ISOLATION AND CHARACTERISATION OF MITOCHONDRIAL PROTEIN FROM *Orthosiphon stamenius* IN NORMAL AND COPPER STRESS CONDITIONS

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Specially dedicated to my beloved husband, ibu, ayah, daughters, son, family members and friends

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ABSTRACT

Orthosiphon stamenius is a species of medicinal herb that can be found in tropical area. Traditionally, the leaves of this plant have been used as diuretics, treatment of rheumatism, kidney and bladder inflammation, as well as abdominal pain and gout. This herbaceous plant requires in-depth research not only for medical field, but also to understand their properties during normal and stress environment. The purpose of this study is initially to search for the most suitable technique for mitochondrial isolation from Orthosiphon stamenius leaves. Three protocols for isolation of mitochondria were performed and the mitochondrial enzyme activities were determined. The highest activity of Cytochrome c Oxidase (1.423 U/g) and the highest mitochondrial protein concentration (2.656 mg/ml) was obtained from mitochondrial isolation using a method from Millar et al. (2007). This protocol was further used for mitochondrial isolation from Orthosiphon stamenius leaves treated with different concentrations of Copper (20 ppm, 60 ppm, 140 ppm). The activity of Cytochrome c Oxidase (4.217 U/g, 4.429 U/g, 4.887 U/g) and the concentration of mitochondrial protein (2.604 mg/ml, 2.623 mg/ml, 4.887 mg/ml) showed significant increase in 20 ppm, 60 ppm and 140 ppm Copper treatment, respectively. Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) has been conducted to check the mitochondrial protein profiles in normal and Copper treated plants. There was no obvious difference obtained in the protein profiles from normal and treated plants. There were six bands clearly visible on the gel for untreated and treated sample. Even though the protein profiles were similar, the increasing activity of Cytochrome c Oxidase and concentration of mitochondrial protein in Copper treated plants showed that plants do response under Copper stress. This study showed interconnected responses of mitochondrial activity and mitochondrial protein when exposed to different level of Copper.

ABSTRAK

Orthosiphon stamenius adalah tumbuhan herba yang biasanya digunakan dalam bidang perubatan tradisional. Ianya boleh didapati di kawasan tropika. Secara tradisi, daun tumbuhan ini boleh digunakan untuk merawat penyakit sendi, buah pinggang, keradangan pundi kencing, sakit perut dan gout. Kajian yang terpeinci mengenai tumbuhan ini diperlukan memandangkan bukan sahaja kerana kepentingannya dalam bidang perubatan malah pemahaman tentang tindakbalas tumbuhan ini dalam keadaan stress dan normal juga perlu diberi perhatian. Tujuan kajian ini adalah untuk memilih satu teknik terbaik untuk pengasingan mitokondria dari daun Orthosiphon stamenius. Tiga teknik untuk pengasingan mitokondria telah dijalankan ke atas daun Orthosiphon stamenius. Aktiviti tertinggi Cytochrome c Oxidase (1.423 U/g) dan kepekatan protein mitokondria tertinggi (2.656 mg/ml) telah diperolehi dari pengasingan mitokondria dari Millar et al. (2007). Teknik ini telah dijalankan ke atas daun Orthosiphon stamenius yang ditambah dengan kepekatan kuprum yang berbeza (20 ppm, 60 ppm, 140 ppm). Aktiviti Cytochrome c Oxidase (4.217 U/g, 4.429 U/g, 4.887 U/g) dan kepekatan protein mitokondria (2.604 mg/ml, 2.623 mg/ml, 4.887 mg/ml) menunjukkan peningkatkan jumlah pada kepekatan kuprum 20 ppm, 60 ppm dan 140 ppm. Kajian ini juga mengunakan natrium sulfat dodecyl polyacrylamide gel elektroforesis (SDS-PAGE) untuk menganalisa profil protein mitokondria dalam keadaan normal dan keadaan ditambah kuprum. Profil protein yang diperolehi adalah sama. Terdapat enam jalur yang terhasil dalam gel bagi sampel yang normal dan sampel yang ditambah kuprum. Walaupun profil protein adalah sama, peningkatan aktiviti Cytochrome c Oxidase dan kepekatan protein mitokondria adalah saling berkaitan dengan peningkatan kepekatan kuprum. Kajian ini menunjukkan terdapat tindak balas mitokondria dan protein mitokondria apabila terdedah kepada beberapa tahap kepekatan kuprum.

