

BIOPROSPECTING FOR CULTURABLE PSYCHROTROPHIC BACTERIA
WITH EXTRACELLULAR HYDROLYTIC ENZYMES ACTIVITY FROM
ARCTIC LAKE SEDIMENTS

IDRIS SHEHU

UNIVERSITI TEKNOLOGI MALAYSIA

BIOPROSPECTING FOR CULTURABLE PSYCHROTROPHIC BACTERIA
WITH EXTRACELLULAR HYDROLYTIC ENZYMES ACTIVITY FROM
ARCTIC LAKE SEDIMENTS

IDRIS SHEHU

A dissertation submitted in partial fulfillment of the
requirements for the award of the degree of
Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering
Universiti Teknologi Malaysia

JANUARY 2015

I dedicate this work to my deceased father and mother: Alhaji Idris Yusuf and Fatima Idris. May Allah forgive their shortcomings and make Jannatul-Firdaus to be their final abode.

ACKNOWLEDGEMENT

My deepest gratitude goes to Almighty Allah for His unquantifiable favour and specifically for keeping me alive to witness this memorable time of my academic pursuit. First and foremost, I wish to express my sincere appreciation to my supervisor Assoc. Prof. Dr. Zaharah Binti Ibrahim, I remain forever grateful for her support, guidance, encouragement and more importantly constructive critics which have contributed immensely to the successful completion of this work. I would also want to acknowledge Assoc. Prof. Dr. Cheah Yoke Kqueen of Universiti Putra Malaysia (UPM) and Prof. Dr. Clement Michael Wong of University Malaysia Sabah (UMS) for providing the arctic samples used in the research. Special thanks and appreciation to my wife Rukayya Muhammad for her patience, understanding and support; you are indeed motivator, and to my children Abdulmajid, NanaFatima (Umma) and Yusuf; I love you all.

To all my senior colleagues in the Environmental Biotechnology Laboratory UTM: Dr. Ivy, Bashir S. Mienda, Ahmad Idi, Hanif, Fahmi, Lam and many others whose names are too numerous to mention, I thank you all. Also, to my friends, and family members whose encouragement inspired me to work harder, I am grateful for their advice and motivation without which this dissertation would not have been the same as presented here. I would also like to thank the developers of the UTM thesis template for making the dissertation writing process a lot easier for me. Those guys are unbelievably awesome! May almighty Allah bless and enrich each and every one for his or her contributions.

ABSTRACT

The ability of cold-adapted microorganisms to produce cold-active enzymes with potential biotechnological applications has recently attracted the attention of scientific community in terms of bioprospecting. This research aimed at assessing the presence and diversity of culturable psychrotrophic bacteria with hydrolytic enzymes activity and their identification using molecular approach. A total of six (6) different Arctic lake sediments were analysed. Mean viable bacterial count ranged from 2.88×10^3 to 5.07×10^5 cfu/g. A total of thirty seven (37) bacterial strains were successfully isolated at 20°C and screened for Amylase, Protease and Lipase activity. Molecular characterization using 16S rRNA gene sequence homology revealed that the isolated bacteria belong to seven genera comprising *Pseudomonas*, *Bacillus*, *Dermacoccus*, *Arthrobacter*, *Janthinobacterium*, *Paenibacillus* and *Chryseobacterium*. Eighteen (18) isolates; representing 49% were found to be positive to at least one of the three enzymes activity tested. Two different strains of *Pseudomonas sp.* (isolates 16D4 and 17D4) were found to be the most potent protease and lipase producing isolates respectively. XIA12 identified as *Bacillus cereus* was found to be the most potent amylase producing bacteria. Study of the growth temperature of the isolated bacteria revealed that most of isolates could grow at temperature above 20°C signifying that the isolates are true psychrotrophs. The findings of this work have confirmed that Arctic environment can serve as an ideal area for biotechnological exploration.

ABSTRAK

Baru-baru ini, keupayaan mikroorganisma sejuk untuk menghasilkan enzim sejuk-aktif yang mempunyai potensi dalam aplikasi bioteknologi telah menarik perhatian para saintifik dari segi bioprospek. Kajian ini bertujuan untuk menilai kehadiran dan kepelbagaian bakteria psychrotrophic yang boleh diternak dengan aktiviti enzim hidrolitik dan pengenalan bakteria menggunakan pendekatan molekul. Sebanyak enam (6) sedimen tasik Artik yang berbeza telah dianalisis. Purata kiraan bakteria yang boleh dicapai adalah di antara 2.88×10^3 hingga 5.07×10^5 cfu/g. Sebanyak tiga puluh tujuh (37) jenis bakteria telah berjaya diasingkan pada 20°C dan ditapis untuk aktiviti Amilase, Protease dan Lipase. Pencirian molekul menggunakan 16S rRNA homologi urutan gen mendedahkan bahawa bakteria yang telah diasingkan tergolong dalam tujuh genera yang terdiri daripada *Pseudomonas*, *Bacillus*, *Dermacoccus*, *Arthrobacter*, *Janthinobacterium*, *Paenibacillus* dan *Chryseobacterium*. Lapan belas (18) bakteria yang telah diasingkan; sebanyak 49% didapati positif untuk sekurang-kurangnya salah satu daripada tiga ujian aktiviti enzim. Dua jenis bakteria yang berbeza daripada golongan *Pseudomonas sp.* (pengasingan 16D4 dan 17D4) masing-masing didapati paling berkesan dalam menghasilkan protease dan lipase. XIA12 telah dikenal pasti sebagai *Bacillus cereus* sebagai bakteria yang paling berkesan dalam menghasilkan amilase. Kajian suhu pertumbuhan bakteria yang telah diasingkan mendedahkan bahawa sebahagian besar daripadanya boleh hidup pada suhu melebihi 20°C menandakan bahawa ianya adalah psychrotrophs yang benar. Hasil kerja ini telah mengesahkan bahawa persekitaran Artik boleh berfungsi sebagai kawasan yang sesuai untuk eksplorasi bioteknologi.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xi
	LIST OF FIGURES	xiii
	LIST OF ABBREVIATIONS AND SYMBOLS	xiv
	LIST OF APPENDICES	xvi
1	INTRODUCTION	1
	1.1 Background of the Study	1
	1.2 Problem Statement	3
	1.3 Objectives of the Research	4
	1.4 Scope of the Research	4
	1.5 Significance and Original Contributions of the Research	5
2	LITERATURE REVIEW	6
	2.1 Microbial Life in Arctic Environment	6
	2.2 Diversity of Cold-adapted Microorganisms	8
	2.2.1 Bacterial Diversity in Arctic Regions	9
	2.3 Psychrotrophic Bacteria	10
	2.4 Biotechnological Potentials of Psychrotrophic Bacteria	11

