AMPLIFICATION OF PARTIAL RICE FLORIGEN FROM MALAYSIAN UPLAND RICE CULTIVAR HITAM AND WAI

ABDULRAHMAN MAHMOUD DOGARA

UNIVERSITI TEKNOLOGI MALAYSIA

AMPLIFICATION OF PARTIAL RICE FLORIGEN FROM MALAYSIAN UPLAND RICE CULTIVAR HITAM AND WAI

ABDULRAHMAN MAHMOUD DOGARA

A dissertation submitted in partial fulfillment of the requirements for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

JANUARY 2015

•

This research work has been dedicated to my late mother Hajiya Ramatu. May ALLAH reward her with Aljannah Firdausi and her soul rest in peace.Ameen.

ACKNOWLEDGEMENT

First and foremost, I give special thanks and glory to the Almighty ALLAH for giving me the grace, wisdom and good health to complete this research work. I am also indebted to my supervisor Dr Zaidah Rahmat who guided and helped greatly towards the completion of this work, may ALLAH reward her abundantly. Ameen

My special appreciation goes to my father Alhaji Mahmoud Dogara and my late Mother Hajiya Ramatu may Allah reward them with Aljannah Firdausi. Ameen

Also to my loving wife, Razeeqa Shafiu for her continuing source of help and encouragement, her unalloyed love, support and understanding, her endurance day and night, always give me the confidence to work very hard while I was in struggle for existence and survival of the fittest.

Finally to all members of Plant Biotechnology Laboratory more especially Shahkila Mohammed Arif for the encouragements all the times.

ABSTRACT

Rice is one of the global sources of food where it is grown in almost 148 million hectares annually with about 11% of the world planted land. Wetland rice is commonly grown and required extra land for good irrigation system. To compensate for land scarcity, upland rice which requires less water consumption is one alternative. Regardless of wetland or upland, rice is still being planted once a year all over the world although it could be harvested faster during short day condition. To meet global rice demand, there is a need to understand the effect of photoperiod on rice florigen and flowering to produce better yield. The aim of the study was to amplify rice florigen from Malaysian upland rice to improve its potential by amplifying Hd3a and RFT1 genes. Following amplification and verification via sequencing, cloning of the purified PCR product and construction of phylogenetic tree was carried out. Malaysian upland rice is grown in long day condition hence only partial RFT1 gene was successfully isolated. The partial gene sequence was aligned with 10 other RFT1 gene belonging to Indica and Japonica varieties and showed that Malaysian upland rice cultivars Hitam and Wai evolved from Japponica cultivars. Findings from this study suggested high similarity of *RFT1* gene between various cultivars and further research on this gene is hoped to provide better understanding of flowering time of Malaysian upland rice for crop improvement.

ABSTRAK

Beras adalah salah satu sumber makanan global di mana setiap tahun ia ditanam di hampir 148 juta hektar iaitu kira-kira 11% daripada tanah dunia. Padi sawah adalah jenis yang biasa ditanam dan memerlukan tanah tambahan untuk sistem pengairan yang baik. Untuk mengimbangi kekurangan tanah, padi tanah tinggi yang tidak memerlukan penggunaan air yang banyak adalah satu alternatif. Tidak kira kawasan paya atau tanah tinggi, padi masih ditanam sekali setahun di seluruh dunia walaupun ia boleh dituai lebih cepat dalam keadaan hari siang yang pendek. Bagi memenuhi permintaan beras global, terdapat keperluan untuk memahami kesan fotokala pada florigen padi untuk menghasilkan hasil yang lebih baik. Tujuan kajian ini adalah untuk mengamplifikasi florigen padi dari padi tanah tinggi Malaysia untuk meningkatkan potensinya dengan amplifikasi gen Hd3a dan RFT1. Setelah proses amplifikasi and pengesahan melalui penjujukan, pengklonan PCR produk tulen dan pembinaan pokok filogenetik telah dijalankan. Padi tanah tinggi Malaysia ditanam dalam keadaan hari siang yang panjang maka hanya sebahagian gen RFT1 berjaya dipencilkan. Urutan jujukan sejajar dengan 10 gen *RFT1* lain daripada variasi Indica dan Japonica menunjukkan bahawa kultivar Hitam dan Wai berasal daripada padi tanah tinggi Malaysia mempunyai evolusi sejajar dengan variasi Japponica. Hasil daripada kajian ini mennunjukkan persamaan yang tinggi antara gen *RFT1* daripada pelbagai kultivar dan penyelidikan lanjut mengenai gen ini diharapkan dapat memberikan pemahaman yang lebih baik daripada masa berbunga padi tanah tinggi Malaysia untuk penambahbaikan tanaman.

