Martinez, D., Larrondo, L. F., Putnam, N., Gelpke, M. D. S., Huang, K., Chapman, J., et al. (2004). Genome sequence of the lignocellulose degrading fungus *Phanerochaete chrysosporium* strain RP78. *Nat Biotech*, 22(6), 695-700.
- Martirani, L., Giardina, P., Marzullo, L., and Sannia, G. (1996). Reduction of phenol content and toxicity in olive oil mill waste waters with the ligninolytic fungus *Pleurotus ostreatus*. *Water Research*, 30(8), 1914-1918.
- McCurdy, M. W., Boardman, G. D., Michelsen, D. L. and Woodby, B. M. (1992). Chemical Reduction and Oxidation Combined with Biodegradation for the Treatment of a Textile Dye., 229-234.
- McGuirl, M. A., and Dooley, D. M. (1999). Copper-containing oxidases. *Current Opinion in Chemical Biology*, 3(2), 138-144.
- Miao, Y. J. (2011). Biological Remediation of Dyes in Textile Effluent : A review on Current Treatment Technology., (pp. 215-230).
- Mohd Zain, N. A., Mohd Suardi, S., and Idris, A. (2010). Hydrolysis of liquid pineapple waste by invertase immobilized in PVA-alginate matrix. *Biochemical Engineering Journal*, 50(3), 83-89.
- Moilanen, A., Lundell, T., Vares, T., and Hatakka, A. (1996). Manganese and malonate are individual regulators for the production of lignin and manganese peroxidase isozymes and in the degradation of lignin by *Phlebia radiata*. *Applied microbiology and biotechnology*, 45(6), 792-799.
- Moreira, M. T., Sierra-Alvarez, R., Lema, J. M., Feijoo, G., and Field, J. A. (2001). Oxidation of lignin in eucalyptus kraft pulp by manganese peroxidase from *Bjerkandera sp.* strain BOS55. *Bioresource Technology*, 78(1), 71-79.
- Mori, T., Kitano, S., and Kondo, R. (2003). Biodegradation of chloronaphthalenes and polycyclic aromatic hydrocarbons by the white-rot fungus *Phlebia lindneri*. *Applied Microbiology and Biotechnology*, 61(4), 380-383.

- Nakamura, Y., Sawada, T., Sungusia, M. G., Kobayashi, F., Kuwahara, M., and Ito, H. (1997). Lignin peroxidase production by *Phanerochaete chrysosporium* immobilized on polyurethane foam. *Journal of Chemical Engineering of Japan*, 30(1), 1-6.
- Nedovic, V., Leskosek-Cukalovic, I., and Vunjak-Novakovic, G. (1999). Immobilized cell technology (ict) in beer fermentation—a possibility for environmentally sustainable and cost-effective process. *University of Belgrade*.
- Nicell, J. A. (2003). McGill University, Montreal, Quebec, Canada. *Chemical Degradation Methods for Wastes and Pollutants: Environmental and Industrial Applications*, 384.
- Nie, G., Reading, N. S. and Aust, S. D. (1999). Relative Stability of Recombinant Versus Native Peroxidase from *Phanerochaete chrysosporium*. *Archives of Biochemistry and Biophysics*, 365,, 328-334.
- Nozaki, K., Beh, C. H., Mizuno, M., Isobe, T., Shiroishi, M., Kanda, T., et al. (2008). Screening and investigation of dye decolorization activities of basidiomycetes. *Journal of Bioscience and Bioengineering*, 105(1), 69-72.
- O'Neill, C., Hawkes, F. R., Hawkes, D. L., Lourenco, N. D., Pinheiro, H. M., and Delee, W. (1999). Colour in textile effluents—sources, measurement, discharge consents and simulation: a review. *Journal of Chemical Technology and Biotechnology*, 74(11), 1009-1018.
- Ohkuma M , M. Y., Johjima T & Kudo T (2001). Lignin degradation and roles of white rot fungi; study on an efficient symbiotic system in fungus-growing termites and its application to bioremediation. *RIKEN Rev Vol. 42*.

- Okino, L., Machado, K., Fabris, C., and Bononi, V. (2000). Ligninolytic activity of tropical rainforest basidiomycetes. *World Journal of Microbiology and Biotechnology*, 16(8-9), 889-893.
- Pandey, A., Singh, P., and Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *International Biodeterioration & Biodegradation*, 59(2), 73-84.
- Paszczynski, A., Huynh, V.-B., and Crawford, R. (1985). Enzymatic activities of an extracellular, manganese-dependent peroxidase from *Phanerochaete chrysosporium*. *FEMS Microbiology Letters*, 29(1), 37-41.
- Pazarlioglu, N. K., Urek, R. O., and Ergun, F. (2005). Biodecolourization of Direct Blue 15 by immobilized *Phanerochaete chrysosporium*. *Process Biochemistry*, 40(5), 1923-1929.
- Pointing, S. (2001). Feasibility of bioremediation by white-rot fungi. *Applied Microbiology and Biotechnology*, 57(1-2), 20-33.
- Radha, K., Regupathi, I., Arunagiri, A., and Murugesan, T. (2005). Decolorization studies of synthetic dyes using *Phanerochaete chrysosporium* and their kinetics. *Process Biochemistry*, 40(10), 3337-3345.
- Radha, K., Sridevi, V., and Kalaivani, K. (2009). Electrochemical oxidation for the treatment of textile industry wastewater. *Bioresource technology*, 100(2), 987-990.
- Rai, H. S., Bhattacharyya, M. S., Singh, J., Bansal, T. K., Vats, P., and Banerjee, U. C. (2005). Removal of Dyes from the Effluent of Textile and Dyestuff Manufacturing Industry: A Review of Emerging Techniques With Reference to Biological Treatment. *Critical Reviews in Environmental Science and Technology*, 35(3), 219-238.