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LIST OF ABBREVIATIONS AND SYMBOLS

ATP	-	Adenosine Tri Phosphate
BN	-	Blue Native
BSA	-	Bovine serum albumin
Cu	-	Copper
CuSO.5H ₂ O	-	Copper (II) sulphate pentahydrate
DNA	-	Deoxyribonucleic Acid
DTT	-	Dithiotheitol
EGTA	-	ethylene glycol-bis-(2-aminoethylether)-n, n'tetraacetic acid
et al.,	-	and Others
Fe	-	Ferum
g	-	Gram
GDC	-	Glycine Decarboxylase Complex
H ₂ O	-	Water
HCl	-	Hydrogen Chloride
HSP	-	Heat Shock Protein
kDa	-	kilo Dalton
М	-	Molar
MCO	-	Metal catalysed oxidation
mg	-	Milligram
min	-	Minute
mL	-	Millilitre
mM	-	Millimolar
MOPS-KOH	-	3-(N-morpholino) propanesulfonic acid-Potassium hydroxide
MPP	-	Mitochondrial Processing Peptidase
nm	-	nanometre

OPGDC	-	2-Oxoglutarete Dehydrogenase Complex
PDC	-	Pyruvate Dehydrogenase Complax
PVP 40	-	Polyvinylpyrrolidone
QCR	-	Cytochrome c Reductase
RNA	-	Ribonucleic Acid
ROS	-	Reactive oxygen species
SDH	-	Succinate Dehydrogenase
SDS	-	Sodium dodecyl sulfate
SDS-PAGE	-	Sodium dodecyl sulfate-polyacrylamide gel
SPSS	-	Statistical Package for the Social Sciences
STDEV	-	Standard deviation
TEMED	-	N,N,N',N'-tetramethylethane-1,2-diamine
TES	-	n-tris (hydroxymethyl) methyl-2-aminoethane-sulphonic acid
ТОМ	-	Translocate of the Outer Mitochondrial Membrane
V	-	Volume
W	-	Weight
Δ	-	Delta
3	-	Epsilon
°C	-	Degree Celsius
μg	-	Microgram
μL	-	Microlitre
μΜ	-	Micromolar

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CHAPTER 1

INTRODUCTION

1.1 Research Background

Mitochondria are important organelles involved in Adenosine Tri Phosphate (ATP) production and many other functions in living organisms. They are found in the cytoplasm and occupy about 20% of the cell's volume (Douce, 1985). The number of mitochondria in a cell depends upon the metabolic requirements of that cell, and may range from a single large mitochondrion to thousands of the organelles (Mader and Windelspecht, 2010). According to Douce (1985), the most common shape for plant mitochondria is that of a rod with hemispherical ends about $2\mu m$ in length and $0.5\mu m$ in diameter. Mitochondria are often called the powerhouses of the cell because they produce most of the ATP utilized by the cell (Mader and Windelspecht, 2010).

Mitochondrion is a double membrane-surrounded organelle, in which cellular respiration takes place. The enzymes of the mitochondrial respiratory chain are located in the inner mitochondrial membrane and synthesize most of the cell's energy equivalents to ATP. Each mitochondrion contains a circular DNA molecule coding for ribosomal and mitochondrial transfer RNA molecules and for several mitochondrial proteins. The inner membrane is folded to cristae comprising the mitochondrial matrix space (Schwab, 2012). They contain enzymes for pyruvate

oxidation, the citric acid cycle, the β -oxidation of fatty acids, and oxidative phosphorylation, as well as the electron transport chain. The matrix of mitochondria is highly concentrated mixture of enzymes. Various enzymes and proteins present in the matrix of mitochondria, which helps in processing and break down carbohydrates and other nutrient molecules obtained from food (Mader and Windelspecht, 2010).

