TOTAL PHENOLIC CONTENT, ANTIOXIDANT AND CYTOTOXIC ACTIVITIES OF *Citrus hystrix* LEAF EXTRACTS

PUSHPALATHA A/P SIVARAJ

UNIVERSITI TEKNOLOGI MALAYSIA

TOTAL PHENOLIC CONTENT, ANTIOXIDANT AND CYTOTOXIC ACTIVITIES OF Citrus hystrix LEAF EXTRACTS

PUSHPALATHA A/P SIVARAJ

A dissertation submitted in partial fulfilment of the requirements for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

JANUARY 2015

I would like to dedicate this thesis to my dearest uncle, MR Ravichandran Ramasamy. Without him, none of this would have happened. Thank you for everything.

AKNOWLEDGEMENT

Thanks to the god Almighty for giving me the chance to successfully finish this project in the given time.

The first person I wish to express my sincere gratitude and appreciation is my research supervisor, Dr. Salehhuddin Bin Hamdan for his continual guidance, critics and advices.

In preparing this dissertation, I was in contact with many people, researchers and many academicians. They have contributed towards my understanding and thoughts. My sincere appreciation goes to all fellow postgraduates especially to Miss Sayang Binti Baba for all the guidance throughout this project. Leaving no stone unturned, more grease to the elbow Musa Ahmed Abubakar for dedicating his time in the completion of my thesis write-up.

Last but not the least, my deepest appreciation goes to my beloved family members and my intimate friends that have been my pillars of strength!

ABSTRACT

Citrus hystrix (C.hystrix) locally known as Limau purut is a culinary flavouring that is common in Malaysia and South East Asian cuisine as well as in herbal preparations. The aim of this study is to evaluate the antiproliferative, antioxidant and total phenolic content of ethanolic and aqueous extract of C.hystrix leaves. Qualitative phytochemical screening was also conducted to detect the presence of phytochemical compounds. The cytotoxic effect of the extracts on colon cancer cells (HT29) were determined by MTT assay. For the antioxidant activity, 2, 2, diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging assay was used. Total phenolic content (TPC) was evaluated by using Folin-Ciocalteu method. Qualitative phytochemical screening was carried using standard procedures. The ethanolic extract of C.hystrix exhibited higher percentage yield of 22.05 % compared to 19.22 % of aqueous during the solvent extraction method. The cytotoxic activity of the extracts showed IC₅₀ values of 0.25 and 3.25 mg/ mL for ethanolic and aqueous extract respectively indicating stronger cytotoxic effects on HT29 cells in ethanolic extract. The TPC was also higher in ethanolic extract compared to aqueous which was 0.12 and 0.104 mg of GA/ g of extract respectively. For the antioxidant activity, both the ethanolic and aqueous extract showed IC_{50} values of > 8 mg/mL in comparison of 0.03 mg/mL of the standard ascorbic acid, indicating low antioxidant activity. Both the ethanolic and aqueous extract of C.hystrix exhibited a weak, negative correlation between TPC and percentage inhibition of DPPH with R² value of 0.2278 and 0.434 respectively. The phytochemical screening of the extracts reveals the presence of several phytochemical compounds namely alkaloid, saponin, flavonoid and fixed oil and fats with maximum presence of phytochemicals in ethanolic compared to aqueous extract of C.hystrix. Relatively, the present result shows that the ethanolic extract of *C.hystrix*, which exhibited higher percentage yield, antiproliferative activity, TPC content and maximum presence of phytochemicals can be used as a potential source of anticancer agent but not a good source of natural antioxidants.

ABSTRAK

Citrus hystrix (C.hystrix) atau dikenali sebagai Limau purut digunakan dalam masakan Malaysia dan Asia Tenggara serta dalam perubatan tradisional. Tujuan kajian ini dilakukan adalah untuk mengesan aktiviti anti- kanser, anti- oksida, jumlah kandungan fenolik serta penyaringan sebatian fitokimia di dalam ekstrak ethanol dan aqueous daun C.hystrix. Aktiviti anti-kanser ekstrak daun limau purut di atas sel kanser kolon (HT29) dianalisis menggunakan kaedah MTT. Aktiviti anti-oksida dianalisis menggunakan kaedah DPPH. Jumlah kandungan fenolik ditentukan menggunakan kaedah Folin- Ciocalteu. Kesemua kehadiran bahan fitokimia ditentukan dengan menggunakan ujian penyaringan bahan fitokimia.Hasil kajian mendapati ekstrak ethanol daun limau purut mempunyai jumlah berat yang lebih tinggi iaitu 22.05 % berbanding ekstrak aqueous iaitu 19.22 %. Ekstrak ethanol juga menunjukkan nilai IC₅₀ yang lebih rendah iaitu 0.25 mg/ mL berbanding ekstrak aqueous iaitu 3.25 mg/ mL dalam ujian anti- kanser terhadap sel kanser kolon .Bagi ujian jumlah kandungan fenolik, ekstrak ethanol menunjukkan nilai lebih tinggi iaitu 0.12 mg GA/ g dibandingkan dengan ekstrak aqueous iaitu 0.104 mg GA/g. Bagi aktiviti anti – oksida kedua- dua ekstrak ethanol dan aqueous menunjukkan nilai IC_{50} > 8 mg/mL dibandingkan dengan nilai peratus inhibisi ascorbic acid iaitu 0.03 mg/ mL yang menunjukkan aktiviti anti- oksida kedua-dua ekstrak daun limau purut adalah rendah.Ujian penyaringan fitokimia telah mengesan beberapa bahan fitokimia iaitu alkaloid, saponin, flavonoid, dan minyak/lemak. Berbanding dengan ekstrak aqueous, lebih banyak bahan fitokimia dikesan di dalam ekstrak ethanol. Kedua- dua ekstrak ethanol dan aqueous daun limau purut menunjukkan polarisasi negatif di antara jumlah kandungan fenolik dan peratus (%) inhibisi DPPH di mana nilai R² adalah 0.2278 bagi ekstrak ethanol dan 0.4344 bagi ekstrak aqueous. Secara keseluruhan, didapati ekstrak ethanol C.hystrix mempunyai jumlah ekstrak tinggi serta mempunyai aktiviti anti – kanser, kandungan fitokimia dan fenolik yang lebih tinggi. C.hystrix berpotensi untuk digunakkan sebagai agen anti-kanser namun bukan sebagai anti-oksida semula jadi.

