IN VITRO ANTIVIRAL ACTIVITY OF Polygonum minus EXTRACTS AGAINST HERPES SIMPLEX VIRUS 1

ALVIN PAUL

UNIVERSITI TEKNOLOGI MALAYSIA

IN VITRO ANTIVIRAL ACTIVITY OF Polygonum minus EXTRACTS AGAINST HERPES SIMPLEX VIRUS 1

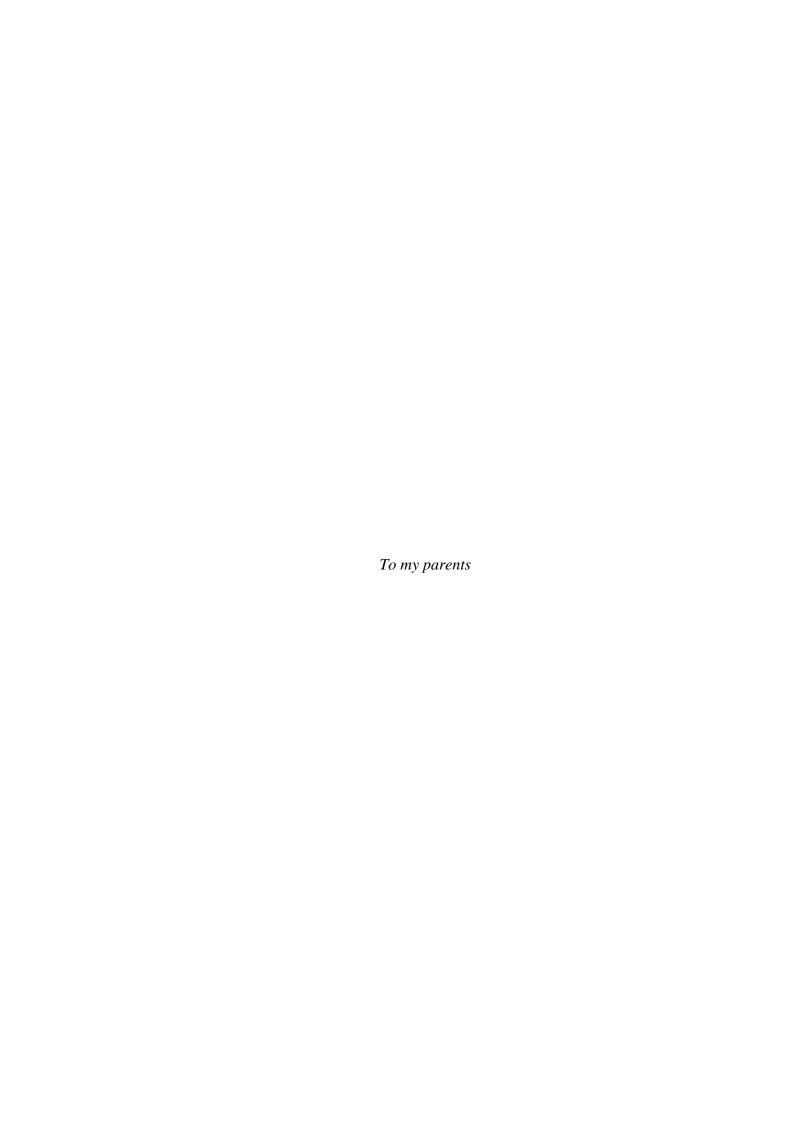
ALVIN PAUL

A dissertation submitted in partial fulfillment of the requirements for award of the degree of

Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering
Universiti Teknologi Malaysia

FEBRUARY 2015



ACKNOWLEDGEMENT

So it is done. First of all, I would like to extend my gratitude to my

supervisor, Dr. Salehhuddin Hamdan, for sharing his expertise and critical advice

throughout this project. He has been instrumental in supporting the project since day

one. Thank you for giving me the opportunity to work with you.

To Ms. Sayang Baba who had taught me the rigors of research work, my

sincerest appreciation. Her acute mentoring and perseverance in dealing with my

folly were tremendous, bar none. A special acknowledgement to Dr. Saleha Shahar

for her recommendations and willingness to discuss experimental protocols with me.

To past and present ATC lab members; Ms. Aqmar Bahaudin, Ms. Asita

Elengoe and Ms. Norsyuhada Jaafar, thank you for the assistance and guidance

during my lab work. Not forgetting friends and fellow class members who had

certainly made the journey more awesome. I am grateful to have known such

wonderful people in the short time studying here.

Finally, to my parents who supported me unconditionally all these years,

thank you. There are no words to express my gratitude. I would not have made it if it

weren't for them.