To prepare a crude mitochondrial fraction by differential centrifugations is not easy because some degree of structural damage has occurred during the isolation procedure (Moore *et al.*, 1993). That is why the isolation of plant mitochondria has always been complicated primarily by biochemical changes that occur during plant cell development and across plant species (Mackenzie, 1994). These techniques, while maintaining mitochondria morphological structure and their functional characteristics have required methods that avoid osmotic rupture of membranes that protect organelles from harmful products released from other cellular compartments (Millar et al., 2007), such as hydrolytic enzymes, phenolic compounds, alkaloids and terpenes (Neuberger, 1985). Most of the problems involved in the isolation of fully functional mitochondria occur during the homogenization phase because of the high shearing forces required to rupture the plant cell wall (Moore *et al.*, 1993). Thus the effects of homogenization on mitochondrial structure and function range from undesirable to totally destructive. The ability to isolate functional mitochondria from plant tissues is a key technique in the study of proteome and metabolic function of the plant mitochondrion (Sweetlove et al., 2007).

Recent advances in proteomic approaches led to the identification of large sets of mitochondrial proteins (Hajek *et al.*, 2004). According to Eubel (2007) one of purpose for mitochondrial isolation is to identify their proteomes and changes in these proteomes during development and environmental stress treatments. One of the threatening stresses is heavy metal. Transition metal ions are essential in biochemical function by being incorporated into or associating with proteins for living cell to functions effectively. For an example in plant mitochondria, key functions of metal cofactors include metabolism, electron transport, ATP synthesis and the detoxification of reactive oxygen species (ROS), however, metal ions can also be highly toxic to cells and cells organelle functions (Tan *et al.*, 2010). The high levels metals become toxic for the cell due to displacement of essential elements for enzymatic functions, interference with functional sites in protein, or enhanced ROS production (Garcia *et al.*, 2014).

The presence of free metal cations, redox active or inactive such as Copper may significantly contribute to the initiation of oxidative stress. Metal-catalysed oxidation (MCO) of protein is mechanisms for metal-linked damage which involved the oxidation of susceptible amino acids (Tan *et al.*, 2010) MCO of protein can be highly specific event when the site of protein oxidation can be defined on the protein surface that binds to metal ions and where proteins are more susceptible to damage if they bind metal ions (Stadman, 1990; Tan *et al.*, 2010). According to Gupta *et al.* (2012), mitochondrial enzymes often require metals as cofactors such Ferum (Fe) and Copper during electron transfer. However, higher concentrations of these essential metals induce ROS production. The presence of free metals ions in plant mitochondria could be crucial in the initiation of oxidative stress, resulting in oxidative damage to respiratory and other mitochondrial proteins.

1.2 Problem Statement

Mitochondria are important organelles that involved in ATP production and many other functions in organisms, including plants. Currently, development of techniques for the isolation of plant mitochondria lack far behind the progress made in the upstream processes such as protein and metabolite detection. As a common herbal plant, there are growing numbers of studies related to *Orthosiphon stamenius* in Malaysia. However, there is no attempt to develop a technique for mitochondrial isolation and analyse the protein profiles from this species yet.

- (i) To deduce the best method for mitochondrial isolation from *Orthosiphon stamenius*.
- (ii) To determine the activity of Cytochrome c Oxidase from isolated mitochondria.
- (iii) To evaluate mitochondrial protein obtained from *Orthosiphon stamenius* under normal and metal treatment conditions.

1.4 Scope of Study

In this research, *Orthosiphon stamenius* plants about three months old were obtained from local supplier. Then, leaves were harvested to obtain and isolate the mitochondria. The crude of mitochondria was isolated using by three different centrifugation methods. First centrifugation method used by Wilson and Chourey (1984). Second and third method used the modification of a protocol by Millar *et al.* (2007), where the mannitol was replaced by sucrose. Total mitochondrial protein quantification for each protocol was determined using Bradford method. The activity of Cytochrome c Oxidase for each samples from each isolation protocol were determined and protein profiles were analysed by Sodium Dodecyl Sulfate Polyacrylamide Gel-Electrophoresis (SDS-PAGE).

Another sample of *Orthosiphon stamenius* about two months old were also obtained from local supplier and was treated with three different concentration of Copper (CuSO.5H₂O). Then, the leaves were harvested after one month. The crude of mitochondria was isolated using the best method. Total mitochondrial protein quantification was determined using Bradford method. The activity of Cytochrome c

Oxidase weas determined and protein profiles were analysed by Sodium Dodecyl Sulfate Polyacrylamide Gel-Electrophoresis (SDS-PAGE).