TABLE OF CONTENTS

CHAPTER

2

TITLE

PAGE

6

DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGEMENTS	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	X
LIST OF FIGURES	xi
LIST OF ABBREVIATIONS/SYMBOLS	xiii
LIST OF APPENDICES	XV

1	INTRODUCTION		
	1.1	Background of Study	1
	1.2	Statements of Problem	3
	1.3	Objectives of Study	4
	1.4	Scope of Study	4

LITERATURE REVIEW

vii

2.1	Citrus hystrix	6
2.2	Antioxidant	9
2.3	Colon cancer	10
	2.3.1 HT-29 Colon cancer cells	12
2.4	Phytochemicals	12
	2.4.1 Phenolic	13
	2.4.2 Flavonoids	15
	2.4.3 Alkaloids	16
	2.4.4 Terpenoids	17
	2.4.5 Tannins	18
	2.4.6 Saponin	19

MATERIALS AND METHODS

3

3.1	Chemicals and reagents 21		
3.2	Experimental design	22	
3.3	Methods	23	
	3.3.1 Culture of HT-29 Colon cancer cells	3 23	
	3.3.2 Preparation of crude plant extract	23	
	3.3.2.1 Ethanol crude extract	23	
	3.3.2.2 Aqueous crude extract	24	
3.4	Cytotoxicity (MTT) assay	24	
3.5	Antioxidant assay	25	
	(DPPH radical scavenging activity)		
3.6	Total Phenolic Content (TPC)	26	
3.7	Qualitative phytochemical screening 27		
	3.7.1 Screening for phenolic compound	27	
	3.7.1.1 Ferric chloride test	27	
	3.7.2 Tannin test	27	
	3.7.3 Saponin/ Froth test	28	
	3.7.4 Fats and fixed oil test	28	

	3.7.5	Terpenoid (Salkowski) tests	28
	3.7.6	Flavonoid test	28
	3.7.7	Alkaloid test	29
3.8	Statist	cical Analysis	29

4 **RESULT AND DISCUSSIONS**

4.1	Extraction of C.hystrix leaves	30
4.2	Cytotoxicity (MTT assay)	31
4.3	Total Phenolic Content (TPC) assay	33
4.4	DPPH radical scavenging activity	36
4.5	Correlation between TPC and DPPH	39
	radical scavenging activity	
4.6	Phytochemical screening	41
	4.6.1 Test for phenolic compounds	42
	4.6.2 Tannin test	43
	4.6.3 Saponin (Froth) test	44
	4.6.4 Fixed oils and fats test	45
	4.6.5 Flavonoids test	46
	4.6.6 Terpenoids test	47
	4.6.7 Alkaloid test	48

5 CONCLUSION AND RECOMMENDATIONS 51

LIST OF REFERENCES	53
APPENDICES	67

30

LIST OF TABLE

TABLE NO.

TITLE

PAGE

2.1	Plant Taxonomy of C.hystrix	7
4.1	The percentage yield of the plant using ethanolic and	
	aqueous extraction	31
4.2	Percentage viability and inhibition of HT-29 colon	32
	cancer cells tested against different concentrations	
	of ethanolic and aqueous extract of C.hystrix leaves	
4.3	IC ₅₀ values of ethanolic and aqueous extract of <i>C.hystrix</i>	33
	leaves	
4.4	Total phenolic contents in the ethanolic and aqueous	36
	extract of C.hystrix leaves expressed in terms of gallic	
	acid equivalent	
4.5	Percentage inhibition of DPPH tested against different	38
	concentrations of ethanolic and aqueous extract of C.hystrix	
	leaves	
4.6	IC ₅₀ values of ethanolic and aqueous extract of <i>C.hystrix</i>	
	leaves	39
4.7	Qualitative analysis of phytochemical constituents of	
	C.hystrix in ethanolic and aqueous extract	42

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Citrus hystrix	6
2.2	Polyps in the colon	11
2.3	Pie chart representing the major groups of plant	13
	phytochemicals	
2.4	Major classes of phenolic compounds in plants	14
2.5	The chemical structures of major classes of flavonoids	16
2.6	Mode of action of berberine, evodiamine, matrine, piperine,	17
	sanguinarine, and tetrandrine on cancer cells	
2.7	Classifications of tannins	19
3.1	Experimental design of the overall scope of study	22
4.1	The graph of percentage inhibition of HT-29 colon cancer ce	11 33
	treated with different concentration of ethanolic and aqueous	1
	extract of C.hystrix leaves	
4.2	Concentration of gallic acid (mg/mL) against absorption at	35
	750 nm	
4.3	Percentage inhibition of the ethanolic and aqueous extracts of	f 39
	C.hystrix leaves and standard on DPPH	
4.4	Correlation between TPC and percentage inhibition of DPPH	
	For ethanolic extract of <i>C.hystrix</i> leaves.	40
4.5	Correlation between TPC and percentage inhibition of DPPH	41
	for aqueous extract of C.hystrix leaves	
4.6	Ferric chloride test of aqueous and ethanolic crude extracts of	f 43
	C.hystrix leaves	
4.7	Gelatin test of aqueous and ethanolic crude extract of C.hystr	rix 44

leaves

4.8	Saponin/ froth test of aqueous and ethanolic extract of	45
	C.hystrix leaves	
4.9	Fixed oil and fats test of aqueous and ethanolic crude extract of	
	C.hystrix leaves	46
4.10	Flavonoid test of aqueous and ethanolic extract of C.hystrix	47
	leaves	
4.11	Terpenoids test of aqueous and ethanolic extract of C.hystrix	48
	leaves	
4.12	Alkaloid test of aqueous and ethanolic extract of C.hystrix	49
	leaves	