TABLE OF CONTENTS

| CHAPTER | TITLE | PAGE |
|---------|---------------------------|------|
| | DECLARATION | ii |
| | DEDICATION | iii |
| | ACKNOWLEDGEMENT | iv |
| | ABSTRACT | V |
| | ABSTRAK | vi |
| | TABLE OF CONTENTS | vii |
| | LIST OF TABLES | X |
| | LIST OF FIGURES | xi |
| | LIST OF ABBREVATIONS | xiii |
| | LIST OF APPENDICES | XV |
| | | |
| 1 | INTRODUCTION | 1 |
| | 1.1 Study background | 1 |
| | 1.2 Problem Statements | 3 |
| | 1.3 Objectives | 4 |
| | 1.4 Scope of Study | 4 |
| | 1.5 Significance of Study | 4 |
| 2 | LITERATURE REVIEW | 6 |

| | EKATUKE KEVIEW | 0 |
|-----|----------------|---|
| 2.1 | Oryza sativa | 6 |

| 2.2 | Rice Florigen | 9 |
|-----|--|----|
| 2.3 | Hd3a gene | 10 |
| 2.4 | RFT1 gene | 11 |
| 2.5 | Mechanism of Hd3a and RFT1 as rice Florigen | 12 |
| 2.6 | Plants DNA Extraction | 13 |
| 2.7 | Nucleic Acid Quantification via Spectrophotometer | 13 |
| 2.8 | Polymerase Chain Reaction | 14 |
| 2.9 | Plants Transformation | 14 |
| 2.1 | Phylogenetic Analysis | 16 |
| | | |
| MF | CTHODOLOGY | 17 |
| 3.1 | Sample Collection | 17 |
| 3.2 | DNA Extraction | 17 |
| 3.3 | Polymerase Chain Reaction | 18 |
| | 3.3.1 Amplification of <i>Hd3a</i> gene | 18 |
| | 3.3.2 Amplification of RFT1 gene | 19 |
| 3.4 | Nucleic Acid Quantitative and Qualitative Analysis | 20 |
| | 3.4.1 Quantification via Spectrophotometer | 20 |
| | 3.4.2 Agarose Gel Electrophoresis | 20 |
| 3.5 | Purification of PCR Product | 21 |
| 3.6 | Blast and Result Analysis | 22 |
| 3.7 | Cloning of the Purified PCR Product into pGEMT | |
| | vector | 22 |
| 3.8 | Preparation of Chemical Competent Cells | 22 |
| 3.9 | Preparation of Electro Competence Cell | 23 |
| 3.1 | OTransformation into Chemical Competence Cells | 23 |
| 3.1 | 1 Transformation into Electro Competence Cells | 24 |

3

| | 3.12 Purification of DNA Plasmid with Centrifugation | 24 |
|---|--|----|
| | 3.13 Phylogenetic Tree Construction | 25 |
| 4 | RESULTS AND DISCUSSION | 27 |
| | 4.1 Genomic DNA Extraction | 27 |
| | 4.2 PCR Results | 28 |
| | 4.2.1 Amplification using Hd3a primers | 29 |
| | 4.2.2 Amplification using RFT1 primers | 29 |
| | 4.3 Cloning of the PCR Product into pGEMT vector | 31 |
| | 4.4 Phylogenetic Analysis | 32 |
| | | |
| 5 | CONCLUSION AND FUTURE WORK | 36 |
| | 5.1 Conclusion | 36 |
| | 5.2 Future Work | 36 |
| | Making Luria Bertani Agar | 50 |
| | Making Luria Broth | 50 |
| | REFERENCES | 37 |
| | APPENDIX A-C | 46 |

LIST OF TABLES

| TABLE NO. | TITLE | PAGE |
|-----------|--|------|
| 2.1 | Photoperiod Respond in rice Plants (Adapted from | |
| | Komiya <i>et al.</i> , 2008) | 9 |
| 3.1 | Sequence of the primer used for amplification of | |
| | Hd3a gene | 19 |
| 3.2 | Sequence of the primer for the amplification of | |
| | RFT1 gene | 20 |

LIST OF FIGURES

FIGURE NO.