- Reddy, C. A. (1995). The potential for white-rot fungi in the treatment of pollutants. *Current opinion in Biotechnology*, 6(3), 320-328.
- Robinson, T., McMullan, G., Marchant, R., and Nigam, P. (2001). Remediation of dyes in textile effluent: A critical review on current treatment technologies with a proposed alternative. *Bioresource Technology*, 77(3), 247-255.
- Rodríguez Couto, S. (2009). Dye removal by immobilised fungi. *Biotechnology Advances*, 27(3), 227-235.
- Sack, U., Hofrichter, M., and Fritzsche, W. (1997). Degradation of polycyclic aromatic hydrocarbons by manganese peroxidase of *Nematoloma frowardii*. *FEMS Microbiology Letters*, 152(2), 227-234.
- Sani, R., Azmi, W., and Banerjee, U. (1998). Comparison of static and shake culture in the decolorization of textile dyes and dye effluents by *Phanerochæte chrysosporium*. *Folia microbiologica*, 43(1), 85-88.
- Sani, R. K., and Banerjee, U. C. (1999). Decolorization of triphenylmethane dyes and textile and dye-stuff effluent by *Kurthiasp*. *Enzyme and Microbial Technology*, 24(7), 433-437.
- Saratale, G., Kalme, S., Bhosale, S., and Govindwar, S. (2007). Biodegradation of kerosene by *Aspergillus ochraceus* NCIM-1146. *Journal of basic microbiology*, 47(5), 400-405.
- Saritha, V., Maruthit, Y. A., and Mukkanti, K. (2009). Potential fungi for bioremediation of industrial effluents. *BioResources*, 5(1), 8-22.
- Schoemaker, H. E., Lundell, T. K., Hatakka, A. I., and Piontek, K. (1994). The oxidation of veratryl alcohol, dimeric lignin models and lignin by lignin peroxidase: The redox cycle revisited. *FEMS Microbiology Reviews*, 13(2–3), 321-331.

- Schoemaker, H. E., and Piontek, K. (1996). On the interaction of lignin peroxidase with lignin. *Pure and applied chemistry*, 68(11), 2089-2096.
- Shary, S., Ralph, S. A., and Hammel, K. E. (2007). New insights into the ligninolytic capability of a wood decay ascomycete. *Applied and environmental microbiology*, 73(20), 6691-6694.
- Shin, M., Nguyen, T., and Ramsay, J. (2002). Evaluation of support materials for the surface immobilization and decoloration of amaranth by *Trametes versicolor*. *Applied microbiology and biotechnology*, 60(1-2), 218-223.
- Singh, D., and Chen, S. (2008). The white-rot fungus *Phanerochaete chrysosporium*: conditions for the production of lignin-degrading enzymes. *Applied microbiology and biotechnology*, 81(3), 399-417.
- Singh, K., Nizam, S., Sinha, M., and Verma, P. K. (2012). Comparative transcriptome analysis of the necrotrophic fungus *Ascochyta rabiei* during oxidative stress: insight for fungal survival in the host plant. *PLoS one*, 7(3), e33128.
- Solís, M., Solís, A., Pérez, H. I., Manjarrez, N., and Flores, M. (2012). Microbial decolouration of azo dyes: A review. *Process Biochemistry*, 47(12), 1723-1748.
- Soloman, P. A., Basha, C. A., Velan, M., Ramamurthi, V., Koteeswaran, K., and Balasubramanian, N. (2009). Electrochemical Degradation of Remazol Black B Dye Effluent. *CLEAN – Soil, Air, Water*, 37(11), 889-900.
- Steffen, K. T., Hatakka, A., and Hofrichter, M. (2002). Removal and mineralization of polycyclic aromatic hydrocarbons by litter-decomposing basidiomycetous fungi. *Applied Microbiology and Biotechnology*, 60(1-2), 212-217.
- Sugiura, T., Yamagishi, K., Kimura, T., Nishida, T., Kawagishi, H., and Hirai, H. (2009). Cloning and homologous expression of novel lignin peroxidase genes in the white-rot fungus *Phanerochaete sordida* YK-624. *Bioscience, biotechnology, and biochemistry*, 73(8), 1793-1798.
- Suhaimi, M. S. (2010). Degradation of Textile Dyes Using Immobilized *Phanerochaete chrysosporium*.
- Sundaramoorthy, M., Kishi, K., Gold, M. H., and Poulos, T. L. (1997). Crystal structures of substrate binding site mutants of manganese peroxidase. *Journal of Biological Chemistry*, 272(28), 17574-17580.