LIST OF ABBREVIATIONS/ SYMBOLS

ANOVA	-	analysis of variance
ATC	-	animal tissue culture
BHA	-	butylated hydroxyl anisole
BHT	-	butylated hydroxyl toluene
CHCl ₃	-	chloroform
CO_2	-	carbon dioxide
DPPH	-	2, 2-diphenyl-1-picrylhydrazyl
H_2SO_4	-	sulphuric acid
GAE	-	gallic acid equivalent
HCl	-	hydrochloric acid
HT-29	-	human colorectal adenocarcinoma cell line
IC	-	inhibitory concentration
K	-	potassium
K562	-	human erythromyeloblastoid leukemia cell line
KB	-	human mouth epidermal carcinoma
L	-	litre
Ml	-	milliliter
Mg	-	milligram
MTT	-	3(4,5- <u>dimethylthiazol</u> -2-yl)-2,5-diphenyltetrazoliumbromide
Molt4	-	human leukemia cells
μg	-	microgram
NCI	-	National Cancer Institute
OD	-	optical density
PBS	-	phosphate buffer saline
PG	-	propyl gallate
P388	-	murine leukemic cell lines

RPM1- 1640	-	Roswell Park Memorial Institute
SPSS	-	Statistical Package for the Social Sciences
TPC	-	total phenolic content
TAC	-	total antioxidant capacity
TPA	-	12-Otetradecanoylphorbol 13-acetate
TBHQ	-	tertiarybutyl hydroquinone
USA	-	United States of America
USDA	-	United States of Department griculture
UV	-	ultraviolet
U937	-	human macrophage cell line
WHO	-	World Health Organization
%	-	percentage
°C	-	degree Celsius
EW	-	extract weight
DW	-	dry weight

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Colon cancer, also known as the colorectal cancer is the fourth leading cause of cancer death worldwide (American Cancer Society, 2014). In Malaysia, a total of 2 866 cases were registered nationwide with the Malaysian National Cancer Registry in 2006 and represent 13.2% of all cancer cases registered (Kong *et al.*, 2010). Patients diagnosed with colorectal cancer often undergoes treatments such as chemotherapy, radiation therapy and surgery (American Cancer Society, 2014).However, chemotherapy works by killing abnormally fast dividing cells and thus effects cells that divide rapidly under normal circumstances, which leads to unwanted side effects (Chueahongthong *et al.*, 2011). Hence, naturally occurring plant compounds that have cancer inhibitory effects but have fewer side effects are a valuable alternative treatment for cancer (Jusoh *et al.*, 2012).

Medicinal plants have played an important role in the treatment of various types of cancers (Valiyari *et al.*, 2012). Numerous plant-derived compounds including taxol from *Taxus brevifolia*, camptothecin from *Camptotheca acuminate*, Decne, vinca alkaloids from *Catharanthus roseus*. Don and podophyllotoxin from

Podophyllum peltuturn are used in clinic all over the world (Valiyari *et al.*, 2012; Chueahongthong *et al.*, 2011).

In this project, the cytotoxic effects of ethanolic and aqueous crude extract of *Citrus hystrix* leaves on colon cancer cells (HT29) were examined. *Citrus hystrix* which is locally known as *Limau purut* or Kaffir lime is one of the traditional medicinal plant that has gain the interest of many researches over the decades due to their potential source of natural antioxidants (Almey *et al.*, 2010; Jamilah *et al.*, 1998; Nurain *et al.*, 2013; Wong *et al.*, 2006; Idris *et al.*, 2008). Besides its leaves, the crude extract of *C.hystrix* such as peels, stems, and juice have shown potential antioxidant activity as well (Ghafar *et al.*, 2010; Chowdhury *et al.*, 2009). *C.hystrix* also exhibits anti-microbial (Chowdhury *et al.*, 2012; Suri *et al.*, 2002) activities.

In regard to cancer research, *Citrus hystrix* extracts have been shown to have anti-proliferative activity on KB (cervical cancer) and P388 (mouse leukemia) cell lines (Manosroi *et al.*, 2006).Glyceroglycolipids in *Citrus hystrix* leaves could inhibit 12-Otetradecanoylphorbol 13-acetate (TPA) and skin carcinogen activities in mice (Murakami *et al.*, 1995). According to research conducted by Ampasavate *et al.* 2010, *C.hystrix* leaves had strong cytotoxic effects on four leukemic cell lines (U937, K562, HL60, Molt 4).

The leaves of *C.hystrix* are used in many Malaysian and South- East Asian regions cuisines, for example in the famous local dishes such as Tom yam, stews, curries, and sauces (Almey *et al.*, 2010; Nurain *et al.*, 2013). It is also well known for its medicinal properties in treating skin disorders (Aziman *et al.*, 2012). Besides, it has also been used in aromatheraphy, nutraceutical, and personal care products (Almey *et al.*, 2010). Due to its various ethnomedical properties, this plant was selected for this study (Almey *et al.*, 2010; Aziman *et al.*, 2012). Also, as the leaves of this plant have been used as a spice in many Thai and Malaysian cuisines, the safety of *C.hystrix* leaves are highly acceptable (Chueahongthong *et al.*, 2011). Till date, little is known concerning the effect of crude extract of *Citrus hystrix* leaves on colon cancer (HT29) cells.

The aim of this project was to determine the cytotoxic effect of ethanolic and aqueous crude extracts of *C.hystrix* leaves on colon cancer cells (HT29), to determine the antioxidant activity and total phenolic content (TPC) of the crude extracts, as well as to determine the phytochemical compounds present in each extract by performing phytochemical screening.

1.2 Statements of Problem

The current medical treatment for colorectal cancer includes polypectomy (removing the polyp) or colon resection (colectomy) through surgery, chemotherapy and radiation therapy (American Cancer Society, 2014). However, some of these methods cause undesired side effects by the non-specific targeting of both normal and cancer cells (Chueahongthong *et al.*, 2011). Thus, research for cancer treatment by using natural products has been increasing rapidly (Valiyari, 2012). Among natural sources, medicinal plants have played an important role in the treatment of many forms of cancer. Numerous studies have identified medicinal plant extracts which not only displayed antioxidant activity, but also cytotoxicity to many forms of cancer (Valiyari, 2012; Chueahongthong *et al.*, 2011). Such compounds include curcumin (Sandur *et al.*, 2007), guava extract, and basil leaf extract (Manosroi *et al.*, 2006).