2.4.1	Roles of Psychrotrophic Bacteria in Bioremediation	11
2.5	Physiological Adaptation of Cold-adapted Microorganisms	13
2.5.1	Microbial Protein Composition and Cold Adaptation	14
2.5.2	Membrane Lipids Composition and Cold Adaptation	15
2.5.3	Other substances of Microbial Cold Adaptation	16
2.6	Microbial Biogeography of Cold Environment	16
2.7	Microbial Hydrolases	18
2.7.1	Advantages of Microbial Enzymes Over Other forms of Enzymes	19
2.7.2	Extracellular Hydrolytic Enzymes of Cold-adapted bacteria	19
2.7.2.1	Lipases	20
2.7.2.2	Proteases	21
2.7.2.3	Amylases	21
2.8	Applications of Microbial Enzymes	22
2.8.1	Microbial Enzymes and Bioremediation	23
2.9	16S rRNA Gene and its Role in Bacterial Identification	23
3	MATERIALS AND METHODS	26
3.1	Experimental Design	26
3.2	Samples Collection	28
3.3	Media Preparation	29
3.4	Enumeration of the Culturable Psychrotrophic Bacteria	29
3.4.1	Preparation of Sample Suspension and Serial Dilution	29
3.4.2	Viable Bacterial Count Using Spread Plate Techniques	30
3.5	Isolation of Pure Culture	30
3.5.1	Preservation of Pure Culture of the Isolates	31

3.6	Grams' Staining	31
3.7	Characterization and Identification of the Isolated Bacteria using 16S rRNA Gene Sequence	31
3.7.1	Genomic DNA Extraction	32
3.7.2	Purity Assessment and Quantification of the Extracted DNA	33
3.7.3	Amplification of 16S rRNA Gene Using Polymerase Chain Reaction	33
3.7.4	Agarose Gel Electrophoresis	34
3.7.5	16S rRNA Gene Sequencing and Phylogenetic Analysis	35
3.8	Screening for Extracellular Hydrolytic Enzymes Activity	36
3.8.1	Amylase Activity	36
3.8.2	Protease Activity	37
3.8.3	Lipase Activity	37
3.9	Determination of growth Temperature Range	38
4	RESULTS AND DISCUSSIONS	39
4.1	Culturable Psychrotrophic Bacterial Enumeration	39
4.2	Isolation of Psychrotrophic Bacteria	40
4.3	Identification of the Isolated Bacteria Using 16S rRNA Gene Sequence	44
4.3.1	16S rRNA Gene Sequences Analysis	45
4.3.2	Phylogenetic Analysis of the Isolated Bacteria	47
4.3.2.1	Phylogenetic Tree	49
4.4	Nucleotide Sequence Accession Number	50
4.5	Hydrolytic Enzymes Activities of the Isolated Bacteria	51
4.5.1	Amylase Activity of the Isolated Bacteria	52
4.5.2	Protease Activity of the Isolated Bacteria	54
4.5.3	Lipase Activity of the Isolated Bacteria	56
4.6	Growth Temperature of the Bacteria	58

5	CONCLUSION AND FUTURE WORK	61
	5.1 Conclusion	61
	5.2 Future work	61
	REFERENCES	63
	APPENDICES	75

LIST OF TABLES

TABLE NO.	TITLE	PAGE
3.1	Sampling Location	28
3.2	Details of PCR Primers Used	34
3.3	PCR Reaction Recipes	34
3.4	PCR Reaction Cycling Parameters	34
4.1	Mean Psychrotrophic Bacterial Count of different Arctic Sediments	39
4.2	Morphological Characteristics of Bacteria Isolated from Arctic lake Sediments	41
4.3	Taxonomic Distribution of Bacteria Isolated from Different Arctic Lake Sediments	46
4.4	Phylogenetic Affiliations of Some Bacteria Isolated from Arctic Lake Sediments	48
4.5	Details of 16S rRNA Gene Partial Sequences Deposited in GenBank Nucleotide Database of NCBI	50
4.6	Occurrence of Bacteria with Hydrolytic Enzymes Activity in Arctic Lake Sediments	51
4.7	Variation of Enzyme Activity Among Amylase Producing Bacteria Based on Coefficient of Hydrolysis	53
4.8	Variation of Proteolytic Activity Among Protease Producing Isolates Based on Coefficient of Hydrolysis	55
4.9	Variation of Lipase Activity Among Lipase Producing Isolates based on Coefficient of Hydrolysis	58
4.10	Growth Temperature of the Enzymes Producing Bacteria Isolated from Arctic Lake Sediment	59

4.11	Summary of Growth Temperature of the Enzymes Producing Bacteria	60
------	-----------------------------------------------------------------	----