TITLE

PAGE

| 2.1 | Evolutionary pathway of two cultivated rice species | |
|-----|--|----|
| | (Adapted from Khush, 1997) | 7 |
| 2.2 | Different Ecologies showing world Rice production. | |
| | (Adapted from Khush, 1997) | 8 |
| 2.3 | Model for <i>Hd3a</i> and <i>RFT1</i> activation in rice showing | |
| | Hd3a and RFT1 are activated in SD condition while | |
| | only RFT1 is activated in LD condition (Adapted by | |
| | Komiya <i>et al.</i> , 2009) | 12 |
| 2.4 | Schematic of Bacterial Transformation (Adapted by | |
| | Tyagi and Mohanty, 2000). | 15 |
| 3.1 | Flow Chat of the Methodology | 26 |
| 4.1 | The Extracted genomic DNA from Hitam and Wai | |
| | from 8, 9, 11 and 14 weeks: Bench Top 1kb DNA | |
| | ladder, H: Hitam, W:Wai | 28 |
| 4.2 | PCR Results of Hitam and Wai from 8 to 11 weeks | |
| | and M: Bench Top 100bp DNA ladder, H: Hitam, | |
| | W:Wai | 28 |
| 4.3 | Amplified <i>Hd3a</i> gene promoter from Hitam and Wai | |
| | both at 11 and 14 weeks and M: Bench Top | |
| | 100bp DNA ladder, H: Hitam, W:Wai | 29 |
| 4.4 | Amplified <i>RFT1</i> gene from Hitam and Wai at 11 and | |
| | 14 weeks and M: Bench Top 100bp DNA ladder, H: | |
| | Hitam, W:Wai | 30 |
| | | |

| 4.5 | Agar plate showing Transformed colony of the | |
|-----|---|----|
| | cloned RFT1 gene | 31 |
| 4.6 | Colonies from cloning after PCR with RFT1 primers | |
| | and M: Bench Top 100bp DNA ladder | 32 |
| 4.7 | Multiple sequences Alignment from 10 RFT1 genes | |
| | from Asian rice of Japonica and Indica variety with | |
| | partial RFT1 gene from Malaysian Upland Rice | |
| | cultivars Hitam and Wai | 33 |
| 4.8 | Phylogenetic tree constructed from nucleotides of | |
| | various RFT1 sequences demonstrate evolutionary | |
| | relationship | |
| | between Malaysian upland rice cultivars Wai and | |
| | Hitam with other Japonica and Indica varieties | 34 |

LIST OF ABBREVATIONS

| RFT1 | - | Rice Flowering Time 1 |
|------|---|---|
| Hd3a | - | Heading date 3a |
| Eh1 | - | Early Heading Date 1 |
| Hd1 | - | Heading Date 1 |
| OsGI | - | Oryza sativa gigantae |
| PCR | - | Polymerase Chain Reaction |
| gDNA | - | Genomic Deoxy Ribonucleic Acid |
| DAS | - | Days After Sowing |
| FT | - | Flowering Time |
| LB | - | Luria Broth |
| LBA | - | Luria Broth Agar |
| μL | - | Microliter |
| mL | - | Mililiter |
| bp | - | Base Pairs |
| QTL | - | Quantitative Trait Loci |
| SAM | - | Shot Apical Meristem |
| UN | - | United Nation |
| FAO | - | Food Agricultural Organization |
| NCB1 | - | National Centre for Biotechnology Information |
| Μ | - | Marker |
| Mg | - | Miligram |

| Kb | - | Kilobase |
|-------|---|-------------------|
| nm | - | Nanometer |
| et al | - | And other persons |

LIST OF APPENDICES

| APPENDIX | TITLE | PAGE | |
|----------|--|------|--|
| Α | Quantification via Spectrophotometer | 46 | |
| В | Graphical Presentation of BLAST Search | 48 | |
| С | LB recipe | 50 | |

CHAPTER 1

INTRODUCTION

1.1 Study background

Plants are generally found in different environment. They have adapted to many external factors with an impact on growth, development and most importantly the ability of the plant to produce food and seed (Brambilla and Fornara, 2013).