Morever, synthetic antioxidants such as butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA), tertiarybutyl hydroquinone (TBHQ) and propyl gallate (PG) are conventional food antioxidants which are being added to foods to prevent the oxidation process of foods during the exposure to environments and increase the shelf life (Jamilah *et al.*, 2011; Wong *et al.*, 2006). Due to increasing regulatory scrutiny, consumer concerns and safety issues concerning synthetic antioxidants, the possibility of natural antioxidants from plants, as an alternative to the synthetic antioxidants are also being actively studied (Almey *et al.*, 2010; Nurain *et al.*, 2013; Jamilah *et al.*, 2011). Due to their natural origin, antioxidant from plants,

does not exhibit side effects and safer for consumption in comparison to synthetic antioxidants (Nurain *et al.*, 2013).

Citrus hystrix leaves have various medical and culinary uses in South East Asia (Almey *et al.*, 2010; Aziman *et al.*, 2012). As the plant is used mostly for cuisine purposes, the cytotoxic effect of the *Citrus hystrix* leaf crude extract on colon cancer cells are studied.

1.3 Research Objectives

Followings are the objectives of this research:

- 1.3.1 To investigate the cytotoxic effect of *C.hystrix* leaf extract on human colon cancer cells (HT29).
- 1.3.2 To determine the antioxidant properties of *C.hystrix* leaf extract.
- 1.3.3 To investigate the total phenolic content of *C.hystrix* leaf extract.
- 1.3.4 To identify the presence of phytochemical compounds present in *C.hystrix* leaf extract by performing phytochemical screening.

1.4 Scope of Research

This project was aimed to determine the cytotoxic effect of *Citrus hystrix* crude extracts on colon cancer cells, to determine the antioxidant and total phenolic content of C. *hystrix* leaves extracts as well as to identify phytochemical compounds present in the crude extract of the leaves. The leaves were extracted by using two

polar solvents namely aqueous and ethanol. Colon cancer cells, (HT-29) were used to test the cytotoxic effect of *C. hystrix* by using the MTT assay. 50% inhibition concentration (IC₅₀) of the active substances was determined as the lowest concentration which reduced cell growth by 50%. The antioxidant activity was measured by using the DPPH free radical scavenging assay and expressed as percentage of DPPH radical inhibition and IC₅₀ values. The total phenolic content was determined by using the Folin- Ciocalteu method and is expressed as mg of GA/g of extract. Following that, phytochemical screening was conducted to determine the presence of phytochemicals such as phenol, tannin, saponin, flavonoid, alkaloid, terpenoids and fixed oils.

REFERENCES

- Abeysinghe, D. C., Li, X., Sun, C., Zhang, W., Zhou, C. and Chen, K. (2007). Bioactive Compounds and Antioxidant Capacities in Different Edible Tissues of Citrus Fruit of Four Species. *Food Chemistry*. 104(4): 1338-1344.
- Aharoni, A., Jongsma, M. A., Kim, T. Y., Ri, M. B., Giri, A. P., Verstappen, F. W.A., Schwab, W. and Bouwmeester, H. J. (2006). Metabolic Engineering of Terpenoid Biosynthesis in Plants. *Phytochemistry Reviews*. 5(1): 49-58.
- Akrout, A., Mighri, H., Krid, M., Thabet, F., Turki, H., El-Jani, H. and Neffati, M. (2012). Chemical Composition and Antioxidant Activity of Aqueous Extracts of Some Wild Medicinal Plants in Southern Tunisia. *International Journal of Life Science and Medical Science*. 2(1): 1-4.
- Aksoy, L., Kolay, E., Ağılönü, Y., Aslan, Z. and Kargıoğlu, M. (2013). Free Radical Scavenging Activity, Total Phenolic Content, Total Antioxidant Status, and Total Oxidant Status of *Endemic Thermopsis Turcica*. Saudi Journal of Biological Sciences. 20(3): 235-239.
- Almey, A. A., Ahmed Jalal Khan, C., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R. and Kamarul Rahim, K. (2010). Total Phenolic Content and Primary Antioxidant Activity of Methanolic and Ethanolic Extracts of Aromatic Plants Leaves. *International Food Research Journal*. 17(4): 1077-1084.

- American Cancer Society. (2014). Colorectal Cancer Facts and Figures. Retrieved January15,2015,from:http://www.cancer.org/research/cancerfactsstatistics/c olorectal-cancer-facts-figures.
- Ampasavate, C., Okonogi, S. and Anuchapreeda, S. (2010). Cytotoxicity of Extracts from Fruit Plants against Leukemic Cell Lines. *African Journal of Pharmacology*. 4(1): 13-21.
- Ayoola, G. A., Coker, H. A., Adesegun, S. A., Adepoju-Bello, A. A., Obaweya, K., Ezennia, E. C. and Atangbayila, T. O. (2008). Phytochemical Screening and Antioxidant Activities of Some Selected Medicinal Plants Used for Malaria Therapy in Southwestern Nigeria. *Tropical Journal of Pharmaceutical Research*.7(3): 1019-1024.
- Aziman, N., Abdullah, N., Noor, Z. M., Zulkifli, K. S. and Kamarudin, W. S. S. W. (2012). Phytochemical Constituents and In Vitro Bioactivity of Ethanolic Aromatic Herb Extracts. *Sains Malaysiana*. 41(11): 1437-1444.
- Bachrach, Z. Y. (2012). Contribution of Selected Medicinal Plants for Cancer Prevention and Therapy. Acta Facultatis Medicae Naissensis. 29(3):117-123.
- Bakar, J. (2011). Phenolics in *Citrus Hystrix* Leaves Obtained Using Supercritical Carbon Dioxide Extraction. *International Food Research Journal*. 18(3): 941-948.
- Barile, E., Bonanomi, G., Antignani, V., Zolfaghari, B., Sajjadi, S. E., Scala, F. and Lanzotti, V. (2007). Phytochemical Screening and Antimicrobial Assessment of Abutilon Mauritianum, Bacopa Monifera and Datura Stramonium.Phytochemistry. 68: 596-603.
- Béjar, L.M., Gili, M., Infantes, B. and Marcott, P.F., (2012). Incidence of Colorectal Cancer and Influence of Dietary Habits in Fifteen European Countries from 1971 to 2002. *Gaceta Sanitaria*.26: 69-73.