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Geographical Map of Arctic Region	8
2.2	Schematic Depiction of Bacterial Growth Rate as a Function of Temperature for Psychrophilic, Psychrotrophic and Mesophilic Microbes.	11
3.1	Experimental Flow Chart	27
3.2	Geographical Map of Svalbard Showing the Sampling Site	28
4.1	Variation in Number of Morphologically Distinct Colonies Isolated from the Arctic Lake Sediments	43
4.2	Pie Chart Showing the Percentages Occurrence of the Representative Bacterial Genera	47
4.3	Phylogenetic Trees Showing Phylogenetic Relationship of the Isolated Bacteria	49
4.4	Starch Agar Plates Showing Amylase activity Test	54
4.5	Skim Milk Agar Plates Showing Protease Activity Test	56
4.6	Tween Agar Plates Showing Lipase Activity Test	58

LIST OF ABBREVIATIONS AND SYMBOLS

<i>ABM</i>	-	Antarctic Bacterial Medium
<i>ABI</i>	-	Application Binary Interface
<i>AFPs</i>	-	Antifreeze Proteins
<i>BLAST</i>	-	Basic Local Alignment Tool
<i>CAPS</i>	-	Cold Acclimation Proteins
<i>cfu/g</i>	-	Colony Forming Unit
<i>CaCl₂</i>	-	Calcium Chloride
$^{\circ}\text{C}$	-	Degree Celcius
<i>DNA</i>	-	Deoxyribonucleic Acids
<i>EDTA</i>	-	Ethylene Diamine Tetraacetic Acid
<i>SMA</i>	-	Skim Milk Agar
<i>g</i>	-	Gram
<i>gb</i>	-	GenBank
<i>g/ml</i>	-	Gram Per Millilitre
<i>h</i>	-	Hour
<i>H_c</i>	-	Coefficient of hydrolysis
<i>H₂O</i>	-	Water
<i>bp</i>	-	Base pairs
<i>kbp</i>		Kilo base pairs
<i>L</i>	-	Litre
<i>Min</i>	-	Minutes
<i>mg/ml</i>	-	Miligram per milliliter
<i>MgSO₄</i>	-	Magnesium Sulphate
<i>ml</i>	-	Mililitres

<i>mM</i>	-	Milimolar
<i>mm</i>	-	Milimetre
<i>M</i>	-	Molar
<i>ng</i>	-	Nanogram
<i>NCBI</i>		National Center for Biotechnology Information
<i>NA</i>	-	Nutrient Agar
<i>NB</i>	-	Nutrient Broth
<i>NaCl</i>	-	Sodium Chloride
<i>NaOH</i>	-	Sodium Hydroxide
<i>ng/μL</i>		Nanogram Per Microliter.
<i>%</i>	-	Percent
<i>PCR</i>	-	Polymerase Chain Reaction
<i>PUFAs</i>		Polyunsaturated Fatty Acids
<i>rpm</i>	-	Revolutions Per Minute
<i>sec</i>	-	Seconds
<i>TAE</i>	-	Tris acetate EDTA
<i>UV</i>	-	Ultraviolet
<i>μL</i>	-	Microlitre
<i>v/v</i>	-	Volume per volume
<i>w/v</i>	-	Weight per volume

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Details of Media and Reagents Used	75
A1	Composition of ABM Medium Used for Bacterial Count and Isolation	75
A2	Composition of Starch Agar Used for the Screening of Amylase Activity.	75
A3	Composition of Skim Milk Agar Used for the Screening of Protease Activity.	76
A4	Composition of Tween Agar Used for the Screening of Lipase Activity.	76
A5	Protocol for the Preparation of Gram's Staining Reagents	76
A6	Gram's Staining Procedure Adopted	77
B	16S rRNA Gene Sequence Data of the Isolated Bacteria	78
C	Details of Some Experimental Results	97
C1	Raw Data of the Screening for Extracellular Hydrolytic Enzymes Activity	97
C2	Pictures of Pure Culture and Gram's Staining Reactions of Some Isolated Bacteria	99
C3	Agarose Gel Picture of the Amplified 16S rRNA Gene of Selected Bacteria.	100

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

The term bioprospecting has been described as a systematic search for novel microorganisms, bioactive compounds, proteins, genes and other products with promising commercial applications (Pascale *et al.*, 2012). The potential for isolating new microbial species for biotechnological applications has initiated a strong interest in the microbiology of cold environment such as Arctic and Antarctic polar regions (Männistö and Häggblom, 2006). Several new bacteria have been isolated, identified and characterized within the past few years, from permanently cold environment. In recent times, potential applications of cold adapted microorganisms and their metabolic products including cold-active enzymes and novel bioactive compounds have been documented (Deming, 2002).

Psychrotrophic bacteria, which are distinct from psychrophilic bacteria due to their ability to grow both at 4°C and at 20°C, have been known to be widely distributed in natural environment and they adapt more to a wider growth temperature range than psychrophiles (Radjasa *et al.*, 2001). A large proportion of the earth's surface is occupied by cold environment such as the Arctic, Antarctic, alpine regions, and abysses. Due to the extremely harsh climatic conditions, these cold environments have been considered for a long time to be extreme and therefore

devoid of life or serving only as repositories for wind-transported microbes trapped in the frozen water (Cowan and Tow, 2004). Although still limited, the growing number of recent studies on the microbial diversity and ecology of cold habitat have changed this view, they have revealed that, even under such hostile conditions associated with polar regions, such areas harbour abundant, live and diverse microbes; particularly psychrotrophs and psychrophiles that may be detected and recovered by cultivation (Suzuki *et al.*, 2001). Recently, increasing attention has been paid to the application of psychrophiles, psychrotrophs, and their cold active enzymes in biotechnology. For example, cold active hydrolases such as lipases, proteinases, amylases and cellulases from these microorganisms have been used as additives in laundry detergents (Suzuki *et al.*, 2001). Potential applications of cold-adapted microorganisms may also include their use as agents for bioremediation at low temperature as well as mitigation greenhouse gas effect.