Cheers! :]

ABSTRACT

The use of medicinal plants as preventative and curative treatment may be integral when a complete treatment is not yet available. Hence, the purpose of this study was to conduct an evaluation on Polygonum minus extracts over its effect against Herpes Simplex Virus 1 (HSV-1) infection in vitro. Methanol, ethanol and aqueous extracts of P.minus were obtained through evaporation under reduced pressure via rotary evaporator. Cytotoxicity testing of P.minus extracts was conducted on Vero cells using MTT assay. DPPH and Folin-Ciocalteu assay were used to evaluate its radical scavenging activity and phenolic content respectively. To better understand its medicinal properties, an in vitro treatment was carried out by means of time-of-addition tests; simultaneous treatment, pre-treatment and posttreatment. Infection with HSV-1 was performed at MOI of 1 and inoculated with methanol and ethanol extracts at its maximum non-toxic dose concentration of 37.50 µg/ml. In simultaneous and pre-treatment, both extract appeared to exert inadequate effect against HSV-1 infected cells (where cell viability recorded well below 60%). However, in post-treatment test, only aqueous extract showed desirable effect where cell viability is maximally retained. This is despite the fact that aqueous extract displayed the lowest radical scavenging activity (IC₅₀ = 146.58 µg/ml, with maximum inhibition at 8 mg/ml concentration), having the lowest phenolic content (61.68 ± 11.621 mg GAE/l at 8 mg/ml concentration), albeit a higher cytotoxicity $(IC_{50} = 408.03 \mu g/ml)$ towards Vero cells compared to methanolic extract. Hence, preliminary finding suggests aqueous *P.minus* extract has curative potential towards HSV-1 infected cells despite its subpar antioxidant activity. Indeed, further studies are required to make clear the exact curative effect of P.minus towards HSV-1 infection before a more conclusive experimental findings be made.

ABSTRAK

Penggunaan tumbuhan ubatan sebagai rawatan pencegahan dan rawatan adalah penting apabila rawatan yang lengkap belum ada. Oleh itu, tujuan kajian ini dijalankan adalah untuk menilai kesan ekstrak *Polygonum minus* terhadap jangkitan virus Herpes Simplex 1 (HSV-1) in vitro. Ekstrak metanol, etanol dan akueus P.minus diperolehi melalui penyejatan di bawah tekanan. Ujian sitotoksiti ekstrak P.minus telah dijalankan ke atas sel-sel Vero menggunakan ujikaji MTT. Manakala, ujikaji DPPH dan Folin-Ciocalteu digunakan untuk menilai aktiviti-memerangkapradikal dan kandugan fenolik. Bagi lebih memahami karakteristik perubatan *P.minus*, rawatan in vitro telah dijalankan melalui ujian berdasarkan masa-penambahan; rawatan serentak, pra-rawatan dan rawatan selepas. Jangkitan HSV-1 telah dilakukan pada MOI 1 dan didedahkan pada metanol dan etanol ekstrak pada kepekatan dos maksima tidak-toksik 37.50 µg/ml. Bagi ujian rawatan-serentak dan pra-rawatan, kedua-dua ekstrak menampakkan kesan perubatan yang tidak mencukupi terhadap sel dijangkiti HSV-1 (ini berikutan sel hidup dicatatkan di bawah 60%). Namun dalam ujian pasca rawatan, hanya ekstrak akueus menunjukkan kesan perubatan di mana sel-sel mampu hidup pada tahap maksima. Ini berikutan hakikat bahawa ekstrak akueus merekodkan aktiviti-memerangkap-radikal terendah (IC₅₀ = 146.58 µg/ml, dengan perencatan maksimum pada kepekatan 8 mg/ml), mempunyai kandungan fenolik yang paling rendah (61.68 ± 11.621 mg GAE/l pada kepekatan 8 mg/ml), walaupun mencatatkan sitotoksiti yang lebih tinggi (IC₅₀ = $408.03 \mu g/ml$) pada sel-sel Vero berbanding dengan ekstrak metanol. Dapatan awal menunjukkan ekstrak akueus *P.minus* mempunyai potensi penyembuhan terhadap sel-sel dijangkiti HSV-1 sungguhpun merekodkan aktiviti antioksida yang rendah. Sememangnya kajian lebih lanjut adalah perlu bagi penjelasan terperinci berkenaan kesan penyembuhan P.minus terhadap HSV-1 jangkitan sebelum kesimpulan yang lebih muktamad boleh dibuat.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE	
	ACKNOWLEDGEMENT	vi	
	ABSTRACT	vii	
	ABSTRAK		
	TABLE OF CONTENTS		
	LIST OF TABLES	xii	
	LIST OF FIGURES		
	LIST OF ABBREVIATIONS	xiv	
	LIST OF APPENDICES	xvi	
1.	INTRODUCTION		
	1.1 Background of Research	1	
	1.2 Problem Statement	3	
	1.3 Research Objectives	3	
	1.4 Scope of Research	3	
	1.5 Significance of Study	4	
2.	LITERATURE REVIEW		
	2.1 Herpes Simplex Virus 1	5	
	2.1.1 Infection	6	
	2.1.2 Treatment	7	
	2.1.2.1 Antiviral Drugs	7	
	2.1.2.2 Drug Resistance	8	
	2.2 Polygonum minus	9	
	2.2.1 Taxonomy	9	
	2.2.2 Plant Description	10	