- Šušla, M., Novotný, Č., Erbanová, P., and Svobodová, K. (2008). Implication of *Dichomitus squalens* manganese-dependent peroxidase in dye decolorization and cooperation of the enzyme with laccase. *Folia Microbiologica*, 53(6), 479-485.
- T. Moreira, M., Feijoo, G., Canaval, J., and M. Lema, J. (2003). Semipilot-scale bleaching of Kraft pulp with manganese peroxide. *Wood Science and Technology*, 37(2), 117-123.
- Takei, T., Ikeda, K., Ijima, H., and Kawakami, K. (2011). Fabrication of poly (vinyl alcohol) hydrogel beads crosslinked using sodium sulfate for microorganism immobilization. *Process Biochemistry*, 46(2), 566-571.
- ten Have, R., Hartmans, S., Teunissen, P. J., and Field, J. A. (1998). Purification and characterization of two lignin peroxidase isozymes produced by *Bjerkanderasp.* strain BOSS55. *FEBS letters*, 422(3), 391-394.
- Thurston, C. F. (1994). The structure and function of fungal laccases. *Microbiology*, 140(1), 19-26.
- Tien, M., and Kirk, T. K. (1983). Lignin-degrading enzyme from the hymenomycete *Phanerochaete chrysosporium* Burds. *Science(Washington)*, 221(4611), 661-662.
- Tien, M., and Kirk, T. K. (1984). Lignin-degrading enzyme from *Phanerochaete chrysosporium*: purification, characterization, and catalytic properties of a unique H<sub>2</sub>O<sub>2</sub>-requiring oxygenase. *Proceedings of the National Academy of Sciences*, 81(8), 2280-2284.
- Vaithanomsat, P., Apiwatanapiwat, W., Petchoy, O., and Chedchant, J. (2010). Production of Ligninolytic Enzymes by White-Rot Fungus *Datronia* sp. KAPI0039 and Their Application for Reactive Dye Removal. *International Journal of Chemical Engineering*, 2010, 6.

- Wang, B.-E., and Hu, Y.-Y. (2007). Comparison of four supports for adsorption of reactive dyes by immobilized *Aspergillus fumigatus* beads. *Journal of Environmental Sciences*, 19(4), 451-457.
- Wariishi, H., Akileswaran, L., and Gold, M. H. (1988). Manganese peroxidase from the basidiomycete *Phanerochaete chrysosporium*: Spectral characterization of the oxidized states and the catalytic cycle. *Biochemistry*, 27(14), 5365-5370.
- Wariishi, H., Valli, K., and Gold, M. H. (1992). Manganese(II) oxidation by manganese peroxidase from the basidiomycete *Phanerochaete chrysosporium*: Kinetic mechanism and role of chelators. *Journal of Biological Chemistry*, 267(33), 23688-23695.
- Wesenberg, D., Kyriakides, I., and Agathos, S. N. (2003). White-rot fungi and their enzymes for the treatment of industrial dye effluents. *Biotechnology Advances*, 22(1), 161-187.
- Wong, F. L., and Abdul-Aziz, A. (2008). Comparative study of poly(vinyl alcohol)-based support materials for the immobilization of glucose oxidase. *Journal of Chemical Technology and Biotechnology*, 83(1), 41-46.
- Wu, L., Ge, G., and Wan, J. (2009). Biodegradation of oil wastewater by free and immobilized *Yarrowia lipolytica* W29. *Journal of Environmental Sciences*, 21(2), 237-242.
- Yang, C.-C., Lue, S. J., and Shih, J.-Y. (2011). A novel organic/inorganic polymer membrane based on poly(vinyl alcohol)/poly(2-acrylamido-2-methyl-1-propanesulfonic acid/3-glycidyloxypropyl trimethoxysilane polymer electrolyte membrane for direct methanol fuel cells. *Journal of Power Sources*, 196(10), 4458-4467.
- Zain, N. A. M., Suhaimi, M. S., and Idris, A. (2011). Development and modification of PVA-alginate as a suitable immobilization matrix. *Process Biochemistry*, 46(11), 2122-2129.

- Zhang, L.-S., Wu, W.-z., and Wang, J.-l. (2007). Immobilization of activated sludge using improved polyvinyl alcohol (PVA) gel. *Journal of environmental sciences*, 19(11), 1293-1297.
- Zhao, X., and Hardin, I. R. (2007). HPLC and spectrophotometric analysis of biodegradation of azo dyes by *Pleurotus ostreatus*. *Dyes and Pigments*, 73(3), 322-325.