- Benyhe, S. (1994). Morphine: New Aspects in the Study of an Ancient Compound. *Life Sciences*.55 (13): 969–979.
- .Benzie, I. F. F., and Strain, J. J. (1996). The Ferric Reducing Ability of Plasma (FRAP) as A Measure of "Antioxidant Power": *The FRAP Assay. Analytical Biochemistry*. 239: 70–76.
- Bouayed, J., and Bohn, T. (2010). Exogenous Antioxidants—Double-Edged Swords in Cellular Redox State: Health Beneficial Effects at Physiologic Doses versus Deleterious Effects at High Doses. Oxidative Medicine and Cellular Longevity. 3(4): 228-237.
- Bruyne, T. D., Pieters, L., Deelstra, H. and Vlietinck, A. (1999). Condensed Vegetable Tannins: Biodiversity in Structure and Biological Activities. *Biochemical Systematics and Ecology*. 27(4): 445-459.
- Butryee, C., Sungpuag, P. and Chitchumroonchokchai, C. (2009). Effect of Processing on the Flavonoid Content and Antioxidant Capacity of *Citrus Hystrix* Leaf. *International Journal of Food Sciences and Nutrition*. 60(S2): 162-174.
- Chan, S.W., Lee, C.Y., Yap, C.F., Wan Aida, W.M. and Ho, C.W. (2009) Optimisation of Extraction Conditions for Phenolic Compounds from Limau Purut (*Citrus Hystrix*) Peels. *International Food Research Journal*.16:203– 213.
- Ching, L. S. and Mohamed, S. (2001). Alpha-Tocopherol Content in 62 Edible Tropical Plants. *Journal of Agricultural and Food Chemistry*. 49(6): 3101-3105.
- Chowdhury, A., Alam, M. A., Rahman, M. S., Hossain, M. A. and Rashid, M. A. (2009). Antimicrobial, Antioxidant and Cytotoxic Activities of *Citrus Hystrix DC*. Fruits. *Dhaka University Journal of Pharmaceutical Sciences*. 8(2): 177-180.

- Chueahongthong, F., Ampasavate, C., Okonogi, S., Tima, S. and Anuchapreeda, S. (2011). Cytotoxic Effects of Crude Kaffir Lime (*Citrus Hystrix*, DC.) Leaf Fractional Extracts on Leukemic Cell Lines. *Journal of Medicinal Plant Research*. 5(14): 3097-3105.
- Cohen, E., Ophir, I. and Shaul, Y. B. (1999). Induced Differentiation in HT29, A Human Colon Adenocarcinoma Cell Line. *Journal of Cell Science*. 112(16): 2657-2666.
- Conforti, F., Sosa, S., Marrelli, M., Menichini, F., Statti, G. A., Uzunov, D., Tubaro, A.T., Menichini, F. and Loggia, R. D. (2008). In Vivo Anti-Inflammatory and In Vitro Antioxidant Activities of Mediterranean Dietary Plants. *Journal of Ethnopharmacology*. 116(1):144-151.
- Cook, N. C. and Samman, S. (1996). Flavonoids—Chemistry, Metabolism, Cardioprotective Effects, and Dietary Sources. *The Journal of Nutritional Biochemistry*, 7(2): 66-76.
- Day, A. J. and Williamson, G. (2001). Biomarkers for Exposure to Dietary Flavonoids: A Review of the Current Evidence for Identification of Quercetin Glycosides in Plasma. *British Journal of Nutrition*, 86(S1): S105-S110.
- Del –Rio, J.A., Obdulio, B.G., Castillo, J., Marin, F.R. and Ortuno, A. (1997).Uses and Properties of Citrus Flavonoids. *Journal of Agricultural and Food Chemistry*. 45(12): 4505-4514.
- Dintinjana, R. D., Petranović, D., Pilčič, G., Ilijic, V., Petranović, D., and Dintinjana,
 M. (2012). Unusual Spreading of Colon Adenocarcinoma into the Right Hip
 Area: A Case Report. *Journal of Cancer Therapy*. 3: 985-988.
- Diplock, A.T., Charleux, J.L., Crozier-Willi, G., Kok, F.J., Rice-Evans, C., Roberfroid, M., Sthal, W. and Vina-Ribes, J. (1998). Functional Food

Science and Defence against Reactive Oxygen Species. *British Journal of Nutrition.* 80: S77-S112.

Dugo, G., and Di Giacomo, A. (Eds.). (2003). Citrus: The Genus Citrus. CRC Press.

- Forgue-Lafitte, M. E., Coudray, A. M., Bréant, B. and Mešter, J. (1989). Proliferation of the Human Colon Carcinoma Cell Line HT29: Autocrine Growth and Deregulated Expression of the C-Myc Oncogene. *Cancer Research*. 49(23): 6566-6571.
- Fotakis, G. and Timbrell, J. A. (2006). In Vitro Cytotoxicity Assays: Comparison of LDH, Neutral Red, MTT and Protein Assay in Hepatoma Cell Lines following Exposure to Cadmium Chloride. *Toxicology letters*. 160(2): 171-177.
- Frutos, P., Hervas, G., García, F. G. and Mantecón, A. R. (2004). Review. Tannins and Ruminant Nutrition. Spanish Journal of Agricultural Research. 2(2): 191-202.
- Garg, A., Garg, S., Zaneveld, L. J. D. and Singla, A. K. (2001). Chemistry and Pharmacology of the Citrus Bioflavonoid Hesperidin. *Phytotherapy Research*. 15(8): 655-669.
- Ghafar, M. F., Prasad, K. N., Weng, K. K. and Ismail, A. (2010). Flavonoid, Hesperidine, Total Phenolic Contents and Antioxidant Activities from Citrus Species. *African Journal of Biotechnology*. 9(3): 326-330.
- Ghazali, M. A. M., Ghanya, A. N., Kesavanarayanan, K. S. and Hazizul, M. (2014). Apoptosis Induction by *Polyonum Minus* is Related to Antioxidant Capacity, Alteration in Expression of Apoptotic-Related Genes and S-phase Cell Cycle Arrest in HepG2 cell line. *BioMed Research International*. 2014.