		2.2.3	Phytoch	emicals	10
	2.3	Medic	inal Plan	ts As Antivirals	12
	2.4	2.4 MTT Assay			14
	2.5	DPPH	[Assay		15
	2.6	Folin-	Ciocalteu	Assay	17
3.	MA	TERIA	LS AND	METHODS	
	3.1	Mater	ials		18
		3.1.1	Chemic	als and Reagents	18
		3.1.2	Plant		19
		3.1.3	Virus		19
		3.1.4	Cell Cu	lture	19
	3.2	Metho	ods		20
		3.2.1	Overvie	w of Methodology	20
		3.2.2	Plant Ex	xtraction	20
			3.2.2.1	Methanol and Ethanol	21
				Extracts	
			3.2.2.2	Aqueous Extracts	21
		3.2.3	Antioxi	dant Evaluation	22
			3.2.3.1	DPPH Assay	22
			3.2.3.2	Total Phenolic Content	22
		3.2.4	Maximu	ım Non-Toxic Dose	23
		3.2.5	In vitro	Antiviral Treatment	24
		3.2.6	Virus ar	nd Cell Culture	25
			3.2.6.1	Propagation of HSV-1	25
			3.2.6.2	End-point Dilution Assay	26
			3.2.6.3	Maintenance of Vero Cells	27
		3.2.7	Statistic	al Analysis	27
4.	RES	SULTS	AND DIS	SCUSSION	
	4.1	Solvent Extraction			28
	4.2	Antio	xidant As	say	30
		4.2.1	DPPH F	Radical Scavenging Activity	30

		4.2.2 Total Phenolic Content	31
	4.3	Cytotoxic Test	34
	4.4	Virus Titer	36
	4.5	In vitro antiviral assay	37
5.	CONCLUSION		
	5.1	Conclusion	42
	5.2	Recommendation	43
REFERENCES			44-54
APPENDICES			
A-D			55-63

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Categories of clinically relevant human herpesviruses	5
2.2	Selected plants known to have antiviral activities	13
2.3	Factors affecting MTT results	14
4.1	Net percentage yield of <i>P.minus</i> extracts	29
4.2	Concentration of <i>P.minus</i> extracts for DPPH IC ₅₀	31
4.3	Total phenolic content of <i>P.minus</i> extracts at 500 μg/ml concentration	32
4.4	50% cytotoxicity of methanol and aqueous extracts	35

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	An electron cryo-tomographic visualization of a HSV virion	6
2.2	Phosphorylation of acyclovir to its active form	8
2.3	Polygonum minus leaves	10
2.4	Reduction of MTT	15
2.5	DPPH reaction to reduced form	16
3.1	Overview of methodology of project	20
4.1	Correlation of determination between phenolic content of <i>P.minus</i> extracts and its radical scavenging activity	33
4.2	Percentage of viable Vero cells following 72 hours inoculation with methanol and aqueous extracts	34
4.3	Image of CPE effect observed following Infection of Vero cells with HSV-1	37
4.4	Overview of treatment test employed	38
4.5	Percentage of cell viability in time-of-addition study	41

LIST OF ABBREVIATIONS

SYMBOLS DESCRIPTION

3D three dimensional

ACV acyclovir

ACV-TP acyclovir-triphosphate

ANOVA analysis of variance

CPE cytopathic effect

CO₂ carbon dioxide gas

DENV2 Dengue virus type 2

dGTP deoxyguanosine triphosphate

DMEM Dulbecco's Modified Eagle's Medium

DMSO Dimethyl sulfoxide

DNA Deoxyribonucleic acid

DPPH 2,2-diphenyl-1-picrylhydrazyl

FBS Fetal Bovine Serum

F-C Folin-Ciocalteu

F-D Folin-Dennis

GAE gallic acid equivalent

HCV Hepatitis C Virus

HIV Human immunodeficiency virus

HHV Human herpesvirus

HSV-1 Herpes simplex virus 1

IC₅₀ half maximal inhibitory concentration

IM Infection media

L15 Leibovitz 15 medium

MNTD maximum non-toxic dose

MOI multiplicity of infection

MTT 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide

RSA radical scavenging activity

SFE supercritical fluid extraction

SFM serum-free media

SPSS Statistical Package for Social Science

TCID₅₀ 50% Tissue culture infective dose

TPC Total Phenolic Content

PBS Phosphate Buffered Saline

pfu plaque forming unit

P.minus Polygonum minus

P/Strep Penicillin-Streptomycin

qRT-PCR quantitative Reverse Transcriptase-Polymerase Chain

Reaction

μl microliter

UAE ultrasound assisted extraction

VSV Vesicular Stomatitis Virus

LIST OF APPENDICES

APPENDIX	TITLE	PAGE	
A	ANOVA of DPPH	55	
	ANOVA of TPC	56	
	ANOVA of MTT	56	
	ANOVA of treatment test	57	
	Levene's test	57	
В	Percentage extract inhibition	58	
	Ascorbic acid standard curve	58	
	Gallic acid standard curve	59	
С	TCID ₅₀ determination based on	60	
	Reed-Muench method		
	MOI calculation	62	
D	Calculation for average yield of extracts	63	