Rice as a cereal plant is one of the source of food in which more than one third of the globe population depend on it. It is grown in almost 148 million hectares land annually which is about 11% of the land used in the world. About 90% rice produced in the world is consumed by Asian that has about 60% of the world population (Khush, 1997). More than 3 billion individuals of the world use rice as their essential food which provide about 80% of the calories needed (Sohrabi *et al.*, 2012).

Rice alone provide more than one fourth of the total calories consumed in the world. It was reported by Food Agricultural Organisation (FAO) (1999) that rice remains the most important staple food in more than 15 countries across the Asia together with the pacific region. It also predominate 10 countries found between the

Caribbean, Latin America, seven countries in North America and Sub Saharan Africa respectively. Rice is a member of family *poaceae* which is made of 2 cultivars. These two species are mainly *Oryza sativa* also known as Asia cultivated rice and is found worldwide while the *Oryza glaberrima* which is found within the west African countries only, which is either planted as wetland or upland.

Wetland rice is a type of rice that is planted in waterlogged area which is difficult to handle. Upland rice is found in most part of the world except Antarctica but is found mostly in Asia, Latin America and Africa. This is the type of rice that is cultivated in dry soil with inadequate water and the yield is very low due to environmental stress (Sohrabi *et al.*, 2012).

Out of 150 million hectares of land utilised for rice cultivation in Asia, only a small proportion of it is upland rice which is around 11% (Sohrabi *et al.*, 2012). Most of the upland rice in Sabah and Sarawak is mainly for home consumption. Due to the environmental stresses, the upland rice does not produce high yield. Therefore improving its yield production is necessary so as to meet the food security in the world. With the recent technology in the area of plant molecular studies, upland rice flowering associated closely with the yield can be increased. Flowering time can be described as heading date in cereal plants, which is the agronomical determinant for acclimatization to precise plantation area and growing seasons for existing species of planted rice.

Photoperiod is the most important factor that helps to provide the plants with suitable signals for flowering. Regulation of flowering is among important processes of plants that relate to its fruit and seed formation. Flowering time is regulated by many factors in environment such like light during the day, water and temperature supply. Flowering is classified into 3 classes according their response to photoperiod. These include long-day plant that produce flowering within the long-day (LD) condition, short day plants that promote flower under short day (SD) condition lastly neutral day plant which operates independently of photoperiod as shown in Table 2.1. Subsequent progress and recent advacement in the understanding of

photoperiodic flowering in *Oryza sativa* helps to reveal some conserved pathways which promotes flowering under short day condition and at the same time suppressed flowering under long day condition.

Genetic factors in addition to environmental factors regulate the time of flowering in plant. Genetic mechanism about photoperiodic response for the long LDP has been studied through the *Arabidopsis* while the short day plants (SDPs) like rice show a remarkable similarity (Yano *et al.*, 2001). Today, the world has depended on rice as a major source of food. Many studies have been carried out to improve its production using conventional approach. Through molecular studies, improvement of rice yield would be achieved. Studies to give background information on flowering genes of Malaysian upland rice could help improve its low yield.

1.2 Problem Statements

The consumption of rice is increasing at the speed of 1.5% per annum while its production presently increase at the speed of 1% per annum only (Jeon *et al.*, 2011). According to the United Nation (UN) estimation, the world population will be 8 billion by 2025 while the present population is 6.7 billion, therefore there is need for the production to increase from 445 million tonnes at present to 486 million tones by 2025 (Jeon *et al.*, 2011). Food Agricultural Organisation estimate by 2050 the world rice requirement will be 524 million tonnes which need yearly increase of 2 million tonnes from the present level of production (Jeon *et al.*, 2011). Upland rice need to be improved so as to utilise the large abundance of land in Asia. To meet with the challenge of producing more rice from the existing land resources, therefore upland rice there is a need to further understand the rice florigen hence this study could serve as flatform for increasing rice yield and seed production.