Furthermore, low temperature environment are abundant on Earth and have been well inhabited by cold-adapted microorganisms most of which can be utilise as cell factories for the production of valuable chemical compounds as well as for clean-up of polluted cold environments (Margesin and Feller, 2010). Likewise, the biomolecules produce by these organisms, particularly proteins and enzymes have already found useful applications in various fields such as medical research, molecular biology, industrial food or feed technologies, detergents and cosmetics (Margesin and Feller, 2010).

In the recent years, the study of cold environments such as arctic and their microorganisms especially bacteria has begun to receive greater attention through which variety of native microbial life forms have been discovered and characterized. (Gesheva and Negoita, 2012). Although arctic environment is sometimes being described as inhospitable, recent applications of molecular methods have revealed a very wide diversity of microbial taxa in the samples taken from such environments; many of which are yet to be cultured and taxonomically unique. Therefore, these environments could be considered to be a suitable environment for the isolation of bacteria capable of producing cold-active enzymes; some of which have been found to be useful in industry (Martínez-Rosales and Castro-Sowinski, 2011).

There have been numerous studies to determine microbial diversity in polar region through culture-dependent methods. For example, bacteria have been recovered from the arctic tundra soils in Siberia, Svalbard, and Canada; majority of which belonged to the phylum *Actinobacteria*, *Bacteroidetes*, *Firmicutes*, and *Proteobacteria* (Kim *et al.*, 2013). It is important to acknowledge that culture-based methods underestimate the true diversity of bacteria associated with environmental samples. However, approaches to investigating microbial ecosystems on the basis of conventional culture methods are important since, via this approach, the ecological role of the cultivated and characterized microbes can be estimated. Furthermore, culturing is necessary to biochemically classify and analyse some physiological features of the organisms for biotechnological exploitation (Galkiewicz *et al.*, 2011).

1.2 Problem Statement

Bioprospecting of cold-adapted microbial resources has been a real challenge and opportunity for contemporary biotechnology. Cold-adapted bioactive compounds recovered from psychrophilic and psychrotrophic microorganisms continue to provide advantages in different areas, such as energy savings; activity at low temperatures; the possibility of challenging reactions with a sufficiently high rate; efficient production with lower cost of processing; thermal protection as well as improving quality of products. The market for bioactive products and industrial enzymes at the moment is growing. Cold-active hydrolytic enzymes such as lipases, proteases amylases etc. have numerous potential applications especially in industries.

Despite an increasing number of microbial diversity assessments of Polar Regions that began to accumulate in recent years, only relatively little is known about the cold-adapted hydrolytic enzymes producing bacteria. Also, early studies of bacteria in Arctic environmental samples have focused much on abundance and diversity and to a certain extent; the influence of climatic conditions on these microbes with relatively few information about the potentials of the organisms

recovered from such environment; in terms of production of bioactive compounds for biotechnological applications. Thus, cultivating and characterizing isolates are believed to be particularly important for providing insight into diversity and bioactivity of these organisms especially; because culture-based methods make the characteristics of microorganisms clear, and also provides several interesting information such as the low temperature adaptability of organisms, elaboration of extracellular enzymes, and many other physiological activities. It is therefore based on this background that this research is designed with the aim of investigating the presence and diversity of psychrotrophic bacteria with extracellular hydrolytic enzymes activity for possible environmental and industrial applications.

1.3 Objectives of the Research

The specific objectives of this research are:

- i. To enumerate and isolate culturable psychrotrophic bacteria associated with Arctic lake sediments using standard culture-based methods.
- ii. To characterize and identify the isolates using 16S rRNA gene sequence homology.
- iii. To screen the isolated bacteria for selected extracellular hydrolytic enzymes activity.

1.4 Scope of the Research

The scope of the research includes: investigating the presence and diversity of culturable psychrotrophic bacteria through enumeration and isolation of bacteria associated with Arctic sediment using standard microbiological methods. Pure cultures of each isolate were identified and characterized using molecular approach

(16S rRNA gene sequence homology), the isolated bacteria were screened for some selected hydrolytic enzymes activity; namely amylase, lipase and protease.

1.5 Significance and Original Contributions of the Research

Industrial processes are carried out under specific conditions which in most cases cannot always be accustomed to an optimal values required for the activity of the enzymes available (Cojoc *et al.*, 2009). The increased use of microbial enzymes especially bacterial extracellular hydrolases in the industry has focused attention on the source identification and recovery of novel enzymes with new desirable properties such as low temperature performance. Such effort is very vital in terms of saving energy as some industrial processes could chiefly be accomplished at near or room temperature (Martínez-Rosales and Castro-Sowinski, 2011). Cold-adapted enzymes have generally been observed to possess high specific activity or catalytic efficiency at low and moderate temperatures relative to enzymes derived from mesophilic or thermophilic microorganisms (Margesin *et al.*, 2008).

The capacity of microbes to synthesize enzymes capable of catalyzing reactions at temperatures near freezing point of water is no small feat. Given the commercial success of these special enzymes, effort is now being directed toward isolating microbes particularly bacteria capable of producing such enzymes in commercial quantity. This research has the potential to make a significant contribution to the existing knowledge of bacterial diversity of arctic ecosystems as well as the potentials of the indigenous microbes in search for a novel biocatalyst with improved properties.