CHAPTER 1

INTRODUCTION

1.1 Background of Research

Herpes simplex virus 1 (HSV-1) is one of nine herpesviruses known to occur in man. The virus is the causative agent of oral herpes infection, of which no cure has been found yet. HSV-1 has the ability to establish latent infection in which virions are reactivated and propagated under certain circumstances (Griffiths, 2014). As a result, a person infected with HSV-1 may experience recurrent symptoms such as lesions near the oral or mucosal regions of body (Whitley and Roizman, 2001; Fatahzadeh *et al.*, 2007). Lifelong infection is thus notable in HSV infected person.

A synthetic drug known as acyclovir is commonly prescribed to control outbreaks of herpes infection (Tan *et al.*, 2013). In brief, acyclovir works by blocking viral DNA synthesis while maintaining cell DNA synthesis. When inside an infected cell, acyclovir is phosphorylated to its active form by the virus' thymidine kinase (Piret and Boivin, 2014). Meanwhile cells not infected with the virus are not subjected to the effects of the drug, attributing to its specificity. However, acyclovir only attacks virus that are active and not on virus in latency (Bdel-Haq and Asmar, 2001) thus making regular drug administration necessary to reduce outbreaks. In addition, there have been cases on the occurrence of drug-resistant HSV strain and is

a major concern especially in severely immunocompromised patients (Griffiths, 2014).

For many years, natural sources have been subjected to screening for antiviral properties. Screening studies are regularly conducted to identify potential medicinal values in natural sources. In this approach, cell biology techniques are employed to identify compounds that may inhibit virus replication. Viruses are inoculated in cell cultures and compounds are subsequently added at IC₅₀ concentration. Evidence for viral inhibition may include cytopathic effect (CPE), cell viability and cell death, among others (Carter and Saunders, 2007). Following successful biological tests, crude extracts of natural source may then be subjected to compound fractionation for further evaluation (Balunas and Kinghorn, 2005). Screening compounds for antiviral properties is particularly useful when dealing with an undocumented natural source and is often the first step to drug discovery.

Polygonum minus is an herb plant of the Polygonaceae family. Due to its sweet, lemony and aromatic character, it is commonly used in traditional Southeast Asia's cooking as a flavor enhancer and interestingly, in folk medicine to treat digestive problems (Qader et al., 2012a). In Malaysia, it is known as 'kesum' in the Malay language (Burkill, 1996). The plant has been reported to exhibit potent antioxidant activity and antimicrobial properties (Faujan et al., 2006; Uyub et al., 2010). Previously, a screening study on 61 ethanolic extracts of medicinal plants in Malaysia found that P.minus was a potent antiviral agent against two types of viruses; HSV-1 and Vesicular Stomatitis Virus (VSV) (Ali et al., 1996). Meanwhile, in this study two other solvent types were included; methanol and aqueous.

1.2 Problem Statement

P.minus has been documented to possess many medicinal properties. However, antiviral properties of methanolic and aqueous *P.minus* extracts against HSV-1 have not been extensively elucidated. This study was conducted to evaluate its potential as an antiviral agent against herpes simplex virus 1 *in vitro*.

1.3 Research Objectives

The objectives are:

- 1. To evaluate antioxidant properties of *P.minus* extracts based on DPPH radical scavenging activity.
- 2. To determine cytotoxicity of *P.minus* extracts on Vero cells based on the MTT assay.
- 3. To evaluate inhibitory effect of *P.minus* extracts against HSV-1 using crystal violet assay based on a time-of-addition study.

1.4 Scope of Research

The first part of the study involved with the preparation of plant extracts using rotary evaporation method followed by determining the antioxidant properties of plant extracts based on DPPH radical scavenging activity and Folin-Ciocalteu assay. Cytotoxicity of plant extracts toward Vero cells were tested to obtain its MNTD concentration. The second part was the quantification of virus stocks using end-point dilution assay. Finally, an *in vitro* antiviral treatment was carried out to

investigate extracts' efficacy as an antiviral agent. The antiviral treatment was done by incubating extracts at its MNTD concentration before virus inoculation (pretreatment), after virus inoculation (post-treatment) and added together with virus (simultaneous-treatment). Cell viability was determined by crystal violet assay.

1.5 Significance of Study

The focus of this study is to evaluate the potential of a local herb in inhibiting viral infection *in vitro*. The study is hoped to help identify potential natural product-based treatment towards herpes infection and at large to spur a continuous interest on finding local plants that may exhibit medicinal properties against infectious diseases.