1.5 Significance of Study

There are several techniques to isolate mitochondria from plants. Studies relating to such methods are imperative and these suitable techniques will determine the potential application their respective industries. By establishing a method for mitochondrial isolation from *Orthosiphon stamenius*, it will enhance our knowledge and open new possibilities for its application in biotechnology. Information on specific mitochondrial protein from the leaves under normal and metal treatment condition is important for further upstream studies related to mitochondrial functions in plants. Furthermore, this will help us to understand better on *Orthosiphon stamenius* response under metal stress.

REFERENCES

- Abdu, A., Aderis, N., Abdul-Hamid, H., Majid, N. M., Jusop, S., Karam, D. S., and Ahmad, K. (2011). Using *Orthosiphon stamineus* B. for Phytoremediation of Heavy Metals in Soils Amended with Sewage Sludge. *American Journal of Applied Sciences*. 8(4), 323-331.
- Appenroth, K. J. (2010). Definition of "heavy metals" and their role in biological systems. *Soil heavy metals*. (pp. 19-29). Berlin Heidelberg: Springer.
- Awale, S., Tezuka, Y., Banskota, A. H., and Kadota, S. (2003). Inhibition of NO production by highly-oxygenated diterpenes of *Orthosiphon stamineus* and their structure-activity relationship. *Biological and Pharmaceutical Bulletin*. 26(4), 468-47.
- Awale, S., Tezuka, Y., Banskota, A. H., Kouda, K., Tun, K. M., and Kadota, S. (2001). Five novel highly oxygenated diterpenes of *Orthosiphon stamineus* from Myanmar. *Journal of natural products*. 64(5), 592-596.
- Bardel, J., Louwagie, M., Jaquinod, M., Jourdain, A., Luche, S., Rabilloud, T., and Bourguignon, J. (2002). A survey of the plant mitochondrial proteome in relation to development. *Proteomics*. 2(7), 880-898.
- Bligny, R., and Douce, R. (1977). Mitochondria of isolated plant cells (*Acer pseudoplatanus* L.) II. Copper deficiency effects on cytochrome c oxidase and oxygen uptake. *Plant physiology*. 60(5), 675-679.

- Bollag, D. M., Edelstein, S., and Rozicky, M. (1996). *Proteins methods*. (2nd ed.)
 Jhon Wiley and Sons Publications.
- Bonjoch, N. P., and Tamayo, P. R. (2001). Protein content quantification by Bradford method. *Handbook of plant ecophysiology techniques* (pp. 283-295). Netherlands: Springer.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical biochemistry*. 72(1), 248-254.
- Braun, H. P., and Schmitz, U. K. (1999). The protein-import apparatus of plant mitochondria. *Planta*. 209(3), 267-274.
- Braun, H. P., Emmermann, M., Kruft, V., and Schmitz, U. K. (1992). Cytochrome c1 from potato: a protein with a presequence for targeting to the mitochondrial intermembrane space. *Molecular and General Genetics MGG*. 231(2), 217-225.
- Brumme, S., Kruft, V., Schmitz, U. K., and Braun, H. P. (1998). New Insights into the Co-evolution of Cytochrome cReductase and the Mitochondrial Processing Peptidase. *Journal of Biological Chemistry*. 273(21), 13143-13149.
- Burkill, I. H. (1966). A dictionary of the economic products of the Malay Peninsula. *A Dictionary of the Economic Products of the Malay Peninsula*. 2(2nd edition).
- Byczkowski, J. Z., and Sorenson, J. R. (1984). Effects of metal compounds on mitochondrial function: a review. *Science of the total environment*. 37(2), 133-162.
- Bykova, N. V., Stensballe, A., Egsgaard, H., Jensen, O. N., and Moller, I. M. (2003).
 Phosphorylation of formate dehydrogenase in potato tuber mitochondria. *Journal of Biological Chemistry*. 278(28), 26021-26030.