REFERENCES

- Alcalde, M., Ferrer, M., Plou, F. J. and Ballesteros, A. (2006). Environmental Biocatalysis: from Remediation with Enzymes to Novel Green Processes. *Trends in Biotechnology*, 24(6), 281-287.
- Amoozegar, M. A., Malekzadeh, F. and Malik, K. A. (2003). Production of Amylase by Newly Isolated Moderate Halophile, *Halobacillus* sp. strain MA-2. *Journal of Microbiological Methods*, 52(3), 353-359.
- Araújo, R., Casal, M. and Cavaco-Paulo, A. (2008). Application of Enzymes for Textile Fibres Processing. *Biocatalysis and Biotransformation*, 26(5), 332-349.
- Ayub, N. D., Tribelli, P. M. and López, N. I. (2009). Polyhydroxyalkanoates are Essential for Maintenance of Redox State in the Antarctic Bacterium *Pseudomonas* sp. 14-3 During Low Temperature Adaptation. *Extremophiles*, 13(1), 59-66.
- Boesenberg-Smith, K. A. Pessarakli, M. M., and Wolk, D. M. (2012). Assessment of DNA yield and Purity: an Overlooked Detail of PCR Troubleshooting. *Clinical Microbiology Newsletter*, 34(1), 1-6.
- Brown, A. E. (2009). *Benson's Microbiological Applications: Laboratory Manual in General Microbiology, Short Version*: McGraw-Hill Higher Education. 82-87

- Carissimi, M., Stopiglia, C., de Souza, T., Corbellini, V. and Scroferneker, M. (2007). Comparison of Lipolytic Activity of *Sporothrix schenckii* Strains Utilizing Olive oil-rhodamine B and Tween 80. *Tecno-Lógica*, 11, 33-36.
- Cavicchioli, R. (2006). Cold-adapted archaea. *Nature Reviews Microbiology*, 4(5), 331-343.
- Cavicchioli, R., Siddiqui, K. S., Andrews, D. and Sowers, K. R. (2002). Low-Temperature Extremophiles and their Applications. *Current Opinion in Biotechnology*, 13(3), 253-261.
- Chintalapati, S., Kiran, M. and Shivaji, S. (2004). Role of Membrane Lipid Fatty Acids in Cold Adaptation. *Cellular and Molecular Biology (Noisy-le-Grand, France)*, 50(5), 631-642.
- Christner, B. C., Skidmore, M. L., Priscu, J. C., Tranter, M. and Foreman, C. M. (2008). Bacteria in Subglacial Environments *Psychrophiles: from Biodiversity to Biotechnology* pp. 51-71
- Chrost, R. J. (1991). Environmental Control of the Synthesis and Activity of Aquatic Microbial Ectoenzymes *Microbial Enzymes in Aquatic Environments* pp. 29-59
- Clarridge, J. E. (2004). Impact of 16S rRNA Gene Sequence Analysis for Identification of Bacteria on Clinical Microbiology and Infectious Diseases. *Clinical Microbiology Reviews*, 17(4), 840-862.
- Cojoc, R., Merciu, S., Popescu, G., Dumitru, L., Kamekura, M. and Enache, M. (2009). Extracellular Hydrolytic Enzymes of Halophilic Bacteria Isolated from a Subterranean Rock Salt Crystal. *Rom Biotechnology Letter*, 14, 4658-4664.
- D'Amico, S., Collins, T., Marx, J. C., Feller, G. and Gerday, C. (2006). Psychrophilic Microorganisms: Challenges for Life. *EMBO Reports*, 7(4), 385-389.

- Deming, J. (2010). Extremophiles: Cold Environments. *Desk Encyclopedia of Microbiology*, 483-493.
- Deming, J. W. (2002). Psychrophiles and Polar Regions. *Current Opinion in Microbiology*, 5(3), 301-309.
- Feller, G. (2007). Life at Low Temperatures: Is Disorder the Driving Force? *Extremophiles*, 11(2), 211-216.
- Feller, G. and Gerday, C. (2003). Psychrophilic Enzymes: Hot Topics in Cold Adaptation. *Nature Reviews Microbiology*, 1(3), 200-208.
- Fendrihan, S. and Negoită, T. G. (2012). *Psychrophilic Microorganisms As Important Source for Biotechnological Processes*: Springer Vienna Pp. 133-172.
- Fierer, N. (2008). Microbial Biogeography: Patterns in Microbial Diversity Across Space and Time. *Assessing Uncultivated Microorganisms: From the Environment to Organisms and Genomes And Back*, 95-115.
- Galkiewicz, J. P., Pratte, Z. A., Gray, M. A. and Kellogg, C. A. (2011). Characterization of Culturable Bacteria Isolated from the Cold-water Coral *Lophelia pertusa*. *FEMS Microbiology Ecology*, 77(2), 333-346.
- Garneau, M. È., Roy, S., Lovejoy, C., Gratton, Y. and Vincent, W. F. (2008). Seasonal Dynamics of Bacterial Biomass and Production in a Coastal Arctic Ecosystem: Franklin Bay, western Canadian Arctic. *Journal of Geophysical Research: Oceans (1978–2012)*, 113(C7).
- Georlette, D., Damien, B., Blaise, V., Depiereux, E., Uversky, V. N., Gerday, C. and Feller, G. (2003). Structural and Functional Adaptations to Extreme Temperatures in Psychrophilic, Mesophilic, and Thermophilic DNA Ligases. *Journal of Biological Chemistry*, 278(39), 37015-37023.