REFERENCES

- Ali, A.M., Mackeen, M.M., El-Sharkawy, S.H., Abdul Hamid J, Ismail, N.H., Ahmad, F., and Lajis, M.N. (1996). Antiviral and cytotoxic activities of some plants used in Malaysian indigenous medicine. *Pertanika Journal of Tropical Agriculture Science*. 19:129-136.
- Aspe, E., and Fernandez, K. (2011). The effect of different extraction techniques on extraction yield, total phenolic, and anti-radical capacity of extracts from *Pinus radiate* Bark. *Industrial Crops and Products*. 34(1):838-844.
- Baharum, S.N., Bunawan, H., Ghani, M.A., Mustapha, W.A.W., and Noor, N.M. (2010). Analysis of the chemical composition of the essential oil of *Polygonum minus* Huds using two-dimensional gas chromatography-time-of-flight mass spectrometry (GC-TOF MS). *Molecules*. 15:7006-7015.
- Balunas, M.J. and Kinghorn, D. (2005). Drug discovery from medicinal plants. *Life Sciences*, 78:431-441.
- Bdel-Haq, N. and Asmar, B. (2001). Anti-herpes viruses agents. *Indian Journal of Pediatrics*. 68:649-654.
- Bernas, T. and Dobrucki, J. (2002). Mitochondrial and nonmitochondrial reduction of MTT: interaction of MTT with TMRE, JC-1, and NAO mitochondrial fluorescent probes. *Cytometry*. 47(4):236-242.
- Blois, M.S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*. 181:1199-1200.

- Burkill, I. (1966). A dictionary of the economic products of the Malay Peninsula, vol. 2. Kuala Lumpur, *Ministry of Agriculture*.
- Bush, L.M., Talledo-Thais, K., Casal-Fernandez, A. and Perez, M.T. (2011). Resistant herpes simplex virus infection and HIV. *Lab Medicine*. 42(8):452-457.
- Carter, J. and Saunders, V. (2007). Virology: Principles and application. *Wiley and Sons Limited*.
- Chayavichitsilp, P., Buckwalter, J., Krakowski, A.C. and Friedlander, S.F. (2009). Herpes Simplex. *Pediatric in Reviews*. 30(4):119-129.
- Conway, J.F. and Homa, F.L. (2011). Nucleocapsid structure, assembly and DNA packaging of herpes simplex virus. In Weller S.K. (Ed.), *Alphaherpesviruses: Molecular Virology*. (pp. 175-193). Norfolk, UK: Caister Academic Press.
- Crozier, A., Jaganath, I.B., and Clifford, M.N. (2006). Phenols, polyphenols and tannins: An overview. In Crozier A., Clifford M.N., and Ashihara H. (Eds.), *Plant Secondary Metabolites: Occurrence, Structure and Role in the Human Diet.* (pp. 1-24). Singapore: Blackwell Publishing Limted.
- De Clerq, E., and Holy, A. (2005). Acyclic nucleoside phosphonates: A key class of antiviral drugs. *Nature Reviews Drug Discovery*. 4:928-940.
- Dent, M., Dragovic-Uzelac, V., Penic, M., Brncic, M., Bosiljkov, T., and Levaj, B. (2013). The effect of extraction solvents, temperature and time on the composition and mass fraction of polyphenols in dalmation wild sage (*Salvia officinalis* L.) extracts. *Food Technology and Biotechnology*. 51(1):84-91.
- Dohner, K., A. Wolfstein, U. Prank, C. Echeverri, D. Dujardin, R. Vallee, and B. Sodeik. (2002). Function of dynein and dynactin in herpes simplex virus capsid transport. *Molecular Biology of the Cell*. 13:2795-2809.

- Eklund, P. C., Langvik, O. K., Warna, J. P., Salmi, T. O., Willfor, S. M., and Sjoholm, R. E. (2005). Chemical studies on antioxidant mechanisms and free radical scavenging properties of lignans. *Organic and Bimolecular Chemistry*. 21:3336-3347.
- Elengoe, A., and Hamdan, S. (2014). Evaluation of hyperthermia effect on cell viability using crystal violet staining, LDH and trypan blue assays. *Advances in Environmental Biology*. 8(3):744-747.
- Elion, G.B. (1983). The biochemistry and mechanism of action of acyclovir. *Journal of Antimicrobial Chemotherapy*. 12:9-17.
- Everett, R. D. (2000). ICP0, a regulator of herpes simplex virus during lytic and latent infection. *Bioessays*. 22:761-770.
- Everette, J.D., Bryant, Q.M., Green, A.M., Abbey, Y.A., Wangila, G.W. and Walker, R.B. (2010). Thorough study of reactivity of various compound classes toward the Folin-Ciocalteu reagent. *Journal of Agricultural and Food Chemistry*. 58:8139-8144.
- Fatahzadeh, M., and Schwartz, R.A. (2007). Human herpes simplex infections: Epidemiology, pathogenesis, symptomatology, diagnosis, and management. *Journal of the American Academy of Dermatology*. 57:737-63.
- Faujan, N.H., Abdullah, N., Abdullah Sani, A. and Babji, A.M. (2007). Antioxidative activities of water extracts of some Malaysian herbs. *ASEAN Food Journal*. 14:61-68.
- Faujan, N.H., Noriham, A., Norrakiah, A.S., and Babji, A.S. (2009). Antioxidant activity of plants methanolic extracts containing phenolic compounds. *African Journal of Biotechnology*. 8(3):484-489.
- Field, H.J., and Hodge, R.A.V. (2013). Recent developments in anti-herpesvirus drugs. *British Medical Bulletin*. 106:213-249.