- Chin, J. H., Abas, H. H., and Sabariah, I. (2008). Toxicity study of Orthosiphon stamineus benth (misai kucing) on Sprague dawley rats. Trop Biomed. 25(1), 9-16.
- Cuypers, A., Koistinen, K. M., Kokko, H., Karenlampi, S., Auriola, S., and Vangronsveld, J. (2005). Analysis of bean (*Phaseolus vulgaris* L.) proteins affected by copper stress. *Journal of plant physiology*. 162(4), 383-392.
- Douce, R. (1985). *Mitochondria in Higher Plants: Structure, Function, and Biogenesis.* (1st ed.). London: Academic Press Inc. (London) Ltd.
- El-Aref, H. M., and Hamada, A. M. (1998). Genotypic differences and alterations of protein patterns of tomato explants under copper stress. *Biologia plantarum*. 41(4), 555-564.
- Eriksson, A., Sjoling, S., Glaser, E., Brennicke, A., and Kuck, U. (1993). A general processing proteinase of spinach leaf mitochondria is associated with the bc1 complex of the respiratory chain. *Plant mitochondria: with emphasis on RNA editing and cytoplasmic male sterility.* 1(1), 299-306.
- Eubel, H., Jansch, L., and Braun, H. P. (2003). New insights into the respiratory chain of plant mitochondria. Supercomplexes and a unique composition of complex II. *Plant physiology*. 133(1), 274-286.
- Eubel, H., Joshual, H., and Millar, A. H. (2007). Isolation and Subfractionation of Plant Mitochondria for Proteomic Analysis. In Thiellement, H., Zivy, M., Damerval, C., and V. Mechin (Ed.) Plant Proteomics. (pp. 49-62). Humana Press.
- Fang, W. C., and Kao, C. H. (2000). Enhanced peroxidase activity in rice leaves in response to excess iron, copper and zinc. *Plant Science*. 158(1), 71-76.
- Farnsworth, N. R., and Soejarto, D. D. (1991). Global importance of medicinal plants. *The conservation of medicinal plants*. 1(1), 25-51.

- Gallagher, C. H., and Reeve, V. E. (1976). Interrelationships of copper, cytochrome oxidase, phospholipid synthesis and adenine nucleotide binding. *Australian Journal of Experimental Biology and Medical Science*. 54(1), 593-600.
- Gallego, S. M., Benavídes, M. P., and Tomaro, M. L. (1996). Effect of heavy metal ion excess on sunflower leaves: evidence for involvement of oxidative stress. *Plant Science*. 121(2), 151-159.
- Garcia, L., Welchen, E., and Gonzalez, D. H. (2014). Mitochondria and copper homeostasis in plants. *Mitochondrion*. 19(1), 269-274.
- Giege, P., Sweetlove, L. J., and Leaver, C. J. (2003). Identification of mitochondrial protein complexes in Arabidopsis using two-dimensional blue-native polyacrylamide gel electrophoresis. *Plant molecular biology reporter*. 21(2), 133-144.
- Gooch, J. W. (2011). *Encyclopedic Dictionary of Polymers*. (2nd ed.). New York: Springer New York.
- Gualberto, J. M., Handa, H., and Grienenberger, J. M. (1995). Isolation and fractionation of plant mitochondria and chloroplasts: specific examples. *Methods in cell biology*. 50(1), 161.
- Gupta, D. K., and Sandalio, L. M. (2012). *Metal toxicity in plants: perception, signaling and remediation*. Germany: Springer.
- Hajek, T., Honys, D., and Capkova, V. (2004). New method of plant mitochondria isolation and sub-fractionation for proteomic analyses. *Plant Science*. 167(3), 389-395.
- Heazlewood, J. L., Howell, K. A., Whelan, J., and Millar, A. H. (2003). Towards an analysis of the rice mitochondrial proteome. *Plant Physiology*. 132(1), 230-242.