- Gesheva, V. and Negoita, T. (2012). Psychrotrophic Microorganism Communities In Soils of Haswell Island, Antarctica, and their Biosynthetic Potential. *Polar Biology*, 35(2), 291-297.
- Gianese, G., Argos, P. and Pascarella, S. (2001). Structural Adaptation of Enzymes to Low Temperatures. *Protein Engineering*, 14(3), 141-148.
- Gianfreda, L. and Rao, M. A. (2004). Potential of Extracellular Enzymes in Remediation of Polluted Soils: a review. *Enzyme and Microbial Technology*, 35(4), 339-354.
- Giudice, A. L., Casella, P., Caruso, C., Mangano, S., Bruni, V., De Domenico, M. and Michaud, L. (2010). Occurrence and Characterization of Psychrotolerant Hydrocarbon-Oxidizing Bacteria from Surface Seawater along the Victoria Land Coast (Antarctica). *Polar Biology*, 33(7), 929-943.
- Groudieva, T., Kambourova, M., Yusef, H., Royter, M., Grote, R., Trinks, H. and Antranikian, G. (2004). Diversity and Cold-Active Hydrolytic Enzymes of Culturable Bacteria Associated with Arctic sea ice, Spitzbergen. *Extremophiles*, 8(6), 475-488.
- Grzymiski, J. J., Carter, B. J., DeLong, E. F., Feldman, R. A., Ghadiri, A. and Murray, A. E. (2006). Comparative Genomics of DNA Fragments from Six Antarctic Marine Planktonic Bacteria. *Applied and Environmental Microbiology*, 72(2), 1532-1541.
- Gupta, R., Gigras, P., Mohapatra, H., Goswami, V. K. and Chauhan, B. (2003). Microbial α -amylases: A Biotechnological Perspective. *Process Biochemistry*, 38(11), 1599-1616.

- Haas, B. J., Gevers, D., Earl, A. M., Feldgarden, M., Ward, D. V., Giannoukos, G. and Sodergren, E. (2011). Chimeric 16S rRNA Sequence Formation and Detection in Sanger and 454-pyrosequenced PCR Amplicons. *Genome Research*, 21(3), 494-504.
- Hall, T. A. (1999). *BioEdit: A User-friendly Biological Sequence Alignment Editor and Analysis Program For Windows 95/98/NT*. Paper presented at the Nucleic acids symposium series.
- Hasan, F., Shah, A. A. and Hameed, A. (2006). Industrial Applications of Microbial Lipases. *Enzyme and Microbial Technology*, 39(2), 235-251.
- Janda, J. M. and Abbott, S. L. (2007). 16S rRNA Gene Sequencing for Bacterial Identification in the Diagnostic Laboratory: Pluses, Perils, and Pitfalls. *Journal of Clinical Microbiology*, 45(9), 2761-2764.
- Joseph, B., Ramteke, P. W. and Thomas, G. (2008). Cold-active Microbial Lipases: Some Hot Issues and Recent Developments. *Biotechnology Advances*, 26(5), 457-470.
- Jungblut, A. D., Lovejoy, C. and Vincent, W. F. (2009). Global Distribution of Cyanobacterial Ecotypes in the Cold Biosphere. *The ISME Journal*, 4(2), 191-202.
- Kato, T., Haruki, M., Imanaka, T., Morikawa, M. and Kanaya, S. (2001). Isolation and Characterization of Psychrotrophic Bacteria from Oil-reservoir Water and Oil Sands. *Applied Microbiology and Biotechnology*, 55(6), 794-800.
- Kavitha, M. and Shanthi, C. (2013). Isolation and Characterization of Cold-active Lipase Producing Pseudomonas sp. 4 from Marine Samples of Tamilnadu Coast. *Research Journal of Biotechnology*, 8(4), 57-62.
- Kawahara, H. (2008). Cryoprotectants and Ice-binding Proteins *Psychrophiles: from Biodiversity to Biotechnology*, pp. 229-246.

- Kevbrina, M., Okhapkina, A., Akhlynin, D., Kravchenko, I., Nozhevnikova, A. and Gal'chenko, V. (2001). Growth of Mesophilic Methanotrophs at Low Temperatures. *Microbiology*, 70(4), 384-391.
- Kim, H. R., Kim, I. H., Hou, C. T., Kwon, K. I. and Shin, B.-S. (2009). Production of a Novel Cold-active Lipase from *Pichia lynferdii* Y-7723. *Journal of Agricultural and Food Chemistry*, 58(2), 1322-1326.
- Kim, H. M., Chae, N., Jung, J. Y. and Lee, Y. K. (2013). Isolation of Facultatively Anaerobic Soil Bacteria from Ny-Ålesund, Svalbard. *Polar Biology*, 36(6), 787-796.
- Krembs, C., and Deming, J. W. (2008). The Role Of Exopolymers In Microbial Adaptation to Sea Ice *Psychrophiles: From Biodiversity to Biotechnology* , pp. 247-264.
- Kumar, D., Kumar, L., Nagar, S., Raina, C., Parshad, R. and Gupta, V. K. (2012). Screening Isolation and Production of Lipase/Esterase Producing *Bacillus sp.* strain DVL2 and Its Potential Evaluation in Esterification and Resolution Reactions. *Archives of Applied Science Research*, 4, 1763-1770.
- Lee, H. K., Ahn, M.-J., Kwak, S. H., Song, W. H. and Jeong, B. C. (2003). Purification and Characterization of Cold-active Lipase from Psychrotrophic *Aeromonas sp.* LPB 4. *Journal of Microbiology-Seoul-*, 41(1), 22-27.
- Madigan, M. T., Martinko, J. M., Dunlap, P. V. and Clark, D. P. (2008). Brock Biology of microorganisms 12th edn. *International Microbiology*, 11, 65-73.
- Männistö, M. K. and Häggblom, M. M. (2006). Characterization of Psychrotolerant Heterotrophic Bacteria from Finnish Lapland. *Systematic and Applied Microbiology*, 29(3), 229-243.