- Fotakis, G., and Timbrell, J.A. (2006). In vitro cytotoxicity assays: Comparison of LDH, neutral red, MTT and protein assay in hepatoma cell lines following exposure to cadmium chloride. *Toxicology Letters*. 160:171-177.
- Ghazali, M.A.M., Al-Naqeb, G., Selvarajan, K.K., Hasan, M.H. and Adam, A. (2014). Apoptosis induction by *Polygonum minus* is related to antioxidant capacity, alterations in expression of apoptotic-related genes, and S-phase cell cycle arrest in HepG2 cell line. *BioMed Research International*. http://dx.doi.org/10.1155/2014/539607.
- Grabmann, J. (2005). Terpenoids as plant antioxidants. In *Vitamins and Hormones*. 72:505-535. Elsevier Inc.
- Griffiths, P.D. (2014). Herpesviruses. Medicine. 42(1):34-38.
- Grunewald, K. and Cyrklaff, M. (2006). Structure of complex viruses and virus-infected cells by electron cryo-tomography. *Current Opinion in Microbiology*. 9:437-442.
- Gunatilaka, A.L. (2012). Plant natural products. In Natanya Civjan (editor). 2-29. *Natural Products in Chemical Biology*, 1st edition. Wiley and Sons, Inc.
- Gupta, R., Warren, T., and Wald, A. (2007). Genital herpes. *Lancet*. 370(9605):2127-2137.
- Hussin, A., Md Nor, N.S. and Ibrahim, N. (2013). Phenotypic and genotypic characterization of induced acyclovir-resistant clinical isolates of herpes simplex virus type 1. *Antiviral Research*. 100(2):306-313.
- Imelda, F., Faridah, D.N., and Kusumaningrum, H.D. (2014). Bacterial inhibition and cell leakage by extract of *Polygonum minus* Huds. leaves. *International Food Research Journal*. 21(2):553-560.

- Jaime, M.F.V., Redko, F., Muschietti, L.V., Campos, R.H., Martino, V.S. and Cavallaro, L.V. (2013). *In vitro* antiviral activity of plant extracts from Asteraceae medicinal plants. *Virology Journal*. 10:245-255.
- Johnston, C., Koelle, D.M., and Wald, A. (2014). Current status and prospects for development of an HSV vaccine. *Vaccine*. 32:1553-1560.
- Karim, B.Y. (1987) Kesom oil: a natural source of aliphatic aldehydes. *Perfumer Flavorist*. 12:27–30.
- Kitazato, K., Wang, Y., and Nobayashi, K. (2007). Viral infectious disease and natural products with antiviral activity. *Drug Discovery Therapy*. 1:14-22.
- Krawczyk, A., Arndt, M.A.E., Grosse-Hovest, L., Weichert, W., Giebel, B., Dittmer, U., Hengel, H., Jager, D., Schnewels, K.E., Eis-Hubinger, A.M., Roggendorf, M., and Krauss, J. (2013). Overcoming drug-resistant herpes simplex virus (HSV) infection by a humanized antibody. *Proceedings of the National Academy of Sciences of the United States of America*. 110(17):6760-6765.
- Kupcsik, L. (2011). Estimation of cell number based on metabolic activity: The MTT assay. In: Stoddart, M.J. (Ed.), *Mammalian cell viability* (pp. 13-19). New York, NY: Humana Press.
- Larder, B.A., Cheng, Y.-C. and Darby, G. (1983). Characterization of abnormal thymidine kinases induced by drug-resistant strains of herpes simplex virus type 1. *Journal of General Virology*. 64:523-532.
- Lattanzio, V., Kroon, P.A., Quideau, S., and Treutter, D. (2008). Plant phenolics secondary metabolites with diverse functions. In Dayf, F. and Lattanzio, V. (editors). *Recent Advances in Polyphenol Research*. 1:1-35. Blackwell Publishing Ltd.