- Heazlewood, J. L., Tonti-Filippini, J. S., Gout, A. M., Day, D. A., Whelan, J., and Millar, A. H. (2004). Experimental analysis of the Arabidopsis mitochondrial proteome highlights signaling and regulatory components, provides assessment of targeting prediction programs, and indicates plant-specific mitochondrial proteins. *The Plant Cell Online*. 16(1), 241-256.
- Himani, B., Seema, B., Bhole, N., Mayank, Y., Vinod, S., and Mamta, S. (2013) Misai Kucing : A Glimpse of Maestro. *Int. J. Pharm Sci. Rev. Res.* 22(2) 55-59.
- Huang, S., Millar, A. H., and Taylor, N. L. (2011). The plant mitochondrial proteome composition and stress response: conservation and divergence between monocots and dicots. *Plant Mitochondria* (pp. 207-239). New York: Springer.
- Huang, S., Taylor, N. L., Narsai, R., Eubel, H., Whelan, J., and Millar, A. H. (2009). Experimental analysis of the rice mitochondrial proteome, its biogenesis, and heterogeneity. *Plant physiology*. 149(2), 719-734.
- Jaganath, I. B., and Ng, L. T. (2000). Herbs. The Green Pharmacy of Malaysia. Kuala Lumpur, Vinpress and Malaysia Agricultural Research and Development Institute. 1(1), 95-99.
- Jansch, L., Kruft, V., Schmitz, U. K., and Braun, H. P. (1995). Cytochrome c Reductase from Potato Does not Comprise Three Core Proteins but Contains an Additional Low-Molecular-Mass Subunit. *European journal of biochemistry*. 228(3), 878-885.
- Jansch, L., Kruft, V., Schmitz, U. K., and Braun, H. P. (1996). New insights into the composition, molecular mass and stoichiometry of the protein complexes of plant mitochondria. *The Plant Journal*. 9(3), 357-368.
- Jansch, L., Kruft, V., Schmitz, U. K., and Braun, H. P. (1998). Unique composition of the preprotein translocase of the outer mitochondrial membrane from plants. *Journal of Biological Chemistry*. 273(27), 17251-17257.

- Jones, C. G., Hare, J. D., and Compton, S. J. (1989). Measuring plant protein with the Bradford assay. *Journal of chemical ecology*. 15(3), 979-992.
- Kabata-Pendias, A. (2011). *Trace elements in soils and plants*. (4th ed.) New York: CRC press.
- Karpova, O. V., and Newton, K. J. (1999). A partially assembled complex I in NAD4-deficient mitochondria of maize. *The Plant Journal*. 17(5), 511-521.
- Keunen, E., Remans, T., Bohler, S., Vangronsveld, J., and Cuypers, A. (2011). Metalinduced oxidative stress and plant mitochondria. *International journal of molecular sciences*. 12(10), 6894-6918.
- Khatun, M. A., Harun-Or-Rashid, M., and Rahmatullah, M. (2011). Scientific Validation of Eight Medicinal Plants Used in Traditional Medicinal Systems of Malaysia: a Review. *American-Eurasian Journal of Sustainable Agriculture*. 5(1), 67-75.
- Kiong, A. L. P., Lai, A. G., Hussein, S., and Harun, A. R. (2008). Physiological responses of Orthosiphon stamineus plantlets to gamma irradiation. American-Eurasian Journal of Sustainable Agriculture. 2(2), 135-149.
- Klodmann, J., Senkler, M., Rode, C., and Braun, H. P. (2011). Defining the protein complex proteome of plant mitochondria. *Plant physiology*. 157(2), 587-598.
- Kruft, V., Eubel, H., Jansch, L., Werhahn, W., and Braun, H. P. (2001). Proteomic approach to identify novel mitochondrial proteins in *Arabidopsis*. *Plant Physiology*. 127(4), 1694-1710.
- Lanzotti, V. (2013). Diterpenes for Therapeutic Use. *Natural Products* (pp. 3173-3191). Berlin Heidelberg: Springer.
- Lithgow, T. (2000). Targeting of proteins to mitochondria. *FEBS letters*. 476(1), 22-26.