1.3 Objectives

1 To optimize PCR Amplification of *Hd3a* and *RFT1* genes using specific primers from Malaysian Upland Rice cultivars Wai and Hitam

2 To clone the ampilified PCR product into intermediate vector

3 To compare the amplified *RFT1* sequence with other known *RFT1* sequences from various *O. sativa* cultivars for creation of phylogenetic tree.

1.4 Scope of Study

Upland rice from the local cultivars were grown up until 14 weeks of age and the leaves harvested at different stages for genomic DNA extraction followed by PCR with specific primers. Amplified PCR product of correct band size was purified and sent for sequencing which was verified and later cloned into *E.coli*. Sequence obtained was aligned with selected *RFT1* gene sequence from NCBI to create phylogenetic tree using the amplified partial sequence.

1.5 Significance of Study

Due to the climate of Malaysia categorized under neutral day or long day condition, rice can only be planted once in a year as it took up until 7 months to obtain the yield. The success in optimizing *Hd3a* and *RFT1* genes from genomic DNA serve as a very important platform on how these genes regulate flowering in

upland rice of Malaysia. To date no such information has been reported on Malaysian upland rice. This serve as a very important platform to see how these genes can be manipulated to adapt to upland environment which in turn will help increase upland rice production in Malaysia. In the long run, manipulation of the gene will aid upland rice to be planted twice in a year hence, increasing its production.

REFERENCES

Babadi, A. A. & Salleh, F. M. (2013). High Quality cDNA synthesis and Amplification of Chalcone Synthase Gene (CHS) from Justicia gendarussa Burm. F. *Jurnal Teknologi*, 64,837-858

Bernier, J., Atlin, G. N., Serraj, R., Kumar, A. & Spaner, D. (2008). Breeding upland rice for drought resistance. *Journal of the Science of Food and Agriculture*, 88, 927-939.

Brack, A. Foreword, (2002). Upland Rice Breeding, *Journal of the Science of Food and Agriculture* 54,751-782.

Brambilla, V. & Fornara, F. (2013). Molecular Control of Flowering in Response to Day Length in Rice. *Journal of integrative plant biology*.55(5): 410-418

Bustin, S., Benes, V., Nolan, T. & Pfaffl, M. (2005). Quantitative real-time RT-PCR-a perspective. *Journal of molecular endocrinology*, 34, 597-601.

Chang, S., Puryear, J. & Cairney, J. (1993). A simple and efficient method for isolating RNA from pine trees. *Plant molecular biology reporter*, 11, 113-116.

Chen, D.-H. & Ronald, P. (1999). A rapid DNA minipreparation method suitable for AFLP and other PCR applications. *Plant Molecular Biology Reporter*, 17, 53-57.

Dai, X., Ding, Y., Tan, L., FU, Y., Liu, F., Zhu, Z., Sun, X., Sun, X., Gu, P. & Cai, H. (2012). LHD1, an Allele of DTH8/Ghd8, Controls Late Heading Date in Common Wild Rice (*Oryza rufipogon*) F. *Journal of Integrative Plant Biology*, 54, 790-799.

Das, M., Harvey, I., CHU, L. L., Sinha, M. & Pelletier, J. (2001). Full-length cDNAs: more than just reaching the ends. *Physiological genomics*, 6, 57-80.

Dingkuhn, M., Jones, M. P., Johnson, D. E. & Sow, A. (1998). Growth and yield potential of Oryza sativa and O. glaberrima upland rice cultivars and their interspecific progenies. *Field Crops Research*, 57, 57-69.

Ebana, K., Shibaya, T., Wu, J., Matsubara, K., Kanamori, H., Yamane, H., Yamanouchi, U., Mizubayashi, T., Kono, I. & Shomura, A. (2011). Uncovering of major genetic factors generating naturally occurring variation in heading date among Asian rice cultivars. *Theoretical and applied genetics*, 122, 1199-1210.

Edwards, K., Johnstone, C. & Thompson, C. (1991). A simple and rapid method for the preparation of plant genomic DNA for PCR analysis. *Nucleic Acids Research*, 19, 1349.