- Margesin, R., and Feller, G. (2010). Biotechnological Applications of Psychrophiles. *Environmental Technology*, 31(8-9), 835-844.
- Margesin, R. and Miteva, V. (2011). Diversity and Ecology of Psychrophilic Microorganisms. *Research in Microbiology*, 162(3), 346-361.
- Margesin, R., Schinner, F., Marx, J. C. and Gerday, C. (2008). *Psychrophiles: from Biodiversity to Biotechnology*: Springer. pp. 211-224
- Martínez-Rosales, C. and Castro-Sowinski, S. (2011). Antarctic Bacterial Isolates that Produce Cold-active Extracellular Proteases at Low Temperature But Are Active and Stable at High Temperature. *Polar Research*, 30.
- Metcalf, A. C., Krsek, M., Gooday, G., Prosser, J. and Wellington, E. (2002). Molecular Analysis of a Bacterial Chitinolytic Community in an Upland Pasture. *Applied and Environmental Microbiology*, 68(10), 5042-5050.
- Michaux, C., Massant, J., Kerff, F., Frère, J. M., Docquier, J. D., Vandenberghe, I. and Charlier, P. (2008). Crystal Structure of a Cold-Adapted Class C β -Lactamase. *FEBS Journal*, 275(8), 1687-1697.
- Mignard, S., and Flandrois, J. P. (2006). 16S rRNA Sequencing in Routine Bacterial Identification: A 30-Month Experiment. *Journal of Microbiological Methods*, 67(3), 574-581.
- Mikhailova, A., Likhareva, V., Khairullin, R., Lubenets, N., Rumsh, L., Demidyuk, I. and Kostrov, S. (2006). Psychrophilic Trypsin-Type Protease From *Serratia Proteamaculans*. *Biochemistry (Moscow)*, 71(5), 563-570.
- Mock, T. and Thomas, D. N. (2008). Microalgae in Polar Regions: Linking Functional Genomics and Physiology with Environmental Conditions *Psychrophiles: From Biodiversity to Biotechnology* pp. 285-312.

- Morgan-Kiss, R. M., Priscu, J. C., Pockock, T., Gudynaite-Savitch, L. and Huner, N. P. (2006). Adaptation and acclimation of photosynthetic microorganisms to permanently cold environments. *Microbiology and Molecular Biology Reviews*, 70(1), 222-252.
- Morita, Y., Nakamura, T., Hasan, Q., Murakami, Y., Yokoyama, K. and Tamiya, E. (1997). Cold-active Enzymes from Cold-adapted Bacteria. *Journal of the American Oil Chemists' Society*, 74(4), 441-444.
- Newman, M. M., Feminella, J. W. and Liles, M. R. (2010). Purification of Genomic DNA Extracted from Environmental Sources for Use in a Polymerase Chain Reaction. *Cold Spring Harbor Protocols*, 2010(2), pdb. prot5383-pdb. prot5383.
- Nogales, B., Moore, E. R., Llobet-Brossa, E., Rossello-Mora, R., Amann, R. and Timmis, K. N. (2001). Combined Use of 16S Ribosomal DNA and 16S rRNA to Study the Bacterial Community of Polychlorinated Biphenyl-Polluted Soil. *Applied and Environmental Microbiology*, 67(4), 1874-1884.
- Nuttall, M. and Callaghan, T. (2000). *Arctic: Environment, People, Policy*: CRC Press.
- Ohgiya, S., Hoshino, T., Okuyama, H., Tanaka, S. and Ishizaki, K. (1999). Biotechnology of Enzymes from Cold-adapted Microorganisms *Biotechnological Applications of Cold-Adapted Organisms* pp. 17-34.
- Pascale, D., Santi, C., Fu, J. and Landfald, B. (2012). The Microbial Diversity of Polar Environments is a Fertile Ground for Bioprospecting. *Marine Genomics*, 8, 15-22.
- Patel, J. B. (2001). 16S rRNA Gene Sequencing For Bacterial Pathogen Identification in the Clinical Laboratory. *Molecular Diagnosis*, 6(4), 313-321.

- Pathom-Aree, W., Nogi, Y., Ward, A. C., Horikoshi, K., Bull, A. T. and Goodfellow, M. (2006). *Dermacoccus barathri* sp. nov. and *Dermacoccus profundus* sp. nov., Novel Actinomycetes Isolated from Deep-Sea Mud of the Mariana Trench. *International Journal of Systematic and Evolutionary Microbiology*, 56(10), 2303-2307.
- Radjasa, O. K., Urakawa, H., Kita-Tsukamoto, K. and Ohwada, K. (2001). Characterization of Psychrotrophic Bacteria in the Surface and Deep-Sea Waters from the Northwestern Pacific Ocean Based On 16s Ribosomal DNA Analysis. *Marine Biotechnology*, 3(5), 454-462.
- Ramaiah, N. (1994). Production of Certain Hydrolytic Enzymes by Psychrophilic Bacteria from the Antarctic krill, Zooplankton and Seawater. Scientific Report of Ninth Indian Expedition to Antarctica, pp. 107-114.
- Ramette, A. and Tiedje, J. M. (2007). Biogeography: An Emerging Cornerstone For Understanding Prokaryotic Diversity, Ecology, and Evolution. *Microbial Ecology*, 53(2), 197-207.
- Ramteke, P. W., Joseph, B. and Kuddus, M. (2005). Extracellular Lipases from Anaerobic Microorganisms of Antarctic. *Indian Journal of Biotechnology*, 4(2), 293-294.
- Reddy, P. V., Shiva Nageswara Rao, S. S., Pratibha, M. S., Sailaja, B., Kavya, B., Manorama, R. R. and Shivaji, S. (2009). Bacterial Diversity and Bioprospecting for Cold-Active Enzymes from Culturable Bacteria Associated with Sediment from a Melt Water Stream of Midtre Lo Enbreen Glacier, An Arctic glacier. *Research in Microbiology*, 160(8), 538-546.
- Reller, L. B., Weinstein, M. P. and Petti, C. A. (2007). Detection and Identification of Microorganisms by Gene Amplification and Sequencing. *Clinical Infectious Diseases*, 44(8), 1108-1114.