- Greiner, K.A., Engelman, K.K., Hall, M.A. and Ellerbeck, E.F. (2004).Barriers to Colorectal Cancer Screening in Rural Primary Care.*Preventive Medicine*. 38: 269-75.
- Guessous, I., Dash, C. and Lapin, P. (2010). Colorectal Cancer Screening. Barriers and Facilitators in Older Persons. *Preventive Medicine*. 50: 3-10.
- Gursoy, N., Sarikurkcu, C., Cengiz, M. and Solak, M. H. (2009). Antioxidant Activities, Metal Contents, Total Phenolics and Flavonoids of Seven Morchella Species. Food and Chemical Toxicology. 47(9):2381-2388.
- Hilmi, I., Hartono, J. L.and Goh, K. L. (2010). Negative Perception in Those at Highest Risk-Potential Challenges in Colorectal Cancer Screening in an Urban Asian Population. Asian Pac J Cancer Prev, 11(3), 815-22.
- Huda-Faujan, N., Noriham, A., Norrakiah, A. S. and Babji, A. S. (2009). Antioxidant Activity of Plants Methanolic Extracts Containing Phenolic Compounds. *African Journal of Biotechnology*. 8 (3): 484-489.
- Idris, N.A., Nor, M.F., Ismail, R., Mohamed, S. and Hassan, Z.C. (2008). Antioxidative Activity of Malaysian Herb Extracts in Refined, Bleached and Deodorized Palm Olein. *Journal of Oil Palm Research*. 20: 517-526.
- Imelda, F., Faridah, D. N. and Kusumaningrum, H. D. (2014). Bacterial Inhibition and Cell Leakage by Extract of *Polygonum Minus Huds* Leaves. *International Food Research Journal*, 21(2).
- Jamilah, B., Man, Y. B. and Ching, T. L. (1998). Antioxidant Activity of *Citrus Hystrix* Peel Extract in Refined Bleached and Deodorized Palm Oleen during Frying of Fish Crackers. *Journal of Food Lipids*. 5(2): 149-157.
- Jamilah, B., Abdulkadir Gedi, M., Suhaila, M. and Md.Zaidul. (2011). Phenolics in *Citrus Hystrix* Leaves Obtained Using Supercritical Carbon Dioxide Extraction. *International Food Research Journal*. 18(3): 941-948.

- Janecka, I. P. (2007). Colon Cancer and Physical Activity: A Content Analysis of Reciprocal Relationship. *Clinical Medicine: Oncology*. 1: 81-89.
- Jusoh, S. A. M., Johan, M. F. and Mohamed, A. S. (2012). Anti-Proliferative and Pro-Apoptotic Effect of *P. sacharosa, E. elatior* and *P. granatum* Aqueous Extract on Human Myeloid Leukaemia: A Preliminary Study. *Asian Journal* of Medical Research. 1(4): 146-151.
- Kaur, C. and Kapoor, H. C. (2001). Antioxidants in Fruits and Vegetables–The Millennium's Health. International Journal of Food Science & Technology. 36(7): 703-725.
- Khanbabaee, K., and van Ree, T. (2001). Tannins: Classification and Definition. *Natural Product Reports*. 18(6): 641-649.
- Koche, D., Shirsat, R., Imran, S. and Bhadange, D. G. (2010). Phytochemical Screening of Eight Traditionally Used Ethnomedicinal Plants from Akola District (MS) India. International *Journal of Pharmacology and Bio Sciences*. 1(4).
- Kong, C. K., Roslani, A. C., Law, C. W., Law, S. C. and Arumugam, K. (2010). Impact of Socio-Economic Class on Colorectal Cancer Patient Outcomes in Kuala Lumpur and Kuching, Malaysia. *Asian Pac J Cancer Prev*, 11(4): 969-974.
- Krishnaiah, D., Devi, T., Bono, A. and Sarbatly, R. (2009). Studies on Phytochemical Constituents of Six Malaysian Medicinal Plants. *Journal of Medicinal Plants Research*. 3(2): 067-072.
- Laohavechvanich, P., Muangnoi, C., Butryee, C. and Kriengsinyos, W. (2010). Protective Effect of Makrut Lime Leaf (*Citrus Hystrix*) in HepG2 cells: Implications for Oxidative Stress. *Science Asia*. 36: 112-117.

- Lavhale, R. R. (2011). Pharmacognostic, Phytochemical Investigation of Polygonum Glabrum Willd Leaves and Screening for Antioxidant Activity. Doctoral Dissertation, KLE University, Karnataka, Belgaum.
- Lasser, K.E., Ayanian, J.Z., Fletcher, R.H. and Good, M.J. (2008). Barriers to Colorectal Cancer Screening in Community Health Centers: A Qualitative Study. *BMC Fam Prac*. 9: 15-22.
- Lim, T. K. (2012). *Citrus Hystrix*. In Edible Medicinal and Non-Medicinal Plants. *Springer Netherlands*. 4: 634-643.
- Lin, Y., Huang, G., Huang, Y., Tzeng, T. R. J. and Chrisey, D. (2010). Effect of Laser Fluence in Laser-Assisted Direct Writing of Human Colon Cancer Cell. *Rapid Prototyping Journal*: 16(3): 202-208.
- Liu, H. Y., Huang, Z. L., Yang, G. H., Lu, W. Q. and Yu, N. R. (2008). Inhibitory Effect of Modified Citrus Pectin on Liver Metastases in a Mouse Colon Cancer Model. *World J Gastroenterol*. 14(48):7386-7391.
- Lu, J. J., Bao, J. L., Chen, X. P., Huang, M. and Wang, Y. T. (2012). Alkaloids Isolated from Natural Herbs as The Anticancer Agents. *Evidence-Based Complementary and Alternative Medicine*.10: 1155.
- Magalhães, L. M., Santos, F., Segundo, M. A., Reis, S. and Lima, J. L. (2010). Rapid Microplate High-Throughput Methodology for Assessment of Folin-Ciocalteu Reducing Capacity. *Talanta*. 83: 441-447.
- Mamta, S. and Jyoti, S. (2012). Phytochemical Screening of Acorus Calamus and Lantana Camara. International Research Journal of Pharmacy. 3(5): 324-326.
- Mangan, J. L. (1988). Nutritional Effects of Tannins in Animal Feeds. *Nutrition Research Reviews*. 1(01): 209-231.