- Luna, C. M., Gonzalez, C. A., and Trippi, V. S. (1994). Oxidative damage caused by an excess of copper in oat leaves. *Plant and Cell Physiology*. 35(1), 11-15.
- Mackenzie, S. (1994). Isolation of plant mitochondria and mitochondrial nucleic acids. Gelvin, S., and Schilperoort, R. Plant Molecular Biology Manual. (pp. 203-214). Netherlands: Springer.
- Mader, S. S. and Windelspecht M. (2010). *Biology*. (11th ed.). United States of America: McGraw-Hill.
- Mariin-Hernandez, A., Gracia-Mora, I., Ruiz-Ramirez, L., and Moreno-Sanchez, R. (2003). Toxic effects of copper-based antineoplastic drugs (Casiopeinas) on mitochondrial functions. *Biochemical pharmacology*. 65(12), 1979-1989.
- Michel, H. (1998). The mechanism of proton pumping by cytochrome c oxidase. Proceedings of the National Academy of Sciences. 95(22), 12819-12824.
- Michel, H. (1999). Cytochrome c oxidase: Catalytic cycle and mechanisms of proton pumping-A discussion. *Biochemistry*. 38(46), 15129-15140.
- Miernyk, J. A., and Randall, D. D. (1987). Some properties of pea mitochondrial phospho-pyruvate dehydrogenase-phosphatase. *Plant physiology*. 83(2), 311-315.
- Millar, H., Considine, M. J., Day, D. A., and Whelan, J. (2001). Unraveling the role of mitochondria during oxidative stress in plants. *IUBMB life*. 51(4), 201-205.
- Millar, A. H., Trend, A. E., and Heazlewood, J. L. (2004). Changes in the mitochondrial proteome during the anoxia to air transition in rice focus around cytochrome-containing respiratory complexes. *Journal of Biological Chemistry*. 279(38), 39471-39478.

- Millar, A. H., Eubel, H., Jansch, L., Kruft, V., Heazlewood, J. L., and Braun, H. P. (2004). Mitochondrial cytochrome c oxidase and succinate dehydrogenase complexes contain plant specific subunits. *Plant molecular biology*. 56(1), 77-90.
- Millar, A. H. (2007). The plant mitochondrial proteome. *Plant Proteomics* (pp. 226-246). Berlin Heidelberg: Springer.
- Millar, A. H., Liddell, A. and Leaver, C. J. (2007). Isolation and Subfractionation of Mitochondria from Plants. In Liza, A. P. and Eric, A. S.(Ed.) Methods in Cell Biology. (pp. 65-90). Academic Press.
- Moller, I. M. (2001). Plant mitochondria and oxidative stress: electron transport, NADPH turnover, and metabolism of reactive oxygen species. *Annual review of plant biology*. 52(1), 561-591.
- Moore, A., Fricaud, A. C., Walters, A. J., and Whitehouse, D. G. (1993). Isolation and Purification of Functionally Intact Mitochondria from Plant Cells. In Graham, J., and Higgins, J. (Ed.) Biomembrane Protocols.(pp. 133-139). Humana Press.
- Neuburger, M. (1985). Preparation of plant mitochondria, criteria for assessement of mitochondrial integrity and purity, survival in vitro. *Higher plant cell respiration* (pp. 7-24). Berlin Heidelberg: Springer.
- Ninfa, A. J., and Ballou, D. P. (1998). Fundamental Laboratory Approaches for Biochemistry and Biotechnology: A Text with Experiments. Fitzgerald Science Press.
- Obata, T., Matthes, A., Koszior, S., Lehmann, M., Araujo, W. L., Bock, R., and Fernie, A. R. (2011). Alteration of mitochondrial protein complexes in relation to metabolic regulation under short-term oxidative stress in Arabidopsis seedlings. *Phytochemistry*. 72(10), 1081-1091.

- Palma, J. M., Gomez, M., Yanez, J., and Del Rio, L. A. (1987). Increased levels of peroxisomal active oxygen-related enzymes in copper-tolerant pea plants. *Plant physiology*. 85(2), 570-574.
- Prasad, M. N. V. (2004). *Heavy metal stress in plants: from biomolecules to ecosystems*. (2nd ed.), India: Springer.
- Prasad, T. K., and Stewart, C. R. (1992). cDNA clones encoding Arabidopsis thaliana and *Zea mays* mitochondrial chaperonin HSP60 and gene expression during seed germination and heat shock. *Plant molecular biology*. 18(5), 873-885.
- Sacchetti, G., Maietti, S., Muzzoli, M., Scaglianti, M., Manfredini, S., Radice, M., and Bruni, R. (2005). Comparative evaluation of 11 essential oils of different origin as functional antioxidants, antiradicals and antimicrobials in foods. *Food chemistry*. 91(4), 621-632.
- Schmidt, O., Pfanner, N., and Meisinger, C. (2010). Mitochondrial protein import: from proteomics to functional mechanisms. *Nature reviews Molecular cell biology*. 11(9), 655-667.
- Schwab, M. (2012). *Encyclopedia of Cancer*. (3th ed.). Berlin: Springer Berlin Heidelberg.
- Srivastava, J., Lambert, J., and Vietmeyer, N. (1996). *Medicinal plants: An expanding role in development*. (1st ed.) World Bank Publications.
- Stadtman, E. R. (1990). Metal ion-catalyzed oxidation of proteins: biochemical mechanism and biological consequences. *Free Radical Biology and Medicine*. 9(4), 315-325.
- Suddee, S., Paton, A. J., and Parnell, J. A. N. (2005). Taxonomic revision of tribe Ocimeae Dumort.(Lamiaceae) in continental South East Asia III. Ociminae. *Kew Bulletin*, 1(1), 3-75.