Fageria, N. & Baligar, V. (2003). Upland rice and allelopathy. *Communications in soil science and plant analysis*, 34, 1311-1329.

Fouladvand, A. (2013). *Optimization of a Rapid DNA Extraction Protocol in Rice*. Nucleic Acid Research, 19, 1350-1353.

Freiberg, C., Fellay, R., Bairoch, A., Broughton, W. J., Rosenthal, A. & Perret, X. (1997). Molecular basis of symbiosis between Rhizobium and legumes. *Nature*, 387, 394-401.

Fulton, T. M., Chunwongse, J. & Tanksley, S. D. (1995). Microprep protocol for extraction of DNA from tomato and other herbaceous plants. *Plant Molecular Biology Reporter*, 13, 207-209.

Goff, S. A., Ricke, D., Lan, T.-H., Presting, G., Wang, R., Dunn, M., Glazebrook, J., Sessions, A., Oeller, P. & Varma, H. (2002). A draft sequence of the rice genome (*Oryza sativa L. ssp. japonica*). *Science*, 296, 92-100.

Hanahan, D. (1983). Studies on transformation of *Escherichia coli* with plasmids. *Journal of molecular biology*, 166, 557-580.

Hansen, G. & Wright, M. S. (1999). Recent advances in the transformation of plants. *Trends in plant science*, 4, 226-231.

HARBERS, M. 2008. The current status of cDNA cloning. *Genomics*, 91, 232-242.

Henson, J. M. & French, R. C. (1993). The polymerase chain reaction and plant disease diagnosis. *Papers in Plant Pathology*, 2,557-561

Hiei, Y., Ohta, S., Komari, T. & Kumashiro, T. (1994). Efficient transformation of rice (*Oryza sativa L.*) mediated by Agrobacterium and sequence analysis of the boundaries of the T-DNA. *The Plant Journal*, 6, 271-282.

Hiratsuka, J., Shimada, H., Whittier, R., Ishibashi, T., Sakamoto, M., Mori, M., Kondo, C., Honji, Y., Sun, C.-R. & Meng, B.-Y. (1989). The complete sequence of the rice (*Oryza sativa*) chloroplast genome: intermolecular recombination between distinct tRNA genes accounts for a major plastid DNA inversion during the evolution of the cereals. *Molecular and General Genetics MGG*, 217, 185-194.

Ishikawa, R., Aoki, M., Kurotani, K.-I., Yokoi, S., Shinomura, T., Takano, M. & Shimamoto, K. (2011). Phytochrome B regulates Heading date 1 (Hd1)mediated expression of rice florigen *Hd3a* and critical day length in rice. *Molecular Genetics and Genomics*, 285, 461-470.

Izawa, T., Takahashi, Y. & Yano, M. (2003). Comparative biology comes into bloom: genomic and genetic comparison of flowering pathways in rice and *Arabidopsis Current opinion in plant biology*, 6, 113-120.

Jain, M., Nijhawan, A., Tyagi, A. K. & Khurana, J. P. (2006). Validation of housekeeping genes as internal control for studying gene expression in rice by quantitative real-time PCR. *Biochemical and biophysical research communications*, 345, 646-651.

Jeon, J.-S., Jung, K.-H., Kim, H.-B., Suh, J.-P. & Khush, G. S. (2011). Genetic and molecular insights into the enhancement of rice yield potential. *Journal of Plant Biology*, 54, 1-9. Khush, G. S. (1997). Origin, dispersal, cultivation and variation of rice. *Oryza: From Molecule to Plant.* Springer, 46(1), 14-22

Kojima, S., Takahashi, Y., Kobayashi, Y., Monna, L., Sasaki, T., Araki, T. & Yano, M. (2002). Hd3a, a rice ortholog of the *Arabidopsis* FT gene, promotes transition to flowering downstream of *Hd1* under short-day conditions. *Plant and Cell Physiology*, 43, 1096-1105.

Komiya, R., Ikegami, A., Tamaki, S., Yokoi, S. & Shimamoto, K. (2008). *Hd3a* and *RFT1* are essential for flowering in rice. *Development*, 135, 767-774.

Komiya, R., Yokoi, S. & Shimamoto, K. (2009). A gene network for longday flowering activates *RFT1* encoding a mobile flowering signal in rice. *Development*, 136, 3443-3450.