- Manosroi, J., Dhumtanom, P. and Manosroi, A. (2006). Anti-Proliferative Activity of Essential Oil Extracted from Thai Medicinal Plants on KB and P388 Cell Lines. *Cancer Letters*. 235(1):114-120.
- Maw, S. S., Mon, M. M. and Oo, Z. K. (2011). Study on Antioxidant and Antitumor Activities of Some Herbal Extracts. World Academy of Science Engineering and Technology. 5: 2011-03-21.
- Middleton, E.J. (1998). Effect of plant flavonoids on immune and inflammatory cell function. *Advances in Experimental Medicine and Biology*.439:175–82.
- Mueller Harvey, I. and McAllan, A. B. (1992). *Tannins:* Their Biochemistry and Nutritional Properties. *Advances in Plant Cell Biochemistry and Biotechnology*. 1: 151-217.
- Murakami, A., Nakamura, Y., Koshimizu, K. and Ohigashi, H. 1995. Glyceroglycolipids from *Citrus Hystrix*, A Traditional Herb in Thailand, Potently Inhibit the Tumor- Promoting Activity of 12-Otetradecanoylphorpol 13-acetate In Mouse Skin. *Journal of Agricultural and Food Chemistry*. 43: 2779-2783.
- Najib Nik A Rahman, N., Furuta, T., Takane, K. and Ali Mohd, M. (1999). Antimalarial Activity of Extracts of Malaysian Medicinal Plants. *Journal of Ethnopharmacology*. 64(3): 249-254.
- National Cancer Institute. (2014). Colon Cancer Treatment. Retrieved January 17, 2015, from: http://www.cancer.gov/cancertopics/pdq/treatment/colon/.
- Nijveldt, R. J., Van Nood, E. L. S., Van Hoorn, D. E., Boelens, P. G., Van Norren, K. and Van Leeuwen, P. A. (2001). Flavonoids: A Review of Probable Mechanisms of Action and Potential Applications. *The American Journal of Clinical Nutrition*.74 (4): 418-425.

- Nomura, A. M., Wilkens, L. R., Murphy, S. P., Hankin, J. H., Henderson, B. E., Pike, M. C. and Kolonel, L. N. (2008). Association of Vegetable, Fruit, and Grain Intakes with Colorectal Cancer: The Multiethnic Cohort Study. *The American Journal of Clinical Nutrition*. 88(3): 730-737.
- Nurain, A., Noriham, M.N. Zainon, W.S.K. Wan Saidatul and S.Z. Khairusy, (2013). Comparative Study of Aqueous and Ethanolic Aromatic Malaysian Herbs Extracts Using Four Antioxidant Activity Assays. *International Journal of Agricultural Research.* 8: 55-66.
- O'Brien, C. A., Pollett, A., Gallinger, S. and Dick, J. E. (2007). A Human Colon Cancer Cell Capable of Initiating Tumour Growth in Immunodeficient Mice. *Nature*. 445(7123): 106-110.
- Okwu, D.E. (2007). Nigerian Medicinal Plants. *Medicinal and Aromatic Plant Science and Biotechnology*. 1(1): 90-96.
- Okwu, D. E. (2008). Citrus fruits: A rich Source of Phytochemicals and Their Roles in Human Health. *International Journal of Chemical Sciences*. 6(2): 451-471.
- Papadopoulou, K., Melton, R. E., Leggett, M., Daniels, M. J. and Osbourn, A. E. (1999). Compromised Disease Resistance in Saponin-Deficient Plants.*Proceedings of the National Academy of Sciences*. 96(22): 12923-12928.
- Pichersky, E. and Gershenzon, J. (2002). The Formation and Function of Plant Volatiles: Perfumes for Pollinator Attraction and Defence. *Current Opinion in Plant Biology*. 5(3): 237-243.
- Pinelo, M., Rubilar, M., Jerez, M., Sineiro, J. and Nunez, M.J. (2005). Effect of Solvent, Temperature, and Solvent-to Solid Ratio on the Total Phenolic Content and Anti Radical Activity of Extracts from Different Components

of Grape Pomace. Journal of Agricultural and Food Chemistry. 53: 2111-2117.

- Pignone, M., Saha,S., Hoerger, T. and Mandelblatt, J. (2002). Cost Effectiveness Analyses of Colorectal Cancer Screening: A Systematic Review for the US Preventive Services Task Force. *Ann Intern Med.* 137: 96-104.
- Piyachaturawat, P., Glinsukon, T. and Chanjarunee A (1985). Antifertility Effect of *Citrus Hystrix DC. Journal of Ethnopharmacol* 13(1):105–110.
- Podolak, I., Galanty, A., and Sobolewska, D. (2010). Saponins as Cytotoxic Agents: a Review. *Phytochemistry Reviews*. 9(3): 425-474.
- Prasad, M.P., Balamurugan, P. and Rajkumar, A.R. (2014). Comparative Phytochemical Analysis of Rutaceae Family (Citrus Species) Extract. International. *Journal of Scientific Research*. 3(4): 2277-8179.
- Prior, R. L., Wu, X. and Schaich, K. (2005). Standardized Methods for the Determination of Antioxidant Capacity and Phenolics in Foods and Dietary Supplements. Journal of Agricultural and Food Chemistry. 53(10): 4290-4302.
- Qader, S. W., Abdulla, M. A., Chua, L. S., Najim, N., Zain, M. M. and Hamdan, S. (2011). Antioxidant, Total Phenolic Content and Cytotoxicity Evaluation of Selected Malaysian Plants. *Molecules*. 16(4):3433-3443.
- Saha, K., Lajis, N. H., Israf, D. A., Hamzah, A. S., Khozirah, S., Khamis, S. and Syahida, A. (2004). Evaluation of Antioxidant and Nitric Oxide Inhibitory Activities of Selected Malaysian Medicinal Plants. *Journal of Ethnopharmacology*. 92(2): 263-267.
- Sandur, S.K, Pandey, M.K, Sung, B., Ahn, K.S., Murakami, A., Sethi, G., Limtrakul, P., Badmaev. V. and Aggarwal, B.B. (2007).Curcumin, Demethoxycurcumin, Bisdemethoxycurcumin, Tetrahydrocurcumn and Turmerones Differentially Regulate Anti-Inflammatory and