- Sun, W., Van Montagu, M., and Verbruggen, N. (2002). Small heat shock proteins and stress tolerance in plants. *Biochimica et Biophysica Acta (BBA)-Gene Structure and Expression*. 1577(1), 1-9.
- Sunderhaus, S., Klodmann, J., Lenz, C., and Braun, H. P. (2010). Supramolecular structure of the OXPHOS system in highly thermogenic tissue of *Arum maculatum*. *Plant Physiology and Biochemistry*. 48(4), 265-272.
- Sweetlove, L., Taylor, N., and Leaver, C. (2007). Isolation of Intact, Functional Mitochondria from the Model Plant Arabidopsis thaliana .In Leister, D., and Herrmann, J. Mitochondria. (pp. 125-136). Humana Press.
- Tan, Y. F., O'Toole N., Taylor, N. L., and Millar, A. H. (2010). Divalent metal ions in plant mitochondria and their role in interactions with proteins and oxidative stress-induced damage to respiratory function. *Plant physiology*. 152(2), 747-761.
- Tanyolac, D., Ekmekci, Y., and Unalan, S. (2007). Changes in photochemical and antioxidant enzyme activities in maize (*Zea mays* L.) leaves exposed to excess copper. *Chemosphere*. 67(1), 89-98.
- Vigani, G. (2012). Discovering the role of mitochondria in the iron deficiencyinduced metabolic responses of plants. *Journal of plant physiology*. 169(1), 1-11.
- Vinit-Dunand, F., Epron, D., Alaoui-Sosse, B., and Badot, P. M. (2002). Effects of copper on growth and on photosynthesis of mature and expanding leaves in cucumber plants. *Plant science*. 163(1), 53-58.
- Weber, K., and Osborn, M. (1969). The reliability of molecular weight determinations by dodecyl sulfate-polyacrylamide gel electrophoresis. *Journal* of Biological Chemistry. 244(16), 4406-4412.

- Werhahn, W., and Braun, H. P. (2002). Biochemical dissection of the mitochondrial proteome from *Arabidopsis thaliana* by three-dimensional gel electrophoresis. *Electrophoresis*. 23(4), 640-646.
- Werhahn, W., Jansch, L., and Braun, H. P. (2003). Identification of novel subunits of the TOM complex from *Arabidopsis thaliana*. *Plant Physiology and Biochemistry*. 41(5), 407-416.
- Wiart, C. (2002) Orthosiphon stamineus Benth. In Wong, F. K. (Ed.) Medicinal Plants of Southeast Asia (pp. 265), Prentice Hall, Kuala Lumpur.
- Wilson, A. J. and Chourey, P. S. (1984). A rapid inexpensive method for the isolation of restrictable mitochondrial DNA from varius plant sources. *Plant Cell Reports*. 3(6), 237-239.
- Wright, C. I., Van-Buren, L., Kroner, C. I., and Koning, M. M. G. (2007). Herbal medicines as diuretics: a review of the scientific evidence. *Journal of Ethnopharmacology*. 114(1), 1-31.
- Yamamoto, Y., and Gaynor, R. B. (2001). Therapeutic potential of inhibition of the NF-κB pathway in the treatment of inflammation and cancer. *The Journal of clinical investigation*. 107(2), 135-142.
- Yruela, I. (2005). Copper in plants. Brazilian Journal of Plant Physiology. 17(1), 145-156.
- Yruela, I. (2009). Copper in plants: acquisition, transport and interactions. Functional Plant Biology. 36(5), 409-430.
- Zaharah, A. (2005). Misai kucing (Orthosiphon stamineus). Penanaman tumbuhan ubatan & beraroma. 14(20), 57-62.