Lee, S., Jeon, J.-S., Jung, K.-H. & An, G. (1999). Binary vectors for efficient transformation of rice. *Journal of Plant Biology*, 42, 310-316.

Monna, L., Lin, H., Kojima, S., Sasaki, T. & Yano, M. (2002). Genetic dissection of a genomic region for a quantitative trait locus, Hd3, into two loci, Hd3a and Hd3b, controlling heading date in rice. *Theoretical and Applied Genetics*, 104, 772-778.

Nagadhara, D., Ramesh, S., Pasalu, I., Rao, Y. K., Sarma, N., Reddy, V. & Rao, K. (2004). Transgenic rice plants expressing the snowdrop lectin gene (gna) exhibit high-level resistance to the whitebacked planthopper (Sogatella furcifera). *Theoretical and Applied Genetics*, 109, 1399-1405.

Ogiso-tanaka, E., Matsubara, K., Yamamoto, S.-I., Nonoue, Y., WU, J., Fujisawa, H., Ishikubo, H., Tanaka, T., Ando, T. & Matsumoto, T. (2013). Natural variation of the *RICE FLOWERING LOCUS T 1* contributes to flowering time divergence in rice. *PloS one*, 8, e75959.

Okayama, H. & Berg, P. (1982). High-efficiency cloning of full-length cDNA. *Molecular and cellular biology*, 2, 161-170.

Park, S. J., Kim, S. L., Lee, S., Je, B. I., Piao, H. L., Park, S. H., Kim, C. M., Ryu, C. H., Park, S. H. & Xuan, Y. H. (2008). Rice Indeterminate 1 (OsId1) is necessary for the expression of *Ehd1* (*Early heading date 1*) regardless of photoperiod. *The Plant Journal*, 56, 1018-1029.

Porebski, S., Bailey, L. G. & Baum, B. R. (1997). Modification of a CTAB DNA extraction protocol for plants containing high polysaccharide and polyphenol components. *Plant molecular biology reporter*, 15, 8-15.

Roongsattham, P., Pongtongkam, P., Thongpan, A., Kaveeta, L., Harinasut, P. & Peyachoknagul, S. (2006). *Hd1*, *Hd3a*, and *Hd6* genes: possible DNA methylation roles in photoperiod sensitive gene regulation of rice KDML 105 (Oryza sativa L.). *Kasetsart J.(Nat. Sci.)*, 40, 462-471.

Saito, K., Linquist, B., Keobualapha, B., Phanthaboon, K., Shiraiwa, T. & Horie, T. (2006). Cropping intensity and rainfall effects on upland rice yields in northern Laos. *Plant and soil*, 284, 175-185.

Sasaki, T., Matsumoto, T., Yamamoto, K., Sakata, K., Baba, T., Katayose, Y., Wu, J., Niimura, Y., Cheng, Z. & Nagamura, Y. (2002). The genome sequence and structure of rice chromosome 1. *Nature*, 420, 312-316.

Seki, M., Narusaka, M., Abe, H., Kasuga, M., Yamaguchi-shinozaki, K., Carninci, P., Hayashizaki, Y. & Shinozaki, K. (2001). Monitoring the expression pattern of 1300 *Arabidopsis* genes under drought and cold stresses by using a full-length cDNA microarray. *The Plant Cell Online*, 13, 61-72.

Sohrabi, M., Rafii, M., Hanafi, M., Siti NOR Akmar, A. & Latif, M. (2012). Genetic Diversity of Upland Rice Germplasm in Malaysia Based on Quantitative Traits. *The Scientific World Journal*, 2012.

Sun, C., Fang, J., Zhao, T., Xu, B., Zhang, F., Liu, L., Tang, J., Zhang, G., Deng, X. & Chen, F. (2012). The histone methyltransferase SDG724 mediates H3K36me2/3 deposition at *MADS50* and *RFT1* and promotes flowering in rice. *The Plant Cell Online*, 24, 3235-3247.

Takahashi, Y., Teshima, K., Yokoi, S. & Innan, H. Shimamoto k (2009) Variations in Hd1 proteins, *Hd3a* promoters, and *Ehd1* expression levels contribute to diversity of flowering time in cultivated rice. *Proceedings of the National Academy of Sciences, USA*, 106, 4555-4560.