- Sato, M., Ramarathnam, N., Suzuki, Y., Ohkubo, T., Takeuchi, M.,and Ochi, H. (1996). Varietal Differences in the Phenolic Content and Superoxide Radical Scavenging Potential of Wines from Different Sources. *Journal of Agricultural and Food Chemistry*. 44(1):37–44.
- Saxena, M., Saxena, J., Nema, R., Singh, D. and Gupta A. (2013). Phytochemistry of Medicinal Plants. *Journal of Pharmacognosy and Phytochemistry*. 1(6): 168-182.
- Schofield, P., Mbugua, D. M. and Pell, A. N. (2001). Analysis of Condensed Tannins: A Review. Animal Feed Science and Technology. 91(1): 21-40.
- Singh, D., Singh, P., Gupta, A., Solanki, S., Sharma, E., and Nema, R. (2012). Qualitative Estimation of the Presence of Bioactive Compound in *Centella Asiatica:* An Important Medicinal Plant. *International Journal of Life Science and Medical Science*. 2(1): 5-7.
- Sparg, S.G., Light, M.E. and Staden, J. (2004). Biological Activities and Distribution of Plant Saponins. *Journal of Ethnopharmacology*.94:219–243.
- Stanković, M. S. (2011). Total Phenolic Content, Flavonoid Concentration and Antioxidant Activity of *Marrubium Peregrinum L*. Extracts. *Kragujevac Journal of Science*. 33(2011): 63-72.
- Sticher, O. (2008). Natural Product Isolation. Natural Product Reports. 25(3):517-554.
- Sun, H.X., Xie, Y., and Ye, Y.P., (2009). Advances in Saponin-Based Adjuvants. *Vaccine*. 27:1787–1796.
- Suri, R., Radzali, M., Aspollah, M., Marziah, S., Arif, Z.J. and Samsumaharto, R.A. (2002). Antibacterial Assay of *Citrus Hystrix* (Limau Purut) Extracts. *Journal of Tropical Medicinal Plants* 3(1):35–42.

- Tachakittirungrod, S., Okonogi, S. and Chowwanapoonpohn, S. (2007). Study on Antioxidant Activity of Certain Plants in Thailand: Mechanism of Antioxidant Action of Guava Leaf Extract. *Food Chemistry*. 103(2): 381-388.
- Takechi, M., Matsunami, S., Nishizawa, J., Uno, C. and Tanaka, Y. (1999). Haemolytic and Antifungal Activities of Saponins or Anti-ATPase and Antiviral Activities of Cardiac Glycosides. *Planta Medica*. 65(6): 585-586.
- United States Department of Agriculture. (2014). Plants Database. Retrieved August 26, 2014, from: http://plants.usda.gov/core/profile?symbol=CIHY2.s
- Valiyari, S. (2012). Dichloromethane and Methanol Extracts of Scrophularia oxysepala Induces Apoptosis in MCF-7 Human Breast Cancer Cells. Advanced Pharmaceutical Bulletin. 2(2): 223.
- Van Wyk, B.E. 2005. *Citrus Hystrix. Food Plants of the World: An Illustrated Guide*. Portland, OR: Timber Press. P. 139.
- Wagner, H. (2000). Bioactive Saponins from Plants: An Update. Studies in Natural Products Chemistry. 21: 633-687.
- Waikedre, J., Dugay, A., Barrachina, I., Herrenknecht, C., Cabalion, P. and Fournet,
 A. (2010). Chemical Composition and Antimicrobial Activity of the
 Essential Oils from New Caledonian *Citrus Macroptera* and *Citrus Hystrix*.
 Chemistry and Biodiversity. 7(4):871-877.
- Wang, Z.T., and Liang, G.Y. (2009). Zhong Yao Hua Xue. Shanghai Scientific & Technical. 31(23): 1987-1991.
- Walton, N.J., Mayer, M.J. and Narbad, A. (2003). Molecules of Interest: Vanillin. *Phytochemistry*. 63: 505-515.

- Willer, A. (2003). Reduction of the Individual Cancer Risk by Physical Exercise. Onkologie. 26(3): 283-289.
- Wong, S. P., Leong, L. P. and William Koh, J. H. (2006). Antioxidant Activities of Aqueous Extracts of Selected Plants. *Food chemistry*, 99(4) :775-783.
- Xu, G., Liu, D., Chen, J., Ye, X., Ma, Y. and Shi, J. (2008). Juice Components and Antioxidant Capacity of Citrus Varieties Cultivated in China. Food Chemistry, 106 (2):545-551.
- Yahya, A. H., Chong, G. H. and Tan, C. P. (2014). The Phytochemical Properties of a New Citrus Hybrid (Citrus Hystrix× Citrus Microcarpa). Science Asia. 40: 121–124.
- Yusoff, H. M., Daud, N., Noor, N. M. and Rahim, A. A. (2012). Participation and Barriers to Colorectal Cancer Screening in Malaysia. *Asian Pacific Journal* of Cancer Prevention. 13: 3983-3987.
- Zhao, J. (2007). Nutraceuticals, Nutritional Therapy, Phytonutrients, and Phytotherapy for Improvement of Human Health: A Perspective on Plant Biotechnology Application. *Recent Patents on Biotechnology*. 1(1):75-97.