- Rendleman, J. (1996). Enzymic Conversion of Malto-oligosaccharides and Maltodextrin into Cyclodextrin at Low Temperature. *Biotechnology and Applied Biochemistry*, 24(2), 129-137.
- Rodrigues, D. F., da C Jesus, E., Ayala-del-Río, H. L., Pellizari, V. H., Gilichinsky, D., Sepulveda-Torres, L. and Tiedje, J. M. (2009). Biogeography of Two Cold-adapted Genera: Psychrobacter and Exiguobacterium. *The ISME Journal*, 3(6), 658-665.
- Rohban, R., Amoozegar, M. A. and Ventosa, A. (2009). Screening and Isolation of Halophilic Bacteria Producing Extracellular Hydrolyses from Howz Soltan Lake, Iran. *Journal of Industrial Microbiology & Biotechnology*, 36(3), 333-340.
- Russell, N. J. (2008). Membrane Components and Cold Sensing *Psychrophiles: From Biodiversity to Biotechnology*, Springer. pp. 177-190.
- Russell, N. J. and Nichols, D. S. (1999). Polyunsaturated Fatty Acids in Marine Bacteria - A Dogma Rewritten. *Microbiology* 145, 767-779.
- Rybalka, N., Andersen, R. A., Kostikov, I., Mohr, K. I., Massalski, A., Olech, M. and Friedl, T. (2009). Testing for Endemism, Genotypic Diversity and Species Concepts in Antarctic Terrestrial Microalgae of the Tribonemataceae (Stramenopiles, Xanthophyceae). *Environmental Microbiology*, 11(3), 554-565.
- Sælensminde, G., Halskau Jr, Ø. and Jonassen, I. (2009). Amino Acid Contacts in Proteins Adapted to Different Temperatures: Hydrophobic Interactions and Surface Charges Play a Key Role. *Extremophiles*, 13(1), 11-20.
- Sharma, R., Chisti, Y. and Banerjee, U. C. (2001). Production, Purification, Characterization, and Applications of Lipases. *Biotechnology Advances*, 19(8), 627-662.

- Shen, L., Yao, T., Xu, B., Wang, H., Jiao, N., Kang, S. and Liu, Y. (2012). Variation Of Culturable Bacteria Along Depth in the East Rongbuk Ice Core, Mt. Everest. *Geoscience Frontiers*, 3(3), 327-334.
- Siddiqui, K. S. and Cavicchioli, R. (2006). Cold-adapted Enzymes. *Annual Review of Biochemistry*, 75, 403-433.
- Singh, C. J. (2003). Optimization of an Extracellular Protease of *Chrysosporium Keratinophilum* and its Potential in Bioremediation of Keratinic Wastes. *Mycopathologia*, 156(3), 151-156.
- Singh, L. and Ramana, K. V. (2006). Isolation and Characterization of Psychrotrophic Antarctic Bacteria from Blue-green Algal Mats and Their Hydrolytic Enzymes. *Fourteenth Indian Expedition to Antarctica, Scientific Report*, 12. pp 199-206
- Sommaruga, R. and Casamayor, E. O. (2009). Bacterial 'Cosmopolitanism' and Importance of Local Environmental Factors for Community Composition in Remote High-Altitude Lakes. *Freshwater Biology*, 54(5), 994-1005.
- Sonan, G., Receveur-Brechot, V., Duez, C., Aghajari, N., Czjzek, M., Haser, R. and Gerday, C. (2007). The linker region plays a key role in the adaptation to cold of the cellulase from an Antarctic bacterium. *Biochemical. Journal*, 407, 293-302.
- Souza, P. M. d. (2010). Application of Microbial A-amylase in Industry-A Review. *Brazilian Journal of Microbiology*, 41(4), 850-861.
- Staley, J. T. and Gosink, J. J. (1999). Poles Apart: Biodiversity and Biogeography of Sea Ice Bacteria. *Annual Reviews in Microbiology*, 53(1), 189-215.
- Stan-Lotter, H. and Fendrihan, S. (2012). *Adaption of Microbial Life to Environmental Extremes*: Springer. pp. 133-157.

- Sutherland, T., Horne, I., Weir, K., Coppin, C., Williams, M., Selleck, M. and Oakeshott, J. (2004). Enzymatic Bioremediation: from Enzyme Discovery to Applications. *Clinical and Experimental Pharmacology and Physiology*, 31(11), 817-821.
- Suzuki, T., Nakayama, T., Kurihara, T., Nishino, T. and Esaki, N. (2001). Cold-active Lipolytic Activity of Psychrotrophic *Acinetobacter* sp. Strain No. 6. *Journal of Bioscience and Bioengineering*, 92(2), 144-148.
- Takeuchi, N. and Kohshima, S. (2004). A Snow Algal Community on Tyndall Glacier in the Southern Patagonia Icefield, Chile. *Arctic, Antarctic, and Alpine Research*, 36(1), 92-99.
- Wells, L. E. (2008). Cold-active viruses *Psychrophiles: from Biodiversity to Biotechnology* , Springer, pp. 157-173.
- Yu, Y., Li, H. R., Zeng, Y. X. and Chen, B. (2011). Bacterial Diversity And Bioprospecting for Cold-Active Hydrolytic Enzymes from Culturable Bacteria Associated with Sediment from Nella Fjord, Eastern Antarctica. *Marine Drugs*, 9(2), 184-195.
- Zachariassen, K. and Lundheim, R. (1999). Applications of Antifreeze Proteins *Biotechnological Applications of Cold-Adapted Organisms*, Springer, pp. 319-332,