Tamaki, S., Matsuo, S., Wong, H. & Yokoi, S. Shimamoto (2007) Hd3a protein is a mobile flowering signal in rice. *Science*, 316, 1033-1036.

Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. (2013). Mega6: molecular evolutionary genetics analysis version 6.0. *Molecular biology and evolution*, 30, 2725-2729.

Thurber, C. S., Reagon, M., Olsen, K. M., Jia, Y. & Caicedo, A. L. (2014). The evolution of flowering strategies in US weedy rice. *American Journal of Botany*, 101, 1737-1747. Toki, S. (1997). Rapid and efficient Agrobacterium-mediated transformation in rice. *Plant Molecular Biology Reporter*, 15, 16-21.

Tsuji, H., Tamaki, S., Komiya, R. & Shimamoto, K. (2008). Florigen and the photoperiodic control of flowering in rice. *Rice*, 1, 25-35.

Tsuji, H., Taoka, K.-I. & Shimamoto, K. (2011). Regulation of flowering in rice: two florigen genes, a complex gene network, and natural variation. *Current opinion in plant biology*, 14, 45-52.

Tyagi, A. K. & Mohanty, A. (2000). Rice transformation for crop improvement and functional genomics. *Plant Science*, 158, 1-18.

Vance, C., Rogelj, B., Hortobágyi, T., De Vos, K. J., Nishimura, A. L., Sreedharan, J., Hu, X., SMITH, B., Ruddy, D. & Wright, P. (2009). Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science*, 323, 1208-1211.

Wang, J.-D., Lo, S.-F., Li, Y.-S., Chen, P.-J., Lin, S.-Y., Ho, T.-Y., Lin, J.-H. & Chen, L.-J. (2013). Ectopic expression of OsMADS45 activates the upstream genes Hd3a and RFT1 at an early development stage causing early flowering in rice. *Botanical Studies*, 54, 12.

Yamamoto, T., Lin, H., Sasaki, T. & Yano, M. (2000). Identification of heading date quantitative trait locus Hd6 and characterization of its epistatic interactions with *Hd2* in rice using advanced backcross progeny. *Genetics*, 154, 885-891.

Yano, M., Katayose, Y., Ashikari, M., Yamanouchi, U., Monna, L., FUSE, T., Baba, T., Yamamoto, K., Umehara, Y. & Nagamura, Y. (2000). Hd1, a major photoperiod sensitivity quantitative trait locus in rice, is closely related to the Arabidopsis flowering time gene CONSTANS. *The Plant Cell Online*, 12, 2473-2483.

Yano, M., Kojima, S., Takahashi, Y., Lin, H. & Sasaki, T. (2001). Genetic control of flowering time in rice, a short-day plant. *Plant Physiology*, 127, 1425-1429.

Ye-yang, F., Chen, C., Ji-rong, W., Shi-hua, C. & Jie-yun, Z. (2011). Quantitative Trait Loci for Yield Traits Located Between *Hd3a* and *Hd1* on Short Arm of Chromosome 6 in Rice. Rice Science, 18(14)

Yu, J., Hu, S., Wang, J., Wong, G. K.-S., Li, S., Liu, B., Deng, Y., Dai, L., Zhou, Y. & Zhang, X. (2002). A draft sequence of the rice genome (*Oryza sativa L*. ssp. *indica*). *science*, 296, 79-92.

Zhao, J., Huang, X., Ouyang, X., Chen, W., DU, A., Zhu, L., Wang, S., Deng, X. W. & LI, S. (2012). *OsELF3-1*, an ortholog of *Arabidopsis EARLY FLOWERING 3*, regulates rice circadian rhythm and photoperiodic flowering. *PloS one*, 7, 3705-3710.

Zhao, K., Wright, M., Kimball, J., Eizenga, G., Mcclung, A., Kovach, M., Tyagi, W., Ali, M. L., Tung, C.-W. & Reynolds, A. (2010). Genomic diversity and introgression in *O. sativa* reveal the impact of domestication and breeding on the rice genome. *PLoS One*, *5*, 780-785.