- Lee, S.H., Tang, Y.Q., Rathkrishnan, A., Wang, S.M., Ong, K.C., Manikam, R., Payne, B.J., Jaganath, I.B., and Sekaran, S.D. (2013). Effects of cocktail of four local Malaysian medicinal plants (*Phyllanthus spp.*) against dengue virus 2. *BMC Complementary and Alternative Medicine*. 13:192.
- Maw, S.S., Mon, M.M., and Oo, Z.K. (2011). Study on antioxidant and antitumor activities of some herbal extracts. *World Academy of Science, Engineering and Technology*. 51:450-455.
- Molyneux, P. (2004). The use of the stable free radical diphenylpicryl-hydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin Journal of Science and Technology*. 26(2):211-219.
- Morfin, F. and Thouvenot, D. (2003). Herpes simplex virus resistance to antiviral drugs. *Journal of Clinical Virology*. 26:29-37.
- Navid, M.N., Laszczyk-Lauer, M.N., Reichling, J., and Schnittzler, P. (2014). Pentacyclic triterpenes in birch bark extract inhibit early step of herpes simplex virus type 1 replication. *Phytomedicine*. 21:1273-1280.
- O'Reilly, D.R., Miller L.K., and V.A. Luckow. (1994). Baculovirus Expression Vectors: A Laboratory Manual. Oxford University Press, New York.
- Penkert, R.R., and Kalejta, R.F. (2011). Tegument protein control of latent herpesvirus establishment and animation. *Herpesviridae*. 2:3.
- Piret, J. and Boivin, G., (2014). Antiviral drug resistance in herpesviruses other than cytomegalovirus. *Review in Medical Virology*. 24:186-218.
- Piret, J. and Boivin, G. (2011). Resistance of herpes simplex viruses to nucleoside analogues: Mechanisms, prevalence, and management. *Antimicrobial Agents and Chemotherapy*. 55(2):459-472.

- Poonia, P., Niazi, J., Chaudhary, G., and Kalia, A.N. (2011). In-vitro antioxidant potential of *Jasminum mesnyi* Hance (leaves) extracts. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 2(1):348-357.
- Pushpa, R., Nishant, R., Navin, K., and Pankaj, G. (2013). Antiviral potential of medicinal plants: An overview. *International Research Journal of Pharmacy*. 4(6):8-16.
- Qader, S.W., Abdulla, M.A., Chua, L.S., Najim, N., Zain, M.M., and Hamdan, S. (2011). Antioxidant, total phenolic content and cytotoxicity evaluation of selected Malaysian plants. *Molecules*. 16:3433-3443.
- Qader, S.W, Abdulla, M.A., Chua, L.S., and Hamdan, S. (2012a). Pharmacological mechanisms underlying gastroprotective activities of the fractions obtained from *Polygonum minus* in Sprague Dawley rats. *International Journal of Molecular Science*. 13:1481-1496.
- Qader SW, Abdulla MA, Chua LS, and Hamdan S. (2012b). Potential bioactive property of *Polygonum minus* Huds (kesum) review. *Scientific Research and Review*. 7(2):90-93.
- Renaud, S., and Lorgeril, M. de (1992). Wine, alcohol, platelets and the French paradox for coronary heart disease. *Lancet*. 339(8808):1523-1526.
- Reynolds, A. E., E. G. Wills, R. J. Roller, B. J. Ryckman, and J. D. Baines. (2002). Ultrastructural localization of the herpes simplex virus type 1 U_L31, U_L34, and U_S3 proteins suggests specific roles in primary envelopment and egress of nucleocapsids. *Journal of Virology*. 76:8939-8952.
- Sagar, B.K. and Singh, R.P. (2011). Genesis and development of DPPH method of antioxidant assay. *Journal of Food Science and Technology*. 48(4):412-422.

- Sanchez-Rangel, J.C., Benavides, J., Heredia, B., Cisneros-Zevallos, L. and Jacobo-Velazquez, D.A. (2013). The Folin-Ciocalteu assay revisited: improvement of its specificity for total phenolic content determination. *Analytical Methods*. 5:5990-5999.
- Singh, S., Singh, R.P. (2008) In vitro methods of assay of antioxidants: An overview. *Food Reviews International.* 24(4):392-415.
- Singleton, V. L., and Rossi, J. A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*. 16:144-153.
- Sodeik, B., M. W. Ebersold, and A. Helenius. (1997). Microtubule-mediated transport of incoming herpes simplex virus 1 capsids to the nucleus. *Journal of Cell Biology*. 136:1007-1021.
- Suedee, A., Tewtrakul, S. and Panichayupakaranant, P. (2014). Anti-HIV-1 integrase activity of *Mimusops elengi* leaf extracts. *Pharmaceutical Biology*. 52(1):58-61.
- Sultana, B., Anwar, F., and Ashraf, M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*. 14:2167-2180.
- Taddeo, B., and B. Roizman. (2006). The virion host shutoff protein (U_L41) of herpes simplex virus 1 is an endoribonuclease with a substrate specificity similar to that of RNase A. *Journal of Virology*. 80:9341-9345.
- Tan, W.C., Jaganath, I.B., Manikam, R., and Sekaran, S.D. (2013). Evaluation of antiviral activities of four local Malaysian Phyllanthus species against Herpes Simplex Viruses and possible antiviral target. *International Journal of Medical Sciences*. 10:1817-1829.

- Tang, L.I.C., Ling, A.P.K., Koh, R.Y., Chye, S.M., and Voon, K.G.L. (2012).
 Screening of anti-dengue activity in methanolic extracts of medicinal plants.
 BMC Complementary and Alternative Medicine. 12:3.
- Teh, S.-S., Bekhit, A.E.-D., and Birch, J. (2014). Antioxidative polyphenols from defatted oilseed cakes: Effect of solvents. *Antioxidants*. 3:67-80.
- Urones, J.G., Marcos, I., Perez, B., and Barcala, P. (1990). Flavonoids from *Polygonum minus. Phytochemistry*. 29 (11):3687-3689.
- Uyub, A.M., Nwachukwu, I.N., Azlan, A.A. and Fariza, S.S. (2010). In-vitro antibacterial activity and cytotoxicity of selected medicinal plant extracts from Penang Island Malaysia on metronidazole-resistant-Helicobacter pylori and some pathogenic bacteria. *Ethnobotany Research and Applications*. 8:95-106.
- Vimala, S., Rohana, S., Rashih, A.A. and Juliza, M. (2011). Antioxidant evaluation in Malaysian medicinal plant: *Persicaria minor* (Huds.) leaf. *Science Journal of Medicine and Clinical Trials*. 1:9-16.
- Wahyuni, T.S., Tumewu, L., Permanasari, A.A., Apriani, E., Adianti, M., Rahman, A., Widyawaruyanti, A., Lusida, M.I., Fuad, A., Soetjipto, N., Fuchino, H., Kawahara, N., Shoji, I., Aoki, C., and Hotta, H. (2013). Antiviral activities of Indonesian medicinal plants in the East Java region against hepatitis C virus. Virology Journal. 10:259.
- Wang, Y., Wang, Q., Qinchang, Z., Zhou, R., Liu, Jinsong, and Peng, T. (2011). Identification and characterization of acyclovir-resistant clinical HSV-1 isolates from children. *Journal of Clinical Virology*. 52:107-112.
- Waterman, P.G. (1992). Roles for secondary metabolites in plants. In Chadwick D.J. and Whelan J. (editors). 255-275. Novartis Foundation Symposium. Ciba Foundation.

- Weyermann, J., Lochmann, D., and Zimmer, A. (2005). A practical note on the use of cytotoxicity assays. *International Journal of Pharmaceutics*. 288:369-376.
- Whitley, R.J., and Roizman, B. (2001). Herpes simplex virus infections. *Lancet*. 357(9267):1513-1518.
- Wiart, C., Kumar, K., Yusof, M.Y., Hamimah, H., Fauzi, Z.M. and Sulaiman, M. (2005). Antiviral properties of Ent-labdenediterpenes of Andrographis paniculata Nees, inhibitors of herpes simplex virus 1. *Phytotherapy Research*. 19:1069-1070.
- Wu, S.F., Lin, C.K., Chuang, Y.S., Chang, F.R., Tseng, C.K., Wu, Y.C. and Lee, J.C.
 (2012). Anti-hepatitis C virus activity of 3-hydroxy caruilignan C from Swieteniamacrophylla stems. Journal of Viral Hepatitis. 19:364-370.
- Yaacob, K.B. (1987). Kesom oil: A natural source of aliphatic aldehydes. *Perfumer Flavorist*. 12:27-30.
- Yang, C.M., Cheng, H.Y., Lin, T.C., Chiang, L.C., and Lin, C.C., (2007). The in vitro activity of geraniin and 1,3,4,6-tetra-O-galloyl-beta-d-glucose isolated from *Phyllanthus urinaria* against herpes simplex virus type 1 and type 2 infection. *Journal of Ethnopharmacology*. 110 (3):555-558.
- Zhao, H., Dong, J., Lu, J., Chen, J., Li, Y., Shan, L., Lin, Y., Fan, W. and Gu, G. (2006). Effects of extraction solvent mixtures on antioxidant evaluation and their extraction capacity and selectivity for free phenolic compounds in barley (*Hordeum vulgare* L.) *Journal of Agricultural and Food Chemistry*. 54:7277-7286.
- Zhong, M.-G., Xiang, Y.-F., Qiu, X.-X., Liu, Z., Kitazato, K. and Wang, Y.-F. (2013). Natural products as a source of anti-herpes simplex virus agents. *Royal Society of Chemistry*. 3:313-328.

Zhu, L., Zhang, Y., and Lu, J. (2012). Phenolic contents and compositions in skins of red wine grape cultivars among various genetic backgrounds and originations. *International Journal of Molecular Sciences*. 13:3492-3510.